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Authors Weghorst, Forrest P Cramer, Karina S

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EMBRYONIC DEVELOPMENT

The evolution of hearing and balance

New genetic tools have allowed researchers to compare how the brainstem auditory and vestibular nuclei develop in embryonic chicks and mice.

FORREST P WEGHORST AND KARINA S CRAMER

Related research article Lipovsek M, Wingate RJ. 2018. Conserved and divergent development of brainstem vestibular and auditory nuclei. *eLife* **7**:e40232. DOI: 10.7554/eLife.40232

he ear is an organ with two main roles hearing and balance - and it relies on mechanically sensitive hair cells to perform both jobs. When the ear detects a sound, it sends signals to clusters of neurons called the brainstem auditory nuclei, and when it registers movement of the head, it sends signals to the brainstem vestibular nuclei. The vestibular nuclei are remarkably similar among vertebrates, from fish to humans (Fritzsch et al., 2014). However, the auditory nuclei display considerable diversity across species. For example, birds and mammals both have a pathway that uses the difference in the time of arrival of a sound at each ear to determine where the noise came from. However, this circuit works differently in these two groups of animals, suggesting that it may have emerged independently multiple times during evolution (Grothe and Pecka, 2014). How, then, did the modern auditory and vestibular nuclei arise?

One strategy to address this question is to explore the embryonic development of vertebrates. Nuclei in the brainstem arise from the embryonic hindbrain, which is remarkably similar across vertebrate species and is divided into segments called rhombomeres (*Di Bonito and Studer, 2017*). Fate mapping studies have been used to test whether cells in the auditory and vestibular nuclei of different vertebrate species derive from the same rhombomeres.

In this technique, embryonic tissue can be labeled with an external marker, such as a dye (for traditional fate mapping), or a fluorescent protein marker (for genetic fate mapping), in order to track the destination of the cells arising from that tissue (Stern and Fraser, 2001; Legué and Joyner, 2010). Previously, traditional fate mapping has been limited to research in birds (which have accessible embryos), while genetic fate mapping has been limited to mam-(which have accessible genomes; mals Cramer et al., 2000; Marín and Puelles, 1995; Kim and Dymecki, 2009).

However, both approaches have technical caveats and they identify progenitors in different ways, which hinders their direct comparison. Traditional fate mapping identifies the fate of all labeled cells within a restricted area of the embryo, which may contain a range of progenitor cell types. In contrast, genetic fate mapping marks only cells that express a certain gene (or genes), but these cells can come from a wider area within the embryo.

Now, in eLife, Marcela Lipovsek and Richard Wingate of King's College London report how they have addressed this dilemma by using vector-based genetic fate mapping in chick embryos (*Lipovsek and Wingate, 2018*). The researchers focused on genes that were only active in certain regions of the embryonic

© Copyright Weghorst and Cramer. This article is distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use and redistribution provided that the original author and source are credited. hindbrain. Plasmid vectors were used to label cells with a fluorescent protein after specific genes in those cells were active. This way, the fate of cells along two anatomical axes (from head to tail, and from back to belly) could be traced. Lipovsek and Wingate studied the same genes that were previously used to construct genetic fate maps of brainstem nuclei in mice, which enabled them to draw direct comparisons between birds and mammals for the first time (**Di Bonito and Studer, 2017**).

Their results confirmed that vestibular nuclei have similar embryonic origins in chicks and mice. In contrast, auditory nuclei that have comparable roles in chicks and mice arise from completely different embryonic tissues. This suggests that birds and mammals used different populations of ancestral cells – often from different rhombomeres – to independently evolve circuits for calculating sound location.

Lipovsek and Wingate provide compelling evidence that the anatomical similarities between vestibular nuclei in birds and mammals are due to common developmental and evolutionary origins. And since their vestibular systems are also homologous to those of fish, it seems that the role of the vestibular organ remained relatively unaffected by our aquatic ancestors' move to land (*Fritzsch et al., 2014*). Conversely, the different developmental origins of auditory nuclei in birds and mammals reflect how each clade solved the problem of hearing on land by adapting to its own ecological niche (*Carr and Christensen-Dalsgaard, 2016*).

Forrest P Weghorst is in the Department of Neurobiology and Behavior, University of California, Irvine, United States thttps://orcid.org/0000-0001-9365-1846

Karina S Cramer is in the Department of Neurobiology

and Behavior, University of California, Irvine, United States cramerk@uci.edu

b https://orcid.org/0000-0003-3793-4862

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References

Carr CE, Christensen-Dalsgaard J. 2016. Evolutionary trends in directional hearing. Current Opinion in Neurobiology 40:111–117. DOI: https://doi.org/10. 1016/j.conb.2016.07.001, PMID: 27448850 Cramer KS, Fraser SE, Rubel EW. 2000. Embryonic origins of auditory brain-stem nuclei in the chick hindbrain. Developmental Biology 224:138–151. DOI: https://doi.org/10.1006/dbio.2000.9779, PMID: 10926755

Di Bonito M, Studer M. 2017. Cellular and molecular underpinnings of neuronal assembly in the central auditory system during mouse development. *Frontiers in Neural Circuits* **11**:18. DOI: https://doi.org/10.3389/ fncir.2017.00018, PMID: 28469562

Fritzsch B, Kopecky BJ, Duncan JS. 2014. Development of the mammalian 'vestibular' system: evolution of form to detect angular and gravity acceleration. In: Romand R, Varela-Nieto I (Eds). Development of Auditory and Vestibular Systems. San Diego: Academic Press. p. 339–367. DOI: https://doi. org/10.1016/B978-0-12-408088-1.00012-9

Grothe B, Pecka M. 2014. The natural history of sound localization in mammals – a story of neuronal inhibition. *Frontiers in Neural Circuits* **8**:116. DOI: https://doi.org/10.3389/fncir.2014.00116, PMID: 25324726

Kim JC, Dymecki SM. 2009. Genetic fate-mapping approaches: new means to explore the embryonic origins of the cochlear nucleus. In: Sokolowski B (Ed). *Auditory and Vestibular Research*. Humana Press. p. 65–85. DOI: https://doi.org/10.1007/978-1-59745-523-7 5

Legué E, Joyner AL. 2010. Genetic fate mapping using site-specific recombinases. *Methods in Enzymology* **477**:153–181. DOI: https://doi.org/10.1016/S0076-6879(10)77010-5, PMID: 20699142

Lipovsek M, Wingate RJ. 2018. Conserved and divergent development of brainstem vestibular and auditory nuclei. *eLife* **7**:e40232. DOI: https://doi.org/10.7554/eLife.40232, PMID: 30566077

Marín F, Puelles L. 1995. Morphological fate of rhombomeres in quail/chick chimeras: a segmental analysis of hindbrain nuclei. *European Journal of Neuroscience* **7**:1714–1738. DOI: https://doi.org/10. 1111/j.1460-9568.1995.tb00693.x, PMID: 7582126 **Stern CD**, Fraser SE. 2001. Tracing the lineage of tracing cell lineages. *Nature Cell Biology* **3**:E216–E218. DOI: https://doi.org/10.1038/ncb0901-e216, PMID: 11533679