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OPEN Sex differences in aggression and its neural substrate in a cichlid fish

Lillian R. Jackson^{1,3}, Mariam Dumitrascu¹ & Beau A. Alward^{1,2,3}✉

Aggression is ubiquitous among social species and can function to maintain social dominance hierarchies. The African cichlid fish *Astatotilapia burtoni* is an ideal study species for studying aggression due to their dominance hierarchy and robust behavioral repertoire. To further understand the potential sex differences in aggression in this species, we characterized aggression in male and female *A. burtoni* in a mirror assay. We then quantified neural activation patterns in brain regions of the social behavior network (SBN) to investigate if differences in behavior are reflected in the brain with immunohistochemistry by detecting the phosphorylated ribosome marker phospho-S6 ribosomal protein (pS6), a marker for neural activation. We found that *A. burtoni* perform both identical and sex-specific aggressive behaviors in response to a mirror assay. Females had greater pS6 immunoreactivity than males in the Vv (ventral part of the ventral telencephalon), a homolog of the lateral septum in mammals. Males but not females had higher pS6 immunoreactivity in the ATn after the aggression assay. The ATn (anterior tuberal nucleus) is a homolog of the ventromedial hypothalamus in mammals, which is strongly implicated in the regulation of aggression in males. Several regions also have higher pS6 immunoreactivity in negative controls than fish exposed to a mirror, implicating a role for inhibitory neural processes in suppressing aggression until a relevant stimulus is present. Male and female *A. burtoni* display both similar and different behavioral patterns in aggression in response to a mirror assay. There are also sex differences in the corresponding neural activation patterns in the SBN. In mirror males but not females, the ATn clusters with the POA, revealing a functional connectivity of these regions that is triggered in an aggressive context in males. These findings suggest that distinct neural circuitry underlie aggressive behavior in male and female *A. burtoni*, serving as a foundation for future work investigating the molecular and neural underpinnings of sex differences in behavior in this species to reveal fundamental insights into understanding aggression.

Aggression is a complex social behavior defined as agonistic acts directed towards others and is used to resolve conflict, maintain social dominance hierarchies, gain access to mates, and defend resources like shelter and food^{1,2}. Although aggression is metabolically costly and oftentimes risky³, it exists in nearly all species⁴. Extensive knowledge has been gained on the molecular and neural mechanisms of aggression in both sexes; however, male intrasexual aggression has been studied more extensively in this regard^{4–7}. Moreover, our most in-depth understanding of the mechanisms of aggression come from a few traditional model organisms such as mice, rats, and fruit flies^{4,8}. To gain a comprehensive understanding of the molecular and neural control of behavior, models are needed in which (1) both sexes perform aggression in similar contexts and, if possible, (2) both sexes perform the same behaviors in those similar contexts⁹. Investigating the same behavior in both sexes allows us to identify the fundamental factors influencing aggression and to further understand processes that shape the neural and molecular basis of social behaviors. However, these goals have been challenging to achieve in current models for studying the molecular and neural control of aggression.

The African cichlid fish *Astatotilapia burtoni* may be an especially useful species in which to gain fundamental insights into the sex differences in aggression and its molecular and neural control^{10–15}. *A. burtoni* exhibit dynamic social interactions and decades of field and laboratory studies have revealed novel insights into the mechanisms governing social behavior. *A. burtoni* are a lekking species in which males will court and mate with females and attack and defend a territory against other males¹⁶. Male *A. burtoni* exist in two dominance states that can be transitioned between depending on the social environment¹⁷. Dominant males are brightly colored, have high levels of circulating sex steroid hormones (androgens and estrogens), and perform reproductive and aggressive behaviors. Aggressive behaviors such as attacks and lateral displays are distinct and easily identifiable in behavioral assays (Fig. 1). In contrast, subordinate males are drably colored, have low levels of circulating sex steroid hormones, and perform submissive behaviors^{18,19}.

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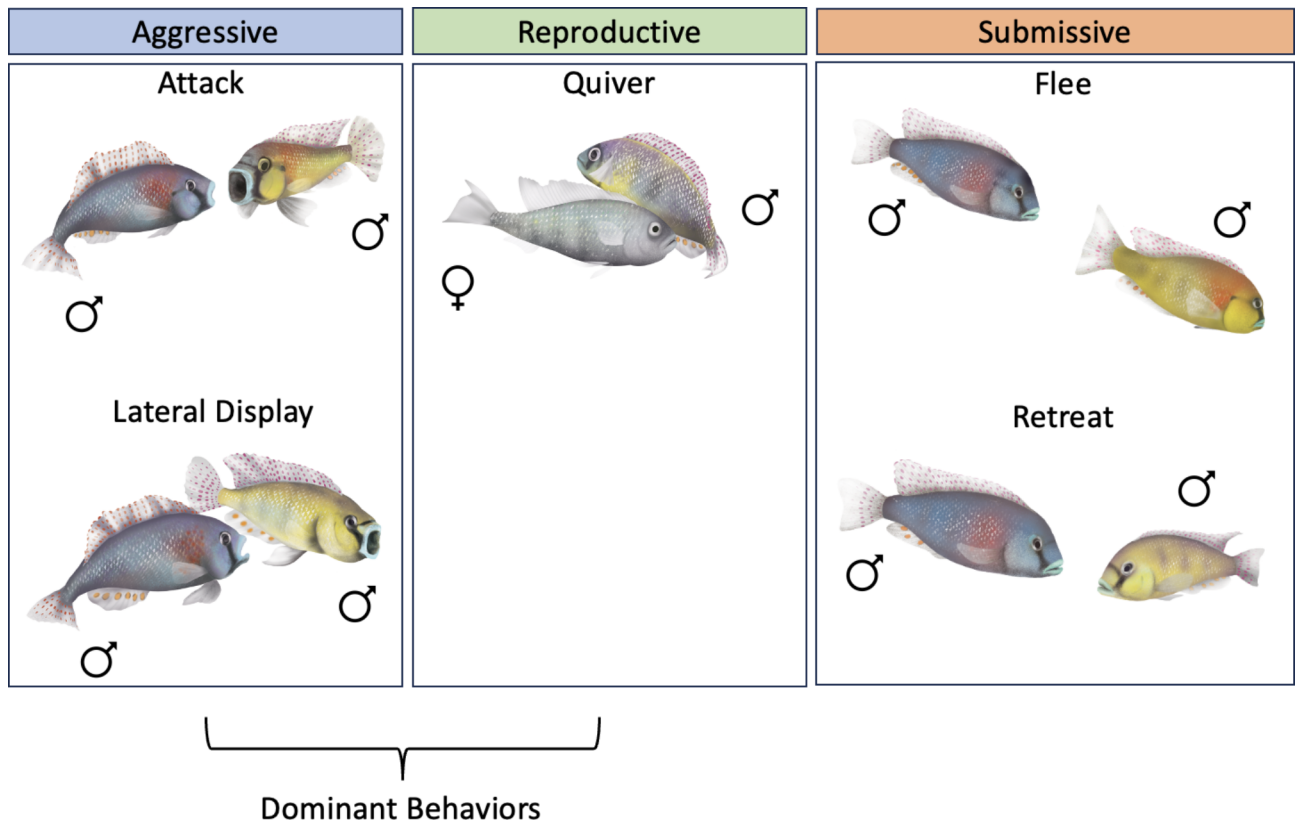


Fig. 1. Typical behaviors characterized in *A. burtoni*. Aggressive and reproductive behaviors are typically performed by dominant males. Submissive behaviors are performed by subordinate males.

Female *A. burtoni* do not naturally form a social hierarchy and typically shoal with other females. However, when in all-female communities, females acquire certain male-typical dominance traits including enhanced aggression²⁰, suggesting some mechanisms regulating male-typical dominance behaviors are present in females. Much like the dominance hierarchy in male *A. burtoni*, female *A. burtoni* dominance is plastic and reversible, with dominant females directing aggressive behaviors like chasing and lateral displays towards subordinate females²⁰. Still, outside of all-female communities, aggressive behavioral patterns in female *A. burtoni* have not been characterized. Aggression in both sexes has, however, been studied in many other teleost species for non-breeding territorial aggression, in which aggression does not differ between males and females. For example, *Gymnotus omarorum* males and females show no differences in aggressive behaviors^{21,22} and in *Stegastes nigricans*, the level of aggression does not differ between males and females but territorial behavior differs during the reproductive period^{23,24}, in which females frequently leave their territories but males remain in theirs to guard eggs.

To definitively determine which behaviors *A. burtoni* females use during aggressive interactions, and whether they are similar to male aggressive behaviors, requires assays in which aggression is displayed by both sexes that is readily observable and quantifiable. Here, we characterize aggressive behaviors in male and female *A. burtoni* using a mirror assay. In *A. burtoni* and other model organisms like zebrafish, mirrors elicit aggressive behavioral responses because the fish treats its reflection as another fish^{25,26}. We reasoned that mirror assays would be especially useful for studying aggression in *A. burtoni* since males alter their aggression if an intruder male is larger, smaller, or the same size (within 5% of other males) in length²⁷. Therefore, to successfully compare aggression in males and females requires the use of a paradigm like the mirror assay, which controls for size given that their reflection should be treated as a fish identical in size to it. However, it is important to note the evidence that cichlids and other fish do not treat a mirror exactly like a live opponent including behavioral responses and neural patterns^{26,28}. We argue that since mirrors are able to elicit robust behavioral responses in *A. burtoni* and we are interested in the mechanisms of the behavioral acts themselves, that it is appropriate to use a mirror assay in this study.

In fish included in the mirror assay, we also investigated neural activation patterns as measured by phosphorylated ribosomes throughout brain regions of the social behavior network (SBN) and the social decision-making network (SDMN), two conserved, interconnected circuits critical for integrating social cues and modulating social behaviors^{29,30}. The phosphorylation of ribosomal protein S6 (pS6) occurs following the activation of neurons via signaling cascades, reflecting an upregulation of protein synthesis^{31–34}. We hypothesized that (1) males and females would perform different aggressive behaviors during a mirror assay and that these differences would be reflected as (2) sex differences in neural activation patterns. Our findings

show suites of convergent and divergent behavioral and neural sex differences in the control of aggression in *A. burtoni*, suggesting independent mechanisms mediate sex differences in behavior in this species in the mirror assay. These results highlight the importance of studying sex differences in diverse organisms to illuminate the fundamental mechanisms underlying the generation of social behavior and suggest *A. burtoni* may be a useful model for understanding the molecular and neural control of sex differences in aggression.

Methods

Animal subjects

Adult *A. burtoni* were bred from a wild-caught stock that originated from Lake Tanganyika, Africa collected in the 1980s^{16,35} and housed in environmental conditions that mimic their natural equatorial habitat (28 °C; pH 8.0; 12:12 h light/dark cycle with full spectrum illumination; constant aeration; debris filtration). Aquaria contained gravel-covered bottoms with terra cotta pots cut in half to serve as shelters. Fish were fed cichlid flakes daily. All experimental procedures were in accordance with Association for Assessment and Accreditation of Laboratory Animal Care standards and approved by the University of Houston Institutional Animal Care and Use Committee (Protocol #202000001). All experimental procedures followed the appropriate ARRIVE guidelines for reporting on experiments involving animals.

Mirror assay behavior recording

Male ($n=8$) and female ($n=9$) *Astatotilapia burtoni* were housed in a pre-assay tank for approximately two weeks prior to being assayed in a mirror assay. One male was removed from the sample due to a corrupted video file. Fish were 7 months – 1 year old at the time of the assay and ranged from in 4.2–6.2 cm in standard length. The pre-assay tank consisted of a 60.5 L plexiglass tank with a clear perforated divider that separated males and females on separate halves to allow males and females to interact but prevent them from mating. Two terra cotta pot shelters were placed on each side of the pre-assay tank. Subjects were housed in groups of 6 fish at a time, including 3 gravid females and 3 dominant males on each side. On average, male triads deviated 0.2 cm in standard length and female triads deviated 0.275 cm in standard length. After two weeks in the pre-assay tank, each subject's behaviors were recorded in a mirror assay. One dominant male and one female were assayed each day until all the subjects in a pre-assay group were assayed (Fig. 2).

The mirror assay tank consisted of a white tub (Sterilite; 400 mm x 318 mm x 12 mm) filled 6 cm deep with UV-sterilized aquaria water with a mirror replacing one side of the assay tank. An opaque cover was first placed on the mirror for a 15-minute habituation period to allow subjects to acclimate to the assay tank. The opaque cover was then removed, and 30 min of behavior was recorded. The water in the mirror assay tank was exchanged between every assay. We also included negative control males ($n=5$) and females ($n=8$) in the same exact pre-assay conditions except the mirror cover was not lifted after the 15-minute habituation period. These negative control fish were held in separate pre-assay tanks than mirror fish. Immediately following the mirror assay, subjects were euthanized in an ice bath for 1–2 min before rapid cervical transection followed immediately by the harvesting of tissue.

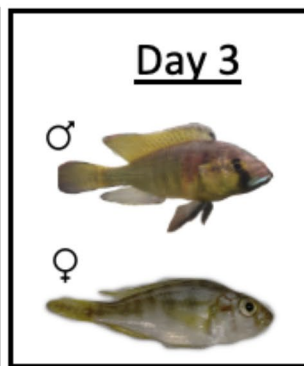
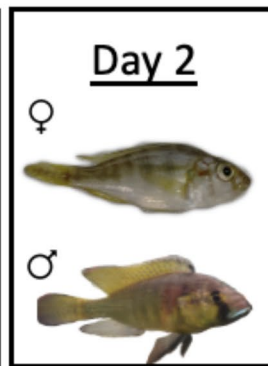
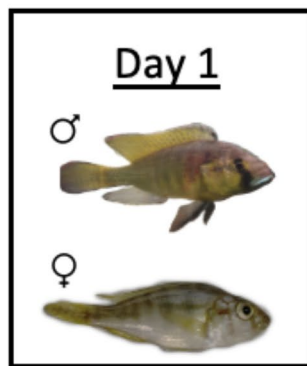
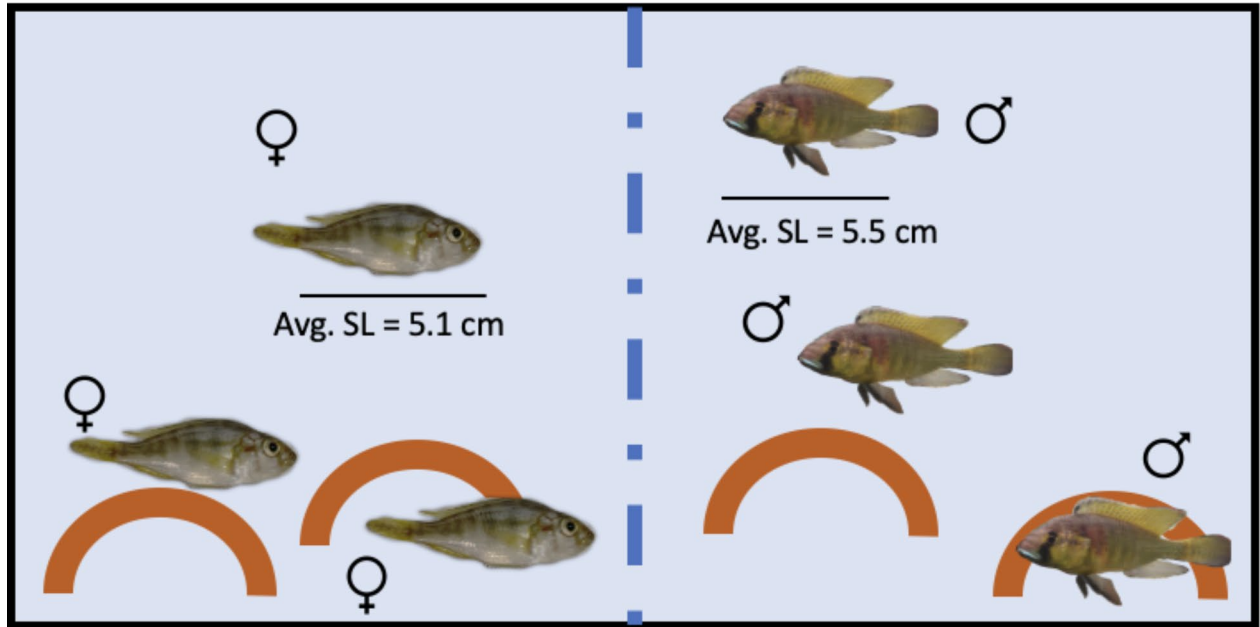
Behavioral analysis

Behavior was recorded using a digital video camera and was quantified using the freely available BORIS (Behavioral Observation Research Interactive Software) version 7.13.9³⁶. Multiple types of behavior were quantified: behaviors considered not to be aggressive or “neutral” (tap, graze); subordinate behaviors (flee, retreat); aggressive behaviors (attack, lateral display, rostral display); and a male-typical reproductive behavior (quiver) (Fig. 1). Taps were defined as making gentle mouth contact with the mirror. Grazing was defined as swimming alongside the mirror for at least 3 s. Fleeing was defined as an abrupt turn and swim away from the mirror. Retreats were defined as a rapid retreat swim from the mirror. Attacks were defined as a rapid swim towards the mirror, making mouth contact, and vigorously wiggling the body. Lateral displays were defined as the presentation of the side of the body towards the mirror, conforming the body at times in a slightly convex shape while lateral to the mirror with erect fins. Lateral displays were further separated into type 1 and type 2 lateral displays to distinguish between more and less vigorous lateral displays respectively. Rostral displays were defined as flaring the opercula towards the mirror without making direct contact with the mirror. Quivers were defined as a rapid vibration of the body towards the mirror while conforming in a convex shape. A group of behaviors were summed together for their mouth movements, termed mouth contact (summed attacks, taps, and rostral displays). Additionally, total behaviors were calculated as the sum of all behaviors and total aggression was the sum of all aggressive behaviors (attacks, lateral displays, rostral displays, and quivers). Furthermore, another behavior was identified that appears to be associated with aggression that we have termed an s-curve display, defined as a curved body conformation facing the mirror at a distance (Table 1).

Tracking and activity data was collected on pre-mirror and mirror assay videos using Noldus EthoVisionXT software. Center-point detection was used to track the subjects throughout the assay tank. Distance moved (cm) was calculated as the accumulated distance the subject moved throughout the assay duration. Velocity (cm/s) was calculated as distance over time. Activity was measured as the average % of pixels changed over time. We defined a zone close to the mirror to measure the proximity of subjects near the mirror, extending 2.4 cm out from the mirror. Zone duration (s) was calculated as the cumulative duration of time spent in the zone. Zone frequency was calculated as the number of times the center-point of the subject entered the zone. Zone latency was defined as the lapse of time until the zone is first entered. Total and mean distances to the zone (cm) were calculated as the cumulative and mean distances from the zone respectively.

A series of dyad assays similar to that used recently in male *A. burtoni*²⁷ were also performed to ensure that female aggressive behaviors observed in the mirror assay were also performed towards a live opponent. Gravid resident females ($n=5$) were housed in a 20.8 L glass tank containing gravel, half of a terra cotta pot,

Pre-assay: 2 weeks



Mirror Assay

Assay 1 ♂ and 1 ♀ each day (alternating order)

Fig. 2. Pre-assay schematic for negative control and mirror fish. Males and females were housed in a tank separated by a perforated divider for 2 weeks. Following this pre-assay period, males and females were assayed in pairs on consecutive days in a mirror assay.

and constant aeration for two days before being introduced to a gravid intruder female. The intruder female was an unfamiliar conspecific that was size-matched within 5% of the resident's standard length. After the intruder female was added to the tank, the following interaction was recorded for one hour and aggressive behaviors performed by the resident female were quantified.

Inter-behavioral intervals

We also calculated inter-behavioral intervals as described in³⁷. The log files containing the results from BORIS were analyzed for inter-behavior intervals (IBI) using custom R software (available at <https://github.com/AlwardLab>). IBI were determined by calculating the time between successive behaviors and averaging across all IBIs within a given log file. Negative control fish performed almost no social behavior so were not included in analyses of temporal patterns of behavior.

Morphological and steroid hormone analysis

Subjects were assessed for standard length (SL), body mass (BM), gonad mass, and gonadosomatic index [GSI = (gonad mass / body mass) * 100]. Average male SL is 5.5 cm and average female SL is 5.1 cm. Male GSI ranged from 0.324 to 1.813 (average GSI = 0.895) and female GSI ranged from 1.074 to 9.122 (average GSI = 5.903). Blood samples were collected with capillary tubes from the caudal vein, centrifuged for 10 min at 5,900 rcf, and the plasma was removed and stored at -80 °C until assayed.

| Behavior | Description | Category |
|------------------------|--|-------------|
| Tap | Fish makes gentle mouth contact with the mirror | Neutral |
| Graze | Fish swims alongside the mirror for at least 3 s | Neutral |
| Flee | Fish makes an abrupt turn and swim away from the mirror | Subordinate |
| Retreat | Fish rapidly retreats from the mirror | Subordinate |
| Jump | Fish moves body out of water | |
| Attack | Fish rapidly swims towards the mirror, making mouth contact, and vigorously wiggling of the body | Aggressive |
| Lateral Display | Fish presents the side of the body towards the mirror, conforming the body at times in a slightly convex shape while lateral to the mirror with erect fins | Aggressive |
| Lateral Display Type 1 | More vigorous lateral display | Aggressive |
| Lateral Display Type 2 | Less vigorous lateral display | Aggressive |
| Rostral Display | Fish flares the opercula towards the mirror without making direct contact with the mirror | Aggressive |
| Quiver | Fish makes a rapid vibration of the body towards the mirror while conforming in a convex shape | Aggressive |
| Mouth contact | Summed number of attacks, taps, and rostral displays | |
| Total behaviors | Sum of all behaviors | |
| Total aggression | Summed number of attacks, lateral displays, rostral displays, and quivers | Aggressive |
| S-curve Display | Fish makes a curved body conformation facing the mirror at a distance | |

Table 1. Ethogram used to score behaviors in BORIS.

Plasma 11-ketotestosterone (11-KT) levels were measured using a commercially available enzyme immunoassay (EIA) kit (Cayman Chemical Company, Ann Arbor, MI, USA). This kit has been extensively validated for use in *A. burtoni*³⁸. For the 11-KT assay, a 2- μ l sample of plasma from each subject was extracted two times using 200 μ l of ethyl ether and evaporated under a fume hood before re-constitution in EIA assay buffer (1:40 dilution). EIA kit protocols were then strictly followed, plates were read at 405 nm using a microplate reader (Biorad), and steroid concentrations were determined based on standard curves. All samples were assayed in duplicate. Intra and inter-assay variability were 13.5% and 10.9%, respectively.

Immunohistochemistry for pS6

Following the cervical transection of subjects, brains were fixed in 4% paraformaldehyde in PBS (pH = 7.4) for 1–2 h and then transferred to 30% sucrose in PBS. Brains were kept in the sucrose solution at 4 °C until they sunk and then embedded in mounting media Neg50 and stored at -80 °C. Brains were sectioned at 30 μ m. We used the representative transverse sections described in Maruska et al.³⁹ as a guide for the sectioning plane of our tissue³⁹. We analyzed neural activation in the brain following an aggression assay vs. a negative control condition using an immunohistochemistry protocol to detect the phosphorylated ribosome marker pS6, similar to that described in Butler et al.⁴⁰.

We performed immunohistochemistry to detect pS6, a proxy of neural activation in the brain⁴¹ that has been used previously in *A. burtoni* brains^{40,42}. After sectioning, the slides were dried in a desiccator for 48 h, then stored at -80 °C. Slides were then dried again in the desiccator for 48 h prior to the immunohistochemistry protocol. Slides were outlined with a hydrophobic barrier using a PAP pen and then immersed in boiling citric acid (10 mM, pH = 6) for 5 min twice. The slides were then rinsed with 1x PBS (pH = 7.4) for 5 min three times. Nonspecific binding was blocked by incubating slides in 1x PBS containing 5% donkey serum and 0.3% Triton X-100 (PBS-T) for 30 min at room temperature. Then slides were incubated in PBS-T containing 0.5% donkey serum with the pS6 primary antibody (1:300, Invitrogen™ Phospho-S6 (Ser244, Ser247) polyclonal antibody) overnight at room temperature. Following the primary antibody incubation, slides were rinsed with 1x PBS for 5 min three times. Slides were then incubated in PBS-T containing 0.5% donkey serum with the donkey anti-rabbit secondary antibody (1:300, Invitrogen Alexa Fluor™ 488) at room temperature for 1 h. The slides were rinsed with DAPI (1:500, Sigma-Aldrich) diluted in PBS for 5 min and then coverslipped with Aqua-Poly/Mount and stored at 4 °C. 9 immunohistochemistry staining batches were run and groups were balanced in each batch.

Quantification of pS6 immunoreactivity

Stained slides for the pS6 immunohistochemistry were imaged using a Nikon Eclipse 80i Microscope MicroFire™ at 20x magnification in GFP and DAPI and quantified in regions of the social behavior network (SBN). The social behavior network consists of interconnected nuclei that are implicated in the production of social behavior^{29,30}. We quantified pS6 immunoreactivity in the following brain regions of the SBN: the ventral part of the ventral telencephalon (Vv) (lateral septum homolog), the supracommissural part of the ventral pallidum (Vs) (medial amygdala and bed nucleus of the stria terminalis partial homolog), subpopulations of the preoptic area (POA): parvocellular preoptic nucleus, anterior part (nPPa), magnocellular preoptic nucleus, magnocellular division (nMMp), magnocellular preoptic nucleus, parvocellular division (nPMp); anterior tuberal nucleus (ATn) (ventromedial hypothalamus homolog), and the periaqueductal gray / central gray (PAG/CG). We additionally quantified pS6 immunoreactivity in the vagal lobe (VL), vagal motor nucleus (Xm), medial part of the dorsal telencephalon, subdivision 3 (Dm-3), and the lateral nucleus of the dorsal telencephalon, granular region (DI-g) due to the involvement of these regions in either processing sensory information or the social decision-making network, which is thought to evaluate the salience of social cues³⁰. Each brain area was identified

with neuroanatomical landmarks present in the DAPI staining and with reference to approximate locations of nuclei described in Maruska et al.³⁹ (Fig. 3). The PAG is notably difficult to identify based on a lack of reliable neuroanatomical markers and pS6 cell density quantified in this region may not be completely representative of the PAG.

The number of pS6 positive cells were quantified for each region of interest (ROI) using ImageJ. A polygon was drawn around the ROI in each hemisphere and each pS6 positive cell within the ROI was counted manually. The cell density for each ROI was calculated per hemisphere (cells/ μm^2) and then the cell density was summed between the hemispheres. This total cell density measure was averaged across two consecutive sections for each region per subject, and the consecutive sections were selected at random using a random number generator. If the subject did not have two consecutive sections for a ROI, the cell density was taken from one section at random. Due to tissue adhesion issues and damage, pS6 cell density in some brain regions could not be quantified for every fish leading to differing sample sizes for the ROI. Unless otherwise noted, this is why sample sizes per group vary for certain ROIs in the Results. Additionally, the three subregions of the preoptic area were averaged together for a total POA measure. The experimenter who performed the cell counting was not blinded to group membership.

Statistical analysis

All statistical tests were performed using Graphpad Prism version 10. Two-way ANOVAs were used to compare (1) male and female aggressive behaviors over time in the mirror assay and (2) effects of sex and mirror condition on behaviors and pS6 immunoreactivity between negative control and mirror males and females. For the analysis of male and female aggressive behaviors over time, behaviors were binned into the habituation period (Pre) and 10-minute periods for the mirror assay. Following a significant main effect or interaction for an ANOVA, Šidák's test was used for multiple comparisons. When normality and equality of variance assumptions were not met, a log transformation of the data was performed before conducting the two-way ANOVA. If log transformations did not correct the data to meet assumptions, we still conducted a two-way ANOVA on the log-transformed data for consistency; this only occurred for two dependent variables (activity and pS6 immunoreactivity in the ATn). We also performed Fisher's tests on the frequency of performance of rostral displays and quivers. Effects were considered significant at $p \leq 0.05$. For several behaviors, log transformations were not feasible due to zeros in the negative control groups. For these behaviors, two-way ANOVAs were conducted, and additional chi-square tests were run. Pearson correlations were used to test for correlations between pS6 immunoreactivity of each ROI for mirror and negative control fish. Missing values were interpolated across all conditions. Heat maps of Pearson correlation coefficients of pS6 immunoreactivity in the ROI for each sex and condition were generated

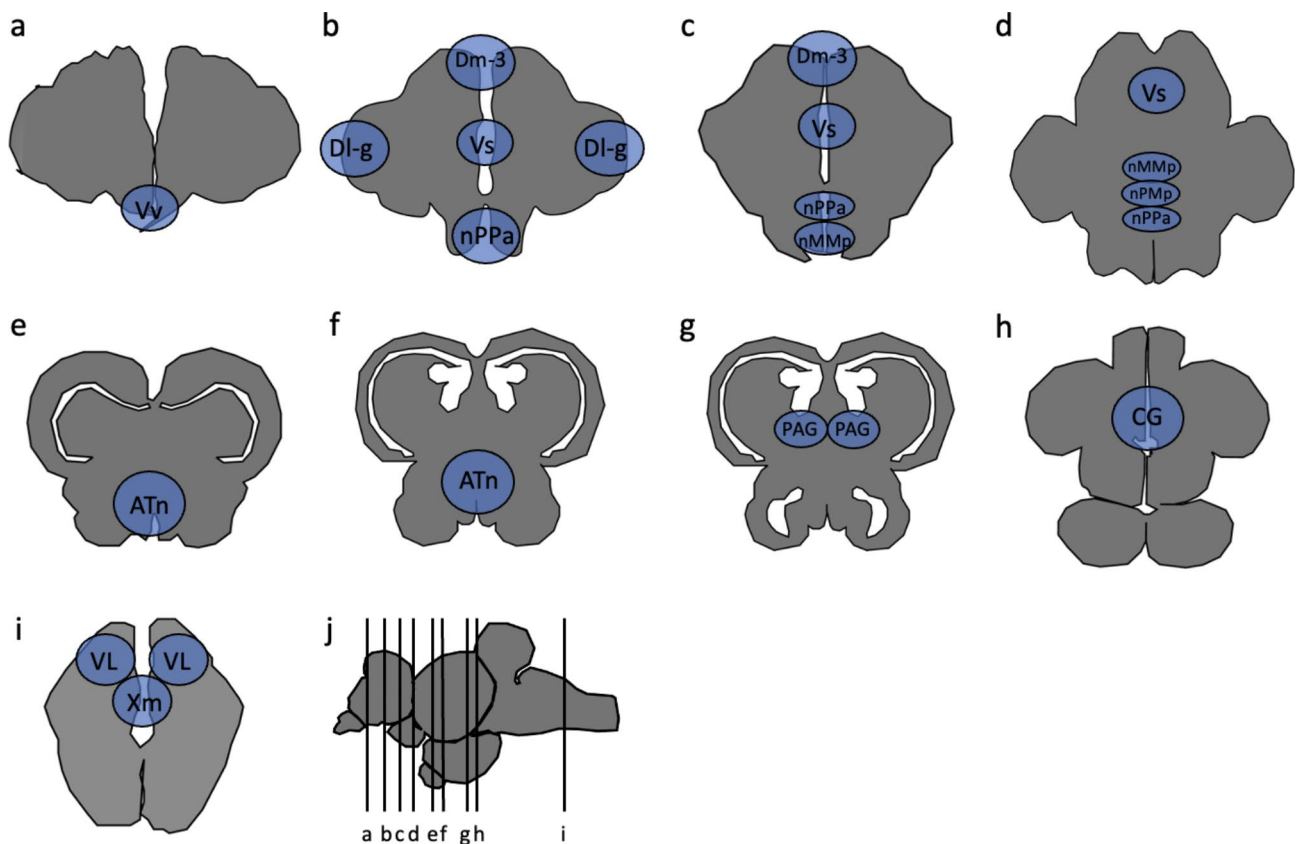


Fig. 3. (a–i) Drawn transverse sections with approximate locations of ROI. (j) Approximate locations of section in the *A. burtoni* brain.

in R (version 4.3.1, R Core Team 2023)⁴³. A principal components analysis (PCA) on all pS6 + cell count data was performed for mirror and negative control conditions. Individual principal component scores were obtained by projecting a new vector onto the PCA space and then correlated to aggressive behaviors in mirror males and females using Spearman correlations. Significant principal components were determined based on the flattening of the scree plot curves.

Results

Male and female *A. burtoni* perform the same and different aggressive behaviors towards a mirror

Contour maps of fish movement over time that were generated in Noldus showed that before the opaque cover was lifted fish swim around the edges of the assay arena (Fig. 4a). However, fish spent the vast majority of their time in front of the mirror once the cover was removed.

There was a significant effect of time but not sex or an interaction on attacks (Table 2; attacks, Two-way ANOVA, time: $p < 0.0001$). There was a significant effect of sex and time but not an interaction on lateral displays (Table 2; lateral displays, Two-way ANOVA, sex: $p = 0.0399$, time: $p = 0.0126$), wherein males performed more lateral displays than females (See videos S3-S4). We also observed a significant effect of sex on rostral displays (Table 2; rostral displays, Two-way ANOVA, sex: $p = 0.0075$), where males produced more rostral displays than females (See video S2). Additionally, there was a significant effect of sex on quivers (Table 2; quivers, Two-way ANOVA, sex: $p = 0.0413$). Specifically, females performed more quivers than males (See example in video S1). Fisher's exact tests showed more males than females performed rostral displays (6/8 males versus 1/9 females; $p = 0.0152$) while more females than males performed quivers (2/8 males versus 8/9 females; $p = 0.0152$). We were surprised to observe females perform a male-typical mating behavior, quivers, in the mirror assay, given the behavior is putatively used as an aggressive display in a mirror assay by females. To confirm females do these behaviors towards another female in a live interaction, we conducted resident intruder tests with focal resident females against intruder females²⁷. Female residents performed quivers, lateral displays, and attacks towards intruder females and also chased them (Fig. S1), indicating quivers are a normal component of the behavioral repertoire of female-female aggression. Across 5 resident females, attacks ranged from 2 to 56 (Average: 22.6, St. Dev.: 22.15), lateral displays ranged from 1 to 30 (Average: 12.8, St. Dev.: 10.76), and quivers ranged from 0 to 9 quivers (Average: 3.2, St. Dev.: 3.77) (Fig. 4).

There were no sex differences in median or mean IBI between mirror males and females (Fig. S2a; Median IBI, Unpaired t-test, $p = 0.1724$, $t = 1.433$, $df = 15$, Fig. S2b; Mean IBI, Mann-Whitney, $p = 0.0927$, $U = 18$).

We compared the behaviors of the NC fish to mirror fish and found across all behaviors NC males and females performed no or very few acts of any behavior types (Fig. S3). Overall, Two-way ANOVAs revealed consistent effects of condition and not sex or an interaction on behaviors performed during the mirror assay (See Supplementary Results for detailed statistical findings; Table S1, Figures S3-S4). This shows the NC fish included in this assay are sufficient as a control group when comparing pS6 immunoreactivity to fish exposed to the mirror (Fig. 5).

There was a significant effect of condition but not sex or an interaction on latency to behave (Table 3; latency to behave, Two-way ANOVA, condition: $p = 0.0408$), wherein mirror males had a longer latency to behave than negative control males (Table 3; latency to behave, Šidák's test, Male Mirror – NC, $p = 0.0259$). There were no significant effects of sex, condition, or sex*condition on latency to aggression (Table 3; latency to aggression, Two-way ANOVA). Additionally, there were no significant effects of sex, condition, or sex*condition on distance moved and velocity (Table 3; distance moved and velocity, Two-way ANOVA of log transformation).

We observed a significant effect of condition but not sex or an interaction on activity (Table 3; activity, Two-way ANOVA of log transformation, condition: $p = 0.0013$). One negative control female outlier was removed based on the Grubb's method ($G = 4.353$). Mirror females have higher activity than negative control females and males follow this same pattern although not significant (Table 3, activity, Šidák's test, Female Mirror – NC, $p = 0.0072$). No significant effects of sex, condition or sex*condition were observed on zone frequency or zone latency (Table 3; zone frequency, Two-way ANOVA of log transformation; zone latency, Two-way ANOVA). There was a significant effect of condition but not sex or an interaction on zone duration (Table 3; zone duration, Two-way ANOVA, condition: $p = 0.0369$) and mirror females had a longer zone duration than negative control females while males did not differ by condition (Table 3, zone duration, Šidák's test, Female Mirror – NC, $p = 0.0285$).

Condition—but not sex or an interaction—had an effect on mean distance and total distance to the zone (Table 3; mean distance to the zone, condition: $p = 0.0032$; total distance to the zone, condition: $p = 0.0030$, Two-way ANOVA of log transformation). Mirror females had a higher mean and total distance to the zone than negative control females and males followed this pattern but was not statistically significant (Table 3, mean distance to the zone: $p = 0.0223$, total distance to the zone: $p = 0.0204$, Šidák's test).

Sex differences and similarities in neural activation patterns of the SBN after aggression

No significant effects of sex, condition, or sex*condition on pS6 immunoreactivity were observed in the POA or subregions nPPa and nPMp, as well as in the Dl-g, Dm-3, and Xm (Table S2, Figure S5; POA, nPPa, nPMp, Dl-g, Dm-3, Two-way ANOVA; Xm, Two-way ANOVA of log transformation). However, there was a significant effect of condition, but not sex or an interaction, on pS6 immunoreactivity in one subregion of the POA, the nMMp (Table 4; nMMp, Two-way ANOVA, condition: $p = 0.0400$) in which negative controls had higher pS6 immunoreactivity in this region on average, but this was not statistically significant. There was a significant effect of sex on pS6 immunoreactivity in the Vv, where females had higher pS6 immunoreactivity compared to males (Table 4; Vv, Two-way ANOVA, sex: $p = 0.0010$).

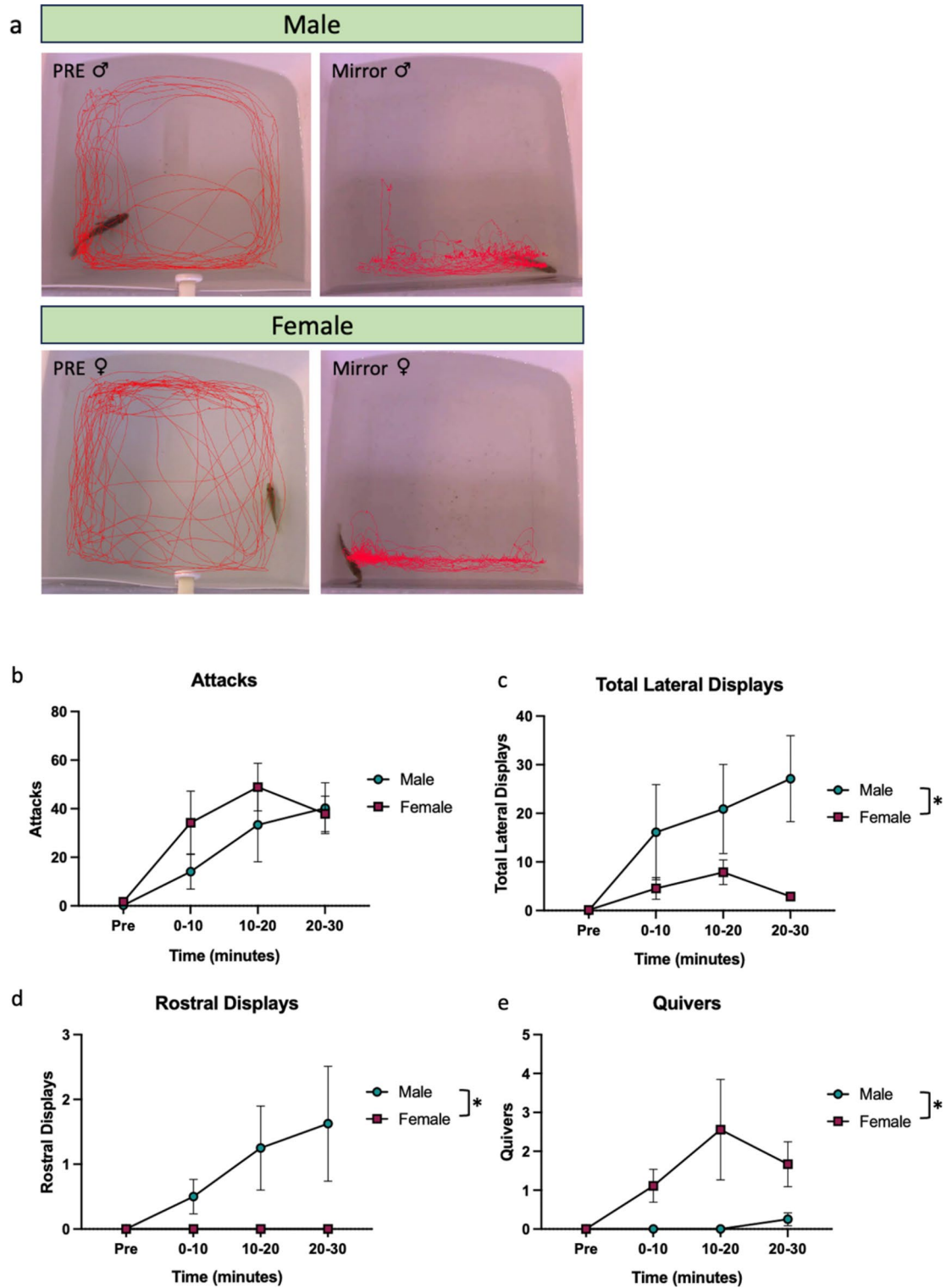


Fig. 4. Male-typical aggressive and courtship behaviors performed by males and females in the mirror assay. (a) Representative track visualization of male and female subjects before (PRE) and throughout (Mirror) a mirror assay trial. (b) Males and females both perform attacks in a mirror assay. (c) (d) Males perform more lateral displays and rostral displays than females. (e) Females perform more quivers than males.

We observed a significant effect of condition on pS6+ cells in the Vs, PAG/CG, and the VL (Table 4; Vs, condition: $p=0.0040$; CG, condition: $p=0.0009$; VL, condition: $p=0.0021$, Two-way ANOVA; PAG, condition: $p=0.0316$, Two-way ANOVA of log transformation). Specifically, in the Vs, PAG/CG, and the VL, female negative controls had higher pS6 immunoreactivity than mirror females while males did not differ by condition

| | Sex | | Time | | Sex*Time | |
|------------------|---------------|---------------|-----------------------|---------------|---------------|--------|
| | F | P | F | P | F | P |
| Attacks | 0.79 1, 15 | 0.3885 | 12.60 2,435, 36,53 | <0.0001 | 1.08 3, 45 | 0.3688 |
| Lateral displays | 5.06 1, 15 | 0.0399 | 4.62 2,353, 35,29 | 0.0126 | 2.40 3, 45 | 0.0805 |
| Rostral displays | 9.54 1, 15 | 0.0075 | 1.95 1,529, 22,93 | 0.1724 | 1.95 3, 45 | 0.1357 |
| Quivers | 4.98 1, 15 | 0.0413 | 3.04 1,306, 19,59 | 0.0882 | 2.76 3, 45 | 0.0530 |

Table 2. Effect of sex and time on aggressive behaviors quantified in a mirror assay.

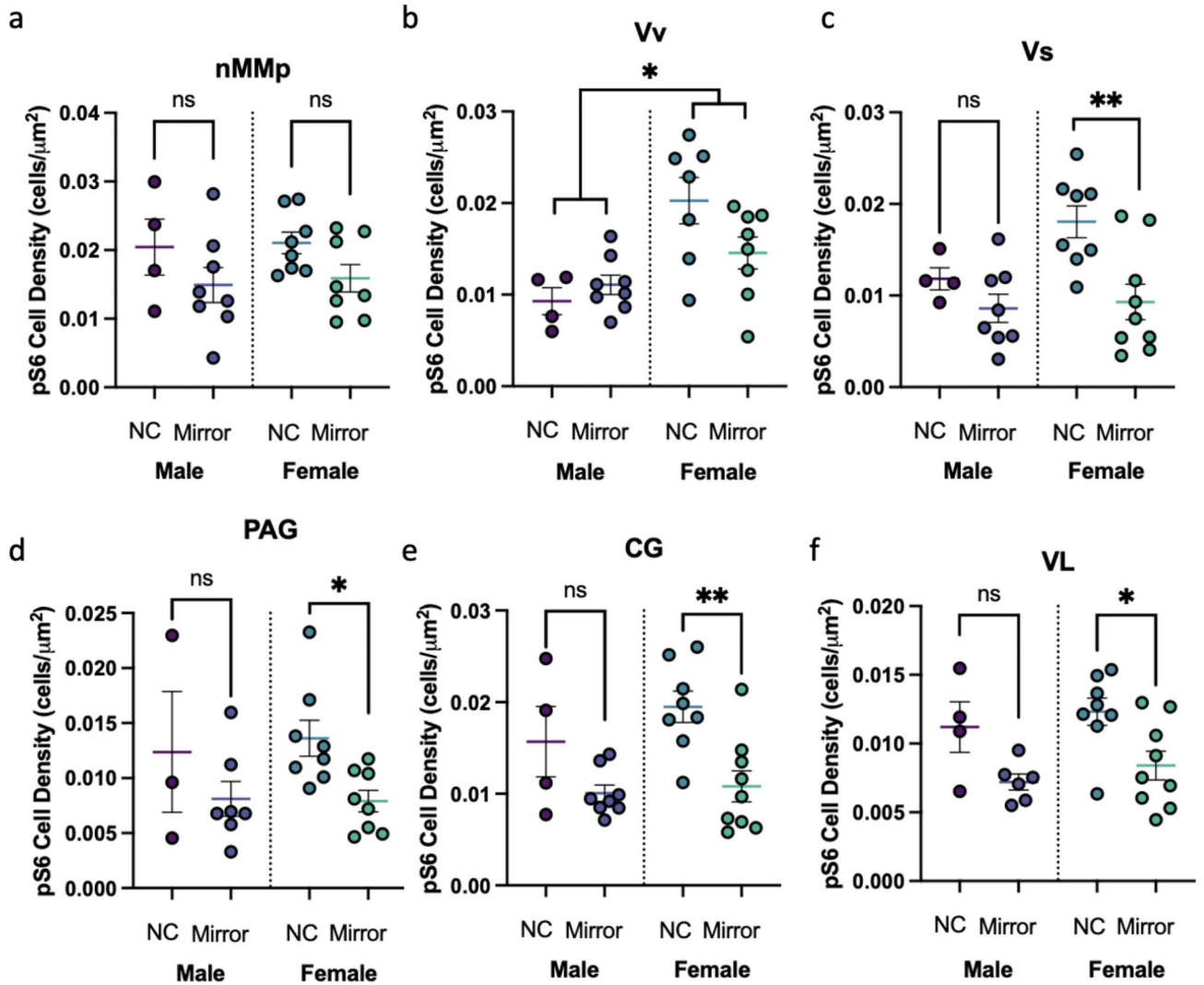


Fig. 5. Sex differences and similarities in neural activation patterns. (a) pS6 immunoreactivity differed by condition in the nMMP, but post-hoc comparisons did not reveal significant differences in pS6 cell density within each sex. (b) Females have higher pS6 immunoreactivity in the Vv than males. (c) (d) (e) (f) pS6 cell density is higher in negative control fish than in mirror fish in the Vs, PAG, CG, and the VL. Multiple comparisons reveal that pS6 cell density is higher in negative control females compared to mirror females for these regions. NC, negative control. ** $p < 0.01$, * $p < 0.05$. ns, not statistically significant.

| | Sex | | Condition | | Sex*Condition | |
|----------------------------|-------------------------|--------|---------------------------|---------------|---------------|--------|
| | F | P | F | P | F | P |
| (1) | | | | | | |
| Latency to behave | 1.62 1, 26 | 0.2149 | 4.63 1, 26 | 0.0408 | 3.63 1, 26 | 0.0679 |
| Latency to aggression | 0.03 1, 17 | 0.8680 | 0.14 1, 17 | 0.7141 | 2.74 1, 17 | 0.1165 |
| Distance moved | 0.03 1, 26 | 0.8537 | 0.07 1, 26 | 0.7970 | 0.06 1, 26 | 0.8036 |
| Velocity | 0.07 1, 26 | 0.7945 | 0.01 1, 26 | 0.9439 | 0.10 1, 26 | 0.7528 |
| Activity | 1.24 1, 25 | 0.2757 | 13.13 1, 25 | 0.0013 | 0.40 1, 25 | 0.5318 |
| Zone frequency | 0.11 1, 26 | 0.7472 | 0.04 1, 26 | 0.8358 | 0.75 1, 26 | 0.3929 |
| Zone latency | 2.38 1, 26 | 0.1351 | 2.12 1, 26 | 0.1573 | 2.26 1, 26 | 0.1452 |
| Zone duration | 3.09 1, 26 | 0.0907 | 4.84 1, 26 | 0.0369 | 1.45 1, 26 | 0.2392 |
| Mean distance to the zone | 1.70 1, 26 | 0.2036 | 10.53 1, 26 | 0.0032 | 0.09 1, 26 | 0.7699 |
| Total distance to the zone | 1.51 1, 26 | 0.2308 | 10.71 1, 26 | 0.0030 | 0.10 1, 26 | 0.7529 |
| | Male Mirror - NC | | Female Mirror - NC | | | |
| | P | | P | | | |
| (2) | | | | | | |
| Latency to behave | 0.0259 | | 0.9776 | | | |
| Activity | 0.1109 | | 0.0072 | | | |
| Zone duration | 0.7684 | | 0.0285 | | | |
| Mean distance to the zone | 0.1230 | | 0.0223 | | | |
| Total distance to the zone | 0.1224 | | 0.0204 | | | |

Table 3. (1) Effect of sex and condition on tracking and activity data quantified in a mirror assay. (2) post-hoc comparisons.

(Table 4, Vs: $p=0.0020$, PAG: $p=0.0408$, CG: $p=0.0024$, VL: $p=0.0181$, Šidák's tests). Mirror males and negative controls did not differ in pS6 immunoreactivity in the Vs, PAG, and CG (Table 4, Vs, PAG, CG, Šidák's tests). Although not statistically significant, male negative controls had higher pS6 immunoreactivity in the VL compared to mirror males (Table 4, VL, Šidák's test, Male Mirror - NC, $p=0.0773$), following the same trend as females in the VL.

A significant sex*condition interaction on pS6 immunoreactivity in the ATn was observed (Fig. 6, Table 4; ATn, Two-way ANOVA of log transformation, interaction: $p=0.0040$). Specifically, males exposed to the mirror had more pS6+ cells in the ATn compared to negative control males, while this was not true for females (Table 4, ATn, Šidák's test, Male Mirror - NC, $p=0.0313$).

No effects of sex or condition on 11KT

There were no significant effects of sex, condition or sex*condition on 11-KT (Table 4, 11-KT, Two-way ANOVA).

Correlated neural activity patterns differ in mirror-exposed and negative control fish

We correlated the pS6 immunoreactivity across regions of the SBN in mirror and negative control fish to investigate the functional connectivity of these regions in response to an aggressive assay. Principal components analysis (PCA) of neural activation patterns revealed two significant components that capture the variability in the data (Fig. 7). For the mirror fish, the first component (PC1, 35.4% explained variance) was primarily weighted by pS6+ cell count in the subregions of the POA (nPPa, nMMp, and nPMp) and the Dm-3 while the second component (PC2, 16.9% explained variance) was weighted primarily by pS6+ cell count in the CG, VL, DL-g, and the Xm and ATn. Variation in male neural activity is mostly captured across PC1, while female neural activation patterns are explained relatively equally across both components. Males and females largely overlap across PC1, but less so across PC2, indicating that PC2 may be more representative of sex differences in neural activation patterns than PC1 in the mirror fish. In the negative control fish, the first principal component (59.3% explained variance) is weighted by the Vs, POA (and subregion nPMp), and the Dm-3. The second component is driven by neural activation of the Vv, ATn, CG, VL, and Xm. Males and females are largely overlapping based on the biplot, suggesting that there are not sex differences in the neural activation patterns captured by PC1 and PC2 in the negative control fish.

| | Sex | | Condition | | Sex*Condition | |
|-------|-------------------------|---------------|---------------------------|---------------|----------------|---------------|
| | F | P | F | P | F | P |
| (1) | | | | | | |
| nMMp | 0.10 1, 24 | 0.7492 | 4.72 1, 24 | 0.0400 | 0.01 1, 24 | 0.9390 |
| Vv | 14.21 1, 23 | 0.0010 | 1.04 1, 23 | 0.3178 | 3.83 1, 23 | 0.0624 |
| Vs | 3.35 1, 25 | 0.0793 | 10.03 1, 25 | 0.0040 | 2.13 1, 25 | 0.1571 |
| PAG | 0.57 1, 22 | 0.4585 | 5.27 1, 22 | 0.0316 | 0.40 1, 22 | 0.5344 |
| CG | 1.41 1, 25 | 0.2459 | 14.06 1, 25 | 0.0009 | 0.64 1, 25 | 0.4308 |
| VL | 1.03 1, 23 | 0.3216 | 11.96 1, 23 | 0.0021 | 0.001 1, 23 | 0.9752 |
| ATn | 0.46 1, 24 | 0.5059 | 0.71 1, 24 | 0.4089 | 10.14 1, 24 | 0.0040 |
| 11-KT | 0.24 1, 26 | 0.6283 | 1.83 1, 26 | 0.1882 | 0.01 1, 26 | 0.9053 |
| | Male Mirror - NC | | Female Mirror - NC | | | |
| | P | | P | | | |
| (2) | | | | | | |
| nMMp | 0.2935 | | 0.2092 | | | |
| Vv | 0.7935 | | 0.0598 | | | |
| Vs | 0.4903 | | 0.0020 | | | |
| PAG | 0.5310 | | 0.0408 | | | |
| CG | 0.1378 | | 0.0024 | | | |
| VL | 0.0773 | | 0.0181 | | | |
| ATn | 0.0313 | | 0.1467 | | | |

Table 4. (1) effect of sex and condition on pS6 immunoreactivity and 11-KT. (2) post-hoc comparisons.

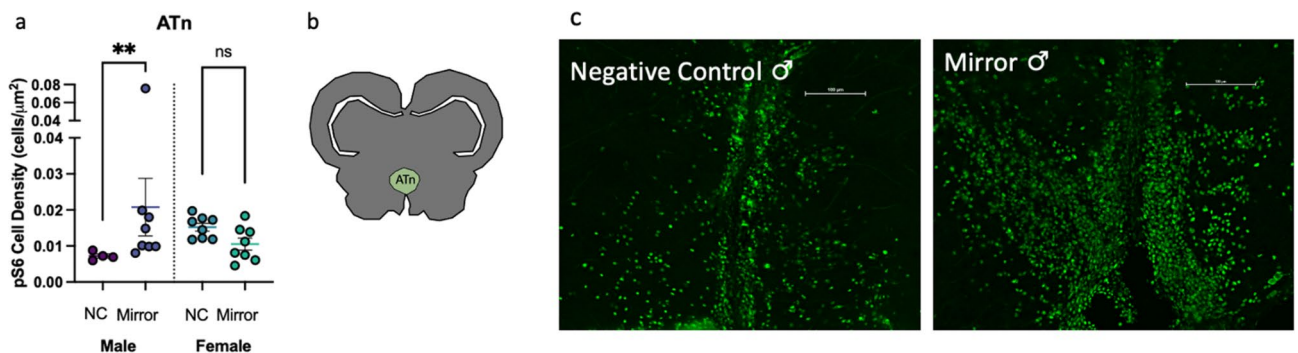


Fig. 6. Males and females have opposite neural activation patterns in the ATn. **(a)** pS6 cell density in the ATn has an interaction effect between sex and condition. **(b)** Representative section of the ATn. **(c)** Representative sections of the ATn stained with pS6 antibody to detect neural activation show lower signal in a male in the control condition compared to a male exposed to a mirror. NC, negative control. Scale bar = 100 μm. ** $p < 0.01$.

To further investigate the correlated pS6 immunoreactivity across the regions of interest, we used Pearson correlations and created heatmaps from the correlation coefficients for mirror and negative control fish with dendrograms to visualize the clustering of neural activity (Fig. 7). For the mirror fish, there is a clear cluster of the POA and subregions (nPPa, nMMp, and nPMp) with a negative correlation to the Dm-3. For the negative control fish, there is a cluster of correlated neural activity between the Vs, Dm-3, ATn, and the Vv, and an additional cluster between the PAG/CG, nMMp, VL, and Xm. We then wondered if these clusters were consistent between males and females within each condition (Fig. 7f). In the mirror males, subregions of the POA (nPPa and nMMp) clustered with the ATn. In the mirror females, there is still a cluster of the POA and subregions (nPPa, nMMp, and nPMp), but the ATn is negatively correlated and clusters with the Dm-3, VL, and Xm. The

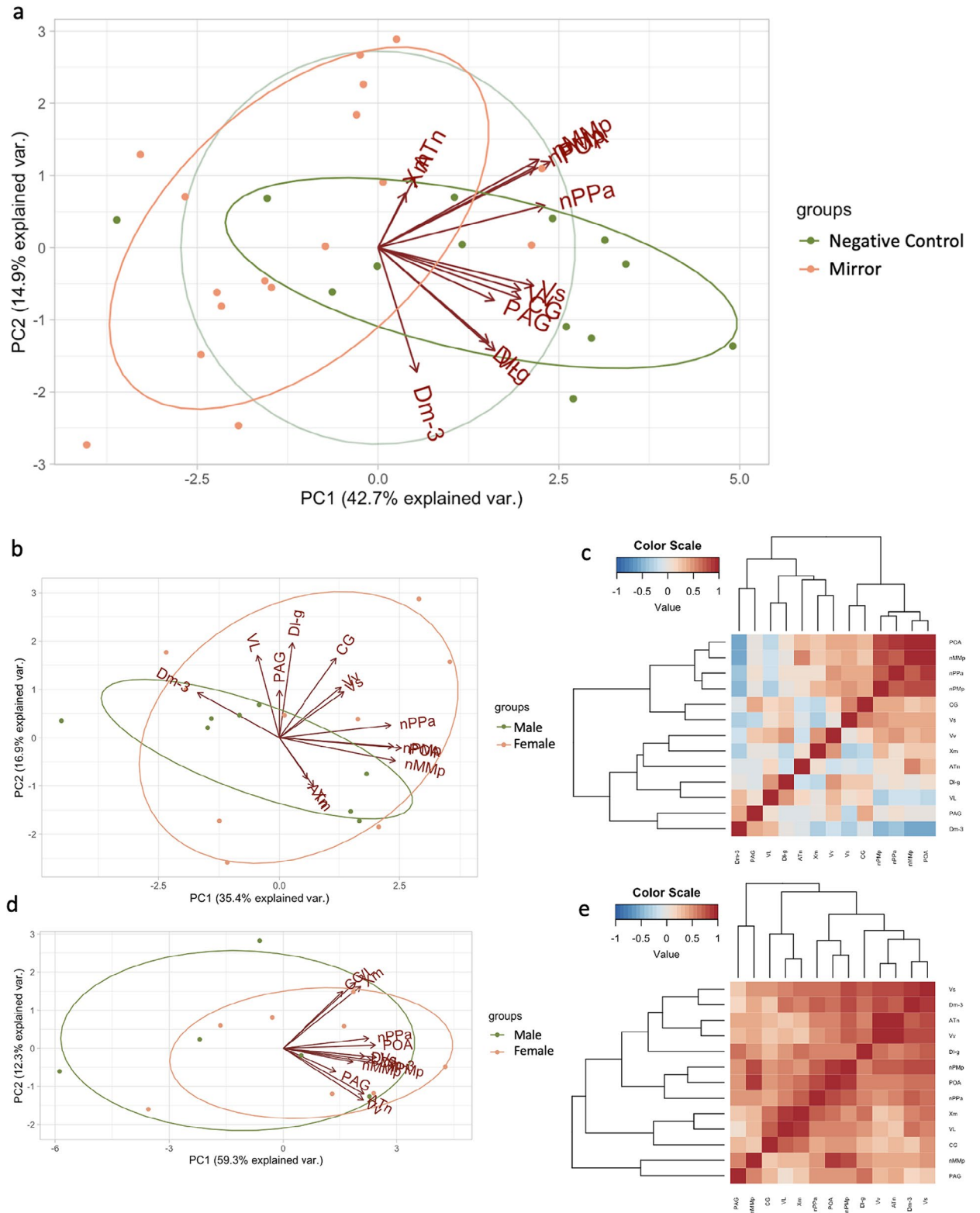


Fig. 7. Neural activation clusters by condition and sex. **(a)** Principal components analysis of pS6 immunoreactivity in mirror and negative control fish **(b)** Principal components analysis of pS6 immunoreactivity in mirror fish. **(c)** Heat map of Pearson correlation coefficients of pS6 immunoreactivity in the ROI of mirror fish. **(d)** Principal components analysis of pS6 immunoreactivity in negative control fish. **(e)** Heat map of Pearson correlation coefficients of pS6 immunoreactivity in the ROI of negative control fish. **(f)** Heat maps of Pearson correlation coefficients of pS6 immunoreactivity in the ROI for each sex and condition. Heat maps were generated in R (version 4.3.1, R Core Team 2023, <https://www.R-project.org/>)⁴³.

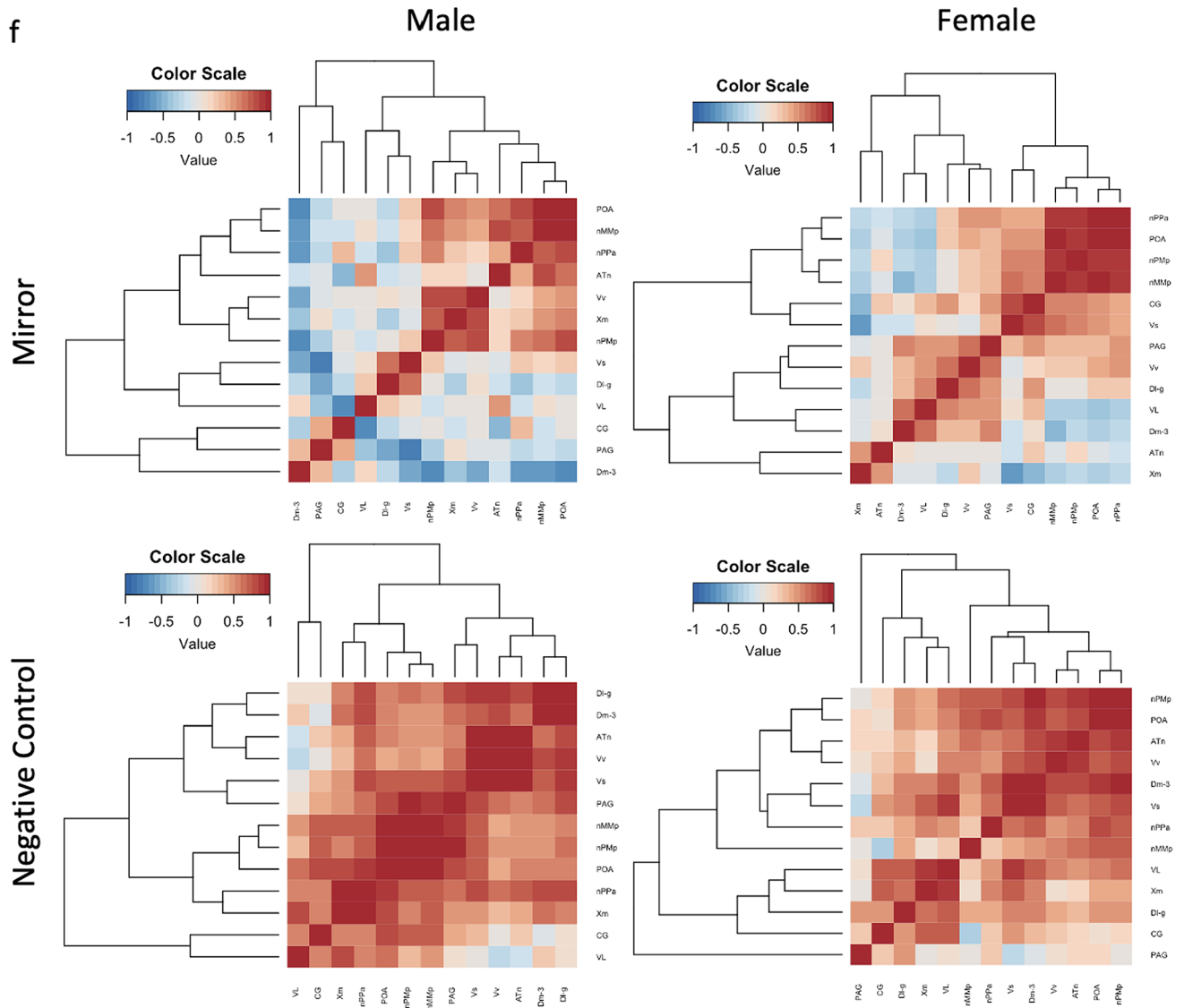


Figure 7. (continued)

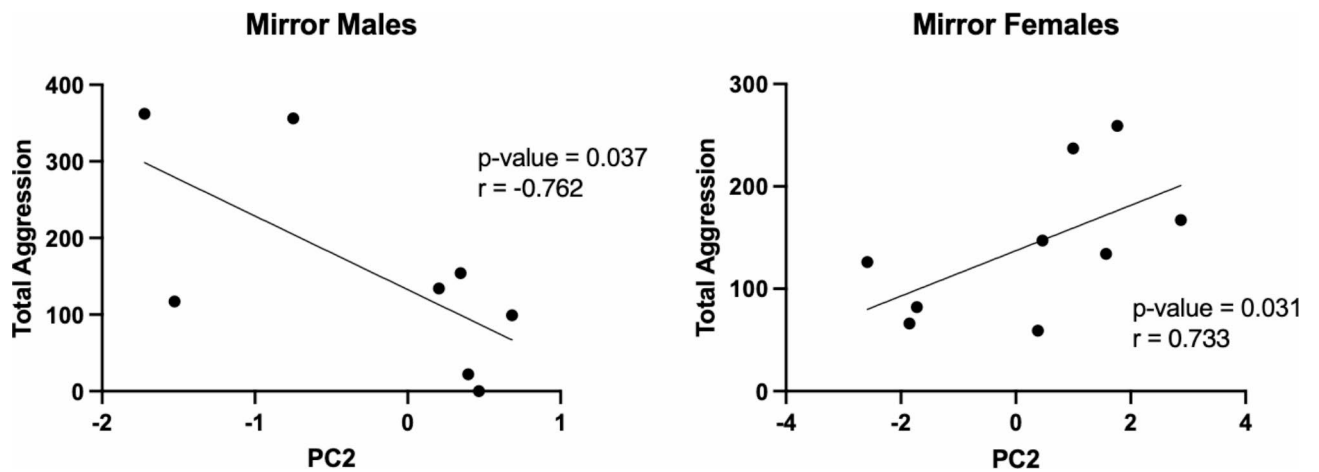


Fig. 8. Correlations of PC2 to total aggression in mirror males and mirror females.

negative control males have correlated activity in the Vs, Vv, ATn, Dl-g, and Dm-3, that is negative correlated to the CG and VL. The female negative controls have a cluster of the POA (nPMp), ATn, and Vv.

With the differences in correlated neural activity between mirror males and females specifically in their opposite relationship to the ATn, we wanted to further investigate if principal component scores of the mirror fish (Fig. 7b) correlated to aggressive behaviors (Fig. 8). We correlated the principal components scores of each mirror fish for PC1 and PC2 to aggressive behaviors and found significant correlations between PC2 with attacks and total aggression in mirror males and females. However, males and females had opposite relationships between these measures - males had a positive correlation between PC2 and attacks/total aggression while females had a negative correlation between PC2 and attacks/total aggression.

Discussion

We aimed to compare male and female intrasexual aggression in *A. burtoni*, a non-traditional model organism of social behavior that has the potential to yield novel insights into the sex differences in the control of social behavior. We determined that in an identical context that elicits aggression, male and female *A. burtoni* perform a mixture of identical behaviors and sex-specific behaviors. Interestingly, quivers, which are performed by males in reproductive contexts, were performed by females in the mirror assay, suggesting it is used as an aggressive signal by females. This has also been found in live interactions of females performing male-typical courtship displays in all-female communities²⁰. As in other species in which females adopt male-like mounting behaviors towards other females to assert dominance, perhaps in teleosts this behavior maintains female social status⁴⁴. Although quivers in males and females are stereotypically identical, quivers in male courtship are often accompanied by additional acoustic information⁴⁵. It is important to consider that there may be other nuanced differences in quivers performed by males and females that are not characterized in this study.

Intrasexual male aggression, and in theory intrasexual female aggression, is thought to follow specific “rules of engagement” between opponents, assessing the resource holding power of the other to minimize risk in conflict⁴⁶. Since *A. burtoni* males alter their aggression depending on the relative size of their opponent, we can safely assume that intrasexual aggression involves a level of opponent assessment. In our study, we find that males exposed to a mirror have a longer latency to behave compared to negative control males. Perhaps this longer duration of time to investigate the mirror serves as a lengthy assessment period since the male is faced with an identically sized opponent. However, we don't see this difference in female *A. burtoni* exposed to a mirror vs. negative controls. Perhaps intrasexual female aggression in *A. burtoni* does not follow the same opponent assessment as it does for *A. burtoni* males, supported by a study in Texas cichlids that suggests that males and females use a different set of rules to settle conflict⁴⁷. However, it is important to note that there are closely related cichlid species in which males and females use similar conflict rules⁴⁸.

One source of variability in our results could be due to the experimental pre-assay housing conditions. Since *A. burtoni* readily form social dominance hierarchies, it is likely that although the males added to the pre-assay tank were all dominant, they formed a new dominance hierarchy throughout the two-week housing period. Some of the changes associated with a dominance status take days to occur fully in *A. burtoni*, including neural and endocrine differences¹⁸. Thus, each fish in these pre-assay groups may be in different physiological states along a dominance continuum, contributing to a variability in the behaviors, hormones, and neural activity patterns measured.

We also revealed a significant difference in condition in activity and zone duration in which mirror females have higher activity and spend more time in the zone near the mirror compared to negative control females. Although not statistically significant, mirror males also have higher activity levels on average than negative controls. Since the change in the environment between the pre-mirror and mirror assay is more pronounced in trials of fish exposed to a mirror compared to the negative controls (opaque cover removed vs. remained on), we expect to see mirror subjects behaving more actively in their assay. Additionally, the mirror allows subjects to be more engaging with their reflection compared to an opaque cover which may be manifesting here in activity levels. Mirror females spending more time near the mirror may reflect an aspect of exploration or investigation that is not necessary for males in this context. In fact, female rats show more exploratory activity than males in a novel environment and have shorter approach latencies to novelty⁴⁹. Perhaps this aspect of elevated exploration in females is consistent with a longer zone duration in mirror females that we have shown here.

We found that 11-KT did not differ between mirror and negative control conditions, which is in line with findings in other teleost species^{50,51} but contrasts with previous findings in male *A. burtoni* where mirror assays were also used^{26,52}. However, as argued by Oliveira and Canario⁵³, the previous mirror assay study in male *A. burtoni*²⁶ has a few issues that could explain the higher 11-KT in mirror fish compared to negative controls, including the confound of exposure to a competing male's chemical cues in the same territory directly before exposure to a mirror. Indeed, the lack of a rise in androgens in a mirror assay that we observed in both males and females has been explained by the fact that there is no clear winner of the fight as is seen against a live opponent because all aggressive acts are perceived to elicit an equal counter response^{15,50,51,53,54}. Despite the absence of an observed rise in androgens, recent work demonstrated the necessity of androgen signaling in male typical but not female typical aggression⁵⁵.

In multiple brain regions we observed impacts of sex and exposure to the mirror on neural activation as measured by pS6 immunoreactivity. For several brain regions no differences in pS6 activation between negative control and mirror fish, including the POA (nPPa and nPMp), Dl-g, Dm-3, and Xm, were observed. The POA plays a role in aggressive and reproductive behaviors across vertebrates³⁰. In *A. burtoni*, there is an increase in *egr1* expression in the nMmp in males after fighting but not after courting, emphasizing the role of this POA subregion in modulating aggression⁵⁶. When we investigated the pS6 immunoreactivity in the POA we expected to see higher pS6 immunoreactivity in the POA in fish exposed to the mirror compared to the negative control condition. Surprisingly, we see no differences in pS6 immunoreactivity as a function of sex or condition for

two of the subregions of the POA (nPPa and nMPp), and although we find a significant effect of condition on pS6 immunoreactivity in the nMPp, it was higher in negative controls. Another study in *A. burtoni* found no effect of social stimulus on *c-Fos* expression in the POA when males were exposed to reproductive or aggressive contexts⁵⁷. Interestingly, the males in the aforementioned study were not engaged in full contact with stimulus fish and therefore not receiving all sensory signals. Hence, it may be that the POA neurons activate in response to integrating several multisensory components of social information, but solely the visual input and behavioral output we have observed in this study is not sufficient for the activation of these neurons. Previous work has emphasized the importance of the integration of chemosensory and visual signals to elicit distinct neural activation patterns compared to the exposure of a unimodal signal^{57,58}. Since our design utilizes a unimodal visual signal, we would hypothesize that neural activation of the POA would differ by condition if fish were exposed to multisensory social cues. Additionally, while the findings indicate involvement of these regions, the specific topographical relevance within these regions is not known. Future studies should attempt to explore whether the involvement of these ROIs in aggression is specific to a subregion or is consistent across the entire region.

The Dl-g (homolog of the medial pallium; hippocampus) is involved in processing sensory input and receiving and processing socially relevant hydrodynamic cues in *A. burtoni*⁵⁹. The Dm (homologous to the basolateral amygdala, blAMY) also receives sensory input including lateral line information^{60–62}. pS6 immunoreactivity in the Dl-g, Dm-3, and Xm did not differ by sex or condition and may indicate that mechanosensory input is similar in the negative control condition and mirror conditions despite distinct differences in behavioral output. Some behavioral measures (distance moved (cm) and velocity (cm/s)) do not differ by sex or condition and may provide evidence of similar mechanosensory input between these groups. It is important to note that the lack of socially relevant hydrodynamic cues that accompany a fight with a real opponent are not present in the mirror assay, and thus regions like the Dl-g, Dm-3, and Xm that process sensory information may not be relevant in an aggressive assay lacking this dynamic sensory input.

One region that did display a sex difference in pS6 immunoreactivity is the Vv. The Vv, homologous to the LS, is thought to integrate internal state with the social environment since this region is shared between the social behavior network and the mesolimbic reward system³⁰. The Vv has also been implicated in processing social information in anxiety-like contexts. In previous work, female *A. burtoni* had higher *cfos* expression in the Vv after watching a preferred male lose a fight⁶³. In this study, we found higher pS6 immunoreactivity in the Vv of females compared to males regardless of condition, which may indicate that the mirror assay is more anxiogenic to females, or that the Vv is more active overall in females than it is in males. Future experiments aimed at testing the role of the Vv in explicitly anxiogenic behavior paradigms in both males and females are thus warranted.

The ventromedial hypothalamus (VMH) has been implicated in mediating aggressive behaviors across several species⁷. In mammals, *cfos* expression in the VMH increases after aggressive interactions in males, and specifically the causal role of the ventrolateral portion of the VMH (VMHvl) in aggression in male mice is well established^{4,64,65}. In *A. burtoni*, the ATn (homologous to the VMH) is likely involved in processing mechanosensory information in male-male aggressive contexts⁵⁹. In our study, pS6 immunoreactivity in the ATn is higher in males exposed to a mirror than in negative controls, supporting the importance of activity in the ATn in mediating male-male aggression in the mirror assay. While pS6 immunoreactivity was not higher in mirror versus control females, this does not preclude the ATn potentially being involved in female aggression in an environment-specific manner. For example, ATn neurons may become activated during female aggression in particular contexts and/or a specific sub-population of neurons in the ATn that remains to be identified regulates female aggression as the ventrolateral portion of the VMH does in male mice. In a previous study in *A. burtoni*, females show context-dependent aggression and neural activation patterns in which *c-fos* expression in the ATn is greater when exposed to brooding females⁶⁶. Perhaps neural activation in the ATn in females is triggered by the visual stimulus of a brooding female, which our study did not capture. Investigations into whether a specific locus within the fish VMH regulates aggression in both male and female *A. burtoni* will be key in disentangling the neural basis of aggression in this species.

Some regions of interest had higher neural activation in negative controls than in mirror fish. These results suggest the potential involvement of inhibitory neurons in suppressing activity in specific aggression circuits. Glutamic acid decarboxylase (*Gad*) is the rate-limiting enzyme that converts glutamate (an excitatory neurotransmitter) into GABA (an inhibitory neurotransmitter). *Gad* exists in two highly conserved isoforms, *gad1* and *gad2*. In *A. burtoni*, the Vs, CG, and VL express both *gad1* and *gad2*⁶⁸. The Vs, PAG/CG and VL show higher pS6 immunoreactivity in negative controls and since *gad1* and *gad2* are both expressed in these regions, we postulate that the inhibitory neurons in these regions are important for suppressing aggressive behaviors. We hypothesize that inhibitory neurons containing GABA are activated in negative control fish whereas lower activation of these neurons permits aggressive behaviors in mirror fish. However, another reasonable explanation for higher neural activation in negative control females is a higher GSI than mirror females (Figure S6). Previous literature shows estrus cycle-related changes in *c-fos* mRNA expression in the female rat brain in response to acute stress⁶⁸, but these estrus cycle-related changes were region specific and *c-fos* expression was not correlated to level of estrogen in the cortex. It is also important to note that the PAG location in this study was only an approximation and may not be completely representative of the PAG given the lack of reliable neuroanatomical markers in this region.

The clusters of correlated neural activity in the mirror fish replicated the loadings in the PCA – in which the POA and subregions cluster together and are negatively correlated to the Dm-3. When we visualized the clusters of correlated neural activity separately in mirror males and females, we found a distinct difference in the relationship of the ATn to the POA present in males and not females (Fig. 7f). This strong correlation of the POA and the ATn present in males and not females reflects a male-specific aggressive state. The core aggression circuit (CAC) is a subnetwork of the SBN that is specialized for aggression and is comprised of the medial amygdala, the

ventrolateral portion of the ventromedial hypothalamus (VMHvl), the ventral premammillary nucleus, and the bed nucleus of the stria terminalis⁴. The POA acts as a link between the CAC and the ventral tegmental area to trigger learned aggressive actions indirectly. Since the ATn and the POA are interconnected, perhaps in male *A. burtoni* aggressive behaviors are initiated by the ATn (VMH homolog) and the POA in a mirror assay, and this is not the case in females. The functional connectivity between the POA and ATn in aggressive males may suggest the presence of a permissive gate that is triggered by the visual stimulation of an aggressive encounter that opens this connection between the ATn and POA. Since this functional connectivity is exclusive to aggressive males, a different mechanism may be triggering aggression in females in response to a mirror.

We also found that PC2 for mirror fish correlated positively to attacks and total aggression in mirror males and correlated negatively to attacks and total aggression in mirror females (Fig. 8). This leads us to believe that the loading variables of PC2 (CG, VL, Xm, Dl-g, and ATn) mediating aggressive responses in males and females in opposite ways, perhaps through neural inhibition in females. These brain regions play a large role in the processing of sensorimotor information and could reflect sex differences in the circuits that function together to perform attacks in *A. burtoni*.

Conclusions

Our results revealed both similar and different behavioral patterns underlying aggressive responses to a mirror in male and female *A. burtoni*. We also observed extensive sex differences in neural activation throughout regions of the SBN in response to our aggression assay, suggesting distinct neurons and neural circuitry underlie aggressive behavior in male and female *A. burtoni*. Interestingly, neural activation in the ATn reflected aggressive state in males but not females, suggesting this region controls aggression in male *A. burtoni* as it does in mice^{4,8,65,69}. Activity in the ATn and the POA appears to be functionally connected in aggressive males, suggesting that the visual input received about an aggressive context in males triggers activity in the ATn and POA to produce a behavioral response to a mirror, but this circuit is absent in female aggression. We also provide evidence that the inhibition of neural activity in a variety of brain regions in the SBN is associated with aggression in *A. burtoni* as well. These findings overall lay a foundation of testable hypotheses for future studies to understand the molecular and neural regulation of sex differences in behavior in *A. burtoni*.

Gaining a fundamental understanding of the molecular and neural basis of aggression requires studies in both sexes in which the conditions and behaviors performed by either sex are similar. In our mirror assay we were able to stimulate aggressive responses in both males and females. The behaviors performed by males were the same as the suite of behaviors they perform in dyadic social interactions, and we confirmed that this is the case for females as well. A significant benefit of assessing aggression in a setup that controls for the environmental and social conditions in males and females is it is presumed that while (1) both sexes are experiencing an elevated motivation to behave aggressively, they are (2) also performing distinct and similar aggressive behavioral acts. Thus, a benefit for the use of *A. burtoni* as a model organism for elucidating the molecular and neural mechanisms governing sex differences in aggression is that the molecular and neural mechanisms of the motivation versus sensorimotor aspects of aggression in both sexes can be studied in the same environment. Therefore, studying sex differences in aggression in *A. burtoni* may lead to new ideas on the fundamental mechanisms controlling social behavior generally, yielding novel hypotheses that can be tested across species.

Data availability

The datasets generated and/or analyzed during the current study are available in the manuscript itself, the Supplementary Information file, and online at <https://github.com/AlwardLab/sexdiffs2024>.

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Author contributions

B.A.A. and L.R.J. designed the study; L.R.J. performed the experiments; L.R.J. and M.D. collected the data; B.A.A. and L.R.J. analyzed the data; L.R.J. made the figures. B.A.A. and L.R.J. wrote and edited the manuscript.

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Declarations

Ethics approval and consent to participate

Experimental procedures were conducted according to the ethical guidelines for the care and use of laboratory animals. Experiments were approved by the University of Houston Institutional Animal Care and Use Committee (Protocol #202000001).

Competing interests

The authors declare no competing interests.

Additional information

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