UC Irvine UC Irvine Previously Published Works

Title

Discovery of CDC42 Inhibitors with a Favorable Pharmacokinetic Profile and Anticancer In Vivo Efficacy.

Permalink https://escholarship.org/uc/item/3db4936c

Journal Journal of medicinal and pharmaceutical chemistry, 67(12)

Authors

Brindani, Nicoletta Vuong, Linh La Serra, Maria <u>et al.</u>

Publication Date

2024-06-27

DOI

10.1021/acs.jmedchem.4c00855

Peer reviewed



Article

Discovery of CDC42 Inhibitors with a Favorable Pharmacokinetic Profile and Anticancer In Vivo Efficacy

Nicoletta Brindani,[#] Linh M. Vuong,[#] Maria Antonietta La Serra, Noel Salvador, Andrea Menichetti, Isabella Maria Acquistapace, Jose Antonio Ortega, Marina Veronesi, Sine Mandrup Bertozzi, Maria Summa, Stefania Girotto, Rosalia Bertorelli, Andrea Armirotti, Anand K. Ganesan,^{*} and Marco De Vivo^{*}



results indicate that these CDC42 inhibitors represent a promising chemical class

toward the discovery of anticancer drugs, with ARN25499 as an additional lead candidate for preclinical development.

INTRODUCTION

CDC42 GTPases (CDC42, RHOJ, RHOQ) have emerged as appealing targets for the rational design of anticancer drugs as they are overexpressed in multiple cancers.^{1–4} Functionally, they play a role in multiple pathways required for tumor progression including cell migration, angiogenesis, and resistance to targeted therapies.^{3,5,6} In addition to its role in cancer development, CDC42 influences cardiovascular physiology, immune system function, nervous system function, and bone remodeling.⁷ CDC42 GTPases are in their active conformation when bound to GTP and switch to their "off" conformation when they hydrolyze GTP to GDP. Also, CDC42 protein activity is finely regulated by (i) guanine nucleotide exchange factors (GEFs), which exchange GDP for GTP to turn the protein "on", (ii) GTPase activating proteins (GAPs), which help switch between "on" and "off" conformations, and (iii) guanine nucleotide dissociation inhibitors (GDIs), which keep GTPases in their "on" state.^{8,9} When the protein binds GTP, it can then activate downstream effectors, leading to altered cytoskeleton organization, polarity, adhesion, and migration, as well as cell proliferation.

On these bases, several approaches have been taken to specifically target CDC42.^{11–14} These include (i) inhibiting CDC42-regulator and effector interactions, (ii) direct inhibition of effector kinases, and (iii) covalent irreversible inhibition of GEF-catalyzed nucleotide exchange.^{11,13} Several studies disclosed new small molecules that target CDC42–GEF interfaces such as ZCL278,¹⁵ ZCL367,¹⁶ CASIN,¹⁷ AZA197,¹⁸ and NSC23766.¹⁹ However, these approaches have failed to make it to the clinic because of lack of specificity

and because of off-target toxicity secondary to the promiscuous activity of GEFs toward multiple Rho family members.^{20,21}

We recently developed a new class of CDC42 GTPase interaction inhibitors.^{22,23} This class was rationally designed to specifically inhibit the interaction between CDC42 and its downstream effectors, differing from the other small molecules that interfere with the CDC42-GEF interaction, thus avoiding potential hematologic side effects.²² In particular, these inhibitors feature a triazine/pyrimidine core functionalized with several 6-membered heteroaryl groups, aniline, and completely saturated piperidine moieties (Scaffold A, Figure 1). In addition, our structure-activity-relationship (SAR) study allowed the identification of the best functionalities on a pyrimidine core that were also retained on the symmetric triazine core, as well as the identification of the influence of different types of masked and free saturated heterocycles groups. Importantly, compounds ARN22089 and ARN25062 (Figure 1) are drug-like CDC42/RHOJ pyrimidine derivatives with a significant ability to inhibit tumors in patient-derived xenografts (PDX) in vivo.²³ On the other hand, compound 1 (Figure 1, which was originally named compound **29** in ref 23) resulted in a promising triazine inhibitor with one-digit

Received:	April 9, 2024				
Revised:	May 7, 2024				
Accepted:	May 13, 2024				
Published:	June 12, 2024				



pubs.acs.org/jmc

Article



Figure 1. Representation of new chemical scaffolds B–D explored starting from initial hits.^{22,23}

Table 1. Modification at R¹ of Triazine Class

Entry	R ¹		IC ₅₀ ^a				
		\mathbb{R}^1	(µM)	(µM)	(µM)	(µM)	(µM)
	HN N N CF3		SKM28	SKMel3	WM3248	A375	SW480
	<u> </u>						
1	1 ^{b,d}	$ \downarrow$	20.7	4.9	3.2	3.7	3.2
2	2d	N N	55 5	17.0	ND ^c	27.4	27.8
2	-		55.5	17.0	ND	27.4	27.0
		N N					
3	3 ^d		7.6	4.2	7.3	8.0	3.7
4	٨d		67	6.4	2.0	1.0	27
4	4		0.7	0.4	5.0	4.0	5.7
	4	HN-N					
5	5 ª	\square	3.4	3.9	3.2	3.1	2.6
6	۲d	$\left\{ \right\} $	10.6	0 1	5.4	66	5.2
0	U	\sim	10.0	0.1	5.4	0.0	5.5
		N					
7	$7^{\rm d}$		26.4	12.9	25.3	13.2	25.0
		l					
8	8 ^d	N= N-	12.8	74	33.7	> 100	ND°
0	0	Ť	12.0	7.4	55.7	- 100	TLD .
0	od		10.0	0.0		0.1	
9	9 "		10.0	9.8	8.0	8.1	5.5
10	10 ^d	Y ^{NH}	ND ^c	> 100	ND ^c	ND ^c	ND ^c
		S					
11	11 ^e	$ \downarrow\rangle$	8.1	5.0	3.9	5.3	6.3
		1					

^{*a*}All IC₅₀ values have a $R^2 > 0.90$. ^{*b*}Compounds initially reported by Brindani et al.²³ ^{*c*}ND = not determined. ^{*d*}The compound features a completely saturated piperidin-4-yl substituent. ^{*e*}The compound features a partially saturated 1,2,3,6-tetrahydropyridin-4-yl substituent.

Entry ^a		R ¹	IC ₅₀ ^b (μM) SKM28	IC ₅₀ ^b (μM) SKMel3	IC ₅₀ ^b (μM) WM3248	IC ₅₀ ^b (μM) A375	IC ₅₀ ^b (μM) SW480
1	12	N C C	7.5	5.8	6.1	5.1	4.8
2	13	F ₃ C	12.3	3.9	4.6	5.1	3.0
3	14	F ₃ CO	6.4	6.0	4.3	5.2	3.0
4	15 (ARN25499)	MeO MeO	13.5	8.1	12.8	9.7	11.2
5	16	MeO MeO	11.1	12.8	21.0	8.0	8.5
6	17 [°]	HO HO	NA ^d	NA ^d	NA ^d	NA ^d	NA ^d
7	18°	HOHO	\mathbf{NA}^{d}	NA ^d	NA ^d	NA ^d	NA ^d
8	19	N ,	12.9	6.4	24.9	16.3	17.1
9	20 (ARN25375)	F ₃ C	10.4	5.6	7.0	4.5	5.9
10	21	F2HCO	13.5	5.2	4.7	4.2	6.8
11	22	MeO ,	8.3	5.1	12.0	9.3	5.8
12	23	MeO N	11.8	10.5	13.1	10.3	11.5

Table 2. Modification at R¹ Aniline Substituents of "Inverted" Pyrimidine Class

^{*a*}Activity of two reference compounds: ARN22089 (IC₅₀–SKM28 = 24.8 μ M,–SKMel3 = 4.2 μ M,–WM3248 = 4.5 μ M,–A375 = 4.9 μ M,–SW480 = 8.6 μ M), ARN25062 (IC₅₀–SKM28 = 6.1 μ M,–SKMel3 = 4.6 μ M,–WM3248 = 9.3 μ M,–A375 = 5.1 μ M,–SW480 = 5.9 μ M). ^{*b*}All IC₅₀ values have a R^2 > 0.90. ^{*c*}Compound as hydrobromide salt. ^{*d*}NA = not active.

micromolar activity in four cancer cell lines (SKMel3, WM3248, A375, SW480). However, it is characterized by low solubility (Sk = 1 μ M).²³ These results prompted us to further investigate the SAR of these new CDC42 interaction inhibitors.

Here, we present 25 additional derivatives that expand this new class of CDC42/RHOJ inhibitors, enhancing their druglike profile. As described in Figure 1, we used compound 1,²³ ARN22089,²² and ARN25062²³ as our starting point for further derivatization of their core scaffold, generating scaffolds of types B–D (Figure 1), which show good potency in five cancer cell lines, and improved metabolic stability on mouse microsomes. Through the resulting SAR, we identified and characterized, in vitro and in vivo, compound 15 (ARN25499), which has (i) excellent kinetic and thermodynamic solubility, (ii) an improved in vivo bioavailability compared to previous leads, and (iii) a significant inhibition of tumor growth in a PDX mouse model.

RESULTS AND DISCUSSION

Exploring the Structure of the New CDC42/RHOJ Inhibitors. To briefly recapitulate our previous drug discovery program on targeting CDC42/RHOJ, we mention that we have recently discovered a new class of trisubstituted pyrimidines (scaffold A, Figure 1, X = CH) as drug-like CDC42/RHOJ inhibitors.²³ Scaffold A features 6-membered (hetero)aryl groups on position 6 of the core, several differently functionalized anilines on position 4, and piperidines on carbon 2. Additionally, we have preliminarily evaluated two triazine counterparts of this pyrimidine class (scaffold A, Figure 1, X = N), showing promising activity in our cell-based assay.²³ This effort allowed us to identify ARN22089 and ARN25062 (Figure 1) as pyrimidine derivatives, and the triazine ARN24928 (Figure 1). All these lead compounds showed favorable PK and in vivo efficacy.²³ This data prompted us to expand this class of triazine inhibitors and to complete the SAR on trisubstituted pyrimidines. Specifically, we previously found that the free 4piperidine moiety represented the best moiety in terms of potency and synthetic feasibility, thus we maintained it in the majority of the derivatives. Moreover to improve the kinetic solubility of the triazine 1 (Sk = 1 μ M, Figure 1)²³ and the metabolic stability in mouse microsomes of selected pyrimidines (ARN22089–27 min, ARN25062–45 min),²³ here we mainly explored: (i) the effect of different heteroaryl cycles of scaffold B; (ii) the inversion of piperidine/aniline positions of scaffold C, and (iii) the embedding of a 3'nitrogen of the aniline moiety in a bicyclic group, as in scaffold D. We assessed the antiproliferative activity of all compounds in four melanoma cancer cell lines that have a BRAFV600E

Table 3. Modification at F	and R	² of "Classic"	Pyrimidine Class
----------------------------	-------	---------------------------	------------------

Entry	Compound Ph $R^2 \xrightarrow{2} N \xrightarrow{4} N \xrightarrow{R^1}$	R ¹	R ²	IC ₅₀ (μM) ^a SKM28	IC50 (µM) ^a SKMel3	IC ₅₀ (μM) ^a WM3248	IC ₅₀ (μM) ^a A375	IC ₅₀ (μM) ^a SW480
1	ARN22089 ^b	N. N.	HN	24.8	4.2	4.5	4.9	8.6
2	24		HN	13.1	5.0	8.6	6.8	5.1
3	25		HN	7.1	6.1	12.6	6.2	6.8
4	26 ^b	, , , , , , , , , , , , , , , , , , ,	HN	12.3	9.5	10.3	9.4	7.9

^{*a*}All IC₅₀ values have a $R^2 > 0.90$. ^{*b*}Compounds initially reported by Jahid et al.²²

mutation (SKM28, SKMel3, WM3248, A375), and a colon cancer line (SW480) with a KRASG12 V mutation (Tables 1-3). These transformed cells have active RHOJ/CDC42 activity that is known to activate downstream Raf-MEK-ERK and PI3K-Akt pathways.^{24,25}

Exploration of Heterocycles on the Triazine Core. In our previous work, we observed that the *m*-trifluoromethylaniline moiety improves the potency of pyrimidine derivatives.²³ Surprisingly, the same modification on a triazine core as in compound 1 dropped the kinetic solubility to 1 μ M but maintained a similar potency compared to the related pyrimidine counterpart.²³ Thus, we explored different heteroaryl substituents R¹ on the triazine core, maintaining the *m*-trifluoromethylaniline. The aim was to improve the solubility and explore the contribution of this substituent to the activity (Table 1). While the substitution of the phenyl group with a pyrimidine group in derivative 2 generally decreased the activity, especially in SKM28 (IC₅₀ = 55.5 μ M, Table 1, entry 2), compared to compound 1, the introduction of a bicyclic heteroaromatic group such as isoquinolyl-, Nmethylindolyl-, indazolyl-, benzofuran-3-yl- in 3-6 restored the activity in the one digit micromolar range in all cancer cell lines (Table 1, entries 3-6). Indeed, among 3-6, the indazolyl group in compound 5 gave the best effect in potency, showing an IC₅₀ in the range of 2.6–3.9 μ M in all five cancer cell lines. We then investigated the role of smaller 5-membered heterocycles. Derivative 7 with N-methylpyrazolyl group maintained the activity in SKM28 but decreased the activity in other cancer cell lines, displaying the IC₅₀ of \sim 3-8 fold higher (Table 1, entry 7) compared to 1. The shifting of the Nmethyl group in the imidazolyl substituent of 8 displayed a negative impact on the antiproliferative activity of 1, annihilating the effect on A375 and decreasing the activity of ~10-fold on WM3248 (Table 1, entry 8). Instead, the presence of free nitrogen of the pyrrolyl substituent of compound 9 restored the activity in all cancer cell lines (Table 1, entry 9). We then evaluated the effect of the geometry of this substituent by introducing a completely saturated pyrrolidinyl group in derivative 10, which resulted in a complete loss of activity compared to its aromatic counterpart 9 (Table 1, entries 9-10). Finally, the introduction of a sulfur heteroatom in the thiophenyl substituent of 11 exhibited one-digit micromolar activity on all cancer cell lines (Table 1, entry 11). Notably, this analogue features the partially saturated 1,2,3,6-tetrahydropyridin-4-yl instead of the typical piperidine

moiety. This was due to synthetic reasons (vide infra), as it is characterized by a slight instability in DMSO, probably due to the susceptibility of thiophene to oxidative processes.

Overall, these data suggested that key features for the activity of this class of CDC42/RHOJ inhibitors are their electronic structure in terms of the molecular size, their conformational freedom, and their number and exact position of the heteroatom in the heterocycle moiety. Specifically, analogues 3-6 pointed out that a heteroaryl bicycle is generally preferable to a monocycle, and derivative 11 suggested a good tolerance of a different conformation of piperidine ring induced by the intracyclic double C-C constraint.

Exploration of "Inverted" Trisubstituted Pyrimidine. The initial goal was to exchange the piperidine and the aniline moieties of ARN22089 and ARN25062 to evaluate the impact of the different orientations of these substituents on the activity. Therefore, we introduced meta-N,N-dimethylaminoand meta-trifluoroaniline moieties on position 2 and the piperidine on position 4 of the pyrimidine core, maintaining a phenyl substituent on position 6. Thus, we produced the analogues 12 and 13, which have comparable antiproliferative activity in all cell lines (Table 2, entries 1, 2). Generally, these new inhibitors have been proven equally potent to the original ARN22089 and ARN25062.23 This result paved the way toward the development of a second type of CDC42/RHOJ inhibitors, where variously substituted anilines were inserted on the C2 of the pyrimidine cycle (Table 2). While the introduction of a meta-trifluoromethoxy phenyl in compound 14, or two substituents as in 15 and 16, were generally well tolerated, the presence of free hydroxyl or carboxylic functional groups on the aniline of 17 and 18 annihilated the activity in the cells (Table 2, entries 3-7). We then investigated the impact of naked or substituted pyridines as in analogues 19-23 to evaluate the role of the basic functionality in this portion of the chemical structure. We found out that the naked 4piperidine in derivative 19 decreased the potency of 2-3 folds compared to ARN25062 (Table 2, entry 8). The insertion of vicinal trifluoromethyl-, difluoromethoxyl-, or methoxyl group in the pyridine ring of compounds 20-22 restored the activity, probably due both to an alteration of the basic strength of the pyridine site and to additional interactions established by fluorine atoms or a methoxy group with the target (Table 2, entries 9-11). A similar, although less relevant, effect was observed with the regioisomer of compound 22, where the

entry	compound	kinetic solubility (Sk) (μM)	thermodynamic solubility (S ThD) $\left(\mu \mathrm{M} ight)^b$	aggregation by NMR (50 μM) ^b	${T_{1/2} \atop (\min)^b}$	$T_{1/2} \underset{(\min)}{\operatorname{microsomal}} b$
1	ARN22089 ^a	250	ND	no	71	27
2	ARN25062 ^a	168	ND	no	>120	45
3	1 ^{<i>a</i>}	1	ND	ND	>120	>60
4	3	6 ± 1	ND	ND	>120	>60
5	4	<1	ND	ND	>120	>60
6	5	1	ND	ND	>120	>60
7	6	12	ND	ND	>120	>60
8	9	237 ± 11	ND	no	>120	>60
9	11	<1	ND	ND	>120	>60
10	12	13 ± 4	ND	ND	60	>60
11	13	<1	ND	ND	>120	>60
12	14	<1	ND	ND	>120	>60
13	15 (ARN25499)	>250	371 ± 5	no	>120	>60
14	15 HCl	244 ± 2	341 ± 38	ND	ND	ND
15	16	245 ± 4	ND	no	89	>60
16	20 (ARN25375)	222 ± 2	106 ± 3	no	>120	>60
17	20 HCl	244 ± 2	1 ± 0	ND	ND	ND
18	21	<2	ND	ND	>120	>60
19	22	238 ± 7	ND	no	>120	>60
20	23	247 ± 6	ND	no	>120	>60
21	24	<1	ND	ND	>120	37
22	25	239 ± 5	ND	yes	>120	>60
23	26	68	ND	ND	>120	>60
a _C			123 bND and latertal			

Table 4. Kinetic and Thermodynamic Solubility, Aggregation by NMR, Plasma, and Microsomal Stability in Mouse of Selected Compounds

¹Compounds initially reported by Brindani et al.²³ ^DND = not detected.

pyridine nitrogen is in *meta*-position and the methoxyl group is in *para* position of the aniline substituent (Table 2, entry 12 compound 23).

Evaluation of Our New Analogues of ARN22089. The investigation of new triazines 2-11 highlighted a positive contribution of bicyclic heteroaryl substituents and a tolerance of partially saturated piperidine moieties. We further proved these results through the compounds 24-26, close analogs of our previous lead compound ARN22089 (Table 3). Derivative 24 maintained the activity of ARN22089, evidencing that a different saturation and conformation of the piperidine do not significantly affect potency (Table 3, entry 2). Moreover, we embedded the meta *N*-methyl amino group of the aniline moiety in an indole group, both in methylated and free forms, generating compounds 25 and 26 (Table 3, entries 3,4). These analogues inhibit the cell viability for all cell lines, confirming a positive effect of a bulkier bicyclic aniline moiety.

Evaluation of Druglike Properties of Our New Leads. After this initial evaluation, we assessed the drug-like profile of this novel RHOJ/CDC42 inhibitors by studying the kinetic solubility, plasma, and phase-I microsomal stability of 18 selected compounds. This was done to define the key chemical elements for a drug-like profile (Table 4). Notably, the first disclosed triazine 1 was very stable in plasma and mouse microsomes (>120 and >60 min, Table 4), but poorly soluble (Sk = 1 μ M). Among the selected triazine derivatives 3–6, 9, and 11, only compound 9 showed an excellent kinetic solubility of 237 ± 11 μ M (Table 4, entry 8), suggesting that the pyrrolyl ring positively impacts the solubility in water. All other triazine inhibitors have a kinetic solubility <12 μ M (Table 4, entries 4–7, 9). On the contrary, the good plasma and microsomal stability of triazine 1 was maintained.

From the first class of trisubstituted pyrimidines, we previously selected ARN22089 and ARN25062 as the most promising compounds for further in vivo studies. Notably, both compounds showed high kinetic solubility (250 and 168 μ M, Table 4), and ARN25062 exhibited ameliorated plasma and microsomal stability (>120 and 45 min, Table 4) compared to ARN22089 (71 and 27 min, Table 4). Here, our goal was to improve the microsomal stability through an expansion of the initial SAR study for the "inverted" trisubstituted pyrimidine class (Table 2). Among the selected nine compounds 12-16 and 20-23 (Table 4, entries 10-13, 15, 16, 17-20), derivatives 15, 16, 20, 22, 23 exhibited excellent kinetic solubility (222 to >250 μ M range). Indeed, the shifting of the N-methylamino aniline from C4 into C2 of the pyridine core as in compound 12 drastically dropped solubility. However, it ameliorated microsomal stability compared to our previous lead ARN22089 (Table 4, entry 10 vs entry 1). The same trend was observed for the solubility and microsomal stability parameters, in the case of the metatrifluoromethylaniline moiety that was shifted from the C4 of the previous candidate ARN25062 to the C2 of the analogue 13 (Figure 1, Table 4, entry 11 vs entry 2). No improvement was observed when the trifluoromethoxy group of 14 replaced the trifluoromethyl group of 13 (Table 4, entry 12). On the other hand, when the meta-trifluoromethyl group was inserted in a pyridin-4-amine as in 20, the solubility was completely restored to 222 \pm 2 μ M value (Table 4, entry 16), suggesting a positive contribution of an additional basic site of the pyridine. Notably, the pyridine analogues 22-23 exhibited excellent kinetic solubility (Table 4, entries 19, 20), except for compound 21 with the 2-difluoromethylpyridine group (Table 4, entry 18). On the other hand, the disubstituted

pubs.acs.org/jmc



Figure 2. (A–E) Example of MST traces of selected compounds **22** (A) and **15** (B). In panel A, there was no change in fluorescence and no binding events. In panel B a good change in fluorescence was displayed, highlighting a potential binding event. (C) Graph displays the difference in normalized fluorescence (ΔF_{norm} [% σ] = F_{hot}/F_{cold}) between protein:compound sample and a protein-only sample. Compounds (C) were tested at 50 μ M toward the activated (loaded with GppNHp) or inactivated (loaded with GDP) His-Cdc42. A single blue asterisk indicates a signal-to-noise ratio above 5, while double red asterisks indicate a signal-to-noise ratio above 12. ΔF_{norm} at 2.5 s (t-jump) and 15 s (thermophoresis signal) were reported. (D,E) example of binding check by ¹⁹F T₂ filter NMR experiments [compound 9-(E)], and by WaterLOGSY [Compound 16-(F)]. Compounds were tested in the absence of protein and in the presence of GppNHp (black) or GDP (gray), and in the presence of activated (loaded with GDP) His-CDC42 (red) or inactivated (loaded with GDP) His-CDC42 (blue). The arrows indicate where a difference in the compound NMR signal is observed after the addition of the protein, highlighting a binding event.

phenyl rings of compounds **15** and **16** improved sensibly the solubility (Table 4, entries 13, 15). Generally, all derivatives of this second class of trisubstituted pyrimidines possess excellent plasma and improved metabolic stability. Specifically, only compounds **12** and **16** showed a moderate decrease in plasma stability.

Moreover, the closest structural analogues 24-26 of ARN22089 were further analyzed (Table 4, entries 21–23). The presence of the double C–C bond in 24 annihilated the solubility, ameliorated the plasma stability, and approximately maintained the same metabolic stability. The solubility was partially recovered through the insertion of the free indole group in 26 with Sk of 68 μ M and completely restored for *N*-methylindole in 25 with Sk 239 ± 5 μ M (Table 4, entries 22–23).

The solubility and the aggregation state of selected compounds 9, 15, 16, 20, 22, 23, and 25 were further evaluated by NMR analysis under experimental conditions employed in microscale thermophoresis (MST) and NMR (Tris buffer– vide infra) according to the SPAM filter approach to avoid false-positive results.²⁶ Since the starting compounds (ARN22089 and ARN25062) showed aggregation at 100 μ M, these six compounds were tested in the binding assay buffer at the maximum concentration of 50 μ M, in the presence of an internal reference (4-trifluoromethyl benzoic acid, 200 μ M). The seven selected compounds were soluble in Tris buffer at least up to 50 μ M (data not shown) in line with kinetics solubility data. Indeed compound 25 showed aggregation at 50 μ M differently from other compounds,

thus it was excluded from further binding evaluation studies (vide infra).

Notably, derivatives 15 (ARN25499) and 20 (ARN25375) resulted in being the best compounds in terms of potency in all cell lines, and kinetic solubility, also showing a strong improvement in microsomal stability. To preliminary evaluate the pH-dependent solubility of these two compounds, we assessed both the kinetic and thermodynamic solubility for both the neutral and the hydrochloride forms of 15 (ARN25499) and 20 (ARN25375) (Table 4, entries 13, 14, 16, 17). The kinetic solubility was maintained for the corresponding hydrochloride forms for both derivatives (Table 4, entries 14, 17). The same behavior has been observed for the thermodynamic solubility of analogue 15, which is >300 μ M for the neutral and salt form (Table 4, entries 13, 14). On the contrary, the hydrochloride form of 20 exhibited a thermodynamic solubility 100-fold lower than the related neutral form (Table 4, entries 16, 17), suggesting a significant pH-dependent behavior for the trifluoromethylpyridine analogue 20.

Microscale Thermophoresis (MST) and NMR for CDC42-Binding Validation. To investigate the binding of the most promising compounds to the target, we have employed microscale thermophoresis (MST) and NMR techniques. Among the previously selected 18 new inhibitors, we further selected the best compounds in terms of potency, drug-like profile, and aggregation by NMR, namely, compounds 9, 15, 16, 20, 22, 23.

pubs.acs.org/jmc



Figure 3. Model structure of compound 15 (ARN25499, panel A) and 20 (ARN25375, panel B) compounds bound to the allosteric drug-binding pocket of CDC42 is reported. The structure of CDC42 is represented as a white surface while the identified drug-binding pocket is shown in both stick and transparent blue surface. 15-ARN25499 and 20-ARN25375 are reported as yellow and green sticks, respectively.



Figure 4. Molecular dynamics (MD) simulations of the protein–ligand complexes. The structural representation of CDC42 in a complex with compound 15 (ARN25499, panel A) or compound 20 (ARN25375, panel B) is reported on the left. CDC42 is represented as a cartoon, while the binding pocket is highlighted as both a stick and a transparent blue surface. Multiple MD snapshots of the 15 (ARN25499, yellow) and 20 (ARN25375, green) binding poses are shown as sticks. On the right, the RMSD over time for the CDC42-ligand complexes is reported with the RMSD running averages highlighted in bold.

We first employed microscale thermophoresis (MST) to retrieve information on the ability of the six selected compounds to engage our target.²⁷ Wild-type His-CDC42 (Ile4-Pro182) was used as a target macromolecule. The protein was activated or inactivated through loading of GppNHp or GDP, respectively. Loading efficiency was evaluated by measuring the protein intact mass with ESI MS⁺.²² Only protein samples with loading efficiency higher than 90% were used in MST experiments. Target proteins were Red-NHS labeled prior to use. Changes in their normalized fluorescence intensity (ΔF_{norm} [%] = F_{hot}/F_{cold}) were recorded. ΔF_{norm} at 2.5 s (t-jump) and 15 s (thermophoresis signal) were evaluated. Compounds were tested for binding at a concentration of 50 μ M in the presence of 0.5% v/v DMSO. Assays were set up in the Tris-HCl buffer. As shown in Figure 2, a change in the fluorescence signal was detected for

compound **15** irrespectively of the CDC42 activation state, displaying good confidence in binding and signal-to-noise ratio higher than 12, both for the active and inactive state of CDC42 (Figure 2, Table S1, Supporting Information). While compounds **9** and **16** led to a fluorescence change only in the presence of the GDP-inactivated CDC42, compound **20** showed binding for both GDP- and GppNHp-CDC42, with a slightly lower confidence in binding than the other compounds (Figure 2, Table S1, Supporting Information). On the other hand, compounds **22**, and **23** did not exhibit binding by MST for both active and inactive states of CDC42 (Figure 2, Table **S1**, Supporting Information). In all the binding events, both the T-jump fluorescence intensity and the thermophoresis signal were modified, except for compound **20** which displayed only a change in the T-jump initial fluorescence intensity.



PK Parameters	ARN25	499 (15)	ARN25375 (20)		
	<i>I.V.</i>	<i>P.O.</i>	<i>I.V.</i>	<i>P.O.</i>	
Cmax (ng/mL)	320	140	643	99	
Tmax (min)	5	120	5	120	
AUC (min*ng/mL)	28937	21686	55116	14320	
t _½ (min)	257	168	115	91	
V _D (L/Kg)	19.7	57.3	7.2	67.2	
CL (mL/min/Kg)	53	236	43	512	
F (%)	2	22	8		

Figure 5. Mouse PK profiles of ARN25499 (15) and ARN25375 (20) following intravenous (I.V.) and oral (P.O.) administration at 3 and 10 mg/ kg, respectively, and the corresponding observed and calculated PK parameters.

The binding of 6 compounds was also evaluated by NMR experiments,²⁸ testing them at 50 μ M in the absence and in the presence of both His-CDC42 (2 μ M) by ¹⁹F T₂ filters^{29,30} (fluorinated compounds) and WaterLOGSY (Water-Ligand Observation with Gradient SpectroscopY)³¹ experiments. In ¹⁹F T₂ Filter experiments, the binding event induces a linebroadening of the ¹⁹F NMR signal of the binding molecule in the presence of the protein, which results in a decrease of its intensity compared to the signals recorded in the absence of protein (as an example see Figure 2D). On the other end, in WaterLOGSY experiments, the binding event is identified by ¹H molecule signal change from negative, in the absence of protein, to less negative or to positive in the presence of protein due to the transfer of magnetization from bulk water to the compound interacting with the macromolecule (examples see Figure 2E).

In the NMR experiments, all 6 compounds bind to the activated form of His-CDC42 and only compound 22 does not bind to the inactivated form (Table S1, Supporting Information), whereas only compounds 9, 15, 16, and 20 bind to the protein in the MST experiments. These results were expected since we know that NMR is highly sensitive even to very weak binders, therefore it is quite common that binding events identified by NMR are not detected by other biophysical techniques.³²

Molecular Modeling for Binding Validation. Based on overall data, we further performed molecular modeling studies of compounds **15** (ARN25499) and **20** (ARN25375) to validate their binding toward the active state of CDC42. We utilized the drug-binding pocket previously identified at the CDC42-PAK protein—protein interface for our computational investigations. Correspondingly, we conducted molecular docking analysis of the new derivative compounds, **15** (ARN25499) and **20** (ARN25375), on the GTP-bound active configuration of CDC42.^{22,23}

Our computational results indicate that both compounds **15** (ARN25499) and **20** (ARN25375) fit inside the effector pocket of CDC42 (see Figure 3A, B), matching with the binding mode earlier proposed for compounds belonging to the previously identified class of CDC42/RHOJ inhibitors (e.g., ARN22089, ARN24928, and ARN25062).^{22,23} As previously described, the pocket possesses a hydrophobic nature that is well-suited for accommodating the phenyl group on C6 of both **15** (ARN25499) and **20** (ARN25375), serving as an anchor point for the binding to the allosteric pocket.²³

To further evaluate the binding configurations, we conducted equilibrium molecular dynamics (MD) simulations of both **15** (ARN25499) and **20** (ARN25375) in complex with CDC42 (Figure 4). Importantly, the binding pose of both **15** (ARN25499) and **20** (ARN25375) in the CDC42-ligand complexes was maintained during 500 ns-long MD simulations (RMSD = 3.40 ± 0.79 Å and 4.25 ± 0.45 for **15** (ARN25499) and **20** (ARN25375), respectively (Figure 4A, B, right panels). Indeed, as shown in Figure 4 (left panels), **15** (ARN25499) and **20** (ARN25375) tightly bind the target pocket throughout the simulation time.

In Vivo Pharmacokinetics of the Selected Follow-Up/ Backup Leads. Based on the overall results and druglike profile of our compounds, the neutral form of compounds 15 (ARN25499) and 20 (ARN25375) were selected as candidates for in vivo pharmacokinetics (PK) studies, given further experiments to assess their in vivo efficacy in animal models (mouse specie) of cancer and also compare the PK profile to the previous lead compound (ARN22089).²³

We tested two different routes of administration: (i) intravenous (I.V.) injection at a concentration of 3 mg/kg (n = 3 animals for each time point), and (ii) oral (P.O.) treatment at a dose of 10 mg/kg (n = 3 animals for each time point, Figure 5). During the PK studies, via either I.V. or P.O. administration, ARN25499 and ARN25375 were well tolerated by all animals, and no treatment-related toxicological signs were observed. While the I.V. profiles of the two inhibitors are



Figure 6. ARN25499 slows the growth of melanoma PDX tumors in NSG mice. (A) Line plot showing the growth curves of the vehicle and ARN24599 treated tumors in mean \pm SEM. A small chunk of tumors was inoculated on either side under the back skin of NSG mice. When tumors reached the initial size of about 150–200 mm³, mice were injected via tail vein with 10 mg/kg ARN25499 or vehicle (9 tumors per group) daily for 2 weeks. The tumor and weight of the mouse were measured every other day with a caliper and weight scale. (B) Scatterplot showing the blue (vehicle) and red (ARN25499) dots representing tumor volume in a millimeter cube at the end of the 2-week treatment. GraphPad Prism 9 was used to generate plot and statistical analysis using 2 way ANOVA (A) and unpaired two-tailed *t* test (B), *****p-value ≤0.0001, 0.0047, respectively. (C) The line plot (mean \pm SD) shows no significant difference in weights between vehicle and ARN25499 treated mice for the 2-week treatment. (D) WM3248 cells were treated with 10 μ M of ARN22089, ARN25375, and ARN25499 for 6 h. The accumulation of pS6 and pERK were measured by immunoblotting. Relative densitometry of pERK and pS6 as compared to unphosphorylated forms of the protein were determined and are reported below each lane. A representative blot of three independent biologic replicates is shown.

comparable, the P.O. profiles exhibited significantly different behaviors consistently with the thermodynamic solubility data of hydrochloride and neutral forms. Indeed, ARN25499 and ARN25375 reached a C_{max} of 320 and 643 ng/mL in 5 min after I.V. administration, respectively, followed by a protracted elimination phase. Both compounds were still detectable after 4 h at a concentration of 74 and 84 ng/mL, respectively. After oral administration (10 mg/kg), both compounds achieved the maximum concentration in 2 h, with C_{max} values of 140 and 99 ng/mL for ARN25499 and ARN25375, respectively. Notably, the elimination phase of ARN25499 was slower than ARN25375-. ARN25499 and ARN25375 were still detectable after 8 h (7 ng/mL) and 4 h (40 ng/mL). Compound ARN25499 showed good exposure with a longer half-life of 257 min (I.V.), and 168 min (P.O.) compared to ARN25375, which showed a half-life of 115 min (I.V.) and 91 min (P.O). In summary, these data indicate that both compounds are well tolerated after a single injection. Remarkably, while ARN25375 exhibited a favorable pharmacokinetic profile of only I.V., with a bioavailability of 8% following its pH-dependent thermodynamic solubility (Table 4), ARN25499 possesses favorable pharmacokinetic profiles of both I.V. and P.O. with bioavailability in mouse of 22% (Figure 5).

In Vivo Efficacy and In Vitro Activity of Selected Follow-Up/Backup Leads. We next test the efficacy of ARN25499 since it has favorable a PK profile, overall. We used the NOD scid gamma (NSG) model and inoculated a chunk of melanoma patient-derived xenograft (PDX) tumor on either

side of the mouse. When the tumors reached a measurable size range, we treated the mice with 10 mg/kg ARN25499 via I.V. daily, for 2 weeks. Drug treatment inhibited the growth of tumors as compared to vehicle (Figure 6A), and ARN25499 treated tumors had a significantly smaller volume as compared with vehicle-treated tumors after 2 weeks of treatment (Figure 6B). Finally, a two-week treatment with the compound showed no adverse effect on the animals and no change in body weight (Figure 6C). Next, we compared the pathway engagement of ARN25499 and ARN25375 with ARN22089 in vitro in WM3248 cells. Our previous reverse phase protein analysis of ARN22089 treated cells indicated that drug treatment inhibited the accumulation of pERK and pS6.²² Following a 6h incubation period, ARN25499 inhibited the accumulation of pERK and pS6 to a greater extent than ARN22089 (Figure 6D).

Chemistry. The triazine analogues 2–11 were obtained through a 4–5 steps synthetic route as depicted in Scheme 1. As previously developed, the order of the steps was crucial to successfully introduce the desired substituents.²³ First, the nucleophilic aromatic substitution (SNAr) of *m*-trifluoromethylaniline on cyanuric chloride 27 allowed the preparation of intermediate 28 with 89% excellent yield, which underwent two sequential Suzuki couplings. Specifically, the first Suzuki employed (1-(*tert*-butoxycarbonyl)-1,2,3,6-tetrahydropyridin-4-yl)boronic acid under classic conditions of Pd(dppf)Cl₂. DCM and aqueous K₂CO₃ in 1,4-dioxane dry at 80 °C to afford common intermediate **30** with 55% yield. Compound

Scheme 1. Synthetic Route toward Trisubstituted Triazines 2-11^a



^{*a*}Reagents and conditions: (a) cyanuric chloride, 3-(trifluoromethyl)aniline, DIPEA, 0 °C; (b) (1-(tert-Butoxycarbonyl)-1,2,3,6-tetrahydropyridin-4-yl)boronic acid, Pd(dppf)Cl₂·DCM, K₂CO₃ 2M, 80 °C 1,4-dioxane; (b') from **29** to **32e**: (1-(tert-Butoxycarbonyl)-1,2,3,6-tetrahydropyridin-4-yl)boronic acid, Pd(dppf)Cl₂·DCM, K₂CO₃ 2M, 80 °C; then Pd(dppf)Cl₂·DCM, benzofuran-3-ylboronic acid, 120 °C; (c) Het-B(OH)₂, Pd(dppf)Cl₂·DCM, K₂CO₃ 2M, 100 °C; (d) HCOONH₄,Pd(OH)₂/C, MeOH or EtOH, reflux; (d') from **32h** to **33h**: Et₃SiH, Pd/C, EtOH, rt; and (e) from compound **32j** to **11**, and from **33a-h** to **2-10**: HCl 4 M 1,4-dioxane solution, 1,4-dioxane, 0 °C to rt.

Scheme 2. Synthetic Route toward "Inverted" Trisubstituted Pyrimidine 12–16, 19–23^a



^aReagents and conditions: (a) phenylboronic acid **35**, $PdCl_2(dppf) \cdot DCM$, K_2CO_3 (2M)aq, 1,4-dioxane dry, 60 °C, then (b) compound **36**, $PdCl_2(dppf) \cdot DCM$, 110 °C; (c) compounds **38a–j**, $Pd(OAc)_2$, (\pm)-BINAP or Xantphos, Cs_2CO_3 , 1,4-dioxane dry, 120 °C; (d) $Pd(OH)_2/C$, NH_4COOH , MeOH dry, 80 °C; and (e) HCl (4 M in 1,4-dioxane), 1,4-dioxane dry.

30 represented a useful building block for the product diversification for this kind of scaffold, since the second Suzuki coupling with a wide range of boronic acids or esters 31a-i under slightly stronger conditions produced trisubstituted triazines 32a-i functionalized with several five-, six-membered, mono- and biheterocycles with 23-79% range yield. Indeed, compound 32e was accessed in a one-pot fashion from monosubstituted triazine 28 with a 45% yield. Then the

reduction and Boc removal transformations got access to final products 2-10. Generally, the reduction was conducted in the presence of HCOONH₄, Pd(OH)₂/C, MeOH, or EtOH under reflux, affording precursors 33a-g and 33h' with 25-92% range yield. Notably these conditions have been proven to be too harsh for the generation of the desired derivative 33h with aromatic pyrrole moiety due to the over-reduction of substrate 32h to 33h', where the pyrrole ring was reduced to a

completely saturated pyrrolidine ring with a very good 86% yield. Thus, we successfully applied milder conditions of reduction in the presence of Et_3SiH , Pd/C in EtOH at room temperature to obtain precursor **33h** starting from **32h** with 60% yield. The double C–C bond of **32** turned out to be strongly dependent on the electronic features of the inserted heterocycle in conjugation with the triazine core.³³ This hypothesis was further supported by substrate **32i**, whose double C–C bond of 1,2,3,6-tetrahydropyridine has not been reduced both with classic conditions under prolonged time (5 days), neither under different conditions (i.e., NiCl₂·H₂O, NaBH₄ in MeOH; or NaBH₄ in the presence of TFA or AcOH in THF at 50 °C). Thus, compound **32i** was directly subjected to BOC deprotection to afford final compound **11** with a 39% yield.

As shown in Scheme 2, we were able to develop a straightforward and efficient 5-step synthetic route for the obtainment of "inverted" trisubstituted pyrimidine 12-16, and 19-23. Since this analogue features the aniline moiety on C2, we performed the one-pot synthesis of intermediate 37 through two sequential Suzuki coupling with phenyl boronic acid 35 and boronic ester 36 in the presence of $PdCl_2(dppf)$. DCM and aqueous K_2CO_3 . For this strategy, we exploited the well-known reactivity order of each position of the pyrimidine halides 34, which follows the general order C4(6) > C2. We drove the insertion of each boronic partner through the temperature regulation: 60 °C for the first phenyl group, and 110 °C for the second tetrahydropyridinyl moiety. Thus, we nicely obtained intermediate 37 with a 55% yield just through one step starting from commercially available 34. This compound represented again our divergent point for the introduction of different aniline substituents on C2 through the Buchwald-Hartwig reaction. Thus, the reaction of 37 with aniline 38a-j in the presence of $Pd(OAc)_2$, (\pm) -BINAP or Xantphos, and Cs₂CO₃ at 120 °C afforded intermediates 39aj with 60-85% range yield. This transformation occurred together with the isomerization of the double C-C bond of tetrahydropyridine producing a mixture of regioisomers, but this did not affect our synthetic plane, since the next step involved the reduction of the aforementioned double bond for the obtainment of precursors 40a-j. The final Boc removal with HCl in dioxane afforded final products 12-16 and 19-23 (Scheme 2).

On the other hand, the unmasking of catechol and salicylic groups in 17 and 18 was easily achieved through the treatment of intermediate 40d-e with BBr₃ in DCM, respectively (Scheme 3). The synthesis of the products 24-26 was carried out following the synthesis strategy previously developed (Scheme 4).²³ Compound 24 was obtained through 3 synthetic steps starting from dichloro-phenyl pyrimidine 41. As previously described, SNAr of 41 with aniline 38a, followed



^aReagents and conditions: (a) BBr₃ (1 M in DCM), DCM dry.

by the Suzuki coupling with 36 afforded intermediate 43a. Thus, the double C–C bond of tetrahydropyridine was maintained in the final product 24 by direct Boc deprotection of 43a with HCl in dioxane (56% yield). Instead, the synthesis of the derivatives 25 and 26 required the additional double C–C reduction step. Thus, SNAr with N-Methyl- and N-Bocaniline 38k–l afforded intermediates 42b–c, respectively. The next Sukuki coupling with 36, double C–C reduction with $Pd(OH)_2/C$ and ammonium formate and Boc deprotection with HCl in dioxane furnished final 25 and 26 with 33 and 16% yields after three steps starting from 36, respectively.

CONCLUSIONS

Based on our previous results on trisubstituted pyrimidines as a promising class of CDC42-PAK interaction inhibitors, we have here reported the design, synthesis, and extensive characterization of a new series of pyrimidine/triazine derivatives to block CDC42. The resulting SAR elucidated the key structural features that enhanced the antiproliferative activity against five cancer cell lines and the overall drug-like profile. Generally, the introduction of bicyclic heterocycles such as isoquinolyl-, indolyl-, indazolyl- and benzofuranyl- favorably impact the compounds' activity, as proven by the set of triazines (1-11), Table 1) and classic pyrimidine derivatives (25–26, Table 3). On the other hand, the shifting of the position of the piperidine moiety as in derivative 12 (Table 2) is well tolerated. This modification prompted the development of a new series of trisubstituted pyrimidines, such as 12-23, which unveil the favorable effect of methoxy and fluorinated groups (Table 3). Taken together, our new data on the inhibitory activity, drug-likeness profile (Table 4), in vitro binding data at the target supported also by computational studies, and the in vivo favorable pharmacokinetic profile, indicate the novel derivative 15 (ARN25499) as the most drug-like candidate of this novel chemical series of CDC42 inhibitors. Indeed, this compound showed improved in vitro metabolic stability compared to our previous leads ARN22089 and ARN25062,²³ and optimal kinetic and thermodynamic solubility in both neutral and salt forms. Importantly, ARN25499 shows also a slightly improved in vivo oral bioavailability and a notable efficacy to inhibit tumor growth in a PDX tumor mouse model. Clearly, since substituted pyrimidine represents a privileged chemical scaffold extensively characterized for its multiple biological properties,³⁴⁻³⁸ additional target selectivity and off-target activity tests will be performed to move into advanced preclinical studies of ARN25499, as similarly already performed for the previous lead ARN22089.²²

In conclusion, these promising findings elevate this compound as an additional lead drug candidate of our program on CDC42 inhibitors, which are now ready for further preclinical characterization and in vivo efficacy studies in other cancer models.

EXPERIMENTAL SECTION

Chemistry. *Chemistry General Considerations.* All the commercially available reagents and solvents were used as purchased from vendors without further purification. Dry solvents were purchased from Sigma-Aldrich. Automated column chromatography purifications were done using a Teledyne ISCO apparatus (CombiFlash Rf) with prepacked silica gel columns of different sizes (from 4 g up to 24 g) and mixtures of increasing polarity of cyclohexane and ethyl acetate (EtOAc) or dichloromethane (DCM) and methanol (MeOH). NMR

pubs.acs.org/jmc

Scheme 4. Synthesis of "Classic" Tri-Substituted Pyrimidine 24-26^a



^{*a*}Reagents and conditions: (a) LiHMDS (1 M in THF), aniline **38a**, **38k–l**, THF dry; (b) compound **36**, $Pd(Cl_2)(dppf) \cdot DCM$, K_2CO_3 (2M)aq, 1,4-dioxane dry; (c) $Pd(OH)_2/C$, NH_4COOH , MeOH dry, 80 °C; and (d) HCl (4 M in 1,4-dioxane dry), 1,4-dioxane dry.

data were collected on 400 or 600 MHz (¹H) and 100 or 150 MHz (¹³C). Spectra were acquired at 300 K, using deuterated dimethyl sulfoxide (DMSO- d_6) or deuterated chloroform (CDCl₃) as solvents. For ¹H NMR, data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, dd = double of doublets, t = triplet, q =quartet, m = multiplet), coupling constants (Hz), and integration. UPLC-MS analyses were run on a Waters ACQUITY UPLC-MS instrument consisting of a single quadrupole detector (SQD) mass spectrometer equipped with an electrospray ionization interface (ESI) and a photodiode array detector (PDA) from Waters Inc. (Milford, MA, USA). PDA range was 210-400 nm. The analyses were performed on an ACQUITY UPLC BEH C_{18} column (50 \times 2.1 mmID, particle size 1.7 μ m) with a VanGuard BEH C₁₈ precolumn (5 \times 2.1 mmID, particle size 1.7 μ m) (for method 1, 2 and 3) and an ACQUITY UPLC HSS T3 (50 \times 2.1 mmID, particle size 1.8 μ M) with a VanGuard HSS T3 precolumn (5×2.1 mmID, particle size 1.8 μ M) (for method 4). The mobile phase was 10 mM NH₄OAc in H₂O at pH 5 adjusted with AcOH (A) and 10 mM NH₄OAc in MeCN- H_2O (95:5) at pH 5 (B). Electrospray ionization in positive and negative modes was applied in the mass scan range of 100-500 Da. Depending on the analysis method used, a different gradient increasing the proportion of mobile phase B was applied. For analysis method 1, the mobile phase B proportion increased from 5 to 95% in 2.5 min. For analysis method 2, the mobile phase B proportion increased from 50 to 100% in 2.5 min. For analysis method 3, the mobile phase B proportion increased from 70 to 100% in 2.5 min. The analysis method 4, the mobile phase B portion increased from 0 to 50% in 2.5 min. The QC analysis was performed starting from a 10 mM stock solution of the test compound in DMSO- d_6 and further diluted 20-fold with MeCN-H₂O (1:1) for analysis. The analyses were performed on a Waters ACQUITY UPLC-MS system consisting of a single quadrupole detector mass spectrometer as described above. The analyses were run on an ACQUITY UPLC BEH C18 column $(100 \times 2.1 \text{ mmID}, \text{ particle size } 1.7 \ \mu\text{m})$ with a VanGuard BEH C₁₈ precolumn (5 \times 2.1 mmID, particle size 1.7 μ m). The mobile phase was 10 mM NH₄OAc in H₂O at pH 5 adjusted with AcOH (A) and 10 mM NH₄OAc in MeCN-H₂O (95:5) at pH 5 (B) with 0.5 mL/ min as flow rate. A linear gradient was applied: 0-0.2 min: 10%B, 0.2-6.2 min: 10-90%B, 6.2-6.3 min: 90-100%B, 6.3-7.0 min: 100%B (QC method). High-resolution mass spectrometry (HRMS) for accurate mass measurements was performed on a Sciex TripleTOF High-resolution LC-MS using a Waters UPLC ACQUITY chromatographic system (from Waters Inc., Milford, MA, USA) coupled to a TripleTOF 5600+ Mass Spectrometer (from Sciex, Warrington, UK) equipped with a NanoSpray III Ion source. The analyses were run on an ACQUITY UPLC BEH C₁₈ column (50 \times 2.1 mmID, particle size 1.7 μ m), using H₂O + 0.1% HCOOH (A) and MeCN + 0.1% HCOOH as mobile phase. All final compounds displayed ≥95% purity as determined by NMR and UPLC-MS (UV at 215 nm) analysis unless otherwise indicated. Compounds 27, 31a-i, 34-36, 38a-j were purchased from Sigma-Aldrich or Fluorochem and used

as such without further purification. Intermediates 28–30, 41, 42–43a were synthesized as previously reported. $^{22,23}_{\rm -}$

General Procedure 1: SNAr. Method A. To a DCM solution (0.5M) of cyanuric chloride (1.3 equiv), corresponding aniline (1.0 equiv) was dropwise added at 0 °C, followed by DIPEA (1.1 equiv). After 10 min, H₂O was slowly added at the same temperature to quench the excess reagent, and the resulting mixture was left to stir for 10–15 min. The organic layer was separated and dried with Na2SO4, the solvent was evaporated, and the resulting crude was purified by silica gel chromatography.

Method B. A solution (0.15 M) of 6-phenyl-2,4-dichloropyrimidine 42 (1.00 equiv) with a suitable aniline of type 38 (1.00 mmol) in THF was cooled to -60 °C. To this solution was added dropwise LiHMDS (1.0 M in THF, 2.5 equiv). After the complete conversion of the starting material, water was added, and the mixture was extracted with EtOAc. The combined organic layers were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. Final normal phase chromatographic purification (Cyclohexane/EtOAc) provided the desired products.

General Procedure 2: Suzuki Cross-Coupling. To a degassed 1,4dioxane solution (0.13 M) of heteroaryl halide (1.0 equiv), suitable boronic acid (1.2 equiv), Pd(dppf)Cl₂·DCM (0.05 equiv) and K₂CO₃ (2 M)aq (2.0 equiv) were added. The resulting mixture was sparged with Argon for a further 10 min and heated at different temperatures in dependence of the substrate: 80 °C for compound **28**, 100 °C for compound **30**, and 120 °C for compound of type **42**. Then, H₂O and EtOAc were added and separated; aqueous layers were extracted two times with EtOAc and collected organic layers were dried over Na₂SO₄. The solvent was evaporated and the resulting crude was purified by silica gel chromatography.

General Procedure 3: Buchwald Reaction for the Obtainment of Compounds **39a**–j. A mixture of compound **37** (1 equiv), proper aniline **38a**–j (1.2 equiv), Pd(OAc)2 (5 mol %), (\pm)-BINAP or Xantphos (5 mol %), Cs₂CO₃ (1.5 equiv) in 1,4-dioxane dry (0.15M) stirred at 120 °C until complete consumption of starting material. After that, water and brine were added, the aqueous layer was extracted with EtOAc, and collected organic layers were dried over Na₂SO₄, filtered, and concentrated under a vacuum. Intermediate 5a-j were purified by silica.

General Procedure 4: Double C–C Reduction. Method A. Under N_2 atmosphere, a suspension of intermediate 32a-i, 39a-j, 43b-c (1 equiv), ammonium formate (6 equiv), $Pd(OH)_2/C$ (20% of starting material weight) in MeOH dry (0.04 M solution) was stirred at reflux temperature until reaction completion. The catalyst was filtered off through a Celite coarse patch, and the resulting filtrate concentrated to dryness at low pressure. Final chromatographic normal phase purification (cyclohexane/EtOAc) afforded pure desired products.

Method B. Reduction with Et_3SiH . Under the N_2 atmosphere, Et_3SiH (10 equiv) was dropwise added to a mixture of substrate **32i** (1.0 equiv) and Pd/C (20% w/w of starting material weight) in EtOH (0.5 M). When no intermediate was detected by UPLC or TLC, the

catalyst was filtered off through a Celite coarse patch, and the resulting filtrate concentrated to dryness at low pressure. The resulting crude was purified by silica gel chromatography.

General Procedure 5: Boc Removal. Under the N_2 atmosphere, HCl (4 M in dioxane) (10 equiv) was added to a solution of intermediate of type 6, 16, 17 (1 equiv) in 1,4-dioxane dry (0.06 M). After completion of reaction, NaOH (2 M) aq was added until pH = 7, an aqueous layer was extracted with EtOAc, and collected organic layers were dried over Na_2SO_4 , filtered, and concentrated under vacuum. Final purification by alumina (DCM/MeOH or DCM/ MeOH·NH₃) afforded the pure desired compound of type 7 and 18.

General Procedure 6: Methyl and Boc Removal. Under the N_2 atmosphere, BBr₃ (1 M in DCM) (6 equiv) was added to a solution of intermediate of type 6 (1 equiv) in CHCl₃ dry (0.06 M). After completion of the reaction, MeOH was added, and the mixture was concentrated under vacuum. The crude was purified by trituration with EtOAc, obtaining pure products.

4-(Piperidin-4-yl)-6-(pyrimidin-5-yl)-N-(3-(trifluoromethyl)phenyl)-1,3,5-triazin-2-amine (2). Compound 2 was synthesized following general procedure 5 using compound 33a (50 mg, 0.10 mmol), HCl 4 M 1,4-dioxane solution (0.25 mL, 1.0 mmol), and 1,4dioxane (1.7 mL). After 24 h, no conversion occurred; thus, HCl 4 M (0.28 mL) was added, and the reaction mixture was stirred overnight. After a further 24 h, HCl 4 M (0.28 mL) was again added, and a standard workup was performed after 24 h. Purification by silica (elution by a gradient from 100/0 to 90/10 DCM/MeOH(NH₃ 1N)) afforded a white solid as the product (25 mg, 62% yield). UPLC-MS: $t_{R} = 0.52 \text{ min (method 1)}$. MS (ESI) m/z: 402.0 [M + H]⁺, calcd for $\tilde{C}_{19}H_{19}F_{3}N_{7}$ [M + H]⁺: 402.4. HRMS (ESI) m/z: 402.1649 calcd for $C_{19}H_{19}F_{3}N_{7}$ [M + H]⁺; found m/z: 402.1660. ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.74 (s, 1H), 9.58 (s, 2H), 9.41 (s, 1H), 8.57–8.29 (m, 1H), 8.00 (s, 1H), 7.63 (s, 1H), 7.44 (d, J = 7.7 Hz, 1H), 3.04 (dt, J = 12.3, 3.4 Hz, 2H), 2.81 (s, 1H), 2.61 (td, J = 12.0, 2.5 Hz, 2H), 2.06-1.86 (m, 2H), 1.79–1.65 (m, 2H). ¹³C NMR (151 MHz, DMSO-d₆) δ 182.0 (Cq), 167.5 (Cq), 164.0 (Cq), 160.8 (CH), 156.5 (CH, 2C), 139.6 (Cq), 130.0 (CH), 129.4 (Cq), 129.3 (q², J_{CF} = 31.7 Hz, Cq), 124.2 (q¹, J_{CF} = 272.8 Hz), 123.8 (CH), 119.4 (CH), 116.6 (CH), 45.8 (CH₂, 2C), 44.9 (CH), 30.8 (CH₂, 2C). ¹⁹F NMR (565 MHz, DMSO- d_6) δ -60.32.

4-(Isoauinolin-5-vl)-6-(piperidin-4-vl)-N-(3-(trifluoromethvl)phenyl)-1,3,5-triazin-2-amine (3). Compound 3 was synthesized following general procedure 5 using compound 33b (68 mg, 0.123 mmol), HCl 4 M 1,4-dioxane solution (0.31 mL, 1.23 mmol), 1,4dioxane (2.0 mL). After 16 h, a standard workup has been performed. Purification by silica (elution by a gradient from 100/0 to 90/10 DCM/MeOH(NH₃ 1N)) afforded a yellowish unclean solid that was subjected to trituration with petroleum ether/EtOAc/MeOH 85:13:2. A pure white solid was collected by trituration (27 mg, 49% yield). UPLC-MS: $t_R = 1.91 \text{ min (method 1)}$. MS (ESI) m/z: 451.0 [M + H]⁺, calcd for $C_{24}H_{22}F_3N_6$ [M + H]⁺: 451.5. HRMS (ESI) m/z: 451.1853 calcd for $C_{24}H_{22}F_3N_6$ [M + H]⁺; found m/z: 451.1861. ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.68 (bs, 1H), 9.43 (s, 1H), 8.89 (bs, 1H), 8.58 (d, J = 6.7 Hz, 2H), 8.52-8.34 (m, 2H), 8.02 (bs, 1H), 7.86 (t, J = 7.7 Hz, 1H), 7.60 (t, J = 7.9 Hz, 1H), 7.42 (d, J = 7.7 Hz, 1H), 3.09 (dd, J = 12.2, 3.4 Hz, 2H), 2.87 (t, J = 12.2 Hz, 1H), 2.67 (td, J = 12.1, 2.5 Hz, 2H), 2.12-1.95 (m, 2H), 1.80 (dd, J = 12.1, 4.0)Hz, 2H). ¹³C NMR (151 MHz, DMSO- d_6) δ 181.8 (Cq), 172.3 (Cq), 164.5 (Cq), 153.7 (CH), 144.4 (CH, 2C), 140.4 (Cq), 134.2 (CH), 133.5 (Cq), 132.7 (Cq), 132.3 (CH), 130.3 (CH), 129.9 (q², J_{CF} = 31.2 Hz, Cq), 129.2 (Cq), 127.2 (CH), 124.7 (q^1 , $J_{CF} = 272.8$ Hz, Cq), 124.2 (CH), 119.7 (CH), 119.0 (CH), 117.0 (CH), 46.0 (CH₂, 2C), 45.1 (CH), 30.9 (CH₂, 2C). ¹⁹F NMR (565 MHz, DMSO-d₆) $\delta - 60.28$

Ethyl-(1H-indol-5-yl)-6-(piperidin-4-yl)-N-(3-(trifluoromethyl)phenyl)-1,3,5-triazin-2-amine (4). Compound 4 was synthesized following general procedure 5 using compound 33c (100 mg, 0.18 mmol), HCl 4 M 1,4-dioxane solution (0.45 mL, 1.8 mmol), and 1,4dioxane (3.0 mL). After 3.5 h, a standard workup has been performed. Purification by silica (elution by a gradient from 95/5 to 80/20 DCM/MeOH (1N NH₃)) afforded not pure compound that has been

subjected to trituration with petroleum ether giving the pure product as a white solid (40 mg, 49% yield). UPLC-MS: $t_R = 1.18$ min (method 2). MS (ESI) m/z: 453.0 [M + H]⁺, calcd for C₂₄H₂₄F₃N₆ $[M + H]^+$: 453.2. HRMS (ESI) m/z: 453.2009 calcd for $C_{24}H_{24}F_3N_6$ $[M + H]^+$; found *m/z*: 453.2018. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.41 (s, 1H), 8.75 (d, J = 1.6 Hz, 1H), 8.61 (s, 1H), 8.28 (dd, J = 8.8, 1.7 Hz, 1H), 7.98 (d, J = 8.3 Hz, 1H), 7.66–7.54 (m, 2H), 7.43 (d, J = 3.1 Hz, 1H), 7.40 (d, J = 7.8 Hz, 1H), 6.60 (d, J = 3.1 Hz, 1H), 3.85 (s, 3H), 3.14–3.02 (m, 2H), 2.78 (t, J = 11.6 Hz, 1H), 2.72–2.57 (m, 2H), 2.06-1.89 (m, 2H), 1.85-1.68 (m, 2H). ¹³C NMR (151 MHz, DMSO-d₆) & 181.1 (Cq), 171.4 (Cq), 164.1 (Cq), 140.3 (Cq), 138.8 (Cq), 131.3 (CH), 129.8 (CH), 129.4 (q^2 , $J_{CF} = 31.2$ Hz, Cq), 127.3 $(q^1, J_{CF} = 214.4 \text{ Hz}, Cq), 125.2 (Cq), 123.4 (CH), 121.8 (CH), 121.3$ (CH), 118.7 (CH), 116.2 (CH), 109.7 (CH), 102.0 (CH), 45.8 (CH₂, 2C), 45.0 (CH), 32.7 (CH₃), 30.8 (CH₂, 2C). ¹⁹F NMR (565 MHz, DMSO- d_6) δ -61.26.

4-(1H-Indazol-5-yl)-6-(piperidin-4-yl)-N-(3-(trifluoromethyl)phenyl)-1,3,5-triazin-2-amine (Compound 5). Compound 5 was synthesized following general procedure 5 using compound 33d (27 mg, 0.05 mmol), HCl 4 M 1,4-dioxane solution (0.125 mL, 0.5 mmol), and 1,4-dioxane dry (0.08 mL). After 6 h, a standard workup has been performed. Purification by silica (elution by a gradient from 99/1 to 85/15 DCM/MeOH(NH₃ 1N)) afforded a white solid as the product (11 mg, 50% yield). UPLC-MS: $t_R = 0.76$ min (method 2). MS (ESI) m/z: 440.0 [M + H]⁺, calcd for $C_{22}H_{21}F_3N_7$ [M + H]⁺: 439.5. HRMS (ESI) m/z: 440.1805 calcd for $C_{22}H_{21}F_3N_7$ [M + H]⁺; found m/z: 440.1814. ¹H NMR (400 MHz, DMSO- d_6) δ 13.37 (s, 1H), 10.49 (s, 1H), 8.93 (s, 1H), 8.50 (s, 1H), 8.43 (dd, J = 8.8, 1.6 Hz, 1H), 8.28 (s, 1H), 8.03 (d, J = 8.1 Hz, 1H), 7.67 (d, J = 8.9 Hz, 1H), 7.62 (t, J = 8.1 Hz, 1H), 7.42 (d, J = 7.7 Hz, 1H), 3.15–3.06 (m, 2H), 2.88-2.78 (m, 1H), 2.74-2.63 (m, 2H), 2.05-1.97 (m, 2H), 1.79 (dd, J = 12.0, 3.9 Hz, 2H). ¹³C NMR (151 MHz, DMSO- d_{δ}) δ 181.1 (Cq), 170.7 (Cq), 164.2 (Cq), 141.8 (Cq), 140.1 (Cq), 135.2 (CH), 129.8 (CH), 129.4 (q^2 , J_{CF} = 31.4 Hz, Cq), 128.1 (Cq), 125.7 (CH), 124.3 (q¹, J_{CF} = 272.3 Hz, Cq), 123.6 (CH), 123.1 (Cq), 122.3 (CH), 118.9 (CH), 116.4 (CH), 110.3 (CH), 45.4 (CH₂, 2C), 44.3 (CH), 30.2 (CH₂, 2C). ¹⁹F NMR (565 MHz, DMSO- d_6) δ -60.28.

4-(Benzofuran-3-yl)-6-(piperidin-4-yl)-N-(3-(trifluoromethyl)phenyl)-1,3,5-triazin-2-amine (Compound 6). Compound 6 was synthesized following general procedure 5 using 33e (102 mg, 0.19 mmol), HCl 4 M 1,4-dioxane solution (0.48 mL, 1.90 mmol), 1,4dioxane (3.2 mL). After 7 h, HCl 4 M (0.24 mL) was added again and the reaction mixture was stirred overnight. Further, HCl 4 M (0.24 mL) has been added and after 16 h standard workup has been performed. The crude product was purified by neutral alumina (elution by gradient DCM/EtOH 100:0 to 90:10) giving the pure product as a white solid (3.5 mg, 4% yield). UPLC-MS: $t_R = 1.27$ min (method 2). MS (ESI) m/z: 440.0 [M + H]⁺, calcd for C₂₃H₂₁F₃N₅O $[M + H]^+$: 440.5. ¹H NMR (400 MHz, DMSO- d_6) δ 10.50 (s, 1H), 8.85 (s, 1H), 8.34 (d, J = 54.9 Hz, 2H), 8.03 (d, J = 8.2 Hz, 1H), 7.72 (d, J = 8.1 Hz, 1H), 7.63 (t, J = 8.0 Hz, 1H), 7.51-7.32 (m, 3H),3.10-3.02 (m, 2H), 2.84-2.73 (m, 1H), 2.68-2.56 (m, 2H), 2.05-1.91 (m, 2H), 1.81–1.67 (m, 2H). ¹³C NMR (151 MHz, DMSO-d₆) δ 181.3 (Cq), 167.8 (Cq), 163.9 (Cq), 155.5 (Cq), 150.7 (CH), 139.9 (Cq), 129.9 (CH), 129.4 (q², J_{CF} = 31.2 Hz, Cq), 125.2 (CH), 124.6 (Cq), 124.3 (q^1 , J_{CF} = 273.2 Hz, Cq), 123.9 (CH, 2C), 122.7 (CH), 119.8 (Cq), 119.2 (CH), 116.5 (CH), 111.8 (CH), 45.9 (CH₂, 2C), 44.9 (CH), 30.9 (CH₂, 2C). ¹⁹F NMR (565 MHz, DMSO-d₆) $\delta - 60.1.$

4-(1-Methyl-1H-pyrazol-4-yl)-6-(piperidin-4-yl)-N-(3-(trifluoromethyl)phenyl)-1,3,5-triazin-2-amine (Compound 7). Compound 7 was synthesized following general procedure 5 using compound 33f (75 mg, 0.14 mmol), HCl, and 4 M 1,4-dioxane solution (0.35 mL, 1.4 mmol), 1,4-dioxane (2.3 mL). After 16 h, a standard workup has been performed. Purification by silica (elution by a gradient from 100/0 to 90/10 DCM/MeOH(1N NH₃)) afforded the pure product as a white solid (43 mg, 76% yield). UPLC-MS: t_R = 1.76 min (method 1). MS (ESI) *m/z*: 404.0 [M + H]⁺, calcd for C₁₉H₂₁F₃N₇ [M + H]⁺: 403.4. HRMS (ESI) *m/z*: 404.1805 calcd for C₁₉H₂₁F₃N₇ [M + H]⁺; found *m/z*: 404.1815. ¹H NMR (600 MHz, DMSO- d_6) δ 10.37 (s, 1H), 8.42 (s, 1H), 8.41 (s, 1H), 8.07–7.96 (m, 2H), 7.58 (t, J = 8.0 Hz, 1H), 7.38 (d, J = 6.0 Hz, 1H), 3.93 (s, 3H), 3.04 (dt, J = 12.3, 3.4 Hz, 2H), 2.72–2.67 (m, 1H), 2.60 (td, J = 12.1, 2.5 Hz, 2H), 1.97–1.85 (m, 2H), 1.69 (qd, J = 12.1, 3.9 Hz, 2H). ¹³C NMR (151 MHz, DMSO- d_6) δ 181.2 (Cq), 167.1 (Cq), 163.9 (Cq), 140.2 (Cq), 139.3 (CH), 133.1 (CH), 129.8 (CH), 129.3 (q², $J_{CF} = 31.3$ Hz, Cq), 124.3 (q¹, $J_{CF} = 272.4$ Hz, Cq), 123.4 (CH), 120.8 (Cq), 118.7 (q³, $J_{CF} = 4.0$ Hz, CH), 116.1 (q³, $J_{CF} = 4.2$ Hz, CH), 45.7 (CH₂), 44.7 (CH), 38.6 (CH₃), 30.6 (CH₂). ¹⁹F NMR (565 MHz, DMSO- d_6) δ –60.31.

4-(1-Methyl-1H-imidazol-5-yl)-6-(piperidin-4-yl)-N-(3-(trifluoromethyl)phenyl)-1,3,5-triazin-2-amine (Compound 8). Compound 8 was synthesized following general procedure 5 using compound 33g (83 mg, 0.17 mmol), HCl, and 4 M 1,4-dioxane solution (0.42 mL, 1.7 mmol) in 1,4-dioxane (2.8 mL). After 2 days, a standard workup has been performed. Purification by silica (elution by a gradient from 98/2 to 90/10 DCM/MeOH(NH₃ 1N)) afforded unclean compound, that was subjected to trituration with pentane/ EtOAc 9/1 furnishing a white solid as the product (48 mg, 72% yield). UPLC-MS: $t_R = 1.65 \text{ min} (\text{method } 2)$. MS (ESI) m/z: 404.0 $[M + H]^+$, calcd for $C_{19}H_{21}F_3N_7[M + H]^+$: 404.4. HRMS (ESI) m/z: 404.1805 calcd for $C_{19}H_{21}F_3N_7$ [M + H]⁺; found m/z: 404.1815. ¹H NMR (600 MHz, DMSO- d_6) δ 10.41 (s, 1H), 8.38 (s, 1H), 7.93 (s, 1H), 7.91 (s, 1H), 7.85 (s, 1H), 7.59 (t, J = 8.0 Hz, 1H), 7.40 (d, J = 7.7 Hz, 1H), 4.04 (s, 3H), 3.14–2.97 (m, 2H), 2.73 (t, J = 11.4 Hz, 1H), 2.63 (t, J = 12.1 Hz, 2H), 2.01–1.89 (m, 2H), 1.70 (qd, J = 12.2, 3.9 Hz, 2H). ¹³C NMR (151 MHz, DMSO-*d*₆) δ 180.9 (Cq), 164.7 (Cq), 163.6 (Cq), 143.8 (CH), 139.9 (Cq), 136.1 (CH), 129.8 (CH), 129.3 (q^2 , J_{CF} = 31.7 Hz, Cq), 129.0 (Cq), 124.2 (q^1 , J_{CF} = 271.8 Hz), 123.8 (CH), 119.1 (CH), 116.4 (CH), 45.6 (CH₂, 2C), 44.5 (CH), 35.2 (CH₃), 30.4 (CH₂, 2C). ¹⁹F NMR (565 MHz, DMSO- d_6) δ -60.28.

4-(Piperidin-4-yl)-6-(1H-pyrrol-2-yl)-N-(3-(trifluoromethyl)phenyl)-1,3,5-triazin-2-amine (Compound 9). Compound 9 was synthesized following general procedure 5 using compound 33h (60 mg, 0.10 mmol), HCl 4 M 1,4-dioxane solution (0.25 mL, 1.0 mmol), and 1,4-dioxane (1.7 mL). After 2 days, a standard workup has been performed. Purification by silica (elution by a gradient from 99:1 to 90:10 DCM/MeOH(NH₃ 1N)) afforded a white solid as the product (15 mg, 38% yield). UPLC-MS: $t_R = 1.91 \text{ min (method 2)}$. MS (ESI) m/z: 389.0 [M + H]⁺, calcd for C₁₉H₂₀F₃N₆ [M + H]⁺: 389.4. HRMS (ESI) m/z: 389.1696 calcd for $C_{19}H_{20}F_3N_6$ [M + H]⁺; found m/z: 389.1705. ¹H NMR (600 MHz, DMSO-*d*₆) δ 11.66 (s, 1H), 10.26 (s, 1H), 8.47 (s, 1H), 8.11–7.92 (m, 1H), 7.57 (t, J = 8.0 Hz, 1H), 7.36 (d, J = 7.7 Hz, 1H), 7.08-7.02 (m, 2H), 6.29-6.24 (m, 1H), 3.05(dd, J = 12.1, 3.4 Hz, 2H), 2.74–2.67 (m, 1H), 2.62 (dd, J = 12.0, 2.5 Hz, 2H), 1.99–1.87 (m, 2H), 1.74 (dd, J = 12.2, 3.8 Hz, 2H). ¹³C NMR (151 MHz, DMSO- d_6) δ 180.8 (Cq), 164.5 (Cq), 163.8 (Cq), 140.4 (Cq), 129.8 (CH), 129.3 (q^2 , J_{CF} = 31.7 Hz, Cq), 128.7 (Cq), 124.3 (q¹, J_{CF}= 271.8 Hz, Cq), 124.2 (CH), 123.3 (CH), 118.5 (q³, J_{CF} = 3.5 Hz, CH), 116.0 (CH), 113.7 (CH), 110.3 (CH), 45.7 (CH₂, 2C), 44.7 (CH), 30.5 (CH₂, 2C). ¹⁹F NMR (565 MHz, DMSO-d₆) $\delta = 60.3$

4-(Piperidin-4-yl)-6-(pyrrolidin-2-yl)-N-(3-(trifluoromethyl)phenyl)-1,3,5-triazin-2-amine (Compound 10). Compound 10 was synthesized following general procedure 5 using compound 33h' (123 mg, 0.21 mmol), HCl 4 M 1,4-dioxane solution (0.53 mL, 2.1 mmol), 1,4-dioxane (3.5 mL). After 16 h, no conversion occurred; thus, HCl 4 M (3.5 mL) was added and the reaction mixture was stirred over the weekend. Then, a standard workup has been performed. Purification by silica (elution by gradient 100/0 to 90/10 DCM/MeOH (1 N NH_3) afforded a yellowish unclean solid that was subjected to trituration with pentane (1.5 mL). The pure product was collected as a white solid after filtration (16 mg, 20% yield). UPLC-MS: $t_R = 1.68$ min (method 1). MS (ESI) m/z: 393.0 [M + H]⁺, calcd for $C_{19}H_{24}F_{3}N_{6}$ [M + H]⁺: 393.4. ¹H NMR (600 MHz, DMSO-d₆) δ 10.46 (s, 1H), 8.40 (s, 1H), 7.92 (d, J = 8.3 Hz, 1H), 7.56 (t, J = 8.0 Hz, 1H), 7.37 (d, J = 7.7 Hz, 1H), 3.99 (s, 1H), 3.10–3.04 (m, 1H), 3.00 (dt, J = 12.3, 3.5 Hz, 2H), 2.86-2.79 (m, 1H), 2.72-2.63 (m, 1H), 2.56 (td, J = 12.1, 2.5 Hz, 2H), 2.16–2.09 (m, 1H), 1.90–1.78

(m, 3H), 1.76–1.69 (m, 2H), 1.63 (qd, J = 12.2, 4.0 Hz, 2H). ¹³C NMR (151 MHz, DMSO- d_6) δ 181.2 (Cq), 173.0 (Cq), 163.8 (Cq), 140.0 (Cq), 129.8 (CH), 129.4 (q², J_{CF} = 31.7 Hz, Cq), 124.2 (q¹, J_{CF} = 272.8 Hz, Cq), 123.4 (CH), 118.9 (CH), 116.2 (CH), 62.9 (CH), 46.9 (CH₂), 45.8 (CH₂, 2C), 44.8 (CH), 32.0 (CH₂), 30.9 (CH₂, 2C), 25.8 (CH₃). ¹⁹F NMR (565 MHz, DMSO- d_6) δ –60.3.

4-(1,2,3,6-Tetrahydropyridin-4-yl)-6-(thiophen-3-yl)-N-(3-(trifluoromethyl)phenyl)-1,3,5-triazin-2-amine (Compound 11). Compound 11 was synthesized following general procedure 5 using compound 32i (48 mg, 0.095 mmol), HCl 4 M 1,4-dioxane solution (0.25 mL, 0.10 mmol), and 1,4-dioxane (1.6 mL). After 16 h, no conversion occurred; thus, HCl 4 M (0.78 mL) was added and the reaction mixture was stirred over the weekend. Then, standard workup has been. Purification by silica (elution by a gradient from 99/1 to 96/4 DCM/MeOH(NH₃ 1N)) afforded unclean compound, that was subjected to trituration with pentane/EtOAc 1/1 furnishing a white solid as the product (15 mg, 39% yield). UPLC-MS: $t_R = 0.90$ min (method 2). MS (ESI) m/z: 404.0 [M + H]⁺, calcd for $C_{19}H_{17}F_{3}N_{5}S [M + H]^{+}: 404.4.$ ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.47 (s, 1H), 8.53 (dd, J = 3.0, 1.2 Hz, 1H), 8.50 (s, 1H), 7.98 (s, 1H), 7.83 (dd, J = 5.0, 1.2 Hz, 1H), 7.72 (t, J = 5.0, 3.1 Hz, 1H), 7.60 (t, J = 8.0 Hz, 1H), 7.53-7.46 (m, 1H), 7.40 (d, J = 7.7 Hz, 1H),3.62-3.54 (m, 2H), 2.99 (t, J = 5.6 Hz, 2H), 2.60-2.53 (m, 2H). ¹³C NMR (151 MHz, DMSO-d₆) δ 171.6, 167.4, 164.8, 140.6, 140.6, 135.6, 134.1, 131.4, 130.3, 129.8 (q², J_{CF} = 31.7 Hz), 128.1, 127.3, 124.8 (q^1 , J_{CF} = 271.8 Hz), 124.0, 119.4 (q^3 , J_{CF} = 3.7 Hz, CH), 116.73 (q^3 , J_{CF} = 4.5 Hz, CH), 45.0 (CH₂), 42.3 (CH₂), 24.7 (CH₂). ¹⁹F NMR (565 MHz, DMSO- d_6) δ -60.29.

N1,N1-Dimethyl-N3-(4-phenyl-6-(piperidin-4-yl)pyrimidin-2-yl)benzene-1,3-diamine (Compound 12). Title compound was synthesized following the general procedure 5 using intermediate 40a (58 mg, 0.12 mmol) and HCl (4 M in dioxane) (0.32 mL) in 1,4dioxane dry (2.1 mL). Purification by alumina (elution by a gradient from 100/0 to 98/2 DCM/MeOH·NH₃ 1N) afforded pure product 12 (15.5 mg, 33% yield). UPLC-MS: $t_R = 1.08 \text{ min} (\text{method } 1)$. MS (ESI) m/z: 374.3 [M + H]⁺, calcd for C₂₃H₂₈N₅ [M + H]⁺: 374.5. HRMS (ESI) m/z: 374.2339 calcd for C₂₃H₂₈N₅ [M + H]⁺; found m/z*z*: 374.2349. ¹H NMR (400 MHz, DMSO- d_6) δ 9.31 (s, 1H), 8.23– 8.10 (m, 2H), 7.58 (s, 1H), 7.55-7.49 (m, 3H), 7.26 (s, 1H), 7.08 (t, I = 7.9 Hz, 1H), 7.06 (s, 1H), 6.35 (dt, I = 7.2, 2.3 Hz, 1H), 3.10-3.00 (m, 3H), 2.92 (s, 6H), 2.69 (tt, J = 11.8, 3.8 Hz, 1H), 2.59 (td, J = 12.0, 2.5 Hz, 2H), 1.83 (d, J = 12.1 Hz, 2H), 1.68 (qd, J = 12.2, 4.0 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 175.2 (Cq), 163.7 (Cq), 160.2 (Cq), 150.9 (Cq), 141.6 (Cq), 137.2 (Cq), 130.6 (CH), 128.7 (CH, 2C), 126.9 (CH, 2C), 107.4 (CH), 105.9 (CH), 105.1 (CH), 103.2 (CH), 46.2 (CH₂, 2C), 44.4 (CH), 40.3 (CH₃, 2C), 31.8 (CH₂, 2C).

4-Phenyl-6-(piperidin-4-yl)-N-(3-(trifluoromethyl)phenyl)pyrimidin-2-amine (Compound 13). The title compound was synthesized following the general procedure 5 previously described using intermediate 40b (45 mg, 0.09 mmol) and HCl (4 M in dioxane) (0.22 mL) in 1,4-dioxane dry (1.5 mL). Purification by alumina (elution by a gradient from 100/0 to 90/10 DCM/MeOH) afforded pure product 13 (16.2 mg, 45% yield). UPLC-MS: $t_R = 1.17$ min (method 2). MS (ESI) m/z: 399.2 $[M + H]^+$, calcd for $C_{22}H_{21}F_{3}N_{4}$ [M + H]⁺: 399.2. HRMS (ESI) m/z: 399.1791 calcd for $C_{22}H_{21}F_{3}N_{4}$ [M + H]⁺; found m/z: 399.1794. ¹H NMR (400 MHz, DMSO- d_6) δ 9.95 (s, 1H), 8.60 (bs, 1H), 8.18 (m, 2H), 7.96 (dd, J =8.3, 1.4 Hz, 1H), 7.30–7.26 (m, 4H), 7.40 (s, 1H), 7.27 (d, J = 7.7 Hz, 1H), 3.06 (dt, J = 12.2, 3.2 Hz, 2H), 2.74 (tt, J = 11.9, 3.2 Hz, 1H), 2.60 (td, J = 12.0, 2.5 Hz, 2H), 1.85 (d, J = 12.5 Hz, 2H), 1.70 (qd, J = 12.2, 4.0 Hz, 2H). ¹³C NMR (101 MHz, DMSO- d_6) δ 175.6 (Cq), 163.9 (Cq), 159.8 (Cq), 141.7 (Cq), 136.8 (Cq), 130.8 (CH), 129.5 (CH), 129.3 (q^2 , J_{CF} = 30.9 Hz, Cq), 128.8 (CH, 2C), 126.9 (CH, 2C), 124.4 (q^1 , J_{CF} = 271.8 Hz, Cq), 122.0 (CH), 117.1 (q^3 , J_{CF} = 3.6 Hz, CH), 114.5 (q^3 , J_{CF} = 4.3 Hz, CH), 106.3 (CH), 46.2 (CH₂, 2C), 44.3 (CH), 31.8 (CH₂, 2C). ¹⁹F NMR (376 MHz, DMSO- d_6) δ - 61.3.

4-Phenyl-6-(piperidin-4-yl)-N-(3-(trifluoromethoxy)phenyl)pyrimidin-2-amine (Compound 14). The title compound was

synthesized following the general procedure 5 previously described using intermediate 40c (70 mg, 0.14 mmol) and HCl (4 M in dioxane) (0.34 mL) in 1,4-dioxane dry (2.3 mL). Purification by alumina (elution by a gradient from 100/0 to 95/5 DCM/MeOH) afforded pure product 14 (23.1 mg, 41% yield). UPLC-MS: $t_R = 2.41$ min (method 1). MS (ESI) m/z: 415.0 [M + H]⁺, calcd for $C_{22}H_{22}F_3N_4O [M + H]^+: 415.2.$ HRMS (ESI) m/z: 415.1740 calcd for $C_{22}H_{22}F_{3}N_{4}O [M + H]^{+}$; found m/z: 415.1747. ¹H NMR (400 MHz, DMSO-d₆) δ 9.91 (s, 1H), 8.22 (s, 1H), 8.17 (m, 2H), 7.70 (dd, J = 8.4, 2.0 Hz, 1H), 7.57-7.51 (m, 3H), 7.41 (t, J = 8.2 Hz, 1H)1H), 7.38 (s, 1H), 6.86 (d, J = 8.0 Hz, 1H), 3.05 (dt, J = 12.0, 2.7 Hz, 2H), 2.72 (tt, J = 11.7, 3.7 Hz, 1H), 2.60 (td, J = 12.1, 2.5 Hz, 2H), 1.84 (d, J = 12.4 Hz, 2H), 1.69 (qd, J = 12.2, 3.9 Hz, 2H). ¹³C NMR (101 MHz, DMSO-d₆) δ 175.6 (Cq), 164.0 (Cq), 159.7 (Cq), 148.7 (Cq), 142.7 (Cq), 136.9 (Cq), 130.8 (CH), 130.0 (CH), 128.8 (CH, 2C), 127.0 (CH, 2C), 120.6 (q¹, J_{CF} = 255.6 Hz, Cq), 117.2 (CH), 112.9 (CH), 110.4 (CH), 106.2 (CH), 46.2 (CH₂, 2C), 44.3 (CH), 31.7 (CH₂, 2C). ¹⁹F NMR (565 MHz, DMSO- d_6) δ – 56.5.

N-(3,4-Dimethoxyphenyl)-4-phenyl-6-(piperidin-4-yl)pyrimidin-2-amine (Compound 15 - ARN25499). The title compound was synthesized following the general procedure 5 using intermediate 40d (30 mg, 0.06 mmol) and HCl (4 M in dioxane) (0.15 mL) in 1,4dioxane dry (1.0 mL). Purification by alumina (elution by a gradient from 100/0 to 95/5 DCM/MeOH) afforded pure compound 15 -ARN25499 (12.4 mg, 38% yield). UPLC-MS: $t_R = 1.79$ min (method 1). MS (ESI) m/z: 391.0 [M + H]⁺, calcd for C₂₃H₂₇N₄O₂ [M + H]⁺: 391.2. HRMS (ESI) m/z: 391.2129 calcd for $C_{23}H_{27}N_4O_2 [M + H]^+$; found m/z: 391.2146. ¹H NMR (400 MHz, DMSO- d_6) δ 9.35 (s, 1H), 8.21–8.11 (m, 2H), 7.78 (d, J = 2.4 Hz, 1H), 7.57–7.49 (m, 3H), 7.26 (dd, J = 8.6, 2.4 Hz, 1H), 7.25 (s, 1H), 6.90 (d, J = 8.7 Hz, 1H), 3.79 (s, 3H), 3.72 (s, 3H), 3.05 (d, J = 12.0 Hz, 2H), 2.70 (tt, J= 11.9, 4.4 Hz, 1H), 2.59 (td, J = 12.0, 2.5 Hz, 2H), 1.82 (d, J = 12.4 Hz, 2H), 1.70 (qd, J = 12.2, 4.0 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 175.2 (Cq), 163.7 (Cq), 160.1 (Cq), 148.5 (Cq), 143.4 (Cq), 137.2 (Cq), 134.8 (Cq), 130.6 (CH), 128.8 (CH, 2C), 126.9 (CH, 2C), 112.4 (CH), 110.4 (CH), 105.0 (CH), 104.3 (CH), 55.9 (CH₃), 55.3 (CH₃), 46.2 (CH₂, 2C), 44.3 (CH), 31.8 (CH₂, 2C).

Methyl 2-Methoxy-5-((4-phenyl-6-(piperidin-4-yl)pyrimidin-2yl)amino)benzoate (Compound 16). The title compound was synthesized following the general procedure 5 previously described using intermediate 40e (51.8 mg, 0.1 mmol) and HCl (4 M in dioxane) (0.25 mL) in 1,4-dioxane dry (1.7 mL). Purification by alumina (elution by a gradient from 100/0 to 95/5 DCM/MeOH) afforded pure compound 16 (17.6 mg, 42% yield). UPLC-MS: $t_R =$ 1.79 min (method 1). MS (ESI) m/z: 419.1 $[M + H]^+$, calcd for $C_{24}H_{27}N_4O_3 [M + H]^+$: 419.2. HRMS (ESI) *m*/*z*: 419.2078 calcd for $C_{24}H_{27}N_4O_3 [M + H]^+$; found *m/z*: 419.2089. ¹H NMR (400 MHz, DMSO- d_6) δ 9.56 (s, 1H), 8.51 (d, J = 2.8 Hz, 1H), 8.18 (m, 2H), 7.85 (dd, J = 9.0, 2.8 Hz, 1H), 7.58–7.48 (m, 3H), 7.29 (s, 1H), 7.13 (d, J = 9.1 Hz, 1H), 3.82 (s, 3H), 3.80 (s, 3H), 3.05 (dt, J = 12.1, 3.2 Hz, 2H), 2.70 (tt, J = 11.8, 3.7 Hz, 1H), 2.59 (td, J = 12.1, 2.5 Hz, 2H), 1.83 (d, J = 12.8 Hz, 2H), 1.69 (qd, J = 12.2, 4.0 Hz, 2H). 1³C NMR (101 MHz, DMSO-*d*₆) δ 175.5 (Cq), 166.0 (Cq), 163.7 (Cq), 160.0 (Cq), 153.0 (Cq), 137.0 (Cq), 133.8 (Cq), 130.7 (CH), 128.8 (CH, 2C), 126.9 (CH, 2C), 123.9 (CH), 121.1 (CH), 119.4 (Cq), 113.2 (CH), 105.3 (CH), 56.1 (CH₃), 51.8 (CH₃), 46.2 (CH₂, 2C), 44.3 (CH), 31.8 (CH₂, 2C).

4-((4-Phenyl-6-(piperidin-4-yl)pyrimidin-2-yl)amino)benzene-1,2-diol Hydrobromide (Compound 17). The title compound was synthesized following the general procedure 6 previously described using intermediate 40d (66 mg, 0.13 mmol) and BBr₃ (1 M in DCM) (0.78 mL, 0.78 mmol) in CHCl₃ dry (2.2 mL). Purification by trituration with EtOAc (1.5 mL) afforded pure compound 8a (16.6 mg, 34% yield). UPLC-MS: $t_R = 2.48 \text{ min (method 4)}$. MS (ESI) *m*/ *z*: 363.0 [M + H]⁺, calcd for C₂₁H₂₃N₄O₂ [M + H]⁺: 363.2. ¹H NMR (400 MHz, DMSO-d₆) δ 9.18 (s, 1H), 8.64 (d, *J* = 10.4 Hz, 1H), 8.37 (q, *J* = 10.5 Hz, 1H), 8.19–8.06 (m, 2H), 7.61–7.48 (m, 3H), 7.22 (d, *J* = 2.5 Hz, 1H), 7.21 (s, 1H), 7.08 (dd, *J* = 8.5, 2.5 Hz, 1H), 6.67 (d, *J* = 8.5 Hz, 1H), 3.41 (d, *J* = 12.9 Hz, 2H), 3.12–3.03 (m, 2H), 2.96 (tt, *J* = 11.4, 3.2 Hz, 1H), 2.13 (dd, *J* = 14.6, 3.7 Hz, 2H), 2.03–1.87 (m, 2H).

2-Hydroxy-5-((4-phenyl-6-(piperidin-4-yl)pyrimidin-2-yl)amino)benzoic Acid Hydrobromide (Compound 18). The title compound was synthesized following the general procedure 6 previously described using intermediate 40e (31.1 mg, 0.06 mmol) and BBr₃ (1 M in DCM) (0.36 mL, 0.36 mmol) in CHCl₃ dry (1.0 mL). Purification by trituration with EtOAc (1.0 mL) afforded pure compound 18 (9.6 mg, 41% yield). UPLC-MS: $t_R = 2.37$ min (method 4). MS (ESI) m/z: 391.0 [M + H]⁺, calcd for C₂₂H₂₃N₄O₃ $[M + H]^+$: 391.4. HRMS (ESI) m/z: 391.1765 calcd for $C_{22}H_{23}N_4O_3$ $[M + H]^+$; found *m/z*: 391.1771. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.87 (s, 1H), 9.57 (s, 1H), 8.74 (d, J = 11.2 Hz, 1H), 8.63 (s), 8.45 (q, J = 10.9 Hz, 1H), 8.22-8.20 (m, 2H), 7.81 (dd, J = 8.9, 2.8 Hz,1H), 7.54–7.50 (m, 3H), 7.33 (s, 1H), 6.94 (d, J = 9.0 Hz, 1H), 3.40 (d, J = 12.4 Hz, 2H), 3.14-3.09 (m, 2H), 2.94 (tt, J = 11.4, 3.3 Hz, 1H), 2.18 (d, J = 13.8 Hz, 2H), 1.95 (qd, 12.4, 2.6 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 172.0 (Cq), 163.9 (Cq), 160.0 (Cq), 157.8 (Cq), 156.0 (Cq), 136.8 (Cq), 132.4 (Cq), 130.8 (CH), 128.8 (CH, 2C), 127.6 (CH), 127.0 (CH, 2C), 120.1 (CH), 116.9 (CH), 112.2 (Cq), 105.2 (CH), 43.0 (CH₂, 2C), 40.2 (CH), 27.0 (CH₂, 2C)

4-Phenyl-6-(piperidin-4-yl)-N-(pyridin-4-yl)pyrimidin-2-amine (Compound 19). The title compound was synthesized following the general procedure 5 previously described using intermediate 40f (70 mg, 0.16 mmol) and HCl (4 M in dioxane) (0.40 mL) in 1,4-dioxane dry (2.7 mL). Purification by alumina (elution by gradient from 100/ 0 to 90/5 DCM/MeOH) afforded pure intermediate 19 (18.5 mg, 35% yield). UPLC-MS: $t_R = 1.88 \text{ min} \pmod{1}$. MS (ESI) m/z: 332.1 $[M + H]^+$, calcd for $C_{20}H_{22}N_5 [M + H]^+$: 332.4. HRMS (ESI) m/z: 332.1870 calcd for C₂₀H₂₂N₅ [M + H]⁺; found m/z: 332.1878. ¹H NMR (400 MHz, DMSO- d_6) δ 10.04 (s, 1H), 8.42–8.32 (m, 2H), 8.23-8.15 (m, 2H), 7.89-7.82 (m, 2H), 7.63-7.50 (m, 3H), 7.44 (s, 1H), 3.06 (dt, J = 11.9, 3.2 Hz, 2H), 2.77 (tt, J = 11.7, 3.7 Hz, 1H), 2.61 (td, J = 12.1, 2.5 Hz, 2H), 1.84 (d, J = 12.4 Hz, 2H), 1.71 (qd, J = 11.0, 2.8 Hz, 2H). ¹³C NMR (101 MHz, DMSO- d_6) δ 175.7 (Cq), 164.2 (Cq), 159.5 (Cq), 149.9 (CH, 2C), 147.4 (Cq), 136.7 (Cq), 130.9 (CH), 128.9 (CH, 2C), 127.1 (CH, 2C), 112.6 (CH, 2C), 107.0 (CH), 46.2 (CH₂, 2C), 44.3 (CH), 31.8 (CH₂, 2C).

4-Phenyl-6-(piperidin-4-yl)-N-(2-(trifluoromethyl)pyridin-4-yl)pyrimidin-2-amine (Compound 20 - ARN25375). The title compound was synthesized following the general procedure 5 previously described using intermediate 40g (70 mg, 0.14 mmol) and HCl (4 M in dioxane) (0.35 mL) in 1,4-dioxane dry (2.3 mL). Purification by alumina (elution by a gradient from 100/0 to 96/4 DCM/MeOH) afforded pure compound 20 (25.1 mg, 45% yield). UPLC-MS: $t_R = 2.05 \text{ min} \pmod{1}$. MS (ESI) m/z: 400.0 [M + H]⁺, calcd for $C_{21}H_{21}F_{3}N_{5}$ [M + H]⁺: 400.4. HRMS (ESI) m/z: 400.1744 calcd for $C_{21}H_{21}F_3N_5$ [M + H]⁺; found m/z: 400.1751. ¹H NMR (400 MHz, DMSO- d_6) δ 10.51 (s, 1H), 8.61 (d, J = 2.1 Hz, 1H), 8.54 (d, I = 5.6 Hz, 1H), 8.20 (dd, I = 6.7, 2.9 Hz, 2H), 7.98 (dd, J = 5.7, 2.1 Hz, 1H), 7.61–7.55 (m, 3H), 7.54 (s, 1H), 3.06 (dt, J = 12.1, 3.2 Hz, 2H), 2.79 (tt, J = 11.8, 3.7 Hz, 1H), 2.61 (td, J = 12.1, 2.5 Hz, 2H), 1.86 (d, J = 11.9 Hz, 2H), 1.71 (qd, J = 12.2, 4.0 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 175.9 (Cq), 164.2 (Cq), 159.3 (Cq), 150.5 (CH), 149.0 (Cq), 147.2 (q², J_{CF} = 32.9 Hz, Cq), 136.5 (Cq), 131.1 (CH), 128.9 (CH, 2C), 127.1 (CH, 2C), 121.9 $(q^1, J_{CF} = 273.4 \text{ Hz}, Cq), 114.7 (CH), 109.1 (q^3, J_{CF} = 3.4 \text{ Hz}, CH),$ 107.8 (CH), 46.2 (CH₂, 2C), 44.2 (CH), 31.8 (CH₂, 2C). ¹⁹F NMR $(376 \text{ MHz}, \text{DMSO-}d_6) \delta - 66.9.$

N-(2-(*Difluoromethoxy*)*pyridin*-4-*yl*)-4-*phenyl*-6-(*piperidin*-4-*yl*)*pyrimidin*-2-*amine* (*Compound* **21**). The title compound was synthesized following the general procedure 5 previously described using intermediate **40h** (70 mg, 0.14 mmol) and HCl (4 M in dioxane) (0.35 mL) in 1,4-dioxane dry (2.3 mL). Purification by alumina (elution by a gradient from 100/0 to 96/4 DCM/MeOH) afforded pure compound **21** (25.1 mg, 45% yield). UPLC-MS: t_R = 2.00 min (method 1). MS (ESI) *m/z*: 398.0 [M + H]⁺, calcd for $C_{21}H_{22}F_2N_5O$ [M + H]⁺: 398.2. HRMS (ESI) *m/z*: 398.1787 calcd for $C_{21}H_{22}F_2N_5O$ [M + H]⁺; found *m/z*: 398.1797. ¹H NMR (400 MHz, DMSO- d_6) δ 10.32 (s, 1H), 8.23–8.14 (m, 2H), 8.07 (d, J = 5.8 Hz, 1H), 7.68 (t, J = 73.4 Hz, 1H), 7.66 (dd, J = 5.8, 1.9 Hz, 1H), 7.64 (d, J = 1.8 Hz, 1H), 7.60–7.53 (m, 3H), 7.50 (s, 1H), 3.06 (dt, J = 12.1, 2.5 Hz, 2H), 2.68 (tt, J = 11.9, 3.6 Hz, 1H), 2.61 (td, J = 12.1, 2.8 Hz, 2H), 1.85 (dd, J = 12.6, 2.4 Hz, 2H), 1.71 (qd, J = 12.2, 3.9 Hz, 2H). ¹³C NMR (101 MHz, DMSO- d_6) δ 175.9 (Cq), 164.3 (Cq), 159.5 (Cq), 159.4 (Cq), 151.1 (Cq), 147.0 (CH), 136.6 (Cq), 131.0 (CH), 129.0 (CH, 2C), 127.1 (CH, 2C), 114.8 (t¹, $J_{CF} = 254.1$ Hz, CH), 110.4 (CH), 107.6 (CH), 97.8 (CH), 46.2 (CH₂, 2C), 44.3 (CH), 31.8 (CH₂, 2C). ¹⁹F NMR (376 MHz, DMSO- d_6) δ – 86.5.

N-(2-Methoxypyridin-4-yl)-4-phenyl-6-(piperidin-4-yl)pyrimidin-2-amine (Compound 22). The title compound was synthesized following the general procedure 5 previously described using intermediate 40i (70 mg, 0.15 mmol) and HCl (4 M in dioxane) (0.37 mL) in 1,4-dioxane dry (2.5 mL). Purification by alumina (elution by a gradient from 100/0 to 95/5 CHCl₃/MeOH) afforded pure compound 22 (21.1 mg, 39% yield). UPLC-MS: $t_R = 1.70$ min (method 1). MS (ESI) m/z: 362.1 [M + H]⁺, calcd for C₂₁H₂₄N₅O $[M + H]^+$: 362.2. HRMS (ESI) m/z: 362.1975 calcd for $C_{21}H_{24}N_5O$ $[M + H]^+$; found *m/z*: 362.1992. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.01 (s, 1H), 8.17 (dd, J = 6.7, 2.8 Hz, 2H), 7.97 (d, J = 5.8 Hz, 1H), 7.58–7.54 (m, 3H), 7.50 (d, J = 1.8 Hz, 1H), 7.44 (s, 1H), 7.37 (dd, J = 5.8, 1.9 Hz, 1H), 3.83 (s, 3H), 3.08 (dt, J = 11.8, 2.8 Hz, 2H), 2.75 (tt, J = 11.8, 3.7 Hz, 1H), 2.60 (td, J = 12.1, 2.5 Hz, 2H), 1.84 (d, J = 12.4 Hz, 2H), 1.70 (qd, J = 12.2, 4.0 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 175.7 (Cq), 164.7 (Cq), 164.1 (Cq), 159.7 (Cq), 149.7 (Cq), 146.7 (CH), 136.8 (Cq), 130.9 (CH), 128.9 (CH, 2C), 127.0 (CH, 2C), 108.0 (CH), 106.9 (CH), 97.0 (CH), 53.0 (CH₃), 46.2 (CH₂, 2C), 44.4 (CH), 31.8 (CH₂, 2C).

N-(6-Methoxypyridin-3-yl)-4-phenyl-6-(piperidin-4-yl)pyrimidin-2-amine (Compound 23). Title compound was synthesized following the general procedure 5 previously described using intermediate 40j (44 mg, 0.09 mmol), HCl (4 M in dioxane) (0.24 mL) in 1,4-dioxane dry (1.5 mL). Purification by alumina (elution by gradient from 100/ 0 to 93/7 DCM/MeOH) afforded pure compound 23 (12.4 mg, 38% yield). UPLC-MS: $t_R = 1.79 \text{ min (method 1)}$. MS (ESI) m/z: 362.0 [M + H]⁺, calcd for $C_{21}H_{24}N_5O$ [M + H]⁺: 362.5. HRMS (ESI) m/z: 362.1975 calcd for $C_{21}H_{24}N_5O [M + H]^+$; found m/z: 362.1985. ¹H NMR (400 MHz, DMSO- d_6) δ 9.48 (s, 1H), 8.68 (d, J = 2.7 Hz, 1H), 8.13 (dd, J = 6.7, 3.0 Hz, 2H), 8.07 (dd, J = 8.9, 2.8 Hz, 1H), 7.57-7.48 (m, 3H), 7.28 (s, 1H), 6.81 (d, J = 8.9 Hz, 1H), 3.83 (s, 3H), 3.07 (d, J = 11.8 Hz, 2H), 2.73 (tt, J = 11.9, 3.6 Hz, 1H), 2.62 (td, J = 12.2, 1.7 Hz, 2H), 1.83 (d, J = 12.0 Hz, 2H), 1.69 (qd, J = 12.3, 4.0 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 174.9 (Cq), 164.6 (Cq), 160.6 (Cq), 159.0 (Cq), 137.5 (CH), 137.4 (Cq), 132.2 (Cq), 131.8 (CH), 131.2 (CH), 129.3 (CH, 2C), 127.4 (CH, 2C), 110.2 (CH), 105.9 (CH), 53.5 (CH₃), 45.1 (CH₂, 2C), 43.1 (CH), 30.1 (CH₂, 2C).

N1,N1-Dimethyl-N3-(6-phenyl-2-(1,2,3,6-tetrahydropyridin-4-yl)pyrimidin-4-yl)benzene-1,3-diamine (Compound 24). The title compound was synthesized following the general procedure 5 previously described using intermediate 43a (60 mg, 0.13 mmol) and HCl (4 M in dioxane) (0.33 mL) in 1,4-dioxane dry (4.4 mL). Purification by alumina (elution by a gradient from 100/0 to 90/10 DCM/MeOH) afforded pure compound 24 (26.5 mg, 56% yield). UPLC-MS: $t_R = 2.06 \text{ min (method 1)}$. MS (ESI) m/z: 372.3 [M + H]⁺, calcd for $C_{23}H_{26}N_5$ [M + H]⁺: 372.2. ¹H NMR (400 MHz, DMSO-d₆) δ 9.42 (s, 1H), 8.07–8.05 (m, 2H), 7.57–7.45 (m, 3H), 7.40 (t, J = 2.3 Hz, 1H), 7.30–7.24 (m, 1H), 7.13 (t, J = 8.1 Hz, 1H), 7.09 (s, 1H), 6.95 (dd, J = 7.8, 1.9 Hz, 1H), 6.40 (dd, J = 8.2, 2.4 Hz, 1H), 3.46 (q, J = 3.2 Hz, 2H), 2.92 (bs, 8H), 2.58 (bs, 2H). ¹³C NMR (101 MHz, DMSO-d₆) δ 163.9 (Cq), 161.1 (Cq), 160.7 (Cq), 150.9 (Cq), 141.0 (Cq), 137.5 (Cq), 135.2 (Cq), 131.8 (CH), 130.1 (CH), 129.0 (CH), 128.8 (CH, 2C), 126.3 (CH, 2C), 107.7 (CH), 106.7 (CH), 103.6 (CH), 99.8 (CH), 45.2 (CH₂), 42.7 (CH₂), 40.2 (CH₃) 2C), 26.1 (CH₂).

1-Methyl-N-(6-phenyl-2-(piperidin-4-yl)pyrimidin-4-yl)-1H-indol-6-amine (Compound 25). The title compound was synthesized following the general procedure 5 previously described using intermediate 44b (50.0 mg, 0.10 mmol), HCl (4 M in dioxane) (0.25 mL) in 1,4-dioxane dry (1.7 mL). Purification by alumina (elution by a gradient from 100/0 to 95/5 DCM/MeOH) afforded pure compound **25** (22.9 mg, 60% yield). UPLC-MS: $t_R = 1.94$ min (method 1). MS (ESI) *m/z*: 384.3 [M + H]⁺, calcd for $C_{24}H_{26}N_5$ [M + H]⁺: 384.2. ¹H NMR (DMSO- d_6) δ 9.58 (bs, 1H), 8.27 (s, 1H), 8.02 (d, *J* = 6.9 Hz, 2H), 7.53–7.47 (m, 4H), 7.24–7.22 (m, 1H), 7.11–7.05 (m, 2H), 6.37 (d, *J* = 3.1 Hz, 1H), 3.77 (s, 3H), 3.07–3.04 (m, 2H), 2.82 (t, *J* = 11.8 Hz, 1H), 2.63 (t, *J* = 12.0 Hz, 2H), 2.00 (d, *J* = 13.0 Hz, 3H), 1.80 (q, *J* = 12.3 Hz, 2H). ¹³C NMR (101 MHz, DMSO- d_6) δ 172.6 (Cq), 161.6 (Cq), 161.1 (Cq), 137.6 (Cq), 136.6 (Cq), 134.5 (Cq), 130.0 (CH), 129.1 (CH), 128.8 (CH, 2C), 126.5 (CH, 2C), 123.7 (Cq), 120.4 (CH), 113.3 (CH), 101.3 (CH), 100.3 (CH), 99.0 (CH), 46.1 (CH_{hb}, 2C), 45.4 (CH), 32.4 (CH₃), 31.9 (CH₂, 2C).

N-(6-Phenyl-2-(piperidin-4-yl)pyrimidin-4-yl)-1H-indol-6-amine (Compound 26). The title compound was synthesized following the general procedure 5 previously described using intermediate 44c (69.0 mg, 0.12 mmol) and HCl (4 M in dioxane) (0.30 mL) in 1,4dioxane dry (2.0 mL). Purification by alumina (elution by a gradient from 100/0 to 95/5 DCM/MeOH) afforded pure compound 26 (14.6 mg, 33% yield). UPLC-MS: $t_R = 1.79$ min (method 1). MS (ESI) m/z: 370.2 [M + H]⁺, calcd for C₂₃H₂₄N₅ [M + H]⁺: 370.2. ¹H NMR (400 MHz, DMSO-d₆) δ 11.04 (s, 1H), 9.43 (s, 1H), 8.02-8.00 (m, 3H), 7.58–7.46 (m, 4H), 7.26 (t, J = 2.7 Hz, 1H), 7.13 (dd, J = 8.5, 1.9 Hz, 1H), 7.03 (s, 1H), 6.37 (t, J = 2.5 Hz, 1H), 3.07 (d, J = 12.0 Hz, 2H), 2.79 (t, J = 11.7 Hz, 1H), 2.68-2.58 (m, 2H), 1.94 (d, J = 12.7 Hz, 2H), 1.78 (qd, J = 12.2, 4.0 Hz, 2H).¹³C NMR (101 MHz, DMSO-*d*₆) δ 172.6 (Cq), 161.6 (Cq), 161.0 (Cq), 137.6 (Cq), 136.1 (Cq), 134.0 (Cq), 129.9 (CH), 128.7 (CH, 2C), 126.4 (CH, 2C), 124.7 (CH), 123.5 (Cq), 119.9 (CH), 113.5 (CH), 103.2 (CH), 101.0 (CH), 98.8 (CH), 46.1 (CH₂, 2C), 45.5 (CH), 31.7 (CH₂, 2C). Compound 26 displayed 93% as determined by UPLC-MS (UV at 215 nm) analysis (see Supporting Information).

tert-Butyl 4-(4-Chloro-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)-3,6-dihydropyridine-1(2H)-carboxylate (Compound **30**). The protocol for the obtainment of intermediate **30** has been modified compared to that previously reported.²³ Compound **30** was synthesized following general procedure 2 using intermediate **28** (1.0 g, 3.247 mmol), (1-(tert-butoxycarbonyl)-1,2,3,6-tetrahydropyridin-4-yl)boronic acid **29** (885 mg, 3.9 mmol), Pd(dppf)Cl₂·DCM (132 mg, 0.162 mmol), K₂CO₃ (aq) 2 M (3.3 mL, 6.49 mmol) and 1,4-dioxane dry (20 mL). After 5 h, a standard workup has been performed. Purification by the gradient (elution by gradient 100/0 to 70/30 cyclohexane/EtOAc) gave the pure product as a white solid (816 mg, 55% yield). UPLC-MS and NMR analysis were consistent with those previously reported.²³

tert-Butyl 4-(4-(Pyrinidin-5-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)-3,6-dihydropyridine-1(2H)-carboxylate (Compound **32a**). Compound **32a** was synthesized following general procedure 2 using compound **30** (200 mg, 0.44 mmol), pyrimidin-5yl boronic acid **31a** (65.3 mg, 0.53 mmol), Pd(dppf)Cl₂·DCM (18.0 mg, 0.022 mmol), K₂CO₃ (2 M) aq (0.44 mL, 0.88 mmol) in 1, 4dioxane dry (3.4 mL). After 1 h, a standard workup has been performed. Purification by silica (elution by gradient 90/10 to 60/40 cyclohexane/EtOAc) afforded the pure product as a white solid (174 mg, 79% yield). UPLC-MS: t_R = 1.87 min (method 2). MS (ESI) *m*/ *z*: 498.0 [M-H]⁻, calcd for C₂₄H₂₃F₃N₇O₂ [M-H]⁻: 498.2. ¹H NMR (400 MHz, DMSO-d₆) δ 10.74 (s, 1H), 9.64 (s, 2H), 9.42 (s, 1H), 8.43 (s, 1H), 8.00 (s, 1H), 7.64 (t, *J* = 8.0 Hz, 1H), 7.57 (s, 1H), 7.45 (d, *J* = 7.8 Hz, 1H), 4.22–4.15 (m, 2H), 3.61–3.54 (m, 2H), 2.68– 2.62 (m, 2H), 1.44 (s, 9H).

tert-Butyl 4-(4-(Isoquinolin-5-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)-3,6-dihydropyridine-1(2H)-carboxylate (Compound **32b**). Compound **32b** was synthesized following general procedure 2 using compound **30** (200 mg, 0.44 mmol), isoquinolin-5yl boronic acid **31b** (91 mg, 0.528 mmol), Pd(dppf)Cl₂-DCM (18.0 mg, 0.022 mmol), K₂CO₃ (2 M) aq (0.44 mL, 0.876 mmol) and 1,4dioxane dry (3.4 mL). After 3 h, a standard workup has been performed. Purification by silica (elution by a gradient from 70/30 to 40/60 cyclohexane/EtOAc) afforded the pure product as a yellowish solid (175 mg, 73% yield). UPLC-MS: $t_R = 2.21$ min (method 2). MS (ESI) *m/z*: 549.0 [M + H]⁺, calcd for $C_{29}H_{28}F_3N_6O_2$ [M + H]⁺: 549.2. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.67 (s, 1H), 9.43 (d, *J* = 1.0 Hz, 1H), 8.90 (s, 1H), 8.65–8.57 (m, 2H), 8.46 (s, 1H), 8.37 (d, *J* = 8.2 Hz, 1H), 8.02 (d, *J* = 7.8 Hz, 1H), 7.89–7.82 (m, 1H), 7.61 (t, *J* = 8.0 Hz, 1H), 7.49–7.39 (m, 2H), 4.23–4.14 (m, 2H), 3.62–3.54 (m, 2H), 2.72–2.65 (m, 2H), 1.44 (s, 9H).

tert-Butyl 4-(4-(1-Methyl-1H-indol-5-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)-3,6-dihydropyridine-1(2H)-carboxylate (Compound **32c**). Compound **32c** was synthesized following general procedure 6 using compound **30** (200 mg, 0.438 mmol), (1-methyl-1H-indol-5-yl)boronic acid 31c (92 mg, 0.526 mmol), Pd(dppf)Cl₂·DCM (17.9 mg, 0.022 mmol), K₂CO₃(aq) 2 M (0.44 mL, 0.876 mmol) in 1,4 dioxane dry (3.4 mL). Purification by silica (elution by a gradient from 100/0 to 70/30 cyclohexane/ EtOAc) afforded the pure product as a yellowish solid (180 mg, 74% yield). UPLC-MS: t_R = 1.50 min (method 3). MS (ESI) *m/z*: 551.0 [M + H]⁺, calcd for C₂₉H₃₀F₃N₆O₂ [M + H]⁺: 551.2. ¹H NMR (400 MHz, CDCl₃) δ 8.89 (d, *J* = 1.6 Hz, 1H), 8.48–8.38 (m, 2H), 7.70 (d, *J* = 7.8 Hz, 1H), 7.58–7.46 (m, 2H), 7.43–7.35 (m, 2H), 7.12 (d, *J* = 3.2 Hz, 1H), 6.63 (d, *J* = 3.3 Hz, 1H), 4.28–4.23 (m, 2H), 3.85 (s, 3H), 3.73–3.60 (m, 2H), 2.86–2.74 (m, 2H), 1.51 (s, 9H).

tert-Butyl 4-(4-(1H-Indazol-5-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)-3,6-dihydropyridine-1(2H)-carboxylate (Compound 32d). Compound 32d was synthesized following general procedure 2 using compound 30 (200 mg, 0.44 mmol), (1H-indazol-6-yl)boronic acid 31d (85.3 mg, 0.528 mmol), Pd(dppf)Cl₂·DCM (18.0 mg, 0.022 mmol), and K₂CO₃ (2 M) aq (0.66 mL, 1.32 mmol) in 1,4-dioxane dry (3.4 mL). After 1.5 h, a standard workup has been performed. Purification by silica (elution by a gradient from 90/10 to 30/70 cyclohexane/EtOAc) afforded pure product that was washed with acetone. Filtration of solid furnished pure product as a white solid (55 mg, 23% yield). UPLC/MS: Rt = 1.97 min (method 2), $[M-H]^{-} = 536.0; [M-H]^{-}$ Calculated for $C_{27}H_{25}F_{3}N_{7}O_{2}$: 536.2. ¹H NMR (400 MHz, DMSO-d₆) δ 13.36 (s, 1H), 10.51 (s, 1H), 8.98 (s, 1H), 8.54-8.43 (m, 2H), 8.27 (s, 1H), 8.02 (d, J = 8.2 Hz, 1H), 7.71–7.59 (m, 2H), 7.51 (s, 1H), 7.42 (d, J = 7.8 Hz, 1H), 4.27–4.15 (m, 2H), 3.63-3.55 (m, 2H), 2.74-2.64 (m, 2H), 1.45 (s, 9H).

tert-Butyl 4-(4-(Benzofuran-3-vl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)-3,6-dihydropyridine-1(2H)-carboxylate (Compound 32e). To a degassed 1,4-dioxane solution (0.13 M, 5.0 mL) of compound 28 (4,6-dichloro-N-(3-(trifluoromethyl)phenyl)-1,3,5-triazin-2-amine) (240 mg, 0.777 mmol), (1-(tert-butoxycarbonvl)-1,2,3,6-tetrahydropyridin-4-vl)boronic acid 29 (211 mg, 0.932 mmol), Pd(dppf)Cl₂·DCM (31.7 mg, 0.0388 mmol) and K₂CO₃(aq) 2 M (0.78 mL, 1.55 mmol) were added. The resulting mixture was sparged with argon for a further 10 min and heated up to 80 °C until consumption of starting material was detected by UPLC (5 h). Then, Pd(dppf)Cl₂·DCM (31.7 mg, 0.0388 mmol) and new boronic acid benzofuran-3-ylboronic acid 31e (151 mg, 0.932 mmol) were added and reaction mixture was heated up to 120 °C until no intermediate was detected by UPLC. Then, H2O and EtOAc were added and separated; aqueous layers were extracted two times with EtOAc and collected organic layers were dried over Na2SO4. The solvent was evaporated and the resulting crude was purified by silica (elution by gradient 100/0 to 70/30 cyclohexane/EtOAc) giving pure product 32e as a white solid (189 mg, 45% yield for two steps). UPLC-MS: t_R = 1.73 min (method 3). MS (ESI) m/z: 538.0 [M + H]⁺, calcd for $C_{28}H_{27}F_{3}N_{5}O_{3}$ [M + H]⁺: 538.4. ¹H NMR (400 MHz, CDCl₃) δ 8.61 (s, 1H), 8.49-8.41 (m, 1H), 8.33 (s, 1H), 7.68 (d, J = 8.1 Hz, 1H), 7.60-7.55 (m, 1H), 7.54-7.46 (m, 2H), 7.43-7.37 (m, 4H), 4.28-4.23 (m, 2H), 3.72-3.64 (m, 2H), 2.77 (s, 2H), 1.52 (s, 9H).

tert-Butyl 4-(4-(1-Methyl-1H-pyrazol-4-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)-3,6-dihydropyridine-1(2H)-carboxylate (Compound **32f**). Compound **32**f was synthesized following general procedure 2 using compound **30** (200 mg, 0.44 mmol), (1-methyl-1H-pyrazol-4-yl)boronic acid (66.5 mg, 0.528 mmol), Pd(dppf)Cl₂.DCM (18.0 mg, 0.022 mmol), K₂CO₃ (2 M) aq (0.44 mL, 0.876 mmol) in 1,4-dioxane dry (3.4 mL). After 1.5 h, a standard workup has been performed. Purification by silica (elution by a gradient from 90/10 to 40/60 cyclohexane/EtOAc) afforded the pure product as a white solid (143 mg, 65% yield). UPLC-MS: $t_R = 0.67$ min (method 3). MS (ESI) *m/z*: 502.0 [M + H]⁺, calcd for $C_{24}H_{27}F_3N_7O_2$ [M + H]⁺: 502.2. ¹H NMR (400 MHz, CDCl₃) δ 8.35 (s, 11H), 8.21 (s, 1H), 8.15 (s, 1H), 7.65 (d, *J* = 8.7 Hz, 1H), 7.51–7.39 (m, 3H), 7.36 (d, *J* = 7.5 Hz, 1H), 4.25–4.17 (m, 2H), 3.98 (s, 3H), 3.67–3.59 (m, 2H), 2.77–2.63 (m, 2H), 1.50 (s, 9H).

tert-Butyl 4-(4-(1-Methyl-1H-imidazol-5-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)-3,6-dihydropyridine-1(2H)-carboxylate (Compound **32g**). Compound **32g** was synthesized following general procedure 2 using compound **30** (100 mg, 0.22 mmol), (1-methyl-1H-imidazol-5-yl)boronic acid **31g** (54.8 mg, 0.26 mmol), Pd(dppf)Cl₂·DCM (9.0 mg, 0.011 mmol), K₂CO₃ (2 M) aq (0.22 mL, 0.44 mmol) in 1,4-dioxane dry (2.0 mL). After 1.5 h, the addition of H₂O promotes the precipitation of a solid that has been filtered and washed with EtOAc, furnishing the desired product as a white solid (65 mg, 60% yield). UPLC-MS: t_R = 1.61 min (method 2). MS (ESI) *m/z*: 502.0 [M + H]⁺, calcd for C₂₄H₂₇F₃Nr₂O₂ [M + H]⁺: 502.2. ¹H NMR (400 MHz, DMSO-d₆) δ 10.43 (s, 1H), 8.01–7.84 (m, 4H), 7.60 (t, *J* = 8.0 Hz, 1H), 7.44–7.35 (m, 2H), 4.21–4.10 (m, 2H), 4.06 (s, 3H), 3.61–3.52 (m, 2H), 2.64–2.58 (m, 2H), 1.44 (s, 9H).

tert-Butyl 4-(4-(1-(tert-Butoxycarbonyl)-1H-pyrrol-2-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)-3,6-dihydropyridine-1(2H)-carboxylate (Compound **32h**). Compound **32g** was synthesized following general procedure 2 using compound **30** (200 mg, 0.44 mmol (1-(*tert*-butoxycarbonyl)-1H-pyrrol-2-yl)boronic acid **31h** (111.4 mg, 0.53 mmol), Pd(dppf)Cl₂-DCM (17.9 mg, 0.022 mmol), K₂CO₃ (2 M) aq (0.44 mL, 0.88 mmol) in 1,4-dioxane dry (3.5 mL). After 1.5 h, the addition of H₂O promotes the precipitation of a solid that has been filtered and washed with AcOEt, furnishing the desired product as a white solid (170 mg, 65% yield). UPLC-MS: t_R = 2.56 min (method 2). MS (ESI) *m/z*: 587.0 [M + H]⁺, calcd for C₂₉H₃₄F₃N₆O₄ [M + H]⁺: 587.2. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.19 (s, 1H), 7.71 (s, 1H), 7.56–7.31 (m, 5H), 6.96 (bs, 1H), 6.28 (t, *J* = 3.3 Hz, 1H), 4.18 (bs, 2H), 3.61 (t, *J* = 5.0 Hz, 2H), 2.69 (s, 2H), 1.50 (s, 9H), 1.47 (s, 9H).

tert-Butyl 4-(4-(Thiophen-3-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)-3,6-dihydropyridine-1(2H)-carboxylate (Compound 32i). Compound 32i was synthesized following general procedure 2 using compound 30 (120 mg, 0.263 mmol), thiophen-3ylboronic acid 31i (40 mg, 0.316 mmol), Pd(dppf)Cl₂·DCM (10.6 mg, 0.013 mmol), and K₂CO₃ (2 M) aq (0.26 mL, 0.526 mmol) in 1,4-dioxane dry (2.0 mL). After 1.5 h, a standard workup has been performed. Purification by silica (elution by a gradient from 95/5 to 80/20 cyclohexane/EtOAc) afforded not a pure product that was washed with acetone. Filtration of solid furnished pure product as a white solid (99 mg, 70% yield). UPLC-MS: $t_R = 1.15$ min (method 3). MS (ESI) m/z: 504.0 [M + H]⁺, calcd for $C_{24}H_{25}F_3N_5O_2S$ [M + H]⁺: 504.2. ¹H NMR (400 MHz, DMSO- d_6) δ 10.50 (s, 1H), 8.54 (d, J = 3.1 Hz, 1H, 8.46 (s, 1H), 8.01 (d, J = 8.3 Hz, 1H), 7.84 (d, J =5.0 Hz, 1H), 7.72 (t, J = 5.0, 3.1 Hz, 1H), 7.60 (t, J = 8.0 Hz, 1H), 7.45 (s, 1H), 7.40 (d, J = 7.8 Hz, 1H), 4.19-4.14 (m, 2H), 3.60-3.52 (m, 2H), 2.66–2.62 (m, 2H), 1.44 (s, 9H).

tert-Butyl 4-(4-(Pyrimidin-5-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)piperidine-1-carboxylate (Compound **33a**). Compound **33a** was synthesized following general procedure 4-Method A using compound **32a** (153 mg, 0.306 mmol), ammonium formate (113.6 mg, 1.8 mmol), Pd(OH)₂/C (30.6 mg) and dry EtOH (5.0 mL) and THF (2.5 mL). After 1 h, a standard workup has been performed. Purification by silica (elution by gradient 80/20 to 50/50 cyclohexane/EtOAc) afforded the pure product as a white solid (51 mg, 32% yield). UPLC-MS: t_R = 1.75 min (method 2). MS (ESI) *m*/*z*: 502.1 [M + H]⁺, calcd for C₂₄H₂₇F₃N₇O₂ [M + H]⁺: 502.2. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.75 (s, 1H), 9.58 (s, 2H), 9.41 (s, 1H), 8.50–8.30 (m, 1H), 8.08–7.88 (m, 1H), 7.69–7.58 (m, 1H), 7.45 (d, *J* = 7.8 Hz, 1H), 4.03 (d, *J* = 13.0 Hz, 2H), 3.05–2.84 (m, 3H), 2.11–1.95 (m, 2H), 1.80–1.63 (m, 2H), 1.42 (s, 9H).

tert-Butyl 4-(4-(Isoquinolin-5-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)piperidine-1-carboxylate (Compound **33b**). Compound **33b** was synthesized following general procedure 4-Method A using compound **32b** (147 mg, 0.26 mmol), ammonium formate (98.4 mg, 1.56 mmol), Pd(OH)₂/C (30 mg) and dry EtOH (4.2 mL) and THF (2.1 mL). After 3.5 h, a standard workup has been performed. Purification by silica (elution by a gradient from 80/20 to 50/50 cyclohexane/EtOAc) afforded the pure product as a white foaming solid (63 mg, 43% yield). UPLC-MS: $t_R = 2.01 \text{ min}$ (method 2). MS (ESI) *m/z*: 549.0 [M-H]⁻, calcd for C₂₉H₂₈F₃N₆O₂ [M-H]⁻: 549.6. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.68 (s, 1H), 9.43 (s, 1H), 8.88 (s, 1H), 8.70–8.28 (m, 4H), 7.98 (d, *J* = 7.8 Hz, 1H), 7.86 (t, *J* = 7.8 Hz, 1H), 7.60 (t, *J* = 8.1 Hz, 1H), 7.42 (d, *J* = 7.8 Hz, 1H), 4.12–3.98 (m, 2H), 3.06–2.87 (m, 3H), 2.13–2.03 (m, 2H), 1.83–1.67 (m, 2H), 1.42 (s, 9H).

tert-Butyl 4-(4-(1-Methyl-1H-indol-5-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)piperidine-1-carboxylate (Compound **33c**). Compound **33c** was synthesized following general procedure 4-Method A using compound **32c** (155 mg, 0.282 mmol), ammonium formate (106.8 mg, 1.69 mmol), Pd(OH)₂/C (31 mg) and dry EtOH (4.0 mL) and THF (3.0 mL). Purification by silica (elution by gradient 95/5 to 70/30 cyclohexane/EtOAc) giving pure product **33c** as a white foaming solid (105.0 mg, 67% yield). UPLC-MS: Rt = 1.42 min (method 3), [M + H]+ = 553.0; [M + H]+ Calculated for C₂₉H₃₂F₃N₆O₂: 553.2. ¹H NMR (400 MHz, DMSOd₆) δ 10.43 (s, 1H), 8.75 (d, J = 1.6 Hz, 1H), 8.61 (s, 1H), 8.28 (dd, J = 8.7, 1.7 Hz, 1H), 7.94 (d, J = 8.0 Hz, 1H), 7.65–7.54 (m, 2H), 7.47–7.37 (m, 2H), 6.60 (d, J = 3.1 Hz, 1H), 4.12–3.98 (m, 2H), 3.85 (s, 3H), 3.05–2.81 (m, 3H), 2.10–1.97 (m, 2H), 1.81–1.66 (m, 2H), 1.43 (s, 9H).

tert-Butyl 4-(4-(1H-Indazol-5-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)piperidine-1-carboxylate (Compound **33d**). Compound **33d** was synthesized following general procedure 4-Method A using compound **32d** (110 mg, 0.205 mmol), ammonium formate (77.6 mg, 1.23 mmol), Pd(OH)₂/C (22 mg), dry MeOH (3.4 mL), and THF (1.7 mL). After 2 h, a standard workup has been performed. Purification by silica (elution by a gradient from 95/5 to 50/50 cyclohexane/EtOAc) afforded the pure product as a white solid (28 mg, 25% yield). UPLC-MS: t_R = 1.80 min (method 2). MS (ESI) *m/z*: 540.1 [M + H]⁺, calcd for C₂₇H₂₉F₃N₇O₂ [M + H]⁺: 540.2. ¹H NMR (400 MHz, DMSO-d₆) δ 13.36 (s, 1H), 10.51 (s, 1H), 8.93 (s, 1H), 8.51 (s, 1H), 8.43 (d, *J* = 9.0 Hz, 1H), 8.27 (s, 1H), 7.99 (s, 1H), 7.71–7.56 (m, 2H), 7.42 (d, *J* = 7.8 Hz, 1H), 4.12–4.00 (m, 2H), 3.04–2.85 (m, 2H), 2.12–1.93 (m, 3H), 1.80–1.66 (m, 2H), 1.43 (s, 9H).

tert-Butyl 4-(4-(Benzofuran-3-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)piperidine-1-carboxylate (Compound **33e**). Compound **33e** was synthesized following general procedure 4-Method A using compound **32e** (170 mg, 0.316 mmol), ammonium formate (100 mg, 1.90 mmol), Pd(OH)₂/C (34 mg), and MeOH dry (3.2 mL). After 1.5 h, a standard workup was performed and the crude product was purified by silica (elution by gradient 90/10 to 85/ 15 cyclohexane/EtOAc) giving the pure product as a white foaming solid (108 mg, 63% yield). UPLC-MS: t_R = 1.63 min (method 3). MS (ESI) *m/z*: 540.0 [M + H]⁺, calcd for C₂₈H₂₉F₃N₅O₃ [M + H]⁺: 540.2. ¹H NMR (400 MHz, CDCl₃) δ 8.60 (s, 1H), 8.47–8.41 (m, 1H), 8.28 (s, 1H), 7.73–7.67 (m, 1H), 7.61–7.54 (m, 1H), 7.51 (t, *J* = 7.9 Hz, 1H), 7.43–7.33 (m, 4H), 4.24 (s, 2H), 3.01–2.83 (m, 3H), 2.20–2.04 (m, 2H), 1.89 (dd, *J* = 12.1, 4.3 Hz, 2H), 1.50 (s, 9H).

tert-Butyl 4-(4-(1-Methyl-1H-pyrazol-4-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)piperidine-1-carboxylate (Compound **33f**). Compound **33f** was synthesized following general procedure 4-Method A using compound **32f** (132 mg, 0.263 mmol), ammonium formate (99.6 mg, 1.58 mmol), Pd(OH)₂/C (26.4 mg) in EtOH dry (4.4 mL) and THF dry (2.2 mL). After 1 h, a standard workup has been performed. Purification by silica (elution by a gradient from 80/20 to 50/50 cyclohexane/EtOAc) afforded the pure product as a white solid (78 mg, 59% yield). UPLC-MS: t_R = 1.68 min (method 3). MS (ESI) *m/z*: 504.0 [M + H]⁺, calcd for C₂₄H₂₉F₃N₇O₂ [M + H]⁺: 504.2. ¹H NMR (400 MHz, DMSO-d₆) δ 10.39 (s, 1H), 8.48–8.37 (m, 2H), 8.04 (s, 1H), 8.04 (d, *J* = 0.7 Hz, 1H), 7.58 (t, *J* = 8.0 Hz, 1H), 7.37 (d, *J* = 7.4 Hz, 1H), 4.06–3.98 (m, 2H), 3.93 (s, 3H), 3.02–2.75 (m, 3H), 2.06–1.88 (m, 2H), 1.74–1.59 (m, 2H), 1.42 (s, 9H).

tert-Butyl 4-(4-(1-Methyl-1H-imidazol-5-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)piperidine-1-carboxylate (Compound **33g**). Compound **33g** was synthesized following general procedure 4-Method A using compound **32g** (90 mg, 0.180 mmol), ammonium formate (68.1 mg, 1.08 mmol), Pd(OH)₂/C (18 mg) in dry MeOH (3.0 mL) and THF (1.5 mL). After 5 h, a standard workup was performed giving pure product without any purification as a white solid (83 mg, 92% yield). UPLC-MS: t_R = 1.47 min (method 2). MS (ESI) *m*/*z*: 504.0 [M + H]⁺, calcd for C₂₄H₂₉F₃N₇O₂ [M + H]⁺: 504.2. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.41 (s, 1H), 8.39 (s, 1H), 7.93–7.89 (m, 2H), 7.86 (d, *J* = 1.1 Hz, 1H), 7.59 (t, *J* = 8.0 Hz, 1H), 7.40 (d, *J* = 7.7 Hz, 1H), 4.09–3.95 (m, 5H), 3.02–2.78 (m, 3H), 2.04–1.93 (m, 2H), 1.74–1.59 (m, 2H), 1.41 (s, 9H).

tert-Butyl 4-(4-(1-(tert-Butoxycarbonyl)-1H-pyrrol-2-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)piperidine-1-carboxylate (Compound 33h). Compound 33h was synthesized following general procedure 4-Method B using compound 32h (100 mg, 0.17 mmol), Et₃SiH (0.27 mL, 1.7 mmol), Pd/C (20 mg) and dry EtOH (3.4 mL). After 3 h, further Et₃SiH (0.27 mL, 1.7 mmol) was added at 0 $^\circ\text{C}\textsc{,}$ and the reaction mixture was left to stir overnight. After the addition of a further 5 equiv of reducing agent, a standard workup has been performed. Purification by silica (elution by gradient 95/5 to 80/20 cyclohexane/EtOAc) afforded the pure product as a white solid (60 mg, 60% yield). UPLC-MS: $t_R = 1.18$ min (method 3). MS (ESI) m/z: 589.0 [M + H]⁺, calcd for C₂₉H₃₆F₃N₆O₄ [M + H]⁺: 588.3. ¹H NMR (400 MHz, DMSO- d_6) δ 10.51 (s, 1H), 8.38 (s, 1H), 7.93 (d, J = 8.4 Hz, 1H), 7.57 (t, J = 8.0 Hz, 1H), 7.46 (t, J = 3.1, 1.7 Hz, 1H), 7.39 (d, J = 7.7 Hz, 1H), 6.90–6.85 (m, 1H), 6.35 (t, J = 3.3 Hz, 1H), 4.07-3.93 (m, 2H), 3.01-2.77 (m, 3H), 2.00-1.87 (m, 2H), 1.72-1.57 (m, 2H), 1.41 (s, 18H).

tert-Butyl 4-(4-(1-(tert-Butoxycarbonyl)pyrrolidin-2-yl)-6-((3-(trifluoromethyl)phenyl)amino)-1,3,5-triazin-2-yl)piperidine-1-carboxylate (Compound **33h**'). Compound **33h**' was synthesized following general procedure 4-Method A using compound **32h** (145 mg, 0.247 mmol), ammonium formate (93.4 mg, 1.48 mmol), Pd(OH)₂/C (29 mg), and dry EtOH (6.0 mL). After 1 h, a standard workup has been performed. Purification by silica (elution by gradient 90/10 to 60/40 cyclohexane/EtOAc) afforded the pure product as a white foaming solid (127 mg, 86% yield). UPLC-MS: t_R = 2.25 min (method 2). MS (ESI) *m/z*: 593.0 [M + H]⁺, calcd for C₂₉H₄₀F₃N₆O₄ [M + H]⁺: 592.3. ¹H NMR (400 MHz, DMSO-d₆) δ 10.47 (s, 1H), 8.36 (s, 1H), 7.91 (d, *J* = 8.4 Hz, 1H), 7.59–7.50 (m, 1H), 7.42–7.34 (m, 1H), 4.64–4.50 (m, 1H), 4.07–3.90 (m, 2H), 3.62–3.38 (m, 2H), 2.99–2.76 (m, 3H), 2.42–2.26 (m, 1H), 2.06–1.76 (m, 5H), 1.72–1.52 (m, 2H), 1.40 (s, 9H), 1.10 (s, 9H).

tert-Butyl 4-(2-Chloro-6-phenylpyrimidin-4-yl)-3,6-dihydropyridine-1(2H)-carboxylate (Compound 37). K_2CO_3 (2 M) aq (2 mL), phenyl boronic acid 35 (256 mg, 2.1 mmol), and $\textrm{PdCl}_2(\textrm{dppf})\cdot$ DCM (163 mg, 0.2 mmol) were added to a solution of trichloropyrimidine 34 (366 mg, 2 mmol) in 1,4-dioxane dry (4 mL) under argon. The reaction mixture stirred at 65 °C for 2 h until complete consumption of starting material 34 (UPLC-MS of not isolated intermediate 2,4-dichloro-6-phenylpyrimidine: $t_R = 1.50$ min (method 2). MS (ESI) m/z: 224.9 [M + H]⁺, calcd for C₁₀H₇Cl₂N₂ [M + H]⁺: 226.1). After that, boronic ester **36** (618 mg, 2 mmol) and a second addition of PdCl₂(dppf)·DCM (82 mg, 0.1 mmol) was added and reaction mixture stirred at 90 $^\circ C$ for other 2h at 90 $^\circ C$ under argon until complete conversion of 2,4-dichloro-6-phenylpyrimidine intermediate. Then water (3 mL) was added and an aqueous layer was extracted with EtOAc (5 mL \times 2). Collected organic layers were dried over Na2SO4, filtered, and concentrated under a vacuum. The desired product 37 was purified by silica eluting by a gradient from 100% cyclohexane to 85/15 cyclohexane/EtOAc, obtaining pure compound 3 (410 mg, 55%). UPLC-MS: $t_R = 2.20$ min (method 2). MS (ESI) m/z: 372.1 [M + H]⁺, calcd for $C_{20}H_{23}ClN_{3}O_{2}$ [M + H]⁺: 372.9. ¹H NMR (400 MHz, CDCl₃) δ 8.12-8.02 (m, 2H), 7.59 (s, 1H), 7.56-7.47 (m, 3H), 7.05 (m, 1H), 4.20 (q, J = 3.0 Hz, 2H), 3.66 (t, J = 5.7 Hz, 2H), 2.64 (m, 2H), 1.49 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 167.7 (Cq), 167.2 (Cq),

161.7 (Cq), 154.8 (Cq), 135.8 (Cq), 133.2 (Cq), 131.8 (CH), 130.7 (CH), 129.2 (CH, 2C), 127.5 (CH, 2C), 109.7 (CH), 80.2 (Cq), 44.1 (CH₂), 40.0 (CH₂), 28.6 (CH₂), 25.4 (CH₃, 3C).

tert-Butyl 4-(2-((3-(Dimethylamino)phenyl)amino)-6-phenylpyrimidin-4-yl)-3,6-dihydropyridine-1(2H)-carboxylate (Compound **39a**). The title compound was synthesized following the general procedure 3 previously described using intermediate 37 (100 mg, 0.27 mmol), aniline **38a** (44 mg, 0.32 mmol), Pd(OAc)₂ (3.0 mg, 0.013 mmol), (\pm)-BINAP (8.4 mg, 0.013 mmol), Cs₂CO₃ (132.0 mg, 0.40 mmol) in 1,4 dioxane (1.8 mL). Purification by silica (elution by gradient from 95/5 to 80/20 cyclohexane/EtOAc) afforded pure intermediate **39a** (89 mg, 70% yield). UPLC-MS: t_R = 2.42 min (method 2). MS (ESI) *m/z*: 472.4 [M + H]⁺, calcd for C₂₈H₃₄N₅O₂ [M + H]⁺: 472.3. ¹H NMR (400 MHz, CDCl₃) δ 8.12–8.02 (m, 2H), 7.55–7.46 (m, 3H), 7.44 (s, 1H), 7.31 (bs, 1H), 7.22 (t, *J* = 7.9 Hz, 1H), 7.19 (s, 1H), 6.96 (s, 1H), 6.93 (s, 1H), 6.48 (d, *J* = 8.2 Hz, 1H), 4.18 (q, *J* = 3.0 Hz, 2H), 3.66 (t, *J* = 5.6 Hz, 2H), 2.69 (m, 2H), 1.51 (s, 9H).

tert-Butyl 4-(6-Phenyl-2-((3-(trifluoromethyl)phenyl)amino)pyrimidin-4-yl)- dihydropyridine-1(2H)-carboxylate (Compound **39b**). Title compound was synthesized following the general procedure 3 previously described using intermediate $_{37}$ (100 mg, 0.27 mmol), aniline **38b** (0.04 mL, 0.32 mmol) in 1,4 dioxane (1.8 mL), Pd(OAc)₂ (3.0 mg, 0.013 mmol), (\pm)-BINAP (8.4 mg, 0.013 mmol), and Cs₂CO₃ (132.0 mg, 0.40 mmol). Purification by silica (elution by gradient from 95/5 to 80/20 cyclohexane/EtOAc) afforded intermediate **39b** (83 mg, 62% combined yield of the two isomers: an isomerization of the double bond occurred during the reaction). UPLC-MS: t_R = 1.53 min (method 3). MS (ESI) *m/z*: 497.2 [M + H]⁺, calcd for C₂₇H₂₈F₃N₄O₂ [M + H]⁺: 497.2. The mixture was used as such in the next step for the obtainment of intermediate **40b**.

tert-Butyl 4-(6-Phenyl-2-((3-(trifluoromethoxy)phenyl)amino)pyrimidin-4-yl)- dihydropyridine-1(2H)-carboxylate (Compound **39c**). The title compound was synthesized following the general procedure 3 previously described using intermediate 37 (100 mg, 0.27 mmol), aniline **38c** (0.04 mL, 0.32 mmol) in 1,4 dioxane (1.8 mL), Pd(OAc)₂ (2.7 mg, 0.012 mmol), (±)-BINAP (7.4 mg, 0.012 mmol), and Cs₂CO₃ (131.0 mg, 0.40 mmol). Purification by silica (elution by gradient from 95/5 to 85/15 cyclohexane/EtOAc) afforded intermediate **39c** (109 mg, 79% combined yield of the two isomers: an isomerization of the double bond occurred during the reaction). UPLC-MS: there are two main peaks related to the isomers α and β of **39c** with double C–C bond shifted t_R = 1.53 and 1.68 min (method 3). MS (ESI) m/z: 513.1 [M + H]⁺, calcd for C₂₇H₂₈F₃N₄O₃ [M + H]⁺: 513.2. The mixture was used as such in the next step for the obtainment of intermediate **40c**.

tert-Butyl 4-(2-((3,4-Dimethoxyphenyl)amino)-6-phenylpyrimidin-4-yl)-dihydropyridine-1(2H)-carboxylate (Compound **39d**). The title compound was synthesized following the general procedure 3 previously described using intermediate **37** (200 mg, 0.54 mmol), aniline **38d** (99.0 mg, 0.64 mmol) in 1,4 dioxane (3.6 mL), Pd(OAc)₂ (6.0 mg, 0.025 mmol), Xantphos (15.5 mg, 0.025 mmol), Cs₂CO₃ (263.0 mg, 0.81 mmol). Purification by silica (elution by a gradient from 80/20 to 75/25 cyclohexane/EtOAc) afforded intermediate **39d** (197.9 mg, 75% combined yield of the two isomers: an isomerization of the double bond occurred during the reaction). UPLC-MS: t_R = 1.97 min (method 2). MS (ESI) *m/z*: 489.1 [M + H]⁺, calcd for C₂₈H₃₃N₄O₄ [M + H]⁺: 489.2. The mixture was used as such in the next step for the obtainment of intermediate **40d**.

tert-Butyl 4-(2-((4-Methoxy-3-(methoxycarbonyl)phenyl)amino)-6-phenylpyrimidin-4-yl)- dihydropyridine-1(2H)-carboxylate (Compound **39e**). The title compound was synthesized following the general procedure 3 previously described using intermediate **37** (200 mg, 0.54 mmol), aniline **38e** (117.0 mg, 0.64 mmol) in 1,4 dioxane (3.6 mL), Pd(OAc)₂ (6.0 mg, 0.025 mmol), Xantphos (15.5 mg, 0.025 mmol), and Cs₂CO₃ (263.0 mg, 0.81 mmol). Purification by silica (elution by a gradient from 100/0 to 75/25 cyclohexane/ EtOAc) afforded intermediate **39e** (236.4 mg, 85% combined yield of the two isomers: an isomerization of the double bond occurred during the reaction). UPLC-MS: t_R = 1.92 min (method 2). MS (ESI) *m/z*: 517.1 $[M + H]^+$, calcd for $C_{29}H_{33}N_4O_5 [M + H]^+$: 517.2. The mixture was used as such in the next step for the obtainment of intermediate **40e**.

tert-Butyl 4-(6-Phenyl-2-(pyridin-4-ylamino)pyrimidin-4-yl)-dihydropyridine-1(2H)-carboxylate (Compound **39f**). The title compound was synthesized following the general procedure 3 previously described using intermediate **37** (100 mg, 0.27 mmol), aniline **38f** (30.4 mg, 0.32 mmol) in 1,4 dioxane (1.8 mL), Pd(OAc)₂ (2.7 mg, 0.012 mmol), Xantphos (6.9 mg, 0.012 mmol), and Cs₂CO₃ (131.0 mg, 0.40 mmol). Purification by silica (elution by a gradient from 100/0 to 95/5 DCM/EtOH) afforded intermediate **39f** (97.8 mg, 85% combined yield of the two isomers: an isomerization of the double bond occurred during the reaction). UPLC-MS: t_R = 1.68 min (method 2). MS (ESI) *m/z*: 430.1 [M + H]⁺, calcd for C₂₅H₂₈N₅O₂ [M + H]⁺: 430.2. The mixture was used as such in the next step for the obtainment of intermediate **40f**.

tert-Butyl 4-(6-Phenyl-2-((2-(trifluoromethyl)pyridin-4-yl)amino)pyrimidin-4-yl)- dihydropyridine-1(2H)-carboxylate (Compound **39g**). The title compound was synthesized following the general procedure 3 previously described using intermediate 37 (100 mg, 0.27 mmol), aniline **38g** (52.3 mg, 0.32 mmol) in 1,4 dioxane (1.8 mL), Pd(OAc)₂ (2.7 mg, 0.012 mmol), (\pm)-BINAP (7.4 mg, 0.012 mmol), and Cs₂CO₃ (131.0 mg, 0.40 mmol). Purification by silica (elution by a gradient from 100/0 to 75/25 cyclohexane/ EtOAc) afforded pure intermediate **39g** (85.6 mg, 80% combined yield of the two isomers: an isomerization of the double bond occurred during the reaction). UPLC-MS: t_R = 2.30 min (method 2). MS (ESI) *m/z*: 498.1 [M + H]⁺, calcd for C₂₆H₂₇F₃N₅O₂ [M + H]⁺: 498.2. The mixture was used as such in the next step for the obtainment of intermediate **40g**.

tert-Butyl 4-(2-((2-(Difluoromethoxy)pyridin-4-yl)amino)-6-phenylpyrimidin-4-yl)- dihydropyridine-1(2H)-carboxylate (Compound **39h**). The title compound was synthesized following the general procedure 3 previously described using intermediate **37** (100 mg, 0.27 mmol), aniline **38h** (51.7 mg, 0.32 mmol) in 1,4 dioxane (1.8 mL), Pd(OAc)₂ (2.7 mg, 0.012 mmol), (\pm)-BINAP (7.4 mg, 0.012 mmol), Cs₂CO₃ (131.0 mg, 0.40 mmol). Purification by silica (elution by a gradient from 100/0 to 80/20 cyclohexane/EtOAc) afforded intermediate **39h** (85.6 mg, 80% combined yield of the two isomers: an isomerization of the double bond occurred during the reaction). UPLC-MS: t_R = 2.30 min (method 2). MS (ESI) *m/z*: 496.1 [M + H]⁺, calcd for C₂₆H₂₈F₂N₅O₃ [M + H]⁺: 496.2. The mixture was used as such in the next step for the obtainment of intermediate **40h**.

tert-Butyl 4-(2-((2-Methoxypyridin-4-yl)amino)-6-phenylpyrimidin-4-yl)-3,6-dihydropyridine-1(2H)-carboxylate (Compound **39**i). Title compound was synthesized following the general procedure 3 previously described using intermediate **37** (100 mg, 0.27 mmol), aniline **38i** (40.0 mg, 0.32 mmol) in 1,4 dioxane (1.8 mL), Pd(OAc)2 (2.7 mg, 0.012 mmol), Xantphos (6.9 mg, 0.012 mmol), and Cs₂CO₃ (131.0 mg, 0.40 mmol). Purification by silica (elution by a gradient from 80/20 to 75/25 cyclohexane/EtOAc) afforded intermediate **39i** (74.1 mg, 60% combined yield of the two isomers: an isomerization of the double bond occurred during the reaction). UPLC-MS: there are two main peaks related to the isomers $t_R = 1.95$ min and 2.05 (method 2). MS (ESI) m/z: 460.1 [M + H]⁺, calcd for C₂₆H₃₀N₅O₃ [M + H]⁺: 460.2. The mixture was used as such in the next step for the obtainment of intermediate **40i**.

tert-Butyl 4-(2-((6-Methoxypyridin-3-yl)amino)-6-phenylpyrimidin-4-yl)-dihydropyridine-1(2H)-carboxylate (Compound **39**). The title compound was synthesized following the general procedure 3 previously described using intermediate 37 (100 mg, 0.27 mmol), aniline **38**j (40.0 mg, 0.32 mmol) in 1,4-dioxane (1.8 mL), Pd(OAc)₂ (2.7 mg, 0.012 mmol), Xantphos (6.9 mg, 0.012 mmol), and Cs₂CO₃ (131.0 mg, 0.40 mmol). Purification by silica (elution by a gradient from 80/20 to 75/25 cyclohexane/EtOAc) afforded intermediate **39**j (82.3 mg, 66% combined yield of the two isomers: an isomerization of the double bond occurred during the reaction). UPLC-MS: there are two main peaks related to the isomers $t_R = 2.01$ min and 2.05 (method 2). MS (ESI) m/z: 460.1 [M + H]⁺, calcd for C₂₆H₃₀N₅O₃ [M + H]⁺: 460.2. tert-Butyl 4-(2-((3-(Dimethylamino)phenyl)amino)-6-phenylpyrimidin-4-yl)piperidine-1-carboxylate (Compound **40a**). The title compound was synthesized following the general procedure 4-Method A previously described using intermediate **39a** (60 mg, 0.127 mmol), Pd(OH)₂/C (12 mg), and NH₄CO₂H (48 mg, 0.76 mmol) in MeOH (3.2 mL). Purification by silica (elution by a gradient from 100/0 to 80/20 cyclohexane/EtOAc) afforded pure intermediate **40a** (59 mg, 98% yield). UPLC-MS: t_R = 2.30 min (method 2). MS (ESI) *m/z*: 474.3 [M + H]⁺, calcd for C₂₈H₃₆N₅O₂ [M + H]⁺: 474.3. ¹H NMR (400 MHz, CDCl₃) δ 8.12–8.02 (m, 2H), 7.52 (s, 1H), 7.51–7.43 (m, 3H), 7.20 (t, *J* = 8.1 Hz, 1H), 7.02 (s, 1H), 6.91 (d, *J* = 8.0 Hz, 1H), 6.48 (d, *J* = 8.3 Hz, 1H), 4.27 (bs, 2H), 2.93–2.70 (m, 3H), 1.97 (d, *J* = 13.0 Hz, 2H), 1.90–1.74 (qd, *J* = 12.8, 4.2 Hz, 2H), 1.49 (s, 9H).

tert-Butyl 4-(6-Phenyl-2-((3-(trifluoromethyl)phenyl)amino)pyrimidin-4-yl)piperidine-1-carboxylate (Compound 40b). The title compound was synthesized following the general procedure 4-Method A previously described using intermediate 39b as an isomeric mixture (70 mg, 0.14 mmol), Pd(OH)₂/C (14 mg), and NH₄CO₂H (53 mg, 0.84 mmol) in MeOH (3.5 mL). Purification by silica (elution by a gradient from 100/0 to 85/15 cyclohexane/EtOAc) afforded pure intermediate 40b (59.3 mg, 85% yield). UPLC-MS: t_R = 1.49 min (method 3). MS (ESI) *m/z*: 499.2 [M + H]⁺, calcd for C₂₇H₃₀F₃N₄O₂ [M + H]⁺: 499.6. ¹H NMR (400 MHz, CDCl₃) δ 8.39 (t, *J* = 2.0 Hz, 1H), 8.10–8.03 (m, 2H), 7.68 (dd, *J* = 8.2, 2.0 Hz, 1H), 7.54–7.47 (m, 3H), 7.44 (t, *J* = 8.0 Hz, 1H), 7.30–7.26 (m, 1H), 7.11 (s, 1H), 4.30 (bs, 2H), 2.90–2.84 (m, 2H), 2.80 (tt, *J* = 11.9, 3.8 Hz, 1H), 1.98 (d, *J* = 13.1 Hz, 2H), 1.80 (qd, *J* = 12.3, 4.4 Hz,2H), 1.49 (s, 9H).

tert-Butyl 4-(6-Phenyl-2-((3-(trifluoromethoxy)phenyl)amino)pyrimidin-4-yl)piperidine-1-carboxylate (Compound 40c). The title compound was synthesized following the general procedure 4-Method A previously described using intermediate 39c as an isomeric mixture (110 mg, 0.21 mmol), Pd(OH)₂/C (22 mg), NH₄CO₂H (79 mg, 1.26 mmol) in MeOH (5.2 mL). Purification by silica (elution by a gradient from 90/10 to 85/15 cyclohexane/EtOAc) afforded pure intermediate 40c (79.5 mg, 72% yield). UPLC-MS: t_R = 1.50 min (method 3). MS (ESI) *m/z*: 515.1 [M + H]⁺, calcd for C₂₇H₃₀F₃N₄O₃ [M + H]⁺: 515.6 ¹H NMR (400 MHz, CDCl₃) δ 8.11–8.01 (m, 3H), 7.58–7.44 (m, 3H), 7.37 (dt, *J* = 8.2, 1.5 Hz, 1H), 7.33 (t, *J* = 8.0 Hz, 1H), 7.09 (s, 1H), 6.86 (d, *J* = 7.9 Hz, 1H), 4.29 (bs, 2H), 2.91–2.83 (m, 2H), (tt, *J* = 11.9, 3.5 Hz, 1H), 1.98 (d, *J* = 13.1 Hz, 2H), 1.79 (qd, *J* = 12.5, 4.3 Hz, 2H), 1.49 (s, 9H).

tert-Butyl 4-(2-((3,4-Dimethoxyphenyl)amino)-6-phenylpyrimidin-4-yl)piperidine-1-carboxylate (Compound 40d). The title compound was synthesized following the general procedure 4-Method A previously described using intermediate 39d as an isomeric mixture (197 mg, 0.40 mmol), Pd(OH)₂/C (39.4 mg), NH₄CO₂H (151.4 mg, 2.5 mmol) in MeOH (10.0 mL). Purification by silica (elution by a gradient from 100/0 to 75/25 cyclohexane/EtOAc) afforded pure intermediate 40d (174.6 mg, 89% yield). UPLC-MS: t_R = 1.91 min (method 2). MS (ESI) *m/z*: 491.1 [M + H]⁺, calcd for C₂₈H₃₅N₄O₄ [M + H]⁺: 491.3. ¹H NMR (400 MHz, CDCl₃) δ 8.10–8.00 (m, 2H), 7.85 (s, 1H), 7.64 (d, *J* = 2.5 Hz, 1H), 7.53–7.45 (m, 3H), 7.04 (dd, *J* = 8.5, 2.4 Hz, 1H), 7.01 (s, 1H), 6.86 (d, *J* = 8.7 Hz, 1H), 4.27 (bs, 2H), 3.92 (s, 3H), 3.88 (s, 3H), 2.89–2.81 (m, 2H), 2.79 (tt, *J* = 11.8, 3.9 Hz, 1H), 1.99 (d, *J* = 13.1 Hz, 2H), 1.80 (qd, *J* = 12.5, 4.4 Hz, 2H), 1.48 (s, 9H).

tert-Butyl 4-(2-((4-Methoxy-3-(methoxycarbonyl)phenyl)amino)-6-phenylpyrimidin-4-yl)piperidine-1-carboxylate (Compound **40e**). The title compound was synthesized following the general procedure 4-Method A previously described using intermediate **39e** as an isomeric mixture (270 mg, 0.52 mmol), Pd(OH)₂/C (54.0 mg), NH₄CO₂H (196.8 mg, 3.1 mmol) in MeOH (13.0 mL). Purification by silica (elution by a gradient from 100/0 to 75/25 DCM/EtOAc) afforded pure intermediate **6**j (371.3 mg, 73% yield). UPLC-MS: t_R = 1.88 min (method 2). MS (ESI) *m/z*: 519.1 [M + H]⁺, calcd for C₂₉H₃₅N₄O₅ [M + H]⁺: 519.2. ¹H NMR (400 MHz, CDCl₃) δ 8.36 (d, *J* = 2.9 Hz, 1H), 8.10–8.05 (m, 2H), 7.75 (dd, *J* = 8.9, 2.9 Hz, 1H), 7.53–7.48 (m, 3H), 7.05 (s, 1H), 7.00 (d, *J* = 9.0 Hz, 1H), 4.28 (s, 2H), 3.93 (s, 3H), 3.92 (s, 3H), 2.84 (m, 3H), 2.00 (d, J = 13.0 Hz, 2H), 1.80 (qd, J = 12.5, 4.3 Hz, 2H), 1.49 (s, 9H).

tert-Butyl 4-(6-Phenyl-2-(pyridin-4-ylamino)pyrimidin-4-yl)piperidine-1-carboxylate (Compound 40f). The title compound was synthesized following the general procedure 4-Method B previously described using intermediate 39f as an isomeric mixture (100 mg, 0.23 mmol), Et₃SiH (0.37 mL, 2.33 mmol), Pd/C (20 mg) in EtOH (3.8 mL). Purification by silica (elution by a gradient from 90/10 to 85/15 cyclohexane/EtOAc) afforded pure intermediate 40f (79.3 mg, 79% yield). UPLC-MS: $t_R = 1.34$ min (method 2). MS (ESI) *m*/*z*: 432.1 [M + H]⁺, calcd for $C_{25}H_{30}N_5O_2$ [M + H]⁺: 432.5. ¹H NMR (400 MHz, CDCl₃) δ 8.43 (d, *J* = 6.4 Hz, 2H), 8.07–8.00 (m, 2H), 7.96 (d, *J* = 6.1 Hz, 2H), 7.51–7–50 (m, 4H), 7.22 (s, 1H), 4.26 (bs, 2H), 2.96–2.76 (m, 3H), 1.97 (d, *J* = 12.6 Hz, 2H), 1.79 (qd, *J* = 12.7, 4.4 Hz, 2H), 1.49 (s, 9H).

tert-Butyl 4-(6-Phenyl-2-((2-(trifluoromethyl)pyridin-4-yl)amino)pyrimidin-4-yl)piperidine-1-carboxylate (Compound **40g**). The title compound was synthesized following the general procedure 4-Method A previously described using intermediate **39g** as an isomeric mixture (100 mg, 0.20 mmol), $Pd(OH)_2/C$ (20 mg), NH_4CO_2H (79 mg, 1.26 mmol) in MeOH (5.2 mL). Purification by silica (elution by a gradient from 100/0 to 85/15 cyclohexane/ EtOAc) afforded pure intermediate **40g** (64.9 mg, 65% yield). UPLC-MS: $t_R = 2.25$ min (method 2). MS (ESI) *m/z*: 500.1 [M + H]⁺, calcd for $C_{26}H_{29}F_3N_5O_2$ [M + H]⁺: 500.5. ¹H NMR (400 MHz, CDCl₃) δ 8.57 (d, J = 5.6 Hz, 1H), 8.40 (d, J = 2.2 Hz, 1H), 8.12–8.01 (m, 2H), 7.69 (dd, J = 5.6, 2.2 Hz, 2H), 7.57–7.48 (m, 3H), 7.21 (s, 1H), 4.30 (bs, 2H), 3.00–2.85 (m, 2H), 2.82 (tt, J = 11.9, 3.6 Hz, 1H), 1.98 (d, J = 12.9 Hz, 2H), 1.81 (qd, J = 12.5, 4.3 Hz, 3H), 1.50 (s, 9H).

tert-Butyl 4-(2-((2-(Difluoromethoxy)pyridin-4-yl)amino)-6-phenylpyrimidin-4-yl)piperidine-1-carboxylate (Compound **40h**). The title compound was synthesized following the general procedure 2-Method A previously described using intermediate **39h** as an isomeric mixture (75 mg, 0.15 mmol), Pd(OH)₂/C (15 mg), NH₄CO₂H (76.3 mg, 1.21 mmol) in MeOH (5.2 mL). Purification by silica (elution by a gradient from 100/0 to 80/20 cyclohexane/EtOAc) afforded pure intermediate **40h** (63.4 mg, 85% yield). UPLC-MS: t_R = 2.21 min (method 2). MS (ESI) *m/z*: 498.1 [M + H]⁺, calcd for C₂₆H₃₀F₂N₅O₃ [M + H]⁺: 498.6 ¹H NMR (400 MHz, CDCl3) δ 8.11–8.02 (m, 3H), 7.61–7.52 (m, 4H), 7.48 (t, *J* = 73.3 Hz, 1H), 7.33 (dd, *J* = 5.8, 1.9 Hz, 1H), 7.20 (s, 1H), 4.30 (bs, 2H), 2.93–2.87 (m, 3H), 2.02 (d, *J* = 12.6 Hz, 2H), 1.81 (qd, *J* = 12.4, 4.4 Hz, 2H), 1.49 (s, 9H).

tert-Butyl 4-(2-((2-Methoxypyridin-4-yl)amino)-6-phenylpyrimidin-4-yl)piperidine-1-carboxylate (Compound **40**i). The title compound was synthesized following the general procedure 4-Method A previously described using intermediate **39i** as an isomeric mixture (80 mg, 0.17 mmol), Pd(OH)₂/C (16 mg), NH₄CO₂H (64.4 mg, 1.02 mmol) in MeOH (5.5 mL). Purification by silica (elution by a gradient from 85/15 to 75/25 cyclohexane/EtOAc) afforded pure intermediate **40i** (64.2 mg, 80% yield). UPLC-MS: t_R = 1.95 min (method 2). MS (ESI) *m*/*z*: 462.1 [M + H]⁺, calcd for C₂₆H₃₂N₅O₃ [M + H]⁺: 462.2. ¹H NMR (400 MHz, CDCl₃) δ 8.04 (m, 3H), 7.64 (s, 1H), 7.55–7.49 (m, 3H), 7.47 (bs, 1H), 7.19 (dd, *J* = 6.0, 2.0 Hz, 1H), 7.15 (s, 1H), 4.29 (bs, 2H), 3.99 (s, 3H), 2.97–2.83 (m, 2H), 2.88 (tt, *J* = 11.8, 3.6 Hz, 1H), 1.97 (d, *J* = 12.5 Hz, 2H), 1.81 (qd, *J* = 12.5, 4.3 Hz, 2H), 1.49 (s, 9H).

tert-Butyl 4-(2-((6-Methoxypyridin-3-yl)amino)-6-phenylpyrimidin-4-yl)piperidine-1-carboxylate (Compound **40***j*). The title compound was synthesized following the general procedure 4-Method A previously described using intermediate **39***j* as an isomeric mixture (80 mg, 0.17 mmol), Pd(OH)₂/C (16 mg), NH₄CO₂H (64.4 mg, 1.02 mmol) in MeOH (5.5 mL). Purification by silica (elution by a gradient from 85/15 to 75/25 cyclohexane/EtOAc) afforded pure intermediate **40***j* (44.9 mg, 56% yield). UPLC-MS: t_R = 1.91 min (method 2). MS (ESI) *m*/*z*: 462.1 [M + H]⁺, calcd for C₂₆H₃₂N₅O₃ [M + H]⁺: 462.6. ¹H NMR (400 MHz, CDCl₃) δ 8.46 (d, *J* = 2.7 Hz, 1H), 8.06–8.00 (m, 2H), 7.98 (dd, *J* = 8.9, 2.8 Hz, 1H), 7.52–7.46 (m, 4H), 7.03 (s, 1H), 6.78 (d, *J* = 9.0 Hz, 1H), 4.27 (s, 2H), 3.95 (s, 3H), 2.93–2.79 (m, 2H), 2.76 (tt, *J* = 11.9, 3.6 Hz, 1H), 1.97 (d, *J* = 13.0 Hz, 2H), 1.78 (qd, *J* = 12.5, 4.2 Hz, 2H), 1.49 (s, 9H).

N-(2-Chloro-6-phenylpyrimidin-4-yl)-1-methyl-1H-indol-6amine (Compound **42b**). The title compound was synthesized following the general procedure 1 previously described using compound **41** (184 mg, 0.82 mmol), aniline **38k** (120 mg, 0.82 mmol), LiHMDS (1.0 M in THF, 2.05 mL) in THF dry (5.5 mL). Purification by silica (elution by a gradient from 100/0 to 95/5 cyclohexane/EtOAc) afforded pure compound **42b** (150.0 mg, 55%). UPLC-MS: t_R = 2.52 min (method 1). MS (ESI) *m/z*: 335.2 [M + H]⁺, calcd for C₁₉H₁₆ClN₄ [M + H]⁺: 335.1. ¹H NMR (400 MHz, CDCl₃) δ 7.91–7.83 (m, 2H), 7.67 (d, *J* = 8.3 Hz, 1H), 7.47–7.36 (m, 3H), 7.34 (d, *J* = 11.3 Hz, 2H), 7.13 (d, *J* = 3.1 Hz, 1H), 7.04 (dd, *J* = 8.3, 1.9 Hz, 1H), 6.88 (s, 1H), 6.54 (d, *J* = 3.1 Hz, 1H), 3.80 (s, 3H).

tert-Butyl 6-((2-Chloro-6-phenylpyrimidin-4-yl)amino)-1H-indole-1-carboxylate (Compound 42c). The title compound was synthesized following the general procedure 1 previously described using compound 41 (100 mg, 0.82 mmol), aniline 38l (112.0 mg, 0.50 mmol), LiHMDS (1.0 M in THF, 2.05 mL) in THF dry (3.3 mL). Purification by silica (elution by a gradient from 100/0 to 95/5 cyclohexane/EtOAc) afforded pure compound 42c (202.0 mg, 96%). UPLC-MS: t_R = 2.17 min (method 2). MS (ESI) *m/z*: 419.3 [M + H]⁺, calcd for C₂₃H₂₀ClN₄O₂ [M + H]⁺: 419.1. ¹H NMR (400 MHz, CDCl₃) δ 8.20 (s, 1H), 7.94–7.88 (m, 2H), 7.64 (d, *J* = 3.7 Hz, 1H), 7.60 (d, *J* = 8.3 Hz, 1H), 7.48–7.37 (m, 3H), 7.27 (s, 1H), 7.18 (dd, *J* = 8.3, 2.0 Hz, 1H), 7.04 (s, 1H), 6.60 (dd, *J* = 3.7, 0.8 Hz, 1H), 1.64 (s, 9H).

tert-Butyl 4-(4-((1-Methyl-1H-indol-6-yl)amino)-6-phenylpyrimidin-2-yl)-3,6-dihydropyridine-1(2H)-carboxylate (Compound **43b**). The title compound was synthesized following the general procedure 2 previously described using intermediate **41** (140.0 mg, 0.42 mmol) boronic ester **36** (155.8 mg, 0.50 mmol), Pd(Cl₂)(dppf)·DCM (34.0 mg, 0.04 mmol), K₂CO₃ (2 M) aq (0.46 mL) in 1,4-dioxane dry (2.8 mL). Purification by silica (elution by a gradient from 100/0 to 75/25 cyclohexane/EtOAc) afforded pure intermediate **43b** (175.0 mg, 86%). UPLC-MS: t_R = 2.31 min (method 2). MS (ESI) *m/z*: 480.6 [M-H]⁻, calcd for C₂₉H₃₂N₅O₂ [M-H]⁻: 480.2. ¹H NMR (400 MHz, CDCl₃) δ 8.02–7.93 (m, 2H), 7.63 (d, *J* = 8.3 Hz, 1H), 7.53 (s, 1H), 7.46–7.40 (m, 3H), 7.23 (bs, 1H), 7.09 (d, *J* = 3.1 Hz, 1H), 7.04 (dd, *J* = 8.3, 1.9 Hz, 1H), 6.99 (s, 1H), 6.90 (s, 1H), 6.51 (dd, *J* = 3.1, 0.8 Hz, 1H), 4.20 (bs, 2H), 3.78 (s, 3H), 3.66 (bs, 2H), 2.84 (bs, 2H).

tert-Butyl 6-((2-(1-(tert-Butoxycarbonyl)-1,2,3,6-tetrahydropyridin-4-yl)-6-phenylpyrimidin-4-yl)amino)-1H-indole-1-carboxylate (Compound 43c). The title compound was synthesized following the general procedure 2 previously described using intermediate 42c (190.0 mg, 0.45 mmol) boronic ester 36 (209.4 mg, 0.67 mmol), Pd(Cl₂)(dppf)·DCM (36.8 mg, 0.05 mmol), K₂CO₃ (2 M) aq (0.68 mL) in 1,4 dioxane dry (5 mL). Purification by silica (elution by a gradient from 100/0 to 85/15 cyclohexane/EtOAc) afforded pure intermediate 43c (217.0 mg, 85%). UPLC-MS: t_R = 2.31 min (method 2). MS (ESI) *m*/*z*: 566.4 [M-H]⁻, calcd for C₃₃H₃₆N₅O₄ [M-H]⁻: 566.3. ¹H NMR (400 MHz, CDCl₃) δ 8.50 (s, 1H), 8.06–8.0 (m, 2H), 7.58 (d, *J* = 3.7 Hz, 1H), 7.55 (d, *J* = 8.3 Hz, 1H), 7.49–7.40 (m, 3H), 7.34 (s, 1H), 7.18 (dd, *J* = 8.3, 2.0 Hz, 1H), 7.02 (s, 1H), 6.90 (s, 1H), 6.57 (d, *J* = 3.7 Hz, 1H), 4.22 (bs, 2H), 3.67 (t, *J* = 5.2 Hz, 2H), 2.85 (bs, 2H), 1.64 (s, 9H), 1.50 (s, 9H).

tert-Butyl 4-(4-((1-Methyl-1H-indol-6-yl)amino)-6-phenylpyrimidin-2-yl)piperidine-1-carboxylate (Compound **44b**). The title compound was synthesized following the general procedure 4-Method A previously described using intermediate **43b** (85 mg, 0.18 mmol), Pd(OH)₂/C (17.0 mg), NH₄CO₂H (68.1 mg, 1.1 mmol) in MeOH (4.5 mL). Purification by silica (elution by a gradient from 100/0 to 70/30 cyclohexane/EtOAc) afforded pure intermediate **44b** (60.0 mg, 69% yield). UPLC-MS: t_R = 2.23 min (method 2). MS (ESI) m/ z: 484.5 [M + H]⁺, calcd for C₂₉H₃₄N₅O₂ [M + H]⁺: 484.6. ¹H NMR (400 MHz, CDCl₃) δ 7.97–7.87 (m, 2H), 7.63 (d, *J* = 8.4 Hz, 1H), 7.50 (s, 1H), 7.45–7.36 (m, 3H), 7.09 (dd, *J* = 3.1, 1.3 Hz, 1H), 7.03 (dd, *J* = 8.3, 1.9 Hz, 1H), 6.94 (bs, 1H), 6.87 (d, *J* = 1.2 Hz, 1H), 6.51 (dd, *J* = 3.1, 0.9 Hz, 1H), 4.23 (bs, 2H), 3.78 (s, 3H), 2.96–2.90 (m, 3H), 2.08 (bs, 2H), 1.93 (qd, *J* = 12.2, 3.9 Hz, 2H), 1.49 (s, 9H).

tert-Butyl 6-((2-(1-(tert-Butoxycarbonyl)piperidin-4-yl)-6-phenylpyrimidin-4-yl)amino)-1H-indole-1-carboxylate (Compound 44c). The title compound was synthesized following the general procedure 4-Method A previously described using intermediate 43c (220.0 mg, 0.39 mmol), Pd(OH)₂/C (44.0 mg), NH₄CO₂H (147.7 mg, 2.3 mmol) in MeOH (5.0 mL). Purification by silica (elution by a gradient from 100/0 to 75/25 cyclohexane/EtOAc) afforded pure intermediate 44c (69.2 mg, 32% yield). The yield has been affected by the formation of the over-reduced indoline derivative, which was separated by desired 44c by purification by silica. UPLC-MS: t_{R} = 1.56 min (method 3). MS (ESI) m/z: 570.4 [M + H]⁺, calcd for $C_{33}H_{40}N_5O_4$ [M + H]⁺: 570.3. ¹H NMR (400 MHz, CDCl₃) δ 8.28 (s, 1H), 8.01–7.93 (m, 2H), 7.59 (d, J = 3.7 Hz, 1H), 7.57 (d, J = 8.3 Hz, 1H), 7.46–7.39 (m, 3H), 7.22 (dd, J = 8.3, 2.0 Hz, 1H), 7.09 (bs), 7.03 (s, 1H), 6.57 (d, J = 3.7 Hz, 1H), 4.23 (bs, 2H), 3.03–2.82 (m, 3H), 2.09 (d, J = 13.3 Hz, 2H), 1.92 (qd, J = 12.5, 3.9 Hz, 2H), 1.63 (s, 9H), 1.49 (s, 9H).

In Vitro and In Vivo Experiments. *Cell Viability*. Cell viability assay was performed as previously described.²² Cancer cell lines were cultured according to the method described in Jahid et al., 2022.²²

In Vivo Efficacy. An in vivo efficacy test was performed as described.^{22,23} All animal experiments were approved by the UC Irvine Institutional Animal Care and Use Committee (IACUC) (AUP-23-116). NOD.Cg-Prkdcscid IL2rgtm1Wjl/SzJ (NSG) mice were purchased from The Jackson Laboratories (stock number 005557); both male and female mice were used in vehicle and inhibitor-treated groups (N = 6). After tumors reached the initial size range of 150-250 mm³, mice were administrated with 10 mg/kg ARN25499, or vehicle via tail vein injection for 2 weeks daily. Tumors were measured every other day with a caliper and tumor volume was calculated ((length \times width \times width)/2). At the end of 2 weeks, tumors were extracted and measured to determine volume (length × width × height). GraphPad Prism9 software was used to generate line and scatterplot graphs and determine significance using two-way ANOVA and unpaired two-tailed t-test. An equal number of males and females were used in the experiments.

In Vitro Treatment and Western Blot Analysis. *Immunoblotting.* Melanoma cells (WM3248) were cultured as described²² and were seeded at 1.5 × 10⁶ cells per 60.8 cm² plate. Sixteen hrs. later, WM3248 cells were treated with 10 μ M of <u>ARN22089</u>, <u>ARN25375</u>, or <u>ARN25499</u> for 6 h. and lysed in Lysis buffer (Cytoskeleton, Inc.) containing a protease inhibitors cocktail. Lysates were then subjected to SDS-PAGE and transferred to PVDF membranes. The expression or phosphorylation of proteins was detected by Western blotting using the following primary Abs: pERK at 1:500, ERK at 1:6000, pS6 at 1:6000, S6 at 1:1000 for 16 h and appropriate HRP-secondary antibody for 2 h (all from Cell Signaling). ImageJ was used to perform densitometry. The experiments were performed in triplicate and representative results are shown.

BIOPHYSICAL METHODS

His-CDC42 Production and Purification. His-CDC42 wild-type (amino acids Ile4-Pro182) was expressed from the pET28a+ vector in *E. coli* BL21 (DE3) cells and purified, GppNHp or GDP-bound, as previously described.^{22,23}

Binding Check of Hit Derivatives by Microscale Thermophoresis. MicroScale Thermophoresis experiments were performed according to the NanoTemper technologies protocols in a Monolith NT.115 Pico (Pico Red/Nano Blue - NanoTemper Technologies). His-CDC42 was RED – NHS labeled and used at a concentration of 10 nM. The compound concentration was 50 μ M throughout all the experiments. DMSO was also constant across samples at 0.5% v/v. Solutions were prepared in 100 mM Trizma base (Sigma) pH 7.5, 40 mM NaCl, 0.05% v/v Tween 20, and incubated 5 min before loading on Premium Capillaries and analysis. Binding was detected at 24 °C, MST power high, and 20% LED power. The MST traces were recorded as follows: 3 s MST power off, 20 s MST power on, and 1 s MST power off. The difference in normalized fluorescence (ΔF_{norm} $[\%o] = F_{hot}/F_{cold})$ between a protein:compound sample and a protein only sample at 1.5–2.5 s and 14.0–15.0 s is calculated and plotted through MO.Affinity analysis v2.3 (NanoTemper Technologies) and GraphPad Prism 8.0.0 (GraphPad Software, San Diego, California USA). Signal-to-noise ratio and response amplitude were used to evaluate the quality of the binding data according to NanoPedia instructions (NanoTemper Technologies). Only a signal-to-noise ratio of more than 5 and a response amplitude of more than 1.5 were considered acceptable, while a signal-to-noise of more than 12 was considered excellent.

NMR Confirmation of Target Engagement. The NMR experiments were performed as reported in Brindani et al. 2023²³ using a 5 mm CryoProbe QCI ¹H/¹⁹F-¹³C/¹⁵N-D quadruple resonance, a shielded z-gradient coil, and an automatic sample changer SampleJet 600 MHz NMR system with temperature control. For all samples, a 1D ¹H NMR experiment was recorded, and the water suppression was obtained using the standard NOESY (nuclear Overhauser effect spectroscopy) presat Bruker pulse sequence, with 64 k data points, a spectral width (sw) of 30 ppm, 64 scans, acquisition time (aq) of 1.835 s, a relaxation delay (d1) of 4 s and a mixing time of 10 ms. The WaterLOGSY experiments were achieved with a 7.5 ms long 180° Gaussian-shaped pulse, aq 0.852 s, mixing time of 1.7 s, relaxation delay of 2 s, and 1024 scans. ¹⁹F T₂ filter experiments were recorded using a CPMG spin-echo scheme with a 35 ms time interval between the 180 pulses and different total lengths (140 and 240 ms, respectively), 32 scans, sw 40 ppm, aq 0.72s, and d1 5 s. The data were multiplied with an exponential window function with 1 Hz line broadening prior to Fourier transformation for ¹H 1D, and ¹⁹F T₂ filter experiments and 2 Hz line broadening for the WaterLOGSY experiments. The solubility of the compounds was evaluated both in PBS and Tris buffer by ¹H 1D experiments and aggregation by WaterLOGSY, testing the compounds in the binding assays buffer at the theoretical concentrations of 10, 25, and 50 in the presence of 200 μ M 4-trifluoromethyl benzoic acid (internal reference).

For the binding experiments, all the compounds were tested at 50 μ M in 20 mM Trizma base (Sigma) pH 7.5, 40 mM NaCl, 5 mM MgCl₂, 5 μ M EDTA, 10% D₂O (for the lock signal), and in the presence of 2 μ M GppNHp or GDP and 2 μ M His-CDC42 (loaded with GDP) or His-CDC42 (loaded with GppNHp). The total amount of DMSO- d_6 in all samples was 1%. All fluorine chemical shifts were referred to the CFCl₃signal in water.

Computational Methods. Molecular Docking. To predict and evaluate the interaction between CDC42 and ARN25499/ARN25375 compounds, we first performed molecular docking. As the receptor structure, we employed the CDC42 protein in complex with the CRIB domain of PAK6 (PDB code 20DB, resolution of 2.4 Å) where we formerly identified a previously unappreciated allosteric pocket at the protein-protein interface.²² The structure was refined by using the Protein Preparation Wizard³⁹ workflow implemented in Maestro Release 2021-3. Specifically, hydrogen atoms were added, and charges and protonation states were assigned titrating the protein at physiologic pH. The steric clashes were relieved by performing a small number of minimization steps until the RMSD of the non-hydrogen atoms reached 0.30 Å. The formerly identified pocket was used to center the grid. Precisely, the cubic grid box of $26 \times 26 \times 26$ Å³ was centered on the previously identified hit compound of the CDC42ligand complex.²² The compound was prepared using LigPrep software implemented in Maestro. First, we added hydrogens and generated ionization states at pH 7.4 \pm 0.5. Then, we generated tautomers and all stereochemical isomers. Finally, we used Glide⁴⁰⁻ to perform the molecular docking, using Extra Precision and retaining a maximum of 20 poses. The best docking pose was chosen for further investigation through classical MD simulation.

Molecular Dynamics Simulations. Molecular dynamics (MD) simulations were performed on the protein–ligand complexes obtained from our docking calculations. The GTP substrate as well as the catalytic Mg²⁺ ion in the active site of the proteins were considered. The systems were hydrated with a 14 Å layer of TIP3P water molecules⁴³ from the protein center. The coordinates of the water molecules at the catalytic center were taken from the PDB X-ray

structure 20DB. Sodium ions were added to neutralize the charge of the systems. The final models are enclosed in a box of $\sim 89 \times 89 \times 89$ Å³, containing \sim 18,800 water molecules, resulting in \sim 59,000 atoms for each system. The AMBER-ff14SB force field⁴⁴ was used for the parametrization of the protein. The parameters for the ligands ARN25499 and ARN25375 were determined via Hartree-Fock calculation, with 6-31G* basis set, convergence criterium SCF = Tight after structure optimization (DFT B3LYP functional; 6-31G* basis set). The Merz-Singh-Kollman scheme⁴⁵ was used for the atomic charge assignment. The GTP and the Mg2+ were parametrized according to Meagher et al. and Allner et al. respectively.^{46,47} Joung-Chetham parameters were used for monovalent ions.⁴⁸ All MD simulations were performed with Amber⁴⁹ and all the systems were objects of the following equilibration protocol. To relax the water molecule and the ions, we performed an energy minimization imposing a harmonic potential of 300 kcal/mol Å² on the backbone, the GTP, and the docked compounds. Then, two consecutive MD simulations in NVT and NPT ensembles (1 and 10 ns, respectively) were carried out, imposing the previous positional restraints. To relax the solute, two additional energy minimization steps were performed imposing positional restraints of 20 kcal/mol Å² and without any restraints, respectively. Such minimized systems were heated up to 303 K with four consecutive MD simulations in NVT (~0.1 ns, 100 K) and NPT ensembles (~0.1 ns, 100 K; ~0.1 ns, 200 K; ~0.2 ns, 303 K), imposing the previous positional restraints of 20 kcal/mol $Å^2$. We used the Andersen-like temperature-coupling scheme⁵⁰ while pressure control was achieved with Monte Carlo barostat at a reference pressure of 1 atm. Long-range electrostatics were treated with the particle mesh Ewald method. We performed an additional MD simulation (~1.5 ns) in the NPT ensemble at 303 K without any restraint to relax the system at such a temperature. Finally, multiple replicas of 500 ns were performed in the NPT ensemble for each system with an integration time step of 2 fs.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.jmedchem.4c00855.

¹H, ¹³C, ¹⁹F NMR spectral data and chromatography analysis of key compounds; MST and NMR analyses for binding evaluation; procedures for aqueous kinetic and thermodynamic solubility, in vitro metabolic stability, and in vitro plasmatic stability; and pharmacokinetic studies (PDF)

Molecular formula strings (CSV)

AUTHOR INFORMATION

Corresponding Authors

- Anand K. Ganesan Department of Dermatology, University of California, Irvine, California 92697, United States; Email: aganesan@uci.edu
- Marco De Vivo Molecular Modeling and Drug Discovery Lab, Istituto Italiano di Tecnologia, 16163 Genova, Italy; orcid.org/0000-0003-4022-5661; Email: marco.devivo@iit.it

Authors

- Nicoletta Brindani Molecular Modeling and Drug Discovery Lab, Istituto Italiano di Tecnologia, 16163 Genova, Italy
- Linh M. Vuong Department of Dermatology, University of California, Irvine, California 92697, United States
- Maria Antonietta La Serra Molecular Modeling and Drug Discovery Lab, Istituto Italiano di Tecnologia, 16163 Genova, Italy; o orcid.org/0000-0001-8732-9965
- Noel Salvador Department of Dermatology, University of California, Irvine, California 92697, United States

Andrea Menichetti – Molecular Modeling and Drug Discovery Lab, Istituto Italiano di Tecnologia, 16163 Genova, Italy

- Isabella Maria Acquistapace Molecular Modeling and Drug Discovery Lab, Istituto Italiano di Tecnologia, 16163 Genova, Italy; © orcid.org/0000-0002-5820-7683
- Jose Antonio Ortega Molecular Modeling and Drug Discovery Lab, Istituto Italiano di Tecnologia, 16163 Genova, Italy
- Marina Veronesi Structural Biophysics Facility, Istituto Italiano di Tecnologia, 16163 Genova, Italy
- Sine Mandrup Bertozzi Analytical Chemistry Facility, Istituto Italiano di Tecnologia, 16163 Genova, Italy
- Maria Summa Translational Pharmacology Facility, Istituto Italiano di Tecnologia, 16163 Genova, Italy
- Stefania Girotto Structural Biophysics Facility, Istituto Italiano di Tecnologia, 16163 Genova, Italy;
 [©] orcid.org/ 0000-0002-0339-6675
- Rosalia Bertorelli Translational Pharmacology Facility, Istituto Italiano di Tecnologia, 16163 Genova, Italy
- Andrea Armirotti Analytical Chemistry Facility, Istituto Italiano di Tecnologia, 16163 Genova, Italy; © orcid.org/ 0000-0002-3766-8755

Complete contact information is available at: https://pubs.acs.org/10.1021/acs.jmedchem.4c00855

Author Contributions

[#]N.B. and L.M.V. contributed equally to this work. **Notes**

The authors declare the following competing financial interest(s): A.K.G., M.D.V., N.B., L.M.V., A.M. are co-inventors on patents related to this work.

ACKNOWLEDGMENTS

We thank the Italian Association for Cancer Research (AIRC) for financial support (IG 23679). We thank S. Venzano for the preparation of the plates with the compounds' solution for the screening.

ABBREVIATIONS

DCM, dichloromethane; DMF, *N*,*N*-dimethylformamide; DMSO, dimethyl sulfoxide; EtOAc, ethyl acetate; MST, microscale thermophoresis; THF, tetrahydrofuran

REFERENCES

(1) Bray, K.; Gillette, M.; Young, J.; Loughran, E.; Hwang, M.; Sears, J. C.; Vargo-Gogola, T. Cdc42 overexpression induces hyperbranching in the developing mammary gland by enhancing cell migration. *Breast Cancer Res.* **2013**, *15* (5), R91.

(2) Murphy, N. P.; Binti Ahmad Mokhtar, A. M.; Mott, H. R.; Owen, D. Molecular subversion of Cdc42 signalling in cancer. *Biochem. Soc. Trans.* 2021, 49 (3), 1425–1442.

(3) Lv, J.; Song, Y. Could cell division cycle protein 42 be a target for lung cancer treatment? *Transl Cancer Res.* **2019**, *8* (1), 312–318.

(4) Lee, S.; Craig, B. T.; Romain, C. V.; Qiao, J.; Chung, D. H. Silencing of CDC42 inhibits neuroblastoma cell proliferation and transformation. *Cancer Lett.* **2014**, 355 (2), 210–216.

(5) Gomez del Pulgar, T.; Benitah, S. A.; Valeron, P. F.; Espina, C.; Lacal, J. C. Rho GTPase expression in tumourigenesis: evidence for a significant link. *Bioessays* **2005**, *27* (6), 602–613.

(6) Haga, R. B.; Ridley, A. J. Rho GTPases: Regulation and roles in cancer cell biology. *Small GTPases* **2016**, 7 (4), 207–221.

(7) Melendez, J.; Grogg, M.; Zheng, Y. Signaling role of Cdc42 in regulating mammalian physiology. *J. Biol. Chem.* **2011**, 286 (4), 2375–2381.

(8) Hercyk, B. S.; Rich-Robinson, J.; Mitoubsi, A. S.; Harrell, M. A.; Das, M. E. A novel interplay between GEFs orchestrates Cdc42 activity during cell polarity and cytokinesis in fission yeast. *J. Cell Sci.* **2019**, *132* (23), No. jcs236018, DOI: 10.1242/jcs.236018.

(9) Gray, J. L.; von Delft, F.; Brennan, P. E. Targeting the Small GTPase Superfamily through Their Regulatory Proteins. *Angew. Chem., Int. Ed. Engl.* **2020**, *59* (16), 6342–6366.

(10) Cotteret, S.; Chernoff, J. The evolutionary history of effectors downstream of Cdc42 and Rac. *Genome Biol.* **2002**, *3* (2), No. REVIEWS0002.

(11) Murphy, N. P.; Mott, H. R.; Owen, D. Progress in the therapeutic inhibition of Cdc42 signalling. *Biochem. Soc. Trans.* 2021, 49 (3), 1443–1456.

(12) Maldonado, M. D. M.; Dharmawardhane, S. Targeting Rac and Cdc42 GTPases in Cancer. *Cancer Res.* **2018**, *78* (12), 3101–3111.

(13) Maldonado, M. D. M.; Medina, J. I.; Velazquez, L.; Dharmawardhane, S. Targeting Rac and Cdc42 GEFs in Metastatic Cancer. *Front. Cell Dev. Biol.* **2020**, *8*, 201.

(14) Xiao, X. H.; Lv, L. C.; Duan, J.; Wu, Y. M.; He, S. J.; Hu, Z. Z.; Xiong, L. X. Regulating Cdc42 and Its Signaling Pathways in Cancer: Small Molecules and MicroRNA as New Treatment Candidates. *Molecules* **2018**, *23* (4), 787 DOI: 10.3390/molecules23040787.

(15) Friesland, A.; Zhao, Y.; Chen, Y. H.; Wang, L.; Zhou, H.; Lu, Q. Small molecule targeting Cdc42-intersectin interaction disrupts Golgi organization and suppresses cell motility. *Proc. Natl. Acad. Sci. U. S. A.* **2013**, *110* (4), 1261–1266.

(16) Aguilar, B. J.; Zhao, Y.; Zhou, H.; Huo, S.; Chen, Y. H.; Lu, Q. Inhibition of Cdc42-intersectin interaction by small molecule ZCL367 impedes cancer cell cycle progression, proliferation, migration, and tumor growth. *Cancer Biol. Ther* **2019**, *20* (6), 740–749.

(17) Liu, W.; Du, W.; Shang, X.; Wang, L.; Evelyn, C.; Florian, M. C.; Ryan, M. A.; Rayes, A.; Zhao, X.; Setchell, K.; Meller, J.; Guo, F.; Nassar, N.; Geiger, H.; Pang, Q.; Zheng, Y. Rational identification of a Cdc42 inhibitor presents a new regimen for long-term hematopoietic stem cell mobilization. *Leukemia* **2019**, *33* (3), 749–761.

(18) Zins, K.; Gunawardhana, S.; Lucas, T.; Abraham, D.; Aharinejad, S. Targeting Cdc42 with the small molecule drug AZA197 suppresses primary colon cancer growth and prolongs survival in a preclinical mouse xenograft model by downregulation of PAK1 activity. J. Transl. Med. **2013**, *11*, 295.

(19) Gao, Y.; Dickerson, J. B.; Guo, F.; Zheng, J.; Zheng, Y. Rational design and characterization of a Rac GTPase-specific small molecule inhibitor. *Proc. Natl. Acad. Sci. U. S. A.* **2004**, *101* (20), 7618–7623.

(20) Muller, P. M.; Rademacher, J.; Bagshaw, R. D.; Wortmann, C.; Barth, C.; van Unen, J.; Alp, K. M.; Giudice, G.; Eccles, R. L.; Heinrich, L. E.; Pascual-Vargas, P.; Sanchez-Castro, M.; Brandenburg, L.; Mbamalu, G.; Tucholska, M.; Spatt, L.; Czajkowski, M. T.; Welke, R. W.; Zhang, S.; Nguyen, V.; Rrustemi, T.; Trnka, P.; Freitag, K.; Larsen, B.; Popp, O.; Mertins, P.; Gingras, A. C.; Roth, F. P.; Colwill, K.; Bakal, C.; Pertz, O.; Pawson, T.; Petsalaki, E.; Rocks, O. Systems analysis of RhoGEF and RhoGAP regulatory proteins reveals spatially organized RAC1 signalling from integrin adhesions. *Nat. Cell Biol.* **2020**, 22 (4), 498–511.

(21) Dütting, S.; Heidenreich, J.; Cherpokova, D.; Amin, E.; Zhang, S. C.; Ahmadian, M. R.; Brakebusch, C.; Nieswandt, B. Critical offtarget effects of the widely used Rac1 inhibitors NSC23766 and EHT1864 in mouse platelets. *J. Thromb. Haemost.* **2015**, *13*, 827–838.

(22) Jahid, S.; Ortega, J. A.; Vuong, L. M.; Acquistapace, I. M.; Hachey, S. J.; Flesher, J. L.; La Serra, M. A.; Brindani, N.; La Sala, G.; Manigrasso, J.; Arencibia, J. M.; Bertozzi, S. M.; Summa, M.; Bertorelli, R.; Armirotti, A.; Jin, R.; Liu, Z.; Chen, C. F.; Edwards, R.; Hughes, C. C. W.; De Vivo, M.; Ganesan, A. K. Structure-based design of CDC42 effector interaction inhibitors for the treatment of cancer. *Cell Rep* **2022**, *39* (4), No. 110760.

(23) Brindani, N.; Vuong, L. M.; Acquistapace, I. M.; La Serra, M. A.; Ortega, J. A.; Veronesi, M.; Bertozzi, S. M.; Summa, M.; Girotto, S.; Bertorelli, R.; Armirotti, A.; Ganesan, A. K.; De Vivo, M. Design, Synthesis, In Vitro and In Vivo Characterization of CDC42 GTPase

Interaction Inhibitors for the Treatment of Cancer. J. Med. Chem. 2023, 66 (8), 5981-6001.

(24) Stengel, K. R.; Zheng, Y. Essential role of Cdc42 in Ras-induced transformation revealed by gene targeting. *PLoS One* **2012**, 7 (6), No. e37317.

(25) Cheng, C. M.; Li, H.; Gasman, S.; Huang, J.; Schiff, R.; Chang, E. C. Compartmentalized Ras proteins transform NIH 3T3 cells with different efficiencies. *Mol. Cell. Biol.* **2011**, *31* (5), 983–997.

(26) Dalvit, C.; Caronni, D.; Mongelli, N.; Veronesi, M.; Vulpetti, A. NMR-based quality control approach for the identification of false positives and false negatives in high throughput screening. *Curr. Drug Discov Technol.* **2006**, *3* (2), 115–124.

(27) Seidel, S. A.; Dijkman, P. M.; Lea, W. A.; van den Bogaart, G.; Jerabek-Willemsen, M.; Lazic, A.; Joseph, J. S.; Srinivasan, P.; Baaske, P.; Simeonov, A.; Katritch, I.; Melo, F. A.; Ladbury, J. E.; Schreiber, G.; Watts, A.; Braun, D.; Duhr, S. Microscale thermophoresis quantifies biomolecular interactions under previously challenging conditions. *Methods* **2013**, *59* (3), 301–315.

(28) Mureddu, L. G.; Vuister, G. W. Fragment-Based Drug Discovery by NMR. Where Are the Successes and Where can It Be Improved? *Front Mol. Biosci* **2022**, *9*, No. 834453.

(29) Dalvit, C.; Flocco, M.; Veronesi, M.; Stockman, B. J. Fluorine-NMR competition binding experiments for high-throughput screening of large compound mixtures. *Comb Chem. High Throughput Screen* **2002**, 5 (8), 605–611.

(30) Dalvit, C.; Piotto, M. (19) F NMR transverse and longitudinal relaxation filter experiments for screening: a theoretical and experimental analysis. *Magn. Reson. Chem.* **2017**, 55 (2), 106–114.

(31) Dalvit, C.; Pevarello, P.; Tato, M.; Veronesi, M.; Vulpetti, A.; Sundstrom, M. Identification of compounds with binding affinity to proteins via magnetization transfer from bulk water. *J. Biomol NMR* **2000**, *18* (1), 65–68.

(32) Dalvit, C. NMR methods in fragment screening: theory and a comparison with other biophysical techniques. *Drug Discov Today* **2009**, *14* (21–22), 1051–1057.

(33) Troll, T. Reduction Potentials of Substituted as-Triazines and s-Tetrazines in acetonitrile. *Electrochim. Acta* **1982**, 27 (9), 1311–1314.

(34) Chiacchio, M. A.; Iannazzo, D.; Romeo, R.; Giofre, S. V.; Legnani, L. Pyridine and Pyrimidine Derivatives as Privileged Scaffolds in Biologically Active Agents. *Curr. Med. Chem.* **2020**, *26* (40), 7166–7195.

(35) Sander, K.; Kottke, T.; Tanrikulu, Y.; Proschak, E.; Weizel, L.; Schneider, E. H.; Seifert, R.; Schneider, G.; Stark, H. 2,4-Diaminopyrimidines as histamine H4 receptor ligands–Scaffold optimization and pharmacological characterization. *Bioorg. Med. Chem.* **2009**, *17* (20), 7186–7196.

(36) Mowbray, C. E.; Bell, A. S.; Clarke, N. P.; Collins, M.; Jones, R. M.; Lane, C. A.; Liu, W. L.; Newman, S. D.; Paradowski, M.; Schenck, E. J.; Selby, M. D.; Swain, N. A.; Williams, D. H. Challenges of drug discovery in novel target space. The discovery and evaluation of PF-3893787: a novel histamine H4 receptor antagonist. *Bioorg. Med. Chem. Lett.* **2011**, *21* (21), 6596–6602.

(37) Hammer, S. G.; Gobleder, S.; Naporra, F.; Wittmann, H. J.; Elz, S.; Heinrich, M. R.; Strasser, A. 2,4-Diaminopyrimidines as dual ligands at the histamine H1 and H4 receptor-H1/H4-receptor selectivity. *Bioorg. Med. Chem. Lett.* **2016**, *26* (2), 292–300.

(38) Natarajan, R.; Anthoni Samy, H. N.; Sivaperuman, A.; Subramani, A. Structure-Activity Relationships of Pyrimidine Derivatives and their Biological Activity - A Review. *Med. Chem.* **2022**, *19* (1), 10–30.

(39) Sastry, G. M.; Adzhigirey, M.; Day, T.; Annabhimoju, R.; Sherman, W. Protein and ligand preparation: parameters, protocols, and influence on virtual screening enrichments. *J. Comput. Aided Mol. Des.* **2013**, *27* (3), 221–234.

(40) Friesner, R. A.; Murphy, R. B.; Repasky, M. P.; Frye, L. L.; Greenwood, J. R.; Halgren, T. A.; Sanschagrin, P. C.; Mainz, D. T. Extra precision glide: docking and scoring incorporating a model of hydrophobic enclosure for protein-ligand complexes. *J. Med. Chem.* **2006**, 49 (21), 6177–6196.

(41) Halgren, T. A.; Murphy, R. B.; Friesner, R. A.; Beard, H. S.; Frye, L. L.; Pollard, W. T.; Banks, J. L. Glide: a new approach for rapid, accurate docking and scoring. 2. Enrichment factors in database screening. J. Med. Chem. 2004, 47 (7), 1750–1759.

(42) Friesner, R. A.; Banks, J. L.; Murphy, R. B.; Halgren, T. A.; Klicic, J. J.; Mainz, D. T.; Repasky, M. P.; Knoll, E. H.; Shelley, M.; Perry, J. K.; Shaw, D. E.; Francis, P.; Shenkin, P. S. Glide: a new approach for rapid, accurate docking and scoring. 1. Method and assessment of docking accuracy. *J. Med. Chem.* **2004**, 47 (7), 1739– 1749.

(43) Jorgensen, W. L.; Chandrasekhar, J.; Madura, J. D.; Impey, R. W.; Klein, M. L. Comparison of simple potential functions for simulating liquid water. *J. Chem. Phys.* **1983**, *79* (2), 926–935.

(44) Maier, J. A.; Martinez, C.; Kasavajhala, K.; Wickstrom, L.; Hauser, K. E.; Simmerling, C. ff14SB: Improving the Accuracy of Protein Side Chain and Backbone Parameters from ff99SB. J. Chem. Theory Comput **2015**, *11* (8), 3696–3713.

(45) Singĥ, U. C.; Kollman, P. A. An approach to computing electrostatic charges for molecules. *J. Comput. Chem.* **1984**, *5* (2), 129–145.

(46) Meagher, K. L.; Redman, L. T.; Carlson, H. A. Development of polyphosphate parameters for use with the AMBER force field. *J. Comput. Chem.* **2003**, *24* (9), 1016–1025.

(47) Allner, O.; Nilsson, L.; Villa, A. Magnesium Ion-Water Coordination and Exchange in Biomolecular Simulations. *J. Chem. Theory Comput* **2012**, *8* (4), 1493–1502.

(48) Joung, I. S.; Cheatham, T. E. Determination of alkali and halide monovalent ion parameters for use in explicitly solvated biomolecular simulations. *J. Phys. Chem. B* **2008**, *112* (30), 9020–9041. third,

(49) Case, D. A.; Aktulga, H. M.; Belfon, K. A. A.; Ben-Shalom, I.; Berryman, J. T.; Brozell, S. R.; Cerutti, D. S.; Cheatham, T. E.; Cisneros, G. A.; Cruzeiro, V. W. D.; Darden, T. A.; Duke, R. E.; Giambasu, G.; Gilson, M. K.; Gohlke, H.; Goetz, A. W.; Harris, R.; Izadi, S.; Izmailov, S. A.; Kasavajhala, K.; Kaymak, M. C.; King, E.; Kovalenko, A.; Kurtzman, T.; Lee, T. S.; LeGrand, S.; Li, P.; Lin, C.; Liu, J.; Luchko, T.; Luo, R.; Machado, M.; Man, V.; Manathunga, M.; Merz, K. M.; Miao, Y.; Mikhailovskii, O.; Monard, G.; Nguyen, H.; O'Hearn, K. A.; Onufriev, A.; Pan, F.; Pantano, S.; Qi, R.; Rahnamoun, A.; Roe, D. R.; Roitberg, A.; Sagui, C.; Schott-Verdugo, S.; Shajan, A.; Shen, J.; Simmerling, C. L.; Skrynnikov, N.; Smith, J.; Swails, J. M.; Walker, R. C.; Wang, J.; Wang, J.; Wei, H.; Wolf, R. M.; Wu, X.; Xiong, Y.; Xue, Y.; York, D. M.; Zhao, S.; Kollman, P. A. *Amber*; University of California: San Francisco, 2022.

(50) Andrea, T. A.; Swope, W. C.; Andersen, H. C. The role of long ranged forces in determining the structure and properties of liquid water. *J. Chem. Phys.* **1983**, *79*, 4576–4584.