# **UC Davis UC Davis Previously Published Works**

## **Title**

Anesthetic Preconditioning Combined with Postconditioning Offers No Additional Benefit Over Preconditioning or Postconditioning Alone

# **Permalink**

<https://escholarship.org/uc/item/3gx6x31h>

**Journal** Anesthesia & Analgesia, 105(2)

# **ISSN**

0003-2999

# **Authors**

Deyhimy, David I Fleming, Neal W Brodkin, Ian G [et al.](https://escholarship.org/uc/item/3gx6x31h#author)

# **Publication Date**

2007-08-01

## **DOI**

10.1213/01.ane.0000267524.71445.e7

## **Copyright Information**

This work is made available under the terms of a Creative Commons Attribution License, available at<https://creativecommons.org/licenses/by/4.0/>

Peer reviewed

# **Anesthetic Preconditioning Combined with Postconditioning Offers No Additional Benefit Over Preconditioning or Postconditioning Alone**

David I. Deyhimy, MD\*

Neal W. Fleming, MD, PhD\*

Ian G. Brodkin, MD†

Hong Liu, MD\*

BACKGROUND: Recent investigations demonstrate that anesthetic preconditioning and postconditioning reduce myocardial infarct size to a degree comparable to that achieved with ischemic preconditioning. We hypothesized that the combination of sevoflurane preconditioning and postconditioning would result in greater preservation of myocardium.

METHODS: Langendorff perfused rat hearts were divided into four groups: control, preconditioning, postconditioning, and preconditioning plus postconditioning. During reperfusion, left ventricular function (left ventricular developed pressure, left ventricular end diastolic pressure, and d*p*/d*t*) were measured. At the end of reperfusion, the infarct sizes were measured with 2,3,5 triphenyltetrazolium chloride staining. Nuclear magnetic resonance was used to measure intracellular pH, Na<sup>+</sup>, and  $\overline{Ca}^{2+}$ .

RESULTS: Left ventricular developed pressure, left ventricular end diastolic pressure, left ventricular  $dp/dt_{\text{max}}$  and  $dp/dt_{\text{min}}$  were significantly improved in the treatment groups when compared with those in the controls. Myocardial infarct size (24%  $\pm$  7%, 16%  $\pm$  8%, and 22%  $\pm$  7% in preconditioning, postconditioning, and pre-plus postconditioning groups versus  $44\% \pm 8\%$  in the control group,  $P < 0.05$ ) and intracellular Na<sup>+</sup> and Ca<sup>2+</sup> were significantly decreased in all experimental groups at the end of reperfusion when compared with those in control. However, there were no differences between these variables in each treatment group.

CONCLUSION: Sevoflurane postconditioning is as effective as preconditioning in protecting myocardial function after global ischemia. The combination of sevoflurane preconditioning and postconditioning offered no additional benefit over either intervention alone.

(Anesth Analg 2007;105:316 –24)

Ince Murry et al. (1) first described ischemic preconditioning (IPC) in dogs, numerous studies have confirmed that periods of myocardial ischemia limit the extent of subsequent myocardial infarction in various animal models. Other forms of preconditioning (pharmacologic) including anesthetic preconditioning (exposure to volatile anesthetics known as APC) have also been described, and studies have

From the \*Department of Anesthesiology and Pain Medicine, University of California, Davis, California; and †Department of Anesthesiology, Vancouver General Hospital, Vancouver, British Columbia, Canada.

Accepted for publication April 5, 2007.

Supported in part by the University of California Davis Health System Research Award, Davis, CA, and Abbott Laboratories' unrestricted grant. Spectrometers were further supported by grant No. RR08206 from the National Institutes of Health, Bethesda, MD.

Part of the results were presented at the American Society of Anesthesiologists 2005 Annual Meeting, October 22–26, 2005, Atlanta, Georgia.

Address correspondence and reprint requests to Hong Liu, MD, Department of Anesthesiology and Pain Medicine, University of California Davis Health System, 4150 v. Street, Suite 1200, Sacramento, CA 95817. Address e-mail to hualiu@ucdavis.edu.

Copyright © 2007 International Anesthesia Research Society DOI: 10.1213/01.ane.0000267524.71445.e7

demonstrated similar efficacy of APC and IPC in protecting the myocardium against prolonged ischemia (2–4). Preconditioning the myocardium results in improved recovery of ventricular function, decreased incidence of arrhythmias and, most importantly, decreased infarct size (IS) (3,5). There are numerous underlying cellular mechanisms through which preconditioning confers protection to the myocardium (2), but one of the key end effects is prevention of intracellular calcium  $([Ca^{2+}]_i)$  accumulation. Calcium accumulation has been repeatedly shown to play a major role in cellular dysfunction and death after ischemia and reperfusion (I/R) (6,7). Preconditioning prevents  $\left[Ca^{2+}\right]_i$  accumulation by activating protective cellular mechanisms, some of which ultimately result in the activation of mitochondrial and sarcolemmal ATP sensitive potassium channels  $(K_{ATP})$ , and Na/H exchangers within myocytes (2,3,8,9).

Although preconditioning is an effective mechanism for protection against myocardial ischemic damage in a laboratory setting, it has significant limitations in a clinical setting, as ischemic episodes in humans are often unpredictable and precipitous. One would have to know in advance that a patient was about to have a myocardial infarction in order to provide appropriately timed preconditioning. Recently, the concept of postconditioning, whereby ischemia or volatile anesthetics are introduced immediately upon reperfusion in an effort to attenuate myocardial I/R injury, has garnered increased attention (10 –14). Postconditioning could have significant clinical applications for patients with continuing myocardial ischemia who arrive in the operating room or cardiac catheterization laboratory for revascularization as it can be implemented at the time of reperfusion.

The goal of this study was to investigate, in an experimental model, whether the combination of sevoflurane preconditioning followed by sevoflurane postconditioning would result in decreasing intracellular  $Na<sup>+</sup>$  and  $Ca<sup>2+</sup>$  concentrations, and decreased myocardial IS by attenuating reperfusion injury, and thus provide greater preservation of myocardial function than either modality alone.

### **METHODS**

This study was approved by the Animal Care Committee of the University of California, Davis (Davis, CA), and all of the experiments were performed within the guidelines for animal care from the National Institutes of Health (Bethesda, MD).

#### Preparation of Isolated Rat Hearts

Male Fischer 344 rats, aged 2–4 mo, were anesthetized with intraperitoneal pentobarbital (50 mg/kg) plus heparin (1000 IU/kg). Once surgical anesthesia was achieved (no response to tail clamp), the hearts were excised via thoracotomy and rapidly transferred to a nonrecirculating Langendorff perfusion system. The aorta was cannulated and the hearts perfused with a Krebs–Henselet bicarbonate solution (127 mM NaCl, 4.75 mM KCl, 1.25 mM  $MgCl_2$ , 2.5 mM CaCl<sub>2</sub>, 25 mM NaHCO<sub>3</sub>, and 10 mM glucose). Perfusion pressure was maintained between 120 and 140 cm  $H_2O$ , and the perfusate was continuously oxygenated with 95% O<sub>2</sub>/5% CO<sub>2</sub> maintaining a pH of 7.40  $\pm$  0.05. Temperature was kept constant at 37°C  $\pm$  0.5°C with a water-jacketed column. A balloon-tipped pressure transducer was placed into the left ventricle (LV) via an incision in the left atrial appendage, and LV end diastolic pressure (LVEDP) was set at 10 cm  $H_2O$ . The hearts were paced at 5 Hz via right atrial pacing wires. All preparations were allowed a 20-min period for equilibration. Global ischemia was achieved by interrupting perfusion to the heart. Pacing was discontinued during the ischemic period.

#### Experimental Design

The rats were randomly divided into four groups consisting of six hearts per group. Three of the groups underwent treatment and one group served as a control (Fig. 1). Rat hearts in the treatment groups were exposed to sevoflurane delivered to the gas mixture at a concentration of 2.5% via a sevotec5

### **Experimental Groups**



Figure 1. Schematic illustration of the experimental protocols used in this study.  $sevo = sevoflurane$ .

variable bypass vaporizer (Datex-Ohmeda, Milwaukee, WI). After the 20-min equilibration period, hearts in the preconditioning group were exposed to sevoflurane (2.5%) for 10 min, followed by a 5-min washout period before 25 min of global ischemia and a 60-min reperfusion. Hearts in the postconditioning group underwent equilibration, then 25 min of global ischemia, followed immediately by sevoflurane (2.5%) exposure for the first 10 min of the 60-min reperfusion period. The preconditioning plus postconditioning group incorporated both protocols. The control group received no treatment before 25 min of global ischemia and none during the 60-min reperfusion period.

#### Myocardial IS

At the end of reperfusion, hearts were removed from the Langendorff apparatus and quickly sectioned into 2-mm slices. The slices were immersed in 2% 2,3,5 triphenyltetrazolium chloride staining solution and placed in a 37°C incubator for 20 min. After incubation, slices were washed with water, placed on petri dishes and then scanned into a computer using Adobe Photoshop software (Adobe, San Jose, CA). Standard computer plainmeteric analysis, using NIH image 1.62 (National Institutes of Health, Bethesda, MD), was performed to determine infarct area. The total area of infarction was divided by the total area of myocardium to yield the percent area of infarction (15).

#### LV Function

LV pressures were measured using a latex balloon filled with water inserted via an incision in the left atrial appendage through the mitral valve. Pressures were recorded using Powerlab 4/20 (ADInstruments, CO Springs, CO). During the equilibration period, LVEDP was set by adjusting the balloon volume to yield a left end diastolic pressure of approximately 10 cm  $H_2O$ . LV pressure measurements included: LV developed pressure (LVDP), calculated as LV end systolic pressure minus LVEDP and LVDP over time, LV d $p/dt_{\text{max}}$  and d $p/dt_{\text{min}}$ .



The unit for left ventricular end systolic pressure (LVESP) data is cm H<sub>2</sub>O. Heart rate (HR) is bpm. Infarct size (IS) expressed as % area of necrosis and the creatine kinase (CK) unit is IU/g dry weight. Data are presented as mean  $\pm$  SD.

\* Treatment groups versus. Control,  $P < 0.05$  is considered significant.  $n = 6$  in each group.

#### Creatine Kinase Analysis

Coronary sinus effluent was collected in 10-min aliquots throughout each experiment for measurement of creatine kinase (CK) as an indicator of myocardial cell damage. The amount of CK released was determined using a CK-10 kit (SIGMA Diagnostics, St. Louis, MO) and a Shimadzu UV-VIS recording photospectrometer (Shimadzu, Columbia, MD). Units are expressed as U/gm dry weight (15).

#### Nuclear Magnetic Resonance Spectroscopy

 $19$ F NMR spectroscopy was performed to measure  $[Ca^{2+}]_i$ . The hearts were loaded for 30 min with perfusate containing the acetoxymethyl ester of 5F-1, 2-bis (2-aminophenoxy)ethane-*N*,*N*,*N*,*N*-tetraacetic acid (FBAPTA) at 2.5  $\mu$ M. FBAPTA was washed out of the extracellular space with control solution before measurement of  $[\text{Ca}^{2+}]_i$ . <sup>19</sup>F spectra were generated from the summed free induction decays of 1500, 45° excitation pulses using  $2K$  word data files and  $\pm 5000$ Hz sweep width.  $[Ca^{2+}]$ <sub>i</sub> was calculated by using:  $[Ca^{2+}]_i = Kd$  [bound]/[free], where the ratio of  $Ca^{2+}$ bound and free FBAPTA concentrations was equal to the ratio of their corresponding peak areas and Kd is 300 nM.

<sup>31</sup>P NMR spectroscopy was assessed from the summed free induction decays of 148, 60° excitation pulses using 4K word data files and  $\pm 4000$  Hz sweep width. Intracellular pH (pH<sub>i</sub>) was calculated from the shift in inorganic phosphorus resonance calibrated at 37°C with reference to control phosphocreatine. Highenergy phosphates were determined as percent of control peak intensity (15).

<sup>23</sup>Na NMR spectroscopy was performed to measure  $intrac{ellular}$  sodium concentrations  $([Na_i])$  by substituting 7.5 mM dysprosium triethylenetetraminehexaacetic acid for NaCl and by adding and  $Ca^{2+}$  2.5 mM (measured by a calcium electrode) to the perfusate. <sup>23</sup>Na spectra were generated from the summed free induction decays of 1000 with excitation pulses at  $90^{\circ}$  using 2K word data files and  $\pm 4000$  Hz sweep widths. The  $[Na_i^+]$  in mEq/kg dry weight was calculated from the calibrated area under the unshifted peak of the <sup>23</sup>Na spectra after subtracting the extracellular peak. At the end of the experiment, the hearts were weighed wet and then dried (at least 48 h) at 65°C to determine dry weight (15).

#### Statistical Analysis

Analysis of variance for repeated measures was used to test differences among groups. When differences among groups were found, a *post hoc* Tukey *t*-test was used to determine where these differences occurred. For all comparisons, differences were considered significant when  $P < 0.05$ . Data are reported as mean  $\pm$  sp.

### **RESULTS**

#### IS

In all three groups exposed to sevoflurane (preconditioning, postconditioning, and pre-plus postconditioning), IS was significantly decreased versus control (Table 1). The preconditioning, postconditioning, and pre-plus postconditioning groups had infarct areas of  $24\%$   $\pm$  7%, 16%  $\pm$  8%, and 22%  $\pm$  7% respectively, compared with the control group of  $44\% \pm 8\%$  ( $P <$ 0.05). Differences among the treatment groups were not statistically significant.

### **CK**

The CK results are listed in Table 1 as the total CK (U/gm dry weight) released during the 60-min reperfusion. APC rat hearts released significantly less CK than control rats:  $341 \pm 51$  vs  $1367 \pm 562$  ( $P < 0.05$ ). Postconditioning hearts released significantly less CK than control hearts:  $378 \pm 67$  vs  $1367 \pm 562$  ( $P < 0.05$ ). Combined pre-plus postconditioning hearts also released significantly less CK (422  $\pm$  89) than control hearts ( $P < 0.05$ ). There were no statistical differences for CK release among the three sevoflurane treatment groups.

#### LV Function

All LV function parameters measured (LVDP% recovery, LVEDP, LV  $dp/dt_{\text{max}}$  and  $dp/dt_{\text{min}}$ ) were better ( $P < 0.05$ ) in the sevoflurane treatment groups (preconditioning, postconditioning, and pre-plus postconditioning) compared with the control group. LVDP is expressed as the percentage of baseline



Figure 2. A. Recovery of left ventricular developed pressure (LVDP) during reperfusion. The LVDP was measured throughout reperfusion as a percentage compared to the baseline LVDP during the equilibration phase prior to ischemia. All experimental groups (preconditioning: closed square; postconditioning: closed triangle; and pre-plus postconditioning: closed circle) had significant ( $P < 0.05$ ) recoveries of LVDP versus control (open square). There were no statistical differences among the three experimental groups. B. Results of left ventricular end diastolic pressure (LVEDP). Units are in cm  $H_2O$ . There were significantly lower diastolic pressures in the preconditioning and pre-plus postconditioning groups during ischemia (*P* 0.05) compared with those in the postconditioning and control groups. There were significantly lower diastolic pressures in the experimental groups during reperfusion  $(P < 0.05)$  compared with those in the control. There were no statistical differences among the three experimental groups during reperfusion. C. Recovery of left ventricular systolic function. LV d $p/dt_{\text{max}}$  values (in cm H<sub>2</sub>O<sub>S</sub><sup>-1</sup>) were measured throughout reperfusion. All experimental groups had significant (*P* < 0.05) recoveries of LV  $dp/dt_{\text{max}}$  versus control. There were no statistical differences among the three experimental groups. D. Recovery of left ventricular diastolic function. LV  $dp/dt_{min}$  values (in cm  $H_2O \cdot s^{-1}$ ) were measured throughout reperfusion. The LV  $dp/dt_{min}$  values were significantly lower in all the experimental groups ( $P < 0.05$ ) versus control. There were no statistical differences among the three experimental groups. \*Experimental groups versus control  $(P < 0.05)$ . #Preconditioning and pre-plus postconditioning versus control and postconditioning ( $P < 0.05$ ). Data presented as mean  $\pm$  sp.  $n = 6$  in each group. pre-plus postconditioning: combined preconditioning and postconditioning.

function recovery after 60 min of reperfusion. All other values reflect function after 60 min of reperfusion. There were no statistically significant differences among the sevoflurane treatment groups for any of the measured LV function parameters. LVDP recovery (% of baseline) was  $42\% \pm 8\%$ , 60%  $\pm 14\%$ , and 55%  $\pm$ 28% for pre-, post-, and pre-plus postconditioning groups, respectively, compared with the control group  $10\% \pm 4\%$  recovery (Fig. 2A). LVEDP values (in cm H<sub>2</sub>O) after reperfusion were 25  $\pm$  10, 24  $\pm$  10, and 21  $\pm$ 6 respectively for the sevoflurane treatment groups

versus  $63 \pm 16$  for the control group (Fig. 2B). LV  $dp/dt_{\text{max}}$  values (in cm  $H_2O \cdot s^{-1}$ ) were 1102  $\pm$  378,  $1598 \pm 614$ , and  $1170 \pm 649$ , respectively, for the sevoflurane treatment groups versus  $491 \pm 259$  for the control group (Fig. 2C). LV dp/dt<sub>min</sub> values (in cm  $H_2O \cdot s^{-1}$ ) were  $-543 \pm 54$ ,  $-822 \pm 201$ , and  $-656 \pm 100$ 388, respectively, for the sevoflurane treatment groups versus  $-355 \pm 161$  for the control group (Fig. 2D). There were no statistically significant differences in heart rate and LV end systolic pressure among all four groups (Table 1).



Figure 3. Ischemia decreased ATP levels during the experiment. The ATP was better preserved in all the experimental groups (preconditioning: closed square; postconditioning: closed triangle; and pre-plus postconditioning: closed circle)  $(P < 0.05)$  versus control (open square). There were no statistical differences among the three experimental groups. \*Experimental groups versus control ( $P < 0.05$ ). Data presented as mean  $\pm$  sp.  $n = 6$  in each group. pre-plus postconditioning: combined preconditioning and postconditioning. ATP adenosine triphosphate.

Figure 4. Ischemia causes an increase in intracellular Ca  $\left[Ca^{2+}\right]_i$  in control hearts (open square) during ischemia and reperfusion and in the postconditioning heart (closed triangle) during ischemia. The increases in  $[\text{Ca}^{2+}]$ , were significantly limited in preconditioning hearts (closed square) and pre-plus postconditioning hearts (closed circle) during ischemia and in all the experimental groups (preconditioning: closed square; postconditioning: closed triangle; and pre-plus postconditioning: closed circle) (*P* 0.05) versus control (open square) during reperfusion. There were no statistical differences among the three experimental groups during reperfusion. \*Experimental groups versus control  $(P < 0.05)$ . #Preconditioning and pre-plus postconditioning versus control and postconditioning  $(P < 0.05)$ . Data presented as mean  $\pm$  sp.  $n = 6$  in each group. pre-plus postconditioning: combined preconditioning and postconditioning.

### ATP Preservation

Results for ATP measurements are shown in Figure 3. Myocardial ischemia led to a significant ( $P < 0.05$ ) decrease in the ATP levels in all groups. The preconditioning and pre-plus postconditioning groups had higher ATP levels at the end of ischemia (34  $\pm$  12 and  $33 \pm 10$ , respectively) than the control and postconditioning groups ( $24 \pm 8$  and  $23 \pm 8$  respectively). At the end of reperfusion, all sevoflurane treatment groups had significantly better recovery of ATP levels than the control group. The preconditioning, postconditioning, pre-plus postconditioning and control group ATP values were  $44 \pm 10$ ,  $42 \pm 8$ ,  $46 \pm 9$ , and  $23 \pm 7$ ,

respectively. Although the treatment groups had statistically significant recovery of ATP versus the control group, there were no significant differences among the treatment groups.

### $\lceil$ Ca<sup>2+</sup> $\rceil$ <sub>i</sub>

The effects of I/R on  $[Ca^{2+}]_i$  are shown in Figure 4. In the control hearts,  $\left[Ca^{2+}\right]_i$  increased from  $260 \pm 16$ at baseline to 1081  $\pm$  177 at the end of ischemia, and to  $1187 \pm 324$  at the end of reperfusion. Exposure to sevoflurane attenuated the increases in  $[Ca^{2+}]$ <sub>i</sub> in preconditioning, postconditioning, and pre-plus postconditioning groups. By the end of reperfusion  $\left[Ca^{2+}\right]$ 



Figure 5. Ischemia causes an increase in intracellular Na  $(Na^+$ <sub>i</sub>) in control hearts (open square) during ischemia and reperfusion and in the postconditioning heart (closed triangle) during ischemia. The increases in Na<sup>+</sup><sub>i</sub> were significantly limited in all the experimental groups (preconditioning: closed square; postconditioning: closed triangle; and pre-plus postconditioning: closed circle) (*P* < 0.05) versus control (open square) during reperfusion. There is no statistical difference between the control and postconditioning groups during ischemia. There were no statistical differences among the three experimental groups during reperfusion. \*Experimental groups versus control  $(P < 0.05)$ . #Preconditioning and pre-plus postconditioning versus control and postconditioning ( $P < 0.05$ ). Data presented as mean  $\pm$  sp.  $n = 6$  in each group. pre-plus postconditioning: combined preconditioning and postconditioning. "We only used one sevoflurane concentration, a higher concentration (1.5 MAC) experiment could show a different result when used in a combined pre- and postconditioning protocol."

levels were  $395 \pm 94$  in the preconditioning group,  $336 \pm 40$  in the postconditioning group, and  $470 \pm 111$ in the pre-plus postconditioning group. The decreases in  $\left[\text{Ca}^{2+}\right]_i$  seen in the three treatment groups were statistically significant compared with those seen in the control group ( $P < 0.05$ ), but there were no statistical differences among the three treatment groups.

## Na<sup>+</sup><sub>i</sub> Accumulation

Changes in  $Na<sup>+</sup><sub>i</sub>$  accumulation that occurred during I/R are shown in Figure 5. In the control hearts, baseline Na<sup>+</sup><sub>i</sub> increased from 30  $\pm$  9 to 104  $\pm$  8 at the end of ischemia and declined to  $33 \pm 3$  at the end of reperfusion. Exposure to sevoflurane attenuated  $\mathrm{Na}^{+}_{\;\;i}$ accumulation in the preconditioning, postconditioning, and pre-plus postconditioning groups. Na<sup>+</sup><sub>i</sub> levels at the end of ischemia and the end of reperfusion were  $64 \pm 3$  and  $27 \pm 2$  in the preconditioning group,  $101 \pm 4$  and  $28 \pm 2$  in the postconditioning group, and  $69 \pm 7$  and  $28 \pm 4$  in the pre-plus postconditioning group. The decreased  $\text{Na}^+$  accumulation at the end of ischemia seen in the three treatment groups was statistically significant when compared with that in the control group ( $P < 0.05$ ), but there were no statistical differences among the three treatment groups.

#### Intracellular Proton Accumulation

At the end of 25 min of sustained ischemia  $pH_i$ decreased in all groups: control  $(5.91 \pm 0.06)$ , preconditioning (6.19  $\pm$  0.10), postconditioning (5.96  $\pm$  0.06),

and pre-plus postconditioning group  $(6.17 \pm 0.10)$ . After 60 min of reperfusion, all groups had a similar recovery of pH<sub>i</sub>:control (7.13  $\pm$  0.10), preconditioning  $(7.21 \pm 0.15)$ , postconditioning  $(7.09 \pm 0.05)$ , and pre-plus postconditioning  $(7.12 \pm 0.09)$  (Table 1). There were no statistical differences among any of the groups at the end of ischemia or at the end of reperfusion.

### **DISCUSSION**

In this study, APC and postconditioning with sevoflurane, alone or in combination, was shown to be equally effective in preventing the increases in Na<sup>+</sup><sub>i</sub> and  $\left[\text{Ca}^{2+}\right]_{i}$ , decreasing IS and preserving ATP and LV function after I/R.

Myocardial ischemia during the perioperative period is a serious risk for patients undergoing both cardiac and noncardiac operations. About 30 – 40% of patients with coronary artery disease experience perioperative myocardial ischemia during noncardiac surgery, increasing the risk for myocardial infarction (16). Anesthetic postconditioning is a potentially useful clinical intervention, as it can be implemented in patients who arrive at the hospital with ischemia and myocardial injury already in progress (17). Anesthetic postconditioning is easy to implement and may be used in combination with a variety of revascularization procedures, such as percutaneous transluminal coronary angioplasty and coronary artery bypass grafting.

Reduction in myocardial IS is the "gold standard" in determining efficacy of preconditioning or postconditioning protocols for myocardial protection against I/R injury (18,19). Numerous studies have demonstrated APC to be an effective intervention against I/R injury (3–5). This study demonstrated that sevoflurane preconditioning, postconditioning, and pre-plus postconditioning all provided similar significant reductions (approximately 50%) in IS when compared with a control group. The combination of sevoflurane preconditioning and postconditioning provided no significant benefit over either intervention alone for any of the measured parameters. We also demonstrated that sevoflurane preconditioning, postconditioning, and pre-plus postconditioning provided similar, significant, reductions in CK release when compared with a control group, correlated with the reduction in IS seen in the treatment groups.

Previous studies have demonstrated that preconditioning not only decreases IS, but also better preserves LV function after I/R (2,20,21). We demonstrated that both sevoflurane preconditioning and postconditioning preserve LV function after I/R to a similar degree. Across all measures of LV function (systolic and diastolic), the sevoflurane-treated groups had significantly better postischemic function when compared with controls. The improved functional recovery correlates with the decreased IS discussed previously. The sevoflurane pre-plus postconditioning group showed no additional improvement over either intervention alone.

Loss of calcium homeostasis with subsequent  $[Ca<sup>2+</sup>]$ <sub>i</sub> accumulation results in cell damage and death (2,7,22–24). Ischemia results in a diminished ATP supply and increases in  $H^+$  production, which stimulates the Na<sup>+</sup>/H<sup>+</sup> exchanger (NHE-1) to transport  $H^+$ out of the cell in exchange for  $Na^+$ , leading to  $Na^+$ <sub>i</sub> accumulation. This results in an increased  $[Ca^{2+}]$ <sub>i</sub> via  $Na^+/Ca^{2+}$  exchanger. In all three of our treatment groups,  $[Ca^{2+}]$ <sub>i</sub> was substantially lower at the end of reperfusion when compared with controls. The lower  $[Ca^{2+}]$ <sub>i</sub> correlated with the improved LV function and decreased IS seen in the treatment groups. There were no statistical differences among the treatment groups, suggesting that similar protective mechanisms may be responsible for the decreased  $[Ca^{2+}]$ <sub>i</sub> accumulation observed for both pre- and postconditioning. Similarly, Na<sup>+</sup><sub>i</sub> levels were substantially lower in the three treatment groups compared with that in the control. A lower level of  $\mathrm{\hat{Na}^+_{i}}$  results in less  $[\mathrm{Ca}^{2+}]_i$  accumulation and presumably improved LV function and decreased IS, all of which were observed in all three treatment groups. Again, there were no statistical differences among the treatment groups suggesting similar protective mechanisms for both preconditioning and postconditioning.

As expected,  $pH_i$  decreased for all groups during ischemia. During reperfusion, all groups showed similar recovery of  $pH_i$  to near normal preischemic levels. The recovery of  $pH_i$  likely involves the NHE-1 and  $Na^+/Ca^{2+}$  exchanger discussed previously. The cell is able to improve pH<sub>i</sub> by transferring  $H^+$  out of the cell at the expense of  $Na<sup>+</sup>$  and  $Ca<sup>2+</sup>$  accumulation. However, other mechanisms may also be involved in maintaining  $pH_i$  and may explain why there was no significant difference among the treatment and control groups. Some studies indicate that preconditioning does not impact certain cellular mechanisms responsible for maintenance of normal  $pH_i$  during  $I/R$  $(25,26).$ 

A multitude of vital cellular mechanisms depend on constant production of ATP by the mitochondria including maintenance of low  $[Ca^{2+}]$ ; levels via the ATP-dependant ion exchangers discussed above. A number of studies have demonstrated that mitochondrial function is often profoundly disrupted by I/R injury, resulting in impairment of cellular respiration and a tremendous decrease in ATP production (2,8,27). APC has been shown to be protective by preserving mitochondrial function and maintaining ATP levels after I/R injury (2,3,5). Our results are consistent with previous findings demonstrating that APC results in preservation of mitochondrial function and maintenance of ATP stores. Both of the preconditioned groups (preconditioning alone and pre-plus postconditioning) had significantly higher ATP levels at the end of ischemia than the control and postconditioning groups. However, by the end of the reperfusion period, all three treatment groups had similar recovery of ATP levels relative to the control. Again, there were no statistically significant differences among the three treatment groups, indicating that the combination of anesthetic pre-plus postconditioning does not provide additional protection over either intervention alone. These findings demonstrate that anesthetic postconditioning preserves mitochondrial function and ATP production to a similar degree as APC alone. Interestingly, the postconditioning group showed similar ATP levels during ischemia to the control group, but recovered ATP levels to those seen in the preconditioned groups by the end of the reperfusion period. This suggests that attenuation or prevention of reperfusion injury is vital for the preservation of mitochondrial function. In addition, the postconditioning intervention, whether ischemic or anesthetic, needs to be implemented immediately upon reperfusion if reperfusion injury is to be prevented (10,28).

Although we are uncertain as to why APC and postconditioning are not additive, we surmise that similar end cellular effectors are induced by either intervention to a comparable degree. Similar studies have been performed with mixed results (13,14,19). Halkos et al. (19) combined IPC and postconditioning without benefit over either intervention alone. Chiari et al. (14) combined IPC and anesthetic postconditioning with isoflurane and found an additive effect versus either intervention alone, suggesting differing mechanisms of myocardial protection between ischemic and anesthetic postconditioning. Finally, Obal et al. (13) were able to show an additive benefit by combining anesthetic pre- and postconditioning with an *in vivo* rat model, which differed from our findings with an *in vitro* rat model.

Both IPC and APC are effective in preserving myocardial function and decreasing IS in numerous species (10,17,18) and volatile anesthetics are powerful mediators of preconditioning and postconditioning (2,11–14,28 –30). Although definitive end-points, such as IS reduction, are difficult to measure in humans, studies using markers of myocardial injury (troponins and CK) indicate less myocardial damage in patients who underwent preconditioning or postconditioning (17,18,20). However, the obvious drawback to preconditioning is lack of foreknowledge of an impending ischemic event. In contrast, postconditioning can be implemented in patients with continuing ischemia at the onset of reperfusion after revascularization procedures, such as coronary angioplasty and stenting, coronary artery bypass grafting, and possibly even organ transplantation.

ANESTHESIA & ANALGESIA

Our study has several significant limitations that should be considered in interpreting the data. We did not specifically evaluate the mechanisms through which our preconditioning or postconditioning groups were able to provide myocardial protection, so it is difficult to draw definitive conclusions as to what they have in common or how they differ. In addition, a number of studies have demonstrated that delayed APC (24 or 48 h) before ischemia is particularly effective in providing myocardial protection as inducible cytoprotective proteins likely provide additional protection against ischemic injury (6,29). Perhaps, if we had a delayed APC in combination with the postconditioning group, we may have seen an additive benefit over preconditioning or postconditioning alone. We did not measure  $\left[Ca^{2+}\right]_{m}$  in this study; however, studies have showed that increased  $[Ca^{2+}]$ <sub>i</sub> correlates with an increased  $\left[\text{Ca}^{2+}\right]_{\text{m}}$  (6,31) Finally, our results are specific to our experimental design with a 25-min period of ischemia and a 60-min reperfusion. Whether these findings can be extrapolated to humans is not known. We did not assess long-term outcome, which may be completely different despite initial similar IS. Perhaps, a more prolonged ischemic insult would show that there is no difference between sevoflurane pre- and postconditioning and that a longer perfusion time, such as 90 min or longer will have a clear demarcation of necrosis. We only used one sevoflurane concentration; a higher concentration (1.5 MAC) experiment could show a different result when used in a combined pre-and postconditioining protocol.

In conclusion, this study demonstrates that sevoflurane administration during the early minutes of reperfusion results in preservation of myocardial function and decreased IS by attenuating I/R injury. Our data show that no additional benefit is obtained when sevoflurane preconditioning and postconditioning are combined over either intervention alone. Sevoflurane postconditioning may be clinically applicable in situations where the potential for I/R injury is of concern.

#### **REFERENCES**

- 1. Murry CE, Jennings RB, Reimer KA. Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium. Circulation 1986;74:1124 –36
- 2. Zaugg M, Lucchinetti E, Uecker M, Pasch T, Schaub MC. Anaesthetics and cardiac preconditioning, Part 1: Signaling and cytoprotective mechanisms. Br J Anaesth 2003;91:551– 65
- 3. Kersten JR, Schmeling TJ, Pagel PS, Gross GJ, Warltier DC. Isoflurane mimics ischemic preconditioning via activation of K(ATP) channels: reduction of myocardial infarct size with an acute memory phase. Anesthesiology 1997;87:361–70
- 4. Chiari PC, Pagel PS, Tanaka K, Krolikowski JG, Ludwig LM, Trillo RA, Puri N, Kersten JR, Warltier DC. Intravenous emulsified halogenated anesthetics produce acute and delayed preconditioning against myocardial infarction in rabbits. Anesthesiology 2004;101:1160-6
- 5. Lutz M, Liu H. Inhaled sevoflurane produces better delayed myocardial protection at 48 versus. 24 hours post exposure. Anesth Analg 2006;102:984 –90
- 6. Steenberg C, Fralix TA, Murphy E. Role of increased cytosolic free calcium concentration in myocardial ischemic injury. Basic Res Cardiol 1993;88:456 –70
- 7. Piper HM. Energy deficiency, calcium overload or oxidative stress: possible causes of irreversible ischemic myocardial injury. Klin Wochenschr 1989;67:465–76
- 8. Zaugg M, Lucchinetti E, Spahn DR, Pasch T, Schaub MC. Volatile anesthetics mimic cardiac preconditioning by priming the activation of mitochondrial K(ATP) channels via multiple signaling pathways. Anesthesiology 2002;97:4 –14
- 9. Wang L, Cheredinichenko G, Hernandez L, Halow J, Camacho SA, Figueredo V, Schaefer S. Preconditioning limits mitochondrial  $Ca(2+)$  during ischemia in ret hearts: role of  $K(ATP)$ channels. Am J Physiol Heart Circ Physiol 2001;280:H2321– 8
- 10. Vinten-Johansen J, Zhao ZQ, Zatta AJ, Halkos ME, Kerendi F. Postconditioning a new link in nature's armor against myocardial ischemia-reperfusion injury. Basic Res Cardiol 2005;100:295–310
- 11. Feng J, Lucchinetti E, Ahuja P, Pasch T, Perriard JC, Zaugg M. Isoflurane postconditioning prevents opening of the mitochondrial permeability transition pore through inhibition of glycogen synthase kinase 3β. Anesthesiology 2005;103:987–95
- 12. Lucchinetti E, da Silva R, Pasch T, Schaub MC, Zaugg M. Anaesthetic preconditioning but not postconditioning prevents early activation of the deleterious cardiac remodeling programme: evidence of opposing genomic responses in cardioprotection by preand postconditioning. Br J Anaesth 2005;95:140 –52
- 13. Obal D, Dettwiler S, Favoccia C, Scharbatke H, Preckel B, Schlack W. The influence of mitochondrial K(ATP)-channels in the cardioprotection of preconditioning and postconditioning by sevoflurane in the rat in vivo. Anesth Analg 2005;101:1252– 60
- 14. Chiari PC, Bienengraeber MW, Pagel PS, Krolikowski JG, Kersten JR, Warltier DC. Isoflurane protects against myocardial infarction during early reperfusion by activation of phosphatidylinositol-3-kinase signal transduction: evidence for anesthetic-induced postconditioning in rabbits. Anesthesiology 2005;102:102–9
- 15. Sniecinski R, Liu H. Reduced efficacy of volatile anesthetic preconditioning with advanced age in isolated rat myocardium. Anesthesiology 2004;100:589 –97
- 16. Mangano DT. Perioperative cardiac morbidity. Anesthesiology 1990;72:153– 84
- 17. Staat P, Rioufol G, Piot C, Cottin Y, Cung TT, L'Huillier I, Aupetit J, Bonnefoy E, Finet G, André-Fouët X, Ovize M. Postconditioning the human heart. Circulation 2005;112:2143– 8
- 18. Zaugg M, Lucchinetti E, Garcia C, Pasch T, Spahn DR, Schaub MC. Anaesthetics and cardiac preconditioning, Part 2: Clinical implications. Br J Anaesth 2003;91:566 –76
- 19. Halkos ME, Kerendi F, Covera JS, Wang NP, Kin H, Payne CS, Sun HY, Guyton RA, Vinten-Johansen J, Zhao ZQ. Myocardial protection with postconditioning is not enhanced by ischemic preconditioning. Ann Thorac Surg 2004;78:961–9
- 20. DeHert SG, Broecke PW, Mertens E, Van Sommeren EW, De Blier IG, Stockman BA, Rogrigus IE. Sevoflurane but not propofol preserves myocardial function in coronary surgery patients. Anesthesiology 2002;97:42–9
- 21. Li F, Hayes JK, Wong KC, Szakacs J. Administration of sevoflurane and isoflurane prior to prolonged global ischemia improves heart function in isolated rat heart. Acta Anaesthesiol Sin 2000;38:113–21
- 22. Tani M, Neely J. Role of intracellular  $Na+$  and  $Ca+$  overload and depressed recovery of ventricular function in reperfused ischemic rat hearts. Circ Res 1989;65:1045–56
- 23. Piper HM, Garcia-Dorado D. Prime cause of rapid myocytes death during reperfusion. Ann Thorac Surg 1999;68:1913–19
- 24. Piper HM, Garcia-Dorado D, Ovize M. A fresh look at reperfusion injury. Cardiovasc Res 1998;38:291–300
- 25. Dennis SC, Gevers W, Opie LH. Protons in ischemia: where do they come from; where do they go? J Mol Cell Cardiol 1991;23: 1077– 86
- 26. Gabel SA, Cross HR, London RE, Steenbergen C, Murphy E. Decreased intracellular pH is not due to increased  $H+$  extrusion in preconditioned rat hearts. Am J Physiol 1997;273:H2257– 62
- 27. Di Lisa F, Bernardi P. Mitochondrial function as a determinant of recovery or death in cell response to injury. Mol Cell Biochem 1998;184:379 –91
- 28. Piper HM, Schafer AC. The first minutes of reperfusion: a window of opportunity for cardioprotection. Cardiovasc Res 2004;61:365–71
- 29. Schlack W, Preckel B, Stunneck D, Thamer V. Effects of halothene, enflurane, isoflurane, sevoflurane, and desflurane on myocardial reperfusion injury in isolated rat heart. Br J Anaesth 1998;81:913–19
- 30. Obal D, Preckel B, Scharbatke H, Müllenheim J, Höterkes F, Thämer V, Schlack W. One MAC of sevoflurane provides protection against reperfusion injury in the rat heart in vivo. Br J Anaesth 2001;87:905–11
- 31. Varadarajan SG, An J, Novalija E, Smart SC, Stowe DF. Changes in  $[Na+]<sub>i</sub>$ , compartmental  $[Ca<sup>2+</sup>]$ , and NADH with dysfunction after global ischemia in intact hearts. Am J Physiol Heart Circ Physiol 2001;280:H280 –93