

# UC Irvine

## UC Irvine Previously Published Works

### Title

Closed Genome Sequence of an Environmental *Aeromonas veronii* Strain from California, United States, with an IncA/C Plasmid Carrying an Extended-Spectrum  $\beta$ -Lactamase Gene, blaVEB-3

### Permalink

<https://escholarship.org/uc/item/38c3k1ds>

### Journal

Microbiology Resource Announcements, 11(3)

### ISSN

2576-098X

### Authors

Lovero, Karissa G  
Mota-Bravo, Luis

### Publication Date

2022-03-17

### DOI

10.1128/mra.01033-21

Peer reviewed



# Closed Genome Sequence of an Environmental *Aeromonas veronii* Strain from California, United States, with an IncA/C Plasmid Carrying an Extended-Spectrum $\beta$ -Lactamase Gene, *bla*<sub>VEB-3</sub>

Karissa G. Lovero,<sup>a</sup>  Luis Mota-Bravo<sup>a</sup>

<sup>a</sup>Department of Ecology and Evolutionary Biology, University of California, Irvine, Irvine, California, USA

**ABSTRACT** We describe the extended-spectrum  $\beta$ -lactamase *bla*<sub>VEB-3</sub> gene found in an IncA/C plasmid in *Aeromonas veronii* strain SW3814, which was collected from a freshwater lake in southern California, United States.

**A***eromonas* bacteria are common in aquatic environments and may serve as reservoirs of antibiotic resistance genes (ARGs) (1–3). These ARGs are often associated with mobile genetic elements (MGEs) (4), which may facilitate their transfer to bacteria of clinical significance (5–7).

*Aeromonas veronii* strain SW3814 was collected from a freshwater lake in southern California (English Springs Park [33.9951N, 117.756W]). Water was filtered with 0.45- $\mu$ m filters (GN-6; Pall) and placed on CHROMagar orientation medium (CHROMagar, Paris, France) containing 4  $\mu$ g/mL cefotaxime (Sigma-Aldrich, St. Louis, MO). A purple isolate, labeled SW3814, was identified as *Aeromonas veronii* using matrix-assisted laser desorption ionization–time of flight (MALDI-TOF) mass spectrometry (Bruker, Billerica, MA). SW3814 was grown (for DNA extraction and tests) overnight in tryptic soy broth (BD Bacto) at 35°C and stored in 25% glycerol at –80°C. Genomic DNA was obtained using a FastPrep homogenizer (MP Biomedicals) with 0.1-mm silica spheres, followed by the DNA extraction method described by Maniatis et al. (8). DNA was quantified with a Qubit fluorimeter (Life Technologies). An Illumina library was prepared with a Nextera DNA Flex library preparation kit, loaded into a 300-cycle high-output flow cell (2  $\times$  150-bp paired-end reads), and run in a MiniSeq instrument with System Suite v2.0.0 (Illumina, San Diego, CA); 2,241,828 Illumina reads were obtained (reads with quality scores of >Q30, 91.1%). Illumina reads were quality filtered using fastp v0.23.1 (9). An Oxford Nanopore Technologies (ONT) (Oxford, UK) library was prepared using SQK-LSK109 and EXP-NBD196 kits, loaded into a FLO-MIN106D flow cell, and run in a MinION ONT device for 75 h. Base calling and quality filtering of ONT reads were conducted using Guppy for GPU v4.5.2; 18,373 ONT reads were obtained (mean size, 12,977 bp; minimum size, 1,000 bp; maximum size, 73,198 bp; reads with quality scores of >Q20, 63.2%). The genome was assembled using Unicycler v0.4.8-beta (10). The genome was annotated by the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) with the best-placed reference protein set and GeneMarkS-2+ v5.3 (11–13). The plasmid copy number was obtained with Unicycler depth. Default parameters were used for all software. The Center for Genomic Epidemiology was used to annotate ARGs using ResFinder v2.1 (14). To determine the association of *bla*<sub>VEB-3</sub> with MGEs, a BLASTn search was performed against the NCBI GenBank nucleotide database using the sequences of *bla*<sub>VEB-3</sub> and IS6100. The top six matches with the greatest query coverage were used to create Fig. 1B.

SW3814 has a chromosome of 4,684,058 bp (GC content, 58.4%; Illumina coverage, 53.8 $\times$ ; ONT coverage, 45.8 $\times$ ), an IncA/C plasmid of 158,496 bp with 2.12 plasmid copies per chromosome (GC content, 51.8%), and a 1,739-bp plasmid with 16.92 copies per chromosome

**Editor** David Rasko, University of Maryland School of Medicine

**Copyright** © 2022 Lovero and Mota-Bravo. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](#).

Address correspondence to Luis Mota-Bravo, [lmota@uci.edu](mailto:lmota@uci.edu).

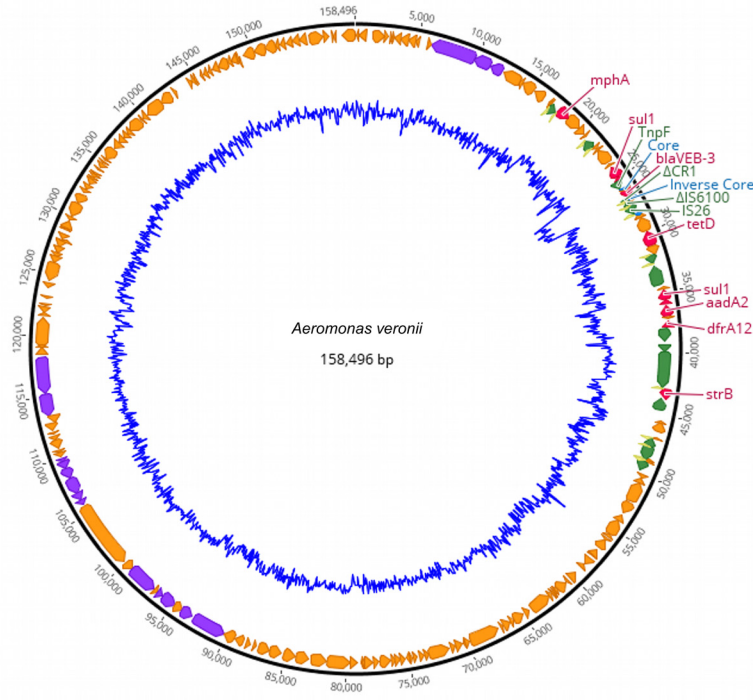
The authors declare no conflict of interest.

**Received** 18 October 2021

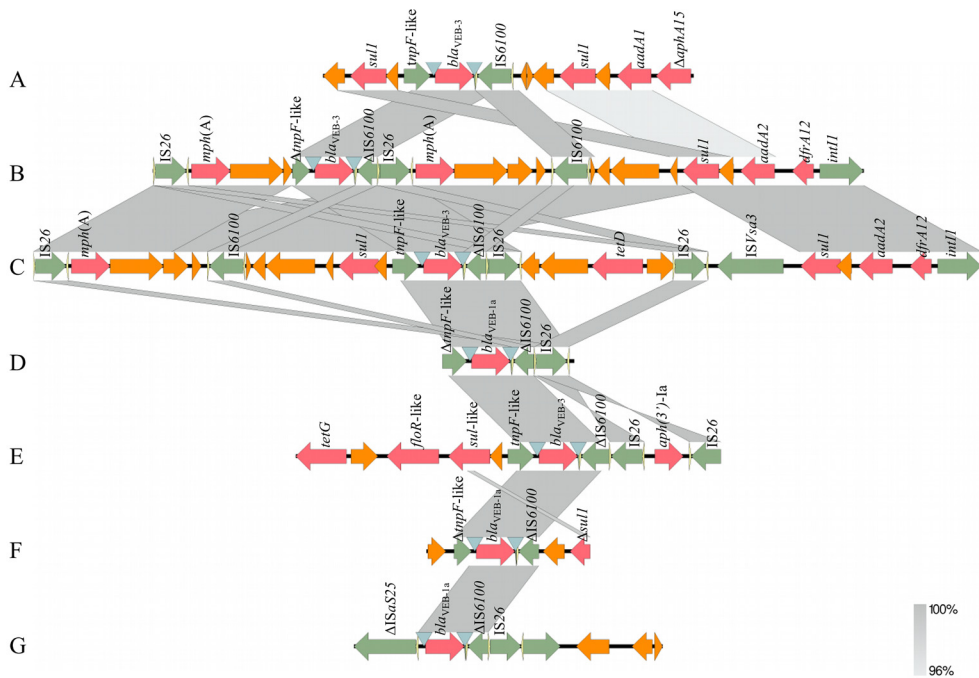
**Accepted** 5 February 2022

**Published** 23 February 2022

**A**



**B**



**FIG 1** (A) Genetic map of p158496, displayed in the outermost ring. Open reading frames are represented by arrows in the direction of transcription; they are color coded according to their putative functions, as follows: purple, conjugation machinery; red, ARGs; green, MGEs; yellow, inverted repeats; orange, all other coding sequences. The inner blue ring displays the GC content of the plasmid. Genes of interest are labeled. This figure was created using Geneious v11.1.5. (B) Schematic representation of isolates from the NCBI GenBank database containing the *bla<sub>VEB</sub>* gene with regions homologous to those found in p158496. Open reading frames are represented by arrows in the direction of transcription; they are color coded according to their putative functions, as follows: red, ARGs; green, MGEs; yellow, inverted repeats; orange, all other coding sequences. Inverted blue triangles represent the recombination core and inverse core sites. The identity between adjacent sequences is shown as gray shading. Δ indicates the truncation of a gene. Descriptions of isolates A through G are presented in Table 1. This figure was created using EasyFig.

**TABLE 1** Descriptions of isolates listed in Fig. 1B

Isolate	GenBank accession no.	Species	Country	Isolation source	Collection date(s)	Genetic location
A	<a href="#">GQ926879.1</a>	<i>Acinetobacter pittii</i>	Taiwan	Blood of hospital patient	1999–2007 <sup>a</sup>	Plasmid
B	<a href="#">CP018201.1</a>	<i>Aeromonas hydrophila</i>	China	Water	2012	Chromosome
C	<a href="#">CP083462.1</a>	<i>Aeromonas veronii</i>	USA	Lake in recreational park	2015	Plasmid
D	<a href="#">HM370390.1</a>	<i>Aeromonas caviae</i>	France	Seine river	2009	Chromosome
E	<a href="#">CP006657.1</a>	<i>Klebsiella pneumoniae</i>	China	Blood of hospital patient	2010	Plasmid
F	<a href="#">HM370392.1</a>	<i>Aeromonas allosaccharophila</i>	France	Seine river	2009	Plasmid
G	<a href="#">HM370391.1</a>	<i>Aeromonas veronii</i>	France	Seine river	2009	Plasmid

<sup>a</sup>Clinical isolates were collected from three hospitals in Taiwan from 1999 to 2007.

(GC content, 56.4%). ARGs found in the IncA/C plasmid were *aadA2*, *bla*<sub>VEB-3</sub>, *sul1*, *tetD*, *dfra12*, *mphA*, and *strB* (Fig. 1A). The plasmid conferred resistance to five classes of antibiotics, classifying the strain as multidrug resistant (15). The *bla*<sub>VEB-3</sub> gene was flanked by MGEs IS6100 and IS26 and a TnpF-like integrase (Fig. 1B). Similar genetic neighborhoods were found in other environmental and clinical bacteria (Fig. 1B and Table 1). The association of the VEB extended-spectrum  $\beta$ -lactamase (ESBL) with MGEs on an IncA/C plasmid highlights the potential for environmental *Aeromonas* strains to harbor and disseminate ARGs.

**Data availability.** The genome sequence data for SW3814 have been deposited in NCBI GenBank under BioProject accession number [PRJNA762937](#), BioSample accession number [SAMN21418941](#), SRA accession number [PRJNA762937](#), and GenBank accession numbers [CP083461](#) (assembled chromosome), [CP083462](#) (plasmid p158496), and [CP083463](#) (plasmid p1739).

## ACKNOWLEDGMENTS

This study was supported by NIH grant GM055246, awarded to L.M.-B.

L.M.-B. conceived the main conceptual ideas and methodology, created a platform for data curation, acquired funding, provided resources, helped supervise and provide project administration, validated the results, and wrote the final manuscript. K.G.L. performed antimicrobial susceptibility testing, formal analysis of the sequenced plasmid, interpretation of the results, visualization, and writing of the manuscript.

## REFERENCES

- Gomes S, Fernandes C, Monteiro S, Cabecinha E, Teixeira A, Varandas S, Saavedra M. 2021. The role of aquatic ecosystems (River Tua, Portugal) as reservoirs of multidrug-resistant *Aeromonas* spp. *Water* 13:698. <https://doi.org/10.3390/w13050698>.
- Jacobs L, Chenia HY. 2007. Characterization of integrons and tetracycline resistance determinants in *Aeromonas* spp. isolated from South African aquaculture systems. *Int J Food Microbiol* 114:295–306. <https://doi.org/10.1016/j.ijfoodmicro.2006.09.030>.
- Piotrowska M, Popowska M. 2014. The prevalence of antibiotic resistance genes among *Aeromonas* species in aquatic environments. *Ann Microbiol* 64:921–934. <https://doi.org/10.1007/s13213-014-0911-2>.
- Piotrowska M, Popowska M. 2015. Insight into the mobilome of *Aeromonas* strains. *Front Microbiol* 6:494. <https://doi.org/10.3389/fmicb.2015.00494>.
- Rhodes G, Huys G, Swings J, McGann P, Hiney M, Smith P, Pickup RW. 2000. Distribution of oxytetracycline resistance plasmids between aeromonads in hospital and aquaculture environments: implication of Tn1721 in dissemination of the tetracycline resistance determinant Tet A. *Appl Environ Microbiol* 66:3883–3890. <https://doi.org/10.1128/AEM.66.9.3883-3890.2000>.
- Schmidt AS, Bruun MS, Dalsgaard I, Larsen JL. 2001. Incidence, distribution, and spread of tetracycline resistance determinants and integron-associated antibiotic resistance genes among motile aeromonads from a fish farming environment. *Appl Environ Microbiol* 67:5675–5682. <https://doi.org/10.1128/AEM.67.12.5675-5682.2001>.
- McIntosh D, Cunningham M, Ji B, Fekete FA, Parry EM, Clark SE, Zalinger ZB, Gilg IC, Danner GR, Johnson KA, Beattie M, Ritchie R. 2008. Transferable, multiple antibiotic and mercury resistance in Atlantic Canadian isolates of *Aeromonas salmonicida* subsp. *salmonicida* is associated with carriage of an IncA/C plasmid similar to the *Salmonella enterica* plasmid pSN254. *J Antimicrob Chemother* 61:1221–1228. <https://doi.org/10.1093/jac/dkn123>.
- Maniatis T, Fritsch EF, Sambrook JK. 1982. *Molecular cloning: a laboratory manual*, p 191–195. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ pre-processor. *Bioinformatics* 34:i884–i890. <https://doi.org/10.1093/bioinformatics/bty560>.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
- Tatusova T, DiCuccio M, Badretdin A, Chetverin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Li W, O'Neill KR, Haft DH, DiCuccio M, Chetverin V, Badretdin A, Coulouris G, Chitsaz F, Derbyshire MK, Durkin AS, Gonzales NR, Gwadz M, Lanczycki CJ, Song JS, Thanki N, Wang J, Yamashita RA, Yang M, Zheng C, Marchler-Bauer A, Thibaud-Nissen F. 2021. RefSeq: expanding the Prokaryotic Genome Annotation Pipeline reach with protein family model curation. *Nucleic Acids Res* 49:D1020–D1028. <https://doi.org/10.1093/nar/gkaa1105>.
- Haft DH, DiCuccio M, Badretdin A, Brover V, Chetverin V, O'Neill K, Li W, Chitsaz F, Derbyshire MK, Gonzales NR, Gwadz M, Lu F, Marchler GH, Song JS, Thanki N, Yamashita RA, Zheng C, Thibaud-Nissen F, Geer LY, Marchler-Bauer A, Pruitt KD. 2018. RefSeq: an update on prokaryotic genome

- annotation and curation. *Nucleic Acids Res* 46:D851–D860. <https://doi.org/10.1093/nar/gkx1068>.
14. Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. *J Antimicrob Chemother* 67:2640–2644. <https://doi.org/10.1093/jac/dks261>.
15. Magiorakos A-P, Srinivasan A, Carey RB, Carmeli Y, Falagas ME, Giske CG, Harbarth S, Hindler JF, Kahlmeter G, Olsson-Liljequist B, Paterson DL, Rice LB, Stelling J, Struelens MJ, Vatopoulos A, Weber JT, Monnet DL. 2012. Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. *Clin Microbiol Infect* 18:268–281. <https://doi.org/10.1111/j.1469-0691.2011.03570.x>.