UC Irvine UC Irvine Previously Published Works

Title

The invasive mosquito species Aedes albopictus: current knowledge and future perspectives

Permalink <https://escholarship.org/uc/item/2xr2j5f6>

Journal Trends in Parasitology, 29(9)

ISSN 1471-4922

Authors

Bonizzoni, Mariangela Gasperi, Giuliano Chen, Xioaguang [et al.](https://escholarship.org/uc/item/2xr2j5f6#author)

Publication Date 2013-09-01

DOI

10.1016/j.pt.2013.07.003

Peer reviewed

The invasive mosquito species Aedes albopictus: current knowledge and future perspectives

Mariangela Bonizzoni¹, Giuliano Gasperi², Xioaguang Chen³, and Anthony A. James⁴

¹ Program in Public Health, Hewitt Hall, Room 3501, University of California, Irvine, CA 92697-3900, USA
² Department of Biology and Biotechnology, University of Pavia, via Ferrata 9, 27100 Pavia, Italy
³ School of California, Irvine CA 92697-3900, USA

One of the most dynamic events in public health is being mediated by the global spread of the invasive mosquito Aedes albopictus. Its rapid expansion and vectorial capacity for various arboviruses affect an increasingly larger proportion of the world population. Responses to the challenges of controlling this vector are expected to be enhanced by an increased knowledge of its biology, ecology, and vector competence. Details of population genetics and structure will allow following, and possibly predicting, the geographical and temporal dynamics of its expansion, and will inform the practical operations of control programs. Experts are now coming together to describe the history, characterize the present circumstances, and collaborate on future efforts to understand and mitigate this emerging public health threat.

The Asian tiger mosquito: origin and spread

The Asian tiger mosquito, Aedes albopictus (Figure 1), is an aggressive, daytime biting insect that is emerging throughout the world as a public health threat following its primary role in recent dengue virus (DENV) and Chikungunya virus (CHIKV) outbreaks [1–3]. Part of its impact on human health is due to its quick and aggressive spread from its native home-range in East Asia and islands of the western Pacific and Indian Ocean [4]. Ae. albopictus has colonized every continent except Antarctica in the past 30–40 years (Figure 2) [4,5]. The first record of this species in Europe (Albania) was in 1979 [5]. Today, Ae. albopictus is present in all countries on the Mediterranean sea, including parts of Turkey and the Middle Eastern states of Lebanon, Israel, and Syria, and is gradually moving north [4,6]. Italy and southern France are the most infested regions [6], but Ae. albopictus also have limited local distribution in southern Switzerland, The Netherlands, Bulgaria, Russia, Belgium, and Germany, confirming predictions of expansions based on climate changes [4].

Ae. albopictus was introduced into the USA at the end of the eighteenth century in Hawaii [7]. The first detection in

460 Trends in Parasitology September 2013, Vol. 29, No. 9

© 2013 Elsevier Ltd. All rights reserved. http://dx.doi.org/10.1016/j.pt.2013.07.003

the continental USA was in 1985 in Texas, and Ae. albopictus is established currently in 866 counties of 26 states. A breeding population of Ae. albopictus was discovered recently in southern California [8,9], and this species also is detected sporadically in Washington and New Mexico (www.cdc.gov/ncidod/dvbid/arbor/albopic_new.htm).

The presence of Ae. albopictus in Central and South America has been documented since the 1980–1990s, primarily in Mexico and Brazil (www.cdc.gov/ncidod/dvbid/ arbor/albopic_new.htm). Records of Ae. albopictus in

Glossary

Abiotic factor: a non-living or physical component of an ecosystem, such as climate or habitat.

Anthropophilic: preferring humans as a blood source.

Arbovirus: animal virus dependent on arthropod vectors for transmission.

Biotic factor: any living component of an ecosystem.

Critical photoperiod: the number of hours of daylight that results in diapause entry for 50% of the insect population, excluding those that do not enter diapause under unambiguous short day-lengths.

Cytoplasmic incompatibility (CI): inability to produce viable offspring as a result of gamete infection by intracellular parasites. In the case of Wolbachia, CI occurs when an infected male mates with an uninfected female (unidirectional CI) or a female infected with a different Wolbachia type (bidirectional CI). When an infected female mates with an infected male, no CI is observed, but the progeny are infected with Wolbachia.

Detritus: particulate matter originating from the disintegration of biological materials such as microbiota and tissues.

Diapause incidence: proportion of the insect population entering diapause under unambiguous short-day lengths.

Exophagous: preferring to feed outdoors; contrast with endophagous (preferring to feed indoors).

Exophilic: preferring to rest outdoors; contrast with endophilic (preferring to rest indoors).

Isofemale selection: selection strategy whereby progeny are derived by mating single females.

Paratransgenesis: genetic modification of a symbiont to affect the vector competence of its host. Symbionts need to be transformed easily without affecting their fitness, they need to spread across host populations, and they need to reside in host organs affected by pathogens.

Polyprotein: a single polypeptide chain that is the primary product of polypeptide synthesis and that is then cleaved into different functional proteins. The genomes of the Flaviviruses, including dengue virus (DENV) and Chikungunya virus (CHIKV), encode a polyprotein that contains three structural and eight non-structural proteins.

Sterile insect technique (SIT): an insect control technique based on the generation of random mutations through chemicals or γ -irradiation that lead to sterility, and release of these insects in the wild to suppress local populations. Vector competence: the ability of an arthropod to acquire, maintain, and transmit pathogens.

Zoophilic: preferring animals to humans as ^a blood source. 1471-4922/\$ – see front matter

Corresponding author: James, A.A. (aajames@uci.edu).

Keywords: Aedes albopictus; public health; arboviruses; expansion; vector control.

Figure 1. Aedes albopictus adult male (A) and female (B). Courtesy of Centro Agricoltura Ambiente, Crevalcore, Italy.

Mexico reflect a widespread distribution starting from the north at the border with Texas in the 1990s, and moving south and east into neighboring countries of Guatemala and Belize [4,6]. The Caribbean islands of Barbados, the Dominican Republic, Trinidad, the Caymans, and Cuba, together with the Central and South American States of El Salvador, Honduras, Panama, Nicaragua, Venezuela, Colombia, and Bolivia, also have documented detections of Ae. albopictus since the late $1990s$ [4,6]. Invasion of Brazil started with focal detection in Rio de Janeiro in 1986, and then in two neighboring states of Espirito Santo and Minas Gerais, and in San Paulo the following year [7,10]. A rapid expansion followed with mosquitoes moving to northern, northeastern, and central regions, reaching the Amazon basin by the early 2000s [11]. Breeding populations of Ae. albopictus have been identified since the late 1990s in the Misiones province of northern Argentina, at the border with Brazil [12]. The current distribution of this

species in Argentina is limited to northeast provinces and includes the neighboring countries of Uruguay and Paraguay [4].

The first detection of Ae. albopictus in Africa was in 1989 when live larvae were detected in the port city of Cape Town, South Africa, in used tires imported from Japan; the infestation was immediately controlled [13,14]. Two years later, it was recorded in Nigeria and dispersed to Cameroon, Equatorial Guinea, and Gabon [1,14].

Once Ae. albopictus is established in an area it is difficult to eradicate, and constant surveillance and appropriate control strategies are required [15]. The widespread distribution of Ae. albopictus outside its native home-range is presumed to have been primarily human-mediated and accidental [16]. Low levels of mitochondrial DNA (mtDNA) diversity and limited phylogeographic genetic differentiation among populations are consistent with dispersal and invasion mediated by human activities, such as continued

Figure 2. Aedes albopictus distribution range. Map indicating the first reports (interception on imports, local captures, and documented endemic populations) of Ae. albopictus by country (political boundaries). Record data are based on published literature [4,6]. In Madagascar, La Réunion, and Hawaii, Ae. albopictus was introduced in the 18th–19th or early 20th centuries.

migration events, commerce in used tires and live plants $('lucky'$ bamboo), and continued propagule pressure $[9,17-]$ 19].

Public health impact of the Asian tiger mosquito

Retrospective studies of the incidence of DENV infections in regions with exclusive or prevalent presence of Ae. albopictus versus the main vector, Aedes aegypti, show the absence of explosive dengue outbreaks [20]. Meta-analysis of published laboratory-based experiments on the relative vector competence (see Glossary) of Ae. albopictus versus Ae. aegypti for DENVs reveal that the former are more susceptible to midgut infection, but more resistant to virus dissemination [20]. These observations led to the conclusion that Ae. albopictus is currently a less efficient vector of DENVs than Ae. aegypti [1]. However, Ae. albopictus was the sole or primary vector of recent (2001–2010) DENV outbreaks in Hawaii, Indian Ocean islands, Central Africa, and southern China [1,2,21,22] and the first DENV (2010) autochthonous transmissions in Europe [3].

Ae. albopictus can transmit several other arboviruses of public health significance, including CHIKV and possibly West Nile virus (WNV) [1,23]. Ae. albopictus was responsible for recent CHIKV epidemics in several islands of the Indian Ocean, Central Africa, and Europe [3,23]. A total of 26 viruses from five different families, including Eastern equine encephalitis, La Crosse, Japanese encephalitis, and Venezuelan equine encephalitis, have been isolated from wild-caught Ae. albopictus, although their role in transmission is uncertain [1]. This species also vectors the filarial nematode Dirofilaria, with transmission occurring primarily between dogs and mosquitoes, but occasionally involving humans [6].

What makes Ae. albopictus a threat? The density of mosquito populations is an important element favoring epidemics in the presence of limited vector competence. This was shown during the yellow fever epidemic in Nigeria in 1987, which was vectored by a high-density, sylvatic Ae. aegypti that had low competence for the virus [24]. Moreover, vector competence is a dynamic status as evidenced by recent CHIKV outbreaks in the Indian Ocean islands of La Réunion, Mauritius, Madagascar, and Mayotte (2005–2007), as well as in Central Africa (2006–2007) and Italy (2007) [23]. Phylogenetic analysis of full-length viral sequences of isolates from these outbreaks revealed an example of rapid (1–2 years) convergent evolution through the independent acquisition of a mutation from Ala to Val at position 226 (A226V) in the E1 gene of the viral polyprotein [23]. This convergent evolution results from a strong selective advantage of the A226V mutation [23], which is associated with improved CHIKV replication and transmission efficiency in Ae. albopictus [25,26]. Ae. albopictus is now the predominant or only CHIKV vector in La Réunion, Mauritius, Italy, Cameroon, and Gabon, supporting the conclusion that the A226V mutation is a viral adaptation to a previously atypical vector with aggressive and rapid dispersal efficiency [23].

The emergence of arboviruses is linked frequently to changes in vector or vertebrate hosts, or both [27]. This continuous evolutionary adaptation is particularly This review summarizes current knowledge of the Asian tiger mosquito Aedes albopictus. Research is necessary to enhance our understanding of Ae. albopictus biology and contribute to the development of efficient and sustainable strategies for population control. An important aspect of future research activities will be the definition and adoption of standardized procedures that allow comparisons of results from different studies and the availability and sharing of wild-caught mosquitoes among the research community.

Phylogeny/taxonomy/genomics and population genetics

- Identification of molecular markers.
- Characterize the genetic diversity of the species, including geographical and temporal aspects, and variability in biological important traits (i.e., vector competence, insecticide resistance, breeding preferences, climatic adaptation and competitive ability).
- Taxonomic status of Ae. albopictus.
- Comparative genomics.

Biology and ecology of Aedes albopictus

- Projected species range.
- Microbiota.
- Ecological influences on competition impact/effects on vector control strategies.
- Diapause, cold- and drought-tolerance.
- Species range expansion.

Vector competence/vectorial capacity

- Collection of epidemiological data.
- Pathogen-resistance phenotypes; identification of markers for vector competence.
- Characterization of arboviruses infecting geographically-different populations and their interaction with arboviruses of public health importance.

Vector control strategies

- Insecticide resistance in Ae. albopictus worldwide populations.
- Biology of Wolbachia in Ae. albopictus: distribution/physiology of endogenous strains and their interaction with strains transferred into Ae. albopictus from other insect species.
- **Genetic engineering of Ae. albopictus: population suppression** and population replacement strategies.
- Mass breeding and quality control.

troubling considering Ae. albopictus competency for multiple arboviruses [1], and emphasizes the importance of understanding and monitoring the competence of geographic populations for different arboviruses (Box 1). In addition, Ae. albopictus shows ecological plasticity in different traits such as larval breeding sites, feeding behavior, and climatic adaptation that increase their potential for spread and adaptation to new environments, and influence their coexistence with other vector species. Increased global movements of goods and humans together with climate change are associated already with the expansion of its range and impact on human health [1,6,16].

Ecological variation

Breeding habitat choice and feeding behavior

Ae. albopictus originated at the edges of forests and bred in natural habitats (tree holes, bamboo stumps, and bromeliads) and consequently was previously considered to be a rural vector [28]. However, this species has adapted well to suburban and urban environments with larvae now breeding in artificial containers (tires, cemetery urns, and water storage containers) (Figure 3) and has become the most

Figure 3. Aedes albopictus breeding sites. Examples include (A) metal containers, (B) terrarium, (C) stone holes, (D) ceramic vessels, (E) plastic containers, (F) gutters, (G) used tire dumps, (H) surface accumulated water, (I) disposable containers, (J) parking poundings, (K) flowerpot trays, and (L) metal containers. Images courtesy of Romeo Bellini (Centro Agricoltura Ambiente, Crevalcore, Italy), Marco E. Metzger (California Department of Public Health, USA), and Xiaoguang Chen (Southern Medical University, Guangzhou, China).

important and sometimes sole vector in urban areas (southern China and Italy) [21,29]. Ae. albopictus feeds aggressively and opportunistically during the day on a wide range of hosts depending on their availability and the environment [30–32]. When offered a choice, this species prefers humans (anthropophilic behavior) [1,30,31], but it also can feed on a large variety of animals (cows, goats, dogs, birds, reptiles, and amphibians) (zoophilic behavior) [30,31]. The opportunistic zoophily enhances the spectrum of pathogens it can vector and the ecological niches it can occupy [1]. Most literature refers to this species as resting and feeding outdoors (exophilic and exophagous), but recent reports provide evidence of geographical variation in this behavior with gravid females being captured indoors in Rome, Italy [32]. Whether this variation is dependent on different capture methodologies needs further investigation because it is difficult to find Ae. albopictus resting in the wild [30,32].

Climatic adaptation

The ability of Ae. albopictus to colonize different ecological niches and hence expand its species range is related partly to its capacity to adapt to seasonal variations through photoperiodic diapause [33,34]. Diapause is an environmentdependent period of dormancy characterized by changes in metabolic pathways, insulin signaling, cell-cycle arrest, and upregulation of stress-response genes [35–37]. When exposed to seasonal short day-length (as in winter time), Ae. albopictus adult females of temperate populations oviposit eggs in which pharate larvae enter into diapause inside the

chorion of the egg until permissive conditions favor the resumption of development [33,34]. Rapid evolution in traits such as the critical photoperiod (seasonal timing of diapause) and diapause incidence has been shown in Ae. albopictus populations invading the USA, and these data support the conclusion that these adaptive phenotypes are a crucial element of the invasion potential [34,37].

Diapausing eggs are larger, contain more total lipid, and are more desiccation-resistant than non-diapause eggs [38,39]. In addition to enhancing overwinter survival, these traits are likely to favor passive dispersal and thereby contribute to invasion success [40]. Analyses of gene expression profiles during diapause identified two genes (phosphoenolpyruvate carboxykinase, Pepck, and proliferating cell nuclear antigen, PCNA) in Ae. albopictus with diapause-related transcriptional profiles conserved across different insect taxa and support the conclusion that diapause and developmental programs are integrated tightly [36]. These results support the existence of a universal diapause 'gene set' that could be disrupted, providing a basis for novel control strategies [36].

The observed ecological variation in the breeding habitat choices, the opportunistic feeding behavior, and the adaptation to different climates may be either the result of plasticity (i.e., one genotype is flexible in its phenotypes) or of local adaptation of different genotypes.

Competitive interactions among species

Competitive interactions of invasive Ae. albopictus depend on the environment, and include both endemic species

(primarily Aedes triseriatus, Aedes atropalpus, and Aedes barberi in the USA and Culex pipiens in Italy) and other invasive species (Ae. japonicus in the USA; Ae. aegypti throughout the American continents, Indian Ocean islands, Bermuda, and tropical Asian countries, and Culex species in Europe and the USA) [40,41]. Most studies focus on competition and coexistence between the two dengue vectors, Ae. albopictus and Ae. aegypti, because of their public health relevance [41]. Ae. aegypti is native to sub-Saharan Africa but has expanded its species range to most tropical and subtropical regions of the world where Ae. albopictus populations also breed [42]. Competition between the two species occurs primarily at the larval stage, although there is new evidence of adult mating interactions influencing the impact of Ae. albopictus on Ae. aegypti [43]. Ae. albopictus tends to be a superior larval competitor, displacing Ae. *aegypti* in suburban and rural sites [1,41]. Furthermore, competition among larvae appears to have asymmetric effects on adult longevity [44]. Abiotic factors such as temperature, rainfall regime, spatial aggregation, and salinity, and biotic factors such as presence of predators, some aspects of detritus input, bacterial communities inhabiting the breeding site, and variation among geographically-distinct mosquito populations, can influence both breeding-site choices and the outcome of competition [45–52]. Seasonality may contribute to coexistence by causing fluctuations in both climate and detritus inputs [50]. Patterns of exclusion and coexistence in Florida are observed in different areas and are correlated with climate and land use, with wetter and cooler conditions favoring Ae. albopictus, and Ae. aegypti remaining prevalent in urban and southern areas of the state [40,41,45]. When Ae. albopictus was introduced to Réunion Island at the beginning of the 20th century it became ubiquitous and displaced Ae. *aegypti*, which is now restricted to the drier western coast of the island [48]. Ae. albopictus populations in this island were shown to be more temperature-tolerant and superior larval competitors than Ae. aegypti [48].

Aedes albopictus microbiota

Facultative and obligate symbiotic relationships between insects and microorganisms are widespread in nature, affect different insect phenotypes including their interaction with pathogens, digestion, protection against natural enemies, and also are exploited for control purposes [53–55]. Ae. albopictus natural populations are infected with Wolbachia pipientis (hereafter referred to as Wolba $chio(56)$. Two Wolbachia strains (wAlbA and wAlbB) were identified and shown to differ in tissue-distribution and density [57]. Wolbachia is the causative agent of cytoplasmic incompatibility (CI) and affects other host traits, including lifespan and vector competence [58,59]. These characteristics fostered the exploration of the use of Wolbachia to aid control of DENV transmission by Ae. aegypti and Ae. albopictus [55,60]. Naturally occurring Wolbachia strains do not affect either CHIKV or DENV replication in Ae. albopictus, but reduce DENV infection of the salivary glands [58]. In addition, recent studies of wildcaught mosquitoes from Madagascar and North America showed the presence of members of the Asaia, Acinetobacter, and Pantoea genera. Members of these Gram-negative

bacteria genera are found in several mosquito species such as Ae. aegypti, Ae. albopictus, and Anopheles stephensi, where they reside primarily in the gut, can be cultured and genetically transformed, and infection persists through generations; these characteristics suggest they are amenable candidates for paratransgenesis [57,61,62].

Aedes albopictus genome and phylogeny

The sequencing and annotation of an arthropod genome generates an important tool for understanding its biology and improving its management. INFRAVEC in Europe (http://www.infravec.eu/index.pl) and the Southern Medical University of Guangzhou (China) have undertaken independently the task of sequencing Ae. albopictus genomes. The genome is organized into three linkage groups (chromosomes) and has a variable estimated haploid DNA content of 0.62–1.66 pg [7]. INFRAVEC is sequencing a strain isolated from an urban environment in the northern Italian city of Rimini. This strain was maintained in a laboratory for 40 generations before multiple rounds of isofemale selections were carried out to reduce heterozygosity before genome sequencing and assembly. The genome effort in China is concentrated on the Foshan strain, which was isolated in Guangdong and has been maintained in the laboratory since 1981. Genome annotations are expected by the end of 2013 (D. Lawson and X. Chen, unpublished data). The availability of genome sequences and annotations of two Ae. albopictus strains of different geographic origins and histories (the Foshan strain is from the native home-range and the Rimini strain is from the 1990 invasion wave into Italy) is important with respect to reports of fluidity in the genome size of individual mosquitoes, most likely resulting from variation of the amount of highly repetitive DNA [7,63]. The genome sequence also will facilitate phylogenetic assessment and allow direct comparisons with other sequenced mosquito vectors, including Anopheles gambiae, Ae. aegypti, and Culex quinquefasciatus, and nine additional anophelinae genomes (vectorbase.org). Additionally, the sequence will contribute to transcriptome analyses which currently are limited to salivary glands, eggs, and embryos prepared for studying diapause [35,36,64].

Vector control strategies

Control of vector populations is the only current strategy for preventing many arbovirus infections, including DENV and CHIKV, because there are no commercially available vaccines nor treatments for these pathogens and their diseases. Vector control strategies can be classified broadly into four categories: (i) environmental modification; (ii) use of chemical compounds, (iii) biological control, and (iv) genetic-based strategies [60]. An integrated approach tailored to local environments and the socio-economic status of the targeted community is expected to result in the most success [65].

Environmental modification

The adoption of screens for windows and the systematic cleanup of water containers that could be exploited as breeding sites are measures proven effective to reduce Ae. albopictus larval populations and impact DENV

transmission [60,66]. Community participation is crucial to the success of these efforts.

Chemical compounds

Various chemical compounds are being used to control larval and adult Ae. albopictus. Insect growth regulators (methoprene, novaluron, and pyriproxifen) and the organophosphate, temephos, which has low mammalian toxicity, low odor, and is available in long-lasting formulations, are applied in different strategies to mosquito breeding sites to reduce larval populations [67–69]. One recent approach proposes the 'auto-dissemination' by adult females attracted to resting spots containing the insecticides and subsequent spread to new breeding sites [29].

Treatment of public areas with high mosquito density with adulticides is a recommended protective measure [70]. Pyrethroids are the most commonly used adulticides because of their low mammalian toxicity and rapid mosquito knockdown [68,69]. Increased use of insecticides for agricultural pest control, for direct control of Ae. albopictus or for control of sympatric vectors (such as other Anophelinae and Culicinae species), has imposed selection pressures on Ae. albopictus populations for increased resistance. Resistance to larvacides, primarily temephos, is documented in Asia (China, Pakistan, Malaysia, and Thailand), Central and South America (Caribbean islands and Brazil) and Europe (Italy and Greece) [7,68,69]. Resistance to DDT (dichlorodiphenyltrichloroethane) and pyrethroids is recognized in Ae. albopictus populations native to Asia, including China,Pakistan,Malaysia andThailand, and is emerging in Africa (Cameroon) [7,68,69,71–74]. The monitoring of insecticide resistance, adoption of standardized procedure for resistance assessment, publication of results through a centralized database (i.e., IRbase available via vectorbase.org), and characterization of biomarkers for understanding resistance would be of great benefit to the rational design of control programs [68,69].

Biological control

Emerging control measures include the use of toxins produced by Bacillus thuringiensis israelensis (Bti) as larvacides, the exploitation of endosymbionts such as representatives of the Asaia, Acinetobacter, and Pantoea genera for paratransgenesis, or Wolbachia and the use of predators [57,61,62,75].

Bti toxins differ from other chemical compounds in that they are specific against mosquitoes and black flies. This specificity makes Bti the most widely used biological insecticide in the USA, and it is commercialized in various formulations with different lasting/releasing properties [76]. Recently, Bti application was shown to have a negative impact on DENV transmission by Ae. albopictus and Ae. aegypti in Malaysia [77]. However, the European Food Safety Authority identified gaps in the knowledge of the safety of Bti, primarily in relation to potential horizontal transfer of genetic material from Bti to other organisms and Bti interference with the analytical systems for quality control of drinking water [78].

Several Wolbachia strains, including wRi, wMelPop, w Pip, and w Mel, were transferred successfully into Ae. albopictus through embryonic microinjection [79–84].

wRi, originally from *Drosophila simulans*, induces either unidirectional or bidirectional CI toward wild type superinfection (i.e., Ae. albopictus is naturally coinfected with two Wolbachia types) [79,80]. The wMel strain of Wolbachia, found naturally in D . melanogaster, reduces susceptibility to DENV and CHIKV, induces immune gene upregulation, and produces bidirectional CI when transinfected into Ae. albopictus [83,84]. Strong CI and anti-dengue resistance also were observed in Ae. albopictus transinfected with w Pip, originally from C. *pipiens*. These promising results led scientists of the Sun Yat-sen University, the Guangzhou Center for Disease Control and Prevention, and Michigan State University to prepare a field trial in Southern China for the implementation of a vector control strategy based on mosquitoes infected with w Pip (Z, X) , unpublished data). Wolbachia-based strategies focused on Ae. aegypti are being implemented already in Australia and are under development in Vietnam, Indonesia, and Brazil (Eliminate Dengue Program, http://www.eliminatedengue. com/). The longterm sustainability of a Wolbachia-based strategy against Ae. albopictus depends on further studies aimed at understanding the: (i) fitness effect of novel Wolbachia strains the mosquitoes, (ii) interaction between the novel strains and the two naturally occurring w AlbA and w AlbB strains, and (iii) mechanism of DENV/CHIKV blocking [57,84].

The use of predators, such as larvivorous fishes, copepods, or the elephant mosquito Toxorhynchites splendens, is an alternative biological control strategy that is suited to controlling species such as Ae. albopictus that breed preferentially in containers. Successful application of this strategy was proven against Ae. aegypti [75]. Several laboratory and field trials support the feasibility of this approach for Ae. albopictus [85–87].

Genetic-based strategies

Two main approaches have been proposed that result in either population reduction (decrease or eliminate the target mosquito population) or population replacement (replace the target population with mosquitoes genetically modified in their vector competence) [88]. So far, most progress has been made in population-reduction strategies, which include both the production of genetically engineered mosquito strains impaired in their ability to fly or the use of sterilizing chemicals or γ -irradiation to induce random mutations leading to sterility in mosquitoes before their release in the wild (sterile insect technique, SIT) [57,89,90]. Population replacement strategies for Ae. albopictus are still in their infancy with the main issues being to: (i) generate mosquitoes with zero viral particles in the salivary glands, (ii) assess the epidemiological impact of these genetically engineered mosquitoes, and (iii) identify methodologies for introgression of the transgene into the target population. Extensive progress has been made in acquiring knowledge in fitness, mating, and quality control of mass-rearing of genetically engineered mosquitoes and in addressing issues related to their safety and community acceptance [91–96]. First releases of Ae. albopictus males sterilized through irradiation were undertaken from 2005 to 2009 in urban and suburban areas of Italy, and these resulted in suppression of the local Ae. albopictus population [90].

Concluding remarks

Ae. albopictus is ranked as one of the top 100 most invasive species worldwide and is being recognized as an increasingly important vector. We estimate that from 1990 to today, the number of people living in countries with documented detections of Ae. albopictus increased from 3.2 to 5.1 billion (http://www.nationmaster.com/graph/peo_poppeople-population&date=2011). During the past 10 years this species was the sole or primary vector for a number of DENV and CHIKV outbreaks in Hawaii, Indian Ocean islands, Central Africa, and southern China, and the first DENV autochthonous transmissions in Europe, emphasizing its public health impact. The public health relevance of this vector mosquito results from its: (i) invasiveness and ecological adaptability, (ii) competence for multiple pathogens, (iii) potential as a bridge vector as a consequence of its opportunistic feeding behavior, and (iv) breeding adaptability to urban, rural, and forest areas. The increase of dengue risk in endemic areas is related primarily to urbanization. The widespread distribution of this species in temperate regions fosters concerns for the introduction of DENV, CHIKV, or other pathogens through travel of a viremic person. In addition, the presence of invasive Ae. albopictus can increase transmission of competent native pathogens, as is happening with Dirofilaria in Italy [3].

We summarize here current knowledge on species distribution, vector competence status, ecological adaptability, and available control strategies. Perspectives for novel vector control strategies and future research needs also are highlighted based on current data gaps. One of the challenges is how to integrate current knowledge and translate it into control approaches. The release of the Ae. albopictus genome annotation is expected to facilitate discovery of molecular markers to characterize the genetic diversity of the species, including geographic and temporal variation and variability among populations in biologically relevant traits (i.e., vector competence, insecticide resistance, breeding preferences, climatic adaptation, and competitive ability). A genome-wide approach is suited to providing a comprehensive view of the genetic basis of a complex phenotype and potentially to identifying its markers as shown by studies on diapause [36]. Additionally, the availability of the genome sequence will facilitate comparative studies across mosquito species. This will result in deeper knowledge and in species-to-species transfer of methodologies and experimental procedures (i.e., transgenesis and SIT) [89,90]. Given the widespread and rapid distribution of Ae. albopictus into regions colonized by indigenous and other invasive vectors, studies of competitive interactions and/or coexistence among species are necessary to assess the impact of each species on public health, identify the cross-species impact of broadly targeted control strategies, and prioritize resources for control. For example, insecticide applications to control Ae. aegypti or Culex species could result in selection for resistance in sympatric Ae. albopictus.

Acknowledgments

We thank the participants of the first International Workshop on Aedes albopictus, the Asian tiger mosquito, held in Pavia (Italy) on March 21–22 2013, with particular mention of Peter Armbruster (Georgetown University), Steven Juliano (Illinois State University), Louis Lambrechts (Institute Pasteur), Daniel Lawson (European Bioinformatics Institute), Patrick Mavingui (Centre National de la Recherche Scientifique, Lyon), and Zhiyong Xi (Michigan State University) for their contributions to this review. We thank Yiji Li for the graphics in Figure 2. This work was supported in part by National Institutes of Health grants R21AI098652-0, U54AI065359, and R01AI083202, and the European Commission Framework Programme FP7 Research Infrastructures project grant 228421.

References

- 1 Paupy, C. et al. (2009) Aedes albopictus, an arbovirus vector: from the darkness to the light. Microbes Infect. 11, 1177–1185
- 2 Rezza, G. (2012) Aedes albopictus and the reemergence of dengue. BMC Public Health 12, 72
- 3 Schafner, F. et al. (2013) Public health significance of invasive mosquitoes in Europe. Clin. Microbiol. Infect. 19, 685–692
- 4 Caminade, C. et al. (2012) Suitability of European climate for the Asian tiger mosquito Aedes albopictus: recent trends and future scenarios. J. R. Soc. Interface 9, 2708–2717
- 5 Benedict, M.Q. et al. (2007) Spread of the tiger: global risk of invasion by the mosquito Aedes albopictus. Vector Borne Zoonotic Dis. 7, 76–85
- 6 Medlock, J.M. et al. (2012) A review of the invasive mosquitoes in Europe: ecology, public health risks, and control options. Vector Borne Zoonotic Dis. 12, 435–447
- 7 Rai, K.S. (1991) Aedes albopictus in the Americas. Annu. Rev. Entomol. 36, 459–484
- 8 Fujioka, K.K. et al. (2012) Discovery of Aedes albopictus (Skuse) in the city of El Monte and the initial response. Proc. Pap. Mosq. Vect. Control Assoc. Calif. 80, 27–29
- 9 Zhong, D. et al. (2013) Genetic Analyasis of invasive Aedes albopictus populations in Los Angeles County, California and its potential public health impact. PLoS ONE 8, e68586
- 10 Santos, R.C. (2003) Updating of the distribution of Aedes albopictus in Brazil (1997–2002). Rev. Saúde Pública 37, 1-4
- 11 Ríos-Velásquez, C.M. et al. (2007) Distribution of dengue vectors in neighborhoods with different urbanization types of Manaus, state of Amazonas, Brazil. Mem. Inst. Oswaldo Cruz 102, 617–623
- 12 Rossi, G.C. et al. (1999) First record of Aedes albopictus (Skuse) from Argentina. J. Am. Mosq. Control Assoc. 15, 422
- 13 Cornel, A.J. and Hunt, R.H. (1991) Aedes albopictus in Africa? First records of live specimens in imported tires in Cape Town. J. Am. Mosq. Control Assoc. 7, 107–108
- 14 Simard, F. et al. (2005) Geographic distribution and breeding site preference of Aedes albopictus and Aedes aegypti (Diptera: Culicidae) in Cameroon, Central Africa. J. Med. Entomol. 42, 726e731
- 15 Holder, P. et al. (2010) A biosecurity response to Aedes albopictus (Diptera: Culicidae) in Auckland, New Zealand. J. Med. Entomol. 47, 600–609
- 16 Tatem, A. et al. (2006) Global traffic and disease vector dispersal. Proc. Natl. Acad. Sci. U.S.A. 103, 6242–6247
- 17 Usmani-Brown, S. et al. (2009) Population genetics of Aedes albopictus (Diptera: Culicidae) invading populations, using mitochondrial nicotinamide adenine dinucleotide dehydrogenase subunit 5 sequences. Ann. Entomol. Soc. Am. 102, 144–150
- 18 Kamgang, B. et al. (2011) Genetic structure of the tiger mosquito, Aedes albopictus, in Cameroon (Central Africa). PLoS ONE 6, e20257
- 19 Delatte, H. et al. (2011) The invaders: phylogeography of dengue and chikungunya viruses Aedes vectors, on the South West islands of the Indian Ocean. Infect. Genet. Evol. 11, 1769–1781
- 20 Lambrechts, L. et al. (2010) Consequences of the expanding global distribution of Aedes albopictus for dengue virus transmission. PLoS Negl. Trop. Dis. 4, e646
- 21 Wu, J.Y. et al. (2010) Dengue fever in mainland China. Am. J. Trop. Med. Hyg. 83, 664–671
- 22 Gasperi, G. et al. (2012) A new threat Looming over the Mediterranean basin: emergence of viral diseases transmitted by Aedes albopictus mosquitoes. PLoS Negl. Trop. Dis. 6, e1836
- 23 de Lamballerie, X. et al. (2008) Chikungunya virus adapts to tiger mosquito via evolutionary convergence: a sign of things to come? Virol. J. 5, 33
- 24 Miller, B.R. et al. (1989) Epidemic yellow fever caused by an incompetent mosquito vector. Trop. Med. Parasitol. 40, 396–399
- 25 Tsetsarkin, K.A. et al. (2007) A single mutation in Chikungunya virus affects vector specificity and epidemic potential. PLoS Pathog. 12, e201
- 26 Vazeille, M. et al. (2007) Two Chikungunya isolates from the outbreak of La Réunion (Indian Ocean) exhibit different patterns of infection in the mosquito, Aedes albopictus. PLoS ONE 2, e1168
- 27 Chevillon, C. et al. (2008) The Chikungunya threat: an ecological and evolutionary perspective. Trends Microbiol. 16, 80–88
- 28 Higa, Y. (2011) Dengue vectors and their spatial distribution. Trop. Med. Health 39, 17–27
- 29 Caputo, B. et al. (2012) The 'auto-dissemination' approach: a novel concept to fight Aedes albopictus in urban areas. PLoS Negl Trop Dis 6, e1793
- 30 Delatte, H. et al. (2010) Blood-feeding behavior of Aedes albopictus, a vector of Chikungunya on La Réunion. Vector Borne Zoonotic Dis. 10, 249–258
- 31 Kamgang, B. et al. (2012) Notes on the blood-feeding behavior of Aedes albopictus (Diptera: Culicidae) in Cameroon. Parasit. Vectors 5, 57
- 32 Valerio, L. et al. (2010) Host-feeding patterns of Aedes albopictus (Diptera: Culicidae) in urban and rural contexts within Rome province, Italy. Vector Borne Zoonotic Dis. 10, 291–294
- 33 Hanson, S.M. and Craig, G.B. (1994) Cold acclimation, diapauses, and geographic origin affect cold hardiness in eggs of Aedes albopictus (Diptera: Culicidae). J. Med. Entomol. 31, 192–201
- 34 Lounibos, L.P. et al. (2003) Asymmetric evolution of photoperiodic diapause in temperate and tropical invasive populations of Aedes albopictus (Diptera: Culicidae). Ann. Entomol. Soc. Am. 96, 512–518
- 35 Poelchau, M.F. et al. (2011) A de novo transcriptome of the Asian tiger mosquito, Aedes albopictus, to identify candidate transcripts for diapause preparation. BMC Genomics 12, 619
- 36 Poelchau, M.F. et al. (2013) Deep sequencing reveals complex mechanisms of diapause preparation in the invasive mosquito, Aedes albopictus. Proc. Biol. Sci. 280, 20130143
- 37 Urbanski, J. et al. (2012) Rapid adaptive evolution of photoperiodic response during invasion and range expansion across a climatic gradient. Am. Nat. 179, 490–500
- 38 Urbanski, J.M. et al. (2010) The molecular physiology of increased egg desiccation resistance during diapause in the invasive mosquito, Aedes albopictus. Proc. Biol. Sci. 277, 2683–2692
- 39 Reynolds, J.A. et al. (2012) Transcript profiling reveals mechanisms for lipid conservation during diapause in the mosquito, Aedes albopictus. J. Insect Physiol. 58, 966–973
- 40 Juliano, S.A. and Lounibos, L.P. (2005) Ecology of invasive mosquitoes: effects on resident species and on human health. Ecol. Lett. 8, 558–574
- 41 Juliano, S.A. (2009) Species interactions among larval mosquitoes: context dependence across habitat gradients. Annu. Rev. Entomol. 54, 37–56
- 42 Brown, J.E. et al. (2011) Worldwide patterns of genetic differentiation imply multiple 'domestications' of Aedes aegypti, a major vector of human diseases. Proc. Biol. Sci. 278, 2446–2454
- 43 Bargielowski, I.E. (2013) Evolution of resistance to satyrization through reproductive character displacement in populations of invasive dengue vectors. Proc. Natl. Acad. Sci. U.S.A. 110, 2888–2892
- 44 Reiskind, M.H. and Lounibos, L.P. (2009) Effects of intraspecific larval competition on adult longevity in the mosquitoes Aedes aegypti and Aedes albopictus. Med. Vet. Entomol. 23, 62–68
- 45 Lounibos, L.P. et al. (2010) Differential survivorship of invasive nosquito species in South Florida cemeteries: do site-specific microclimates explain patterns of coexistence and exclusion? Ann. Entomol. Soc. Am. 103, 757–770
- 46 Leisnham, P.T. and Juliano, S.A. (2012) Impacts of climate, land use, and biological invasion on the ecology of immature Aedes mosquitoes: implications for La Crosse emergence. Ecohealth 9, 217–228
- 47 Yee, D.A. et al. (2013) Implications of saline concentrations for the performance and competitive interactions of the mosquitoes Aedes aegypti (Stegomyia aegypti) and Aedes albopictus (Stegomyia albopictus). Med. Vet. Entomol. http://dx.doi.org/10.1111/mve.12007
- 48 Bagny Beilhe, L. et al. (2013) Ecological interactions in Aedes species on Réunion Island. Med. Vet. Entomol. http://dx.doi.org/10.1111/j.1365-2915.2012.01062.x
- 49 Ponnusami, L. et al. (2008) Diversity of bacterial communities in container habitats of mosquitoes. Microb. Ecol. 56, 593–603
- 50 O'Neal, P.A. and Juliano, S.A. (2013) Seasonal variation in competition and coexistence of Aedes mosquitoes: stabilizing effects of egg mortality or equalizing effects of resources? J. Anim. Ecol. 82, 256–265
- 51 Leisnham, P.T. et al. (2009) Intrapopulation divergenece in competitive interactions of the mosquito Aedes albopictus. Ecology 90, 2405–2413
- 52 Leisnham, P.T. and Juliano, S.S. (2010) Intrapopulation differences in competitive effect and response of the mosquito Aedes aegytpi and resistance to invasion of a superior competitor. Oecologia 164, 221–230
- 53 Weiss, B. and Aksoy, S. (2011) Microbiome influences on insect host vector competence. Trends Parasitol. 27, 514–522
- 54 Ramirez, J.L. et al. (2012) Reciprocal tripartite interactions between the Aedes aegypti midgut microbiota, innate immune system and dengue virus influences vector competence. PLoS Negl. Trop. Dis. 6, e1561
- 55 Iturbe-Ormaetxe, I. et al. (2011) Wolbachia and the biological control of mosquito-borne disease. EMBO Rep. 12, 508–518
- 56 Sinkins, S.P. et al. (1995) Wolbachia pipientis: bacterial density and unidirectional cytoplasmic incompatibility between infected populations of Aedes albopictus. Exp. Parasitol. 81, 284–291
- 57 Zouache, K. et al. (2009) Persistent Wolbachia and cultivable bacteria infection in the reproductive and somatic tissues of the mosquito vector Aedes albopictus. PLoS ONE 4, e6388
- 58 McMeniman, C.J. et al. (2009) Stable introduction of a life-shortening Wolbachia infection into the mosquito Aedes aegypti. Science 323, 141–144
- 59 Mousson, L. et al. (2012) The native Wolbachia symbionts limit transmission of dengue virus in Aedes albopictus. PLoS Negl. Trop. Dis. 6, e1989
- 60 McGraw, E.A. and O'Neill, S.L. (2013) Beyond insecticides: new thinking on an ancient problem. Nat. Rev. Microbiol. 11, 181–193
- 61 Valiente-Moro, C. et al. (2013) Diversity of culturable bacteria including Pantoea in wild mosquito Aedes albopictus. BMC Microbiol. 13, 70
- 62 Minard, G. et al. (2013) Prevalence, genomic and metabolic profiles of Acinetobacter and Asaia associated with field-caught Aedes albopictus from Madagascar. FEMS Microbiol. Ecol. 83, 63–73
- 63 Severson, D.W. and Behura, S.K. (2012) Mosquito genomics: progress and challenges. Annu. Rev. Entomol. 57, 143–166
- 64 Arca', B. et al. (2007) An insight into the sialome of the adult female mosquito Aedes albopictus. Insect Biochem. Mol. Biol. 37, 107–127
- 65 WHO (2004) Global Strategic Framework for Integrated Vector Management (Document WHO_CDS_CPE_PVC_2004), World Health **Organization**
- 66 Fonseca, D.M. et al. (2013) Area-wide management of Aedes albopictus: gouging the efficacy of traditional integrated pest control measures against urban container mosquitoes. Pest Manag. Sci. http:// dx.doi.org/10.1002/ps.3511
- 67 WHO (2011) Global Insecticide Use for Vector-Borne Disease Control, a 10 Year Assessment (2000–2009) (5th edn) (Document WHO/HTM/ NTD/VEM/WHOPES/2011.6), World Health Organization
- 68 Ranson, H. et al. (2010) Insecticide resistance in dengue vectors. TropIKA.net J. 1, (http://journal.tropika.net)
- 69 Vontas, J. et al. (2012) Insecticide resistance in the major dengue vectors Aedes albopictus and Aedes aegypti. Pestic. Biochem. Physiol. 104, 126–131
- 70 WHO and the European Mosquito Control Association (2011) Guidelines for the Control of Invasive Mosquitoes and Associated Vector-Borne Diseases on the European continent. WHO/EMCA conference on the vector-related risk of introduction of Chikungunya and Dengue fever and the spread of Ae. albopictus and Ae. japonicus within Europe, World Health Organization http://www.emca-online.eu/ documents/visitors/WHO2011_revised.pdf
- 71 Kamgang, B. et al. (2011) Insecticide susceptibility of Aedes aegypti and Aedes albopictus in Central Africa. Parasit. Vectors 4, 79
- 72 Khan, H.A. et al. (2011) First report of field evolved resistance to agrochemicals in dengue mosquito, Aedes albopictus (Dipera: Culicidae) from Pakistan. Parasit. Vectors 4, 146
- 73 Wan-Norafikah, O. et al. (2013) Susceptibility of Aedes albopictus Skuse (Diptera Culicidae) to permetrhin in Kuala Lumpur, Malaysia. Asian Biomed. 7, 51–62
- 74 Chuaycharoensuk, T. et al. (2011) Frequency of pyrethorid resistance in Aedes aegypti and Aedes albopictus (Diptera: Culicidae) in Thailand. J. Vector Ecol. 36, 204–212
- 75 Chang, M.S. et al. (2011) Challenges and future perspective for dengue control in the Western Pacific Region. West. Pac. Surveill. Response J. 2, 9–16
- 76 Sulaiman, S. et al. (1997) Field evaluation of Vectobac G, Vectobac 12AS and Cactimos WP against the dengue vector Aedes albopictus in tires. J. Vector Ecol. 22, 122–124
- 77 Tan, A.W. et al. (2012) Spray application of Bacillus thuringiensis israelensis (Bti strain AM65-52) against Aedes aegypti (L.) and Ae. albopictus Skuse populations and impact on dengue transmission in a dengue endemic residential site in Malaysia. Southeast Asian J. Trop. Med. Public Health 43, 296–310
- 78 European Food Safety Authority (2013) Conclusion on the peer review of the pesticide risk assessment of the active substance Bacillus thuringiensis israelensis AM65-52. EFSA J. 11, 3054
- 79 Xi, Z. et al. (2006) Interspecific transfer of Wolbachia into the mosquito disease vector Aedes albopictus. Proc. Biol. Sci. 273, 1317–1322
- 80 Fu, Y. et al. (2010) Artificial triple Wolbachia infection in Aedes albopictus yields a new pattern of unidirectional cytoplasmic incompatibility. Appl. Environ. Microbiol. 76, 5887–5891
- 81 Suh, E. et al. (2009) Pathogenicity of life-shortening Wolbachia in Aedes albopictus after transfer from Drosophila melanogaster. Appl. Environ. Microbiol. 75, 7783–7788
- 82 Calvitti, M. et al. (2010) Characterization of a new Aedes albopictus (Diptera: Culicidae)–Wolbachia pipientis (Rickettsiales: Rickettsiaceae) symbiotic association generated by artificial transfer of the wPip strain from Culex pipiens (Diptera: Culicidae). J. Med. Entomol. 47, 179–187
- 83 Blagrove, M.S. et al. (2012) Wolbachia strain wMel induces cytoplasmic incompatibility and blocks dengue transmission in Aedes albopictus. Proc. Natl. Acad. Sci. U.S.A. 109, 255–260
- 84 Blagrove, M.S. et al. (2013) A Wolbachia wMel transinfection in Aedes albopictus is not detrimental to host fitness and inhibits Chikungunya Virus. PLoS Negl. Trop. Dis. 7, e2152
- 85 Soumare, M.K. and Cilek, J.E. (2011) The effectiveness of Mesocyclops longisetus (Copepoda) for the control of container-inhabiting mosquitoes in residential environments. J. Am. Mosq. Control Assoc. 27, 376–683
- 86 Nyamah, M.A. et al. (2011) Fiels observation on the efficacy of Toxorhynchites spendens (Wiedemann) as biological agent against Aedes albopictus (Skuse) larvae in a cemetery. Trop. Biomed. 28, 312–319
- 87 Dieng, H. et al. (2002) A laboratory and field evaluation of Macrocyclops distinctus, Megacyclopss viridis and Mesocyclops pehpeiensis as control agents of the dengue vector Aedes albopictus in a peridomestic area in Nagasaki, Japan. Med. Vet. Entomol. 16, 285–291
- 88 Terenius, O. et al. (2008) Molecular genetic manipulation of vector mosquitoes. Cell Host Microbe 4, 417–423
- 89 Labbé, G.M. et al. (2012) Female-specific flightless (fsRIDL) phenotype for control of Aedes albopictus. PLoS Negl. Trop. Dis. 6, e1724
- 90 Bellini, R. et al. (2013) Pilot field trials with Aedes albopictus irradiated sterile males in Italian urban areas. J. Med. Entomol. 50, 317–325
- 91 Benedict, M.Q. et al. (2011) Safety of genetically modified mosquitoes. JAMA 305, 2069–2070
- 92 James, A.A. (2011) Genetics-based field studies prioritize safety. Science 331, 398
- 93 James, S. et al. (2011) Ecology. Mosquito trials. Science 334, 771–772
- 94 Subramaniam, T.S. et al. (2012) Genetically-modified mosquito: the Malaysian public engagement experience. Biotechnol. J. 7, 1323–1327
- 95 Facchinelli, L. et al. (2013) Field cage studies and progressive evaluation of genetically-engineered mosquitoes. PLoS Negl. Trop. Dis. 7, e2001
- 96 Bellini, R. et al. (2013) Mating competitiveness of Aedes albopictus radio-sterilized males in large enclosures exposed to natural conditions. J. Med. Entomol. 50, 94–102