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Review

Corticotropin-Releasing Factor Family: A Stress Hormone-Receptor System's Emerging Role in Mediating Sex-Specific Signaling

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Abstract: No organ in the body is impervious to the effects of stress, and a coordinated response from all organs is essential to deal with stressors. A dysregulated stress response that fails to bring systems back to homeostasis leads to compromised function and ultimately a diseased state. The components of the corticotropin-releasing factor (CRF) family, an ancient and evolutionarily conserved stress hormone-receptor system, helps both initiate stress responses and bring systems back to homeostasis once the stressors are removed. The mammalian CRF family comprises of four known agonists, CRF and urocortins (UCN1–3), and two known G protein-coupled receptors (GPCRs), CRF₁ and CRF₂. Evolutionarily, precursors of CRF- and urocortin-like peptides and their receptors were involved in osmoregulation/diuretic functions, in addition to nutrient sensing. Both CRF and UCN1 peptide hormones as well as their receptors appeared after a duplication event nearly 400 million years ago. All four agonists and both CRF receptors show sex-specific changes in expression and/or function, and single nucleotide polymorphisms are associated with a plethora of human diseases. CRF receptors harbor N-terminal cleavable peptide sequences, conferring biased ligand properties. CRF receptors have the ability to heteromerize with each other as well as with other GPCRs. Taken together, CRF receptors and their agonists due to their versatile functional adaptability mediate nuanced responses and are uniquely positioned to orchestrate sex-specific signaling and function in several tissues.

Keywords: BNST; cardiovascular; CRF; diabetes; GPCR; gut; inflammatory bowel disease; pancreas; reproduction; sexually dimorphic; urocortins

1. Introduction: The Corticotropin-Releasing Factor and Receptor Family

Corticotropin-releasing factor/hormone (CRF or CRH), urocortins, their paralogs, urotensin-I and sauvagine, and their two known G protein-coupled receptors, CRF₁ and CRF₂, are a family of ancient peptide hormones found in all chordates (Figure 1A–C). A diuretic hormone found in the fruit fly, *Drosophila melanogaster*, shows homology with the mammalian CRF and urocortins and likely served as a potential peptide precursor in early metazoan ancestry nearly 800 million years ago (Figure 1A,B). The mammalian CRF receptors and the fly diuretic hormone receptors appear to have evolved from a common ancestral G protein-coupled receptor, *seb-2*, which is present in worms, including *Caenorhabditis elegans* (Figure 1C). Phylogenetically, urotensin-I appears to have evolved first and is found in brain regions that evolved earlier and regulated physiological functions such as

osmo- and vasoregulation [1,2]; these functions are still shared by all homologs of the urotensin family. CRF is phylogenetically a younger peptide that evolved after duplication events [1,2] and is more prevalent in younger and higher order brain areas. CRF has overlapping physiological functions with the three urocortins, and together, these peptides regulate glucocorticoid and catecholamine release, energy metabolism, reproduction, cardiovascular, and immune functions, to list a few [3].

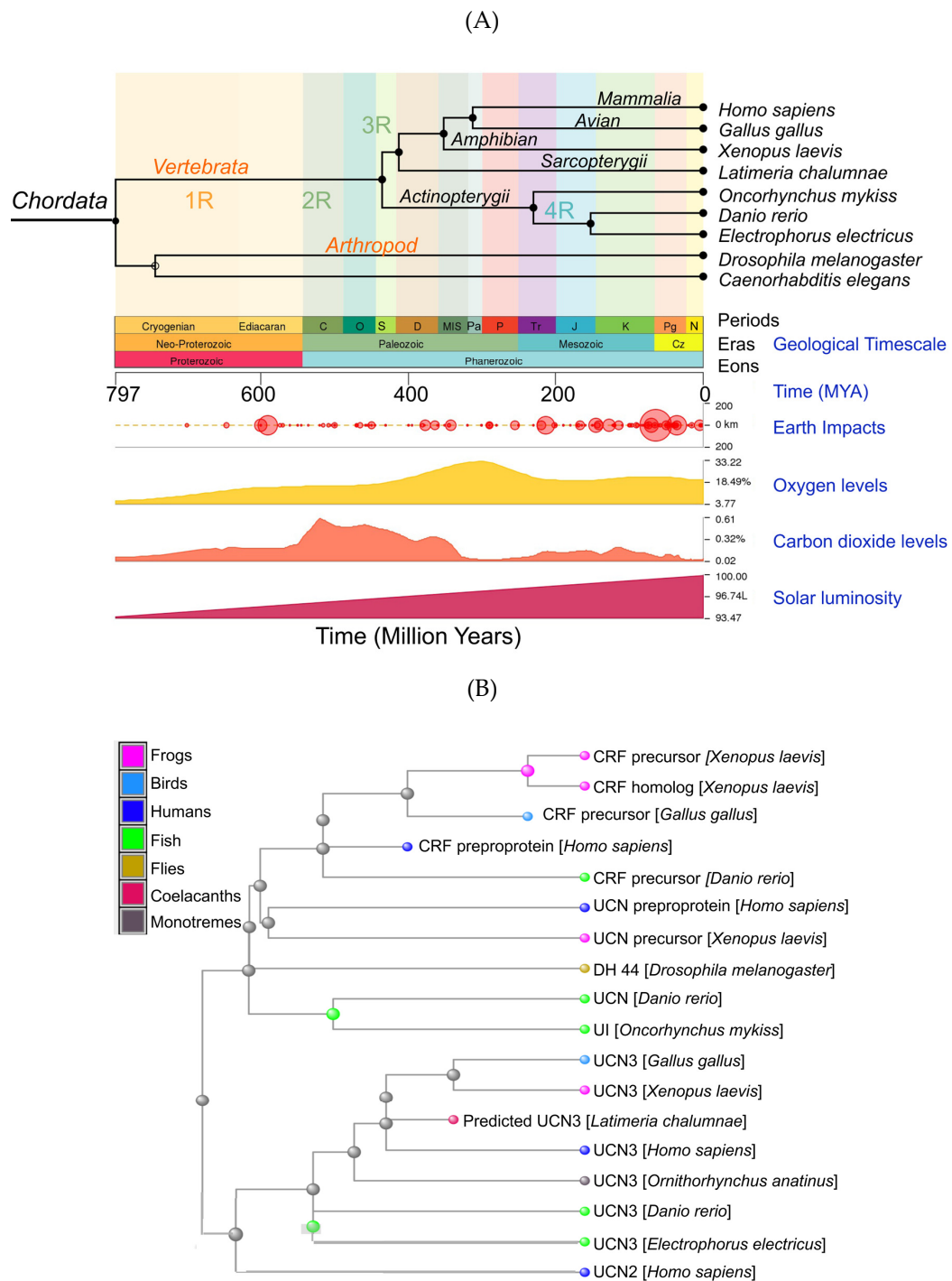


Figure 1. Cont.

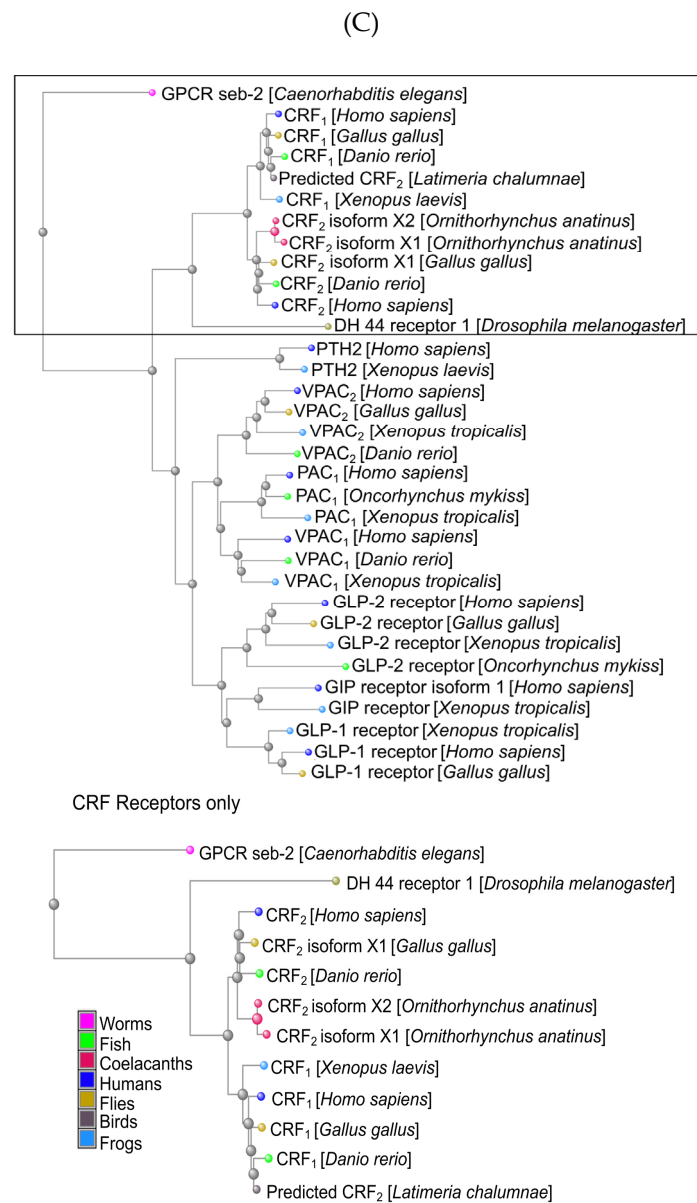


Figure 1. Origin of the corticotropin-releasing factor (CRF) family of peptide hormones: (A) Phylogenetic tree of showing evolution of prototype urotensin-I (UI), urocortins, and CRF with respect to geological times (in million years). The phylogenetic tree was generated using the TimeTree, which is a public knowledge base as described by Kumar et al. [4]. Prototype diuretic peptide hormones are present in the fruit fly, *Drosophila melanogaster*. Putative rounds of whole genome duplication events are depicted as 1R–4R. Earth impact events, changes in oxygen and carbon dioxide levels, as well as solar luminosity that might have influenced evolution are also shown. (B) A cluster dendrogram of the CRF family of peptides was generated in Cobalt using the National Center for Biotechnology Information (NCBI) database. These data suggest that UCN3 and UCN2 evolved earlier than UCN1 and CRF, with UCN3 and UCN2 being more homologous to each other than to CRF and UCN1. UCN3 appears to be most conserved evolutionarily. (C) Cluster dendrograms of CRF receptors and their orthologs. Boxed Inset: CRF receptors are more closely related to diuretic hormone 44 (DH44) receptors found in flies and arose from a common GPCR ancestral protein, seb-2, which is present in worms. CRF receptor orthologs: GIP: gastric inhibitory polypeptide receptor; GLP-1/2 R: glucagon-like peptide 1/2 receptors; PTH2: parathyroid hormone 2 receptor; PAC₁: pituitary adenylate cyclase-activating polypeptide type I receptors; VPAC_{1/2}; vasoactive intestinal polypeptide receptors 1 and 2.

The mammalian CRF family comprises of four known agonists, CRF and three urocortins (UCN1–3), and two known G protein-coupled receptors (GPCRs), CRF₁ and CRF₂ (Figure 2). In mammals, CRF is primarily responsible for regulating and/or initiating stress responses via activation of the hypothalamic–pituitary–adrenal (HPA) axis [5], whereas urocortins are involved in the recovery response to stress [6]. In peripheral organs, components of the CRF system are synthesized locally as in vivo RNA interference (RNAi) silenced its expression and altered gut function [7–9]. CRF and UCNs exhibit both autocrine and paracrine actions. While the actions of the CRF system in several processes are extensively researched as summarized in excellent reviews and references therein [3,4], the sex-specific modalities of the system are poorly understood. Female animals are generally not used in experiments, which led to the understanding of the role of the CRF system based on data primarily obtained from experiments based on male animals. Given that a myriad of diseases and disorders affecting tissues and organs in which the CRF system is expressed show sex-specific manifestation of diseases and outcomes, the need to characterize and elucidate the action and function of the CRF system in both sexes is needed. For example, epidemiological data suggest that the incidence of mental health and functional gastrointestinal disorders is higher in women than men [10,11], whereas rates of heart disease are higher in men than women [12,13], but how the actions of the CRF system differs between males and females in these organs is poorly understood. In this review, we will first touch upon the components of the CRF system and next review literature about what is known about the sex-specific actions of these hormones and their receptors.

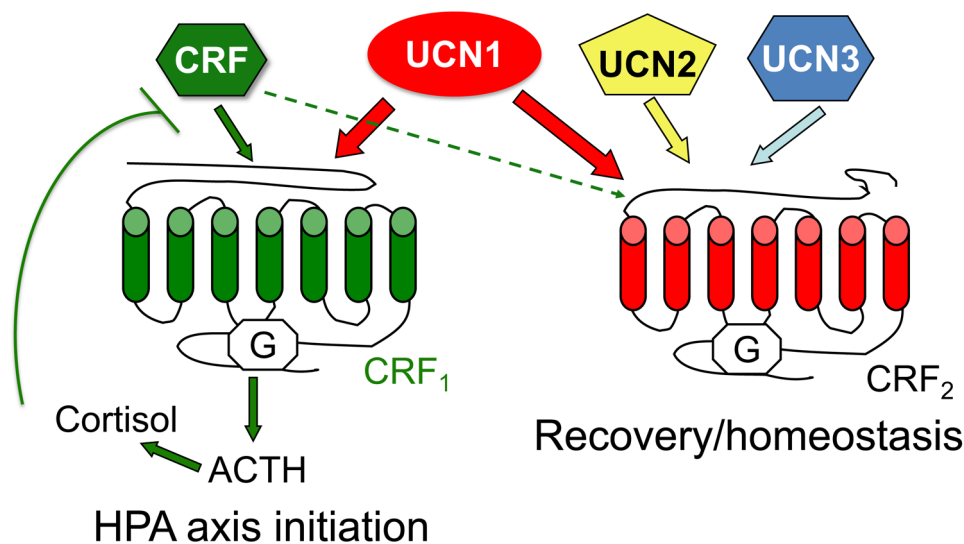


Figure 2. CRF family of peptide hormones and their two G protein-coupled receptors (GPCR) receptors, CRF₁ and CRF₂. Based on in vitro binding assays, CRF binds mainly to CRF₁, whereas urocortin 1 (UCN1) binds to both receptors, but with 10-fold higher affinity than other ligands. UCN2 and UCN3 bind exclusively to CRF₂. The activation of CRF₁ receptors by CRF in the hypothalamus initiates the hypothalamic–pituitary–adrenal (HPA) axis, ultimately resulting in the release of glucocorticoids (cortisol in primates and corticosterone in rodents) from the adrenal cortex. Activation of CRF₂ by urocortins is key for recovery from stress responses and returning to homeostasis.

2. The Hypothalamic–Pituitary–Adrenal (HPA) Axis

In the brain, CRF is synthesized by the paraventricular nucleus (PVN) and stored in the median eminence region of the hypothalamus [3]. In response to stress, CRF is released into the portal circulation from the median eminence, which in turn stimulates the secretion of adrenocorticotrophic hormone (ACTH) from the anterior pituitary into the bloodstream. ACTH goes on to regulate the release of glucocorticoids (cortisol in humans and corticosteroids in rodents) from the adrenal cortex. While urocortins are not known to directly activate the HPA axis, UCN1 activates the HPA axis by

stimulating CRF production [14]. Elevated levels of cortisol during the stress response result in an increased concentration of amino acids in the blood, the increased release of fatty acids, and increased blood glucose levels as a result of stimulating the breakdown of non-carbohydrate sources. Elevated glucocorticoid levels as seen during chronic stress can have several adverse effects on physiology that are well-documented [15]. HPA axis function is also implicated in biology of addiction and alcohol dependence [16]. While genomic and non-genomic actions of glucocorticoids [17,18] that result in increased intracellular levels of Ca^{2+} are also well known, the mechanisms behind this increased Ca^{2+} levels in neurons that are less studied include extrusion or clearance of toxic levels of intracellular Ca^{2+} , which can ultimately lead to neuronal cell death [19,20]. Moreover, hormones of the hypothalamic–pituitary–adrenal and gonadal axes regulate a plethora of functions that include immune, metabolic, and reproduction [21,22].

3. Looking Beyond the Fight or Flight Stress Response

The body's integration of the nervous and endocrine systems in response to stress is critical to maintaining homeostasis, a term coined by Walter Cannon in 1929 [23]. The evolutionarily conserved survival mechanism that coordinates the reactions to stress in vertebrates was termed as the "fight or flight" response by Hans Selye [24,25], but coordination of stress responses requires orchestration of several organs and their functions [15]. The hypothalamus simultaneously activates the adrenal cortex via the HPA axis to release glucocorticoids and the sympathetic nervous system, which in turn secretes the hormones epinephrine and norepinephrine. Together, these circulating hormones in the bloodstream lead to many physiological changes such as elevated heartbeat, heart rate, and blood pressure; an increase in blood glucose levels; dilation of air passages; vasodilation near skeletal muscles and vasoconstriction around digestive organs; and dilation of pupils [26]. This response is referred to as the short-term alarm stage. The endocrine system, on the other hand, is responsible for sustaining the stress response in the long-term resistance stage.

A body of literature suggests that the components of the CRF system regulate several more biological processes other than activation of the HPA axis. For example, in rodents, CRF modulates feeding behavior under conditions of stress [27–29]. The injection of UCN1–3 in rodent brains decreases food intake after 90 min (in food-deprived animals) or 4–5 h (in *ad lib* fed). In these studies, effects were transient and seen at a very high dose [30,31]. All of these studies used only male rats; thus, it is not clear if UCNs have any long-lasting effect on food intake, or in female animals. UCN2 infusion over days also modulated food intake in male mice [32,33]. Thus, nutrient sensing appears to be an ancient function that UCN2/3 and CRF_2 receptors still mediates as shown by others and us [30–34]. Urocortins are known to regulate vascular permeability [35]. In pregnancy, it has profound vasodilatory effects on the utero-placental and feto-placental circulations [36,37]. The placenta, an organ of fetal origin, is key for survival for the developing fetus inside a mother's womb. During pregnancy, the placenta serves as an additional site for CRF synthesis and release [38,39] and CRF serves as a "placental clock" that determines the onset of labor [40,41]. The role of UCN2 and UCN3 in glucose homeostasis and diabetes is emerging [32–34,42,43], whereas CRF_2 , the predominant peripherally expressed receptor, is a key mediator of sexually dimorphic responses in diabetes phenotype [34]. The role of UCN1, UCN2, and CRF_2 is also emerging in several gastrointestinal diseases, as discussed in later sections.

4. Gene Structure and Tissue Distribution of CRF and Urocortins

The human *CRH* gene is located on chromosome 8 and contains two exons. The human *UCN* (or *UCN1*) gene is located on chromosome 2 and contains two exons, and the entirety of the coding region is contained within the second exon. The *UCN2* gene is located on chromosome 3 and contains two exons. The least studied member of the CRF family is *UCN3*; it is located on chromosome 10 and consists of two exons. In mice, the *Crh* gene is located on chromosome 3 and consists of two exons. The *Ucn1* gene is located on chromosome 5 in mice and contains two exons. The *Ucn2* gene is located on chromosome 9 and contains one exon, and the mouse *Ucn3* gene is located on chromosome 13

and contains two exons. The genes for the *CRH* family of peptide hormones evolved after a series of gene duplication events (Figure 1A), and the phylogenetic evolution of CRF receptor paralogs is nicely summarized by Lovejoy et al. [2].

Interestingly, progeny from homozygous *Crh*^{-/-} mice (knockout of *Crh* gene) die within 12 h of birth due to lung dysplasia and atrophy of the adrenal gland [6]. *Crh*^{-/-} mice display marked fetal glucocorticoid deficiency and show impaired, sexually dimorphic stress responses (Table 1) [5]. However, progeny from *Crh*^{-/-} mice can be rescued by exogenous glucocorticoid administration and once rescued, the pups develop into adults with normal growth, fertility, and longevity [5]. Bhargava et al. reported that the PVN-specific elimination of *Crh* using RNAi alters stress-induced levels of ACTH hormone, but it did not affect basal ACTH levels [44], confirming that PVN-specific CRF plays a key role in the initiation of stress responses, but it is redundant for housekeeping functions. In agreement with PVN-specific RNAi findings, hypothalamus-specific *Crh* knockout mice have decreased basal, diurnal, and stress-activated plasma corticosterone secretion as well as basal plasma ACTH levels. In addition, these mice show adrenal atrophy and have noticeably reduced anxiety-like behaviors [45]. Mice overexpressing *Crh*, both in the brain and peripheral tissues (*Crh*-OE or *Crh*-Tg), have increased hypothalamic CRF and circulating corticosterone levels and show a phenotype resembling Cushing's syndrome [46–49]. Taken together, these data suggest that either a lack of or overexpression of CRF hormone results in impaired stress, immune responses, and altered gastrointestinal function [5,15,21,50–52].

Table 1. Phenotypic characteristics of transgenic modifications in genes for components of the CRF family of peptide hormones and receptors.

Genotype	Transgenic Modification	Gross Phenotype and Biological Characteristics
<i>Crh</i> ^{-/-}	Knockout [5]	Progeny from homozygous <i>Crh</i> ^{-/-} die within 12 h after birth due to lung hyperplasia. Can be rescued after intervention with exogenous glucocorticoids. Marked fetal glucocorticoid deficiency, but normal postnatal growth, fertile.
<i>Crh</i> -OE or <i>Crh</i> -Tg	Constitutive overexpression [46]	Elevated basal plasma corticosterone, normal basal ACTH, and HPA axis responses to stress.
<i>Crh</i> -OE or <i>Crh</i> -Tg	Overexpression [49]	Cushing's syndrome-like symptoms. Increased hypothalamic CRF expression, increased circulating corticosterone and adrenocorticotrophic hormone (ACTH). Delayed and attenuated HPA axis responses to stress.
<i>Ucn1</i> ^{-/-}	Knockout [53,54]	Significant auditory deficit and acoustic startle responses in both lines. Increased anxiety-like responses in one line, but not the other. No overt phenotypic abnormality. Fertile.
<i>Ucn2</i> ^{-/-}	Knockout [55,56]	Metabolic phenotype suggesting role in glucose homeostasis. No other overt phenotypic abnormality. Fertile. Female, but not male knockout show increased basal ACTH, corticosterone rhythm, decreased fluid intake, and depression-like behavior.
<i>Ucn3</i> ^{-/-}	Knockout [57]	Regulation of glucose homeostasis. No change in anxiety- or depression-like behaviors. No other gross phenotype. Fertile.
<i>Ucn1</i> ^{-/-} / <i>Ucn2</i> ^{-/-}	Double knockout [58]	Impaired recovery responses to stress. Fertile.
<i>Ucn1</i> ^{-/-} / <i>Ucn2</i> ^{-/-} / <i>Ucn3</i> ^{-/-}	Triple knockout [6]	Reveal urocortins' role in stress recovery; increased anxiety-like behavior in response to stress. May have impaired cardiac and reproductive functions.
<i>Crhr1</i> ^{-/-}	Knockout [59]	Low plasma corticosterone, impaired HPA axis responses, decreased anxiety-like responses. Progeny from heterozygous mice are fertile and without overt phenotype, but male progeny show 15% mortality. Progeny from homozygous female dam die 2 days after birth. Adrenal gland atrophy.
<i>Crhr2</i> ^{-/-}	Knockout [60,61]	Hypertensive, increased anxiety-like behavior, hypersensitive to HPA axis stress responsive, fertile, no other overt phenotype.
<i>Crhr1</i> ^{-/-} / <i>Crhr2</i> ^{-/-}	Double knockout [62]	Sexually dimorphic stress responses: female mice have reduced anxiety-like behavior, whereas male mice show increased anxiety-like behavior.

Two *Ucn1* null (*Ucn1*^{-/-}) mice lines have been developed independently [53,54]. *Ucn1*^{-/-} mice are fertile and do not show any gross overt phenotypic abnormality. Curiously, male progeny from one of the lines display significant increases in anxiety-like behavior [53], whereas the male progeny in the second line do not differ from their wild-type littermates with regards to anxiety-like behavior [54]. Both lines harbor deletion of the coding exon and used identical genetic background (mixed 129S/C57BL6), thereby confounding results and suggesting that either animal housing environment or the sex of the researcher (or both) contribute to anxiety-like behavior [63]. However, both lines of *Ucn1*^{-/-} mice (of both sexes) have significant auditory deficits and acoustic startle responses (Table 1). Plasma membrane Ca²⁺ ATPase pumps (PMCA) are responsible for the extrusion of Ca²⁺ from neurons, glia, and other cell types and are regulated by glucocorticoids as well as stressors [19,20,64]. Mice null for *Pmca1* are embryonic lethal, whereas heterozygous *Pmca1* mice display no overt phenotype [65]. Interestingly, spontaneous mutations in the *Pmca2* gene (A→G transition) resulting in glycine-to-serine substitutions in a highly conserved region of the PMCA2 protein causes deafness and imbalance in mice [66]. In agreement with these findings, mice null for *Pmca2* are deaf and heterozygous mice have significant hearing loss [67]. PMCA1 expression is repressed by glucocorticoids in the hippocampus of male rats [20]. In addition, various stressors, such as restraint and cold stress, repress hippocampal PMCA1 expression [20]. The treatment of cultured hippocampal cells with glucocorticoids repress its expression and function, resulting in high intracellular Ca²⁺ levels [Ca²⁺]_i and neuronal cell death due to faulty Ca²⁺ extrusion mechanisms [19]. Thus, the loss of PMCA function results in elevated [Ca²⁺]_i in the inner ear canals, which contributes to hearing loss in these mice. UCN1 has been shown to modulate Ca²⁺ signaling via the activation of CRF receptors [68,69]. Taken together, these data suggest that Ca²⁺ signaling mediated by PMCA might be defective in the auditory canals of *Ucn1*^{-/-} mice, explaining this phenotype. The elimination of UCN1 expression in the vasculature enhances the capillary leak and migration of immune cells [35], confirming its role in vasoregulation.

Ucn2 null mutant (*Ucn2*^{-/-}) mice are viable and fertile but show a metabolic phenotype suggesting a role for UCN2 in glucose homeostasis [55]. Furthermore, the exogenous administration of UCN2 improves glucose clearance in diabetic male mice and acts as an insulin sensitizer in the skeletal muscles of obese male mice [32,33]. *Ucn3* null mice (*Ucn3*^{-/-}) are also viable and fertile, and similar to UCN2, UCN3 is involved in the regulation of glucose homeostasis [57], as characterized using male mice null for *Ucn3*. Double and triple knockouts of urocortins are also reported [6,58]. These mice are viable (Table 1), but they reveal a key role of urocortins in stress recovery [6]. However, these knockout studies do not report an exhaustive characterization of morphology and function of all organs, and it is possible that urocortins knockouts may have impaired cardiac, liver, and reproductive functions. The readers are also referred to excellent reviews that summarize phenotypes for gene knockouts of the CRF family of peptide hormones and their two receptors [70–72].

Both CRF and UCN1 are secreted as pro-peptide precursors that are 196 and 122 amino acids long, respectively [73]. The mature CRF peptide in humans is a 41-amino acid peptide hormone that is most closely related to UCN1, a 40-amino acid peptide, sharing approximately 45% amino acid identity with human and rat sequences. The human UCN2 has a size of 38 amino acids, sharing 38% amino acid identity with rat CRF and 42% amino acid identity with rat UCN1. The UCN3 consists of 38 amino acids and shares 26% amino acid identity with CRF and 34% amino acid identity with UCN1. Human CRF and all three urocortins also share homology with CRF and urocortins found in all other classes of vertebrata that include amphibian (frog), actinopterygii (zebra fish and teleost), and avian (chicken) (Figure 3).

Amino Acid Sequence Alignment

CRF	[<i>Homo sapiens</i>]	128	SPAALAEK--GAR	--NALGGHQ	-----EAPERERRSEE-	--PPISLDLTPHLLRVELEMARAEQLA	QQAHSNRKLMIEIGK-	196
UCN	[<i>Homo sapiens</i>]	67	GPGRRLGLGT---	-----	-----AGERPRR-DN-	--PPLSIDLTPHLLRVELEMARAEQLA	ERAEQNRIIFDSVGGK-	124
CRF	[<i>Danio rerio</i>]	84	--QLQLTQ-RLL	--EGKVGNI[6]	YALRALDMEKERRRSEE-	--PPISLDLTPHLLRVELEMARAEQMA	QQAHSNRKMEIFPGK-	162
UCN2	[<i>Homo sapiens</i>]	21	PVTFPTQLRPQ[5]	TPRPAASES[5]	TWPAASGMSCPRIHP-	SRIVSLDVPFGLLOILLEQARAAAR	EQATTNARLLARVGHG	112
UCN3	[<i>Homo sapiens</i>]	69	SSEE-----[8]	FPISGARGG[4]	RYRYVSOAQDPGRKPDQT[6]	TKFTLSLDVPTNIMNLFNIAKAKNLR	QQAANARLLMAQIGRR[1]	161
DH44	[<i>D. melanogaster</i>]	24	TAORGAVGAGGAA[2]	SGAAAGGAE[6]	TNGYPLDPTGTRNSODD[6]	NKPSLSIVNPLDVLQRLLLEIARRM[4]	ROVELNRRLLKRVGKR[232]	356
CRF	[<i>Gallus gallus</i>]	99	AAALQGS--GSP	--EGDEGAG	-----EAVEKRRSEE-	--PPISLDLTPHLLRVELEMARAEQLA	QQAHSNRKLMIEIGK-	167
UCN	[<i>Danio rerio</i>]	191	APQQDNSLE----	-----	-----ELSEFSKR-NDD	--PPISLDLTPHLLRVELEMARAEQMA	EQAELNRRKLVDEVGK-	250
UCN3	[<i>Danio rerio</i>]	67	SEEP-----[5]	TPASK----	-----YRYSLOTQLRSKLYRNS[6]	ISQVTLSDVPTNIMNLFNIAKAKNLR	AKADDNARLLAQIGRR[1]	147
UCN3	[<i>Gallus gallus</i>]	67	EEEEEEAEERGG[2]	FPFEG----	-----HYRYTSOAVGVKTYQNR[6]	TKVTLSLDVPTNIMNLFNIAKAKNLR	AKAANARLLMAQIGRR[1]	154
CRF	[<i>Xenopus laevis</i>]	80	AVQQLQAQWSSQ	--PGMRAAS[6]	PYSAQEPDPTKARRAE-	--PPISLDLTPHLLRVELEMARAEQIA	QQAHSNRKLMIDIIIGK-	162
UI	[<i>Oncorhynchus mykiss</i>]	106	APFEGNSLE----	-----	-----EPVELTKR-NDD	--PPISLDLTPHLLRVELEMARAEQMA	EQAELNRRKLVDEVGK-	165
UCN	[<i>Xenopus laevis</i>]	92	TAFQEGIPDESAK	-----	-----GFLFAFSDRLKR-ED	--PPISLDLTPHLLRVELEMARAEQIA	QQAHSNRKLVDEVGK-	158
UCN3	[<i>Xenopus laevis</i>]	67	QEEMDEEDEKKEK[2]	FPQAAA----	-----RYRYLSOAVGVKTYQNR[6]	TKFTLSLDVPTNIMNLFNIAKAKNLR	AKAANARLLMAQIGRR[1]	154
CRF	[<i>Xenopus laevis</i>]	85	AVQQLQAQWSSQ	--PGQRAAA[6]	PYSAQEPDPTKARRAE-	--PPISLDLTPHLLRVELEMARAEQIA	QQAHSNRKLMIDIIIGK-	167
UCN3	[<i>Latimeria chalumnae</i>]	70	QPEEGQEEVVRGK[8]	FPAS----	-----HYRYLSOAVGVKTYQNR[6]	TKFTLSLDVPTNIMNLFNIAKAKNLR	AKAANARLLMAQIGRR[1]	162
UCN3	[<i>E. electricus</i>]	59	SQES-----[5]	LPESN----	-----YKFLSOTLLRSKMYRNS[6]	NQVTLSDVPTNIMNLFNIAKAKNLR	AKAANARLLAQIGRR[1]	139
UCN3	[<i>O. anatinus</i>]	69	WEREEGEEEDG-[7]	LPSSGRDGA[6]	HSRLLSQALWFKRRHPSK[6]	TKVTLSLDIPTSIMVLFMTAKEQSLQ	DKANARLLMAQIGRR[1]	170

				Accession #
CRF	[<i>Homo sapiens</i>]	183	QQAHSNRKLMIEIGK-	196 NP_000747.1
UCN	[<i>Homo sapiens</i>]	111	RAEQNRIIFDSVGGK-	124 NP_003344.1
CRF	[<i>Danio rerio</i>]	149	QQAHSNRKMEIFPGK-	162 NP_001007380.1
UCN2	[<i>Homo sapiens</i>]	98	QATTNARLLARVGHG	112 NP_149976.1
UCN3	[<i>Homo sapiens</i>]	146	QQAANARLLMAQIGRR[1]	161 NP_444277.2
DH44	[<i>Drosophila melanogaster</i>]	110	QVELNRRLLKRVGKR[232]	356 NP_649922.2
CRF	[<i>Gallus gallus</i>]	154	QQAHSNRKLMIEIGK-	167 NP_001116503.1
UCN	[<i>Danio rerio</i>]	237	QQAELNRRKLVDEVGK-	250 XP_005160887.2
UCN3	[<i>Danio rerio</i>]	132	KADDNARLLAQIGRR[1]	147 NP_001076423.1
UCN3	[<i>Gallus gallus</i>]	139	KAANARLLMAQIGRR[1]	154 XP_001231711.1
CRF	[<i>Xenopus laevis</i>]	149	QQAHSNRKLMIDIIIGK-	162 NP_001165681.1
UI	[<i>Oncorhynchus mykiss</i>]	152	QQAELNRRKLVDEVGK-	165 NP_001117815.1
UCN	[<i>Xenopus laevis</i>]	145	QQAQNRIIFDSVGGK-	158 NP_001086429.1
UCN3	[<i>Xenopus laevis</i>]	139	KAANARLLMAQIGRR[1]	154 XP_017948238.1
CRF	[<i>Xenopus laevis</i>]	154	QQAHSNRKLMIDIIIGK-	167 NP_001165680.1
UCN3	[<i>Latimeria chalumnae</i>]	147	KAANARLLMAQIGRR[1]	162 XP_005997394.1
UCN3	[<i>Electrophorus electricus</i>]	124	KAANARLLAQIGRR[1]	139 XP_026884884.1
UCN3	[<i>Ornithorhynchus anatinus</i>]	155	KANARLLMAQIGRR[1]	170 XP_007655165.1

Figure 3. A comparison of examples of primary amino acid sequence from each group of CRF and urocortin precursor peptides found in the Protein data bank and generated using the multiple alignment tool from NCBI Blast. Red and blue are alignment columns with no gaps, whereas grey residues are with gaps. Highly conserved amino acids are shown in red and less conserved amino acids are shown in blue based on the residues' relative entropy thresholds.

CRF is found in the parvocellular PVN of the hypothalamus, central nucleus of the amygdala, the bed nucleus of the stria terminalis (BNST), and the hindbrain regions in the central nervous system, amongst other regions [1–3]. The medial BNST regulates sexual behavior, whereas the lateral BNST regulates stress-related functions. In rats, no differences between the sexes are seen in the distribution of immunoreactive (IR) vasoactive intestinal polypeptide neurons that are in close proximity to CRF-IR neurons [74]. In male rats, lesions of BNST significantly decrease CRF mRNA expression in the medial parvocellular PVN [75]. When exposed to a myriad of stressors, CRF expression shows sexually dimorphic responses in the rodent brains. CRF expression in the amygdala, but not BNST or the hippocampus, differs between male and female mice exposed to predator odor [76]. However, in heterozygous knock-in mice for glutamic acid decarboxylase (GAD67), the enzyme responsible for the conversion of glutamate to GABA, exposure to predator order increases CRF expression in the BNST of male but not female mice compared with mice exposed to control odor [76]. In response to acute restraint stress exposure, CRF mRNA expression increases in the BNST of male, but not female rats, whereas transcription factor c-fos mRNA expression increases in female rats alone [77]. Exposure to chronic variable mild stress results in increases in total CpG (cytosine-guanine nucleotides) methylation of *Crh* gene in female but not male rats [78].

In the periphery, CRF is expressed in the various regions of the gut, including the enteric neurons, skin, adrenal glands, and the placenta [2,3]. While CRF expression is reported in several peripheral tissues, it remains unclear if CRF is synthesized locally in peripheral cells or is transported from the brain via circulation. RNAi experiments demonstrated for the first time that the knockdown of CRF in gut tissue severely decreased the expression of CRF-IR in the enteric neurons and the mucosa [8], suggesting local transcription and translation in these cell types. The gut-specific knockdown of CRF also prevents toxin-induced inflammation and demonstrates a role for locally transcribed and synthesized CRF and UCN2 in the regulation of both basal and stress-induced gut motility [8,9]. UCN1 is predominantly expressed in the Edinger Westphal and the supraoptic nuclei in the brain. In addition, UCN1-IR is also reported in the BNST [79]. In response to maternal separation, UCN-IR neurons show sexually dimorphic expression of c-Fos that is concomitant with dampening of the corticosterone responses in female but not male rats [79]. In the periphery, UCN1 is found in the

intestine, cardiac myocytes, thymus, skin, spleen, and immune cells. UCN2 is expressed in the hypothalamus, brainstem, and spinal cord. In the periphery, it is found along the gastrointestinal tract, heart, blood cells, and adrenal gland [80–82]. UCN3 is found in the hypothalamus and amygdala, the intestine, and the pancreas [81,83–85]. Urocortins are distributed throughout the neuroaxis and peripheral tissues [7,8,33,36,42,61,80,83,86–93] with UCN1 being abundantly expressed in early and late gestational tissues [94–96].

5. Gene Structure and Tissue Distribution of CRF Receptors

CRF receptors also evolved via a series of unknown duplication events [1,2] and belong to the class B family of GPCRs (Figure 1C). CRF₁ and CRF₂ are products of two separate genes [97,98] and have several splice variants expressed in various central and peripheral tissues. In humans, CRF₁ is encoded by the *CRHR1* gene, which contains 14 exons across 20 kb and is located on the long arm of chromosome 17. The entire length of *CRHR1* is approximately 51.55 kb. The entire gene product is termed CRF₁-β, which is a 444-amino acid protein receptor that is involved in signaling properties and impaired agonist binding [97]. Removing exon 6—a section of *CRHR1* that contains the information for a 29-amino acid peptide—from the mRNA results in the translation of CRF₁-α, which is a 415-amino acid long receptor that is the predominant functional receptor isoform. CRF₁-α has high binding affinities for CRF and UCN1, facilitating their actions, and it is expressed in specific central and peripheral tissues throughout the body (Figure 2). CRF₁ is also known to have six other subtypes, as a result of extensive alternative splicing; these are classified “c–h” and are present in humans and rodents, with several subtypes detailed to be nonfunctional. The subtypes all have a deletion of exon 6 and one or more exon excision, with the exception of subtype h, which has an insertion of a cryptic exon in the space separating exon 5 and 6. In mice, 13 exons are reported for the *Crhr1* gene. CRF₁ knockout mice (*Crhr1*^{−/−}) are known to display chronic glucocorticoid deficiency as a result of interference with the HPA axis [59]. In addition, *Crhr1*^{−/−} mice exhibit reduced anxiety-related behavior. Whereas heterozygous *Crhr1* mice are fertile, progeny from the homozygous *Crhr1*^{−/−} cross die within 48 h after birth with lung dysplasia [59]. The promoter region of the *Crhr1* gene harbors several binding sites for transcription factors such as SP1 and AP1 and hormone response elements that bind glucocorticoid, progesterone, and estrogen receptors [99].

CRF₂, which is encoded by the *CRHR2* gene, has three variants: α, β, and γ. The human *CRHR2* gene is 50 kb in length and contains 15 exons. Whereas CRF₂-α and CRF₂-β are found in both humans and rodents, CRF₂-γ is only found in the limbic region of the human brain. The three functional subtype mRNAs are made by utilizing an alternate 5′ exon 1 that connects onto a common set of exons. α, β, and γ variants have the same C-terminus and transmembrane domains [100]. The differences emerge in their N-terminal extracellular domains. CRF₂-α has 34 amino acids at the N terminus and CRF₂-β has 61 amino acids at the N terminus, whereas CRF₂-γ has only 20. In summary, the α variant is a 411-amino acid receptor, the β variant is a 431-amino acid receptor, and the γ variant is a 397-amino acid receptor [101,102]. CRF₂-γ shows no significant homology to the other two variants [101]. The three functional subtypes of CRF₂ all contain different promoters and splicing control regulation [100]. Moreover, the three variants’ diverse expression is connected to the regulation of expression in distinct tissues. Two independent lines for CRF₂ knockout mice (*Crhr2*^{−/−}) have been generated [60,61]. Male *Crhr2*^{−/−} mice are hypertensive, have increased ACTH and glucocorticoid release, i.e., are hypersensitive to the HPA axis-mediated stress responses, and show increased anxiety-like behavior [60,61]. The promoter region of *Crhr2* gene also harbors several binding sites for SP1 as well as for glucocorticoid and estrogen receptors [100,103]. *Crhr1/Crhr2* double knockout mice display sexually dimorphic HPA axis stress responses (Table 1): female mice have decreased anxiety-like behavioral responses, whereas male mice show increased anxiety-like behavior [62].

CRF₁ and CRF₂ share 70% amino acid sequence similarity overall and nearly 80% identity in their transmembrane and intracellular domains. However, at the amino acid level, CRF₁ and CRF₂ exhibit low homology at the N-terminus: approximately 47% [100]. CRF receptors evolved from an

ancestral GPCR *seb-2* that is found in worms, including *C. elegans* (Figure 4). The receptors diverged after a gene duplication event to diuretic hormone (DH) receptors. Two DH44 receptors are reported in the fruit fly, *Drosophila melanogaster*, and other orthologs are found throughout the phyla (Figure 1C). Amino acid sequence alignment reveals a high sequence identity between DH44 receptors CRF₁ and CRF₂ (Figure 4). The two receptors are generally distinguished by their location in the central nervous system [2] and peripheral nervous system, in addition to their ability to differentially bind UCN1-3 and CRF [104]. CRF₁ protein expression is sparse and not sexually dimorphic at postnatal days 0–21 in the PVN, but adult male mice show higher CRF₁ levels in the PVN than female mice [105]. In rats, the sexually dimorphic expression of CRF receptors is seen in several brain regions with no two brain areas being alike [106]. CRF₁ binding in various brain regions is similar in both male and female juvenile rats, but it is higher in the cingulate cortex of adult female rats compared with male rats. In contrast, CRF₂ binding is greater in the BNST of adult male compared with female rats, irrespective of age [106].

Gonadectomy in young mice (6 weeks old) results in a significant decrease in CRF₁-IR cells in the PVN of male but not female mice. CRF₁ cells co-express estrogen receptor alpha (ER α) and androgen receptors, suggesting that circulating gonadal sex hormones differentially regulate CRF₁ expression and function. Restraint stress induces a sexually dimorphic pattern of neural activation in PVN with male mice showing higher CRF₁-IR compared with female mice [105]. In gonadectomized young adult male mice, treatment with an androgen receptor agonist increases the expression of CRF₂ mRNA expression in various limbic regions and the lateral septum [103]. Thus, androgens might dampen stress responses and reduce anxiety-like behaviors in males of a species via mechanisms that involve increasing the expression of CRF₂. In monogamous prairie voles, significantly higher CRF₂ binding is reported in the BNST of males compared with females, suggesting a role for CRF₂ in pair bonding [107]. The evolution, expression, distribution, and functions of CRF, urocortins, and their receptors are addressed in several excellent reviews and the references therein [1–3,15,52,70–72].

Accession #	Amino Acid Sequence Alignment	
NP_004373.2	1	M--GGHPQLRLVKALL--- --LLG-----LNPVSASLQD-QHC
NP_001874.2	1	M-----DAA----- --LHsllEancSLALAEELLDDGWG
XP_696346.3	1	MSRILHPQVLTWITFLIS RAT[6]ALLLL-----STNLARTLI-LWN
NP_989652.1	1	MVPGPRPALLLLFLQ- --- AFLLW-----DSPVAASIQE-QYC
XP_015136531.1	1	M-----AGRAPQRSLPA AAT[1]ALLLLpivLHCesSVNLPLSVLLEKYC
NP_001122749.1	1	MNPSISTAGAVVDDPQMS [11]ESF[6]ACFYFf1mLVEhyKDRYVASLCEDTFG [25]EFALNSS [5]NH [20]AA [17]
XP_005994697.1	1	MYT----- --TI NI TG 9
XP_002667894.2	1	M-----DASLQPFLEE FGD -----LNC-----TLLDAFQ
AAF58250.1	1	MSDHHN-----IDSVNA SGS [3]-----LdLhnlDGIgesVLEQLCLV
AAF58501.4	1	MA---DDDLRALVDSLDD ASQ [3]AKVIANfsVDMlqRASALIGAQQGSSG
NP_001079049.1	1	MLLAKTPC---LLLVQ- --- VIAAG-----ISFALTSLQD-Q-C
NP_004373.2	42	LQCNASVDLIGTCWPRSAGQLVVRPCPAFFYG---VRYNTTNNGYRECLANGSWA
NP_001874.2	39	-YCNTTLDQIGTCWPRSAGALVERPCPEYFNG---VKYNTTRNAYRECLENGTTWA
XP_696346.3	57	LFCNTSIDGIGTCWPRSAGEVVSRPCPEFLG---VRYNTTNNVYRECLANGTTWA
NP_989652.1	47	PQCNASVDLIGTCWPRSAGVGLVARPCPEYFG---VRYNTTNNGYRECLANGSWA
XP_015136531.1	55	LVCNATDQIGTCWPRSAGKLVVRPCPEFFNG---IKYNTTKNAYRECLANGTTWA
NP_001122749.1	144	SPCPPTDNGW-NCFDSATPG-VVFKQCPNYLYGgsnlKTYDYLRSQKVCRSNGWAT [8]BHTDYTCGMSNGDV-----
XP_005994697.1	10	PYCNASVDLIGTCWPRSAGVLLIARPCPEYFNG---VRYNTTNNIYRECFPNGTTWA
XP_002667894.2	42	VYCNATDDEIGTCWPRSNSGRIERPCPEYFNG---VKYNTTRIAAYRECLENGTTWA
AAF58250.1	52	-HCLTQFDSI-LCWPRTAGTLAVLQCMDELQG---IHYDSSKNATRFCHANGTWE
AAF58501.4	82	LQCPSSFDSV-LCWPRTAGSLAVLPCPEEFGK---VHYDITDNTATRFCHANGTWD
NP_001079049.1	42	LACNASIDMIGTCWPRTAGQVVARPCPEYFNG---VQYNTTGNVYRECHLNGSWA
NP_004373.2	113	KVHYHVAVIINYLGHGISLVALLVAFVFLRLRSIRCLRN-IHWNLIS
NP_001874.2	110	DLHYRIALVVNYLGHCVSVAALVAAPFLFLALRSIRCLRN-VIHWNLIT
XP_696346.3	128	KLHYHIAVIINYLGHGISLVALLVAFVFLMRLRSIRCLRN-IHWNLIT
NP_989652.1	118	KLHYHIAVIINYLGHCVSGLTLLVAFVFLMRLRSIRCLRN-IHWNLIT
XP_015136531.1	126	AHHYKIALIINYLGHGISVGLIIVAFVFLMRLRSIRCLRN-VIHWNLIT
NP_001122749.1	220	---EARILAGLLTYSASVIFLIPAVFLMRLRSIRCLRN-IHWNLIT [6]AFYLIITVSLFVVDNADPLSSQVQNHLLP
XP_005994697.1	81	KLHYHIAVIINYLGHGISLVALLVAFVFLMRLRSIRCLRN-IHWNLIT
XP_002667894.2	113	PMHYKIALIINYLGHCVSVALIIVAFVFLMRLRSIRCLRN-IHWNLIT
AAF58250.1	123	EVIVLEPTIIYYIGYTLSSLVLSLALIVFAYFKELRCLRN-TIHANLFF
AAF58501.4	155	SPNVELPAIIYAGGYFLSFAITLVVALIIFLSPKDLRCLRN-TIHANLFL
NP_001079049.1	113	KVHYHIAVINFLGHGISLVALLVAFVFLMRLRSIRCLRN-IHWNLIT
NP_004373.2	189	RLVTA--YNYFVHTNFFWMPGEGCYLHTAIVLTYSDTKLRKWMFICIGWVFPPIIIVAWAIGKLY---YDNE
NP_001874.2	185	RCITTI--FNYFVHTNFFWMPVEGCVLHTAIVMTYSTERLRKCLFLFIGWCIPFPIIIVAWAIGKLY---YENE
XP_696346.3	204	RLVTA--YNYFVHTNFFWMPGEGCYLHTAIVLTYSDTKLRKWMFICIGWVFPPIIIVAWAIGKLY---YDNE
NP_989652.1	194	RLVTA--YNYFVHTNFFWMPGEGCVLHTAIVLTYSDTKLRKWMFICIGWVFPPIIIVAWAIGKLY---YDNE
XP_015136531.1	201	RCITTV--YNYFVHTNFFWMPVEGCVLHTAIVMTYSDTKLRKWMFIFIGWCIPCIIVAWAIGKLY---YENE
NP_001122749.1	299	RLVTSIQLRVLRLTNFTWMLAEAVLWRLHTAQHSEGETLRSYKVICWGVPGVTVVYIFVRSLL---NDV [2]-CWIE
XP_005994697.1	157	RLVTA--YNYFVHTNFFWMPGEGCYLHTAIVLTYSDTKLRKWMFICIGWVFPPIIIVAWAIGKLY---YDNE
XP_002667894.2	188	RLITTI--YNYFVHTNFFWMPVEGCVLHTAIVMTYSDTKLRKWMFIFIGWCIPCIIVAWAIGKLY---YENE
AAF58250.1	197	IALLITL--FHFFLTNFFWMLVEGLYLYMLVVKVTSFGDNLRFNIYASIGWGGPALFVVTWAVAKSLvtvYSTP [5]NCPWM
AAF58501.4	230	ITLVIL--PQFYVLTNFFWMPVEGLYLYLTVLVTQSSDNISFIIYALIGWGPCAVCILVWSIAKAFaphLENE [7]DCAM
NP_001079049.1	189	RLVTA--HNYFVHTNFFWMPGEGCYLHTAIVLTYSDTKLRKWMFICIGWVFPPIIIVAWAIGKLY---YDNE
NP_004373.2	262	KRPGVYTDYIYQGPMLVLLINIFLFLNIVRILMTKLRASSTSETIYQRKAVKATLVLLPLLGITYMLFFVNPGEDEVSR
NP_001874.2	258	KEPGDLVDYIYQGPMLVLLINIFLFLNIVRILMTKLRASSTSETIYQRKAVKATLVLLPLLGITYMLFFVNPGEDDLQ
XP_696346.3	277	KRAGIYTDYIYQGPMLVLLINIFLFLNIVRILMTKLRASSTSETIYQRKAVKATLVLLPLLGITYMLFFVNPGEDEISQ
NP_989652.1	267	KRAGVYTDYIYQGPMLVLLINIFLFLNIVRILMTKLRASSTSETIYQRKAVKATLVLLPLLGITYMLFFVNPGEDEISR
XP_015136531.1	274	KEPGKYIDYIYQGPVILVLLINIFLFLNIVRILMTKLRASSTSETIYQRKAVKATLVLLPLLGITYMLFFVNPGEDEISQ
NP_001122749.1	374	NTVAVIEWMIITPSSLAMGVNLLGLLIVLIVLKKLRCDPHEIRIYQRKAVGALMLIPVFGVQOLTI----YRFSN
XP_005994697.1	230	KRPGVYTDYIYQGPMLVLLINIFLFLNIVRILMTKLRASSTSETIYQRKAVKATLVLLPLLGITYMLFFVNPGEDEVSR
XP_002667894.2	261	KEPGKYVDYIYQGPVILVLLINIFLFLNIVRILMTKLRASSTSETIYQRKAVKATLVLLPLLGITYMLFFVNPGEDEISQ
AAF58250.1	278	QET--HVDWYIYQGPVAVLIINLTFLLRMWVLTIKLRSANTVETQYRKAALKLVLLPLFGITYLVLAGPSESGLMG
AAF58501.4	313	RES---HIDWIFKVPASLALLVNLVFLIRIMWVLTIKLRSANTLETRQYKASKALLVLLPLFGITYLVLLVLTGP--EQGISR
NP_001079049.1	262	KKAGVYTDYIYQGPVILVLLINIFLFLNIVRILMTKLRASSTSETIYQRKAVKATLVLLPLLGITYMLFFVNPGEDEISR
NP_004373.2	342	VVFIYFNSFLESFQGFVSVFYCFNLSEVR SAIRKRWRHQDKHSIRARVARAMSI P TSPTRVFSHSI
NP_001874.2	338	IMFIYFNSFLESFQGFVSVFYCFNVEVR SAVRKRWRHQDHSIRVPMARAMSI P TSPTRISFHSI
XP_696346.3	357	IVFIYFNSFLESFQGFVSVFYCFNLSEVR SAVRKRWRHQDKHSIRARVARAMSI P TSPTRVFSHSI
NP_989652.1	347	IVFIYFNSFLESFQGFVSVFYCFNLSEVR SAVRKRWRHQDKHSIRARVARAMSI P TSPTRVFSHSI
XP_015136531.1	354	IVFIYFNSFLESFQGFVSVFYCFNLSEVR SAARKRWRHQDHSIRVVRARAMSI P TSPTRISFHSI
NP_001122749.1	449	TYVQVTDQSLNGLQGMFVSFIVCYTNRSVV ECVLKFWSSHQEKRALGAECRQMSI [2]- -SGKLLRPPQ
XP_005994697.1	310	IVFIYFNSFLESFQGFVSVFYCFNLSEVR SAVRKRWRHQDKHSIRARVARAMSI P TSPTRVFSHSI
XP_002667894.2	341	IVFIYFNSFLESFQGFVSVFYCFNLSEVR SAARKRWRHQDHSIRARVARAMSI P TSPTRISFHSI
AAF58250.1	356	HMFVAVLRAVLLSTQGFVSVLYFYCFNLSEVR NALRHHITWRDTRTQLNQNRYTT [19]S [14]TTTTLVQOHP
AAF58501.4	390	NLFFAIRAFLLISTQGFVSVLYFYCFNLSEVR [19]SSIKNRHRHASKDYSLSRSTESLRT S TSPITPGHYE
NP_001079049.1	342	IVFIYFNSFLESFQGFVSVFYCFNLSEVR SAVRKRWRHQDKHSIRARVARAMSI P TSPTRISFHSI
NP_004373.2	410	K QSTAV 415 CRF ₁ [Homo sapiens]
NP_001874.2	406	K QTAAV 411 CRF ₂ [Homo sapiens]
XP_696346.3	425	K QSSAV 430 CRF ₁ [Danio rerio]
NP_989652.1	415	K QSSAV 420 CRF ₁ [Gallus gallus]
XP_015136531.1	422	K QTAAV 427 CRF ₂ [Gallus gallus]
NP_001122749.1	517	T BHHVL 522 GPCR seb-2 [Caenorhabditis elegans]
XP_005994697.1	378	K QSTAV 383 CRF ₂ [Latimeria chalumnae]
XP_002667894.2	409	K QTTAV 414 CRF ₂ [Danio rerio]
AAF58250.1	457	L [42]EENSV 504 DH-44R1 [Drosophila melanogaster]
AAF58501.4	-	- - - - - DH-44R2 [Drosophila melanogaster]
NP_001079049.1	410	K QSSAI 415 CRF ₁ [Xenopus laevis]

Figure 4. A comparison of examples of primary amino acid sequence from CRF receptors and their closely related GPCRs, seb-2, and diuretic hormone 44 receptors found in the Protein data bank and generated using the multiple alignment tool from NCBI Blast. Red and blue are alignment columns with no gaps, whereas grey residue are aligned with gaps. Highly conserved amino acids are shown in red, and less conserved are shown in blue based on the residues' relative entropy thresholds.

6. Agonist-Induced CRF Receptor-Regulatory Protein Supercomplexes

In vitro ligand-binding studies reveal that CRF binds with high affinity to CRF₁ and has very low affinity for CRF₂ [104]. UCN1 binds with 10–40-fold higher affinity to CRF₁ than CRF and shows equal binding affinities for both CRF₁ and CRF₂ (Figure 2). UCN2 and UCN3 bind exclusively to CRF₂, with UCN3 exhibiting a higher affinity than UCN2 [104,108]. CRF₁ binds with higher affinity to UCN1 than CRF; however, this binding affinity does not translate to signaling efficacy [68]. Meanwhile, CRF₁ does not internalize and signal when stimulated with UCN2 or UCN3, whereas, in the presence of CRF₂, the two receptors heteromerize, internalize, and signal in response to these ligands [69].

The current ligand–receptor complex model is one in which the C-terminal domain of CRF binds to the N-terminal domain of the CRF₁ receptor. The N-terminus can then bind to the J-domain formed by the seven-transmembrane receptor that couples to various G proteins to signal [109,110]. The receptors' ligand selectivity is induced by the presence of a single nucleotide difference in the extracellular N-terminal domains of the receptors CRF₁ and CRF₂ [108]. The agonist selectivity of CRF₁ and CRF₂ can be attributed to the variance in homology at the N-terminus, which is involved in ligand binding [108] as well as differences in amino acid residues within the four agonists [104]. When exposed to excessive concentrations of agonist or ligand, CRF₁ receptor internalization and desensitization occurs, which is a response that is common to most GPCRs. G-protein coupled receptor kinase 3 phosphorylates the CRF₁ receptor, which is followed by the β arrestin2 protein trafficking the CRF₁ receptor from the membrane into the cell via endocytosis [111], although the association of CRF receptors with β arrestins is very transient and unlike that of class B GPCRs [68]. This process represents the internalization of the CRF₁ receptors, which are later marked for degradation or sent back to the plasma membrane i.e., recycled. CRF₁ receptors traffic and signal differentially when bound to CRF versus UCN1 [68]. When bound to CRF, CRF₁ receptors endocytose and recycle to the cell surface via fast recycling Rabs and are resensitized within 2–4 h of ligand stimulation. In contrast, UCN1-bound CRF₁ receptors traffic via slower recycling Rabs, and approximately 20% of the receptors are targeted for degradation. Furthermore, endothelin-converting enzyme 1 is involved in the dissociation of UCN1-bound CRF₁ receptors that couple to G_q but not to G_s [68].

Both CRF receptors harbor either pseudo or cleavable signal peptides [69,112–116]. These signal peptides are probably cleaved in a cell- or context-specific manner. For example, the signal peptide of CRF₁ is reportedly cleaved in one study, but the N-terminal tag of CRF₁ is not cleaved in others. CRF₂- α is shown to harbor a pseudo signal peptide that can be converted to a cleavable one [113], whereas CRF₂- β harbors a cleavable signal peptide [69]. Trafficking and signaling of CRF receptors has been described in several excellent reviews previously [109,110].

Upon ligand stimulation, GPCRs or associated proteins are post-translationally modified and can exist as super complexes in lipid rafts [117]. CRF₂ binds at least four different ligands in its N-terminal extracellular domain and can potentially bind other ancillary proteins via its C-terminal PDZ-domain or intracellular 3 (i3) loop. Several cytoskeleton-associated proteins interact with the i3 loop of GPCRs including F-actin and filamin A [118–120]. Yeast two-hybrid screens have been extensively used to identify such interactions, but a major caveat of such a screen is that it detects only binary interactions between two binding partners. CRF₂ interacts with F-actin and uses the F-actin cytoskeleton to traffic from the endoplasmic reticulum to the cell surface [69]. In conditions where the polymerization of F-actin was prevented, CRF₂ receptors fail to reach the plasma membrane, whereas CRF₁ receptor trafficking to the plasma membrane remains unaffected, suggesting that the two receptors use divergent pathways to travel to the cell membrane from internal compartments [69].

CRF₁ can form homo-oligomers at the plasma membrane and in intracellular compartments [116,121]. However, as a result of pseudo signal peptide existence in the receptor, CRF₂- α acts as a monomer [116]. The configuration of heteromers of CRF receptors with other GPCRs has been suggested to regulate the integration of various systems, such as HPA activation and drug abuse, for instance. Direct allosteric interaction among CRF₁ and vasotocin VT2 receptor (VT2R) receptors is suggested in studies, using heterodimerization as a way to explain CRF potentiation by arginine vasotocin [122]. CRF₁ is known

to hetero(di)merize with vasopressin receptor V1b to mediate synergistic actions of vasopressin and CRF [123]. As a heteromeric partner of serotonin receptor, 5HT₂R, CRF₁ responds to serotonin, but not CRF, to signal via IP3 [124]. CRF modulatory action in the ventral tegmental area has been linked to the CRF₁ activation of certain signaling pathways (phospholipase C, protein kinase A and C (PLC, PKA, and PKC)) in addiction studies and evidence of CRF₂- α and dopamine receptor 1 interaction in HEK293T cells provides corroboration of joint action between dopamine neurotransmitter and CRF in neurons [121]. The two CRF receptors have the ability to form heteromeric complexes in association with regulatory proteins. CRF₁ and CRF₂- β can combine with other proteins to create heteromeric complexes in HEK293 cells co-expressing both receptors and in vivo in the pancreases of male mice [69]. These heteromeric complexes provide insight into the diverse and nuanced function achieved by members of the CRF system.

The sex-specific trafficking of CRF₁ receptors is reported in one study. This study used brain slices to show that in response to swim stress, male rats have higher CRF₁ receptor internalization and association with β arrestin2, whereas female rats do not [125]. In contrast to male rats, stressed female rats have a higher proportion of receptors on the plasma membrane of neurons in the locus-coeruleus (LC)-norepinephrine system than the unstressed rats, indicating the opposite trafficking pattern in females. The LC-norepinephrine system is the part of the brain thought to be in charge of regulating hyperarousal in patients with stress-related psychiatric disorders. A probable explanation for these sex-specific trafficking directions in internalization suggests differences in the relationship between CRF₁ receptors and β arrestin2 [111,125]. Thus, studies addressing the sex and context-specific trafficking and signaling of CRF receptors will help in understanding the differential and nuanced effect of this system in a myriad of diseases that range from psychiatric to cardiovascular and from gastrointestinal and metabolic to reproductive diseases.

7. Sex-Specific Responses Mediated by Urocortins in Health and Disease

Urocortins are regulatory factors of the cardiovascular system, and the effects of UCNs in the periphery are largely mediated by the activation of CRF₂. Studies suggest that the urocortins have the following effect on the cardiovascular system: vasodilation, cardioprotection against ischemia–reperfusion injury, and positive inotropic and lusitropic effects. Understanding the roles and mechanisms of UCNs are significant since they can be targets for treatment modalities for coronary heart disease, hypertension, and congestive heart failure [83,86,88,91,126–129]. UCNs have positive effects in the treatment of heart failure. UCN2 when injected peripherally acting via CRF₂ raises left ventricular ejection fraction and cardiac output levels in the treatment of heart failure patients, corresponding with a decrease in vascular resistance [129]. UCNs also protect against ischemia as a result of a decrease in infarct size [88], as well as amelioration of the ventricular function by lessening myocardial damage and obstructing apoptosis pathways [127]. The pre-conditioning and post-conditioning of neonatal rat cardiomyocytes with UCN1 after a period of ischemia–reperfusion injury decreased apoptosis and necrosis, suggesting that UCN1 exerts cardioprotective effects [130].

The findings from a study that ascertained the effects of UCN2 in male rats with heart failure reveal that increased CRF₂ receptor expression in the heart is correlated with a higher survival rate, without aggravating stress-related behavior [128]. The results from this aforementioned study are consistent with other studies showing that UCN2 and UCN1 in ventricular myocytes increase the rate and effectiveness of cardiac myocyte contractility [131,132]. UCNs serve to enhance cardiac myocyte contractility, since UCNs increase the concentration of intracellular calcium ions [129,132]. An increased concentration of Ca²⁺ in the cytosol of cardiac myocytes results in increased contraction of the heart. When CRF₂ is inhibited, the inotropic and lusitropic effects of UCNs are removed. UCN2, the cognate ligand for CRF₂ is known to improve cardiac function and recover the amplitude of Ca²⁺ transients by inhibiting the expression and interaction between store-operated Ca²⁺ channels, such as Ora1, and TRPC5 [133].

The sex differences in coronary risk and treatment are well documented [13], yet the biological mechanism is poorly understood. In a population-based prospective study with a sample size of 34,000 people, with a roughly even distribution of men and women, it was found that men are twice as likely as women to have a myocardial infarction, even after accounting for traditional risk factors such as blood pressure, serum lipid levels, and body mass index, among others [134]. The authors conclude that given the minor changes in incidence risk in myocardial infarction between the pre- and postmenopausal stage in women, with women still at lower incidence than men, it is unlikely that changes in female hormone levels influence the risk of myocardial infarction [134].

The differences in CRF receptor signaling and trafficking between men and women could partly explain how biological sex affects cardiovascular health and other diseases in which these receptors and their ligands are involved. Irrespective of the tissue/organ, CRF₂ expression appears to be protective and beneficial, whereas loss or inability to increase CRF₂ expression leads to pathology. Should the effect of UCNs be sex-specific in cardiovascular tissue and CRF receptors be upregulated in premenopausal or cycling females compared to males, then sexual dimorphism in the CRF system may have significant implications. A broader understanding of the sex differences regarding CRF₂ and the urocortins in the cardiovascular system and experiments that include female subjects are crucial to uncovering the underlying mechanisms.

8. Sex-Specific Differences in Psychiatric and Related Disorders

Psychiatric disorders such as post-traumatic stress disorder (PTSD), depression, and anxiety both have common origins in disrupted HPA axis and corticolimbic circuits. The hypersecretion of CRF and disruptions in processes downstream from the initial CRF are characteristic of many psychiatric disorders. The HPA axis is regulated by the CRF pathway, which when disturbed can affect how the stress response is both initiated and received. In humans, ACTH levels are increased further in women than in men, suggesting greater susceptibility to CRF. Therefore, the manipulation of CRF levels distresses the HPA axis more in women than in men [135]. The LC-norepinephrine system mentioned earlier is another target of the CRF pathway, and sex differences in LC neurons suggest that there are important considerations to be made in the treatment of stress-related psychiatric disorders. Patients suffering from psychiatric disorders such as PTSD and anxiety disorders experience hyperarousal, which is mediated by the LC-norepinephrine system. This system is often activated along with the HPA axis and at least partially regulated by CRF [136–138]. Animal studies show that female rats have more LC neurons than male rats, which could play a role in the differences in the manifestation of stress-related disorders [139,140]. LC dendrites appear to be more complex and extensive in female rats, with morphological analysis indicating longer and further radiating LC dendritic trees in females compared to males [141]. With denser, branching, and more complex dendrites, female rodents, and by extrapolation, human females are probably better equipped to process more emotional stimuli and initiate a greater arousal response.

CRF administration in the brain promotes grooming (anxiety-like relieving behavior) in both sexes, but at a greater magnitude in female than male mice. This discrepancy is greatest when ovarian hormones were highest in female mice, indicating a possible relationship between hormone levels and CRF-induced activation of grooming [142]. In another study that induced CRF overexpression in early development in mice, both sexes display anxiety-related behavior in adulthood, with the behavior being more pronounced in female mice. When stimulated with a secondary stressor in adulthood, female mice show avoidance regardless of history of CRF overexpression, whereas only male mice affected by CRF overexpression earlier show significant avoidance [143]. Studies show that neurons in the Edinger–Westphal nucleus co-express estrogen receptor (ER β) and UCN1 immunoreactivity in both rat and mouse tissues [143]. mRNA levels of UCN1 in the Edinger–Westphal nucleus of male mice are approximately 10 times higher than in the Edinger–Westphal nucleus of female mice in diestrus and 1.6 times higher than in female mice in proestrus. Estrogen decreases the transcriptional activity of the UCN1 promoter via ER β [144]. UCN1 immunoreactivity was also increased in the mid-brain

region of male rats exhibiting alcohol preference behavior compared with controls, whereas UCN1 expression did not differ between control and alcohol-consuming female rats [145]. Others report a decrease in levels of UCN1 in pregnant compared with non-pregnant female rats [146].

In human studies, male suicide victims with major depression show higher UCN1 levels in the brains compared with healthy subjects, whereas no changes in UCN1 levels are noted between female suicide victims and control [147]. Interestingly, UCN1 and brain-derived nerve growth factor appear to be expressed in the opposite direction in a sex-specific manner in suicide victims with major depression. Single nucleotide polymorphisms (SNPs) in the *CRHR2* gene in human studies have been shown to associate with PTSD symptoms in women alone [148]. SNPs in the *CRHR2* gene are strongly associated with lifetime PTSD diagnosis in women but not men [148]. Corticolimbic circuitry is differentially activated more in women than in men, which could indicate why traumatic and/or stressful events have a greater effect. Women show increased susceptibility to stress disorders including PTSD and depression possibly due to changes in neuronal plasticity [149]. That same vulnerability is explicit in the role of CRF₂ in reducing the effect of those stress disorders with the diminishing of symptoms associated with the *CRHR2* gene.

Genetic vulnerability for major depressive disorder is also been linked to the *CRHR1* gene [150]. In a population study of Han Chinese, three SNPs within the *CRHR1* gene were identified. Haplotype with alleles G-G-T, for respectively the SNPs of rs1876828, rs242939, and rs242941, is significantly over-represented in major depressive patients compared to healthy control, suggesting that the carriers of this haplotype might have an increased probability of developing major depression in Han Chinese population. In another study, the same three haplotype-tagged SNPs of *CRHR1*—rs242939, rs1876828, and rs242941—also associate with a genetic risk for depression during pregnancy and post-partum [151]. SNPs in *CRHR1* are shown to contribute to the development of irritable bowel syndrome (IBS) [152]. Three SNPs in *CRHR1* (rs110402, rs242924, and rs7209436) were studied and IBS was associated with the major allele for these SNPs. Studies have found a significant interaction between SNP rs110402 of the *CRHR1* gene and the environment (childhood trauma) regarding the development of depression in adults [153,154]. Subsequent studies show that the protective effect of the rs110402 A allele against developing depression after childhood trauma is observed in men but not in women [155]. Interestingly, women allelic carriers of rs110402 SNP did not have altered cortisol response to the dexamethasone/CRF test, but male rs110402 allele carriers show decreased cortisol responses compared to male GG genotype carriers. Thus, environment and biological sex display complex interaction for the development of psychiatric disorders in humans.

9. CRF System in Sex-Specific Regulation of Gut Function and Pathology

While epidemiological data suggest that functional gastrointestinal diseases are more prevalent in women than men, these diseases also present sex-specific symptoms [10]. However, only recently have studies focused on elucidating mechanisms that might explain some of these sex differences in disease manifestation [156]. Exogenous administration of CRF and urocortins in the brain can lead to stress-related alterations of gut motor function, and the injection of CRF antagonists can prevent the effects of stressors, which provides evidence for the significant role of CRF receptors in the regulation of stress-induced alterations in gastrointestinal motility. CRF initiates the inhibitory effect of gastric transit and motility by acting upon CRF₂ [157,158]. Stressful episodes often precede flares associated with inflammatory bowel disease (IBD). In experimental animal models of colitis, a role for urocortins and CRF₂ has been elucidated. Global knockout of the CRF₂ receptor function or gut-specific elimination of CRF₂ with RNAi exacerbates inflammation in experimental models of colitis [7,71,92]. Gut-specific elimination of CRF₂, but not CRF₁, skews tumor-necrosis factor- α (TNF- α) and extracellular signal-regulated kinase (ERK) signaling [92]. Intriguingly, the administration of UCN1 in male *Crhr2* heterozygous mice with colitis ameliorates inflammation and increases survival. In sharp contrast, in female *Crhr2* heterozygous mice, UCN1 administration increases mortality [92]. The increase in the survival and amelioration of colitis symptoms in *Crhr2* heterozygous male mice

is attributed to UCN1's ability to decrease levels of TNF- α , interleukin (IL)-6, and IL-1 β as well as restore mitogen-activated protein kinase (MAPK) and heat shock protein 27 signaling in a sex-specific manner [92]. CRF signaling in the gut also acts upon mast cells to regulate inflammation as well as feeds back to the hypothalamus to regulate anxiety-like behavior in a rat model of functional dyspepsia [159]. In patients with diarrhea-predominant IBS, disease severity correlates with CRF₂ receptors in B7+ mast cells/extracellular vesicles [160]. Mast cells are important targets as their pliability permits fast and selective immune responses as well as regulate gut permeability [161]. Mast cells harbor CRF receptors, and the two receptors mediate opposite signaling in immune cells [50].

While studies in human samples have demonstrated a role for the components of the CRF system in the regulation of immune responses, few have ascertained sex-specific actions of these hormone receptors. CRF regulates macrophage function by increasing lipopolysaccharide-controlled cytokine production in a CRF receptor-dependent manner [162,163]. In human studies, colons from IBD patients show altered expression levels of UCN1, UCN2, and CRF₂ [87,92,164–166]. CRF₂-IR is found in goblet and immune cells in the duodenum and colons of human subjects with levels of UCN1 and CRF₂ increasing in male but not female patients with Crohn's disease [92]. Furthermore, the spatiotemporal distribution of UCN1–CRF₂ differs between male and female subjects, where co-localization is seen in gut samples from male but not female Crohn's disease patients [92].

The sex-specific role of UCN1 and CRF₂ is reported in the pathogenesis of pancreatic inflammation [93]. While treatment with caerulein induces pancreatitis in both male and female mice, after an identical dose of caerulein, C57BL6 female mice show less severe pancreatitis than C57BL6 male mice [93]. Female mice show less necrosis, zymogen granules, and vacuolization than male mice, but they have similar levels of edema and neutrophil infiltration as male mice [93]. In female *Crhr2*^{-/-}, caerulein induces more severe inflammation compared to male mice [93]. Sex-specific differences in rough endoplasmic reticulum (RER) ultrastructure along with signaling in unfolded protein response and autophagy are also reported [93]. While pancreatic acinar cell ultrastructure from saline-treated male and female mice is very similar, the induction of pancreatitis resulted in more severe RER ultrastructure damage in the acinar cells of female compared with male mice (Figure 5, left versus middle panel). In male mice lacking a functional CRF₂ receptor (*Crhr2*^{-/-}), pancreatitis causes severe and more dramatic distortion of RER cisternae, mitochondrial swelling, and unusual autophagic bodies compared to C57BL6 mice of both sexes as well as *Crhr2*^{-/-} female mice (Figure 5, top right panel). Interestingly, an in-between phenotype is seen in heterozygous *Crhr2* mice i.e., mice haploinsufficient for CRF₂ receptor function. These findings suggest to us that cellular stress downregulates CRF₂ receptors, putting an undue burden on organelles such as the RER and mitochondria. RER is the main organelle responsible for protein synthesis and quality control, whereas mitochondria regulate cellular respiration and oxidative stress. These damaged RER and mitochondria structure/function would explain the increase in accumulation of misfolded proteins, altered secretory capacity, and oxidative stress. If this ultrastructure damage occurs in other cell types as well, it would provide a logical explanation as to why stress exacerbates the accumulation of misfolded proteins and increases oxidative stress in other pathologies, such as neurological diseases. Interestingly, UCN1 ameliorates pancreatitis severity in male but not in female mice by restoring several of the above-mentioned signaling pathways. In another study, UCN2 decreases pancreatitis symptoms and inflammation via nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) pathways [167].

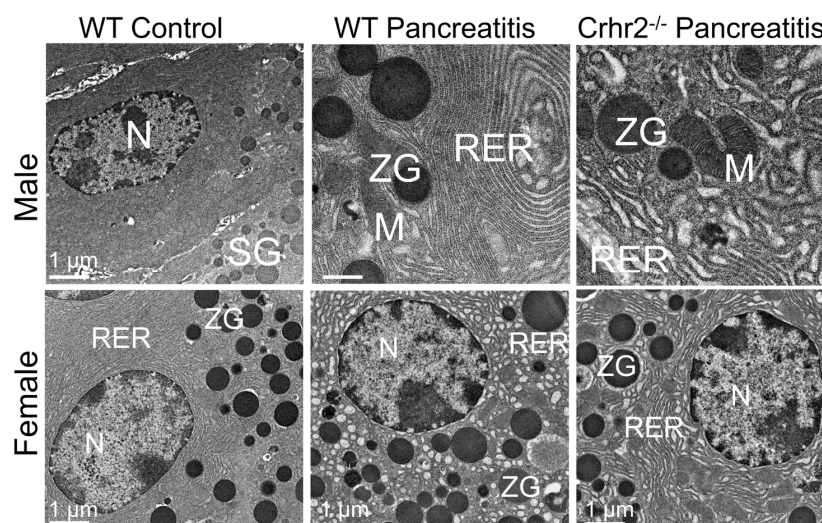


Figure 5. Electron micrographs showing representative pancreatic acinar cell sections from control and pancreatitis male and female mice. Endoplasmic reticulum (RER) shows whorling and the distortion of cisternae; mitochondrial (M) swelling is also evident. N: nucleus; SG: secretory granules; WT: wild-type; ZG: zymogen granules. Scale bar: 1 μ M. Modified from Kubat et al. [93].

Globally, 14 million more men are diagnosed with type 2 diabetes than women, despite more women being obese than men, as reflected by their body mass index measurements [168]. Interestingly, impaired glucose tolerance is more common in women than in men, independent of age. Stress is known to cause an elevation in blood glucose levels, diabetes, fatty liver, obesity, and metabolic syndrome [169]. Prospective longitudinal cohort studies spanning >10 years show that *perceived stress* increased the risk of developing diabetes in men by approximately 2-fold and had a 1.4 higher odds ratio than women. Perceived stress is also associated with increased insulin resistance. The same study reports that excessive work and work reward imbalance is also associated with a nearly 2-fold increased risk for diabetes in men but not women [170]. Short duration or poor initiation of sleep is also associated with an increased risk of diabetes in both men and women. A recent genome-wide association study of approximately 890,000 people with type 2 diabetes (DIAGRAM consortium) [171] identified several novel diabetes susceptibility loci including the stress receptor corticotropin-releasing factor receptor 2 (*Crhr2*). Thus, stress, whether emotional or physical, is highly associated with unhealthy lifestyle and behavior that includes eating behaviors, low exercise levels, smoking and alcohol abuse, and poor sleep.

Meta-analysis studies reveal that diabetic women are at 50% greater risk for cardiovascular-related mortality and 30% greater risk for stroke than diabetic men. In a clinical trial on patients with diabetes and vascular disease (ADVANCE-ON), glucose control with gliclazide resulted in less frequently end-stage kidney disease and subgroup analysis by sex shows that the risk reduction was only significant in men [172]. Women also have a higher risk of experiencing severe hypoglycemia with insulin therapy. Female sex, statin use, smoking, diabetes duration, body weight, and initial HbA1c all predict earlier failure of any “dual” treatment [172]. Despite this mounting evidence of sex-differences in the manifestation of metabolic disease in humans, very little is known about the sex-differences in metabolic signaling pathways.

Recent evidence from experimental models of diabetes suggests that urocortin 2 and 3 (UCN2–3) might be involved in the regulation of insulin secretion and blood glucose levels [33,43]. Islets lacking UCN3 show increased insulin secretion [43] and the exogenous administration of UCN2 normalized glucose levels in two distinct murine models of diabetes [33] and increased glucose uptake in the skeletal muscles of obese mice [32]. Studies from our laboratory have examined sex-specific actions of CRF₂ receptor in diabetes [34]. In this study, we find that *male Crhr2*^{-/-} and *Crhr2* haploinsufficient mice have worse glucose and insulin tolerance, microvesicular hepatic steatosis, and dyslipidemia than

female *Crhr2* mice, strongly suggesting that the *Crhr2* locus modulates the sex-specific expression of multiple genes influencing diabetes and metabolic syndrome in humans and mice. In agreement with our observations, a systematic study that characterized the effects of sex and body weight on metabolic outcomes in C57BL/6 mice found that male mice had worse glucose clearance on a high-fat diet than female mice [173]. Furthermore, this study used nearly 300 mice per sex to characterize metabolic phenotype on breeder's chow and high-fat diet (HFD) to create a reference database [173].

10. CRF System in Reproduction

In the brain, CRF mRNA expression in the PVN is reported to be higher in female than male rats [174]. This discrepancy is associated with the estrous cycle, with CRF expression highly correlating with sex hormone levels [174]. Higher rates of depression are noted after puberty and before menopause in women, further supporting the probability of ovarian hormones playing a role in susceptibility to stress-related psychiatric disorders. Studies find that estrogen promotes CRF expression in the PVN, whereas androgens inhibit CRF expression in rodents [175]. A human study shows that men with higher levels of estrogen than controls have a greater number of hypothalamic CRF neurons, but the predicted decrease in CRF neurons in women after menopause are not found, suggesting that other factors besides sex hormones are at play [176]. The sexually dimorphic BNST brain region also shows CRF-IR that is regulated by gonadal sex steroids in the female mice [177]. As discussed earlier, both estrogens and androgens modulate expression levels of CRF₁ and CRF₂ receptors in rodents of both sexes. While expression of both ligands and receptor follows cycling of estrogen levels, androgens appear to dampen stress responses and reduce anxiety-like behaviors in male rodents by increasing expression of CRF₂ [103,105], which is consistent with our observations, *vis-à-vis*, a role for CRF₂ in exerting protective effects in inflammation and metabolic stress in peripheral tissues [7,34,92,93].

The CRF system's importance to maternal–fetal stress patterns and embryo implantation further emphasizes the role it plays, specifically in females. In pregnancies with preeclampsia, CRF levels are increased, whereas CRF binding protein levels are at a lower level than in normal pregnancies, indicating the possibility of a greater proportion of free CRF having an effect on the pathology of preeclampsia [178]. With generally higher concentrations of free CRF in normal pregnancies as well, those heightened levels are known to increase placental vasodilation. Fetal urocortin levels measured at term without labor are lower than those at term with labor and in preterm labor [94]. UCN1 also plays a similar role, increasing uterine contractility and inducing the production of ACTH and prostaglandin PGE₂ to vasodilate the placenta [90]. UCN2 and UCN3 could be vital to the development of the trophoblast and the continuation of pregnancy, but increased concentrations in preeclampsia placentae could represent a reaction to oxidative stress [179].

UCN1 is important for embryo implantation [90,180,181]. UCN1 is abundantly expressed in early and late gestational tissues [179]. In pregnancy, UCN1 regulates vascular tone [36,37]. Reduced circulating levels of UCN1 are associated with increased uterine artery resistances and may regulate uterine artery tone [182]. Urocortin directly and indirectly triggers myometrial contractility, which induces uterine contractions [37,90]. We have shown involvement of UCN1 in regulation of microvascular permeability [35]. Others describe its potential role in preeclampsia [179]. Blood flow regulation is key for the overall health of the placenta and brain development in the fetus. Thus, impaired UCN1 levels may result in prenatal complications and have significant implications for adverse pregnancy outcomes, such as preterm birth, low body weight, and neonatal morbidity.

Human studies that examined UCN1 in fetal cord blood or maternal plasma at birth also found significant relationships of UCN1 to risk of preterm birth, as well as associations with greater neonatal morbidity [183–185]. Two studies that used amniotic fluid acquired during midgestation found a minimal relationship of UCN1 to other stress measures or to birth outcomes [186,187]. In contrast, two other studies that examined levels in amniotic fluid found a relationship of higher UCN1 levels to preterm birth [188,189]. One study sampling maternal serum when labor was initiated did not find an association of UCN1 to risk of preterm birth [190]. Clearly, findings have not been consistent regarding

the potential role of UCN1 in predicting adverse birth outcomes, which is probably due to variability in the timing of sample collection. However, it has been suggested that sampling in the second trimester (when the studies of amniotic fluid took place) may lead to less reliable results [187]. It has been suggested that the predictive values of the entire CRF system reach clinical reliability between 26 and 31 weeks of gestation [191]. Thus, research is needed that uses samples during the third trimester rather than midgestation to increase reliability in predicting outcomes. In agreement with these published observations about the role of UCN1, we have also found that placental/fetal UCN1 levels inversely correlate with perceived stress score (PSS) in a cohort of healthy pregnant women. As PSS increased, placental UCN1 levels decreased (Figure 6). Higher PSS indicates higher levels of stress, and lower UCN1 could lead to an insufficient vasodilation of the placenta. Excess vasoconstriction of the placenta is a known possible cause of preeclampsia, so combined with the indication that stress in mothers can affect the placenta during pregnancy, this suggested relationship warrants further exploration.

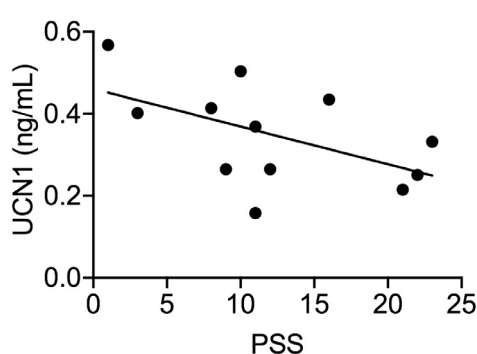


Figure 6. A scatterplot of the correlation between perceived stress score (PSS) and placental UCN1 levels. Analysis revealed that placental UCN1 levels were negatively correlated with PSS scores ($r = 0.531$, $p = 0.07$). Higher PSS scores indicate higher levels of stress experienced during the third trimester of pregnancy. Nulliparous women ($n = 12$) were recruited at the University of California San Francisco (UCSF) Mission Bay Hospital under UCSF's institutional review board protocol # 16-10957. All women carried to term and completed PSS questionnaires between 37 and 40 weeks of gestation. Placentae were collected after delivery, and UCN1 levels were measured using ELISA.

In the ovarian cycle, UCN1 targets steroidogenic luteal cells during luteal regression by binding to the CRF receptors. UCN1 is expressed in mid- and late-phase corpus luteum luteinized granulosa and thecal cells [96]. CRF and CRF₁ receptor mRNA are found in higher levels in regressing corpus luteum than in the pregnant corpus luteum or the mid-luteal phase. These studies have not found evidence of UCN2 and UCN3 having a part in the ovary [192,193]. UCN1 mRNA is expressed during the endometrial phases of the menstrual cycle, with the highest concentrations of UCN1 and mRNA in the late secretory endometrial phase [194]. CRF and UCN1 could also play a role in endometriosis, as peritoneal endometriosis tissues with activated mast cells staining for both. This, along with heightened urocortin levels in endometriomas, indicates a relationship with the symptoms of endometriosis, and whether that correlation exists between low fertility, inflammation, fibrosis, or spontaneous abortions is still unknown [195].

The length of human gestation appears to be decided by the combination of the UCN1 from the fetus and the CRF from the placenta. In post-term pregnancy, CRF and UCN1 levels are lower than in term pregnancy, and the concentrations of both neuropeptides decrease as the induction-delivery interval increases [95]. Serum UCN1 measured from preterm deliveries and term deliveries were not significantly different, so maternal UCN1 does not seem to be a reliable marker [190]. Another study showed that low UCN1 levels in amniotic fluid at midterm are suggestive of preterm birth [188]. A recent study showed that a retroviral long terminal repeat transposon-like element 1B (THE1B) controls placental expression of CRF in humans and other anthropoid primates [196]. This element is not present in prosimians and non-primates, including rodents such as mice and rats. Placental

CRF in turn influenced gestational age and birth timing in human pregnancies [196]. Placental, not hypothalamic (or brain), CRF expression is associated with gestational age in humans in this study. Taken together with results from several vertebrate experimental models and human studies, these data suggest that CRF and urocortins are key players in maintaining healthy pregnancy and birth outcomes.

11. Summary

The CRF family of peptide hormones and their two known CRF receptors regulate several physiological functions besides the well-characterized regulation of the HPA axis responses (Figure 7). Stressors in life are unavoidable. Stress response to physical, psychological, environmental or cellular stressors has two arms: *initiation* and *recovery*. CRF binding to CRF₁ receptors is primarily responsible for initiating stress responses via the HPA axis and the release of glucocorticoids, whereas urocortins binding to peripherally expressed CRF₂ receptors are responsible for the recovery response to stress and subjected to feedback regulation by glucocorticoids. We think that this system is in “over-drive”, and the inability of the system to return to homeostasis via mechanisms that largely involve CRF₂ receptors puts undue burden on this system. The myriad of molecular and cellular signals and functions that this system orchestrates in multiple organs for a balanced stress response is an overwhelming task. Some of these signals and functions are shared between members of the two known biological sexes, whereas others are distinct. This ancient system’s role in regulating sexually dimorphic signaling responses in a plethora of pathophysiology is emerging and will be key for developing therapeutic strategies that will be successful in a sex and gender-specific manner.

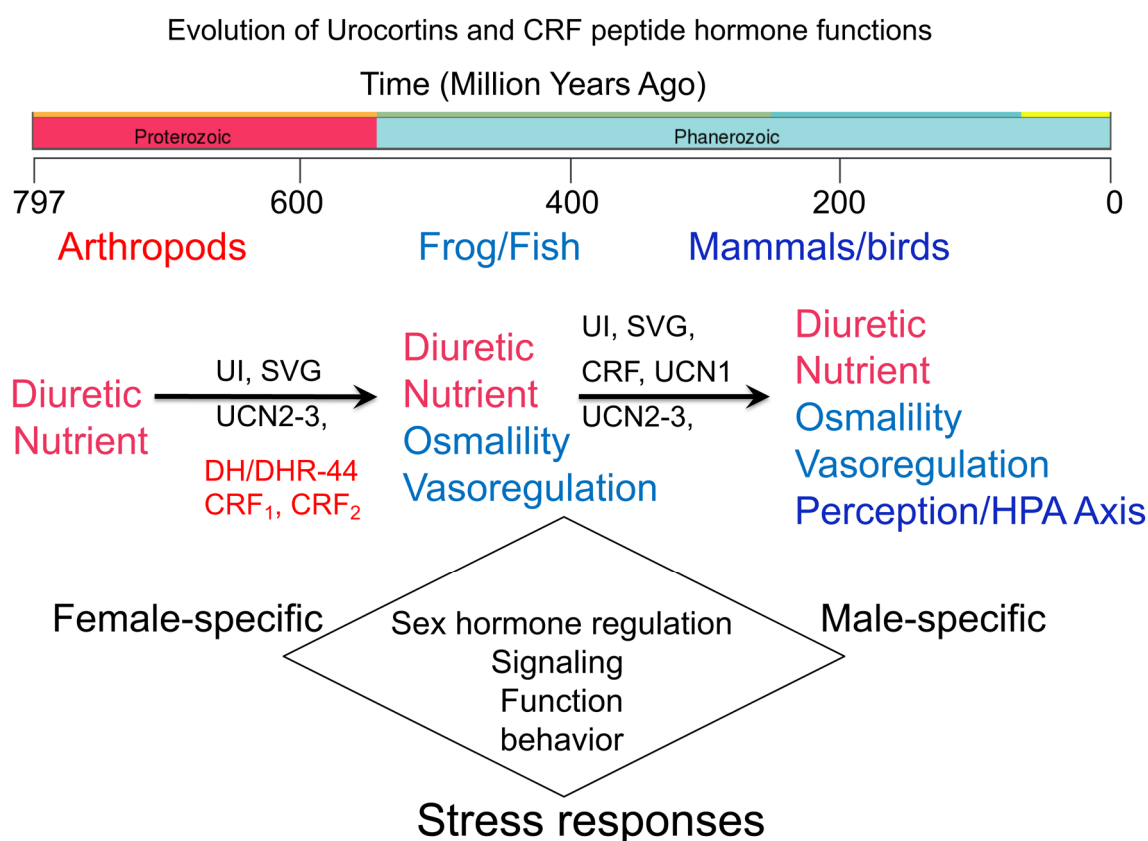


Figure 7. A summary schema showing the evolution of CRF and urocortins hormones and their functions.

Diuretic hormones (DH44) and DH44 receptors in the fruit flies probably served as prototypes for urocortin 3 and 2, and CRF receptors, respectively, which first appeared in Actinopterygii (fish) and Sarcopterygii after a genome-wide duplication event nearly 400 million years ago. Distinct CRF, urotensin I (UI), urocortins, and sauvagine peptides first seemed to have appeared nearly 500 million years ago. The innervation of hypothalamic cells by CRF family members occurred in class Actinopterygii along with appearance of urocortin-like peptides. The release of CRF into the portal circulation appeared in Sarcopterygii, whereas UI-like and sauvagine peptides transitioned to being expressed in the skin in the amphibians. The innervation and descending of CRF fibers in the spinal cord seemed to have occurred nearly 290 million years ago [2]. As the peptides evolved, the functions appeared to have diversified to include osmo and vasoregulation, as well as regulation of HPA axis-mediated stress responses. By virtue of having sex hormone response elements in the promoter regions of all four hormones and two receptors, sex-specific differences in the expression and function mediated by the CRF system are noted.

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