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Title

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Journal

International Journal of Hygiene and Environmental Health, 218(1)

ISSN

1438-4639

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Publication Date

2015

DOI

10.1016/j.ijheh.2014.08.003

Peer reviewed

Published in final edited form as:

Int J Hyg Environ Health. 2015 January ; 218(1): 91–98. doi:10.1016/j.ijheh.2014.08.003.

Ratio of cord to maternal serum PCB concentrations in relation to their congener-specific physicochemical properties

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Abstract

The aim was to characterize placental transfer of some congeners of polychlorinated biphenyls (PCBs) and to relate human *in utero* exposure to these pollutants to their physicochemical properties. We included into the study 1134 births during the period 2002–2003 from two highly PCB contaminated districts in eastern Slovakia. Concentrations of 15 PCB congeners (IUPAC No. 28, 52, 101, 123⁺¹⁴⁹, 118, 114, 153, 105, 138⁺¹⁶³, 167, 156⁺¹⁷¹, 157, 180, 170, and 189) in umbilical cord (C) and maternal serum (M) were determined. The C/M ratios were significantly related, either positively or inversely depending on parameter, to the logarithm of partition coefficient octanol-water (K_{OW}), to fusion enthalpy at the melting point, molecular weight, water solubility, total surface area of the molecule, solvent accessible surface area, melting point, molar volume, and molecular electronegativity distance vector. We found an inverse association between $\log K_{OW}$ and lipid adjusted $\log C/M$ (const= 1.078, $b_1 = -0.179$, $p < 0.001$, $R^2 = 0.039$). Parameters evaluated were interrelated except fusion enthalpy at the melting point and electron affinity vs. solubility. We discuss the possible role of cholesterol as a transplacental transporter of PCBs.

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The authors declare no competing financial interest.

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Keywords

polychlorinated biphenyls; placenta; placental transfer; partition coefficient octanol water; physicochemical parameters

Introduction

The sensitivity of developing organism to the effects of environmental pollutants during the prenatal period has been amply documented (Fox et al., 2012; Winneke, 2011; Parent et al., 2011; Gore, 2010; Dickerson et al., 2011; Wigle et al., 2008; Sly and Flack, 2008). Polychlorinated biphenyls (PCBs) (ATSDR, 2000) are members of the group of persistent organic pollutants (POPs) and are important with respect to bioaccumulation in environmental media, persistence in the environment and toxic properties. They have been detected in fetuses (De Koning and Karmaus, 2000; Berg et al., 2010) where they can exert adverse effects (Ulbrich and Stahlmann, 2004; Boucher et al., 2009). To reach the fetus they must cross the placenta. PCBs, as a group, easily pass the placental barrier (Park et al., 2008; Linderholm et al., 2007; Correia Carreira et al., 2011; Bergonzi et al., 2009) by simple diffusion due to their electronegativity, high lipophilicity and moderate molecular weight. However PCBs in the environment are a mixture of congeners, each of which is characterized by its own physicochemical properties and toxicity. The knowledge of rules of transplacental transfer is important for protection of developing organism. The speed and the extent of compound-transfer from the maternal to fetal side depend on the physicochemical and structural characteristics of the chemical as well as the physical characteristics of the maternal-placental-embryonic-fetal unit (Giaginis et al., 2009; Giaginis et al., 2011; Myren et al., 2007). Pollutant properties such as molecular weight, lipid solubility and protein binding could also determine the transfer of pollutants from mother to fetus to a great extent (Myllynen et al., 2009). Kinetics of placental transfer of several POPs in humans have only recently been described (Needham et al., 2011; Suzuki et al., 2005; Tsukimori et al., 2013; Porpora et al., 2013), however we did not find any data on correlation of placental transfer of POPs to their physicochemical parameters. In a recent study on placental transfer of POPs any correlation between the maternal/cord serum concentration ratios and chemical properties of these pollutants such as molecular weight, molar volume, number of halogen substituents or log octanol water partition coefficient (K_{ow}) were found (Vizcaino et al. 2014). A close relationship between the physicochemical properties encoded in the molecular structure and the ability of PCBs to mimic natural hormones may reflect toxic responses they elicit in biological systems (Puri et al., 2003). It is known that of these factors the lipophilicity, mostly expressed as the K_{OW} , drives the kinetics of environmental pollutants in many biological systems (Hawker and Connell, 1988; Isnard and Lambert, 1988; Paasivirta et al., 1999; van Gestel et al., 1985; Woodburn et al., 1987). The aim of our study was to determine how is related the placental transfer of individual PCB congeners to their physicochemical properties. Besides transfer by simple diffusion, closely related to lipid solubility, transport of PCBs by carrier lipids was considered. In this connection we discussed which lipid components of serum may be involved in PCB transport across the placenta.

Materials and methods

We included into the study 1134 births during the period 2002–2003 from two districts (Michalovce and Svidnik) in eastern Slovakia highly contaminated by PCBs (Hertz-Picciotto et al., 2003). The characteristics of infants and mothers participating in the study have been described elsewhere (Hertz-Picciotto et al., 2003; Sonneborn et al., 2008; Sonneborn et al., 2008; Park et al., 2008). All women provided written informed consent, and the study protocol was approved by institutional review boards at the University of California–Davis and the Slovak Medical University, Bratislava. Concentrations of 15 PCB congeners (IUPAC No. 28, 52, 101, 105, 114, 118, 123⁺¹⁴⁹, 138⁺¹⁶³, 153, 156⁺¹⁷¹, 157, 167, 170, 180, and 189) in the umbilical cord and maternal serum were determined using solid phase extraction and high resolution gas chromatography with micro electron capture detection as already described (Conka et al., 2005; Petrík et al., 2006). For quality control, a solvent blank and recovery sample (fortified porcine serum) were analyzed with each batch of 10 human serum samples. Recovery was checked using PCB 174 as internal standard and PCB 103 served as a syringe standard. Certified reference material, Mackerel oil (CRM no. 350, Community bureau of reference, Brussels, Belgium) was used to verify the analytical procedure every 3 months as described earlier (Conka et al., 2005). LODs for individual PCB congeners and samples were evaluated using the ratio of noise/peak height in GC chromatogram and standardized for unit sample amount. We restricted our analyses to serum samples with PCB concentrations \geq LOD. A useful marker of placental transfer of chemicals is the ratio between concentration in cord serum (C) to that in maternal serum (M) (Abballe et al., 2008; Covaci et al., 2002; Patayová et al., 2013; Vizcaino et al. 2014). C/M ratios were calculated for PCB congeners for which a sufficient number of pairs (>133) were available from samples with concentrations \geq LOD. We considered this number satisfactory as concentrations of PCBs are strongly intercorrelated in maternal and cord serum (Ayotte et al., 2003).

We report both lipid adjusted and wet weight PCB concentrations. We estimated total serum lipids using the enzymatic summation method (Akins et al., 1989). Serum total cholesterol (TC), nonesterified cholesterol (FC), triglycerides (TG), and phospholipids (PL) were assayed by automated, enzymatic methods and total lipids (TL) were calculated from the expression $TL = 1.677 * (TC - FC) + FC + TG + PL$. We gathered data on physicochemical parameters of individual PCB congeners from available literature sources. We used linear regression to examine the association between log C/M ratio for PCB congeners and their physicochemical parameters (SPSS 16, Softonic International S.L.).

Results

We present information on the number of analyzed serum samples and number of samples with concentrations \geq LOD and geometric means of lipid adjusted (ng/g lipid) and wet weight (ng/mL) concentrations of PCB congeners in cord and maternal blood serum in Table 1. It can be seen that at the low end of M values, with PCB congeners 28, 52, 101, and 189, the lipid adjusted C values $>$ M values. On the other hand, at the high end of M values, C values $<$ M values. The difference between the wet weight adjusted concentrations in cord blood and maternal blood serum reflects much lower lipid content in cord serum (median

2.46 g/L) compared to maternal serum (median 10.17 g/L). For PCB congeners 105, 114, 123, and 157 the percentage of samples with concentrations below the detection limit was unacceptably high and therefore these congeners were not considered for statistical treatment. For the remaining PCB congeners the geometric means of lipid and wet weight C and M concentrations, ranked in order of increasing values of M.

When searching for the physicochemical parameters we have found data on K_{OW} (Hawker and Connell, 1988; Woodburn et al., 1987; De Bruijn and Hermens, 1990; Li et al., 1992; Lü et al., 2007; Mackay et al., 1992), water solubility (Abramowitz and Yalkowsky, 1990; Dunnivant and Elzerman, 1988; Huang and Hong, 2002; Makino, 1998; Yalkowsky and Valvani, 1979; Yalkowsky et al., 1983), total surface area of the molecule (Hawker and Connell, 1988; De Bruijn and Hermens, 1990; Yalkowsky and Valvani, 1979), solvent accessible surface area (Makino, 1998), electron affinity (Makino, 1998), melting point (Abramowitz and Yalkowsky, 1990; Yalkowsky and Valvani, 1979), enthalpy of fusion at the melting point (Puri et al., 2003), molar volume (Yalkowsky and Valvani, 1979; Huang et al., 1993; Liu et al., 2008; Shiu et al., 1986) and molecular electronegativity distance vector (Liu et al., 2008; Qin et al., 2008) (Table 2).

We found by bivariate regression analysis (Table 3) that C/M values are statistically significantly related to the physicochemical parameters found, except electron affinity. We illustrate the relationship between lipid adjusted log C/M and log K_{OW} in Figure 1 as this parameter is playing a central role in the behavior of many xenobiotics in biological systems (Hawker and Connell, 1988). It shows that the placental transfer of PCB congeners is decreasing with their increasing lipophilicity (const = 1.078, $b_1 = -0.179$, $p < 0.001$, $R^2 = 0.039$).

Discussion

In the current study we have shown that the ratio of cord to maternal serum concentration of PCB congeners is inversely related to their lipophilicity expressed as K_{OW} . We expected a positive relationship between these variables in agreement with rules governing passage of lipid soluble xenobiotics through lipid bilayer membranes (Balaz, 2009) and assumptions on placental transfer of lipophilic substances by diffusion (Suzuki et al., 2005; Tsukimori et al., 2013). We confirmed findings that the overall effect of the physicochemical properties on behavior in biological systems is difficult to predict (Needham et al., 2011) and transfer of PCB congeners from maternal to fetal side is more complex than the simple diffusion of the free fraction governed by parameters as molecular weight, pKa, protein binding and lipid solubility (Pacifci and Nottoli, 1995; Audus, 1999). The predictive potency of lipid solubility has already been questioned in human toxicokinetics (Tonnelier et al., 2012). A clue for explaining this controversy may be at least partly in a variety of transporters expressed in the placenta which can facilitate transfer of xenobiotics (Evseenko et al., 2006). Depending on the localization and function, transporters may either increase or decrease xenobiotic transfer towards fetal circulation (Myllynen and Vähäkangas, 2013; Vähäkangas and Myllynen, 2009). Due to structural resemblance of PCBs to thyroxin the most important candidates are transporters of thyroxin. Thyroid hormone can cross placenta and maternal thyroxin is crucial for normal development of fetal brain and other organs. To cross placenta

thyroxin uses various transport mechanisms (Patel et al., 2011; Mortimer et al., 2012; Landers et al., 2013a; 2013b; Landers et al., 2009; Feldt-Rasmussen and Rasmussen, 2007; Richard et al., 2012). With regard to other potential transporters we found data on binding of PCBs to albumin (Han et al., 2013; Becker and Gamble, 1982), serum transport proteins-transferrin and thyroid-binding globulin (Cheek et al., 1991; Marchesini et al., 2008) and plasma lipoproteins and proteins (Borlakoglu et al., 1990; Becker and Gamble, 1982). Fatty acids have been suggested as a transporter for structurally related dioxins and furans (Koppe et al., 1992). Lipid transport is complex, involves a variety of molecules present on the syncytiotrophoblast surface, and requires their coordinated interaction with other intracellular molecules in different placental cellular compartments. Currently, by far, all of the molecules and processes involved in transplacental lipid transfer have not been identified (Desoye et al., 2011; Larqué et al., 2013; Gil-Sánchez et al., 2012; Herrera et al., 2006). We discuss the hypothesis of transplacental transport by lipids carrying PCB molecules by analyzing behavior of concentration of serum lipid components in maternal and infant's blood in Figure 2 and Table 4. From the lipid components only total cholesterol in cord serum significantly correlated with that in maternal serum ($r_s=0.045$, $p<0.001$) and can thus be considered as transporter of PCBs.

The C/M ratio, a marker of placental transfer of PCBs used in this study, suffers from potential disadvantage as inter-individual variations due to endogenous and/or exogenous factors. Indeed in our previous study (Patayová et al., 2013) we have shown that the anthropometric, socioeconomic, and maternal health factors are associated with functioning of the important part of the body system, the placenta. The strong interrelation between the different physicochemical parameters makes difficult the identification of a "primer" association between one of them and the measured outcomes (materno-fetal transfer rate). With this objective we analyzed our data with multiple linear regression. However interrelation between variables precluded to obtain valid results. Extremely high values of the Variance Inflation Factor (how2stats, 2011) confirmed multicollinearity. Lipophilicity determines the behavior of many chemicals in biological systems (Balaz, 2009) and in agreement bioconcentration of PCBs in environmental media is dependent on K_{OW} values (Eisler and Belisle, 1996; Paasivirta et al., 1999; Hawker and Connell, 1985a; 1985b; Noegrohati and Hammers, 2008; Padmanabhan et al., 2006; Hope et al., 1998). The role of K_{OW} in functioning of the blood-brain barrier, one of the most important and sophisticated biological systems, was confirmed in a recently published model relating transfer parameters across it to the physical chemical properties of 70 structurally diverse compounds (Zhang et al., 2010). For placenta a similar general model has not yet been published. Moreover, most modeling efforts have focused on placental drug transfer (Hutson, 2011; Hutson et al., 2011) with less attention to environmental toxins (Needham et al., 2011).

Our main finding that the transfer of PCB congeners across human placenta decreases with increasing K_{OW} contrasts to enhanced passage of substances across the blood-brain barrier (Hou and Xu, 2003; Levin, 1980) with increasing K_{OW} . It has to be noted however, that K_{OW} of substances studied in blood brain barrier transfer were several orders of magnitude smaller than K_{OW} s of PCB congeners evaluated currently.

There is an indication from our data that PCB congeners occurring in serum at low concentrations in a typically environmental PCB congener mixture have a C/M ratio >1 compared to more abundant congeners with a C/M ratio <1. However, the significance of this observation may be questioned due to detection limits in the current study. A similar greater exposure to offspring from lower than from higher doses has been described (Chen et al., 2001) for placental transfer of non-ortho substituted PCB congeners.

Our observation on inverse relationship between transfer of PCB congeners through the human placental barrier and lipophilicity, is in agreement with PCB mother fetus transfer data in marine mammals. In grey seal (*Halichoerus grypus*) the transfer from inner blubber to maternal serum was selective and strongly depended on the log K_{OW} value of the compounds, with less lipophilic compounds being more efficiently released. These results indicate that compounds with a high log K_{OW} and thus with a high lipophilicity are less easily transferred into the bloodstream (van den Berghe et al., 2012). In southern elephant seals (*Mirounga leonina*) lactational transfer rates were dependent on the log K_{OW} values of the analytes measured, less lipophilic compounds being more readily transferred to the pups by the lactational route (Miranda Filho et al., 2009). In sea lions the fetus blubber to mother partition ratio of PCBs decreased with increasing K_{OW} (Greig et al., 2007). In common dolphin the percentage of transfer declines inversely with the number of chlorines paralleled by increase of lipophilicity (Borrell and Aguilar, 2005). In a study on arctic beluga whales (*Delphinapterus leucas*) a single physicochemical parameter, log K_{OW}, largely explained the transplacental transfer for PCBs with congeners having a log K_{OW} <6.5 preferentially transferred to the fetus (Desforges et al., 2012). This parameter has also been examined in Zebrafish (*Brachydanio rerio*), in which a curvilinear relationship was observed between bioconcentration of PCBs and log K_{OW} based upon data covering the log K_{OW} range 5.06–8.18. Highest bioconcentration resulted at about a log K_{OW} of 7.38, above which the degree of bioconcentration decreased (Fox et al., 1994). The physicochemical background to these events can be found in the diffusion limitation of the exchange between adipose and blood, which steeply decreased as a function of the K_{OW} for the 13 PCBs studied (Levitt, 2010). In agreement with our findings was found a decrease of partition ratio between lipid-based concentrations of PCB congeners in milk and maternal serum in regard to the number of chlorine substitutions of each congener measured (Needham et al., 2011). A similar trend was observed with cord/maternal ratios (Vizcaino et al., 2014).

That PCB congener specific C/M values are related to several physicochemical parameters was expected as many physicochemical parameters of PCB congeners are interrelated (Hawker and Connel, 1988; De Bruijn and Hermens, 1990; Shiu and Mackay, 1986; Patil, 1991; Mackay et al., 1980; Inoue et al., 2006; Miller et al., 1985; Banerjee et al., 1980; Chiou et al., 2005; Opperhuizen et al., 1988; Silla et al., 1992). We confirmed this by pairwise Spearman rank correlation (Table 5) except electron affinity and fusion enthalpy at the melting point which were partly or completely unrelated to other parameters.

Acknowledgments

This project has been funded by the U.S. National Institutes of Health grants R01-CA96525 and K12-ES019852, the European Commission through the 7FP project OBELIX (No. 227391) and the project “Center of Excellence of

Environmental Health”, ITMS No. 26240120033, based on the supporting Operational Research and Development Program financed from the European Regional Development Fund.

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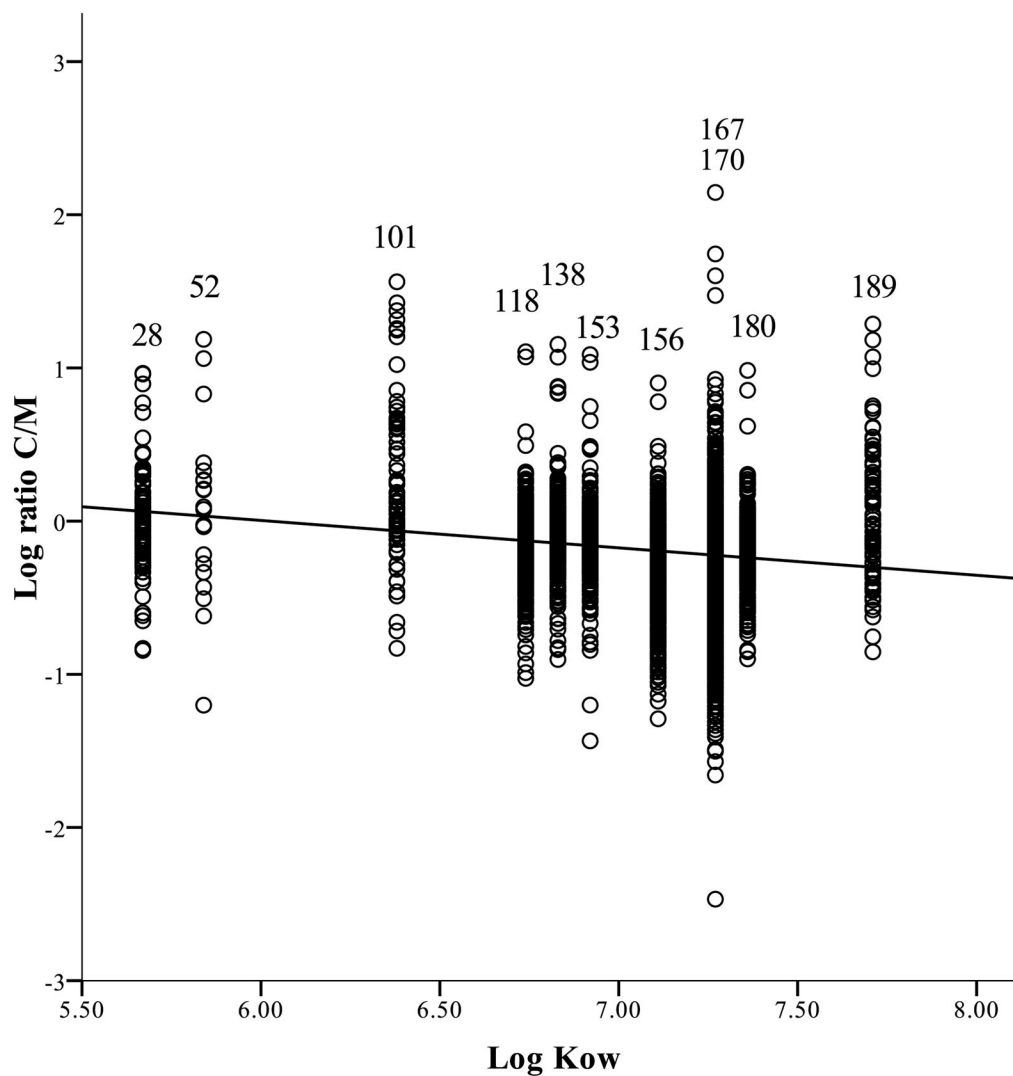


Figure 1. Relationship between the logarithm of the ratio of cord/maternal PCB congener lipid adjusted serum concentration (C/M) and log octanol-water partition coefficient (K_{OW}). The numbers in the figure denote PCB congener. The relationship is characterized by slope = -0.179 , $p < 0.001$, $R^2 = 0.039$ and. R^2 stands for coefficient of determination and p for significance.

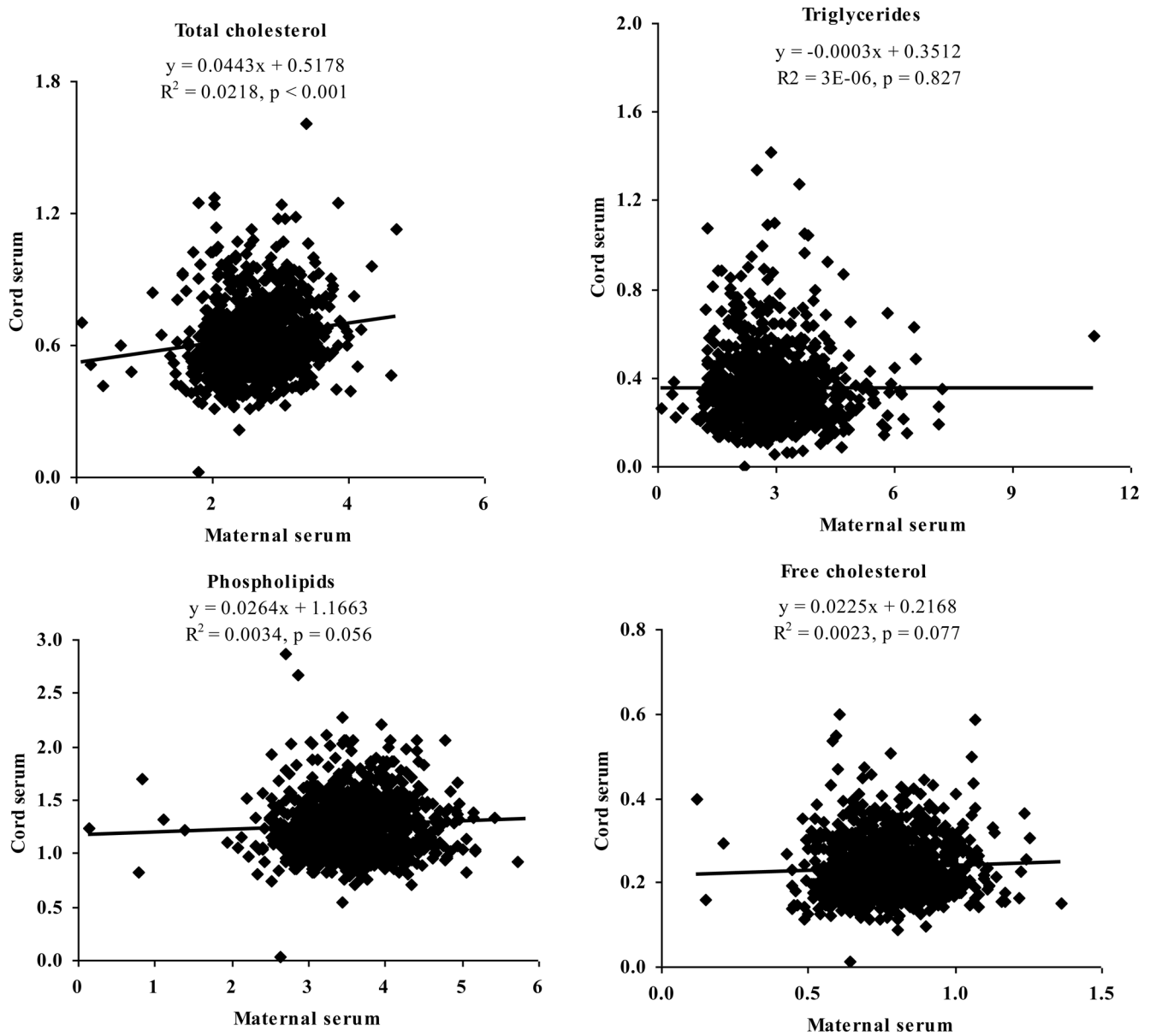


Figure 2.
 Relationship between concentration (g/L) of main lipid components in cord blood serum and in maternal blood serum.

Table 1

Numbers of analyzed serum samples from cord and maternal blood and numbers of serum samples with concentrations of PCB congeners LOD (limit of detection) and geometric means of concentration of PCB congeners in cord and maternal blood serum.

PCB congener	28		52		101		105		114	
	Mother	Cord	Mother	Cord	Mother	Cord	Mother	Cord	Mother	Cord
Total number	1053	1063	1094	1065	1101	1078	1101	1076	1101	1079
Number LOD	409	136	159	133	212	212	207	77	146	47
Concentration ng/g lipids	9.69	9.86	5.23	7.31	6.05	8.71	6.23	5.31	4.36	5.8
Concentration ng/mL	0.1	0.02	0.06	0.02	0.06	0.02	0.06	0.01	0.05	0.01
PCB congener	118		123+149		138+163		153		156+171	
Total number	1101	1080	1101	1079	1101	1080	1101	1081	1101	1080
Number LOD	920	750	154	133	1101	1074	1101	1079	1051	729
Concentration ng/g lipids	12.95	8.65	3.35	4.98	93.84	77.14	146.35	109.95	13.69	6.36
Concentration ng/mL	0.13	0.02	0.03	0.01	0.93	0.19	1.46	0.27	0.14	0.02
PCB congener	157		167		170		180		189	
Total number	1101	1080	1101	1080	1101	1080	1101	1080	1101	1080
Number LOD	208	76	571	194	1099	1066	1101	1079	339	133
Concentration ng/g lipids	3.57	5.25	9.3	6.64	57.29	35.76	134.33	90.09	4.41	4.92
Concentration ng/mL	0.04	0.01	0.1	0.02	0.57	0.09	1.33	0.22	0.05	0.01

Table 2

The values of physicochemical parameters of 15 PCB congeners.

PCB congener	Log octanol-water partition and Connell, 1988)	Solubility (Makino, 1998)	Molecular weight	Total surface area and Connell, 1988)	Solvent accessible surface area (Makino, 1998)	Electron affinity (Makino, 1998)	Melting point and Yalkowsky, 1990)	Fusion enthalpy at melting point (Puri et al., 2003)	Molar volume (Shiu et al., 1986)	Molecular electronegativity distance vector (Qin et al., 2008)
28	5.67	-6.22	257.54	230.83	121.44	-0.703	347	24.3	247.3	4.16
52	5.84	-7	291.99	235.84	130.98	-0.49	329	15.8	268.2	4.62
101	6.38	-7.8	326.43	251.62	139.72	-0.721	340	27	289.1	5.23
105	6.65	-7.52	326.43	259.41	137.6	-0.855	398	20.8		5.13
114	6.65	-7.5	326.43	259.41	137.87	-0.942	392	19.9		4.97
118	6.74	-7.33	326.43	262.04	139.3	-0.922	392	20.4	289.1	5.37
123+149	6.74	-7.42	326.43	262.04	137.6	-0.855	398	26	310	5.25
138+163	6.83	-8.38	360.90	264.76	146.86	-0.747	382	21	310	5.58
153	6.92	-8.49	360.88	267.39	148.61	-0.773	412	19.2	310	5.84
156+171	7.11	-8.64	395.32	273.15	153.14	-0.869	425		310	5.56
157	7.18	-8.25	360.88	275.01	146.79	-0.954	414	29.1	310	5.6
167	7.27	-8.21	360.88	277.64	148.25	-1.009	408	22.5		5.85
170	7.27	-8.9	395.32	277.74	154.41	-0.826	405	23.1		5.72
180	7.36	-9.1	395.32	280.37	155.95	-0.884	372	22.1		6
189	7.71	-8.72	395.32	290.61	156.02	-1.072	431	31.3		6.03

Table 3

Regression analysis between cord/maternal serum PCB concentration ratios (C/M) and physicochemical parameters characteristic for individual PCB congeners. Included were serum samples with concentrations LOD (limit of detection) of 11 PCB congeners. R² stands for coefficient of determination and p for statistical significance.

Physicochemical parameters	C/M	R ²	Constant	Slope	p
Log octanol-water partition coefficient	Wet weight	0.032	6.455	-0.126	<0.001
	Lipid adjusted	0.014	6.425	-0.010	<0.001
Electron affinity	Wet weight	0.026	-0.739	0.030	<0.001
	Lipid adjusted	0.014	-0.732	0.003	<0.001
Fusion enthalpy at the melting point	Wet weight	0.016	21.012	0.652	<0.001
	Lipid adjusted	0.010	21.14	0.066	<0.001
Melting point	Wet weight	0.062	371.635	-10.985	<0.001
	Lipid adjusted	0.035	369.27	-1.047	<0.001
Total surface area	Wet weight	0.032	253.787	-3.673	<0.001
	Lipid adjusted	0.014	252.913	-0.305	<0.001
Molecular weight	Wet weight	0.019	324.314	-7.361	<0.001
	Lipid adjusted	0.007	322.445	-0.549	<0.001
Solubility	Wet weight	0.011	-7.599	0.121	<0.001
	Lipid adjusted	0.003	-7.566	0.008	<0.001
Molar volume	Wet weight	0.019	287.812	-4.466	<0.001
	Lipid adjusted	0.007	286.679	-0.333	<0.001
Solvent accessible surface area	Wet weight	0.017	138.702	-1.773	<0.001
	Lipid adjusted	0.006	138.245	-0.129	<0.001
Molecular electronegativity distance vector	Wet weight	0.018	31.71	-0.509	<0.001
	Lipid adjusted	0.007	31.594	-0.041	<0.001

Table 4

Descriptive statistics on concentration of main lipid components (g/L) in cord and mother serum.

	Total cholesterol g/L		Phospholipids g/L		Free cholesterol g/L		Triglycerides g/L		Total lipids g/L	
	Cord	Mother	Cord	Mother	Cord	Mother	Cord	Mother	Cord	Mother
Number	1075	1066	1075	1067	1075	1067	1075	1067	1065	1065
Mean	0.63	2.61	1.26	3.55	0.23	0.77	0.35	2.83	2.52	10.23
Standard Deviation	0.16	0.53	0.26	0.58	0.07	0.15	0.17	1.01	0.52	1.98
Median	0.62	2.57	1.23	3.54	0.22	0.76	0.31	2.70	2.46	10.17
Minimum	0.02	2.70	0.03	0.15	0.01	0.12	0.05	0.10	0.13	1.75
Maximum	1.61	4.71	2.87	5.72	0.60	1.36	2.02	11.05	5.34	20.17
Geometric Mean	0.62	2.54	1.23	3.49	0.23	0.76	0.32	2.65	2.47	10.03

Table 5

Pairwise Spearman rank correlation between physicochemical parameters characterizing 15 PCB congeners. N stands for number, r_s for correlation coefficient and p for statistical significance.

Physico-chemical parameters	Molecular weight	Log octanol-water partition coefficient	Solubility	Total surface area	Solvent accessible surface area	Electron affinity	Melting point	Fusion enthalpy at the melting point	Molar volume	Molecular electronegativity distance vector
	N	15	15	15	15	15	15	14	9	15
Molecular weight	r_s	0.908	-0.962	0.916	0.955	-0.446	0.607	0.284	0.862	0.855
	P	<0.001	<0.001	<0.001	<0.001	0.095	0.016	0.326	0.003	<0.001
Log octanol-water partition coefficient	N	15	15	15	15	15	15	14	9	15
	r_s		-0.856	0.999	0.909	-0.667	0.683	0.320	0.880	0.956
	P		<0.001	<0.001	<0.001	0.007	0.005	0.265	0.002	<0.001
Solubility	N	15	15	15	15	15	15	14	9	15
	r_s			-0.866	-0.951	0.336	-0.522	-0.244	-0.844	-0.839
	P			<0.001	<0.001	0.221	0.046	0.401	0.004	<0.001
Total surface area	N	15	15	15	15	15	15	14	9	15
	r_s				0.913	-0.652	0.681	0.322	0.880	0.952
	P				<0.001	0.008	0.005	0.262	0.002	<0.001
Solvent accessible surface area	N	15	15	15	15	15	15	14	9	15
	r_s					-0.463	0.573	0.257	0.780	0.912
	P					0.082	0.026	0.374	0.013	<0.001
Electron affinity	N	15	15	15	15	15	15	14	9	15
	r_s						-0.659	-0.306	-0.633	-0.565
	P						0.008	0.288	0.067	0.028
Melting point	N	15	15	15	15	15	15	14	9	15
	r_s							0.326	0.780	0.608
	P							0.255	0.013	0.016
Fusion enthalpy at the melting point	N	14	14	14	14	14	14	14	8	14
	r_s								0.255	0.297
	P								0.542	0.303

Physico-chemical parameters	Molecular weight	Log octanol-water partition coefficient	Solubility	Total surface area	Solvent accessible surface area	Electron affinity	Melting point	Fusion enthalpy at the melting point	Molar volume	Molecular electronegativity distance vector
	N								9	9
Molar volume	r _s									0.844
	P									0.004
	N									15
Molecular electronegativity distance vector	r _s									
	P									

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