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Medial temporal lobe subregional morphometry using high resolution MRI in Alzheimer's Disease

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Abstract

Autopsy studies of Alzheimer's Disease (AD) have found that neurofibrillary tangle (NFT) pathology of the medial temporal lobe (MTL) demonstrates selective topography with relatively stereotyped subregional involvement at early disease stages, prompting interest in more granular measurement of these structures with in vivo MRI. We applied a novel, automated method for measurement of hippocampal subfields and extrahippocampal MTL cortical regions. The cohort included cognitively normal (CN) adults (n=86), early Mild Cognitive Impairment (EMCI; n=43), late MCI (LMCI; n=22), and mild AD (n=40) patients from the Alzheimer's Disease Neuroimaging Initiative (ADNI). For pseudolongitudinal analysis of the continuum from preclinical to mild AD dementia, the groups were further divided according to amyloid status based on PET. Specific subregions associated with the early NFT pathology of AD were more sensitive to preclinical and early prodromal AD than whole hippocampal volume while more diffuse involvement was found in later stages. In particular, BA35, the first region associated with NFT deposition, was the only region to discriminate preclinical AD from amyloid negative CN adults ("normal aging"). In general, patterns of atrophy in the pseudolongitudinal analysis largely recapitulated Braak staging of NFTs within the MTL.

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Data used in preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). As such, the investigators within the ADNI contributed to the design and implementation of ADNI and/or provided data but did not participate in analysis or writing of this report. A complete listing of ADNI investigators can be found at: http://adni.loni.usc.edu/wp-content/uploads/how_to_apply/ADNI_Acknowledgement_List.pdf

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Introduction

Alzheimer's Disease (AD) is a progressive neurodegenerative condition in which impairments of episodic memory are an early hallmark symptom. Memory loss is likely driven, in part, by the well-described early neurofibrillary tangle (NFT) pathology in the medial temporal lobe (MTL), the central anatomic structure supporting episodic memory (Arnold, Hyman, Flory, Damasio, & Van Hoesen, 1991; Braak & Braak, 1991; Squire, 2004). While measurements of the hippocampus have proven to be one of the most reliable biomarkers of AD, both for prediction of cognitive decline in Mild Cognitive Impairment (MCI) and for tracking disease progression, more granular measurement of MTL substructures may prove to enhance sensitivity at different disease stages (Pluta, Yushkevich, Das, & Wolk, 2012).

Indeed, in early stages, NFT pathology does not uniformly involve the MTL and hippocampus (Arnold et al., 1991; Braak & Braak, 1991). The first region thought to display this pathology is perirhinal cortex (PRC), particularly Brodmann Area 35 (BA35), which Braak and Braak referred to as transentorhinal cortex (Braak & Braak, 1991). This is then followed by involvement of the entorhinal cortex (ERC) and then cornu ammonis 1 (CA1) subfield of the hippocampus proper. As such, more global measures of MTL structure may obscure these more specific regional changes at early stages.

With the goal of developing more sensitive measures of early AD pathology, a number of *in vivo* approaches for measurement of hippocampal subfields and cortical MTL structures have been developed using MRI (Apostolova et al., 2006; Iglesias et al., 2015; Kerchner et al., 2010; S. G. Mueller & Weiner, 2009; Van Leemput et al., 2009; Wisse et al., 2012; Yushkevich et al., 2015; Zeineh, Engel, Thompson, & Bookheimer, 2003). However, high field strength (7T MRI), relatively long acquisition times (~30 min), and labor intensive manual annotation limit practical utility of some of these methods for larger cohorts and clinical populations. Use of a high in-plane resolution (~0.4×0.4×2mm³), but anisotropic, T2-weighted MRI sequence orthogonal to the long axis of the hippocampus, has proven a useful compromise, as these scans can be obtained in approximately 8 minutes at 3T field strength. These scanning parameters allow for visualization of the internal structures of the hippocampus, not visible on a typical research-quality T1-weighted scan (1×1×1mm³).

While a number of manual protocols for segmentation of the above T2-weighted sequence have been published (La Joie et al., 2013; S. G. Mueller, Schuff, Raptentsetsang, Elman, & Weiner, 2008; S. G. Mueller et al., 2007; Shing et al., 2011; Yushkevich et al., 2015), our group developed an automated technique for labeling hippocampal subfields and cortical MTL regions [(Pluta et al., 2012; Yushkevich et al., 2015; Yushkevich et al., 2010); see (Iglesias et al., 2015) for a different automated approach)]. Using this method, measurement of select subregions, including CA1 and BA35, have demonstrated greater sensitivity to MCI status than whole hippocampal volumes (Pluta et al., 2012; Yushkevich et al., 2015).

The current manuscript describes the relative involvement of different MTL subregions across a range of cognitive impairment associated with AD. In addition to assessing the relative sensitivity of these measures in clinically classified groups, we explored whether

regional atrophy patterns correspond to distribution of NFT deposition described in autopsy studies. To do so, we examined differences in regional MTL atrophy across the continuum from cognitively normal (CN) without evidence of AD pathology, or "amyloid negative" based on florbetapir-PET, to preclinical AD (CN adults with evidence of cerebral amyloid, or "amyloid positive"), prodromal AD ("amyloid positive" MCI patients), and early AD dementia. The current contribution represents the most comprehensive analysis of morphometric data measured from an MTL-dedicated sequence obtained in a multi-site manner as part of ADNI.

Materials and Methods

Participants

Data used in the preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). The ADNI was launched in 2003 by the National Institute on Aging (NIA), the National Institute of Biomedical Imaging and Bioengineering (NIBIB), the Food and Drug Administration (FDA), private pharmaceutical companies and non-profit organizations, as a \$60 million, 5-year public-private partnership. The primary goal of ADNI has been to test whether serial magnetic resonance imaging (MRI), positron emission tomography (PET), other biological markers, and clinical and neuropsychological assessment can be combined to measure the progression of mild cognitive impairment (MCI) and early Alzheimer's disease (AD). Determination of sensitive and specific markers of very early AD progression is intended to aid researchers and clinicians to develop new treatments and monitor their effectiveness, as well as lessen the time and cost of clinical trials.

The Principal Investigator of this initiative is Michael W. Weiner, MD, VA Medical Center and University of California – San Francisco. ADNI is the result of efforts of many co-investigators from a broad range of academic institutions and private corporations, and subjects have been recruited from over 50 sites across the U.S. and Canada. The initial goal of ADNI was to recruit 800 subjects but ADNI has been followed by ADNI-GO and ADNI-2. To date these three protocols have recruited over 1500 adults, ages 55 to 90, to participate in the research, consisting of cognitively normal older individuals, people with early or late MCI, and people with early AD. The follow up duration of each group is specified in the protocols for ADNI-1, ADNI-2 and ADNI-GO. Subjects originally recruited for ADNI-1 and ADNI-GO had the option to be followed in ADNI-2. For up-to-date information, see www.adni-info.org.

For this study, subjects were selected from the subset of the ADNI 2 participants in which high-resolution T2-MRI were collected at the 20 sites using scanners developed by Siemens Medical Systems. This acquisition was developed after initiation of ADNI 2 data collection as a sub-project with the goal of comparing the efficacy and added value of MTL sub-regional measurements relative to whole hippocampal and other standard volumetric measurements (Iglesias et al., 2015; S. Mueller et al., 2013). Clinical categorization of study participants into cognitively normal (CN), early MCI (EMCI), late MCI (LMCI), and AD were as described in detail at www.adni-info.org and used standard clinical criteria for MCI and AD (Landau et al., 2012; McKhann, Drachman, Folstein, Katzman, & Price, 1984;

Petersen, 2004). MCI patients were all amnestic, either single-or multi-domain. Division into early and late MCI groups was based solely on education-adjusted scores for the delayed paragraph recall subscore of the Wechsler Memory Scale-Revised Logical Memory II (WMS-LM II). Thus, EMCI patients straddled the boundary between normal memory and LMCI.

In light of the delayed initiation of the high-resolution T2-MRI substudy, many of the included MRI's did not correspond to baseline clinical assessments (n=149 of 191). In these instances, diagnosis and psychometric testing closest to the time of the scan was utilized. As MCI patients were not systematically divided into EMCI and LMCI at these follow-up visits, we used the same WMS-LM II cutoff to dichotomize into these two categories. Further, the control group included subjects designated as significant memory concern, a category added after the initiation of ADNI2 recruitment based on the presence of subjective memory complaints and a Cognitive Change Index score 16 (Saykin et al., 2006), but psychometric testing within the normal range. As we did not observe clear differences in control participants with or without such subjective complaints, these groups were collapsed into the CN group.

Psychometric Testing

The following measures were included in the current analysis: Wechsler Memory Scale-Revised Logical Memory I & II (Wechsler, 1987), Rey Auditory Verbal Learning Test (Rey, 1964), the Trail Making Test [Trails A and Trails B] (Reitan, 1958), category fluency [Animals] (Butters, Granholm, Salmon, Grant, & Wolfe, 1987), and Boston Naming Test (Kaplan, Goodglass, & Weintraub, 1983). AVLT measures include 5 immediate memory trials and 5- and 30-min delayed recall trials and a 30-min delayed recognition measure. Additionally, the Clinical Dementia Rating (CDR) scale (Morris, 1993) was obtained in all participants. Demographic and clinical characteristics of the subject pool is described in Table 1.

MRI Image acquisition

All subjects were scanned with a T1-MRI protocol optimized for best contrast to noise in a feasible acquisition time (Jack et al., 2008; Leow et al., 2006). Raw data had an acquisition matrix of $192 \times 192 \times 166$ and voxel size $1.25 \times 1.25 \times 1.2 \text{ mm}^3$. Zero-filled reconstruction (i.e., sinc interpolation) resulted in a 256×256 matrix and voxel size of $0.9375 \times 0.9375 \times 1.2 \text{ mm}^3$. In addition, T2-MRI with high in-plane resolution in the oblique coronal orientation prescribed to be perpendicular to the long axis of the hippocampal formation was also acquired (Figure 1). The MRI sequence parameters were: TR/TE 8020/50 ms, $0.4 \times 0.4 \times 2.0 \text{ mm}^3$ resolution, min 24 slices, acquisition time: 8.1 min. Further details on ADNI imaging protocols can be found at http://adni.loni.usc.edu/methods/documents/mri-protocols/.

Image processing

Image quality of high-resolution T2-MRI was assessed by a manual rater using the following criteria: 1) Each image was assigned a quality rating between 0 and 5 based on the severity of noise, motion and ringing artifacts. Images with a rating >2 were included in the

study. 2) Images where part of the posterior hippocampal complex was not included because of a field of view that was too far anterior were excluded. For each participant, data from the earliest available MRI session that met the selection criteria above were used. High-resolution T2-MRI was labeled using the Automatic Segmentation of Hippocampus Subfields (ASHS) software package (Yushkevich et al., 2015; Yushkevich et al., 2010). Briefly, this method utilizes a multi-atlas label fusion technique in combination with a learning-based error correction module to produce a fully automated segmentation of hippocampal subfields along the entire length of the hippocampal formation, as well as segmentation of some MTL cortical regions. Candidate segmentations of a subject's MRI are obtained using high-dimensional mapping to multiple manually labeled atlas images. These are then fused into a consensus segmentation, taking into account the degree of similarity between the subject image and atlas images. Patterns of systematic segmentation errors introduced in this procedure are learned a priori using training data, and are corrected in a further post-processing step, to generate the final segmentation.

Cornu Ammonis (CA1/2/3) and dentate gyrus (DG) subfields of the hippocampal formation, subiculum, and entorhinal (ERC), parahippocampal (PHC), and perirhinal (PRC) cortices (BA35 and BA36) were labeled in each subject. The segmentation protocol used for atlas set generation and reliability of automated labeling for these subregions was previously reported (Yushkevich et al., 2015). Further details of the protocol can be found in supplementary material for this prior report: http://onlinelibrary.wiley.com/store/10.1002/hbm.22627/asset/supinfo/hbm22627-sup-0001-suppinfo.docx? v=1&s=7ce9ee4821c571dcbf6ad9981cc9682c41350fac. An example of the automatic segmentation is shown is Figure 1.

Voxelwise cortical thickness maps for the entire brain was generated from T1-MRI using a diffeomorphic registration based thickness estimation method described in (Das, Avants, Grossman, & Gee, 2009), as implemented in the ANTS Cortical Thickness pipeline, evaluated in (Tustison et al., 2014). Briefly, a one-to-one mapping between the white-gray matter interface and the gray matter-cerebrospinal fluid interface is derived and thickness is defined as the distance between corresponding points. T1 and T2 images were registered with FLIRT software (Jenkinson & Smith, 2001), as part of the hippocampal subfield segmentation pipeline (ASHS) [see (Yushkevich et al., 2015)]. We visually verified that there were no registration failures. Thickness maps were then integrated over the cortical labels obtained from T2-MRI to generate average cortical thickness in the ERC and PRC subregions. Whole hippocampal volumes were generated from the anatomical image of each subject using the multi-atlas label fusion technique previously described (Wang & Yushkevich, 2013).

Florbetapir-PET

Florbetapir-PET scanning consisted of four 5-minute dynamic frames, acquired 50 to 70 minutes after injection and is described in detail on-line (adni.loni.ucla.edu/about-data-samples/image-data/). Processed Florbetapir PET data was made publicly available by ADNI and used the following automated processing pipeline (adni.loni.ucla.edu/research/pet-post-processing/). Briefly, after co-registration with T1-weighted MRI, an SUVR was

calculated by taking the mean uptake over voxels in four regions of interest (ROIs), frontal, anterior/posterior cingulate, lateral parietal, and lateral temporal cortices, and then dividing these regions by mean uptake over voxels of the entire cerebellum. A composite measure taking the mean SUVR of the four cortical ROIs was used to determine whether a participant was "amyloid positive" or "amyloid negative." We used a previously established cutoff, 1.11, for this measure (Landau et al., 2012). SUVR values of this composite measure are included in Table 1.

Statistical Analysis

Morphometric measurements from MTL subregions were entered into statistical analysis using IBM SPSS 22.0. For hippocampal subfields, volumes produced by ASHS were used. Because of the uncertainty associated with the lateral boundaries of MTL cortical structures, cortical thickness was used for ERC, PRC, and PHC regions instead of volumes. To reduce the number of comparisons, mean values of right and left hemispheres were calculated for each MTL region. Additionally, given poorer reliability of CA2 and CA3, based on our prior work (Yushkevich et al., 2015), a combined CA label was utilized. Consistent with the fact that CA1 accounts for most of this volume, results were highly similar whether we used CA1 or the entire CA region.

The discriminative ability of the imaging biomarkers derived from high-resolution T2-MRI were examined by comparing volumes or cortical thickness between CN group and each of the three patient groups: EMCI, LMCI and AD. Further comparisons were made across the spectrum of AD by comparing amyloid negative CN adults with amyloid positive CN participants (i.e. preclinical AD), EMCI (i.e. early prodromal AD), LMCI (i.e. late prodromal AD), and AD (i.e. mild AD dementia). All values were adjusted by age and education based on linear regression in amyloid negative CN adults. For volumes, intracranial volume produced by ASHS was also used to adjust values. Direct group comparisons used one-sided t-tests.

Results

Clinical Group Discrimination

Results of the comparisons of symptomatic patients versus CN adults are tabulated in Table 2. Average hippocampal subfield volumes as well as thickness in MTL cortex were progressively smaller in EMCI, LMCI and AD relative to controls. Importantly, we found that subregion-specific measurements were sensitive to the earliest symptomatic disease stage, EMCI, but not whole hippocampal volume. Perirhinal cortex thickness, particularly that of BA36 [t(70.1)=4.0, p<.001)], had a strong group effect when compared to controls. All other extrahippocampal MTL regions were significantly different as well, but with smaller effect sizes [ERC: t(127)=3.0, p<0.01; BA35:t(63.6)=2.4, p<0.05; PHC: t(127)=2.0, p<0.05]. Hippocampal regions were generally more marginally significant [subiculum: t(127)=2.3, p<.05; CA: t(62.6)=1.7, p<0.05; DG: t(61.5)=1.8, p<0.05]. Notably, whole hippocampal volume was not significantly different between these groups [t(59.9)<1, p>0.1)]

In LMCI, while both BA35 [t(106)=5.3, p<.001)] and BA36 [t(106)=4.1, p<.001)] continued to be quite sensitive to clinical status, CA [t(106)=7.2, p<.001)] displayed the largest effect size. Indeed, the pattern of atrophy appeared to be much more diffuse at this stage, with DG [t(106)=5.7, p<0.001], SUB [t(106)=5.2, p<0.001], and whole hippocampal volume [t(24.6)=4.6, p<0.001] demonstrating significant group discrimination. Similarly, at the AD stage, most regions displayed highly significant group differences when compared to controls with CA having the largest effect size, but only marginally relative to whole hippocampal volume.

Groups Dichotomized by Amyloid Status

Our primary analysis was to examine the relative effect on MTL subregional structure of AD along the continuum of preclinical disease to mild dementia. To so, a "normal aging" group (amyloid negative CN adults) was compared to preclinical AD (amyloid positive CN adults), early and late prodromal AD (amyloid positive EMCI and LMCI, respectively) to mild AD dementia (amyloid positive AD) based on amyloid status using florbetapir PET (Table 3). We only included florbetapir scans that were obtained within 1-year of the MRI scan (mean: 135.6 ± 157.1 days) used for MTL subregion analysis, and, thus, some individuals in the prior analysis were excluded. In light of the 8 ROIs examined per group comparison, statistical effects that would survive Bonferroni correction (p<0.006) were noted with bold font in Table 3.

The only region to reach significance in the preclinical AD group relative to "normal aging" was BA35 [t(62)=2.0, p<0.05]. No other region approached significance. Interestingly, in absolute terms measured by percent difference, BA35 (8.1%) and PHC (5.2%) displayed the greatest atrophy in the preclinical AD (all other regions < 1%). Indeed, several regions, including CA and whole hippocampus volume was actually somewhat larger in the preclinical AD group.

In contrast, in the early prodromal AD stage most regions displayed some degree of decreased volume or thickness relative to the "normal aging" group. All extrahippocampal MTL regions reached significance [BA35: t(21)=2.9, p<0.01; BA36: t(59)=3.4, p<0.001; ERC: t(59)=2.9, p<0.01; PHC: t(59)=3.5, p<0.001]. While all of the hippocampal subregions were also significant, these effects were more marginal [subiculum: t(59)=1.8, p<0.05); CA: t(20.0)=1.8, p<0.05; DG: t(20.0)=2.0, p<0.05]. Notably, whole hippocampal volume did not significantly differ with the "normal aging" group, but did display a nonsignificant trend [t(18.4)=1.5, p=0.07]. Extrahippocampal MTL regions also displayed generally greater percent differences with the "normal aging" group than hippocampal structures ($\sim12-18\%$ vs $\sim6-8\%$).

In late prodromal AD and mild AD dementia there was much more diffuse involvement of MTL structures and all regions reached statistical significance in comparison with "normal aging". Indeed, CA was the region of largest effect size in the late prodromal AD comparison [t(53)=6.7, p<0.001] and both CA and whole hippocampus best discriminated groups in mild AD dementia [t's=7.0, p's<0.001]. Nonetheless, even in the latter group, BA35 displayed the greatest percent difference with the "normal aging" group. Figure 2 plots the evolution of atrophy in select regions across disease stage with this pseudo-

longitudinal data. Note that adjustment for gender had no substantive impact on these comparisons.

We also calculated the estimated sample size required to discriminate the amyloid positive groups from the "normal aging" group with a significance criterion of 0.05 (one-sided) and a power of 0.8 to correctly reject the null hypothesis if it is false. We assumed equal samples sizes from both groups (see Table 3). As suggested by the above statistical comparisons, extrahippocampal MTL regions would allow for smaller sample sizes in preclinical and early prodromal AD than hippocampal subfields and whole hippocampal volume while this advantage would be lost during the late prodromal and mild dementia stages. Importantly, BA35 appeared to allow for detection of disease effects with a reasonable number of participants in these very early stages (i.e. n=86 in preclinical AD and n=30 in prodromal AD).

Finally, we examined whether there were statistically significant differences in discrimination between the subfields with largest effect sizes versus whole hippocampus by comparing areas under the curve (AUC). We did find the BA35 had a significantly higher AUC than whole hippocampus in discriminating "normal aging" from preclinical AD (.65 vs. .44, p< 0.05). In early prodromal AD, PHC displayed a trend towards significantly greater discrimination (.76 vs. .60, p=0.08). In late prodromal AD, AUC's favored BA35 (.93) and CA (.93) relative to whole hippocampus (.89), but these differences did not reach significance (p>0.1).

Discussion

There are two primary findings from the current work. The first is that more granular measurement of MTL subregions provides enhanced sensitivity to early stages of AD compared to whole hippocampal measurements. The second is that the pattern of atrophy within the MTL associated with different degrees of AD severity, from preclinical to dementia, generally recapitulates the topographic pattern of NFT deposition reported in the histology literature. Indeed, regions associated with the earliest NFT pathology, particularly PRC and ERC, displayed significant atrophy in preclinical and early prodromal AD (Braak & Braak, 1991). Taken together, these data hold promise for the value of MTL subregional measurements as a biomarker sensitive to the earliest neurodegenerative effects of disease that could serve an important role in tracking disease progression in preclinical and prodromal AD intervention studies. As this represents the first experience with our fully automated segmentation pipeline in a multi-site context, the present findings also support the practicality of this approach in future clinical studies.

Selective involvement of MTL subregions in preclinical and prodromal AD

A primary motivation for measurement of subregions within the MTL is the potential increased sensitivity to early AD relative to whole hippocampal measurement (Fox et al., 1996; Jack, Petersen, O'Brien, & Tangalos, 1992; Jack et al., 2011). Indeed, several MTL subregions displayed stronger discrimination between preclinical and early symptomatic patients from CN adults, but caution must be applied as the power to detect statistical differences in discrimination was lacking once groups were divided based on amyloid status.

Nonetheless, BA35 was the only region to discriminate preclinical AD from amyloid negative CN adults ("normal aging") while BA35/36, ERC, and PHC discriminated early prodromal AD from this group. In the latter comparison, hippocampal subfields displayed more modest discrimination and whole hippocampus trend level discrimination. By later clinical stages, this advantage was less evident. Such differences in sensitivity to these early disease changes have implications for size of cohorts that may need to be recruited in the context of an intervention study. This is demonstrated in the sample size calculations in which, for example, 17 individuals per group (34 total) would be needed for a reasonably powered study discriminating early prodromal AD from "normal aging" with BA35 measurements while 52 per group (104 total) would be required with whole hippocampal volume.

While this is the first study to comprehensively examine MTL subregional change across varying degrees of clinical progression, other work has supported the general principle of enhanced sensitivity to MCI with measurement of specific subregions (Iglesias et al., 2015; La Joie et al., 2013; S. G. Mueller et al., 2010). For example, La Joie and colleagues reported stronger group effects for CA1 than other subfields or volume of entire hippocampal formation in a manual tracing study (La Joie et al., 2013). Similarly, Mueller et al. found a CA1-2 transition zone to most strongly distinguish these groups (S. G. Mueller et al., 2010). In the current analysis, we also found CA1 to be the hippocampal subfield that tended to be associated with the most atrophy in MCI and AD. Notably, the above studies did not include PRC measurements, which would be expected to precede CA1 involvement. Indeed, in a different MCI cohort from here, we reported strongest group discrimination with CN adults for both CA1 and BA35 (Pluta et al., 2012; Yushkevich et al., 2015). While beyond the scope of the current analysis, it is also worth noting that the BA35 thinning found in the preclinical group also appeared to relate to performance on AVLT delayed memory measures (5-minute delayed recall: r=.52, p=0.014; recognition hits minus false alarms: r=.51, p=0.015), supporting potential early cognitive consequences of this subtle atrophy.

Advantages of subregions relative to more global measures of hippocampal volume are less evident in later clinical stages, which is not unexpected given the relatively more diffuse involvement of MTL structures with disease progression.(Braak & Braak, 1991)

Nonetheless, examination of absolute differences in individual subregions suggests that there still may remain some degree of selectivity, but without translating into greater statistical significance. The increased precision of measurement of the entire hippocampus versus that of smaller subregions, particularly in later disease stages, likely mitigates against the value of the larger absolute differences detected in these regions.

Measurement of Perirhinal Cortex

There is relatively little work on *in vivo* measurement of PRC despite the medial portion of this structure, the 'transentorhinal' region, being associated with the earliest NFT pathology (Braak & Braak, 1991). A number of reasons likely account for this limited attention, including anatomic variability, disagreement about boundaries, and even nosology, as there is inconsistency in whether the PRC is defined as including both BA35 and BA36

(Augustinack et al., 2013; Ding & Van Hoesen, 2010). Nonetheless, Augustinack and colleagues recently developed an automated surface-based labeling method of BA35 derived from annotation of very high-resolution post-mortem scans (Augustinack et al., 2013). They applied this approach to ADNI data and also found very strong group discrimination between MCI patients and controls.

We utilized an in vivo atlas in which BA35 and BA36 were separately labeled and guided by description of the anatomic variability of these structures defined by sulcal patterns reported in the exhaustive histological work of Ding and Van Hoesen (Ding & Van Hoesen, 2010). Our prior work has suggested good reliability of automatic segmentations with manual ones (Yushkevich et al., 2015). Errors are largely driven by differences in the number of slices labeled anterior to the head of the hippocampus, as the atlas protocol artificially limits these segmentations to just one slice rostral to the hippocampal head. As such, the entire structure is not included and the anterior extent is dependent on the size of the hippocampus, which is further influenced by disease status. This produces confounds for measurement of volume, which, for example, may be underestimated in an individual with a smaller hippocampal head. In light of this issue and some degree of uncertainty in the medial and lateral boundaries of these regions using sulcal pattern heuristics (see Figure 3) that may not completely conform to cytoarchitecture, we measured mean cortical thickness, which likely limits the impact of these errors relative to volume measurements.

In prior work, we have also utilized thickness measures or "normalized" volumes based on the number of slices included in the segmentation, essentially a measure of area (Yushkevich et al., 2015). However, the latter does not account for uncertainty in the lateral boundaries or the fact that individuals with the same overall volume may differ along the anterior-posterior and medial-lateral dimensions. Notably, we previously found evidence of strong discrimination between MCI and controls in BA35 using thickness in a single-site study, but used a T2-derived measure of thickness (see Yuchkevich et al., 2015) rather than the T1-extracted thickness used here. The anisotropic nature and susceptibility to motion artifact of the T2 scans make this approach challenging, particularly in the ADNI dataset, which is somewhat noisier for this sequence relative to that obtained in our prior study (Yuchkevich et al., 2015). We did examine both methods in the current dataset and found the T1 data more sensitive. We continue to work to improve these measurements both with regard to acquisition quality and analysis approach (Xie et al., 2014).

Pattern of atrophy recapitulates Braak staging

In addition to examining group statistics, we also calculated absolute percent change in volume and thickness across the spectrum of AD relative to "normal aging". This cross-sectional, pseudo-longitudinal analysis is akin to the inference that Braak and Braak originally used to stage temporal ordering of NFT deposition in autopsy data (Braak & Braak, 1991). Our findings do appear to qualitatively match the expected pattern of regional involvement. Specifically, BA35 displayed the greatest percent difference (8.1%) in preclinical AD while all other regions were less than 1% smaller or even slightly larger than the control group. The only exception was PHC, which was 5.2% thinner. In early prodromal AD, ERC and other extrahippocampal MTL regions displayed more significant cortical

thinning, while hippocampal regions were more modestly affected. By late prodromal AD and mild AD dementia, hippocampal subfields were more prominantly atrophied with CA being the most affected although the degree of absolute volume change was only marginally more than DG and subiculum. These findings broadly match the spread of NFT's initiated in transentorhinal cortex (BA35) and spreading to adjacent cortical MTL areas and then into the hippocampus proper focused in CA1 (Miller et al., 2015). However, one should be cautious in over-interpretation of these findings given their qualitative nature.

Interestingly, we found the BA36 portion of PRC displayed similar or better discrimination than BA35 in early prodromal AD. This result is somewhat unexpected given the greater NFT burden earlier in the latter. Despite being frequently grouped together as PRC, these two regions differ in cytoarchitecture with BA35 and 36 being best classified as periallocortex-proisocortex and isocortex, respectively (Augustinack et al., 2013; Ding & Van Hoesen, 2010). It is worth noting that a more precise timing of BA36 involvement relative to BA35 is lacking in the literature, but clearly both regions are significantly affected in later symptomatic disease (Arnold et al., 1991; Braak & Braak, 1991; Van Hoesen, Augustinack, Dierking, Redman, & Thangavel, 2000). Importantly, BA35 displayed a larger degree of cortical thinning in absolute terms compared to BA36 across the spectrum from preclinical AD to mild AD dementia. The small size of BA35 and its location along the collateral sulcus likely contributes to increased variability, which may reduce its discriminatory power relative to BA36.

Limitations

While this represents the largest cohort of MCI/AD patients utilizing this MTL-specific T2weighted MRI acquisition, the sample size remains modest, particularly when groups were divided based on amyloid status. This limits inference about the degree to which subregions are dissociable in their sensitivity to disease stage. Indeed, despite differences in apparent effect size, only BA35 significantly differed from whole hippocampal volume in discrimination of preclinical AD while PHC displayed a trend towards such a difference in the early prodromal phase. It is also important to point out that a number of comparisons were made for each group comparison and not all significant effects survive Bonferroni correction. However, this analysis was not entirely exploratory and many of the reported findings are consistent with prediction based on Braak staging and the expected association with progressive atrophy in these regions. Further, the cross-sectional nature of the study makes arguments about sensitivity to disease progression tentative. For example, the fact that the groups are well-matched in age does not reflect the fact that in a longitudinal cohort both age and disease would contribute to change. Increased sample size and longitudinal analyses would also allow for more detailed examination of the predictive value of these measures and relationship to cognitive change.

The accuracy of thickness measurements in PRC, PHC, and ERC is limited by accuracy of gray matter segmentation in T1-MRI, as well as that of the respective ROI labels in T2-MRI. BA35, in particular, has lower segmentation reliability due to its small size and proximity to the collateral sulcus. Gray matter segmentation may also be affected by errors in resolving sulcal CSF in this region. These factors introduce higher variance in the thickness

measurements. However, these limitations also represents potential for improvements, and we continue to work on refined methodologic approaches to enhance reliability, such as additional strategies to measure thickness and combined use of T1- and T2-weighted sequences in the segmentation process (Xie et al., 2014; Yushkevich et al., 2015). Further, there are on-going collaborative efforts to use multimodal data, including histology, to develop a harmonized hippocampal subfield protocol, which has the potential to improve validity of these measures (hippocampalsubfields.com).

Conclusion

This manuscript describes use of an automated tool for segmentation of subregions within the MTL. Consistent with the selective topography of NFTs in early AD, subregional measurement appeared to offer enhanced sensitivity at preclinical and early prodromal disease stages compared to whole hippocampal volumes. As such, this may be a valuable biomarker for therapeutic trials in these populations.

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• We determined the pattern of MTL subregional atrophy along the continuum from preclinical AD to mild dementia

- We applied a novel, automated method for measurement of hippocampal subfields and extrahippocampal MTL cortical regions
- Medial perirhinal cortex (BA35) was the only region sensitive to preclinical AD
- Pattern of atrophy along the continuum of disease severity recapitulated
 Braak staging of NFTs within the MTL
- More granular MTL measurement enhances sensitivity to early AD stages relative to whole hippocampal volume

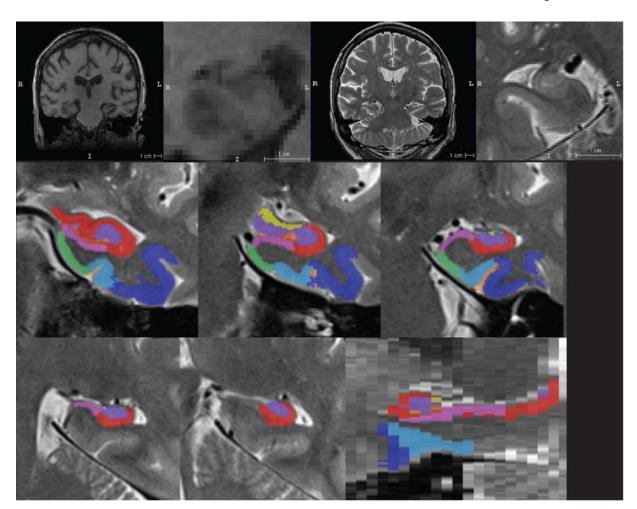


Figure 1.Top row from left to right: T1-MRI, T1-MRI zoomed in to hippocampus in coronal view, T2-MRI, zoomed in to hippocampus in oblique coronal view. Middle and bottom row show an example of automatic segmentation MTL subregions.

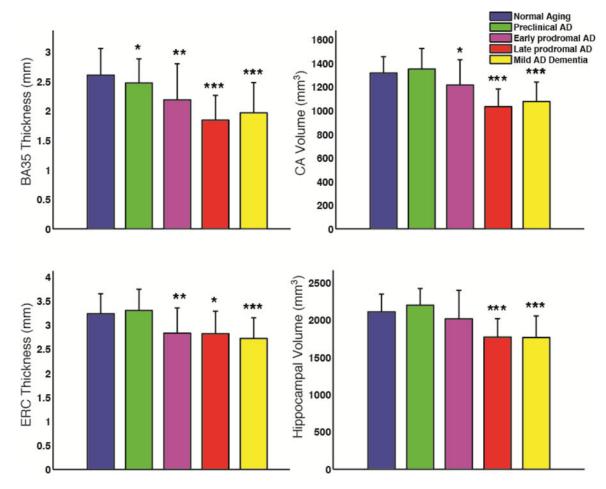
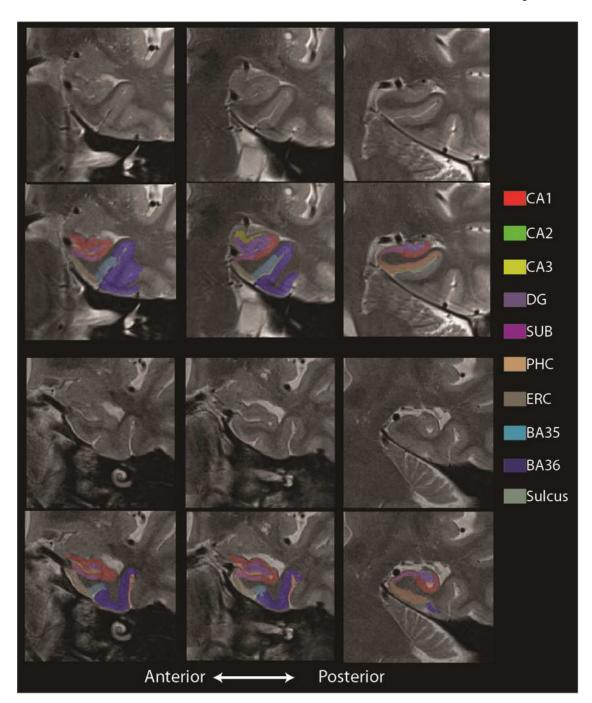


Figure 2. Group comparison of select MTL subregional measurements adjusted for age, as well as ICV when volume measurements. Statistical significance is based on comparison with the "normal aging" (CN Am-) group. Error bars reflect 1 standard deviation. *: p<0.05, **: p<0.01, ***:p<0.001.



Example of automatic segmentation in two individuals at several slices along the anterior-posterior axis. In particular, these cases exhibit the heuristic rules and variability of lateral boundaries in BA35 and BA36 based on collateral sulcus depth (top two rows: deep collateral sulcus; bottom two rows: shallow collateral sulcus)

Table 1

Group demographic, psychometric, and biomarker data.

	CN (n=86)	EMCI (n=43)	LMCI (n=22)	AD (n=40)
Age	74.3 (6.9)	74.4 (7.1)	74.1 (7.4)	74.3 (8.7)
Education	16.9 (2.2)	16.7 (2.9)	16.7 (2.8)	16.6 (2.4)
LM Delay	14.7 (3.3)	12.0 (3.5) ***	4.3 (3.2) ***	1.5 (2.9) ***
Trails A (sec)	32.0 (12.9)	36.9 (16.6)	38.4 (17.8)	55.4 (31.4) ***
Trails B (sec)	84.6 (62.1)	92.7 (48.7)	111.24 (62.6)	171.3 (90.3)***
BNT Total	28.5 (2.0)	27.5 (3.4)	27.0 (3.9)	25.5 (6.0) ***
Category Fluency	21.4 (5.7)	18.7 (5.1)**	16.8 (5.0) **	12.2 (4.6) ***
AVLT (Trials 1–5)	47.8 (10.9)	43.0 (12.7)*	31.5 (9.3) ***	23.4 (8.3) ***
AVLT 5-min Delay	9.8 (3.5)	7.8 (3.8) **	4.6 (3.1) ***	1.4 (1.9) ***
AVLT 30-min Delay	8.2 (4.3)	6.1 (4.8)*	1.9 (2.7) ***	0.5 (1.7)***
Florbetapir PET (SUVR)	1.13 (0.3)	1.16 (0.2)	1.29 (.21) ***	1.36 (.21)***

All statistics are in comparison to CN group;

* p<0.05;

** p<0.01;

*** p<0.001.

Note that 1 LMCI, and 3 AD patients did not complete Trails B and 1 AD did not complete the AVLT 30-min delayed recall.

Table 2

Comparisons to control group after adjustment for age and education, as well as ICV for volume measurements.

	CDV / 0.0	T1. (CT / 42)		17 (10)
~.	CN (n=86)	EMCI (n=43)	LMCI (n=22)	AD (n=40)
CA				
Volume (SD) mm ³	1323.5 (147.4)	1261.9 (213.6)	1059.0 (182.0)	1027.8 (190.4)
t-value		1.7	7.2	8.7
p-value		<0.05	< 0.001	< 0.001
Dentate Gyrus				
Volume (SD) mm ³	786.8 (86.9)	747.8 (129.4)	654.9 (127.4)	628.0 (116.5)
t-value		1.8	5.7	8.5
p-value		< 0.05	< 0.001	< 0.001
Subiculum				
Volume (SD) mm ³	440.0 (61.2)	413.7 (62.5)	365.3 (53.0)	344.4 (51.6)
t-value		2.3	5.2	8.6
p-value		< 0.05	< 0.001	< 0.001
ERC				
Thickness (SD) mm	3.25 (.42)	2.99 (.54)	2.78 (0.56)	2.68 (0.44)
t-value		3.1	4.3	7.0
p-value		< 0.01	< 0.001	< 0.001
BA35				
Thickness (SD) mm	2.58 (0.45)	2.32 (.63)	1.99 (0.53)	2.02 (0.55)
t-value		2.4	5.3	5.6
p-value		< 0.05	< 0.001	< 0.001
BA36				
Thickness (SD) mm	3.35 (0.41)	2.99 (.51)	2.91 (0.55)	2.78 (0.56)
t-value		4.0	4.1	5.7
p-value		2e ⁻⁵	4e ⁻⁵	2e ⁻⁷
РНС				
Thickness (SD) mm	2.25 (.45)	2.06 (.61)	1.84 (.40)	1.96 (.53)
t-value		2.0	3.8	3.1
p-value		< 0.05	< 0.001	< 0.01
Нірро				
Volume (SD) mm ³	2144.6 (218.8)	2074.3 (364.8)	1758.9 (381.3)	1691.0 (307.6)
t-value		1.2	4.6	8.4
p-value		>0.1	< 0.001	< 0.001

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Table 3

Comparison to CN, Amyloid Negative group; all values adjusted for age and education and volumes adjusted for ICV

(SD) mm ³ 1311.8 (125.9) ence from CN Size mm (SD) mm ³ 784.7 (74.3) ence from CN Size size contact from CN size s						
(SD) mm³ 1311.8 (125.9) ence from CN Size Gyrus (SD) mm³ 784.7 (74.3) ence from CN size Size (SD) mm³ 437.8 (56.9) ence from CN ss (SD) mm 3.23 (.45) ence from CN Size Size Size Size Size Size Size Size		CN (n=44) Amyloid Neg	CN (n=20) Amyloid Pos	EMCI (n=17) Amyloid Pos	LMCI (n=11) Amyloid Pos	AD (n=26) Amyloid Pos
Size Gyrus Size	YC.					
Size Gyrus Gyrus (SD) mm³ Size um (SD) mm³ 437.8 (56.9) ence from CN ence from CN Size	Volume (SD) mm ³	1311.8 (125.9)	1340.2 (176.9)	1208.7 (224.3)	1018.3 (148.2)	1069.2 (160.6)
Size Gyrus Gyrus (SD) mm³ 784.7 (74.3) ence from CN size	% difference from CN		-2.2	7.9	22.4	18.5
Size Gyrus (SD) mm³ 784.7 (74.3) ence from CN Size (SD) mm³ 437.8 (56.9) ence from CN ss (SD) mm 3.23 (.45) ence from CN Size	-value		<1	1.8	2.9	0.7
Gyrus 784.7 (74.3) ence from CN 784.7 (74.3) um 784.7 (74.3) slize 81.2 ence from CN 437.8 (56.9) ss (SD) mm 3.23 (.45) ss (SD) mm 3.23 (.45) ence from CN 81.23 (.45)	-value		>0.1	<0.05	<0.001	<0.001
Gyrus 784.7 (74.3) ence from CN 784.7 (74.3) size 437.8 (56.9) sence from CN 83.23 (.45) ss (SD) mm 3.23 (.45) ence from CN 3.23 (.45)	sample Size		N/A	80	8	12
size Size cnc from CN Size cnc from CN Size ss (SD) mm³ 437.8 (56.9) cnc from CN ss (SD) mm 3.23 (.45) cnc from CN ss (SD) mm Size	Dentate Gyrus					
Size (SD) mm³ (SE)	Volume (SD) mm ³	784.7 (74.3)	780.6 (103.2)	715.8 (133.4)	634.6 (106.1)	661.3 (94.6)
Size (SD) mm ³ 437.8 (56.9) ence from CN ss (SD) mm 3.23 (.45) ence from CN ss (SD) mm Size	% difference from CN		0.5	8.7	19.2	15.7
Size (SD) mm³ 437.8 (56.9) ence from CN Size ss (SD) mm 3.23 (.45) ence from CN	-value		<1	2.0	5.5	6.1
Size (SD) mm³ 437.8 (56.9) ence from CN Size ss (SD) mm 3.23 (.45) ence from CN	-value		>0.1	<0.05	<0.001	<0.001
(SD) mm ³ 437.8 (56.9) ence from CN Size ss (SD) mm 3.23 (.45) ence from CN size size size size size size size size	sample Size		>1000	64	12	14
(SD) mm ³ 437.8 (56.9) ence from CN Size ss (SD) mm 3.23 (.45) ence from CN	subiculum					
Size Size ss (SD) mm 3.23 (.45) snce from CN	Volume (SD) mm ³	437.8 (56.9)	442.1 (72.4)	407.7 (64.0)	357.8 (47.9)	360.5 (43.8)
Size ss (SD) mm 3.23 (.45) ence from CN	% difference from CN		-0.1	6.9	18.3	17.7
Size ss (SD) mm 3.23 (.45) ence from CN Size	-value		<1.0	1.8	4.3	0.9
Size ss (SD) mm 3.23 (45) ence from CN Size	-value		>0.1	<0.05	<0.001	< 0.001
ss (SD) mm 3.23 (.45) ence from CN	sample Size		N/A	102	14	14
ss (SD) mm 3.23 (45) ence from CN	ERC					
ence from CN	ľhickness (SD) mm	3.23 (.45)	3.27 (.44)	2.84 (.55)	2.89 (.48)	2.71 (.45)
Size	% difference from CN		-1.2	12.1	10.5	16.1
Size	-value		<1	2.9	2.2	4.7
	-value		>0.1	<0.01	<0.05	<0.001
	Sample Size		N/A	42	46	20
BA35	3A35					
Thickness (SD) mm 2.70 (.40) 2.48 (.4)	Thickness (SD) mm	2.70 (.40)	2.48 (.41)	2.22 (.64)	1.86 (.46)	2.00 (.50)

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	CN (n=44) Amyloid Neg	CN (n=20) Amvloid Pos	EMCI (n=17) Amvloid Pos	LMCI (n=11) Amvloid Pos	AD (n=26) Amvloid Pos
% difference from CN	•	8.1	17.8	31.1	26.0
t-value		2.0	2.9	6.0	6.4
p-value		<0.05	<0.01	<0.001	<0.001
Sample Size		98	34	10	12
BA36					
Thickness (SD) mm	3.35 (.48)	3.34 (.33)	2.92 (.52)	2.94 (.53)	2.66 (.48)
% difference from CN		0.0	12.8	12.2	20.6
t-value		<1.0	3.4	2.8	6.4
p-value		>0.1	<0.01	<0.01	<0.001
Sample Size		>1000	36	40	14
PHC					
Thickness (SD) mm	2.31 (.43)	2.19 (.42)	1.89 (.43)	1.76 (.34)	2.00 (.58)
% difference from CN		5.2	18.2	23.8	13.4
t-value		<1	3.5	4.0	2.5
p-value		>0.1	<0.01	<0.001	<0.05
Sample Size		312	28	14	0/
Hippo					
Volume (SD) mm ³	2156.8 (176.2)	2193.5 (234.1)	2003.0 (402.1)	1768.2 (252.5)	1748.4 (266.3)
% difference from CN		-1.7	7.1	17.9	6.81
t-value		<1.0	1.5	0.9	0.7
p-value		>0.1	0.07	<0.001	<0.001
Sample Size		N/A	104	10	10

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