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Permalink

<https://escholarship.org/uc/item/3130w5nz>

Journal

Nature Human Behaviour, 5(12)

ISSN

2397-3374

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Publication Date

2021-12-01

DOI

10.1038/s41562-021-01119-3

Peer reviewed



Published in final edited form as:

Nat Hum Behav. 2021 December ; 5(12): 1744–1758. doi:10.1038/s41562-021-01119-3.

Resource Profile and User Guide of the Polygenic Index Repository

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Author contributions

D.J.B., D.C., A.O., and P.T. designed and oversaw the study. A.O. supervised all analyses and led the writing of the manuscript. J.B. was the lead analyst, responsible for the GWAS and MTAG analyses, quality control of GWAS summary statistics, and the PGI validation analyses. C.A.P.B. was responsible for quality control of genotype data and the construction of PGIs. G.G., N.W., H.J., and M.B. assisted with analyses. G.G. conducted the illustrative application and wrote the Python code. N.W. designed and implemented the algorithm used to generate Figure 1. R.K.L. ran a meta-analysis of general risk tolerance omitting validation datasets. P.T. derived the measurement-error-correction estimator. A.K., D.A.H., and the 23andMe Research Group conducted genome-wide association analyses for 23andMe. The following authors shared genotype data to enable dataset participation in the Repository: K.M.H. for Add Health; D.W.B., A.C., D.L.C., T.E.M., R.P., K.S., and B.S.W. for Dunedin and E-Risk; A.S. and O.A. for ELSA; L.M. and T.E. for ECGUT; W.G.I. and M.M. for MCTFR; R.A. and P.K.E.M. for STR; T.T.M., K.P.H., and E.M.T. for TTP; and P.H. and J.F. for WLS. More details about dataset-level contributions are in the Supplementary Note. D.W.B. conducted PGI validation analyses in Dunedin and E-Risk. R.A., A.Y., J.P.B., P.K., S.O., M.J., P.V., M.N.M., and D.L. contributed to study design. All authors contributed to and critically reviewed the manuscript. D.J.B., A.O., D.C. and P.T. made especially major contributions to the writing and editing.

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Code availability

The software used for the measurement-error correction is available at https://github.com/JonJala/pgi_correct. The code for constructing PGIs and principal components, the code for the illustrative application, and the code for analyzing the data displayed in Figure 1 is at <https://www.thessgac.org/pgi-repository>.

Competing interests

D.A.H., A.K., and members of the 23andMe Research Team are current or former employees of 23andMe, Inc. and hold stock or stock options in 23andMe. The authors declare no other competing interests.

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Abstract

Polygenic indexes (PGIs) are DNA-based predictors. Their value for research in many scientific disciplines is rapidly growing. As a resource for researchers, we used a consistent methodology to construct PGIs for 47 phenotypes in 11 datasets. To maximize the PGIs' prediction accuracies, we constructed them using genome-wide association studies—some not previously published—from multiple data sources, including 23andMe and UK Biobank. We present a theoretical framework to help interpret analyses involving PGIs. A key insight is that a PGI can be understood as an unbiased but noisy measure of a latent variable we call the “additive SNP factor.” Regressions in which the true regressor is the additive SNP factor but the PGI is used as its proxy therefore suffer from errors-in-variables bias. We derive an estimator that corrects for the bias, illustrate the correction, and make a Python tool for implementing it publicly available.

The ability to predict complex outcomes from genotype data alone is rapidly increasing. The main catalyst behind the increases is the success of genome-wide association studies¹ (GWAS). GWAS estimate the relationship between a trait, called a “phenotype,” and each of millions of genetic variants. The “summary statistics” (coefficients and standard errors) from GWAS can be used to construct a DNA-based predictor of the phenotype, calculated essentially as a coefficient-weighted sum of allele counts^{2,3}. There are a variety of terms used for such DNA-based predictors. In this paper, we will refer to them as “polygenic indexes” (see Box).

As GWAS sample sizes have grown, coefficients are estimated more precisely, enabling the construction of more predictive PGIs. One example is the PGI for educational attainment. The original PGI was constructed from a GWAS of ~100,000 individuals and predicted ~2% of the variance in years of schooling across individuals⁴. The third and most recent PGI for educational attainment (EA) predicts ~12% of the variance⁵. Qualitatively similar patterns have been observed in PGIs for other complex-trait phenotypes^{1,6}, including height, fertility, personality traits, and risk of many common diseases.

PGIs became mainstream in human genetics remarkably quickly. While predictive genetic indexes have a long history in plant and animal genetics⁷, the idea of using GWAS summary statistics to generate a PGI for humans was first proposed in 2007². The first study to empirically construct and validate a PGI was a GWAS of bipolar disorder and schizophrenia

published in 2009³. Soon thereafter, command of methods used to construct PGIs became a standard part of the skill repertoire of analysts specializing in genome-wide data.

Today, PGIs are profoundly impacting research across the disciplinary spectrum. In medicine, much of the discussion revolves around their potential use as tools for identifying individuals who could benefit from enhanced screening and preventive therapies⁸. Though much uncertainty remains about their ultimate clinical utility⁹, one recent study of polygenic risk for five common diseases concluded that the science is sufficiently far along to contemplate incorporating polygenic prediction into clinical care¹⁰. Researchers working at the intersection of the social and natural sciences have articulated visions of how PGIs could be productively leveraged in a number of ways to advance knowledge about important questions^{11–13}. Already, the various iterations of the EA PGI have been used, among other things, to trace out pathways for genetic influences that develop with age¹⁴ and through school¹⁵, study assortative mating^{16,17}, trace recent migration patterns^{18,19}, and improve analyses of the relationship between education and earnings²⁰. As PGIs become more predictive and available for more phenotypes, potential applications will multiply, and novel areas of research are likely to open up.

To depict the rapid growth in research using PGIs, Figure 1 shows the percentage of PGI-related papers presented at the annual meetings of the Behavior Genetics Association. The percentage increased from zero in 2009 to 20% in 2019. The figure also shows how the percentages of papers classified as candidate-gene studies and twin/family/adoption studies—two other commonly used approaches—have evolved over time. The declining fraction of candidate-gene studies in the figure is consistent with the hypothesis of a paradigm shift, with candidate-gene-based approaches gradually being displaced by PGI-based approaches¹³. This shift occurred, at least in part, because PGIs are not subject to some well-known methodological limitations of candidate-gene studies^{21–23}.

In this paper, we hope to promote productive behaviour-genetic research using PGIs in three ways. First and most centrally, we make a broad array of PGIs available via a Polygenic Index Repository, covering a number of datasets that may be useful to social scientists. By constructing the PGIs ourselves and making them available as variables downloadable from the data providers, our resource eliminates a number of roadblocks for researchers who would like to use PGIs in their research, as we detail below. The Repository contains PGIs for 47 phenotypes. To maximize prediction accuracy of the PGIs, we meta-analysed summary statistics from multiple sources, including several large-scale GWASs conducted in UK Biobank and the personal genomics company 23andMe. 23andMe shared summary statistics from 37 separate association analyses, 9 of which have not been reported previously. Therefore, almost all PGIs in our initial release perform at least as well as currently available PGIs in terms of prediction accuracy. We will update the Repository regularly with additional PGIs and datasets.

Second, we present a theoretical framework for interpreting associations with a PGI. Using this framework, we show that a PGI can be understood as an unbiased but noisy measure of what we call the “additive SNP factor,” which is the best linear predictor of the phenotype from the measured genetic variants. Because the PGI is a noisy measure, regressions that

use the PGI as an explanatory variable suffer from errors-in-variables bias. Since different papers use different versions of a PGI, the magnitude of this bias varies. We hope that the theoretical framework helps establish a common language for discussions about the interpretation of PGIs and their effect sizes.

Third, we propose an approach that improves the interpretability and comparability of research results based on PGIs: to use in place of ordinary least squares (OLS) regression, we derive an estimator that corrects for the errors-in-variables bias. (We are aware of four papers to date that have implemented a measurement-error correction along the lines we propose here^{24–27}. Our approach is most similar to that of ref.²⁶, who develops a nearly identical framework using a psychometrics modeling approach but focuses on the univariate case.) The estimator produces coefficients in units of the standardized additive SNP factor, which has a more meaningful interpretation than units of some particular PGI. We illustrate by applying the estimator to multivariate and gene-by-environment regressions from a recently published paper²⁰. We make a Python command-line tool publicly available for implementing the estimator.

For a less technical description of the paper and of how PGIs should—and should not—be interpreted, see the Frequently Asked Questions (FAQs in Supplementary Information).

Results

The Polygenic Index Repository

The Polygenic Index Repository is a resource that addresses several practical obstacles that researchers interested in using PGIs must often confront. These include:

1. Constructing PGIs from individual genotype data can be a time-consuming process, even for researchers trained to work with large datasets.
2. Since the prediction accuracy of a PGI is increasing in the sample size of the underlying GWAS, it is generally desirable to generate PGI weights from GWAS summary statistics based on the largest available samples. However, privacy and IRB restrictions often create administrative hurdles that limit access to summary statistics and force researchers to trade off the benefit of summary statistics from a larger sample against the costs of overcoming the hurdles. In practice, researchers often end up constructing PGIs using only publicly available summary statistics.
3. Publicly available GWAS summary statistics are sometimes based on a discovery sample that includes the target cohort (or close relatives of cohort members) in which the researcher wishes to produce the PGI. Such sample overlap causes overfitting, which can lead to highly misleading results⁹. (Sometimes, when GWAS consortia provide summary statistics upon request from a GWAS that is restricted so as to exclude the cohort, this barrier is surmounted at low cost.)
4. Because different researchers construct PGIs from GWAS summary statistics using different methodologies, it is hard to compare and interpret results from different studies.

We overcome #1 by constructing the PGIs ourselves and releasing them to the data providers, who in turn will make them available to researchers. This simultaneously addresses #2 because we use all the data available to us that may not be easily available to other researchers or to the data providers, including genome-wide summary statistics from 23andMe. Using these genome-wide summary statistics from 23andMe is what primarily distinguishes our Repository from existing efforts by data providers to construct PGIs and make them available, such as the effort by the Health and Retirement Study (<https://hrs.isr.umich.edu/data-products/genetic-data/products#pgs>). It also distinguishes our Repository from efforts to make publicly available PGI weights directly available for download²⁸ (although we also do that, for weights constructed without 23andMe data). To deal with #3, for each phenotype and each dataset, we construct a PGI from GWAS summary statistics that excludes that dataset. We overcome #4 by using a uniform methodology across the phenotypes. In Methods, we detail how the Repository disseminates the PGIs, as well as the principal components of the genetic data in each dataset (which often should be used as controls for ancestry; see Supplementary Methods).

Figure 2 depicts the algorithm that determined which PGIs we constructed. In a preliminary step, we obtained GWAS summary statistics for a comprehensive list of 53 candidate phenotypes (see Supplementary Tables 1 and 2, meta-analysed the summary statistics for each candidate phenotype, and calculated the expected R^2 for an out-of-sample regression of each candidate phenotype on a PGI derived from its GWAS summary statistics. We calculated this expected R^2 from the GWAS summary statistics (see Methods for details). If it exceeded $R^2 = 0.01$, then we used the meta-analysis output to construct a PGI for the phenotype. We call these the “single-trait PGIs.” For each candidate phenotype, we also identified a list of supplementary phenotypes: any other phenotype whose pairwise genetic correlation with the candidate exceeds 0.6 in absolute value. For each candidate with at least one supplementary phenotype, we then calculated the out-of-sample expected R^2 of a PGI derived from a joint analysis of the candidate and supplementary phenotype summary statistics. If the expected R^2 exceeded 0.01, then we used the joint-analysis output to construct a “multi-trait PGI” for the phenotype. When both single-trait and multi-trait PGIs are available, the multi-trait PGI generally has greater predictive power, but the single-trait PGI may be better suited for some applications (see Supplementary Methods).

For each of the 47 phenotypes for which we constructed a single-trait and/or multi-trait PGI, Table 1 lists the total sample size included in the GWAS summary statistics (Total N), followed by the sample-size contributions from three separate sources. For comparison, we also report the sample size of the largest GWAS whose summary statistics are in the public domain (Public N). With three exceptions, Total N exceeds Public N . Two exceptions are height and BMI, where our UKB sample inclusion filters lead to a slightly smaller sample size than the Public N . The remaining exception is cognitive performance, where the sample size of our GWAS is smaller due to overlap between the discovery sample in the largest GWAS with publicly available summary statistics and some of our Repository cohorts. For the remaining phenotypes, the gains in sample size relative to the public N are often substantial, and driven by our inclusion of summary statistics from large-scale GWASs conducted in 23andMe, UKB, or both. Table 1 also shows the 36 and 35 phenotypes for which we created single-trait and multi-trait PGIs, respectively.

We created PGIs for these phenotypes in 11 Repository cohorts that shared their individual-level genetic data with us (regardless of whether the phenotype itself is measured in the cohort). Table 2 lists the datasets and some of their basic characteristics. Each data provider will make these PGIs available to researchers through their own data access procedures (see Supplementary Note).

The UK Biobank is among the 11 cohorts included in the Polygenic Index Repository. Because of its large sample size (see Table 2), the UK Biobank contributes substantially to the available sample for the GWAS for many phenotypes. We therefore did not want to exclude the entire UK Biobank from the GWASs used to create the PGIs. Instead, we split the UK Biobank sample into three equal-sized partitions. We ran three 1/3-sample GWASs for each phenotype. To create the PGI for each partition, we included results from the other two partitions in the meta-analysis. Consequently, researchers can conduct analyses of a PGI in any one of the partitions and obtain unbiased results. However, we caution researchers against conducting analyses in two or three of the partitions and meta-analyzing across partitions; because the other partitions are used to create the PGI, the results obtained across different partitions (although individually unbiased) will be correlated. Meta-analysis standard errors will therefore be anticonservative, and this bias can be substantial (see Methods). Therefore, to maximize the usefulness of our PGIs for research involving related individuals or brain-scan data, we assigned to the same partition all pairs of individuals that are related up to second degree (and some pairs of third degree), as well as all individuals with brain-scan data.

For validating the predictive power of the PGIs, we used five cohorts for which we had access to individual-level genetic and phenotypic data: the Health and Retirement Study, a representative sample of Americans over the age of 50; the Wisconsin Longitudinal Study, a sample of individuals who graduated from high school in Wisconsin in 1957; the Dunedin Multidisciplinary Health and Development Study, a sample of residents of Dunedin, New Zealand, born in 1972–1973; the Environmental Risk (E-Risk) Longitudinal Twin Study, a birth cohort of twins born in England and Wales in 1994–1995; and the UKB (our third partition). The top panel of Figure 3 shows the observed R^2 and 95% confidence intervals for the single-trait PGIs in one or more validation cohorts, depending on which had a measure of the phenotype. Height, BMI, and educational attainment are shown separately because the y -axis scale is different. The bottom panel of Figure 3 shows the difference between the R^2 of the single-trait Repository PGI and that of a PGI we constructed using the largest non-overlapping GWAS whose summary statistics are in the public domain. The Repository PGIs are almost always at least as predictive as the PGIs based on publicly available GWAS results. For the corresponding results for the multi-trait PGIs, see Supplementary Figure 1. The multi-trait PGIs are usually at least as predictive as the single-trait PGIs (Supplementary Figure 1C and Supplementary Table 3).

We have written a User Guide (reproduced in the Supplementary Methods) that will be distributed by participating cohorts along with the Repository PGIs. It discusses interpretational issues, including those relevant for whether researchers should use the single-trait or multi-trait PGIs when both are available.

Theoretical Framework for Polygenic Indexes

To help interpret PGIs, we lay out a theoretical framework. Denote individual i 's phenotype value by y_i^* . Denote individual i 's allele count at genetic variant j by $x_{ij}^* \in \{0, 1, 2\}$. Without loss of generality, we use a mean-centered transformation of the phenotype and allele counts, such that $y_i \equiv y_i^* - E(y_i^*)$ and $x_{ij} \equiv x_{ij}^* - E(x_{ij}^*)$ for each SNP j . We denote the vector of mean-centered allele counts at J genetic variants by $\mathbf{x}_i = (x_{i1}, x_{i2}, \dots, x_{iJ})'$. As a benchmark, consider the standardized best linear predictor of the phenotype based on the allele counts:

$$g_i \equiv \frac{\mathbf{x}_i' \boldsymbol{\gamma}}{sd(\mathbf{x}_i' \boldsymbol{\gamma})},$$

where

$$\boldsymbol{\gamma} = \arg \min_{\tilde{\boldsymbol{\gamma}}} E[(y_i - \mathbf{x}_i' \tilde{\boldsymbol{\gamma}})^2].$$

That is, the optimal weight vector $\boldsymbol{\gamma}$ is the vector of coefficients from the population regression of y_i on \mathbf{x}_i . This population regression may also include control variables; we omit them here to avoid cluttering notation, but in the Supplementary Methods we extend the framework to include them and explain why they do not affect the results in this paper. In the User Guide (also in the Supplementary Methods), we explain how control variables do matter for the interpretation of a PGI.

When the set of genetic variants in \mathbf{x}_i is all variants in the genome, g_i is referred to as the “standardized additive genetic factor.” The variance in the phenotype explained by g_i is called the “(narrow-sense) heritability,” often the object of interest in twin, family, and adoption studies that draw inferences without access to molecular genetic data.

In studies with molecular genetic data—our focus here—the set of genetic variants in \mathbf{x}_i is restricted to those measured or imputed from the single-nucleotide polymorphisms (SNPs) assayed by standard genotyping platforms (and which pass quality-control filters). In that case, the variance in the phenotype explained by g_i is called the “SNP heritability”²⁹, which we denote h_{SNP}^2 . We will refer to g_i as the standardized “additive SNP factor.”

Since the population regression cannot be run, the vector $\boldsymbol{\gamma}$ is unknown, so g_i cannot be constructed empirically. What can be constructed empirically is a “polygenic index (PGI),” \hat{g}_i , which is a standardized, weighted sum of allele counts using some other weight vector $\hat{\boldsymbol{\gamma}}$ calculated from GWAS summary statistics:

$$\hat{g}_i \equiv \frac{\mathbf{x}_i' \hat{\boldsymbol{\gamma}}}{sd(\mathbf{x}_i' \hat{\boldsymbol{\gamma}})}.$$

In general, $\hat{\boldsymbol{\gamma}}$ will not be equal to $\boldsymbol{\gamma}$ because $\hat{\boldsymbol{\gamma}}$ is calculated from GWAS summary statistics that are estimated in a finite sample. The key observation for our framework is that when $\hat{\boldsymbol{\gamma}}$

is calculated using standard methods (that include all the SNPs in \mathbf{x}_j), such as LDpred³⁰ and PRS-CS³¹, the resulting PGI can be expressed as

$$\hat{g}_i = \frac{(g_i + e_i)}{\rho},$$

where e_i is mean-zero estimation error that is uncorrelated with g_i and $\rho \equiv sd(\mathbf{x}_i\hat{\gamma})/sd(\mathbf{x}_i\gamma)$ is a scaling factor that standardizes \hat{g}_i . In words, the PGI is a standardized, noisy measure of the additive SNP factor, where the noise is classical measurement error.

One way to characterize the amount of measurement error is the value ρ . In Methods, we show that

$$\rho^2 = 1 + Var(e_i) = \frac{h_{SNP}^2}{R^2} \geq 1,$$

where h_{SNP}^2 is the SNP heritability (the predictive power of g_i) and R^2 is the fraction of variance explained in a regression of the phenotype y_i on the PGI \hat{g}_i (the predictive power of \hat{g}_i). The ratio h_{SNP}^2/R^2 is greater than or equal to one because the weights that define g_i maximize the variance explained in y_i and therefore any other weights—including those used to construct the PGI—explain at most h_{SNP}^2 of the variation. Furthermore, the amount of measurement error ρ would achieve its minimum value of one only if the PGI weights were based on GWAS summary statistics from an infinite sample.

Across studies, ρ^2 varies. For example, R^2 depends on the sample size of the GWAS underlying the PGI weights and the method of constructing PGI weights (e.g., LDpred vs. PRS-CS). However, ρ^2 can usually be estimated using estimates of h_{SNP}^2 and R^2 from the sample at hand or other samples that are sufficiently similar.

Measurement-Error-Corrected Estimator for PGI Regressions

Typical research with a PGI involves running a regression with the PGI as an explanatory variable and reporting results in units of standard deviations of the PGI. This approach, however, has two shortcomings. First, it is often unclear how to interpret these units, which depend on the amount of measurement error. Second and relatedly, the effect sizes are not comparable across PGIs that differ in their amount of measurement error.

We argue that such a regression should be interpreted as aiming to approximate a regression with the standardized additive SNP factor as the explanatory factor. The PGI serves as an empirically feasible proxy for the standardized additive SNP factor. An analysis of the standardized additive SNP factor has a clearer interpretation than an analysis of the PGI and puts results in comparable units, regardless of which specific PGI was used in the analysis. Here we extend known results from errors-in-variables models to derive a consistent estimator for the coefficients from a regression with the standardized additive SNP factor as an explanatory variable.

The “theoretical regression” is what we call a regression with the (unobserved) standardized additive SNP factor as an explanatory variable. Consider an OLS regression of a phenotype ϕ_i on the standardized additive SNP factor g_i , a vector of covariates z_i , and a vector w_i of interactions between g_i and a subset of the regressors in z_i (possibly all of them):

$$\phi_i = g_i\beta_g + z_i\zeta_g + w_i\delta_g + \epsilon_{g,i}, \quad (1)$$

where the g subscripts indicate that these are parameters from the theoretical regression. (Note that the phenotype ϕ_i need not be the same phenotype y_i for which the standardized additive SNP factor is the best linear predictor. For example, some papers have studied the relationship between the PGI for educational attainment and test scores at younger ages¹⁴. Note also that the covariates in z_i may be measured with error; equation (1) represents whatever regression is run by a researcher except that g_i is measured without error.) The “feasible regression” is what we call the regression using the PGI \hat{g}_i in place of g_i :

$$\phi_i = \hat{g}_i\beta_{\hat{g}} + z_i\zeta_{\hat{g}} + \hat{w}_i\delta_{\hat{g}} + \epsilon_{\hat{g},i}, \quad (2)$$

where \hat{w}_i is the vector of interactions with \hat{g}_i in place of g_i . We denote the vectors of coefficients from the theoretical and feasible regressions by $\alpha_g \equiv (\beta_g, \zeta_g, \delta_g)'$ and $\alpha_{\hat{g}} \equiv (\beta_{\hat{g}}, \zeta_{\hat{g}}, \delta_{\hat{g}})'$, respectively.

In what follows, we sketch the derivation of an estimator for α_g (for details, see the Supplementary Methods). The derivation assumes that the error in the PGI, e_i , is uncorrelated with z_i and w_i . In the Supplementary Methods, we show that this condition holds exactly if the PGI weights $\hat{\gamma}$ are unbiased estimates of γ . We also show that if the PGI weights $\hat{\gamma}$ are estimated using LDpred-inf—as is true for the Repository PGIs—then the bias in our estimator due to plausible violations of this condition will typically be negligible.

Extending the standard formula for errors-in-variables bias³² in a multivariate regression to this setting, and under the assumption that e_i is uncorrelated with z_i and w_i , the feasible-regression coefficients can be shown to be biased:

$$\alpha_{\hat{g}} = P(V_g + \Omega)^{-1}V_g\alpha_g \neq \alpha_g, \quad (3)$$

where is $P \equiv \begin{pmatrix} \rho I_1 + |w| & 0 \\ 0 & I_{|z|} \end{pmatrix}$, $I_{|x|}$ is the identity matrix with the dimensionality of x , V_g is the variance-covariance matrix of $(g_i, w_i, z_i)'$, and Ω is the component of the variance-covariance matrix of $(\hat{g}_i, \hat{w}_i, z_i)'$ that is due to error (see Supplementary Methods). In the special case of a univariate regression, in which the only covariate is a constant term, equation (3) implies that the regression slope coefficient $\beta_{\hat{g}}$ converges to $\frac{1}{\rho}\beta_g$. This is a familiar form of attenuation bias, in which the degree of attenuation toward zero is greater the larger the amount of measurement error. In the multivariate case, however, the amount of attenuation bias for $\beta_{\hat{g}}$ will also depend on the covariance matrix of g_i with z_i . Moreover, the other coefficients, $\zeta_{\hat{g}}$ and $\delta_{\hat{g}}$, will be biased as well, not necessarily toward zero. For example,

a covariate whose coefficient in equation (1) is zero can have a coefficient in equation (2) that is non-zero, leading to an incorrect rejection of the null hypothesis (Abel (2017), unpublished manuscript).

The idea underlying our “corrected” estimator follows immediately from equation (3) by inverting the bias term:

$$\boldsymbol{\alpha}_{corr} = \mathbf{V}_g^{-1}(\mathbf{V}_g + \boldsymbol{\Omega})\mathbf{P}^{-1}\boldsymbol{\alpha}_{\hat{g}} = \boldsymbol{\alpha}_g. \quad (4)$$

This expression is called a regression-disattenuation estimator. It cannot be implemented directly, however, because \mathbf{V}_g involves the variance and covariances of the unobserved standardized additive SNP factor g_j . However, the variance and covariances involving g_j differ from analogous terms involving \hat{g}_j only due to measurement error, and the amount of measurement error is given by ρ . Therefore, the variance and covariances involving g_j can be inferred from estimable quantities. In the Supplementary Methods, we derive an expression for $\boldsymbol{\alpha}_{corr}$ in terms of ρ and population parameters that can be estimated consistently using the observed data. That expression is stated in Methods. We implement that version of the estimator. In the Supplementary Methods, we also derive standard errors for the regression coefficients, under the assumption that ρ is known.

If the PGI is uncorrelated with the covariates, then the estimator will inflate the naïve OLS estimate $\hat{\beta}_g$ and its standard error by the factor ρ . If, in addition, the covariates are uncorrelated with each other, then the estimator will also inflate $\hat{\delta}_g$ and its standard error by the factor ρ . Because the regression coefficients and standard errors are inflated by the same factor, the t -statistics and p -values for the corrected estimates will be identical to those for the uncorrected estimates. Correlation between the PGI and the covariates and correlation among the covariates will lead to deviations from this “rule of thumb” adjustment (and can lead to the adjustment being different across regression coefficients and standard errors).

In the univariate case where ρ is estimated within the same dataset as the PGI analysis is conducted, we show that while uncertainty in \hat{R}^2 causes downward bias in the standard error, uncertainty in \hat{h}_{SNP}^2 causes upward bias, and the net effect is likely to be standard errors that are slightly conservative. We conjecture that the standard errors will also typically be conservative in multivariate settings. If the ρ estimate is from a different dataset, then ignoring the uncertainty in ρ will unambiguously cause the standard errors to be anticonservative. In such settings and in settings when there is meaningful uncertainty in the estimate of ρ , we recommend that users calculate bootstrapped standard errors. The bootstrapped standard errors correctly account for uncertainty in ρ and will be larger than the unbootstrapped standard errors.

We provide a Python command-line tool that implements the measurement-error correction based on a user-specified value of ρ . The package can also estimate ρ by calculating estimates of h_{SNP}^2 (using the GREML method^{29,33} or, for larger datasets, BOLT-REML³⁴) and R^2 . The package can calculate standard errors either treating ρ as known or (at some computational cost) by bootstrapping. When possible, we recommend users estimate ρ

within the dataset they use to analyse the PGI. If the dataset is too small to reliably estimate ρ or lacks a measure of the phenotype corresponding to the PGI, an estimate of ρ from another dataset can be used under the assumption of perfect genetic correlation of the phenotype across datasets. In the Polygenic Index Repository, we provide pre-specified estimates of ρ for three participating datasets for which we have access to the phenotypic data corresponding to the PGI: HRS, WLS, and the third partition of UKB (see Supplementary Table 4). For many of the cohorts, the standard error on the h_{SNP}^2 estimate is large, so we recommend a value of ρ based on existing h_{SNP}^2 and R^2 estimates from a larger sample.

Although our estimator is derived for an OLS estimation framework, it will be approximately correct for logistic regression³⁵ and survival models³⁶ as long as the coefficient on the standardized additive SNP factor, β_g , is not too large. For example, applying a measurement-error correction that would be correct for OLS will be a very accurate approximation for the coefficient in a survival model when the hazard ratio associated with a one-standard deviation difference in the variable measured without error is 1.11³⁶. However, the correction is roughly 20% too small when the hazard ratio is 1.65³⁶.

Illustrative Application

To illustrate our proposed measurement-error correction, we apply it to several analyses reported in a recent paper relating educational attainment (and labour market outcomes) to a PGI for educational attainment²⁰. The paper uses data from the HRS, one of our validation cohorts ($N = 8,537$; 58.3% female; median age = 83). As a preliminary analysis, the paper reports some straightforward tests of the relationship between educational attainment (EA) and the EA PGI. In Panel A of Table 3, we reproduce their univariate regression of EA on the PGI and their multivariate regression that additionally includes controls for mother's and father's EA. In the univariate regression, shown in column (1), a 1-standard-deviation increase in the PGI is associated with 0.844 (95% confidence interval = [0.793, 0.895]) additional years of schooling. This association is reduced to 0.619 (95% confidence interval = [0.572, 0.666]) years in column (2), once the controls are included.

The measurement-error-corrected univariate regression is shown in column (3) of Panel A. We estimate that a 1-standard-deviation increase in the additive SNP factor is associated with 1.318 (95% confidence interval = [1.238, 1.398]) additional years of schooling. Relative to the PGI coefficient in column (1), this coefficient is larger by a factor of $1.318 / 0.844 = 1.56$. In the regression with controls for parental education, shown in column (4), we estimate a corrected coefficient of 1.104 (95% confidence interval = [1.022, 1.186]) additional years. Relative to column (2), this is an increase by a factor of $1.104 / 0.619 = 1.78$. Since for EA in the HRS, $\hat{h}_{SNP}^2 \approx 0.25$ and $\hat{R}^2 \approx 0.10$, according to the rule of thumb mentioned above, both coefficients should be expected to have increased by a factor of 1.58 ($\approx \sqrt{0.25/0.10}$). The increase is larger than that from column (2) to (4) due to the positive correlations between the PGI, the controls, and the dependent variable.

The results in Panel A illustrate a general implication of the measurement-error correction for mediation analyses: the correction deflates estimates of how much covariates mediate

the effect of the PGI. There have been several mediation analyses in which researchers study how much the coefficient on a PGI is reduced when control variables—which are usually positively correlated with both the PGI and the dependent variable—are added to the regression^{37–39}. Going from column (1) to (2), the drop in the coefficient on the PGI would lead a researcher to conclude that parental education mediates $(0.844 - 0.619) / 0.844 = 27\%$ of the effect of the PGI. Going from column (3) to (4) shows the corrected estimate of mediation is only $(1.318 - 1.104) / 1.318 = 16\%$. The drop is larger for the uncorrected regressions because in those regressions, the control variables are proxying for part of the additive SNP factor that is not well captured by the PGI. Therefore, studies that do not correct for measurement error will tend to overestimate the extent to which the control variables mediate the effect of the PGI.

The results in Panel B illustrate a fairly general implication of the measurement-error correction for PGI-by-environment interaction analyses: in contrast to how it affects mediation estimates, the correction tends to increase the magnitude of PGI-by-environment interaction estimates. A main result of Papageorge and Thom is about two such interactions: a higher PGI is associated with a weaker relationship between childhood SES and high school completion but a stronger relationship between childhood SES and college completion²⁰. Columns (1) and (2) reproduce two specifications that show this result: a regression of high school completion on the PGI, self-reported childhood SES, their interaction, and controls; and the analogous regression for college completion. The key finding is that the interaction term is negative in column (1) but positive in column (2). As shown in columns (3) and (4), once the additive SNP factor is considered instead of the PGI, the interaction coefficients for both the high school and college regressions move farther away from zero, strengthening the main result of the paper. In general, PGI-by-environment interaction studies that do not correct for measurement error will tend to underestimate the magnitude of the interaction because the interaction term will tend to be attenuated by the measurement error. Note, however, that this conclusion may not hold if other regressors are correlated with the interaction term.

Discussion

We described the initial release of the Polygenic Index Repository, which contains PGIs for 47 phenotypes. A major goal of this effort is to disseminate PGIs with greater predictive power than the PGIs typically used. To maximize prediction accuracy of the PGIs, we meta-analysed data from multiple sources, including 23andMe and the UK Biobank.

We also derived a measurement-error-corrected estimator that can be used instead of OLS regressions where the independent variables include a PGI or a PGI and its interactions. While some lack of comparability of results across studies is inevitable (e.g., due to differences across samples in SNP heritabilities), one goal of both the Repository and the proposed estimator is to increase comparability. For example, when constructing the PGIs, we applied to each cohort uniform sets of inclusion criteria for individuals and markers in the genotype data. The estimator contributes to improving comparability by putting regression coefficients in units of the additive SNP factor, regardless of the predictive power of the particular PGI available to the researchers.

Because genetic associations are easily misinterpreted, researchers who use PGIs should be especially careful to understand and convey the appropriate interpretation of their findings. For example, it is important to keep in mind that PGI associations may be mediated by environmental factors, and these factors may be modifiable. To facilitate understanding of these and other interpretational issues, we have written a User Guide that cohorts will distribute to users of the Repository PGIs (see Supplementary Methods).

As more GWAS summary statistics become available in the years ahead, and better methods for constructing PGIs are developed, we plan to update the Repository regularly with more predictive PGIs that leverage these advances. For example, future releases will incorporate PGIs of novel phenotypes for which it is not currently feasible to construct PGIs with meaningful predictive power. We emphasize, however, that although PGIs have attained levels of predictive power that can be useful to researchers, the limited heritability of behavioural phenotypes such as those in the Repository implies that the PGIs will never be able to predict any individual's phenotype with much precision. Additionally, since GWAS summary statistics have only been available in large samples of individuals from European ancestries, currently available PGIs have limited portability to individuals of non-European ancestries⁴⁰. In future releases of the Repository, once sufficient data becomes available to create PGIs that have non-negligible predictive power for other ancestry groups, we will update the Repository to contain such PGIs.

Methods

The polygenic indexes (PGIs) shared through the Repository are based on summary statistics from three types of sources: GWASs conducted in UK Biobank (UKB), GWASs conducted in samples of volunteer research participants from 23andMe, and other published genome-wide association studies (GWAS). In Section I below, we begin by detailing how the Repository facilitates researchers' access to the PGIs. In Section II, we describe how the summary statistics used in our main analyses were generated, quality-controlled and meta-analysed to generate a set of files used as inputs into construction of the single-trait and multi-trait PGIs. In Section III, we define and justify the R^2 criterion we used to determine which PGIs to include in the first release of the Repository. We then describe quality-control filters applied to the individual-level genotype data supplied by each Repository cohort. We conclude by describing the methods used to construct the cohort PGIs. In Section IV we state our measurement-error-corrected estimator and its standard error in terms of estimable quantities. Section V describes our estimation of ρ in the HRS, WLS and UKB. Section VI describes the data underlying Figure 1.

I. PGI Dissemination Strategies

Recall from the main text that the PGI Repository aims to overcome four obstacles: (1) Constructing PGIs can be time-consuming and requires specialized knowledge; (2) Researchers face administrative hurdles in accessing all the genome-wide summary statistics for constructing PGIs; (3) Publicly available summary statistics may include the target dataset (which should be omitted when constructing the PGI); and (4) PGIs are often constructed with a variety of methods and idiosyncratic analysis decisions.

Disseminating PGIs through participating datasets—As described in the main text, we overcome all four obstacles by constructing PGIs that include both publicly available data and restricted data (including genome-wide summary statistics from 23andMe) and releasing them to the data providers, who in turn will make them available to researchers. We omit the participating dataset from the summary statistics used to construct that dataset's PGIs, and we construct all the PGIs using a uniform methodology. We similarly construct principal components (PCs) of the genome-wide data using a uniform methodology and release them to the data providers to make available. Our methodology is described below in Sections II and III. The code we used for constructing the PGIs and PCs is posted on the SSGAC website (<https://www.thessgac.org/pgi-repository>).

To access the PGIs and PCs in a dataset, researchers will need to follow the usual data access procedures for that dataset (typically including a Data Use Agreement and approval from an IRB). The current procedures for each dataset are in the Supplementary Note, and up-to-date procedures will be maintained on the SSGAC website.

Data providers can join the Repository if: (i) they share their individual-level genetic data with the SSGAC so that we can construct the PGIs and PCs on our secure servers; and (ii) they have procedures by which external researchers can gain access to the dataset.

Disseminating PGI weights based on public data—For datasets not participating in the Repository, we cannot overcome obstacles (1)-(4). However, for researchers who wish to construct the PGIs in a non-participating dataset, we facilitate this effort by posting on the SSGAC website (www.thessgac.org/data) the weights underlying the PGIs constructed from publicly available data (i.e., that have no 23andMe data). As mentioned above, we also post the code we used for constructing the PGIs and PCs (<https://www.thessgac.org/pgi-repository>). In addition, we refer researchers to 23andMe's "Publication Dataset Access Request Form" (<https://research.23andme.com/dataset-access/#how-to>), which allows researchers to gain access to GWAS results used in published papers (after signing a Data Use Agreement with 23andMe). In this way, researchers can gain access to the same 23andMe data that we used, and use it to construct the PGIs that are wholly or partly based on data from 23andMe.

II. Summary Statistics

UKB GWAS—Supplementary Table 5 lists all UKB phenotypes for which we ran GWASs. Before running the GWASs, we filtered out poor-quality genotypes: (i) samples identified as putatively carrying sex-chromosome configurations that are neither XX nor XY, (ii) samples identified as outliers in heterozygosity and missingness rates, (iii) samples whose sex inferred from sex chromosomes does not match self-reported gender, and (iv) samples with missing sex, birth year, genotyping batch, or PC information. We also restricted the sample to individuals we will refer to as of "European ancestries," defined as the first genetic PC provided by UKB being greater than 0 and individual self-reporting to be of "British", "Irish", or "Any other white background."

In order to make PGIs for the UK Biobank (UKB) without having to exclude the entire UKB from the discovery GWAS, we split the UK Biobank sample into three equal-sized

partitions and, for each partition, used the summary statistics from the other two partitions when generating its PGI. The first partition (UKB1) is composed of UKB participants with brain-scan data (as indicated by data field 12188), all pairs of UKB participants related up to second degree, and the pairs of relatives of third-degree relatedness with greatest relatedness. Pairs of individuals of third-degree relatedness were ordered based on the maximum relatedness coefficient they have with another participant and assigned to the first partition in decreasing relatedness order until the partition was full. Remaining individuals with third-degree relatives were assigned to the second partition. Finally, individuals with no third degree or closer relatives were randomly assigned to the second (UKB2) or third (UKB3) partition.

For all phenotypes in Supplementary Table 5, we ran three separate GWASs, one for each partition. Briefly, each GWAS in UKB was conducted using mixed-linear models implemented by the software BOLT-LMM⁴¹. The dependent variable in each analysis is a phenotype that has been residualized on sex, a third-degree polynomial in birth year (defined as $(birthyear - 1900)/100$), their interactions, 106 genotyping batch dummies, and the first 40 of the PCs released by the UK Biobank. Details on how each phenotype is coded are provided in Supplementary Table 5. For the variance-component estimation in BOLT-LMM (but not the association analyses), we restricted the set of markers to the set of 622,788 hard-called SNP genotypes that remained after filtering for 1% minor allele frequency and 60% imputation accuracy and pruning with an r^2 threshold of 0.3. Our subsequent association analyses were performed on imputed SNP dosages provided by UKB.

Using the UK Biobank split-sample PGI—Splitting the UKB into thirds as described above increases the predictive power of the PGI within each third (relative to omitting the UKB from the GWAS sample). Researchers may desire to conduct analyses that simultaneously include individuals from different partitions of the data or to meta-analyse results across different partitions. Such analyses will produce estimates that are unbiased, but the standard errors will be incorrectly calibrated. To see why, consider a linear model

$$Y_i = X_i\beta + \varepsilon_i,$$

where X_i is a vector of covariates that includes a PGI. Imagine that the data (Y, X) include individuals from different partitions of the data. As a result of the sample-splitting procedure above, $\text{Cov}(X_i, \varepsilon_j) = 0$, which implies that the OLS estimator for β will be unbiased. However, because some of the individuals in the data were used to generate the PGI for other individuals in the data, $\text{Cov}(X_i, \varepsilon_j) \neq 0$ whenever individuals i and j are in different partitions. As a result,

$$\begin{aligned} \text{Var}(\hat{\beta}) &= \text{Var}\left[(X'X)^{-1}X'Y\right] \\ &= \text{Var}\left[(X'X)^{-1}X'\varepsilon\right] \end{aligned} \tag{5}$$

$$\neq (\mathbf{X}'\mathbf{X})^{-1}\mathbf{X}'\text{Var}(\boldsymbol{\epsilon})\mathbf{X}(\mathbf{X}'\mathbf{X})^{-1}. \quad (6)$$

The expression (6) is the standard general formula for the sampling variance of OLS estimates. It is not equal to (5) due to the correlation between $(\mathbf{X}'\mathbf{X})^{-1}\mathbf{X}'$ and $\boldsymbol{\epsilon}$. If we knew the correlation between these two vectors, we could calculate correct standard errors in this setting, but the correlation structure is complex, and we are unaware of any current method that produces correct standard errors. For this reason, we recommend that researchers only do analyses on sets of individuals within a partition. If researchers choose to do analyses with individuals across different partitions, they should include the strong caveat that their standard errors may be poorly calibrated.

23andMe GWAS—Our analyses use summary statistics from GWASs conducted by 23andMe in samples of European-ancestry volunteer research participants for 37 different phenotypes. Supplementary Table 6 provides an overview of these summary statistics. 28 out of the 37 are from previously published studies^{5,42–55}. For these, we cite the original study in the column labelled “Citation”. The remaining 9 are based on previously unreported GWASs. Two of these GWASs are for phenotypes (Subjective Well-Being and Risk) for which GWASs had been previously published by 23andMe but with a smaller sample. The remaining summary statistics have not been previously published by 23andMe. Supplementary Table 6 describes the details of the association model used for each phenotype. For details on 23andMe’s genotyping and imputation, see Supplementary Tables 17 and 18 in Lee et al.⁵

Quality control of summary statistics—We applied a uniform set of quality-control filters to each original file with summary statistics (both those from previously unpublished and previously published GWASs). We closely followed the quality-control pipeline detailed in section 1.5.1 of Okbay et al.³⁷ and implemented in the software EasyQC⁵⁶. Our QC protocol departed from Okbay et al. in the following steps:

- We used data from the Haplotype Reference Consortium reference panel (r1.1)⁵⁷ to check for strand misalignment, allele mismatch, chromosome and base pair position concordance, and allele frequency discrepancies (instead of using data from the 1000 Genomes Phase 1⁵⁸). (Mapping file and allele frequency data were downloaded from the EasyQC website, from the following urls, respectively: https://homepages.uni-regensburg.de/~wit59712/easyqc/HRC/HRC.r1-1.GRCh37.wgs.mac5.sites.tab.rsid_map.gz, <https://homepages.uni-regensburg.de/~wit59712/easyqc/HRC/HRC.r1-1.GRCh37.wgs.mac5.sites.tab.cptid.maf001.gz>.)
- For simplicity and uniformity, we applied a more conservative imputation accuracy filter of 0.7 to all input files irrespective of the software that was used for imputation.
- We applied a uniform minor allele frequency filter of 0.01 to all input files. Stricter filters varying by sample size were not necessary because the studies that we analysed were much larger than some of those in Okbay et al.

- We filtered out standard-error outliers. To do so, we first estimated the standard deviation ($\hat{\sigma}_y$) of the phenotype in each input file by regressing the reported standard errors on the following approximation to the standard error of a coefficient estimated by OLS when the phenotype is standardized:

$$SE_{pred, j} = \frac{1}{\sqrt{N}} \times \frac{1}{\sqrt{2 \times MAF_j \times (1 - MAF_j)}},$$

where MAF_j is the minor allele frequency of SNP j and N is the GWAS sample size. We filtered out markers with $\frac{SE_{pred, j}}{SE_j} < \frac{\hat{\sigma}_y}{2}$ or $\frac{SE_{pred, j}}{SE_j} > 2\hat{\sigma}_y$.

This filter allowed us to identify and remove markers for which the reported GWAS sample size deviated considerably from the sample size implied by the marker's standard error. This filter was particularly relevant for publicly available summary statistics, where marker-specific sample sizes were typically not reported. (Having an accurate number for the sample size is important for LDpred³⁰.)

Before each filtered file was cleared for subsequent meta-analyses, we also prepared and visually inspected a number of diagnostic plots, as described in Okbay et al. Our final analyses are limited to files whose diagnostic plots did not suggest any anomalies. Finally, we examined the genetic correlation between input files (estimated using the LDSC software package⁵⁹) for each phenotype to make sure phenotype coding was in the same direction across 23andMe, UKB, and published studies. Supplementary Table 7 summarizes the number of SNPs dropped in each filtering step in the files that passed all diagnostic checks.

Single-Trait Input GWAS—In this section, we describe the construction of single-trait input GWASs used in several of our downstream analyses, including as inputs for the single-trait and multi-trait PGIs. The single-trait input GWAS for a phenotype is obtained by meta-analysing summary statistics from up to three sources of information: analyses in UKB, analyses in 23andMe, and summary statistics from a previously published study of the phenotype^{5,42,49,50,52,60–73}. The input GWAS for a phenotype is the same across most cohorts. However, when there is overlap between a Repository cohort and cohorts that contributed to summary statistics from previously published studies, or in order to construct a PGI for a UKB partition that is based on summary statistics including the rest of the UKB sample, we restrict the meta-analyses to summary statistics based on non-overlapping data. Details on the construction of single-trait input GWAS are in Supplementary Table 8.

To illustrate the general procedure, consider the single-trait input GWAS for neuroticism in ELSA and EGCUT. Supplementary Table 8 shows that the largest meta-analysis of neuroticism (NEURO1) yielded a final sample of $N = 484,560$ individuals by combining data from UKB ($N = 361,688$), 23andMe ($N = 59,206$) and a previously published study ($N = 63,666$). Since the column does not indicate any overlap with ELSA, the single-trait input GWAS for neuroticism in ELSA is the set of summary statistics from this meta-analysis. EGCUT, however, is listed in Supplementary Table 8 as overlapping with the NEURO1 meta-analysis. The reason is that EGCUT contributed to the summary statistics of the

previously published study (it is one of the cohorts in de Moor et al.⁶⁶). To eliminate overlap, EGCUT's single-trait input is therefore generated by meta-analysing the summary statistics from UKB ($N = 361,688$) and 23andMe ($N = 59,206$) only. This restricted meta-analysis is listed in the table as NEURO2. Similarly, the largest single-trait input GWAS for neuroticism includes the UKB, so all three UKB partitions are listed as overlapping with it. To eliminate overlap, the single-trait input for each UKB partition (which are labelled NEURO3, NEURO4, and NEURO5) is generated by meta-analysing 23andMe, de Moor et al., and the remaining two UKB partitions.

Each input GWAS is conducted by meta-analysing the relevant input files in MTAG⁷⁴. All analyses are conducted allowing for sample overlap and setting all genetic correlations equal to unity. However, we allow the SNP-heritability parameter to vary across input files. Even though MTAG produces a separate output file for each input file, the assumption of perfect genetic correlation ensures that the SNP coefficients in each output file are a constant multiple of each other (hence the PGIs generated by the output files are the same). In all analyses that follow, we adopt the convention of designating the output file with the highest estimated SNP heritability as the input GWAS (this matters for the expected R^2 calculation but nothing else). The details of the heritability estimation are described below, in the subsection "Criterion for Inclusion in Repository" in Section III.

Multi-Trait Input GWAS—For several phenotypes in the first-wave release of the Repository, we provide multi-trait PGIs. Here, we describe the multi-trait input GWAS used to generate each of these.

In a first step, we used LDSC⁵⁹ to estimate genetic correlations between the phenotypes in Supplementary Table 8. For phenotypes with multiple single-trait input GWAS files, we used the version with the largest Total N . This restriction leaves 53 single-trait input GWAS files, each of which is associated with a distinct phenotype. Because there may be sample overlap between the meta-analysed summary statistics, we used GWAS-equivalent sample sizes as reported by MTAG when estimating genetic correlations. (This was the case for Age First Birth, Number Ever Born (men), Number Ever Born (women), and Asthma/Eczema/Rhinitis. For the first three phenotypes, we meta-analysed the publicly available summary statistics from Barban et al.⁷³, which included the first release of UKB, with UKB full release. Similarly, for Asthma/Eczema/Rhinitis, we meta-analysed publicly available summary statistics from Ferreira et al.⁴⁹, which included the first release of UKB, with UKB full release.) The set of pairwise genetic correlations is reported in Supplementary Table 9.

In a second step, we identified each Repository phenotype's supplementary phenotypes. A phenotype is supplementary to a target phenotype (and vice versa) if the pairwise genetic correlation between the phenotypes exceeds 0.6 in absolute value. Under this definition, the estimates in Supplementary Table 9 identify each target phenotype's supplementary phenotypes. These are listed in the column "Input files" of Supplementary Table 10 (set to "No Supplementary Phenotypes" if the phenotype has genetic correlation less than 0.6 with all other phenotypes). For 37 of the 53 Repository phenotypes, we identified at least one supplementary phenotype.

In a final step, for each of these 37 phenotypes, and for each Repository cohort, we ran a multivariate MTAG analysis on the target phenotype together with its supplementary phenotypes, using the version of the target phenotype and each supplementary phenotype for which the cohort is listed in the column “Repository Datasets Sumstats are Used For” in Supplementary Table 8. (In some cases, the same version of the target phenotype and each supplementary phenotype were used for more than one cohort; in those cases, we ran the MTAG analysis only once for that group of cohorts.)

Each MTAG analysis produces multiple output files—one for the target phenotype and one for each of the supplementary phenotypes—but we only retain the summary statistics for the target phenotype. In what follows, we refer to each such file as a multi-trait input GWAS.

For multi-trait MTAG analyses, in order to understand which traits drive results from using multi-trait PGIs, in Supplementary Table 10, we report the average weight that MTAG assigned to each input file in the multi-trait MTAG analyses. These weights may vary by SNP when there is variation in the sample size across SNPs, but the average weights summarize the relative contributions to predictive power.

III. Constructing Repository PGIs

Criterion for Inclusion in Repository—The previous section described how we generated single-trait and multi-trait input GWASs from which it is straightforward to generate single-trait and multi-trait PGIs for a large number of phenotypes. We now describe how we determined, for each candidate phenotype, whether to include neither the single- nor multi-trait PGI, both PGIs, or one of the two in the initial release of the Repository. The structure of our algorithm is outlined in Figure 2. This section provides the details.

For both single- and multi-trait PGIs, we limited the initial set of PGIs released to those with an out-of-sample expected R^2 above 1%. While the threshold itself is arbitrary, the decision to have a threshold was driven by two considerations: the value of a PGI for research is increasing in its predictive power, and we worried that a PGI with low predictive power could cause more harm than good if researchers are tempted to conduct underpowered studies.

We calculated the expected predictive power of each PGI (that might potentially be included in the Repository) using the following formula from Daetwyler et al.⁷⁵:

$$E(R^2) = \frac{(h_{SNP}^2)^2}{h_{SNP}^2 + \frac{M}{N}},$$

where h_{SNP}^2 is the phenotype’s SNP heritability, M is the effective number of independent SNPs which we assume to be equal to 60,000⁹, and N is the GWAS sample size for the phenotype.

We first used the formula above to project the expected predictive power of each potential single-trait PGI. Our projections for the 53 potential PGIs and the underlying parameter

values assumed are shown in Supplementary Table 1. We set h_{SNP}^2 equal to the SNP heritability estimated by LDSC in the summary statistics from the single-trait input GWAS file with the largest sample size for a phenotype. We set N equal to the GWAS-equivalent sample size reported in the MTAG output. For the 37 phenotypes with at least one supplementary phenotype, we generated similar projections for the multi-trait PGIs, using the Multi-Trait Input GWAS files instead. The results of the 37 projections, and the underlying parameter values assumed, are shown in Supplementary Table 2.

We find that our criterion results in 47 phenotypes with at least one PGI in the Repository (see Figure 2). For 12 phenotypes, our procedure results in the release of a single-trait PGI but no multi-trait PGI; these are the phenotypes with no supplementary phenotypes. For 11 other phenotypes, our procedure results in the release of a multi-trait PGI but no single-trait PGI; these are typically phenotypes without large GWASs but for which we have multiple supplementary phenotypes with large GWASs. Finally, our procedure yields 24 phenotypes with both single- and multi-trait PGIs that satisfy our inclusion criterion (Table 1) and 6 phenotypes for which neither PGI qualifies.

Genotyping and Imputation in Repository Cohorts—Genotyping was performed using a range of commercially available arrays. Cohorts were encouraged to upload genotypes imputed against the 1000 Genomes Phase 3⁷⁶ or HRC⁵⁷ imputation panels. Some cohorts provided only genotyped SNPs or data imputed against an older panel. In those cases, we performed the imputation against the HRC reference panel (version 1.1) using the Michigan Imputation Server⁷⁷. Supplementary Table 11 provides study-specific details on the genotyping arrays, pre-imputation quality control filters, imputation software used, and reference samples.

Genotype Data QC in Repository Cohorts—We restricted the set of markers to the SNPs present in the third phase of the international HapMap project (HapMap 3)⁷⁸ in order to reduce computational burden (relative to using all reported SNPs) while keeping a set of markers that covers most of the common variation in individuals with European ancestries.

Subject-level QC in Repository Cohorts—We restricted the samples to individuals with European ancestries. Exclusion criteria were based on the first four principal components of the genetic data. In order to obtain the principal components, for each cohort, we first converted the imputed genotype dosages for HapMap3 SNPs into hard calls. We then merged the data with all samples from the third phase of the 1000 Genomes Project, restricting to SNPs that had a call rate greater than 99% and minor allele frequency greater than 1% in the merged sample. We calculated the principal components (PCs) in the 1000 Genomes subsample and projected these onto the remaining individuals in the merged data. In order to select European-ancestry samples, we plotted the first four PCs against each other and visually identified the individuals that cluster together with the 1000 Genomes EUR sample.

Creation of PCs in Repository Cohorts—In the Repository cohorts, before constructing PCs, we removed markers with imputation accuracy less than 70% or

minor allele frequency less than 1%, as well as markers in long-range LD blocks (chr5:44mb-51.5mb, chr6:25mb-33.5mb, chr8:8mb-12mb, chr11:45mb-57mb). Next, we restricted the sample to individuals with European ancestries, as described immediately above. We further pruned the markers to obtain a set of approximately independent markers, using a 1Mb rolling window (incremented in steps of 5 variants) and an R^2 threshold of 0.1. We used this set of markers to estimate a genetic relatedness matrix. We identified all pairs of individuals with a relatedness coefficient greater than 0.05 as calculated by Plink1.9⁷⁹. We excluded one individual from each pair, calculated the first 20 PCs for the resulting sample of unrelated individuals using Plink 1.9, and projected the PCs onto the sample of unrelated individuals. In HRS, we re-labeled the PCs in sets of five in order to address identifiability concerns.

Constructing PGIs—All PGIs in the initial release of the Repository were constructed in Plink2⁷⁹ using imputed genotype probabilities. Prior to constructing the PGIs, we adjusted the SNP weights for linkage disequilibrium (LD) using LDpred³⁰. We estimated the LD patterns using genotype data from the public release of the HRC Reference Panel (version 1.1) after applying the following quality-control filters. First, we limited the set of variants to HapMap3 SNPs and filtered out variants with genotyping call rate <0.98 and individuals with genotype missingness rate >0.02. Next, we calculated the genomic relatedness matrix and dropped one individual out of each pair with relatedness coefficient >0.025. We clustered the remaining individuals based on their identity-by-state distances using Plink1.9 and dropped an individual if the Z -score corresponding to their distance to their nearest neighbour is less than -5 . In the remaining sample that we fed into LDpred for LD estimation, there were 1,214,408 SNPs and 14,028 individuals. At the coordination step of LDpred, we used the option “--max-freq-discrep” in order to exclude markers that have a frequency discrepancy greater than 0.1 between the summary statistics and genotype data. We also used the “--z-from-se” option so that Z statistics were obtained from the GWAS coefficient estimates and their standard errors rather than from P values (the default) because the latter led to issues in LDpred for markers with extremely small P values. For each PGI, we used the LD window recommended by Vilhjalmsón et al.³⁰, i.e., the number of markers common between the LD reference data, cohort genotype data and summary statistics left after the remaining LDpred quality control filters (MAF > 0.01, no allele mismatch, no ambiguous alleles), divided by 3,000. The fraction of causal markers was set to 1 for each phenotype to ensure consistency across phenotypes.

Prediction Analyses—We conducted a validation exercise for our PGIs in the HRS ($N = 10,978$; 57% female; median age = 82), WLS ($N = 8,937$; 52% female; median age = 82), Dunedin ($N = 887$; 49% female; 1972 birth cohort), E-Risk ($N = 1,968$; 51% female; 1994 birth cohort), and UKB (third partition; $N = 148,662$; 54% female; median age = 71) cohorts. The HRS sample used in our validation exercise (2006–2010) is smaller than the HRS sample for which we are releasing PGIs (2006–2012) because we only had access to phenotype data in the former. Supplementary Table 12 describes the phenotypes used as outcomes in these analyses for all cohorts except UKB. The UKB phenotypes are described in Supplementary Table 5. (The UKB phenotypes used in the prediction exercise differ slightly from the GWAS phenotypes described in Supplementary Table 5 in that they were

not residualized on the PCs and genotyping batch dummies. Instead, we have controlled for these covariates in the regressions when calculating incremental R^2 as described below.) As a general rule, if a single measurement in time was available, we residualized the phenotype on a second-degree polynomial in age, sex, and their interactions. If multiple measurements were available, we either did the same residualization in each wave and took the mean across waves or we took the maximum across waves and then residualized on birth year, sex, and their interactions.

Supplementary Table 3 shows the results from the prediction analyses. The incremental R^2 was calculated as the difference in explained variance when adding the PGI to a regression of the residualized phenotype on the first 10 principal components of the genetic data. In the UKB prediction analyses, we included an additional 10 principal components and 106 genotyping batch dummies. We obtained 95% confidence intervals around the incremental R^2 's by bootstrapping with 1000 repetitions. Supplementary Table 3 also shows the predictive power of “public PGIs”, which are PGIs constructed using our Repository pipeline based on the largest publicly available GWAS on the phenotype that does not have sample overlap with the prediction cohort^{4,5,62–70,72,37,73,80–84,42,49,50,52,55,60,61} (we also use <http://www.nealelab.is/uk-biobank/>). The details of the input GWAS used for each validation cohort for the construction of the “public PGIs” are in Supplementary Table 13.

IV. Measurement-Error-Corrected Estimator

Equation (4) in the main text gives an expression for our measurement-error-corrected estimator, but it cannot be implemented directly because \mathbf{V}_g and $\mathbf{\Omega}$ are based on unobserved variables. In the Supplementary Methods we derive an equivalent expression in terms of variables that can all be consistently estimated using sample analogues:

$$\boldsymbol{\alpha}_{\text{corr}} = \mathbf{P} \begin{bmatrix} \frac{1}{\rho^2} \boldsymbol{\Sigma}_G & \boldsymbol{\Sigma}_{\hat{G}, z} \\ & \boldsymbol{\Sigma}_z \end{bmatrix}^{-1} \mathbf{V}_g \boldsymbol{\alpha}_{\hat{g}}, \quad (7)$$

where

$$\boldsymbol{\Sigma}_G \equiv \begin{bmatrix} 1 & \rho^2 \text{Cov}(\hat{\mathbf{w}}, \hat{g}_i) \\ \rho^2 \boldsymbol{\Sigma}_{\hat{\mathbf{w}}} - (\rho^2 - 1) \boldsymbol{\Sigma}_{\text{int}, z} \end{bmatrix}$$

$\boldsymbol{\Sigma}_{\hat{G}, z} \equiv \text{Cov}[(\hat{g}_i, \hat{\mathbf{w}}_i), \mathbf{z}_i]$, $\boldsymbol{\Sigma}_z \equiv \text{Var}(\mathbf{z}_i)$, $\boldsymbol{\Sigma}_{\hat{\mathbf{w}}} \equiv \text{Var}(\hat{\mathbf{w}}_i)$, $\boldsymbol{\Sigma}_{\text{int}, z} \equiv \text{Var}(\mathbf{z}_{\text{int}, i})$, and $\mathbf{z}_{\text{int}, i}$ is the vector of the covariates that are interacted with g_i to form the vector \mathbf{w}_i .

To obtain standard errors for $\boldsymbol{\alpha}_{\text{corr}}$, we calculate

$$\text{Var}(\boldsymbol{\alpha}_{\text{corr}}) = \mathbf{C} \mathbf{A}_{\hat{g}} \mathbf{C}', \quad (8)$$

where $\mathbf{A}_{\hat{g}} \equiv \text{Var}(\boldsymbol{\alpha}_{\hat{g}})$ and

$$C \equiv P \begin{bmatrix} \frac{1}{\rho^2} \Sigma_G & \Sigma_{\hat{G}, z} \\ & \Sigma_z \end{bmatrix}^{-1} V_g. \quad (9)$$

The standard errors are the square root of the diagonal of $\text{Var}(\mathbf{a}_{\text{corr}})$. Note that equations (7)–(9) are written in terms of population variance-covariance matrices, model coefficients, and the parameter ρ . To implement this correction, we replace each of these terms with its sample counterpart.

V. Estimation of ρ in HRS, WLS and UKB

We estimated the value of ρ for all PGIs satisfying the criterion for inclusion in the Repository in three of our validation datasets: HRS, WLS and UKB (partition 3). Recall from the main text that ρ is defined as

$$\rho = \sqrt{\frac{h_{SNP}^2}{R^2}},$$

where h_{SNP}^2 is the SNP heritability and R^2 is the fraction of variance explained in a regression of the phenotype on the PGI.

In order to estimate h_{SNP}^2 and R^2 , we first took the residualized phenotypes described in section “Prediction Analyses” and additionally residualized these on 20 PCs in HRS and WLS, and 40 PCs and batch effects in UKB3. We did the same for the PGIs. In HRS and WLS, we estimated h_{SNP}^2 with genomic-relatedness-matrix restricted maximum likelihood (GREML) implemented in GCTA v1.93.0beta^{29,33} using HapMap3 SNPs with MAF > 1%. Prior to the h_{SNP}^2 estimation, we dropped one individual from each pair with a relatedness greater than 0.025. We estimated R^2 as the explained variance in a simple regression of the residualized phenotype on the residualized PGI. Standard errors for R^2 , h_{SNP}^2 , and ρ were estimated with a 100-block jackknife procedure.

In UKB3, because of the large sample size, we faced computational constraints. We therefore used the REML implementation in BOLT v2.3³⁴ (with the `--remlNoRefine` option). Moreover, we estimated standard errors only for three phenotypes: friend satisfaction, educational attainment, and height. We chose these three phenotypes so as to have one each corresponding to a single-trait PGI with low (friend satisfaction), medium (educational attainment) and high predictive power (height).

Supplementary Table 4 lists the estimates of ρ for HRS, WLS and UKB3, along with the underlying h_{SNP}^2 and R^2 estimates and standard errors where available.

VI. Categorization of BGA Annual Meeting Presentations

To obtain the data for Figure 1, we first created a dataset containing the titles, authors, and abstracts of all presentations at the 2009–2019 Behavior Genetics Association Annual Meetings. The information about the presentations is printed each year in issue six of the association journal Behavior Genetics. There were 2,034 presentations in this initial dataset. Included in the initial dataset were 36 symposia and 5 papers that were submitted as a part of symposia; all 41 of these are omitted from the final dataset. The final dataset contains a total of 1,993 presentations.

After some trial-and-error and visual inspection of several dozen abstracts, we arrived at the algorithm below for categorizing studies:

- We categorized a presentation as a “PGI study” if the title or the abstract contains at least one of the following keywords: ‘PGS’, ‘PRS’, ‘PGRS’, ‘polygenic score’, ‘polygenic risk score’, ‘genetic risk score’, ‘GRS’.
- We categorized a presentation as a “twin, family, or adoption study” if it satisfies at least one of the following conditions:
 - The abstract contains ‘twin’ at least twice.
 - The title contains the word ‘twin’.
 - The title or abstract contain at least one of the following keywords: ‘twin registry’, ‘center for twin research’, ‘twin project’, ‘twin panel’, ‘twin study at the’, ‘twin study (LTS)’, ‘(RFAB) twin study’, ‘twin register’, ‘twin pairs’, ‘nonidentical twins’, ‘identical twins’, ‘pairs of twins’, ‘twin sample’, ‘MZ’, ‘DZ’, ‘monozygotic’, ‘dizygotic’, ‘pairs of twins’, ‘adopted’, ‘adoptee’, ‘adoptive’, ‘adoption design’, ‘biological parent’, ‘adoptive parent’, ‘adoption-sibling’, ‘genetically-unrelated’, ‘genetically-related’, ‘siblings reared together’, ‘siblings reared apart’, ‘mother and child’, ‘father and child’, ‘parent and child’, ‘intergenerational’, ‘transracial’, ‘biometric’, ‘path analy’, ‘Cholesky’, ‘children-of-twins’, ‘children of twins’, ‘common environment’, ‘unique environment’, ‘ACE’, ‘ACDE’.
- We categorized a presentation as a “candidate-gene study” if it satisfies at least one of the following conditions:
 - The title contains ‘candidate gene’ or at least one of the following candidate gene keywords: ‘HTR2’, ‘MAOA’, ‘5-HTT’, ‘5HTT’, ‘DRD’, ‘SLC6’, ‘BDNF’, ‘COMT’, ‘TPH’, ‘MTHFR’, ‘APOE’, ‘DTNBP1’, ‘DBH’, ‘ABCB1’, ‘VNTR’, ‘CRHR’, ‘AKT’, ‘NRG’, ‘AVP’, ‘rs0’, ‘rs1’, ‘rs2’, ‘rs3’, ‘rs4’, ‘rs5’, ‘rs6’, ‘rs7’, ‘rs8’, ‘rs9’.
 - The abstract contains at least one of the above candidate-gene keywords.
 - The abstract contains ‘candidate’ at least twice and ‘candidate gene’ at least once.

However, a presentation was removed from the candidate-gene study category if the abstract contains GWAS keywords: ‘wide association analysis’, ‘wide association study’, ‘GWAS’.

To quantify how accurately the algorithmic classifications predict categorizations based on human evaluations, we asked two researchers with expertise in behaviour genetics to categorize 65 randomly sampled presentations. The raters worked independently, without any external assistance, and based their categorizations solely on information supplied about the title and abstract. Each rater assigned three yes/no labels—representing candidate-gene study; twin, family or adoption study; or PGI study—to each presentation. Raters sought to make labelling decisions consistent with the labels’ typical usage in the literature. We defined “agreement” on a presentation as an identical judgment about each of the three labels (i.e., if the raters disagreed about any of the three categories, they were considered as not agreeing). Even under this strict definition, we found an interrater agreement of 94%. The agreement between the algorithm’s and one rater’s categorizations was 86%, and that between the algorithm’s and the other rater’s categorizations was 83%.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgements

We thank C. Shulman for helpful comments. This research was carried out under the auspices of the Social Science Genetic Association Consortium (SSGAC). This research was conducted using the UK Biobank Resource under application number 11425. J.B. was supported by the Pershing Square Fund of the Foundations of Human Behavior, awarded to D.L.; H.J., M.B., D.C., and P.T. by the Ragnar Söderberg Foundation (E42/15), to D.C.; C.A.P.B., P.K., and A.O. by an ERC Consolidator Grant (647648 EdGe), to P.K.; H.J., M.B., A.Y., J.P.B., M.N.M., D.C., D.J.B., and P.T. by Open Philanthropy (010623-00001), to D.J.B.; C.A.P.B., R.A., and S.O. by Riksbankens Jubileumsfond (P18-0782:1), to S.O.; C.A.P.B. and S.O. by the Swedish Research Council (2019-00244), to S.O.; G.G., N.W., and D.J.B. by the NIA/NIH (R24-AG065184 and R01-AG042568), to D.J.B.; D.J.B. by the NIA/NIH (R56-AG058726), to T. Galama; T.T.M., K.P.H., and E.M.T.D. by the NIH/NICHD R01-HD083613 to E.M.T.D. and R01-HD092548 to K.P.H.; P.T. by the NIA/NIMH (R01-MH101244-02 and U01-MH109539-02), to B. Neale. The study was also supported by the NIA/NIH (K99-AG062787-01, P.T.); Netherlands Organisation for Scientific Research VENI (016.Veni.198.058, A.O.); the Swedish Research Council (421-2013-1061, M.J.); the Government of Canada through Genome Canada and the Ontario Genomics Institute (OGI-152, J.P.B.); the Social Sciences and Humanities Research Council of Canada (J.P.B.); the European Union (MP1G118418R, T.E.); the Estonian Research Council (PRG1291, T.E.); the National Health and Medical Research Council (GNT113400, P.M.V.); and the Australian Research Council (P.M.V.). We thank the following consortia for sharing GWAS summary statistics: Reproductive Genetics (ReproGen) Consortium for age at first menses; Genetics of Personality Consortium (GPC) for neuroticism, extraversion, and openness; Psychiatric Genomics Consortium (PGC) for ADHD and depressive symptoms; Tobacco and Alcohol Genetics (TAG) Consortium for cigarettes per day and ever smoker; International Genomics of Alzheimer’s Project (IGAP) for Alzheimer’s disease, GWAS & Sequencing Consortium of Alcohol and Nicotine use (GSCAN) for cigarettes per day, ever smoker and drinks per week; Genetic Investigation of Anthropometric Traits (GIANT) Consortium for height and BMI; and Cognitive Genomics (COGENT) Consortium for cognitive performance. We thank the Neale Lab for making UK Biobank GWAS results available for asthma, cannabis use, COPD, hayfever, life satisfaction (family, finance, friend, work), loneliness, migraine, nearsightedness, number ever born (men, women), religious attendance, self-rated health and subjective well-being. We thank the research participants and employees of 23andMe for making this work possible. A full list of acknowledgements is provided in the Supplementary Note.

Data availability

For how to access the Repository PGIs and other data from each participating dataset, see Supplementary Note; an up-to-date list of participating datasets and data access procedures is maintained at <https://www.thessgac.org/pgi-repository>. For each phenotype

that we analyse, we report GWAS and MTAG summary statistics and PGI (LDpred) weights for all SNPs from the largest discovery sample for that analysis, unless the sample includes 23andMe. SNP-level summary statistics from analyses based entirely or in part on 23andMe data can only be reported for up to 10,000 SNPs. Therefore, if the largest GWAS or MTAG analysis for a phenotype includes 23andMe, we report summary statistics for only the genome-wide significant SNPs from that analysis. In addition, we report summary statistics for all SNPs from a version of the largest GWAS analysis that excludes 23andMe. Finally, we also report summary statistics and PGI (LDpred) weights that the “public PGIs” are based on. These summary statistics and PGI weights can be downloaded from <https://www.thessgac.org/pgi-repository>. The data underlying Figure 1 is also available at <https://www.thessgac.org/pgi-repository>. Researchers at non-profit institutions can obtain access to the genome-wide summary statistics from 23andMe used in this paper by completing the 23andMe Publication Dataset Access Request Form, available at <https://research.23andme.com/dataset-access/>.

References

1. Visscher PM et al. 10 Years of GWAS Discovery: Biology, Function, and Translation. *Am. J. Hum. Genet* 101, 5–22 (2017). [PubMed: 28686856]
2. Wray NR, Goddard ME & Visscher PM Prediction of individual genetic risk to disease from genome-wide association studies. *Genome Res.* 17, 1520–1528 (2007). [PubMed: 17785532]
3. Purcell SM et al. Common polygenic variation contributes to risk of schizophrenia and bipolar disorder. *Nature* 460, 748–752 (2009). [PubMed: 19571811]
4. Rietveld CA et al. GWAS of 126,559 individuals identifies genetic variants associated with educational attainment. *Science* (80-.). 340, 1467–1471 (2013).
5. Lee JJ et al. Gene discovery and polygenic prediction from a genome-wide association study of educational attainment in 1.1 million individuals. *Nat. Genet* 50, 1112–1121 (2018). [PubMed: 30038396]
6. Cesarini D & Visscher PM Genetics and educational attainment. *npj Sci. Learn* 2, 4 (2017). [PubMed: 30631451]
7. Wray NR, Kemper KE, Hayes BJ, Goddard ME & Visscher PM Complex trait prediction from genome data: Contrasting EBV in livestock to PRS in humans. *Genetics* 211, 1131–1141 (2019). [PubMed: 30967442]
8. Green ED & Guyer MS Charting a course for genomic medicine from base pairs to bedside. *Nature* 470, 204–213 (2011). [PubMed: 21307933]
9. Wray NR et al. Pitfalls of predicting complex traits from SNPs. *Nat. Rev. Genet* 14, 507–515 (2013). [PubMed: 23774735]
10. Khera AV et al. Genome-wide polygenic scores for common diseases identify individuals with risk equivalent to monogenic mutations. *Nat. Genet* 50, 1219–1224 (2018). [PubMed: 30104762]
11. Belsky DW & Harden KP Phenotypic Annotation: Using Polygenic Scores to Translate Discoveries From Genome-Wide Association Studies From the Top Down. *Curr. Dir. Psychol. Sci* 28, 82–90 (2019).
12. Benjamin DJ et al. The Promises and Pitfalls of Genoecomics. *Annu. Rev. Econom* 4, 627–662 (2012). [PubMed: 23482589]
13. Freese J The Arrival of Social Science Genomics. *Contemp. Sociol. A J. Rev* 47, 524–536 (2018).
14. Belsky DW et al. The Genetics of Success: How Single-Nucleotide Polymorphisms Associated With Educational Attainment Relate to Life-Course Development. *Psychol. Sci* 27, 957–972 (2016). [PubMed: 27251486]
15. Harden KP et al. Genetic associations with mathematics tracking and persistence in secondary school. *npj Sci. Learn* 5, 1 (2020). [PubMed: 32047651]

16. Robinson MR et al. Genetic evidence of assortative mating in humans. *Nat. Hum. Behav* (2017). doi:10.1038/s41562-016-0016
17. Yengo L et al. Imprint of Assortative Mating on the Human Genome. *Nat. Hum. Behav* 2, 2, 948–954 (2018). [PubMed: 30988446]
18. Abdellaoui A et al. Genetic correlates of social stratification in Great Britain. *Nat. Hum. Behav* 3, 1332–1342 (2019). [PubMed: 31636407]
19. Domingue BW, Rehkopf DH, Conley D & Boardman JD Geographic Clustering of Polygenic Scores at Different Stages of the Life Course. *RSF Russell Sage Found. J. Soc. Sci* 4, 137 LP–149 (2018).
20. Papageorge NW & Thom K Genes, Education, and Labor Market Outcomes: Evidence from the Health and Retirement Study. *J. Eur. Econ. Assoc* 18, 1351–1399 (2020). [PubMed: 32587483]
21. Rietveld CA et al. Replicability and Robustness of Genome-Wide-Association Studies for Behavioral Traits. *Psychol. Sci* 25, 1975–1986 (2014). [PubMed: 25287667]
22. Hewitt JK Editorial policy on candidate gene association and candidate gene-by-environment interaction studies of complex traits. *Behav. Genet* 42, 1–2 (2012). [PubMed: 21928046]
23. Duncan L & Keller M A critical review of the first 10 years of candidate gene-by-environment interaction research in psychiatry. *Am. J. Psychiatry* 168, 1041 (2011). [PubMed: 21890791]
24. Beauchamp JP Genetic evidence for natural selection in humans in the contemporary United States. *Proc. Natl. Acad. Sci. U. S. A* 113, 7774–9 (2016). [PubMed: 27402742]
25. Kong A et al. Selection against variants in the genome associated with educational attainment. *Proc. Natl. Acad. Sci* 114, E727–E732 (2017). [PubMed: 28096410]
26. Tucker-Drob EM Measurement Error Correction of Genome-Wide Polygenic Scores in Prediction Samples. *bioRxiv* 165472 (2017). doi:10.1101/165472
27. DiPrete TA, Burik CAP & Koellinger PD Genetic instrumental variable regression: Explaining socioeconomic and health outcomes in nonexperimental data. *Proc. Natl. Acad. Sci* 115, E4970 LP–E4979 (2018). [PubMed: 29686100]
28. Lambert SA et al. The Polygenic Score Catalog: an open database for reproducibility and systematic evaluation. *medRxiv* 2020.05.20.20108217 (2020). doi:10.1101/2020.05.20.20108217
29. Yang J et al. Common SNPs explain a large proportion of the heritability for human height. *Nat. Genet* 42, 565–569 (2010). [PubMed: 20562875]
30. Vilhjálmsson BJ et al. Modeling Linkage Disequilibrium Increases Accuracy of Polygenic Risk Scores. *Am. J. Hum. Genet* 97, 576–592 (2015). [PubMed: 26430803]
31. Ge T, Chen C-Y, Ni Y, Feng Y-CA & Smoller JW Polygenic prediction via Bayesian regression and continuous shrinkage priors. *Nat. Commun* 10, 1776 (2019). [PubMed: 30992449]
32. Spearman C The Proof and Measurement of Association between Two Things. *Am. J. Psychol* 15, 72–101 (1904).
33. Yang J, Lee SH, Goddard ME & Visscher PM GCTA: A tool for genome-wide complex trait analysis. *Am. J. Hum. Genet* 88, 76–82 (2011). [PubMed: 21167468]
34. Loh PR et al. Contrasting genetic architectures of schizophrenia and other complex diseases using fast variance-components analysis. *Nat. Genet* 47, 1385–1392 (2015). [PubMed: 26523775]
35. Rosner B, Spiegelman D & Willet WC Correction of Logistic Regression Relative Risk Estimates and Confidence Intervals for Random Within-Person Measurement Error. *Am. J. Epidemiol* 136, 1400–1403 (1992). [PubMed: 1488967]
36. Hughes M Regression dilution in the proportional hazards model. *Biometrics* 49, 1056–1066 (1993). [PubMed: 8117900]
37. Okbay A et al. Genome-wide association study identifies 74 loci associated with educational attainment. *Nature* 533, 539–542 (2016). [PubMed: 27225129]
38. Stergiakouli E et al. Association between polygenic risk scores for attention-deficit hyperactivity disorder and educational and cognitive outcomes in the general population. *Int. J. Epidemiol* 46, dyw216 (2016).
39. Elliott ML et al. A Polygenic Score for Higher Educational Attainment is Associated with Larger Brains. *Cereb. Cortex* 29, 3496–3504 (2018).

40. Martin AR et al. Human Demographic History Impacts Genetic Risk Prediction across Diverse Populations. *Am. J. Hum. Genet* 100, 635–649 (2017). [PubMed: 28366442]
41. Loh P-R et al. Efficient Bayesian mixed-model analysis increases association power in large cohorts. *Nat. Genet* 47, 284–290 (2015). [PubMed: 25642633]
42. Day FR et al. Shared genetic aetiology of puberty timing between sexes and with health-related outcomes. *Nat. Commun* (2015). doi:10.1038/ncomms9842
43. Lo M-T et al. Genome-wide analyses for personality traits identify six genomic loci and show correlations with psychiatric disorders. *Nat. Genet* 49, 152–156 (2016). [PubMed: 27918536]
44. Sanchez-Roige S et al. Genome-wide association study of alcohol use disorder identification test (AUDIT) scores in 20 328 research participants of European ancestry. *Addict. Biol* 24, 121–131 (2019). [PubMed: 29058377]
45. Sanchez-Roige S et al. Genome-wide association study of delay discounting in 23,217 adult research participants of European ancestry. *Nat. Neurosci* 21, 16–20 (2018). [PubMed: 29230059]
46. Warrier V et al. Genome-wide analyses of self-reported empathy: Correlations with autism, schizophrenia, and anorexia nervosa. *Transl. Psychiatry* 8, 1–10 (2018). [PubMed: 29317594]
47. Hu Y et al. GWAS of 89,283 individuals identifies genetic variants associated with self-reporting of being a morning person. *Nat. Commun* 7, 1–9 (2016).
48. Hinds DA et al. A genome-wide association meta-analysis of self-reported allergy identifies shared and allergy-specific susceptibility loci. *Nat. Genet* (2013). doi:10.1038/ng.2686
49. Ferreira MA et al. Shared genetic origin of asthma, hay fever and eczema elucidates allergic disease biology. *Nat. Genet* 49, 1752–1757 (2017). [PubMed: 29083406]
50. Demontis D et al. Discovery of the first genome-wide significant risk loci for attention deficit/hyperactivity disorder. *Nat. Genet* 51, 63–75 (2019). [PubMed: 30478444]
51. Pasman JA et al. GWAS of lifetime cannabis use reveals new risk loci, genetic overlap with psychiatric traits, and a causal influence of schizophrenia. *Nat. Neurosci* (2018). doi:10.1038/s41593-018-0206-1
52. Liu M et al. Association studies of up to 1.2 million individuals yield new insights into the genetic etiology of tobacco and alcohol use. *Nature Genetics* 51, 237–244 (2019). [PubMed: 30643251]
53. Hyde CL et al. Identification of 15 genetic loci associated with risk of major depression in individuals of European descent. *Nat. Genet* 48, 1031–6 (2016). [PubMed: 27479909]
54. Pickrell JK et al. Detection and interpretation of shared genetic influences on 42 human traits. *Nat. Genet* 48, 709–717 (2016). [PubMed: 27182965]
55. Karlsson Linnér R et al. Genome-wide association analyses of risk tolerance and risky behaviors in over 1 million individuals identify hundreds of loci and shared genetic influences. *Nat. Genet* 51, 245–257 (2019). [PubMed: 30643258]
56. Winkler TW et al. Quality control and conduct of genome-wide association meta-analyses. *Nat. Protoc* 9, 1192–1212 (2014). [PubMed: 24762786]
57. McCarthy S et al. A reference panel of 64,976 haplotypes for genotype imputation. *Nat. Genet* 48, 1279–1283 (2016). [PubMed: 27548312]
58. Abecasis GR et al. A map of human genome variation from population-scale sequencing. *Nature* 467, 1061–1073 (2010). [PubMed: 20981092]
59. Bulik-Sullivan BK et al. An atlas of genetic correlations across human diseases and traits. *Nat. Genet* 47, 1236–1241 (2015). [PubMed: 26414676]
60. Perry JRB et al. Parent-of-origin-specific allelic associations among 106 genomic loci for age at menarche. *Nature* 514, 92–97 (2014). [PubMed: 25231870]
61. Stringer S et al. Genome-wide association study of lifetime cannabis use based on a large meta-analytic sample of 32 330 subjects from the International Cannabis Consortium. *Transl. Psychiatry* 6, e769 (2016). [PubMed: 27023175]
62. Furberg H et al. Genome-wide meta-analyses identify multiple loci associated with smoking behavior. *Nat. Genet* 42, 441–447 (2010). [PubMed: 20418890]
63. Wray NR et al. Genome-wide association analyses identify 44 risk variants and refine the genetic architecture of major depression. *Nat. Genet* 50, 668–681 (2018). [PubMed: 29700475]

64. Doherty A et al. GWAS identifies 14 loci for device-measured physical activity and sleep duration. *Nat. Commun* 9, 1–8 (2018). [PubMed: 29317637]
65. van den Berg SM et al. Meta-analysis of Genome-Wide Association Studies for Extraversion: Findings from the Genetics of Personality Consortium. *Behav. Genet* 46, 170–182 (2016). [PubMed: 26362575]
66. de Moor MHM et al. Meta-analysis of genome-wide association studies for neuroticism, and the polygenic association with Major Depressive Disorder. *JAMA Psychiatry* 72, 642–650 (2015). [PubMed: 25993607]
67. de Moor MHM et al. Meta-analysis of genome-wide association studies for personality. *Mol. Psychiatry* 17, 337–349 (2012). [PubMed: 21173776]
68. Okbay A et al. Genetic variants associated with subjective well-being, depressive symptoms, and neuroticism identified through genome-wide analyses. *Nat. Genet* 48, 624–633 (2016). [PubMed: 27089181]
69. Locke AEA et al. Genetic studies of body mass index yield new insights for obesity biology. *Nature* 518, 197–206 (2015). [PubMed: 25673413]
70. Wood AR et al. Defining the role of common variation in the genomic and biological architecture of adult human height. *Nat. Genet* 46, 1173–1186 (2014). [PubMed: 25282103]
71. Kunkle BW et al. Genetic meta-analysis of diagnosed Alzheimer’s disease identifies new risk loci and implicates A β , tau, immunity and lipid processing. *Nat. Genet* 51, 414–430 (2019). [PubMed: 30820047]
72. Trampush JW et al. GWAS meta-analysis reveals novel loci and genetic correlates for general cognitive function: a report from the COGENT consortium. *Mol. Psychiatry* 22, 336–345 (2017). [PubMed: 28093568]
73. Barban N et al. Genome-wide analysis identifies 12 loci influencing human reproductive behavior. *Nat. Genet* 48, 1462–1472 (2016). [PubMed: 27798627]
74. Turley P et al. Multi-trait analysis of genome-wide association summary statistics using MTAG. *Nat. Genet* 50, 229–237 (2018). [PubMed: 29292387]
75. Daetwyler HD, Villanueva B & Woolliams JA Accuracy of Predicting the Genetic Risk of Disease Using a Genome-Wide Approach. *PLoS One* 3, e3395 (2008). [PubMed: 18852893]
76. Auton A et al. A global reference for human genetic variation. *Nature* 526, 68–74 (2015). [PubMed: 26432245]
77. Das S et al. Next-generation genotype imputation service and methods. *Nat. Genet* 48, 1284–1287 (2016). [PubMed: 27571263]
78. The International HapMap 3 Consortium et al. Integrating common and rare genetic variation in diverse human populations. *Nature* 467, 52–58 (2010). [PubMed: 20811451]
79. Chang CC et al. Second-generation PLINK: rising to the challenge of larger and richer datasets. *Gigascience* 4, 1–16 (2015). [PubMed: 25838885]
80. Yengo L et al. Meta-analysis of genome-wide association studies for height and body mass index in ~700000 individuals of European ancestry. *Hum. Mol. Genet* 27, 3641–3649 (2018). [PubMed: 30124842]
81. Savage JE et al. Genome-wide association meta-analysis in 269,867 individuals identifies new genetic and functional links to intelligence. *Nat. Genet* 50, 912–919 (2018). [PubMed: 29942086]
82. Howard DM et al. Genome-wide meta-analysis of depression identifies 102 independent variants and highlights the importance of the prefrontal brain regions. *Nat. Neurosci* 22, 343–352 (2019). [PubMed: 30718901]
83. Jones SE et al. Genome-wide association analyses of chronotype in 697,828 individuals provides insights into circadian rhythms. *Nat. Commun* 10, 1–11 (2019). [PubMed: 30602773]
84. Nagel M et al. Meta-analysis of genome-wide association studies for neuroticism in 449,484 individuals identifies novel genetic loci and pathways. *Nat. Genet* 50, 920–927 (2018). [PubMed: 29942085]

Box.**Note on Terminology**

In this paper, we use the term “polygenic index” instead of the commonly used terms “polygenic score” and “polygenic risk score.” Most of us prefer the term polygenic index because we are persuaded by the argument that it is less likely to give the impression of a value judgment where one is not intended. The term polygenic index was first proposed by Martha Minow at a meeting of the Trustees of the Russell Sage Foundation.

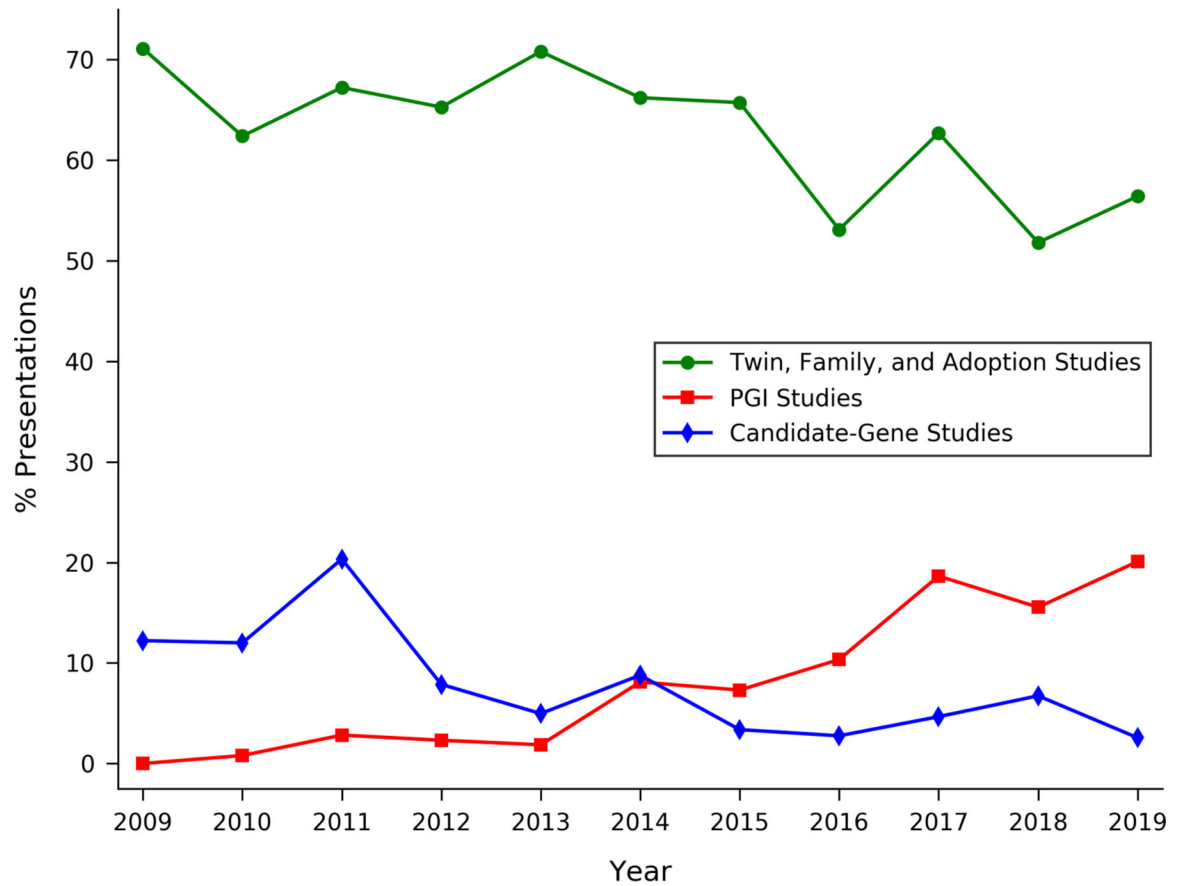


Figure 1. Type of study in presentations at Behavior Genetics Association Annual Meetings.

Notes: For a description of the data underlying this figure, see Methods. Out of 1,993 presentations in total (over the 2009–2019 period), the percentages that are in exactly 0, 1, 2, or 3 categories are 26.6%, 67.6%, 5.5%, and 0.2%, respectively.

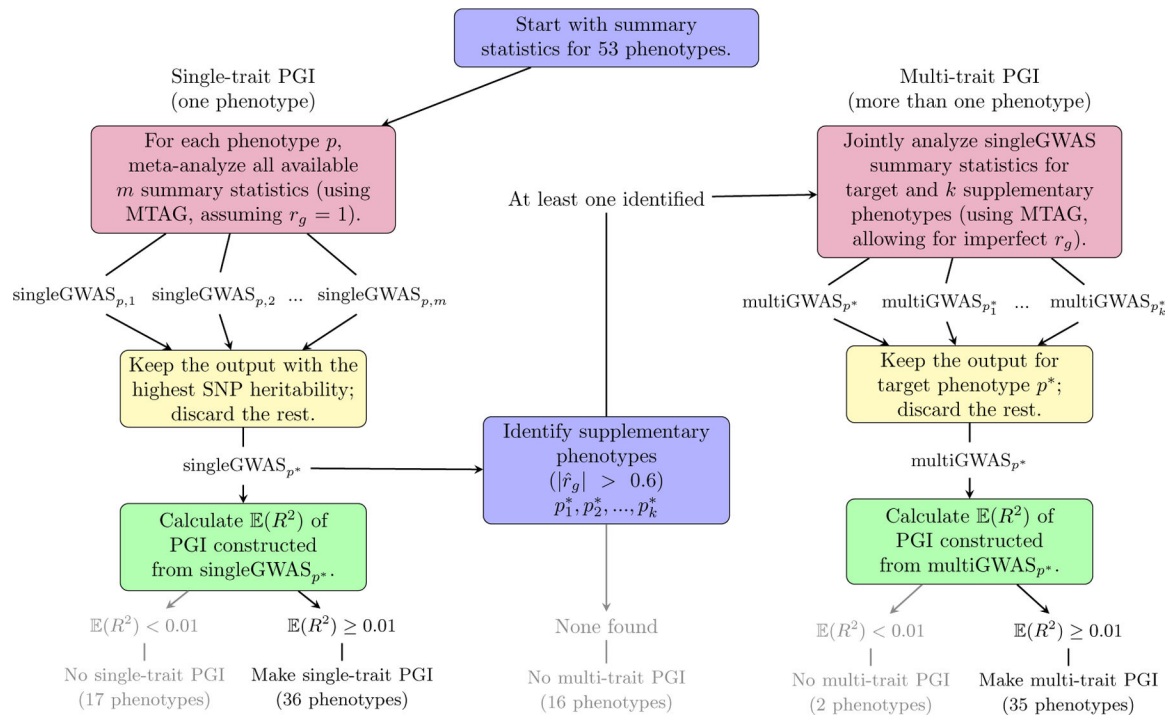


Figure 2. Algorithm determining which single-trait and multi-trait PGIs were generated for the Repository.

Notes: See Table 1 for the 36 single-trait PGIs and 35 multi-trait PGIs included in the Repository.

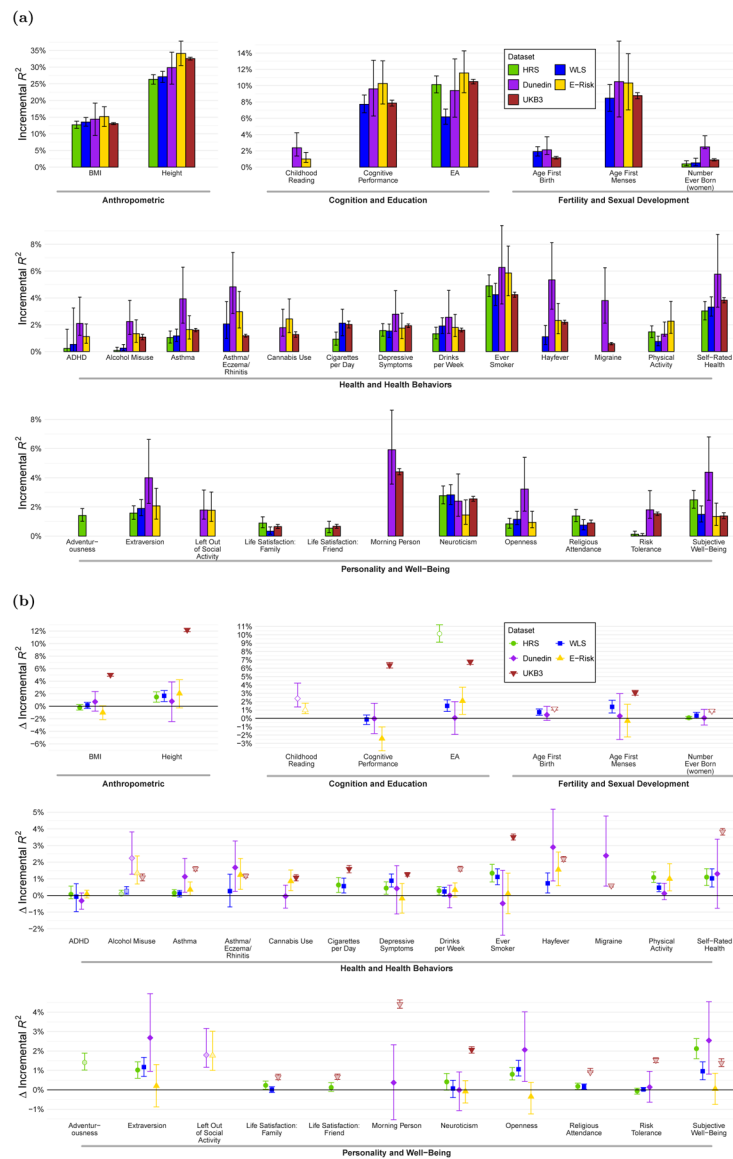


Figure 3. Predictive power of Repository single-trait PGIs.

Notes: Error bars show 95% confidence intervals from bootstrapping with 1,000 repetitions.

Panel (A): Incremental R^2 from adding Repository’s single-trait PGI to a regression of the phenotype on 10 principal components of the genetic relatedness matrix for HRS, WLS, Dunedin and E-Risk, and on 20 principal components and 106 genotyping batch dummies for UKB. Prior to the regression, phenotypes are residualized on a second-degree polynomial for age or birth year, sex, and their interactions (see Supplementary Tables 5 and 12). For the sample sizes of the GWAS that the PGIs are based on, see Supplementary Table 8. Panel (B): Difference in incremental R^2 between Repository single-trait PGI and PGI constructed from publicly available summary statistics using our Repository pipeline. (Note that the latter do not include PGI directly available from datasets, such as the ones accessible from the HRS website.) If no publicly available summary statistics are available for a phenotype, then the difference in incremental R^2 is equal to the incremental R^2 of

the single-trait PGI and is represented by an open circle. “Cigarettes per Day” in Dunedin was omitted from the Figure because the confidence interval (−5.99% to 0.94%) around the point estimate (−2.38%) required extending the y-axis substantially, making the figure hard to read. For the GWAS sample sizes of the PGIs based on publicly available summary statistics, see Supplementary Table 13.

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Table 1.

Repository phenotypes and GWAS sample sizes

Phenotype	GWAS Sample Size				PGIs Released			Suppl. Phenotypes
	Total	23andMe	UKB	Other	Public N	Single	Multi	
Anthropometric								
1 Body Mass Index (BMI)	760,630	-	438,476	322,154	795,640	X		
2 Height	698,334	-	445,054	253,280	709,706	X		
Cognition and Education								
3 Childhood Reading	172,503	172,503	-	-	-	X		
4 Cognitive Performance	260,354	-	225,056	35,298	269,867	X	X	5, 6, 7
5 Educational Attainment	1,047,538	365,536	-	682,002	766,345	X	X	4, 6, 8, 33, 45
6 Highest Math	430,439	430,439	-	-	-	X	X	4, 5, 7, 8, 33
7 Self-Rated Math Ability	564,692	564,692	-	-	-	X	X	4, 6
Fertility and Sexual Development								
8 Age First Birth	407,884	9,370	156,733*	241,781	241,781	X	X	5, 6, 11, 12, 19, 22
9 Age First Menses (Women)	329,345	76,831	-	252,514	252,514	X	X	10
10 Age Voice Deepened (Men)	55,871	55,871	-	-	-		X	9
11 Number Ever Born (Men)	260,991	-	168,056*	92,935	165,492		X	8, 12
12 Number Ever Born (Women)	399,803	-	188,208*	211,595	211,595	X	X	8, 11
Health and Health Behaviors								
13 Alcohol Misuse	151,067	19,407	131,660	-	-	X	X	24
14 Allergy - Cat	46,646	46,646	-	-	-		X	15, 16, 17, 18, 26
15 Allergy - Dust	46,646	46,646	-	-	-		X	14, 16, 17, 18, 26
16 Allergy - Pollen	46,646	46,646	-	-	-		X	14, 15, 17, 19, 26
17 Asthma	445,965	-	445,965	-	361,141	X	X	14, 15, 16, 18, 26
18 Asthma/Eczema/Rhinitis	685,716	135,538	307,609*	242,569	242,569	X	X	14, 15, 16, 17, 26
19 Attention Deficit Hyperactivity Disorder (ADHD)	117,754	62,380	-	55,374	55,374	X	X	8, 22
20 Cannabis Use	202,180	22,771	144,112	35,297	117,911	X		
21 Cigarettes per Day	340,140	76,186	-	263,954	263,954	X		
22 Chronic Obstructive Pulmonary Disease (COPD)	445,965	-	445,965	-	91,787		X	8, 19, 30
23 Depressive Symptoms	942,579	307,354	404,984	230,241	500,199	X	X	30, 40, 43, 47

Phenotype	GWAS Sample Size				PGIs Released		Suppl. Phenotypes	
	Total	23andMe	UKB	Other	Public N	Single	Multi	
24 Drinks per Week	941,287	403,938	-	537,349	537,349	X	X	13
25 Ever Smoker	1,255,948	623,146	-	632,802	632,802	X		
26 Hayfever	445,963		445,963	-	360,527	X	X	14, 15, 16, 17, 18, Eczema [†]
27 Migraine	693,993	283,985	410,008	-	361,194	X		
28 Nearsightedness	367,906	191,843	176,063	-	360,677	X		
29 Physical Activity	357,039	265,934	-	91,105	91,105	X		
30 Self-Rated Health	1,203,099	758,713	444,386	-	359,681	X	X	22, 23, 37
Personality and Well-Being								
31 Adventurousness	557,923	557,923	-	-	-	X	X	46
32 Cognitive Empathy	46,861	46,861	-	-	-	X	X	Agreeableness [†]
33 Delay Discounting	23,217	23,217	-	-	-	X	X	5, 6
34 Extraversion	122,255	59,225	-	63,030	63,030	X	X	35
35 Left Out of Social Activity	507,804	507,804	-	-	-	X	X	34, 38, 40, 47
36 Life Satisfaction: Family	168,313	-	168,313	-	118,818	X	X	38, 39, 47
37 Life Satisfaction: Finance	169,051	-	169,051	-	119,394		X	30, 40, 47
38 Life Satisfaction: Friends	168,001	-	168,001	-	118,649	X	X	35, 36, 39, 47
39 Life Satisfaction: Work	115,038	-	115,038	-	82,190		X	36, 38, 47
40 Loneliness	439,525	-	439,525	-	355,583		X	23, 35, 37, 43, 47
41 Morning Person	493,043	91,967	401,076	-	449,734	X		
42 Narcissism	452,535	452,535	-	-	-	X		
43 Neuroticism	484,560	59,206	361,688	63,666	380,060	X	X	23, 40, 47
44 Openness	76,551	59,176	-	17,375	17,375	X		
45 Religious Attendance	444,842	-	444,842	-	360,063	X	X	5
46 Risk	1,427,867	969,309	-	458,558	466,571	X	X	31
47 Subjective Well-Being	1,022,510	728,752	169,219	124,539	204,978	X	X	23, 35, 36, 37, 38, 39, 40, 43

Notes:

* For Age First Birth, Number Ever Born (Men), Number Ever Born (Women) and Asthma/Eczema/Rhinitis, the publicly available summary statistics include the first release of UKB. Therefore, there is sample overlap between our UKB GWAS and publicly available summary statistics. For these phenotypes, in the UKB column, we report the UKB sample size excluding samples from the publicly available GWAS.

For Eczema and Agreeableness, both the single- and multi-trait PGI_s had an expected predictive power less than 0.01, so they were used only as supplementary phenotypes for other phenotypes. Therefore, they are not included in the table and are not represented by a number. The GWAS sample for Eczema consists of only UKB, with $N = 440,177$. The GWAS sample for Agreeableness consists only of 23andMe, with $N = 59,176$.

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Table 2.

Datasets participating in the Repository

Dataset	<i>N</i>	Country	Population- or Family-based
Dunedin Multidisciplinary Health and Development Study	887	New Zealand	Population
English Longitudinal Study of Ageing (ELSA)	7,310	UK	Population
Environmental Risk (E-Risk) Longitudinal Twin Study	2,316	UK	Family
Estonian Genome Center, University of Tartu (EGCUT)	51,719	Estonia	Population
Health and Retirement Study (HRS)	12,090	USA	Population
Minnesota Center for Twin and Family Research (MCTFR)	7,654	USA	Family
National Longitudinal Study of Adolescent to Adult Health (Add Health)	5,689	USA	Family
Swedish Twin Registry (STR)	38,072	Sweden	Family
Texas Twin Project	556	USA	Family
UK Biobank (UKB)	445,985	UK	Population
Wisconsin Longitudinal Study (WLS)	8,949	USA	Family

Notes: The “*N*” column gives the number of participants in each dataset for whom the PGIs in Table 1 are supplied in the initial release of the Repository (i.e., those who passed the subject-level exclusion filters described in Methods). “Population- or Family-based” refers to how individuals were recruited to the dataset.

Table 3.

Application of measurement-error correction

Panel A. Association Between EA and the PGI, Without and With Controls for Parental EA

	Original		Corrected	
	(1)	(2)	(3)	(4)
EA PGI	0.844 (0.026)	0.619 (0.024)	1.318 (0.041)	1.104 (0.042)
Father's EA	-	0.154 (0.010)	-	0.112 (0.010)
Mother's EA	-	0.176 (0.011)	-	0.141 (0.012)
# Obs.	8,537	8,537	8,537	8,537

Panel B. Interaction Between PGI and Family SES Predicting High School and College Completion

	(1)	(2)	(3)	(4)
	High school	College	High school	College
EA PGI	0.095 (0.008)	0.055 (0.008)	0.166 (0.014)	0.103 (0.014)
Family SES	0.069 (0.009)	0.031 (0.010)	0.063 (0.009)	0.034 (0.010)
EA PGI X Family SES	-0.047 (0.009)	0.068 (0.010)	-0.084 (0.015)	0.101 (0.016)
# Obs.	8,387	8,387	8,387	8,387

Notes: Each column reports estimated regression coefficients, with standard errors in parentheses. Panel A: Columns (1) and (2) replicate results from Papageorge and Thom's Table 2 columns 1 and 2. Panel B: Columns (1) and (2) replicate results from Papageorge and Thom's Table B.2 panel B columns 2 and 4. Panels A and B: Columns (3) and (4) apply our measurement-error-corrected estimator to the feasible-regression results in Columns (1) and (2). A value of $\rho = 1.52$ was used in the correction. All regressions include indicators for birth year, sex, interactions of birth year and sex, and 10 principal components of the genetic data (coefficients not reported). The regressions in Panel B also control for mother and father's educational attainment and an indicator for whether these values are missing (these data are missing for 2000 individuals). Our panel B regressions differ from Papageorge and Thom as we do not include a cubic of the PGI as control variables. We omitted the cubic terms because our measurement-error-corrected estimator does not account for non-linear transformations of the PGI.