Anoxic carbon flux in photosynthetic microbial mats as revealed by metatranscriptomics

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Abstract

Photosynthetic microbial mats possess extraordinary phylogenetic and functional diversity that makes linking specific pathways with individual microbial populations a daunting task. Close metabolic and spatial relationships between *Cyanobacteria* and *Chloroflexi* have previously been observed in diverse microbial mats. Here we report that an expressed metabolic pathway for the anoxic catabolism of photosynthate involving *Cyanobacteria* and *Chloroflexi* in microbial mats can be reconstructed through metatranscriptomic sequencing of mats collected at Elkhorn Slough, Monterey Bay, CA. In this reconstruction, *Microcoleus* spp., the most abundant cyanobacterial group in the mats, ferment photosynthate to organic acids, CO_2 and H_2 through multiple pathways, and an uncultivated lineage of the *Chloroflexi* take up these organic acids to store carbon as polyhydroxyalkonates (PHAs). The metabolic reconstruction is consistent with metabolite measurements and single cell microbial imaging with flu**o**rescence *in situ* hybridization and NanoSIMS.

Introduction

Linking biogeochemical processes observed in natural microbial communities with their associated metabolic pathways and assigning these pathways to specific groups remains a daunting task, particularly as community complexity increases (Kunin *et al.,* 2008a). Photosynthetic microbial mats found in marine and hypersaline environments are stratified microbial communities that harbor extraordinary phylogenetic and functional diversity, harboring up to 42 phyla, including many candidate divisions (Dillon *et al.,* 2009, Kunin *et al.,* 2008b, Ley *et al.,* 2006, Nicholson *et al.,* 1987). Extensive measurements have been performed on these communities, documenting biogeochemical cycling of carbon, nitrogen and sulfur (Canfield and Des Marais 1993, Decker *et al.,* 2005, Hoehler *et al.,* 2001, Hoehler *et al.,* 2002, Jorgensen and Des Marais 1986). The microbial populations involved in these biogeochemical cycles have been primarily inferred based on the physiology of isolates and expression of individual functional genes (Green *et al.,* 2008, Omoregie *et al.,* 2004, Orphan *et al.,* 2008). However, a comprehensive understanding of element cycling in the mats requires reconstruction of complete metabolic pathways expressed by specific microbial groups.

Photosynthetic microbial mats perform light driven carbon fixation during daylight hours (Bebout *et al.,* 2002). Photosynthate is accumulated, often as glycogen, through both oxygenic and anoxygenic photosynthesis (Nicholson *et al.,* 1987). This photosynthate is fermented to organic acids and H_2 under anoxic conditions at night (Burow *et al.,* 2012, Hoehler *et al.,* 2002). The flux through these fermentation products varies up to four orders of magnitude between day and night (Hoehler *et al.,* 2001). Recent studies of mats collected at Elkhorn Slough, Monterey Bay, CA has provided

persuasive evidence that *Microcoleus* spp., the most abundant cyanobacterial group in the mats, are the dominant fermenters (Burow *et al.*, 2012). However, the pathways through which *Microcoleus* spp. ferment photosynthate are unknown, as are microbial groups and pathways that may further metabolize the organic acids derived from fermentation. Relative measures of activity comparing rRNA transcripts to genes in samples from the Elkhorn Slough mats have indicated that *Chloroflexi* are among the most active microbial group under dark, anoxic conditions (Burow *et al.*, 2012). Studies in thermophilic mat systems collected in Yellowstone National Park, which are much less complex than marine and hypersaline mats, have suggested a link between anoxic fermentation of accumulated photosynthate by *Cyanobacteria* and carbon uptake by *Chloroflexi* (van der Meer *et al.,* 2003, van der Meer *et al.,* 2005, van der Meer *et al.,* 2007). The intimate physical association of *Cyanobacteria* and *Chloroflexi* was demonstrated in hypersaline mats collected from Guerrero Negro, Baja California, where fluorescence *in situ* hybridization was used to visualize *Chloroflexi* filaments within the exopolysaccharide sheaths of *Microcoleus* spp. (Ley *et al.,* 2006). The close links between *Cyanobacteria* and *Chloroflexi* observed in other microbial mat environments suggest that this association may be a general phenomenon observed in mats and that this association may play an important role in anoxic carbon cycling.

Metatranscriptomic analysis has been used to characterize active microbial communities in aquatic and terrestrial environments (Frias-Lopez *et al.,* 2008, Gilbert *et al.,* 2008, Poretsky *et al.,* 2005, Turnbaugh *et al.,* 2010, Urich *et al.,* 2008). Gene expression from microbial assemblages has been correlated with biogeochemical data and the phylogeny of expressed genes affiliated with specific populations (Hollibaugh *et al.,*

2011, Liu *et al.,* 2011, McCarren *et al.,* 2010, Mou *et al.,* 2011, Poretsky *et al.,* 2009, Poretsky *et al.,* 2010, Shi *et al.,* 2011). In engineered microbial systems where complexity is reduced, the expression of specific pathways has been defined and correlated with environmental conditions (He *et al.,* 2010). Metatranscriptomics of microbial mats offer the possibility of defining key metabolic pathways in the biogeochemical cycling of carbon and of establishing a link in the carbon cycle between *Cyanobacteria* and *Chloroflexi*.

Here, we demonstrate that metatranscriptomic data obtained from Elkhorn Slough microbial mat samples collected under dark anoxic conditions were dominated by transcripts from *Cyanobacteria* and *Chloroflexi*. This density of sequenced transcripts allowed us to reconstruct pathways for anoxic carbon catabolism that link fermentation in the *Cyanobacteria* with organic acid uptake and carbon storage in the *Chloroflexi*. These metabolic pathway inferences were supported by ^{13}C-acetate isotopic labeling studies of the Elkhorn Slough mat coupled with single cell microbial imaging.

Methods

Sample site and collection

The sampling site was located in the Elkhorn Slough estuary in Central California, USA, at 36N 48' 46.61" (decimal degree Lat 36.81295) and -121S 47' 4.89" (decimal degree Long -121.7847). Microbial mats were sampled on 10th of January 2009 and transported to a greenhouse facility at NASA Ames Research Center within 1 to 2 hours. In the greenhouse, mat pieces were placed in acrylic aquaria transparent to ultraviolet radiation and covered with water for ca. 20 h before the beginning of a diel cycle study (Bebout *et al.,* 2002, Burow *et al.,* 2012). Diel cycle studies were carried out under natural solar irradiance, and the water temperature was kept constant at ca. 18 °C (*in situ* average). The data presented result from diel cycle studies carried out on the 12th/13th January and 13th/14th January 2009.

Biogeochemical assays

Replicate vials were prepared identically as follows: small sub-cores (11 mm diameter, 15 mm depth or vertically sectioned for depth profile analyses) were cut from whole sections of intact microbial mat and placed in serum vials with 4 mL of field site water. Serum vials were capped with butyl rubber stoppers. The 10.5 mL headspace of the serum vials was left as air for light/daytime incubations and was thoroughly flushed with N_2 (gas and liquid phase degassed) for dark/nighttime incubations. H_2 was allowed to accumulate in three replicate vials and was repeatedly sampled at specific time points over the course of the diel experiment. Organic acids were allowed to accumulate in replicate vials (separate to vials in which H_2 was measured), with three vials sacrificed at each time point. Sections of microbial mat were also incubated individually in triplicate

vials as described above after sectioning to identify the location of H_2 production. Sectioning using a sterile scalpel blade yielding the following mat layers; 0-2 mm, 2-4 mm and 4-15 mm. To measure H_2, 25 μL of headspace gas was withdrawn by volumetric syringe and injected directly onto a gas chromatograph with quantification by a mercuric oxide detector. To analyze organic acids, the entire liquid phase (4 mL) was sampled (with the associated incubation sacrificed). Liquid was filtered through 0.2 μm syringe-driven filters for storage in glass vials at -20^{O}C. Organic acids (C1-5) were quantified via high-pressure liquid chromatography (Albert and Martens 1997).

Nucleic acid extraction from Elkhorn Slough mat cores

Ten mat cores of 1 cm diameter were flash frozen in liquid nitrogen at multiple time points during the diel cycle and stored at -80^{o}C until further processing. RNA of two samples (BN; 9:00 pm, 12th January 2009; 4 hrs after sunset, and EN; 7:00 am, 13th January 2009, 20 min before dawn) was extracted from the uppermost 2 mm of 5 mat cores by combining phenol-chloroform extraction with parts of the RNeasyMini kit (Qiagen, Valencia, CA, USA). Per core, biomass was transferred in a tube containing 0.5 ml RLT ^{TM} buffer and homogenized using a rotor-stator homogenizer (Omni International, Kennesaw, GA, USA). The suspension was then bead-beated with zirconium beads (200 μm, OPS Diagnostics, Lebanon, **NJ**, USA) and the cell debris and beads pelleted. The supernatant was extracted with phenol-chloroform-isoamyl alcohol (125:24:1, pH 4.5, Ambion, Austin, TX, USA) The aqueous phase was run through the gDNA eliminator spin column to remove genomic DNA **and** further purified following the RNeasyMini kit protocol. Extracted RNA was treated with DNase using the TURBO

DNA-free kit according to protocol (Ambion). **Amplification, sequencing and sequence analysis of 16S rRNA genes and transcripts is described in Supplemental Information.**

Selective removal of rRNA and cDNA synthesis

The MICROB*Enrich* and MICROB*Express*™ Kits (Ambion) were used to remove the ribosomal RNAs (rRNA) by a subtractive hybridization approach, thereby enriching the messenger RNA (mRNA) in the total RNA pool. In the MICROB*Enrich* protocol, 7 additional eukaryotic capture probes, and in the MICROB*Express*™ Kit, 5 additional cyanobacterial capture probes were used. The sequences can be found in Supplementary Information. ~400 ng of mRNA were linearly amplified with the MessageAmp II-Bacteria Kit (Ambion) according to the manufacturer's instructions. 4 µg of the amplified, antisense RNA (aRNA) were converted to double-stranded cDNA with random hexamers in multiple replicates using the SuperScript® Double-Stranded cDNA Synthesis Kit (Invitrogen, Carlsbad, CA, USA) and purified with DNA Purification Spin Columns.

The quality and quantity of total RNA, mRNA, aRNA and cDNA were verified by measurements on the NanoDrop-1000 Spectrophotometer (NanoDrop Technologies, Wilmington, DE), with the Qubit fluorometer (Life Technologies, Grand Island, NY, USA) and the Bioanalyzer (Agilent Technologies, Santa Clara, CA, USA).

Metatranscriptomic sequencing and analysis

Double stranded cDNA of the two samples (BN, 12th January 2009, 9:00 pm and EN,

13th January 2009, 7:00 am) were sequenced at the Department of Energy Joint Genome Institute (JGI, Walnut Creek, CA, USA) on a 454 Genome Sequencer FLX Instrument (454 Life Sciences, **Branford, CT, USA**) following standard procedures. After sequencing, ribosomal RNA reads were detected by BLAST search and removed from the data set. Non-rRNA sequences were analyzed for replicate sequences using the open-source program CD-HIT provided as a web-based tool at http://microbiomes.msu.edu/replicates according to a previously described protocol (Gomez-Alvarez *et al.,* 2009, Li and Godzik 2006). Replicates were defined as sequences sharing greater than 99% nucleotide identity, with an allowable length difference of 2 bp, and a requirement that the first 3 bp of the replicate sequences be identical.

The BLASTX program in BLAST version 2.2.21 was used with dereplicated non-rRNA reads as query sequences against all amino acid sequences in the microbial RefSeq release 45 (January 2011) database (Altschul *et al.,* 1990, Pruitt *et al.,* 2009). RefSeq BLAST hits with bitscores below 40 were removed, and results were processed using MEGAN version 4.32 with default settings (Huson *et al.,* 2007). Taxonomic assignments based on NCBI taxonomy and functional assignments based on SEED and KEGG were analyzed using graphical tools within MEGAN (Ogata *et al.,* 1999, Overbeek *et al.,* 2005).

Completeness of sampling estimates were made using Chao and ACE estimators in MOTHUR version 1.21.1 (Schloss *et al.,* 2009). Read clusters used as input for MOTHUR were generated in two ways. Firstly, reads with a top blastx hit to the same RefSeq protein sequence were considered part of the same mRNA transcript cluster and used to estimate absolute coverage. Secondly, reads unambiguously assigned by MEGAN

to a single SEED functional category or species (as defined by NCBI taxonomy) were considered part of the same functional or species cluster and used to calculate functional and species **sampling completeness** respectively.

FISH-SIMS

Small sub-cores (11 mm diameter, 15 mm depth) of the microbial mats were cut from whole sections of intact microbial mat and placed in serum vials with 4 mL water obtained from the field site. Serum vials were capped with butyl rubber stoppers and the headspace of the vials (10.5 mL) was thoroughly flushed with N_2 (gas and liquid phase degassed) for dark/nighttime incubations. A stable isotope tracer, [2-^{13}C]acetate (99% ^{13}C, Sigma, St. Louis, USA), was added to the liquid phase of vials containing live cores at a final concentration of 0.2 mM. As a control, cores pretreated with 4 mM paraformaldehyde (final concentration) were also incubated with 0.2 mM [2-^{13}C]acetate. The cores were incubated under dark, anoxic conditions for 10 hours. The incubation was stopped by adding 4 mM paraformaldehyde to the live cores. Preliminary NanoSIMS analysis revealed that single cells in the live cores were labeled but not in the killed cores pretreated with paraformaldehyde. The method used to label *Chloroflexi* with fluorescent oligonucleotides was previously described (Woebken *et al.,* 2012). *Chloroflexi* were targeted using equimolar amounts of the FISH probes CFX1223 and GNSB-941 (Björnsson *et al.,* 2002). High-resolution secondary ion mass spectrometry (SIMS) was performed at LLNL with a Cameca NanoSIMS 50. The $^{13}C/^{12}C$ ratio was measured using $^{12}C_2$ and $^{13}C^{12}C$ and correcting for the dimer abundances (Finzi-Hart *et al.,* 2009)

Data Deposit

The dereplicated metatranscriptomic sequences with rRNA sequences removed can be downloaded from IMG/M (http://img.jgi.doe.gov/cgi-bin/m/main.cgi) under the names Elkhorn Slough cyanobacterial mat night (9pm Metatranscriptome CGUI) (BN library) and Elkhorn Slough cyanobacterial mat Day (7am transcriptome CGUN) (EN library) at the tab "FASTA nucleic acid file for all scaffolds". 16S rRNA gene sequences obtained in this study are deposited under GenBank accession numbers JX002103– JX002655.

Results

Microbial Mat Characterization

H_2 production was >10x higher at night in the upper phototrophic layer (0-2 mm) in the Elkhorn Slough mat compared to H_2 produced during the day or in the lower mat layers (Supplemental Figure 1A). Acetate, formate and propionate were concomitantly produced with H_2, whereas lactate, butyrate and valerate were not detected (200 nM detection limit; Supplemental Figure 1B). H_2 and organic acid measurements observed for the microbial mat sampled in January 2009 were similar to those previously reported from this field site in November 2009 (Burow *et al.,* 2012).

The microbial community composition in the upper layer was assessed using amplicon pyrosequencing of the V8 region of SSU rRNA genes from a time point taken before sunrise (EN; 0700, January 13, 2009); sequences grouping with 16 distinct phyla were observed (Supplemental Table 1 and Figure 1A). The active microbial community was investigated by amplicon pyrosequencing of the V8 region of SSU rRNA from cDNA derived from RNA extracted at the same time point (EN) and at the beginning of the dark period (BN; 2100; January 12, 2009). The active communities were subsets of the total number of phyla (11 and 13 phyla, respectively) and dominated by *Cyanobacteria*. *Chloroflexi* sequences were present at low abundance in both the DNA and cDNA pyrotag libraries.

Nearly full-length 16S rRNA gene sequences were consistent with amplicon pyrosequencing of the cDNA (Figure 1A). Cyanobacterial sequences clustered into a few groups (Figure 2), of which the majority were related to *Microcoleus chthonoplastes* PCC 7420 and a cyanobacterium recently isolated from the Elkhorn Slough mats (ESFC-1)

(Woebken *et al.,* 2012). The *Chloroflexi*-related sequences were only distantly related to cultured representatives, but were closely related **to** sequences from uncultivated *Chloroflexi* recovered from hypersaline environment at Guerrero Negro, Baja California (Figure 3) (Ley *et al.,* 2006).

Metatranscriptome Sequence Analysis

A total of 590,783 and 492,302 metatranscriptome sequence reads were generated for the BN and EN time points of which 43% (BN) and 32% (EN) were non-ribosomal sequences (Supplemental Table 2). After removal of these rRNA sequences and de-replication, 244,004 (BN) and 144,017 (EN) sequences remained for further analysis.

Coverage of the total pool of unique mRNA transcripts in the samples was analyzed in three ways: absolute, functional, and species-level taxonomic. Absolute coverage was estimated by BLAST against the microbial RefSeq database, where reads yielding top BLAST hits from the same RefSeq amino acid sequence were considered part of the same mRNA transcript. This RefSeq BLAST is likely to underestimate transcript diversity, as only 64.6% (BN) and 68.7% (EN) of reads for each sample showed significant similarity (bitscore > 40) to any RefSeq sequence (Supplemental Table 3). Collector's curves showed that sampling of the total number of unique mRNA transcripts is far from complete (Supplemental Figure 2A).

Estimations of functional and species-level coverage were obtained using the unambiguous assignment by MEGAN of reads to SEED functional categories or species. Although only 37.9%-40.4% of reads were given a functional assignment and 15.6%-22.9% given an unambiguous species level assignment, both time points were sampled

relatively completely with respect to the functional and species-level **diversity presently described in RefSeq** (Supplemental Table 3, Supplemental Figure 2B). These results are likely a reflection of the limited scope of the SEED and RefSeq databases compared to the full natural range of functions and species, rather than of the true completeness of sampling. Moreover, although functions and species were well sampled independently, by no means were all the functions for every species sampled as well. However, these results do mean that these metatranscriptomic samples are broadly representative of those functions and species present in both the sample and SEED/RefSeq databases.

Abundant taxa in metatranscriptomes

The taxonomic composition of the microbial mat was assessed using **the MEGAN software package for all** protein-coding transcripts. Both metatranscriptomes were dominated by reads assigned to *Cyanobacteria* and *Chloroflexi*, the former constituting 22-47% of the reads, the latter 15-33% (Figure 1B). *Proteobacteria* and *Bacteroidetes*-affiliated reads were the next abundant groups. A large proportion of the reads were recruited to the *M. chthonoplastes* PCC 7240 genome. Of the 8,294 predicted protein-coding genes in the *M. chthonoplastes* genome, 4,030 (49%) were detected in the BN library and 1,821 (24%) were detected in the EN library. Estimates of richness using Chao and ACE indices predicted the expression of 4,978 and 4,948 *M. chthonoplastes* protein-coding genes in the BN transcriptome, suggesting 81% (Chao) or 80% (ACE) coverage. These indices predicted 62% (Chao) and 48% (ACE) coverage in the EN transcriptome. Transcripts associated with *Microcoleus* spp. represented 24% (n = 37,524) of all BN transcripts with matches to the RefSeq database (n = 157, 525) and 8% (n = 7, 581) of all EN transcripts with matches to the RefSeq database (n = 98,894).

These reads are not necessarily all derived from a single *Microcoleus* relative, and may reflect a diverse pool of *Microcoleus* spp. that are all better related to *M. chthonoplastes* PCC 7240 than to any other sequenced cyanobacterial genome. Unlike the *Microcoleus* spp., the *Chloroflexi* populations present in the mat samples did not have a closely related genome for fragment recruitment. Within the metatranscriptome reads affiliated with the phylum *Chloroflexi*, 96% were assigned to the class *Chloroflexi* at both time points, and ca. 85% of those reads further to the order *Chloroflexales* (BN; n = 18,769 and EN; n = 26,981, respectively; Supplemental Figure 3).

Anoxic Carbon Catabolism by Cyanobacteria and Chloroflexi

Transcript reads from *Microcoleus* spp. associated with carbohydrate metabolism were overrepresented when compared to carbohydrate reads observed for the whole community (BN; n = 18% and EN; n = 22%; **Supplemental Figure 4**). To elucidate the carbon catabolic pathways in the *Microcoleus* populations in the mat samples, fermentation pathways for conversion of glycogen were reconstructed from the *M. chthonoplastes* genome (Figure 4A; Supplemental Table 4-5). Genes for glycogen phosphorylase and the Embden-Meyerhof-Parnas (EMP) pathway coded for proteins for the conversion of stored photosynthate, glycogen, to pyruvate. Genes for pyruvate transformation via three pathways were identified: to lactate (lactate dehydrogenase), to formate and acetyl-CoA (pyruvate-formate lyase) and to acetyl-CoA and CO_2 (pyruvate-ferredoxin oxidoreductase). Genes to transform acetyl-CoA to ethanol (alcohol/aldehyde dehydrogenase) and acetate (acetate kinase) were identified. No annotated phosphotransacetylase was detected and no reads that cluster with other cyanobacterial

phosphotransacetylases were identified in the transcript reads. The catalytic [NiFe] Hox hydrogenase was identified as part of a predicted seven gene operon with NAD(P)H dehydrogenase subunits (*hoxEFUYH*), a predicted aldehyde/alcohol dehydrogenase (*adhE*) and a hypothetical protein. Transcripts for all the genes described above were observed in the BN and EN libraries (Figure 5, Supplemental Table 6). Higher numbers of reads for the fermentative pathway genes were observed in the BN sample, consistent with the higher level of *Microcoleus* expression at the beginning of the dark period. The *Microcoleus* genes were the majority of all the cyanobacterial-associated reads assigned to genes in the fermentation pathway.

A number of *Chloroflexales*-associated reads were assigned as acetate- and lactate-permeases, consistent with the hypothesis that these microbes take up the organic acids excreted during fermentation by *Microcoleus* spp. (Figure 4B; Supplemental Table 4-6). Organic acid uptake is often associated with production of polyhydroxyalkonoates (PHAs) that act as carbon storage compounds and uptake of acetate has been shown to be involved in the conversion of glycogen to PHAs in bacteria responsible for enhanced biological **phosphorus** removal (EBPR) (Mino *et al.,* 1998). Pathways for the biosynthesis of PHAs were reconstructed from multiple *Chloroflexales* genomes and transcripts were identified for *phaA* (acetoacetyl-CoA transferase), *phaB* (acetoacetyl-CoA reductase) and *phaC* (PHA synthase) in both the BN and EN libraries (Figure 5). The identification of *Chloroflexales*-associated transcripts for glycogen phosphorylase in both libraries implies that reducing equivalents required for PHA production in these bacteria derive from the breakdown of glycogen. Transcripts for acetyl-CoA synthetase (*acs*), which converts acetate to acetyl-CoA, were also detected in both libraries.

Accumulation of glycogen by the *Chloroflexi* is proposed to occur during anoxygenic photosynthesis to store carbon and reducing equivalents for dark, anoxic metabolism (Sirevag and Castenholz 1979, Taffs *et al.,* 2009). These reducing equivalents are generated by glucose metabolism through the EMP pathway to produce pyruvate, which is oxidized to acetyl-CoA by pyruvate dehydrogenase, which is detected in both libraries.

^{13}C-Acetate Uptake by Chloroflexi Observed by FISH-SIMS

The reconstructed pathway described above suggested that the *Chloroflexi* in the Elkhorn Slough mat assimilated organic acids under dark, anoxic conditions. **Single cell analysis using NanoSIMS was used to test the specific hypothesis that acetate, the most abundant organic acid observed in the mats, was assimilated by the *Chloroflexi***. Mat samples were incubated with 0.2 mM ^{13}C-acetate during the dark portion of the diel cycle **and *Chloroflexi* cells** in these samples were stained by the CARD-FISH procedure with *Chloroflexi*-specific oligonucleotide probes (Figure 6A) and identified by confocal laser scanning microscopy. **This analysis revealed a morphological diversity consistent with the phylogenetic diversity detected in nearly full-length 16S rRNA sequences from Elkhorn Slough mats (Figure 3), in which the sequence identity of the *Chloroflexi* phylotypes ranged from 76.6%-89.6%.** The identified *Chloroflexi* were targets for NanoSIMS analysis measuring ^{13}C/^{12}C in individual bacterial cells. These FISH-SIMS experiments demonstrated specific uptake of ^{13}C-lableled acetate into *Chloroflexi* cells (δ^{13}C = 260 ±35‰; n=40), with the observed ^{13}C-label in *Microcoleus* (-15.8 ±3.0‰; n=12) and other cyanobacterial cells (-14.0 ±2.8‰; n=12) close to natural abundance (Figure 6B). **Additional single cells were**

enriched in ^{13}C above natural abundance, but did not stain with the *Chloroflexi*-specific probes.

Discussion

In this study, we have taken a highly diverse microbial system and used metatranscriptomic sequencing to identify genes in the two most abundant taxa, *Cyanobacteria* and *Chloroflexi*, which are responsible for anoxic catabolism of photosynthate. The success of this reconstruction is remarkable because of the relatively modest amount of sequencing (~400 MB) used. It is highly unlikely that a comparable amount of metagenomic sequencing would have provided a resolved metabolic pathway because the diversity of the community would prevent assembly of large contiguous sequences (Kunin *et al.*, 2008b). Metatranscriptomic sequencing focuses on the most highly expressed genes, **which can resolve important metabolic processes.** Central to the success of this strategy was the availability of isolate genome sequences for *Microcoleus* and the *Chloroflexales* that facilitated metabolic pathway reconstruction and fragment recruitment of transcripts. A notable discrepancy in the metatranscriptomic data obtained from Elkhorn Slough samples is the difference in relative abundance of the rRNA transcripts, which identify *Cyanobacteria* as the dominant active population, compared to the mRNA transcripts, which fluctuate between *Cyanobacteria* and *Chloroflexi* as the dominant populations. This phenomenon has been observed, though less dramatically, in metatranscriptomic studies of a microbial mat from Mushroom Spring in Yellowstone (Liu *et al.*, 2011). The dominance of cyanobacterial rRNA may reflect the synthesis of large numbers of ribosomes by cyanobacterial cells, which have

larger cell sizes in comparison to other bacterial cells. Cell size and ribosome content have previously been directly correlated in prokaryote cells (Ecker and Kokaisl 1969). Alternatively, it could reflect an amplification bias in the PCR reaction with cDNA from the mats. However, since two different forward primers were used for pyrotag sequencing and Sanger clone libraries, this explanation is not very likely.

Combining the metatranscriptomic data and metabolic pathway reconstruction with metabolite and single cell measurements enabled us to propose an anoxic catabolic pathway for carbon fixed during the light period by oxygenic and anoxygenic photosynthesis. In this pathway, photosynthate accumulated during the day is stored as glycogen by both *Microcoleus* spp., derived from oxygenic photosynthesis and the *Chloroflexi*, derived from anoxygenic photosynthesis. Under dark anoxic conditions, *Cyanobacteria* depolymerize and ferment the glycogen, excreting H_2 and organic acids and generating ATP for cellular maintenance. *Chloroflexi* depolymerize glycogen, generating reducing equivalents to transform organic acids cross-fed from the *Cyanobacteria* to PHAs, generating ATP for cellular maintenance. Our analysis does not exclude other pathways for anoxic metabolism of organic acids, such as sulfate reduction by δ-proteobacteria, but these pathways could not be completely reconstructed from the metatranscriptomic datasets (Risatti *et al.,* 1994).

Reconstruction of fermentation pathways from the *M. chthonoplastes* genome identified multiple pathways for pyruvate fermentation, all of which were expressed in the mats samples analyzed by metatranscriptomics. Therefore the predominant organic acid end products of pyruvate fermentation were predicted to be acetate, formate and lactate. Acetate and formate were observed, consistent with the predicted activity of

pyruvate ferredoxin oxiodoreductase (PFR), pyruvate formate-lyase (PFL) and pyruvate dehydrogenase complex (PDC). However, no lactate was observed, consistent with both the low number of observed lactate dehydrogenase transcripts and the detection of reads for lactate permeases from the *Chloroflexi*. Propionate production was also observed, which suggests that *Microcoleus* may have a pathway to ferment pyruvate to propionate, similar to pathways present in propinobacteria (Himmi *et al.,* 2000). Interestingly, no genes in the *Microcoleus* genome or cyanobacterial transcripts in the two datasets could be assigned as a phosphotransacetylase despite the identification of multiple pathways in *Microcoleus* to produce acetyl-CoA and the production of high levels of acetate in the mat samples. This observation suggests that an unassigned protein may substitute for phosphotransacetylase in *Microcoleus* fermentation. The recruitment of metatranscriptomic reads provides persuasive evidence that H_2 is generated by a NAD(P)H-dependent Hox hydrogenase, consistent with previous studies which utilized PCR and microarray-based methods (Burow *et al.,* 2012, Marshall *et al.,* 2012) The expression of PFR may imply that reducing equivalents for H_2 are generated by oxidation of pyruvate to acetyl-CoA, as is observed for [FeFe] hydrogenases (Schut and Adams 2009). However, the specific interactions between PFR and the Hox hydrogenase are unknown, despite the co-occurrence of PFR with Hox in all available cyanobacterial genomes (Carrieri *et al.,* 2011).

An important hypothesis arising from analysis of the metatranscriptomic data was that the *Chloroflexi* should take up organic acids under dark, anoxic conditions. The linkage of organic acid uptake to anoxic conversion of glycogen to PHA has previously been observed in bacteria performing EBPR during wastewater treatment (Mino *et al.,*

1998). The postulated link between fermentation products excreted by *Microcoleus* and carbon compounds taken up by *Chloroflexi* is consistent with observations of cross feeding between these populations in other microbial mats. Glycogen fermentation and organic acid excretion by *Cyanobacteria* has been demonstrated in thermophilic *Synechoccocus*-dominated mats found in alkaline siliceous hot springs (Octopus Spring and Mushroom Spring) in Yellowstone National Park using both ^{14}C-labeling and transcript analysis, which demonstrated upregulation of *Synechoccocus* fermentation genes (Nold and Ward 1996, Steunou *et al.,* 2006). In these mats, thermophilic *Chloroflexales* related to *Roseiflexus castenholzii* are also abundant and pulse labeling experiments with $^{13}CO_2$ and $^{14}CO_2$ provided evidence for the transfer of cyanobacterial-derived photosynthate from *Synechoccocus* to the *Roseiflexus* populations (van der Meer *et al.,* 2005). The intermediacy of acetate was demonstrated by lipid analysis of *Synechoccocus* mat samples incubated with ^{13}C-acetate. However, this acetate incorporation was observed during the morning, in contrast to observations with the *Microcoleus*-dominated mats described here, and may be directly linked to photoheterotrophic growth. The cross-feeding of *Synechoccocus* and *Roseiflexus* populations in the Yellowstone mats has served as the basis for developing *in silico* models of mat carbon metabolism that are broadly consistent with the anoxic carbon metabolism observed at Elkhorn Slough (Taffs *et al.,* 2009). Though accumulation of PHAs by mat dwelling *Chloroflexi* has never been directly demonstrated, studies of microbial mats found in the Great Sippewissett Salt Marsh in Massachusetts, USA which contain *Chloroflexi* populations, have shown that PHAs accumulate at night and decrease during the day (Rothermich *et al.,* 2000). The accumulation of PHA in the dark was

putatively linked to bacterial breakdown of glycogen, which supports the model for the Elkhorn Slough mats derived from metatranscriptomic data. Additionally, studies of carbon metabolism in *Chloroflexus auarantiacus* have shown that this model organism produces both glycogen and PHA (Sirevag and Castenholz 1979).

Acetate uptake and incorporation into PHA was demonstrated in single cells of uncultured *Accumulibacter* spp., a microbial population important in EBPR, by MAR-FISH using ^{14}C-labeled acetate (Burow *et al*., 2008). Evidence for acetate uptake by single cells affiliated with the *Chloroflexi* was obtained by application of FISH-SIMS to the mats samples using ^{13}C-labeled acetate and *Chloroflexi*-specific oligonucleotide probes. **Other single cells that were not affiliated with the *Chloroflexi* were observed with enriched ^{13}C/^{12}C ratios, indicating that additional bacterial groups assimilated acetate under anoxic conditions.** Future work will focus on cultivating *Chloroflexi* from the Elkhorn Slough mats to test the proposed linkage of acetate uptake to glycogen conversion to PHA **and identifying other group involved in the anoxic metabolism of organic acids derived from fermentation.**

In conclusion, **metatranscriptomic sequencing of samples obtained from microbial mats has demonstrated that *Cyanobacteria* and *Chloroflexi* are the most active groups under dark, anoxic conditions. Genomic reconstruction and fragment recruitment of transcripts was used to reconstruct a pathway for the catabolism of photosynthate that linked fermentation by *Microcoleus* with PHA production by the *Chloroflexi*. This work highlights the utility of metatranscriptomics to define pathways for element cycling in complex microbial communities.**

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Figure Legend

Figure 1. A: Comparison of nearly full-length and pyrotag amplicon 16S rRNA sequences of the phototrophic layer from Elkhorn Slough mat analyzed using the RDP classifier with a confidence threshold of ≥ 80%. Unassigned reads are sequences that could not be classified with a confidence threshold of ≥ 80% on the phylum level.
B: Taxonomic classification of metatranscriptome reads analyzed by MEGAN using the RefSeq database (NCBI). Unassigned reads have an unknown taxonomic affiliation.

Figure 2. Neighbor joining tree of *Cyanobacteria*-related nearly full-length 16S rRNA sequences from cDNA samples recovered from BN (SIUS) and EN (SIUT) samples. OTU_{98} representatives of OTUs that contain more than one sequence (numbers of sequences per OTU in parenthesis) are depicted in the tree. Bootstrap values calculated with the PhyML algorithm that were ≥ 50% are displayed in the tree. Scale bar represents 10% estimated sequence divergence.

Figure 3. **Neighbor joining tree** of *Chloroflexi*-related nearly full-length 16S rRNA sequences from cDNA samples BN (SIUS) and EN (SIUT). Elkhorn Slough mat sequences group with sequences retrieved from other microbial mat sites and are only distantly related with cultured *Chloroflexi*. Bootstrap values calculated with the PhyML algorithm that were ≥ 50% are displayed in the tree. Scale bar represents 10% estimated sequence divergence.

Figure 4. A. Reconstruction of anoxic low-molecular-weight metabolite producing pathways based on the *M. chthonoplastes* genome. Abbreviations: glycogen

phosphorylase (GP), lactate dehydrogenase (LDH), pyruvate-formate lyase (PFL), alcohol/aldehyde dehydrogenase (ADH), pyruvate:ferredoxin oxidoreductase (PFR), bidirectional [NiFe] hydrogenase (HOX), and acetate kinase (ACK). Phosphotransacetylase (PTA) was not identified. B. Reconstruction of glycogen to PHA conversion pathway in sequenced *Chloroflexales* genomes. Abbreviations: acetyl-CoA synthetase (ACS), glycogen phosphorylase (GP), Embden-Meyerhof-Parnas pathway (EMP), pyruvate dehydrogenase complex (PDC), acetoacetyl-CoA transferase (PhaA), acetoacetyl-CoA reductase (PhaB), polyhydroxyalkonoate synthase (PhaC). Detailed assignments for these genes are presented in the Supplementary Information (Supplemental Table 4-5). Quantitative measurements of recruited reads from BN and EN datasets are presented in Figure 5 and Supplemental Table 6.

Figure 5. Comparison of reads involved in carbon catabolic pathways in BN and EN libraries. Reads representing transcripts for the *Microcoleus spp.* fermentation pathway and the *Chloroflexales* glycogen to PHA conversion. The reads are presented as a relative proportion of the total reads assigned to this SEED functional category. Pha (*phaCAB* and HOX (*hoxEFUYH*) represent the sum of multiple transcripts. Actual numbers of reads recruited to the genes in each category are presented in the Supplemental Information (Supplemental Table 6).

Figure 6. A. ^{13}C-acetate uptake under dark, anoxic conditions within the microbial community of the Elkhorn Slough mat. (A) Scanning-electron micrographs (SEM). (B) CARD-FISH micrographs using CF mix probes (CFX1223 and GNSB-941) targeting *Chloroflexi* in green. (C) Ion micrographs of ^{13}C/^{12}C ratios of the same community members imaged by SEM and CARD-FISH. Scale bar = 5 μm. B. ^{13}C-acetate uptake

under dark, anoxic conditions by different microbial groups in the Elkhorn Slough mat. *Chloroflexi* (n = 40) were identified using CF mix CARD-FISH probes. *Microcoleus* spp. (n = 12) and other *Cyanobacteria* (n = 12) were identified by their morphological characteristics. Error bars represent the standard error of the mean. Note that the probes do not distinguish *Chloroflexi* from *Chloroflexales* so these results are discussed at the phylum level.

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