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Article

Low Expression of IL-15 and NKT in Tumor Microenvironment Predicts Poor Outcome of MYCN-Non-Amplified Neuroblastoma

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Abstract: Immune tumor microenvironment (TME) in neuroblastoma (NBL) contributes to tumor behavior and treatment response. T cells and natural killer (NK) cells have been shown to play important roles in the neuroblastoma TME. However, few reports address the clinical relevance of natural killer T cells (NKTs) and interleukin-15 (IL-15), one of the crucial cytokines controlling the activation and expansion of NK/NKT cells, in NBL. In this study, we examined NKT immunoscores and IL-15 expression in both MYCN-amplified and MYCN-non-amplified NBL to correlate with clinical outcomes such as event-free survival (EFS) and overall survival (OS). From Gene Expression Omnibus (GEO) datasets GSE45480 ($n = 643$) and GSE49711 ($n = 493$), we found that NKT immunoscore and IL-15 expression were both significantly lower in MYCN-amplified NBL, and similar results were observed using our clinical NBL samples ($n = 53$). Moreover, NBL patients (GEO dataset GSE49711 and our clinical samples) with both lower NKT immunoscore and IL-15 expression exhibited decreased EFS and OS regardless of MYCN gene amplification status. Multivariate analysis further showed that the combination of low NKT immunoscore and low IL-15 expression level was an independent prognostic factor for poor EFS and OS in our NBL patients. These findings provide the rationale for the development of strategy to incorporate IL-15 and NKT cell therapy into the treatment regimen for neuroblastoma.

Keywords: neuroblastoma; tumor microenvironment; NKTs; IL-15; immunoscore; prognosis

1. Introduction

Neuroblastoma (NBL) is a neuroectodermal solid tumor occurring in early childhood with 20~50 cases per million individuals [1]. Disease course and treatment outcome for NBL vary significantly depending on clinical features and molecular risk factors. The International Neuroblastoma Risk Group (INRG) provides a consensus pretreatment classification scheme for risk-stratified therapy according to the stage, age, histology, MYCN (v-myc myelocytomatosis viral related oncogene, neuroblastoma derived (avian)) status, and other

genetic aberrations of NBL patients [2]. Clinical outcomes for the low-risk and intermediate-risk groups have been excellent when treated with surgery and limited chemotherapy. However, despite the multi-modality treatment with surgery, intensive chemotherapy, stem cell transplantation, and radiotherapy, prognosis for high-risk patients has remained dismal. While the addition of anti-disialoganglioside (GD2)-based immunotherapy has increased event-free and overall survival in patients with high-risk NBL [3], there is still room for improvement.

New advances in immunotherapy have recognized the contribution of immune tumor microenvironment (TME) to NBL development and progression [4,5]. Most aggressive NBLs have amplification of the MYCN oncogene [6], which is associated with poor survival [7,8], and analysis of TARGET transcriptomic data has revealed that fewer natural killer (NK) and T cells are present in the TME of MYCN-amplified NBL [9]. In particular, MYCN expression has been shown to inversely correlate with the expression of ligands for NK-cell-activating receptors [10], suggesting that MYCN amplification may have an adverse impact on the activation of NK cells, which are important players in antibody-based immunotherapy [11]. In addition, T cells in the TME of NBL, as defined by the presence of CD3⁺, CD4⁺, and CD8⁺ subpopulations, are associated with favorable clinical outcomes in MYCN-amplified NBL [12], and another study has similarly reported that increased cytotoxic tumor infiltration lymphocytes (TILs) in MYCN-non-amplified NBL is associated with the presence of activated NK/T cells and improved outcomes [13].

In comparison, few studies have addressed the role of natural killer T cells (NKTs) in the NBL immune microenvironment. While Metelitsa et al. has reported that the presence of NKTs within the primary tumor site is associated with improved prognosis in patients with NBL [14–16]; its prognostic values for MYCN-non-amplified NBL versus MYCN-amplified NBL remain to be elucidated. NKTs are CD1d-restricted in that they can be directly cytotoxic against CD1d⁺ cells [17]. Although the majority of human tumors express low levels of CD1d to escape from NKT cytotoxicity, NKTs have been shown to overcome immune evasion by modulation of the TME via inhibition of tumor-associated macrophages (TAMs) [14].

Immunotherapy with dinutuximab, a monoclonal antibody (mAb) against GD2, in combination with granulocyte-macrophage colony-stimulating factor (GM-CSF) [18] and interleukin-2 (IL-2) [19] has been approved for the treatment for high-risk NBL, as GM-CSF and IL-2 are known to augment antibody-dependent cellular cytotoxicity (ADCC) by granulocyte/monocyte and NK/NKTs, respectively. Recent preclinical studies in murine models have demonstrated that the combination of IL-15 and NKTs enhanced anti-tumor activity against NBL [18–20]. Although IL-15 is a key player in NK/NKTs development and homeostatic maintenance [21,22], the role of IL-15 in the NBL TME has not been evaluated. In this study, we examined the expression of NKTs and IL-15 in MYCN-amplified and MYCN-non-amplified NBL and explored their prognostic potentials for NBL.

2. Materials and Methods

2.1. Data Source and Processing

Gene expression profiles and clinical information were obtained from Gene Expression Omnibus (GEO) databases (GEO accession: GSE49711 and GSE45480), and survival data for GSE49711 were acquired from the R2 platform (<https://hgserver1.amc.nl/cgi-bin/r2/main.cgi> (accessed on 25 January 2021)). Specifically, GSE49711 contains RNA-seq data from 498 NBL patients, while GSE45480 include microarray data from 649 NBL patients. Cell type enrichment scores for NKTs (NKT immunoscores) were calculated from gene expression data using the xCell web tool developed by Aran et al. (<http://xcell.ucsf.edu/> (accessed on 25 January 2021)) [23]. Clinical characteristics of the patients involved in GSE49711 and GSE45480 has been previously described [23].

2.2. Neuroblastoma (NBL) Samples

Archival samples of cDNA derived from a previous collection of primary tumors from 53 NBL patients were used for the study [24]. These patients were enrolled in studies between 1986 and 1995, and we obtained the samples through the Children's Oncology Group, the Pediatric Oncology Group, and the Cooperative Human Tissue Network. The study was conducted according to the guidelines of the Declaration of Helsinki, and approved by the Institutional Review Board of Chang Gung Memorial Hospital, Taiwan (protocol code 201701442A3; 11 December 2017). Clinicopathologic information of the participants were summarized in Table 1 and are representative of the NBL population in general. The 5-year event-free survival (EFS) of these patients was 58.8% with a median follow-up time of 3.1 years, while the 5-year overall survival (OS) of these patients was 62.0% with a median follow-up time of 3.47 years.

Table 1. Clinical and pathological characteristics of 53 neuroblastoma (NBL) patients.

Characteristics	Number (%)
Age at diagnosis	
<1.5 years	32 (60.4)
≥1.5 years	21 (39.6)
INSS stage	
1	7 (13.2)
2	10 (18.9)
3	10 (18.9)
4	20 (37.7)
4s	6 (11.3)
Risk group	
Low and intermediate	30 (56.6)
High	23 (43.4)
Histology ^a	
Favorable	19 (35.8)
Unfavorable	13 (24.5)
MYCN	
Non-amplified	42 (79.2)
Amplified	11 (20.7)
Progression/Event	
No	34 (64.2)
Yes	19 (35.8)
Death from disease	
Alive	37 (69.8)
Dead	16 (30.2)

^a Data not available in 21 patients.

2.3. Real-Time Reverse-Transcription Polymerase Chain Reaction

Total RNA was isolated from NBL specimens using TRIzol (Invitrogen, Carlsbad, CA, USA), and RNA (1 µg) was converted to cDNA using the High-Capacity cDNA Reverse Transcription Kit (Applied Biosystems, Foster, CA, USA) in accordance with the manufacturer's protocol. The human IL-15 qPCR primer pair was purchased from OriGene (CAT#: HP233499, Rockville, MD, USA), whereas the Vα24 (5'-CTGGAGGGAAAGAACTGC-3', 5'-TGTCAGGGAAACAGGACC-3') and the glyceraldehyde-3-phosphate dehydrogenase (GAPDH) (5'-CCACTCCTCCACCTTT-3', 5'-ACCACCCTGTTGCTGT-3') primer pairs were designed as described by Grose et al. [25]. Expression levels of Vα24 and IL-15 were determined via quantitative RT-PCR with the SYBR Green Real-Time PCR Master Mix using the Applied Biosystems 7500 Fast Real-Time PCR System, with GAPDH serving as the endogenous control. In accordance with the manufacturer's protocol, 10 ng of

cDNA template were added for each 10 μ L qRT-PCR reaction. The fluorescent signals were analyzed by Applied Biosystems 7500 Software v2.0.6.

2.4. Statistical Analysis

Relative levels of V α 24 (a member of the subfamily of the T cell receptor which conserved expressed on NKTs) and IL-15 mRNAs were expressed as $-\Delta\text{Ct}$ after subtracting the Ct of the reference gene, GAPDH, from that of the genes of interest. Their prognostic values were evaluated via receiver operating characteristic (ROC) area under the curve (AUC) analysis, and the Youden index (sensitivity + specificity – 1) was calculated to determine optimal cut-off values for high versus low gene expression levels. Survival curves were plotted using the Kaplan–Meier method, with the log-rank test applied for comparison. The Cox proportional hazards regression model was employed to identify independent prognostic factors. Statistical computations were performed with Prism 7.0 (GraphPad Software, La Jolla, CA, USA) and SPSS V22.0 (IBM, Armonk, NY, USA) software.

3. Result

3.1. NKTs and IL-15 Expression Were Increased in the Tumor Microenvironment of MYCN-Non-Amplified NBL

As IL-15-related cell signaling regulates NKT proliferation, differentiation, and survival [26–28], we analyzed two large NBL datasets, GSE45480 ($n = 643$) and GSE49711 ($n = 493$), to elucidate the clinical relevance of IL-15 and NKT expression. As shown in Figure 1, IL-15 expression level was significantly higher in MYCN-non-amplified patients than in MYCN-amplified patients in both GSE45480 (median expression value: 3.05 vs. 2.81, $p < 0.001$) (Figure 1A) and GSE49711 (median log FPKM: 12.4 vs. 11.5, $p < 0.001$) (Figure 1B). Consistent with previous reports [16], NKT immunoscores were higher in the MYCN-non-amplified group than the MYCN-amplified group in both GSE45480 (median NKT immunoscore: 0.2494 versus 0.2019, $p < 0.001$) (Supplementary Figure S1A) and GSE49711 (median NKT immunoscore: 0.0761 versus 0.0393, $p < 0.001$) (Supplementary Figure S1B). To validate these findings, we evaluated the expression levels of V α 24 and IL-15 in our 53 NBL clinical samples and found that they were both higher in the MYCN-non-amplified group, although the differences did not reach statistical significance due to small sample size ($p = 0.15$ and 0.19 , respectively) (Supplementary Figure S2).

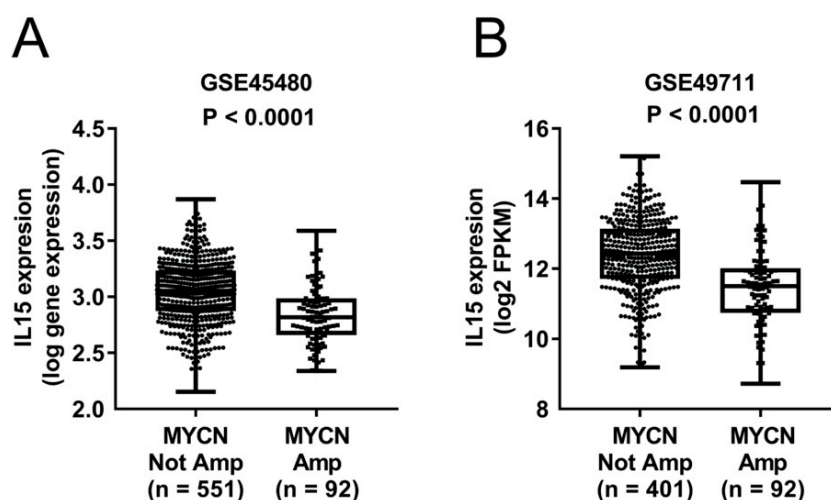


Figure 1. Higher expression of IL-15 in MYCN-non-amplified than MYCN-amplified NBL. IL-15 RNA expression levels were shown as boxplots of log gene expression in GSE45480 (A) and log2 fragments per kilobase of exon per million fragments mapped (FPKM) expression in GSE49711 (B). *t*-test was applied for statistical comparison.

3.2. Lower NKT Immunoscore or IL-15 Expression Level Is Associated with Decreased Event-Free and Overall Survival in NBL Dataset

We next examined the prognostic values of the NKT immunoscore and IL-15 expression level using the GSE49711 dataset. Kaplan–Meier survival analysis showed that NBL patients with low NKT immunoscores exhibited significantly poorer EFS (41.2 versus 71.7%; $p < 0.0001$) and OS (63.2 versus 81.6%; $p < 0.0001$) than those with high NKT immunoscores over the course of follow-up (Figure 2A,D). Similarly, a low level of IL-15 expression was significantly associated with decreased EFS (51.9 versus 66.8%, $p = 0.0006$) and OS (63.2 versus 81.6%, $p < 0.0001$) compared to high level of IL-15 expression (Figure 2B,E). To further evaluate the combined effect of the NKT immunoscore and IL-15 expression level, patients were stratified into three groups: both-high, both-low, and others, for Kaplan–Meier survival analysis. As shown in Figure 2C, patients in the both-low cohort had significantly inferior EFS than those in the both-high group ($p < 0.0001$) at 1 year (62.5 vs. 90.2%), 3 years (37.4 vs. 77.9%), and 5 years (33.5 vs. 75.9%). In addition, patients with both low NKT immunoscore and IL-15 expression had significantly poorer OS than those in the both-high group ($p < 0.0001$) at 1 year (79.9 vs. 99.5%), 3 years (47.2 vs. 94.1%), and 5 years (47.2 vs. 89.4%) (Figure 2F).

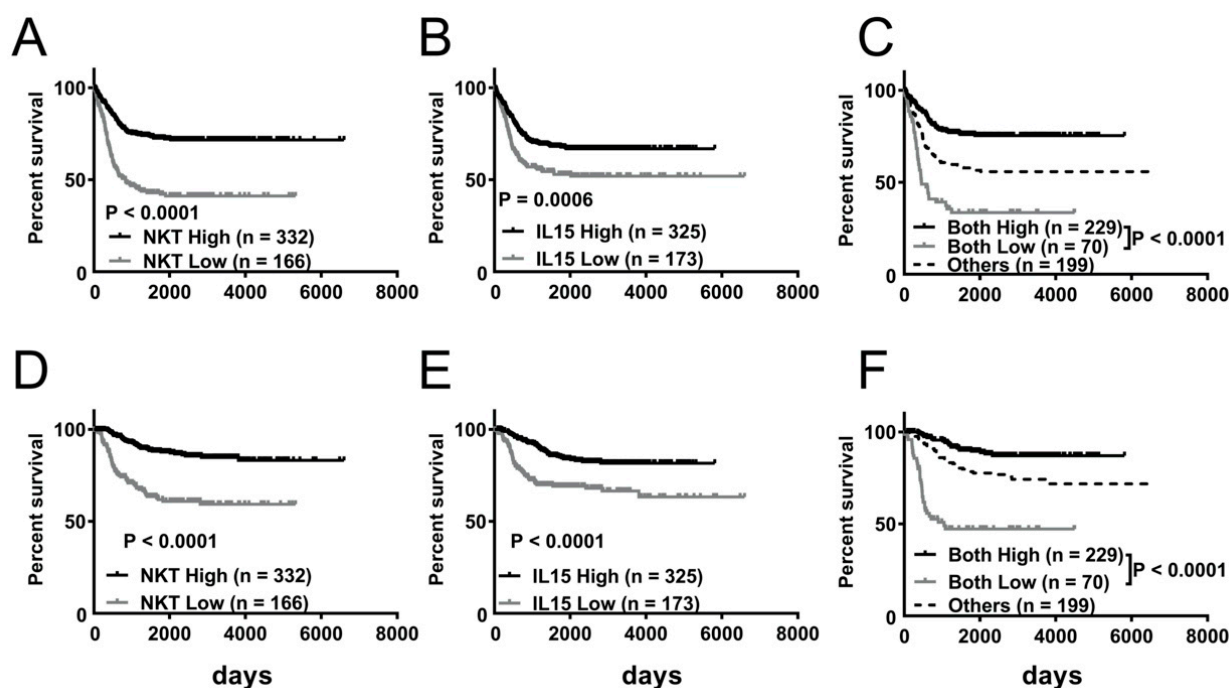


Figure 2. Survival benefit of greater abundance of natural killer T cell (NKT) and IL-15 expression in NBL dataset GSE49711. Kaplan–Meier curves showing event-free survival (EFS) (A–C) and overall survival (OS) (D–F) of NBL according to NKT immunoscore (A,D), IL-15 expression (B,E), and combined NKT immunoscore and IL-15 expression (C,F). Log-rank test was applied for statistical comparison.

3.3. Survival Benefit of Greater Abundance of NKT Alone or in Combination with Higher IL-15 Expression in MYCN-Non-Amplified NBL

Furthermore, we examined the prognostic values of the NKT immunoscore and IL-15 expression level in MYCN-non-amplified patients from GSE49711. Kaplan–Meier survival analysis showed that MYCN-non-amplified NBL patients with low NKT immunoscores had a significantly inferior EFS (51.2 versus 74.5%; $p < 0.0001$) and OS (76.7 versus 85.6%; $p = 0.01$) compared to those with high NKT immunoscores (Figure 3A,D). Similar trends of association between lower IL-15 expression and worse EFS and OS were noted although not statistically significant ($p = 0.39$ and 0.08 , respectively) (Figure 3B,E). To further investigate the combined effect of the NKT immunoscore and IL-15 expression level in the MYCN-non-

amplified subgroup, patients were again stratified into three groups as mentioned above for Kaplan–Meier survival analysis. As shown in Figure 3C, both-low patients exhibited significantly shorter EFS than those in the both-high group ($p = 0.002$) at 1 year (76.9 vs. 90.7%), 3 years (60.9 vs. 79.3%), and 5 years (51.5 vs. 77.2%). In addition, these patients with concurrently low NKT immunoscores and IL-15 expression level had significantly lower OS than those in the both-high group ($p = 0.001$) at 1 year (92.3 vs. 100%), 3 years (72.2 vs. 96.1%), and 5 years (72.2 vs. 91.2%) (Figure 3F).

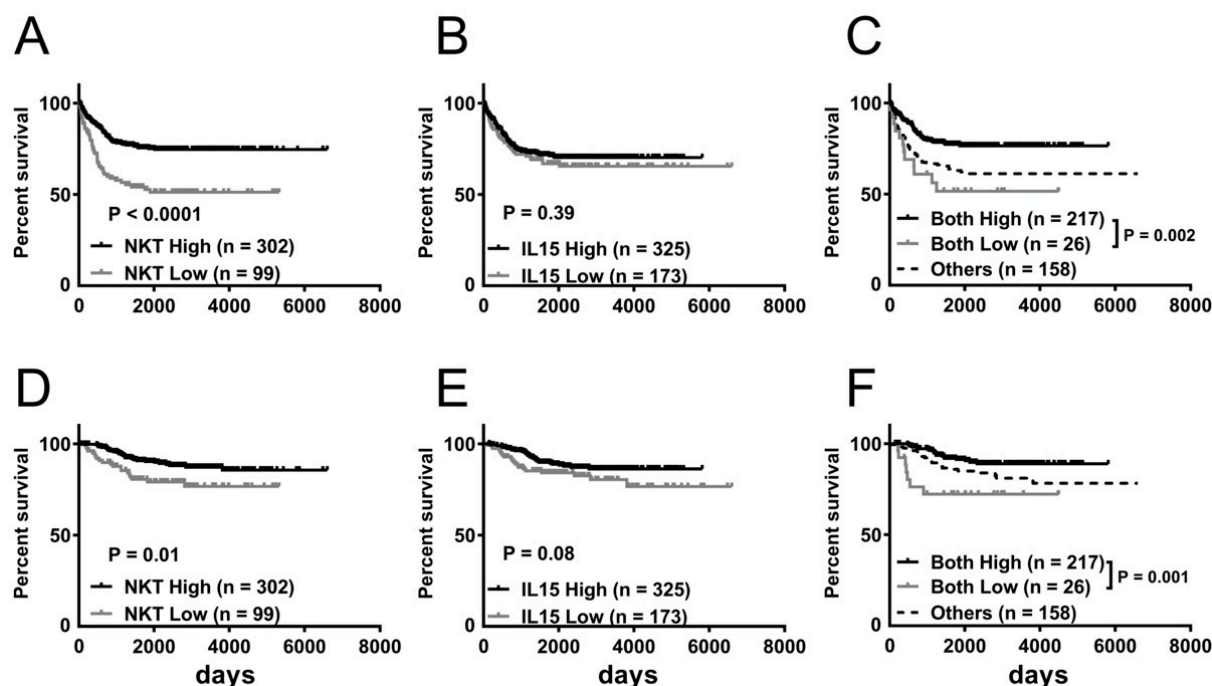


Figure 3. Survival benefit of greater abundance of NKT and IL-15 expression in MYCN-non-amplified NBL dataset GSE49711. Kaplan–Meier curves showing EFS (A–C) and OS (D–F) of MYCN non-amplified NBL according to NKT immunoscore (A,D), IL-15 expression (B,E), and combined NKT immunoscore and IL-15 expression (C,F). Log-rank test was applied for statistical comparison.

3.4. Lower Expression of Vα24 or IL-15 Is Associated with Decreased Overall Survival in NBL Patients

To validate the results of our *in silico* analysis of transcriptomic data, we performed correlative analysis between expression levels of Vα24 and IL-15 from 53 primary NBL samples and the patients' clinical outcome. Kaplan–Meier survival analysis revealed that NBL patients with low Vα24 expression had significantly decreased EFS (38.4 versus 94.4%; $p = 0.0012$) and OS (46.3 versus 94.4%; $p = 0.0076$) compared to those with high Vα24 expression (Figure 4A,D). Similarly, low IL-15 expression was significantly associated with inferior EFS (40.6 versus 83.9%; $p = 0.0045$) and OS (51.1 versus 81.3%, $p = 0.042$) compared to high IL-15 expression (Figure 4B,E). As in our *in silico* analysis, the combined prognostic impact of Vα24 and IL-15 expression levels was determined by stratifying patients for Kaplan–Meier survival analysis. Our results showed that patients with concurrently low Vα24 and IL-15 expressions had significantly lower EFS (32.9 versus 100%; $p = 0.0007$) and OS (45.7 versus 100%; $p = 0.0079$) than those in the both-high group (Figure 4C,F).

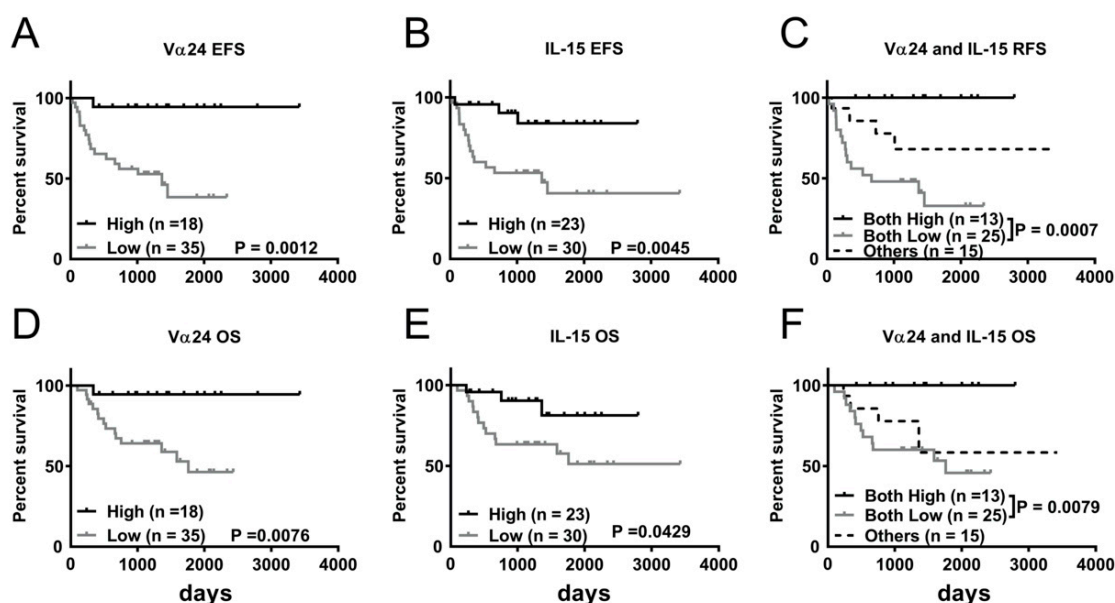


Figure 4. Survival benefit of greater abundance of NKT and IL-15 expression in tumor tissue of NBL patients. Kaplan–Meier curves showing EFS (A–C) and OS (D–F) of 53 NBL patients according to Vα24 (A,D), IL-15 (B,E), and IL-15 + Vα24 combined (C,F) expression. Log-rank test was applied for statistical comparison.

Furthermore, univariate Cox regression analysis of various parameters associated with EFS and OS in NBL revealed that in addition to well-known prognostic factors such as INSS stage, risk category, and age at diagnosis, low expression of Vα24 ($p = 0.012$), low expression of IL-15 ($p = 0.011$), and concurrently low expressions of Vα24 and IL-15 ($p = 0.003$) were all found to be significant predictors of poor EFS (Table 2). In addition, low expression of Vα24 ($p = 0.03$) and concurrently low expressions of Vα24 and IL-15 ($p = 0.03$) were also significant predictive factors of poor OS (Table 3). However, upon multivariate Cox regression analysis, the patient's risk category ($p = 0.003$ and 0.015 , respectively) and low expression of IL-15 ($p = 0.012$ and 0.027 , respectively) in combination with low expression of Vα24 remained the only independent predictors of EFS and OS ($p = 0.009$ and 0.045 , respectively) (Tables 2 and 3).

Table 2. Cox regression analyses of the various factors associated with event-free survival in NBL patients ($n = 53$).

Overall Survival			
Variables	HR (95% CI)	Favorable/Unfavorable	<i>p</i>
Univariate analysis			
Stage	7.64 (2.42–24.15)	1, 2, 3, 4S/4	0.001
Risk group	31.15 (4.09–237.38)	Low, Middle/High	0.001
Age at diagnosis_1.5	5.08 (1.63–15.79)	<1.5 years/≥1.5 years	0.005
Vα24	9.51 (1.26–72.03)	Low/High	0.03
IL-15	3.39 (0.97–11.93)	Low/High	0.05
Vα24 and IL-15	3.55 (1.14–11.01)	All Low/Others	0.03
Multivariate analysis			
Stage	5.04 (0.49–51.34)	1, 2, 3, 4S/4	0.17
Risk group	95.60 (4.59–189.12)	Low, Middle/High	0.003
Age at diagnosis_1.5	1.62 (0.47–5.59)	<1.5 years/≥1.5 years	0.45
Vα24	6.21 (0.80–48.09)	Low/High	0.08
IL-15	4.18 (1.18–14.80)	Low/High	0.027
Vα24 and IL-15	8.14 (1.05–63.06)	All Low/Others	0.045

Abbreviations: HR, hazard ratio; 95% CI, 95% confidence interval.

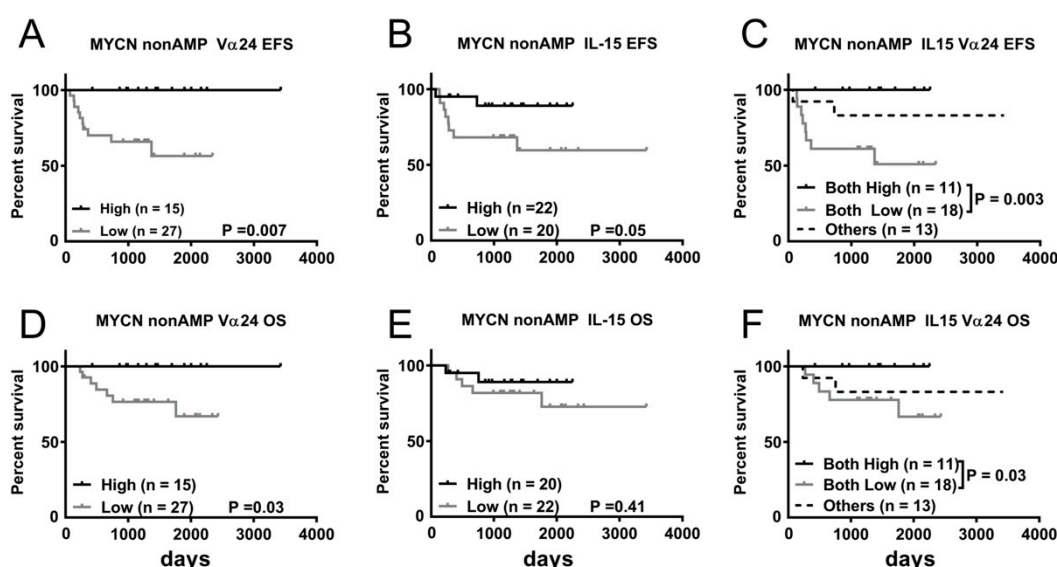
Table 3. Cox regression analyses of the various factors associated with overall survival in NBL patients ($n = 53$).

Event-Free Survival			
Variables	HR (95% CI)	Favorable/Unfavorable	<i>p</i>
Univariate analysis			
Stage	3.69 (1.43–9.49)	1, 2, 3, 4S/4	0.007
Risk group	6.63 (2.18–20.15)	Low, Middle/High	0.001
Age at diagnosis_1.5	3.85 (1.46–10.17)	<1.5 years/≥1.5 years	0.006
Vα24	13.24 (1.76–99.68)	Low/High	0.012
IL-15	5.00 (1.46–17.21)	Low/High	0.011
Vα24 and IL-15	5.50 (1.81–16.6)	All Low/Others	0.003
Multivariate analysis			
Stage	8.51 (0.83–87.84)	1, 2, 3, 4S/4	0.07
Risk group	21.14 (1.79–249.09)	Low, Middle/High	0.015
Age at diagnosis_1.5	2.43 (0.73–8.08)	<1.5 years/≥1.5 years	0.147
Vα24	10.26 (1.33–79.36)	Low/High	0.026
IL-15	4.95 (1.43–17.12)	Low/High	0.012
Vα24 and IL-15	15.55 (1.99–121.41)	All Low/Others	0.009

Abbreviations: HR, hazard ratio; 95% CI, 95% confidence interval.

3.5. Lower Expression of Vα24 or IL-15 Is Associated with Inferior Overall Survival in MYCN-Non-Amplified NBL Patients

We next evaluated the prognostic effects of Vα24 and IL-15 expression levels in MYCN-non-amplified NBL patients. Kaplan–Meier survival analysis indicated that MYCN-non-amplified NBL patients with low Vα24 expression had significantly worse EFS (56.4 versus 100%; $p = 0.007$) and OS (66.9 versus 100%; $p = 0.03$) than those with high Vα24 expression (Figure 5A,D). Low IL-15 expression was associated with poorer EFS with borderline significance but was not significantly associated with OS ($p = 0.05$ and 0.41 , respectively) compared to high IL-15 expression (Figure 5B,E). We also evaluated the combined prognostic value of Vα24 and IL-15 expression levels by stratifying MYCN-non-amplified NBL patients into three groups as described above for Kaplan–Meier survival analysis. As shown in Figure 5C,F, patients with concurrently low Vα24 and IL-15 expressions tend to have inferior EFS (50.9 versus 100%; $p = 0.003$) and OS (66.6 versus 100%; $p = 0.003$) compared to those in the both-high group.

**Figure 5.** Survival benefit of greater abundance of Vα24 and IL-15 expression in tumor tissues of MYCN-non-amplified NBL patients. Kaplan–Meier curves showing EFS (A–C) and OS (D–F) of 53 MYCN-non-amplified NBL patients according to IL-15 (A,D), Vα24 (B,E), and IL-15 + Vα24 combined (C,F) expression. Log-rank test was applied for statistical comparison.

3.6. Lower NKT Immunoscore Is Associated with Inferior Overall Survival in MYCN-Amplified NBL Dataset

Kaplan–Meier survival analysis of MYCN-amplified NBL patients in GSE49711 showed that those with low NKT immunoscores had significantly worse OS than their high NKT immunoscore counterparts (30.0 versus 57.0%; $p = 0.03$), although there was only a trend for shorter EFS ($p = 0.23$) (Supplementary Figure S3D). The low IL-15 expression group showed a similar trend for adverse EFS and OS ($p = 0.15$ and 0.20 , respectively) (Supplementary Figure S3B,E). Patients who were low for both NKT immunoscores and IL-15 expression level had significantly worse OS than those in the both-high group ($p = 0.03$) at 1 year (70.9 vs. 84.5%), 3 years (26.9 vs. 73.5%), and 5 years (26.9 vs. 71.3%) (Supplementary Figure S3F). As shown in Supplementary Figure S3C, there was a similar trend for the worse EFS in the both-low group ($p = 0.13$).

4. Discussion

In this study, we demonstrated that lower expression of both NKTs (V α 24) and IL-15 is significantly associated with poor EFS and OS in NBL patients as well as those in the MYCN-non-amplified NBL subgroup. Moreover, in MYCN-amplified NBL, lower expression of both NKTs and IL-15 significantly correlates with worse OS. These findings provide novel insights for the risk stratification of NBL and suggest that strategies to enhance either NKTs or IL-15 may improve clinical outcomes for high-risk NBL. In addition, association of MYCN amplification with advanced tumor stage and disease progression in NBL has long been known [7,29]. Studies of the MYCN transgenic mouse model have shown that MYCN is sufficient to drive murine NBL tumorigenesis [30]. MYCN is a multifaceted transcription regulator in NBL that can activate genes sustaining growth and repress genes driving differentiation [31]. Furthermore, MYCN in NBL could influence immune surveillance by modulating the infiltration of T cells [9,32], NK cells [10], and NKTs [15], thus supporting the role of MYCN as an immunosuppressive oncogene in high-risk NBL patients. Although MYCN is distinct from MYC (c-myc) [33,34], they are known to share prominent but incomplete redundancy [31]. Recent studies indicate that MYC not only acts as an oncogene but also directly regulates immune responses by various mechanisms that facilitate immune evasion and immunosuppression [35–37]. However, whether MYCN in NBL exerts a similar effect on immune regulation as MYC does in other cancers remains to be investigated.

There is currently no report on the clinical relevance of IL-15 in NBL. We are the first to show that lower IL-15 expression is significantly associated with worse EFS and OS in NBL patients. This is in line with previous reports that IL-15 is an independent prognostic marker for patients with prostate cancer [38], breast cancer [39], and lung adenocarcinoma [40]. We are also the first to report the association between IL-15 and MYCN, although the molecular mechanism remains obscure. IL-15 and IL-2 are known to have similar biologic properties due to shared receptor signaling components (IL-2 receptor β and γ c), but they differ in their distinctive high-binding-affinity α -chain receptors [41]. While the combination of IL-2 and anti-GD2 has been shown to enhance NK cell-mediated ADCC against NBL [42], IL-2-related toxicities remain a clinical challenge [3]. Compared to IL-2, IL-15 does not provoke activation-induced cell death (AICD), induces less regulatory T cell expansion, and is not associated with capillary leak syndrome [43]. Given these advantages, IL-15 has been used as a monotherapy or add-on strategy for cancer treatment [44,45]. The substitution of IL-15 for IL-2 has led to significant preclinical antitumor activity that exceeded the effect of IL-2 in an orthotopic patient-derived xenograft (PDX) model of NBL [20], and co-expression of IL-15 with GD2-specific chimeric antigen receptor (CAR) has enhanced the therapeutic efficacy of CAR-NKTs or CAR-Ts in xenograft mouse models of NBL [18,46]. Taken together, the demonstrated benefit of combining IL-15 with immunotherapy is in line with our findings that high IL-15 expression significantly correlates with better clinical outcomes in NBL patients.

In a study of 98 NBL patients, NKT level as assessed by V α 24 expression is higher in MYCN-non-amplified than MYCN-amplified cases, with higher expression associated with better clinical outcomes [16]. However, the prognostic significance of IL-15 expression in NBL has not been explored. Low levels of NKTs in the TME are associated with worse outcome in this study, which is in agreement with previous reports [16,47] and in line with the cold tumor hypothesis [5] that lack of infiltrating NKTs in TME predicts poor outcome in NBL. However, this is the first study to further analyze NKT expression in the NBL TME according to the tumor's MYCN status, demonstrating that fewer NKTs were associated with worse EFS and OS in MYCN-non-amplified NBL and with worse OS in MYCN-amplified NBL. Although little is known regarding the interaction between NKTs and MYCN in NBL, Metelitsa et al. has shown that MYCN regulates the localization of NKTs to the site of disease in NBL and that NKT infiltration is preferentially found in MYCN-non-amplified NBL in a CCL2-dependent manner [16]. NKTs have the capacity to mount strong anti-tumor responses and have thus become a major focus in the development of effective cancer immunotherapy. It has been reported that IL-15 protects antigen-activated NKTs from suppression by tumor-conditioned hypoxic tumor-associated macrophages [48]. Moreover, IL-15 can expand and activate NK, NKTs, and memory CD8⁺ T cells in TME, leading to tumor destruction [49]. The positive impacts of IL-15 on NK and T cells, in addition to NKT cells, may account for the greater power of combined NKT and IL-15 in predicting outcome of MYCN-non-amplified NBL. Novel strategies to enhance NKT activity by the incorporation of IL-15 in GD2-based immunotherapy, such as anti-GD2 antibody or GD2-specific CAR-Ts or CAR-NKs, are being actively pursued in a phase 1 trial of autologous NKTs engineered to co-express a GD2-specific CAR with IL-15 in three children with relapsed or resistant NBL (NCT03294954). Currently, no dose-limiting toxicities have been observed, and objective response with regression of metastatic bone lesions has been noted in one patient [50]. However, in view of the extremely high cost of CAR-Ts, CAR-NKs, and CAR-NKTs cell therapies, the combination of NKT activators with IL-15 might offer a less expensive alternate approach. We previously reported α -galactosyl ceramide (α -GC) and its analogs as strong activators of the anti-cancer effects of NKTs [51,52], and combining α -GC or its analog with IL-15 may be a promising add-on approach for anti-GD2 immunotherapy in high-risk NBL.

5. Conclusions

This study shows that in NBL patients with MYCN-non-amplified tumors, lower expression of NKTs (V α 24) or IL-15 alone or in combination are independent prognostic factors for poor EFS, and lower expression of IL-15 alone or in combination with low NKTs (V α 24) are independent predictors for worse OS. These findings support the incorporation of IL-15 into various immunotherapeutic strategies for the treatment of neuroblastoma.

Supplementary Materials: The following are available online at <https://www.mdpi.com/2075-4426/11/2/122/s1>, Table S1: Relationships between the expression levels of V α 24 and IL-15 and various clinicopathologic characteristics, Figure S1: The relative abundance of NKT subtype was greater in MYCN-non-amplified NBL, Figure S2: Expression levels of V α 24 and IL-15 in NBL tissues, Figure S3: Kaplan–Meier curves showing EFS and OS of MYCN-amplified NBL according to NKT immunoscore and IL-15 expression in GSE49711.

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Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.

Data Availability Statement: The GEO data presented in this study are openly available in GSE49711 and GSE45480. The archival sample data presented in this study are available on request from the corresponding author. The data are not publicly available due to ethical.

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References

1. Stiller, C.A.; Parkin, D.M. International variations in the incidence of neuroblastoma. *Int. J. Cancer* **1992**, *52*, 538–543. [\[CrossRef\]](#)
2. Pinto, N.R.; Applebaum, M.A.; Volchenboum, S.L.; Matthay, K.K.; London, W.B.; Ambros, P.F.; Nakagawara, A.; Berthold, F.; Schleiermacher, G.; Park, J.R.; et al. Advances in Risk Classification and Treatment Strategies for Neuroblastoma. *J. Clin. Oncol.* **2015**, *33*, 3008–3017. [\[CrossRef\]](#)
3. Yu, A.L.; Gilman, A.L.; Ozkaynak, M.F.; London, W.B.; Kreissman, S.G.; Chen, H.X.; Smith, M.; Anderson, B.; Villablanca, J.G.; Matthay, K.K.; et al. Anti-GD2 antibody with GM-CSF, interleukin-2, and isotretinoin for neuroblastoma. *N. Engl. J. Med.* **2010**, *363*, 1324–1334. [\[CrossRef\]](#)
4. Borriello, L.; Seeger, R.C.; Asgharzadeh, S.; DeClerck, Y.A. More than the genes, the tumor microenvironment in neuroblastoma. *Cancer Lett.* **2016**, *380*, 304–314. [\[CrossRef\]](#) [\[PubMed\]](#)
5. Blavier, L.; Yang, R.M.; DeClerck, Y.A. The Tumor Microenvironment in Neuroblastoma: New Players, New Mechanisms of Interaction and New Perspectives. *Cancers* **2020**, *12*, 2912. [\[CrossRef\]](#) [\[PubMed\]](#)
6. Thompson, D.; Vo, K.T.; London, W.B.; Fischer, M.; Ambros, P.F.; Nakagawara, A.; Brodeur, G.M.; Matthay, K.K.; DuBois, S.G. Identification of patient subgroups with markedly disparate rates of MYCN amplification in neuroblastoma: A report from the International Neuroblastoma Risk Group project. *Cancer* **2016**, *122*, 935–945. [\[CrossRef\]](#) [\[PubMed\]](#)
7. Brodeur, G.M.; Seeger, R.C.; Schwab, M.; Varmus, H.E.; Bishop, J.M. Amplification of N-myc in untreated human neuroblastomas correlates with advanced disease stage. *Science* **1984**, *224*, 1121–1124. [\[CrossRef\]](#)
8. Tang, X.X.; Zhao, H.; Kung, B.; Kim, D.Y.; Hicks, S.L.; Cohn, S.L.; Cheung, N.K.; Seeger, R.C.; Evans, A.E.; Ikegaki, N. The MYCN enigma: Significance of MYCN expression in neuroblastoma. *Cancer Res.* **2006**, *66*, 2826–2833. [\[CrossRef\]](#)
9. Zhang, P.; Wu, X.; Basu, M.; Dong, C.; Zheng, P.; Liu, Y.; Sandler, A.D. MYCN Amplification Is Associated with Repressed Cellular Immunity in Neuroblastoma: An In Silico Immunological Analysis of TARGET Database. *Front. Immunol.* **2017**, *8*, 1473. [\[CrossRef\]](#) [\[PubMed\]](#)
10. Brandetti, E.; Veneziani, I.; Melaiu, O.; Pezzolo, A.; Castellano, A.; Boldrini, R.; Ferretti, E.; Fruci, D.; Moretta, L.; Pistoia, V.; et al. MYCN is an immunosuppressive oncogene dampening the expression of ligands for NK-cell-activating receptors in human high-risk neuroblastoma. *Oncoimmunology* **2017**, *6*, e1316439. [\[CrossRef\]](#)
11. Erbe, A.K.; Wang, W.; Carmichael, L.; Kim, K.; Mendonca, E.A.; Song, Y.; Hess, D.; Reville, P.K.; London, W.B.; Naranjo, A.; et al. Neuroblastoma Patients' KIR and KIR-Ligand Genotypes Influence Clinical Outcome for Dinutuximab-based Immunotherapy: A Report from the Children's Oncology Group. *Clin. Cancer Res.* **2018**, *24*, 189–196. [\[CrossRef\]](#) [\[PubMed\]](#)
12. Mina, M.; Boldrini, R.; Citti, A.; Romania, P.; D'Alicandro, V.; De Ioris, M.; Castellano, A.; Furlanello, C.; Locatelli, F.; Fruci, D. Tumor-infiltrating T lymphocytes improve clinical outcome of therapy-resistant neuroblastoma. *Oncoimmunology* **2015**, *4*, e1019981. [\[CrossRef\]](#) [\[PubMed\]](#)
13. Wei, J.S.; Kuznetsov, I.B.; Zhang, S.; Song, Y.K.; Asgharzadeh, S.; Sindiri, S.; Wen, X.; Patidar, R.; Najaraj, S.; Walton, A.; et al. Clinically Relevant Cytotoxic Immune Cell Signatures and Clonal Expansion of T-Cell Receptors in High-Risk MYCN-Not-Amplified Human Neuroblastoma. *Clin. Cancer Res.* **2018**, *24*, 5673–5684. [\[CrossRef\]](#)
14. Song, L.; Asgharzadeh, S.; Salo, J.; Engell, K.; Wu, H.W.; Sposto, R.; Ara, T.; Silverman, A.M.; DeClerck, Y.A.; Seeger, R.C.; et al. Vα24-invariant NKT cells mediate antitumor activity via killing of tumor-associated macrophages. *J. Clin. Investig.* **2009**, *119*, 1524–1536. [\[CrossRef\]](#)
15. Song, L.; Ara, T.; Wu, H.W.; Woo, C.W.; Reynolds, C.P.; Seeger, R.C.; DeClerck, Y.A.; Thiele, C.J.; Sposto, R.; Metelitsa, L.S. Oncogene MYCN regulates localization of NKT cells to the site of disease in neuroblastoma. *J. Clin. Investig.* **2007**, *117*, 2702–2712. [\[CrossRef\]](#)
16. Metelitsa, L.S.; Wu, H.W.; Wang, H.; Yang, Y.; Warsi, Z.; Asgharzadeh, S.; Groshen, S.; Wilson, S.B.; Seeger, R.C. Natural killer T cells infiltrate neuroblastomas expressing the chemokine CCL2. *J. Exp. Med.* **2004**, *199*, 1213–1221. [\[CrossRef\]](#)
17. Bendelac, A.; Rivera, M.N.; Park, S.H.; Roark, J.H. Mouse CD1-specific NK1 T cells: Development, specificity, and function. *Annu Rev. Immunol.* **1997**, *15*, 535–562. [\[CrossRef\]](#)
18. Xu, X.; Huang, W.; Heczey, A.; Liu, D.; Guo, L.; Wood, M.; Jin, J.; Courtney, A.N.; Liu, B.; Di Pierro, E.J.; et al. NKT Cells Coexpressing a GD2-Specific Chimeric Antigen Receptor and IL15 Show Enhanced In Vivo Persistence and Antitumor Activity against Neuroblastoma. *Clin. Cancer Res.* **2019**, *25*, 7126–7138. [\[CrossRef\]](#) [\[PubMed\]](#)

19. Heinze, A.; Grebe, B.; Bremm, M.; Huenecke, S.; Munir, T.A.; Graafen, L.; Frueh, J.T.; Merker, M.; Rettinger, E.; Soerensen, J.; et al. The Synergistic Use of IL-15 and IL-21 for the Generation of NK Cells From CD3/CD19-Depleted Grafts Improves Their ex vivo Expansion and Cytotoxic Potential Against Neuroblastoma: Perspective for Optimized Immunotherapy Post Haploidentical Stem Cell Transplantation. *Front. Immunol.* **2019**, *10*, 2816. [[CrossRef](#)] [[PubMed](#)]
20. Nguyen, R.; Moustaki, A.; Norrie, J.L.; Brown, S.; Akers, W.J.; Shirinifard, A.; Dyer, M.A. Interleukin-15 Enhances Anti-GD2 Antibody-Mediated Cytotoxicity in an Orthotopic PDX Model of Neuroblastoma. *Clin. Cancer Res.* **2019**, *25*, 7554–7564. [[CrossRef](#)]
21. Matsuda, J.L.; Gapin, L.; Sidobre, S.; Kieper, W.C.; Tan, J.T.; Ceredig, R.; Surh, C.D.; Kronenberg, M. Homeostasis of V α 14i NKT cells. *Nat. Immunol.* **2002**, *3*, 966–974. [[CrossRef](#)]
22. Baev, D.V.; Peng, X.H.; Song, L.; Barnhart, J.R.; Crooks, G.M.; Weinberg, K.I.; Metelitsa, L.S. Distinct homeostatic requirements of CD4+ and CD4- subsets of V α 24-invariant natural killer T cells in humans. *Blood* **2004**, *104*, 4150–4156. [[CrossRef](#)]
23. Aran, D.; Hu, Z.; Butte, A.J. xCell: Digitally portraying the tissue cellular heterogeneity landscape. *Genome Biol.* **2017**, *18*, 220. [[CrossRef](#)]
24. Lin, R.J.; Lin, Y.C.; Chen, J.; Kuo, H.H.; Chen, Y.Y.; Diccianni, M.B.; London, W.B.; Chang, C.H.; Yu, A.L. microRNA signature and expression of Dicer and Drosha can predict prognosis and delineate risk groups in neuroblastoma. *Cancer Res.* **2010**, *70*, 7841–7850. [[CrossRef](#)]
25. Huh, J.Y.; Kim, J.I.; Park, Y.J.; Hwang, I.J.; Lee, Y.S.; Sohn, J.H.; Lee, S.K.; Alfadda, A.A.; Kim, S.S.; Choi, S.H.; et al. A novel function of adipocytes in lipid antigen presentation to iNKT cells. *Mol. Cell. Biol.* **2013**, *33*, 328–339. [[CrossRef](#)]
26. Gamero, A.M.; Ussery, D.; Reintgen, D.S.; Puleo, C.A.; Djeu, J.Y. Interleukin 15 induction of lymphokine-activated killer cell function against autologous tumor cells in melanoma patient lymphocytes by a CD18-dependent, perforin-related mechanism. *Cancer Res.* **1995**, *55*, 4988–4994. [[PubMed](#)]
27. Gordy, L.E.; Bezbradica, J.S.; Flyak, A.I.; Spencer, C.T.; Dunkle, A.; Sun, J.; Stanic, A.K.; Boothby, M.R.; He, Y.W.; Zhao, Z.; et al. IL-15 regulates homeostasis and terminal maturation of NKT cells. *J. Immunol.* **2011**, *187*, 6335–6345. [[CrossRef](#)] [[PubMed](#)]
28. Kennedy, M.K.; Glaccum, M.; Brown, S.N.; Butz, E.A.; Viney, J.L.; Embers, M.; Matsuki, N.; Charrier, K.; Sedger, L.; Willis, C.R.; et al. Reversible defects in natural killer and memory CD8 T cell lineages in interleukin 15-deficient mice. *J. Exp. Med.* **2000**, *191*, 771–780. [[CrossRef](#)]
29. Seeger, R.C.; Brodeur, G.M.; Sather, H.; Dalton, A.; Siegel, S.E.; Wong, K.Y.; Hammond, D. Association of multiple copies of the N-myc oncogene with rapid progression of neuroblastomas. *N. Engl. J. Med.* **1985**, *313*, 1111–1116. [[CrossRef](#)]
30. Weiss, W.A.; Aldape, K.; Mohapatra, G.; Feuerstein, B.G.; Bishop, J.M. Targeted expression of MYCN causes neuroblastoma in transgenic mice. *EMBO J.* **1997**, *16*, 2985–2995. [[CrossRef](#)] [[PubMed](#)]
31. Huang, M.; Weiss, W.A. Neuroblastoma and MYCN. *Cold Spring Harb. Perspect. Med.* **2013**, *3*, a014415. [[CrossRef](#)]
32. Layer, J.P.; Kronmuller, M.T.; Quast, T.; van den Boorn-Konijnenberg, D.; Efferm, M.; Hinze, D.; Althoff, K.; Schramm, A.; Westermann, F.; Peifer, M.; et al. Amplification of N-Myc is associated with a T-cell-poor microenvironment in metastatic neuroblastoma restraining interferon pathway activity and chemokine expression. *Oncimmunology* **2017**, *6*, e1320626. [[CrossRef](#)] [[PubMed](#)]
33. Schwab, M.; Alitalo, K.; Klempnauer, K.H.; Varmus, H.E.; Bishop, J.M.; Gilbert, F.; Brodeur, G.; Goldstein, M.; Trent, J. Amplified DNA with limited homology to myc cellular oncogene is shared by human neuroblastoma cell lines and a neuroblastoma tumour. *Nature* **1983**, *305*, 245–248. [[CrossRef](#)] [[PubMed](#)]
34. Kohl, N.E.; Kanda, N.; Schreck, R.R.; Bruns, G.; Latt, S.A.; Gilbert, F.; Alt, F.W. Transposition and amplification of oncogene-related sequences in human neuroblastomas. *Cell* **1983**, *35*, 359–367. [[CrossRef](#)]
35. Casey, S.C.; Baylot, V.; Felsher, D.W. MYC: Master Regulator of Immune Privilege. *Trends Immunol.* **2017**, *38*, 298–305. [[CrossRef](#)] [[PubMed](#)]
36. Casey, S.C.; Baylot, V.; Felsher, D.W. The MYC oncogene is a global regulator of the immune response. *Blood* **2018**, *131*, 2007–2015. [[CrossRef](#)]
37. Marinkovic, D.; Marinkovic, T. The new role for an old guy: MYC as an immunoplayer. *J. Cell. Physiol.* **2020**. [[CrossRef](#)] [[PubMed](#)]
38. Blum, D.L.; Koyama, T.; M’Koma, A.E.; Iturregui, J.M.; Martinez-Ferrer, M.; Uwamariya, C.; Smith, J.A., Jr.; Clark, P.E.; Bhowmick, N.A. Chemokine markers predict biochemical recurrence of prostate cancer following prostatectomy. *Clin. Cancer Res.* **2008**, *14*, 7790–7797. [[CrossRef](#)]
39. Vilsmaier, T.; Heidegger, H.H.; Schroder, L.; Trapp, E.; Zehni, A.Z.; Rack, B.; Janni, W.; Mahner, S.; Weissenbacher, T.; Jeschke, U.; et al. Interleukin 15 and Eotaxin correlate with the outcome of breast cancer patients vice versa independent of CTC status. *Arch. Gynecol. Obstet.* **2020**. [[CrossRef](#)]
40. Dong, Y.; Liu, Y.; Bai, H.; Jiao, S. Systematic assessment of the clinicopathological prognostic significance of tissue cytokine expression for lung adenocarcinoma based on integrative analysis of TCGA data. *Sci. Rep.* **2019**, *9*, 6301. [[CrossRef](#)]
41. Fehniger, T.A.; Caligiuri, M.A. Interleukin 15: Biology and relevance to human disease. *Blood* **2001**, *97*, 14–32. [[CrossRef](#)] [[PubMed](#)]
42. Frost, J.D.; Hank, J.A.; Reaman, G.H.; Friedrich, S.; Seeger, R.C.; Gan, J.; Anderson, P.M.; Ettinger, L.J.; Cairo, M.S.; Blazar, B.R.; et al. A phase I/IB trial of murine monoclonal anti-GD2 antibody 14.G2a plus interleukin-2 in children with refractory neuroblastoma: A report of the Children’s Cancer Group. *Cancer* **1997**, *80*, 317–333. [[CrossRef](#)]
43. Waldmann, T.A. The biology of interleukin-2 and interleukin-15: Implications for cancer therapy and vaccine design. *Nat. Rev. Immunol.* **2006**, *6*, 595–601. [[CrossRef](#)]

44. Van den Bergh, J.M.; Van Tendeloo, V.F.; Smits, E.L. Interleukin-15: New kid on the block for antitumor combination therapy. *Cytokine Growth Factor Rev.* **2015**, *26*, 15–24. [[CrossRef](#)] [[PubMed](#)]
45. Waldmann, T.A. Interleukin-15 in the treatment of cancer. *Expert Rev. Clin. Immunol.* **2014**, *10*, 1689–1701. [[CrossRef](#)] [[PubMed](#)]
46. Chen, Y.; Sun, C.; Landoni, E.; Metelitsa, L.; Dotti, G.; Savoldo, B. Eradication of Neuroblastoma by T Cells Redirected with an Optimized GD2-Specific Chimeric Antigen Receptor and Interleukin-15. *Clin. Cancer Res.* **2019**, *25*, 2915–2924. [[CrossRef](#)] [[PubMed](#)]
47. Hishiki, T.; Mise, N.; Harada, K.; Ihara, F.; Takami, M.; Saito, T.; Terui, K.; Nakata, M.; Komatsu, S.; Yoshida, H.; et al. Invariant natural killer T infiltration in neuroblastoma with favorable outcome. *Pediatr. Surg. Int.* **2018**, *34*, 195–201. [[CrossRef](#)]
48. Liu, D.; Song, L.; Wei, J.; Courtney, A.N.; Gao, X.; Marinova, E.; Guo, L.; Heczey, A.; Asgharzadeh, S.; Kim, E.; et al. IL-15 protects NKT cells from inhibition by tumor-associated macrophages and enhances antimetastatic activity. *J. Clin. Investig.* **2012**, *122*, 2221–2233. [[CrossRef](#)]
49. Zhang, S.; Zhao, J.; Bai, X.; Handley, M.; Shan, F. Biological effects of IL-15 on immune cells and its potential for the treatment of cancer. *Int. Immunopharmacol.* **2021**, *91*, 107318. [[CrossRef](#)] [[PubMed](#)]
50. Heczey, A.; Courtney, A.N.; Montalbano, A.; Robinson, S.; Liu, K.; Li, M.; Ghatwai, N.; Dakhova, O.; Liu, B.; Raveh-Sadka, T.; et al. Anti-GD2 CAR-NKT cells in patients with relapsed or refractory neuroblastoma: An interim analysis. *Nat. Med.* **2020**, *26*, 1686–1690. [[CrossRef](#)] [[PubMed](#)]
51. Chang, Y.J.; Huang, J.R.; Tsai, Y.C.; Hung, J.T.; Wu, D.; Fujio, M.; Wong, C.H.; Yu, A.L. Potent immune-modulating and anticancer effects of NKT cell stimulatory glycolipids. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 10299–10304. [[CrossRef](#)] [[PubMed](#)]
52. Hung, J.T.; Huang, J.R.; Yu, A.L. Tailored design of NKT-stimulatory glycolipids for polarization of immune responses. *J. Biomed. Sci.* **2017**, *24*, 22. [[CrossRef](#)] [[PubMed](#)]