

UC Davis

UC Davis Previously Published Works

Title

Methane Transport during a Controlled Release in the Vadose Zone

Permalink

<https://escholarship.org/uc/item/3jr2b656>

Journal

Vadose Zone Journal, 17(1)

ISSN

1539-1663

Authors

Felice, Mark
de Sieyes, Nick
Peng, Juan
et al.

Publication Date

2018

DOI

10.2136/vzj2018.02.0028

Peer reviewed

Original Research

Core Ideas

- Vadose zone CH₄ migration was densely monitored during a controlled-release scenario.
- Soil moisture strongly influenced vadose zone CH₄ fate and CH₄ oxidation.
- Soil CH₄ fate appears primarily controlled by biological CH₄ oxidation.

M. Felice, N. de Sieyes, J. Peng, R. Schmidt, M. Buelow, K. Scow, and D. Mackay, Dep. of Land, Air, and Water Resources, Univ. of California, One Shields Ave., Davis, CA 95616; P. Jourabchi, Golder Associates Ltd., Suite 200, 2920 Virtual Way, Vancouver, BC, V5M 0C4 Canada. *Corresponding author (mfelice@ucdavis.edu).

Received 1 Feb. 2018.

Accepted 13 May 2018.

Supplemental material online.

Citation: Felice, M., N. de Sieyes, J. Peng, R. Schmidt, M. Buelow, P. Jourabchi, K. Scow, and D. Mackay. 2018. Methane transport during a controlled release in the vadose zone. *Vadose Zone J.* 17:180028. doi:10.2136/vzj2018.02.0028

© Soil Science Society of America.
This is an open access article distributed under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Methane Transport during a Controlled Release in the Vadose Zone

Mark Felice,* Nick de Sieyes, Juan Peng, Radomir Schmidt, Maya Buelow, Parisa Jourabchi, Kate Scow, and Douglas Mackay

Shallow, small-rate releases of ethanol-blended fuels from underground storage tanks (USTs) may be quite common and result in subsurface CH₄ generation. However, vadose zone transport of CH₄ generated from these fuel releases is poorly understood, despite the potential to promote vapor intrusion or create explosion hazards. In this study, we simulated shallow CH₄ generation with a controlled subsurface CH₄ release from July 2014 to February 2015 to characterize subsurface CH₄ migration and surface emissions and to determine environmental controls on CH₄ fate and transport. July 2014 through November 2014 was an extended period of drought followed by precipitation during December 2014. Throughout the experiment, under varied CH₄ injection rates, CH₄ formed a radially symmetrical plume around the injection point. Surface efflux during the drought period of the experiment was relatively high and stable, with approximately 10 to 11 and 34 to 52% of injected CH₄ reaching the ground surface during the low- and high-rate injections, respectively. Following the period of precipitation and increased soil moisture, efflux dropped and stabilized at approximately 1% of injected CH₄, even as soil moisture began to decrease again. Tracer and inhibitor experiments and estimates of soil diffusivity suggest that microbial CH₄ oxidation was responsible for the observed drop in efflux. The decrease in efflux only after soil moisture increased suggests a strong environmental control over the transport and oxidation of vadose zone CH₄.

Abbreviations: BGS, below ground surface; DFE, difluoroethane; PVC, polyvinyl chloride; TDR, time domain reflectometry; UST, underground storage tank.

Despite concerns about CH₄ generation at petroleum-hydrocarbon-contaminated sites (Jewell and Wilson, 2011; Ma et al., 2014; Rasa et al., 2013; Sihota et al., 2013; Spalding et al., 2011; Wilson et al., 2012), many aspects of CH₄ transport in vadose zone soils are poorly understood, particularly in the case of small-rate, shallow gasoline vapor releases. While increased regulation of underground storage tank (UST) construction and installation has resulted in fewer large releases of liquid fuel, small-rate gasoline vapor releases still occur and often go undetected and overlooked due to their smaller release rates. The result is a limited understanding of the environmental and health risks associated with small-rate releases of these fuels and their degradation products, such as CH₄.

Large crude oil and fuel releases to groundwater are well studied, but these differ substantially from small-rate shallow fuel vapor releases. Small-rate shallow fuel vapor releases present a potential point source of CH₄ generation in the vadose zone just below the ground surface, in contrast to more commonly studied scenarios where CH₄ is generated in groundwater contaminant plumes. Small-rate fuel vapor releases may be quite common. For example, a survey of California USTs found that >60% of surveyed tanks were leaking fuel at rates undetectable by the leak detection technology at that time (highest rates ranged from 7.6×10^{-4} to 1.5 L d^{-1} of liquid fuel, with others were at or below $7.6 \times 10^{-4} \text{ L d}^{-1}$) (Young and Golding, 2002). In these cases, leaks typically occur due to pressure fluctuations during fuel dispensing or tank refilling, and occur at fittings on the top of the UST, often at fill-vent risers. This places the leaks at very shallow depth, approximately 1 m below the ground surface (Young and Golding, 2002).

Methane generation may occur anywhere that organic compounds are degraded under anaerobic conditions and frequently occurs at fuel-contaminated sites due to high C inputs from fuel. Fuel stored in USTs is frequently biofuel, which can be up to 95% ethanol or gasoline blended with ethanol (10–85%). Ethanol is a highly labile C source that can be rapidly degraded, depleting O₂ and exacerbating the generation of CH₄ (Ma et al., 2013; Powers et al., 2001).

Methane may have several negative health and safety impacts at the local scale: the generated CH₄ may (i) pose an asphyxiation or explosion hazard under a limited set of circumstances if it becomes trapped and accumulates to sufficient concentration in a structure or enclosed space overlying the contaminated zone (Jewell and Wilson, 2011; Ma et al., 2014; Nelson et al., 2010; Sihota et al., 2013; Wilson et al., 2012), (ii) increase the risk of vapor intrusion by creating pressure gradients that drive advective transport of toxic fuel constituents (Ma et al., 2014; Sihota et al., 2013; Wilson et al., 2012), and (iii) create higher O₂ demand that will deplete O₂ otherwise available for degradation of fuel constituents (Jewell and Wilson, 2011; Ma et al., 2012, 2014; Sihota et al., 2013; Wilson et al., 2012). These effects may be exacerbated by the shallow depth of small-rate vapor releases, which allows less residence time for the degradation of CH₄ or volatile organic compounds during migration to surface or subsurface structures. Numerical model simulations of vapor intrusion from a benzene source below a basement or slab-on-grade building have identified depth as one important factor in controlling benzene vapor intrusion risk (Abreu and Johnson, 2006). In addition to the potential negative health and safety impacts, CH₄ is a potent greenhouse gas contributing to climate change, with a global warming potential approximately 28 times higher than CO₂, and trailing only CO₂ in radiative forcing (IPCC, 2013).

Methane transport through the vadose zone occurs by advection and/or diffusion in the vapor phase, attenuated by processes that transfer mass from vapor to other phases or that degrade CH₄. For small-rate fuel releases, it is likely that diffusion is the dominant gas transport mechanism (Johnson et al., 2006; Lundegard and Johnson, 2006; Ma et al., 2014; Sihota et al., 2013). Soil physical properties such as porosity and moisture status strongly influence diffusive transport processes but also influence biological CH₄ oxidation, an important control on CH₄ fate (Gebert et al., 2011).

Methane-oxidizing bacteria (methanotrophs) are ubiquitous in soils and can oxidize CH₄ to CO₂. However, CH₄ oxidation kinetics are influenced by the microbial communities present (Aronson et al., 2013) and their interaction with environmental factors, such as soil moisture (Bender and Conrad, 1995; Gullledge and Schimel, 1998; van den Pol-van Dasselaar et al., 1998), soil structure (Hütsch, 1998), and soil N levels (Maxfield et al., 2008; Seghers et al., 2003). Because methanotrophs rely on gas transport to supply their C source, they are particularly sensitive to changes in soil moisture and soil structure, with a lower optimum soil moisture content than many other bacteria, also preferring coarser

textured and higher porosity soils (Boeckx et al., 1997; Dörr et al., 1993; Gebert et al., 2011). Given the effect of soil physical properties and methanotroph community structure, CH₄ oxidation efficiency varies widely, with soils in some studies showing almost no CH₄ attenuation while others show complete oxidation of CH₄ prior to reaching the ground surface (Börjesson et al., 2007; Chanton et al., 2009; Le Mer and Roger, 2001; Sass et al., 1992). For example, a review of landfill CH₄ emission studies reported that, on average, studies found that 36% of the CH₄ produced was oxidized before reaching the atmosphere, but values varied from net CH₄ consumption to no CH₄ oxidation (Chanton et al., 2009). A study measuring CH₄ emissions and production in rice (*Oryza sativa* L.) paddies found that an average of 73% of CH₄ produced was oxidized, but again, the numbers were highly variable, ranging from 0 to >90% (Sass et al., 1992).

A better understanding of CH₄ fate and transport in the shallow vadose zone is essential for assessing the risk posed by CH₄ associated with shallow small-rate ethanol-blended fuel releases. A number of studies have addressed shallow subsurface transport of CH₄ originating from leaking natural gas lines (Okamoto and Gomi, 2011; Yan et al., 2015). However, these experiments simulated significantly higher CH₄ release rates than expected CH₄ generation rates at small-rate ethanol-blended fuel releases, and very few studies have examined the impact of soil and environmental properties on CH₄ fate and transport at the field scale under controlled conditions, which is a unique opportunity afforded by our study. The objectives of this study were to: (i) characterize subsurface CH₄ migration and transformation from a controlled field release at shallow depths, including the areal extent of the CH₄ plume; (ii) measure the lag time for CH₄ oxidation to begin; (iii) determine the fraction of subsurface CH₄ reaching the atmosphere; and (iv) measure seasonal variability and identify environmental controls of CH₄ migration. We hypothesized that biological CH₄ oxidation would begin within several days of introducing CH₄ to the soil and that this biological activity would limit the efflux of CH₄ to the atmosphere. We also hypothesized that CH₄ migration would be tied to soil moisture, with decreased efflux during wet periods.

Materials and Methods

Site Characterization

The CH₄ injection experiment was conducted at a field site located in the Putah Creek Riparian Reserve, approximately 2.25 km south of the University of California campus at Davis. The soil is mapped as riverwash (Soil Survey Staff, 2017), with a measured texture of silt loam in the upper 0.25 m of soil and from approximately 0.95 to 1.5 m below ground surface (BGS), the maximum depth of monitoring. The texture is silt from approximately 0.25 to 0.95 m BGS. Weighted averages ± standard deviations for solid density, bulk density, and porosity are 2.66 ± 0.04 g cm⁻³, 1.38 ± 0.14 g cm⁻³, and 0.48 ± 0.05, respectively. See Supplemental Material for complete site characterization.

Experimental Methods

Injection and Monitoring Network

As shown in Fig. 1, during the experiment, the site was instrumented with an array of narrow-diameter, custom-built, stainless steel drive points installed at 0.5, 1.0, and 1.5 m BGS to allow gas injection and sampling of soil gas, and an array of polyvinyl chloride (PVC) collars for the measurement of surface gas efflux with a portable gas analyzer described below. The site was also instrumented with temperature sensors installed at regular depth intervals throughout the soil profile (details below).

Gas Injection

Gas was injected in the center of the monitoring network at the 1-m depth. The injection initially alternated between 1000 mL d⁻¹ of a mix of 25% CH₄ and 75% Ne and 1000 mL d⁻¹ of pure CH₄ to observe CH₄ transport under different injection rates. The Ne was initially included as a conservative tracer but was found to be below method detection limits in soil gas and surface efflux when injected at the rates described above. The experiment also included periods during which 1,1-difluoroethane (1,1-DFE) was injected as a conservative tracer or ethene was injected as a methanotroph inhibitor to help identify the role of biological CH₄ oxidation. Injection periods are summarized in Table 1. The 25 and 100% CH₄ injections are equivalent to the CH₄ generated from the degradation of 3.8 × 10⁻³ and 1.5 × 10⁻² L d⁻¹, respectively, of liquid E10 gasoline blend releases, assuming complete conversion of all ethanol to CH₄. Typical low-rate fuel releases documented in a field survey of California USTs fell between 7.6 × 10⁻⁴ and 1.5 L d⁻¹ (Young and Golding, 2002).

Soil Sampling

Cores extending to 1.5 m BGS were collected throughout the experiment for moisture content analysis. Pre-release samples were also used for bulk density measurements. The pre-release cores were collected volumetrically by driving a 1.7-cm i.d. and 1.9-cm o.d. aluminum tube into the soil at 7.5-cm intervals, except for the first interval, which was 3.8 cm. To reduce compaction and

improve recovery, a pilot hole for the aluminum tube was produced using a hand auger for samples taken below 26.3 cm. The tubing was trimmed to remove soil from the overlying layers that may have contaminated the sample. Subsequent samples not used for bulk density analysis were collected directly with a 5-cm-diameter auger. All samples were sealed in plastic bags and placed on ice, then stored at 4°C until further processing. Throughout the experiment, sampling events included samples collected adjacent to the injection point and in randomly selected background locations outside of the area with detectable soil gas CH₄ and CH₄ efflux. On the final two sampling events (16–17 Dec. 2014 and 28 Jan. 2015), a transect was sampled including four points extending from directly adjacent to the injection point to approximately 2.4 m distal to the injection point.

Soil Analysis

Soils collected during soil sampling events were analyzed for particle size distribution, mineral density, bulk density, organic C content, and moisture retention characteristics. A Beckman Coulter LS 230 laser diffraction particle size analyzer was used to analyze the distribution of particle sizes from Core B11, the closest core to the experimental plot. The top 1.5 m of the core was subsampled in triplicate at 9-cm intervals. Each triplicate subsample was then itself analyzed in triplicate, and the results for all runs of all replicates were averaged for analysis.

Organic C content and moisture retention curves for soils sampled at three depths between the surface and 1.5 m were analyzed by the University of California–Davis Analytical Laboratory using loss-on-ignition (Nelson and Sommers, 1996, p. 1004) and pressure plate (10, 33, 100, 500, 1000, and 1500 kPa; Klute, 1986) methods, respectively.

Efflux Monitoring

Real-time surficial monitoring of CO₂ and CH₄ efflux was conducted using a LI-COR LI-8100A automated soil CO₂ flux system (LI-COR Biosciences) connected to a Los Gatos Research Ultraportable Greenhouse Gas Analyzer. Efflux monitoring

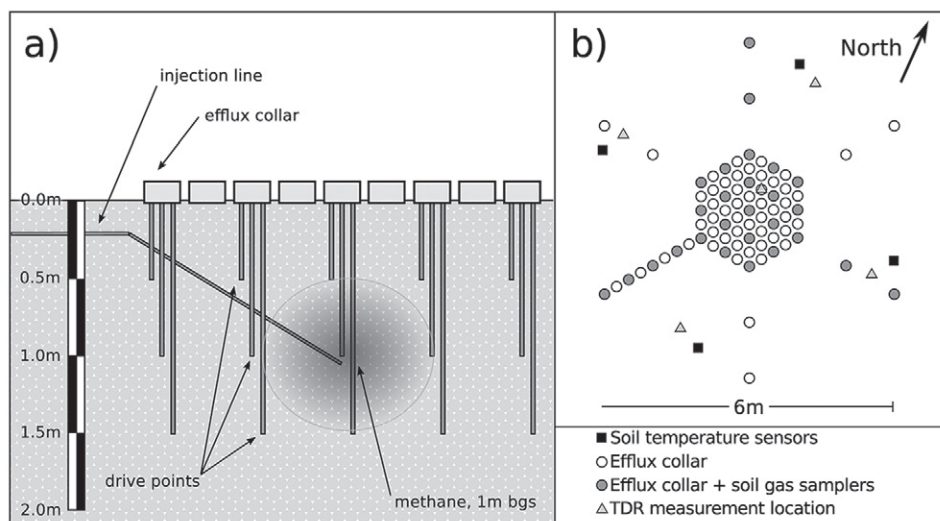


Fig. 1. (a) Vertical cross-section and (b) plan view schematics of the CH₄ injection and monitoring network.

Table 1. Methane injection periods and composition of injected gas throughout the experiment. The gas mixture was injected at a target rate of 1000 mL d⁻¹ during all injection periods.

Injection period	Gas composition	Injection dates	Injection duration
			d
First low-rate CH ₄ injection	25% CH ₄ , 75% Ne	7–22 July 2014	15
First high-rate CH ₄ injection	100% CH ₄	23 July–19 Sept. 2014	58
Second low-rate CH ₄ injection	25% CH ₄ , 75% Ne	19 Sept.–17 Oct. 2014	28
Second high-rate CH ₄ injection	100% CH ₄	17 Oct. 2014–3 Feb. 2015	109
Conservative tracer injection	100% 1,1-DFE†	3–27 Feb. 2015	24
CH ₄ /tracer co-injection	50% CH ₄ , 50% 1,1-DFE	27 Feb.–13 Mar. 2015	14
CH ₄ /CH ₄ -oxidation inhibitor co-injection	50% CH ₄ , 50% ethene	13–23 Mar. 2015	10

† DFE, difluoroethane.

locations were established using a grid of concentric hexagonal rings centered on the ground surface directly above the gas release location and extending out to a radius of 104 cm, as depicted in Fig. 1. A less dense array of collars was installed along the 60° “arms” of the network outside of the densely monitored central zone.

Soil Gas Monitoring

Custom soil gas samplers were manufactured from 6.4-mm o.d. 316 stainless steel straight tubing. The ends were sealed with stainless screw nails, and side ports were drilled into a vertical groove approximately 2 cm from the sealed end to minimize clogging during installation. At all locations, clusters of samplers were hammered by hand to depths of approximately 0.5, 1.0, and 1.5 m BGS; the locations of all clusters are shown in Fig. 1. At the release point, additional points were installed to approximately 0.25 and 1.75 m BGS. An additional drive point for releasing gas into the subsurface was driven diagonally to 1.0 m BGS directly below the central efflux collar. The tops of all drive points were finished with gas-tight polypropylene valves and stainless steel quick-connect fittings.

The sampling of drive points was conducted using a custom hand vacuum pump system (see the Supplemental Material). This hand pump system was used to purge 80 mL of gas, equivalent to more than three sampler volumes, from each sampler. During sampling, needled inlets and outlets were inserted through the septa on a 20-mL sample vial, and the gas flow from the sampler was routed through the inlet–outlet, thereby purging the vial with soil gas. Once 80 mL of soil gas had been purged through the vial, the inlet and outlet needles were removed and the soil gas sample was labeled, stored in the dark, and returned to the laboratory for analysis.

Samples were run on an Agilent 6890N gas chromatograph outfitted with an Agilent CP7429 column and both a thermal conductivity detector (TCD) and a flame ionization detector (FID) connected to an Agilent 7694 headspace autosampler (Agilent Technologies). Concentrations of O₂, N₂, Ne, and CO₂ were analyzed using the TCD, while ethene, 1,1-DFE, and CH₄ were analyzed using the FID. The detection limits for O₂, CO₂, ethene, 1,1-DFE, and CH₄ were 1.5, 0.0075, 0.00098, 0.00050, and 0.00016%, respectively.

Temperature Monitoring

Soil temperature was logged using five strings of iButton temperature loggers (Maxim Integrated) installed into PVC casings installed at the site (Fig. 1b). Temperature sensors in the four strings at the perimeter of the site were spaced at 34, 64, 94, 156, and 224 cm BGS. Temperature sensors in the string near the center of the site were spaced at 17, 37, 56, 79, 99, 119, 140, 161, 180, 200, 221, and 230 cm BGS.

Moisture Monitoring

Moisture content was determined gravimetrically for all depths sampled during soil sampling events throughout the duration of the experiment by loss of moisture after oven drying 5- to 15-g subsamples of soil for at least 48 h. Starting in September 2014, the moisture content of surficial sediments to 0.1 m BGS was analyzed in the field every 4 d using a hand-held GS3 time-domain reflectometry (TDR) probe with a ProCheck handheld reader (Decagon Devices). For the TDR measurements, a custom calibration curve of probe response vs. soil moisture content was created using site sediments, pursuant to manufacturer instructions. Surface soil measurements were made routinely at one location inside the central efflux monitoring network and four locations adjacent to temperature loggers (Fig. 1b).

Results

Soil Moisture and Precipitation

No precipitation fell from the beginning of the experiment (July 2014) until 24 Sept. 2014 (Fig. 2), during which time soil moisture at 2 cm decreased (Fig. 3). At 100 and 145 cm, soil moisture increased from April to June 2014 and decreased in September. Precipitation intensity and frequency increased significantly and peaked from the end of November until the end of December, which was accompanied by increases in soil moisture. Moisture at 2 cm was temporally coupled with precipitation, peaking in December as precipitation peaked. Changes in moisture at depth were delayed relative to precipitation events, with moisture at 100 and 145 cm peaking at the end of the experiment in February 2015.

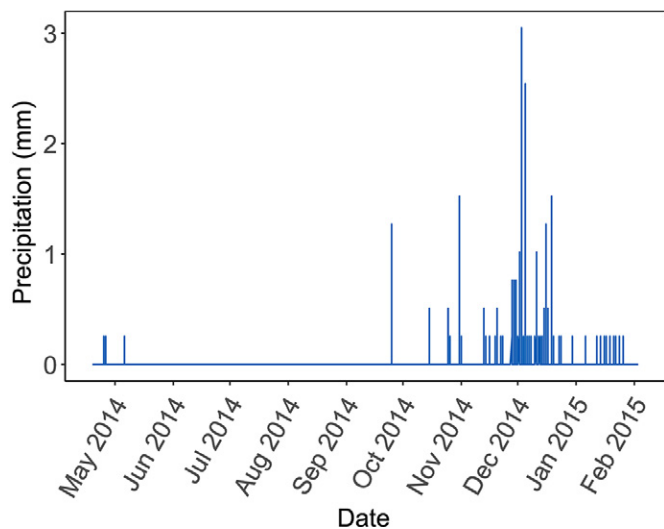


Fig. 2. Precipitation events between the first pre-experimental soil sampling (9 Apr. 2014) and conclusion of the final 100% CH₄ injection (3 Feb. 2015). Data were obtained from the weather station at the Campbell Tract, Davis, CA, approximately 5.2 km (3.2) miles north-west of the experiment site.

Minimum, maximum, and average soil moisture content at these depths are shown in Table 2. Monitoring of surficial soils by TDR probe began in September 2014 and showed similar trends to the soil sampling (Fig. 3).

Soil Gas: Methane

At monitoring points located away from the injection point, elevated CH₄ was detected in the soil gas shortly after the start of injection and responded rapidly to changes in injection rate (Fig. 4). This implies that CH₄ migration was very rapid. The highest CH₄ concentrations were measured at the injection point, with concentrations decreasing radially. Methane was regularly present at low concentrations above the method detection limit as far as 3 m from the injection point. Soil gas profiles from the low- and high-rate injection periods are shown in Fig. 5.

Methane concentrations appeared to be stable during the first 25 and 100% CH₄ injection periods (Fig. 4). Methane concentrations were also stable during the second 25% injection period. The average CH₄ concentration at the injection point was 0.024 ± 0.003 and $0.032 \pm 0.005\%$ during the two 25% CH₄ injection periods, and 0.012 ± 0.013 and $0.19 \pm 0.019\%$ during the 100% CH₄ injection periods. Based on average soil gas CH₄ concentrations from each injection period, the fourfold higher injection rate

Table 2. Range in volumetric soil moisture content measured at various depths during the experiment. Moisture content was measured by oven drying.

Depth	Min.	Max.	Avg.
cm	%		
2	2.8	36.0	13.3
100	5.2	27.5	14.3
145	5.5	18.3	14.8

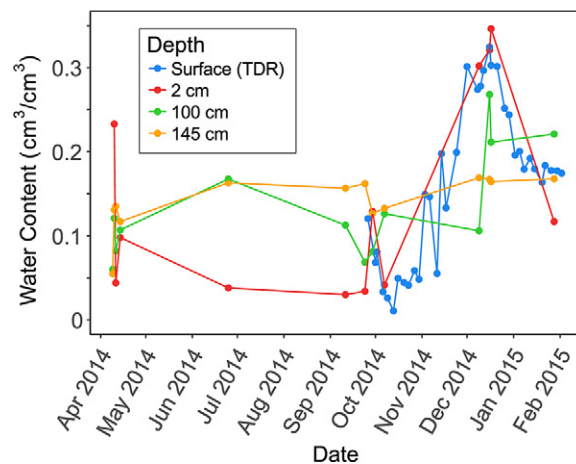


Fig. 3. Volumetric soil water content measured between the first pre-experimental soil sampling (9 Apr. 2014) and conclusion of the final 100% CH₄ injection (3 Feb. 2015). Water content at 2, 100, and 145 cm was determined by oven drying. Water content at the soil surface was determined from the average time domain reflectometry (TDR) probe reading from five sampling locations.

initially resulted in a 2.9- to 7.3-fold increase in vapor concentrations above and below the injection point. The greatest vapor concentration increases occurred at the 0.25- and 1-m depths.

While soil gas CH₄ concentrations remained relatively stable throughout most of the experiment, they began to change in November 2014 during the final 100% CH₄ injection period. The trends in soil gas CH₄ concentration varied depending on depth and lateral distance from the injection point. All sampling locations distal to the injection point showed a similar temporal pattern, where concentrations increased and peaked in early December 2014 followed by declining concentrations until the end of the experiment. The change in surface moisture content (TDR) also peaked in early December and then declined until the end of the experiment (Fig. 3). However, at the injection point, soil gas CH₄ concentrations rose steadily until the end of the experiment,

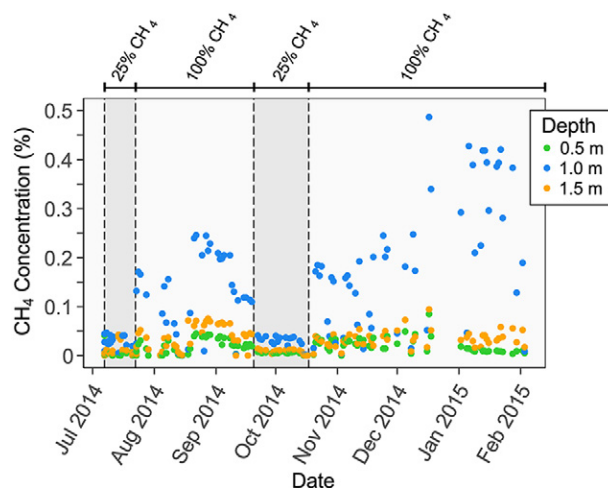


Fig. 4. Subsurface CH₄ concentrations at the central monitoring point (Ring 0,0) throughout the CH₄ injection experiment (7 July 2014–3 Feb. 2015).

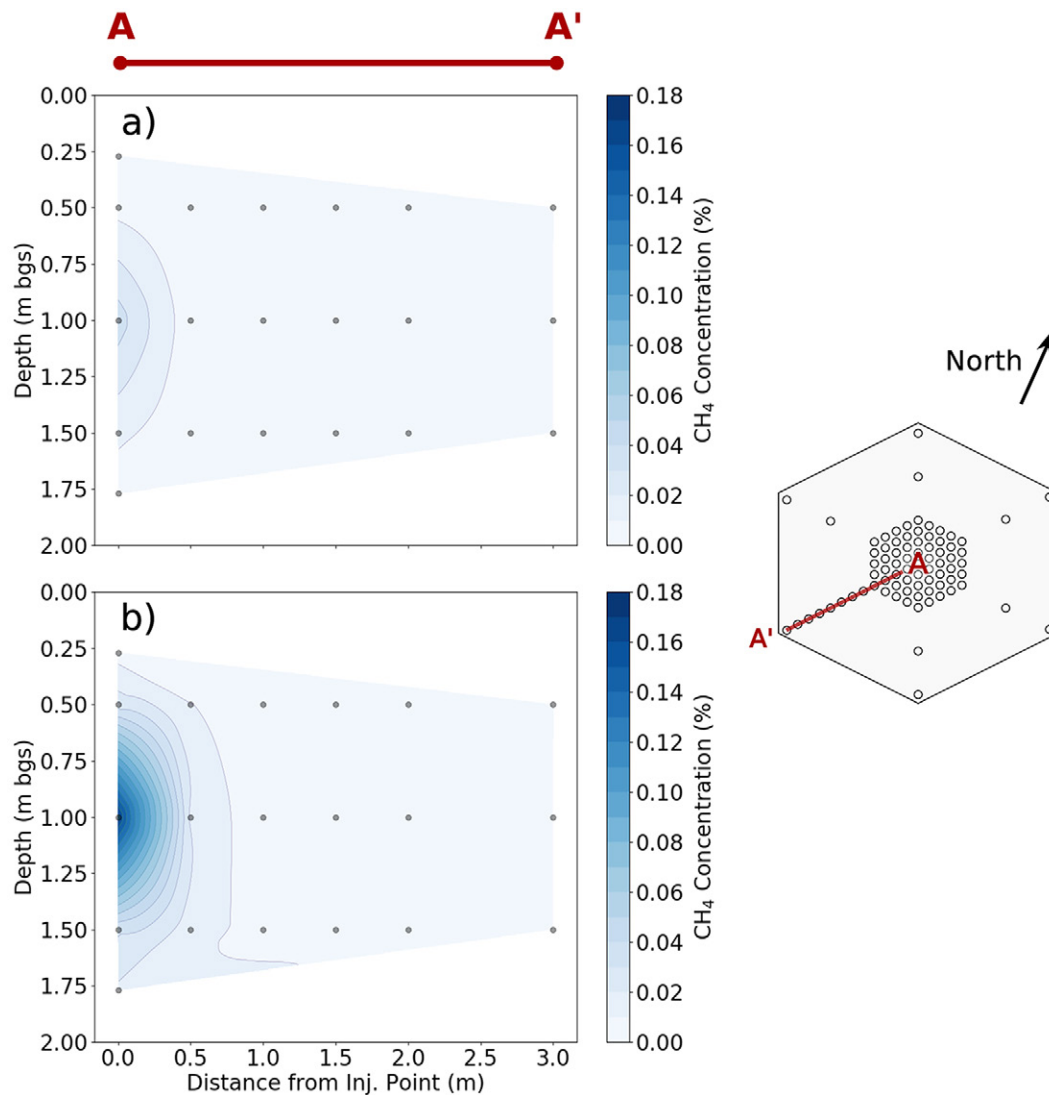


Fig. 5. Soil gas profiles during the second (a) 25% (8 Oct. 2017) and (b) 100% (23 Oct. 2017) CH₄ injection periods. Circles represent sampling points. Interpolations estimated by Delauney triangulation.

even as soil moisture began to decline (Fig. 4). During this period, the cross-sectional area of the plume increased.

Efflux

Prior to CH₄ injection, CH₄ efflux ranged from -0.0008 to $0.0006 \mu\text{mol m}^{-2} \text{s}^{-1}$ ($-0.0003 \mu\text{mol m}^{-2} \text{s}^{-1}$ average), indicating some net CH₄ emission from the soil to the atmosphere, but more frequently net movement of CH₄ from the atmosphere to the soil (Fig. 6). There was no clear pattern in the distribution of CH₄ efflux during this period. Methane was detected at the surface as far as 1 m from the injection point (the most distal monitoring point during the initial injection) within 1 d of beginning injection and reached steady state within 3 d of beginning injection or changing the injection mix (Fig. 7). Sitewide CH₄ efflux reflected the injection rate or mix, averaging 0.0018 and $0.0015 \mu\text{mol m}^{-2} \text{s}^{-1}$ during the first and second 25% CH₄ injection periods, respectively, and 0.0060 and $0.0065 \mu\text{mol m}^{-2} \text{s}^{-1}$ during the first and second 100% CH₄ injection periods. The

fraction of injected CH₄ reaching the surface as efflux was also correlated to the injection rate. An average of 11 ± 1 and $10 \pm 1\%$ of the injected CH₄ was detected as efflux during the two 25% CH₄ injection periods, while 52 ± 6 and $34 \pm 6\%$ was detected during the two 100% CH₄ injection periods. However, the fraction of efflux declined throughout the final 100% CH₄ injection period from $>50\%$ to $<2\%$.

Methane efflux was highest directly above the injection point, with rates decreasing radially (Fig. 6). Very low positive CH₄ efflux was periodically detected at 3 m from the injection point throughout most of the experiment, but efflux at these 3-m monitoring points fluctuated between low-magnitude positive and negative values. The areal extent of positive CH₄ efflux remained consistent throughout most of the final 100% CH₄ injection period, extending to the outer monitoring points approximately 1.5 m from the injection point. However, CH₄ efflux decreased dramatically at the end of the injection period in January 2015, with positive CH₄ efflux detected to approximately 0.5 m from the injection point

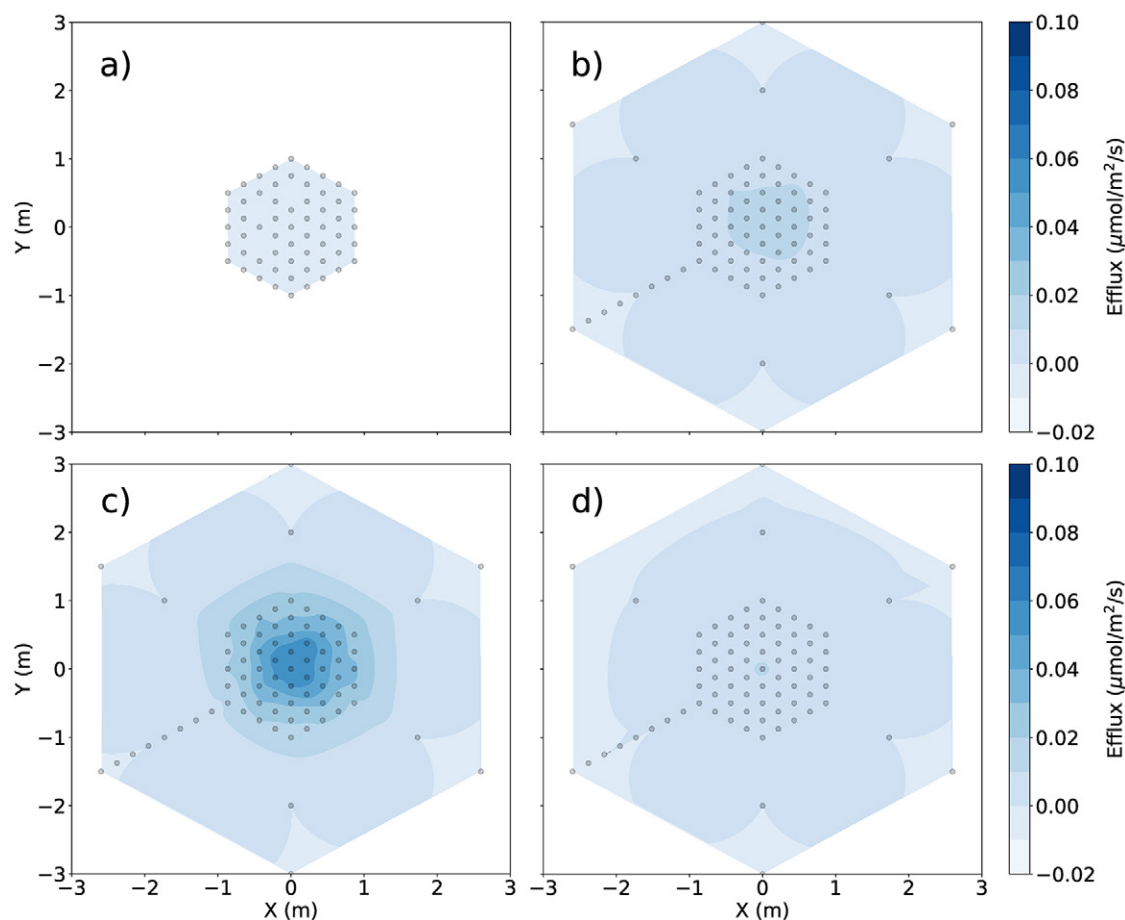


Fig. 6. Contour plots of efflux snapshots taken (a) prior to the start of CH_4 injection (30 Apr. 2014), (b) during 25% CH_4 injection (8 Oct. 2014), (c) during 100% CH_4 injection (11 Sept. 2014), and (d) during 100% CH_4 injection following a period of precipitation and declining efflux (15 Jan. 2015). Negative efflux indicates net consumption of CH_4 by the soil, and positive efflux indicates net emissions of CH_4 to the atmosphere. Sampling points are represented by circles. Contouring estimated using Delaunay triangulation.

and negative efflux detected in some areas. The temporal changes in CH_4 efflux during the final 100% CH_4 injection period followed a similar trend at all sampling locations (Fig. 7).

Carbon dioxide efflux did not form a discernible spatial pattern and did not correspond spatially to CH_4 efflux. Carbon dioxide did show significant seasonal variation, with efflux increasing during the latter stages of the experiment when soil moisture was high.

1,1 Difluoroethane and Ethene Injection

In February 2015, 1 wk of injecting 100% 1,1-difluoroethane (1,1-DFE) as a conservative tracer appeared to induce some inhibition of CH_4 oxidation. During the 100% 1,1-DFE injection, CH_4 efflux was higher above the injection point and decreased outward, switching from net CH_4 emission to net CH_4 consumption at the outer edges of the monitoring network (Supplemental Fig. S4). This contrasts with the pre-release efflux, which showed negative rates across the site, with no spatial pattern clearly associated with the CH_4 injection point.

When CH_4 was co-injected with an equal volume of ethene as a CH_4 -oxidation inhibitor or 1,1-DFE, efflux was not detectable, but

soil gas concentrations of CH_4 were lower than both ethene and 1,1-DFE concentrations at corresponding sampling locations, indicating that these gases were undergoing different processes. Soil gas CH_4 concentrations were approximately 20 and 61% of soil gas 1,1-DFE and ethene concentrations, respectively (Supplemental Fig. S5).

Discussion

Efflux: Methane

Negative efflux observed prior to CH_4 injection shows that this soil was a net CH_4 sink (Fig. 6a) and suggests an active community of CH_4 -oxidizing bacteria with similar oxidation rates to those observed in situ for grasslands in several other studies (0.000043 – $0.0010 \mu\text{mol m}^{-2} \text{s}^{-1}$) (Kammann et al., 2001; Rong et al., 2015; van den Pol-van Dassel et al., 1998). However, CH_4 consumption rates were lower than those observed in pine ($0.0032 \mu\text{mol m}^{-2} \text{s}^{-1}$) and hardwood ($0.0030 \mu\text{mol m}^{-2} \text{s}^{-1}$) forests (Gulledge et al., 2004). The positive CH_4 efflux observed shortly after the beginning of CH_4 injection shows that the existing methanotroph community was unable to fully oxidize the added CH_4 inputs.

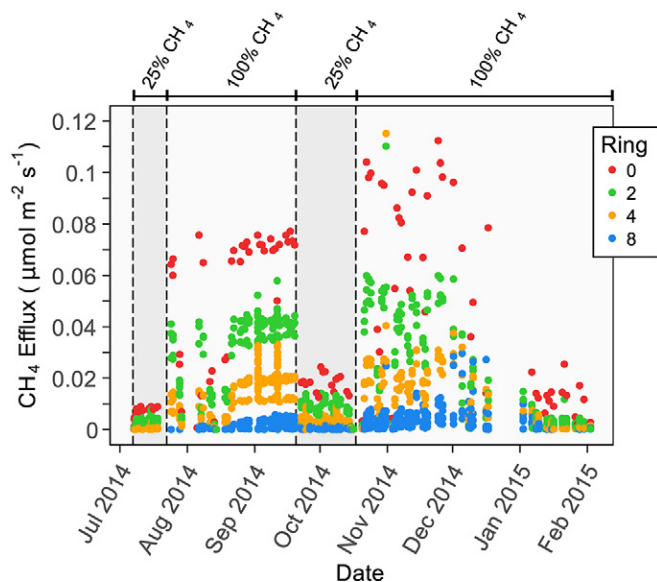


Fig. 7. Efflux throughout the duration of the experiment. Rings 2, 4, and 8 represent the efflux collars placed hexagonally around the injection point (Ring 0) with circumradii of 0.5, 1.0, and 2.0 m, respectively.

Methane efflux observed in this study during the injection period was within the range of CH_4 efflux detected at dryland sites with low to moderate levels of CH_4 seepage (Etiopie and Klusman, 2010) but lower than in studies of landfills, rice paddies, or catastrophic fuel releases with subsurface CH_4 generation (Table 3). The relative effluxes are consistent with relative CH_4 inputs, which in this experiment fall between the well-studied low inputs of atmospheric CH_4 and the high inputs of landfills, wetlands, and catastrophic fuel spills.

Despite the relatively low CH_4 inputs, the increase in CH_4 efflux relative to the injection rate suggests that the CH_4 -oxidizing capacity of the methanotroph community was already saturated at the lower 25% CH_4 injection rate under the conditions at that time and was unable to significantly increase the oxidation rate even with increased CH_4 inputs. Other researchers have observed the inability of methanotrophs to oxidize additional CH_4 in

proportion to increased inputs. A study of two landfills found that the percentage of generated CH_4 that was oxidized increased as CH_4 emissions decreased, fitting an exponential decay curve (Chanton et al., 2011). A similar exponential function related the fraction of CH_4 oxidized as a function of total CH_4 production in a modeling study of the effects of climate, landfill cover type, and CH_4 emission rates in California (Abichou et al., 2010). Another study, which examined landfill and volcanic pasture and forest soils in New Zealand, suggested that the specific methanotroph community was responsible for an observed decline in CH_4 oxidation efficiency as CH_4 concentrations increased. High-affinity Type II methanotrophs in the forest soil were present, and these soils showed a decline in CH_4 oxidation efficiency as CH_4 concentrations increased to relatively low concentrations of 0.004%, whereas landfill cover soils adapted to high CH_4 concentrations, and pasture soils dominated by Type I methanotrophs showed relatively constant or increasing CH_4 oxidation efficiency to concentrations as high as 0.75%. Kinetics suggest that a methanotroph population consisting of only 10% high-affinity methanotrophs could account for the observed trends (Tate et al., 2012).

Changes in CH_4 efflux under steady injection rates were related to the onset of precipitation and increased soil moisture content. During the dry periods, efflux rapidly reached steady state and remained stable. However, efflux began to decline in December 2014 following periods of precipitation and increased soil moisture content (Fig. 7). Moisture may have decreased efflux through two mechanisms: (i) directly through increasing the water-filled pore space and thus decreasing soil gas diffusivity; and (ii) increasing CH_4 oxidation activity by providing optimal conditions for methanotroph activity. Estimates of soil diffusivity under different moisture contents reveal the potential impact of soil moisture on physical transport processes.

The Millington–Quirk relationship can be used to estimate the effective diffusion coefficient in soil gas (D_s) from the total porosity (f), air-filled porosity (f_a), and the free-air diffusion coefficient (D_0):

$$D_s = \frac{f_a^{10/3}}{f^2} D_0 \quad [1]$$

Table 3. Comparison of efflux measured during this and other studies.

Efflux	Study setting	Reference
$\mu\text{mol m}^{-2} \text{s}^{-1}$		
0.49–9.3	landfill	Mønster et al. (2015)
0.44	cropland	Raich and Schlesinger (1992)
2	landfill	Czepiel et al. (1996)
0.19	rice paddy	Chen et al. (2013)
1.4	above denatured fuel-grade ethanol release	Sihota et al. (2013)
24	above denatured fuel-grade ethanol release	Sihota et al. (2013)
0.004–0.04	drylands with low-moderate microseepage	Etiopie and Klusman (2010)
0.003†	present study (25% CH_4 injection)	present study
0.011†	present study (100% CH_4 injection)	present study

† Efflux above the injection point averaged across all sampling dates at respective injection level.

Using this equation, the estimated effective diffusion coefficient for CH₄ dropped by a factor of 3.4 from the driest to the wettest period. Other field studies have shown this effect of moisture as well. For example, moisture content and air permeability were the most important parameters influencing CH₄ and CO₂ transport through soil from a landfill, based on sensitivity analysis of numerical transport models (Poulsen et al., 2001). However, we observed a drop in efflux as a fraction of injected CH₄ of approximately 25 times, a much larger decrease than predictions based solely on changes in soil gas diffusivity, which strongly suggests that physical transport processes are not the sole control on CH₄ efflux. Additional support for the hypothesis that microbial CH₄ oxidation is involved in reducing CH₄ efflux is the observation that the fraction of injected CH₄ measured as efflux remained consistently low through the end of the experiment, even as the soil began to dry out and diffusivity increased.

While we hypothesize that the increased moisture content following precipitation events relieved moisture stress, allowing higher CH₄ oxidation rates, theoretically, a drop in diffusivity could potentially reduce CH₄ oxidation rates if it sufficiently limited O₂ available to CH₄-oxidizing bacteria. However, optimal CH₄ oxidation rates are achieved when the moisture content is low enough to allow adequate CH₄ and O₂ diffusion but high enough to eliminate physiological moisture stress. The moisture content observed at our site during the period of decreased efflux fell within the optimal moisture content for CH₄ oxidation found by several studies (Bender and Conrad, 1995; Gulledge and Schimel, 1998; Schnell and King, 1996; van den Pol-van Dasselaar et al., 1998; Whalen and Reeburgh, 1996).

Efflux: Carbon Dioxide

Carbon dioxide efflux during this experiment was generally within or below the range of values for natural soil respiration reported in the literature. For example, a review of soil respiration studies found efflux of 1.16 μmol m⁻² s⁻¹ in temperate grasslands and 1.43 μmol m⁻² s⁻¹ in croplands (Raich and Schlesinger, 1992). Near denatured ethanol spills in southwestern Minnesota, natural soil respiration generated CO₂ efflux of 3.7 to 7 μmol m⁻² s⁻¹ (Sihota et al., 2013). A study of a petroleum pipeline rupture site found natural soil respiration contributing CO₂ efflux of 0.4 to 5.1 μmol m⁻² s⁻¹ (Sihota et al., 2016). Due to the relatively small C inputs from CH₄ in this study, changes in CO₂ efflux above natural soil respiration may have been difficult to detect. Assuming complete oxidation of all injected CH₄, 1000 mL d⁻¹ (0.47 μmol s⁻¹) of injected CH₄ could generate as much as 0.47 μmol s⁻¹ of CO₂. Assuming a uniform distribution of CO₂ efflux across an area of 1 to 12.6 m², which is the areal extent of the efflux monitoring network, CO₂ efflux from CH₄ oxidation should be in the range of 0.02 to 0.5 μmol m⁻² s⁻¹, which is much smaller than the natural spatial variation in CO₂ efflux observed at the site. For example, one initial pre-experiment monitoring found CO₂ efflux between 2.28 and 6.19 μmol m⁻² s⁻¹.

Therefore, the lack of a clear spatial relationship between CH₄ efflux and CO₂ in this study is not surprising.

Soil Gas: Methane

The subsurface CH₄ concentrations at the depths and distances distal to the injection point also tracked with soil moisture content, but unlike efflux, the soil CH₄ concentrations appeared to increase during the periods of highest precipitation and surface soil moisture, which is consistent with diffusion limitation caused by increasing water-filled pore space limiting gases from migrating away from the injection point. Interestingly, subsurface CH₄ concentrations at the injection point continued to increase and remained high at the end of the experiment, whereas CH₄ concentrations at distal points began to decrease again as the soil began to dry out near the end of the experiment. Because changes in soil gas diffusivity in a particular uniform soil should change only through changes in moisture content and water-filled pore space, we would expect the soil gas concentrations distal to the injection point to be inversely related to soil moisture content, which appears true throughout most of the experiment. However, the data appear inconsistent with this trend during the final month of the experiment, when soil gas concentrations begin to decline despite declining soil moisture content. This suggests that another mechanism, such as microbial CH₄ oxidation may also be involved.

During the co-injection of CH₄ and 1,1-DFE or ethene, 1,1-DFE and ethene concentrations were higher than corresponding CH₄ concentrations (Supplemental Fig. S5). We expected physical transport processes to have a similar effect on CH₄, 1,1-DFE, and ethene, so the lack of concordance in concentrations between CH₄ and these other two gases suggests biological CH₄ oxidation. Both 1,1-DFE and ethene are much more recalcitrant than CH₄. No data on 1,1-DFE biodegradation were available, and ethene degradation rates of 22.3 to 136 pmol g⁻¹ soil h⁻¹ with 0.002 to 0.005% headspace have been reported (Zechmeister-Boltenstern and Smith, 1998). The relative concentrations of CH₄ are higher in the ethene co-injection than the 1,1-DFE co-injection, which may be the result of some ethene degradation or slight inhibition of CH₄ oxidation. One study demonstrated complete methanotroph inhibition in a landfill cover soil at 0.1% ethene (Chan and Parkin, 2000), and another showed total inhibition of atmospheric CH₄ oxidation in volcanic forest soils at 0.002% (Xu and Inubushi, 2009), while ethene was present at concentrations ranging from below the detection limit to approximately 0.45% in this study. Positive CH₄ efflux during the 100% 1,1-DFE injection period compared with negative efflux during pre-experiment characterization suggests that 1,1-DFE inhibited atmospheric CH₄ oxidation. Although 1,1-DFE has not been reported as a methanotroph inhibitor, difluoromethane is an effective methanotroph inhibitor (Miller et al., 1998).

Conclusions

Methane migration occurred rapidly, with surface efflux and soil gas concentrations achieving a steady state within several

days of the start of or changes in injection rates. Soil moisture conditions appeared to control CH₄ efflux, which was probably a combination of biological CH₄ oxidation and reduced soil gas diffusivity due to increased water-filled pore space. While our study suggests a larger role for biological CH₄ oxidation, numerical model simulations could be used to further elucidate the controls on CH₄ fate and transport in this system.

This experiment was conducted in the context of assessing the risks posed by CH₄ generated in the vadose zone from the degradation of ethanol released during shallow small-rate fuel releases. Our CH₄ injection rate represented an intermediate severity of small-rate fuel release from a UST that would go undetected with standard leak detection systems (3.8×10^{-3} – 1.5×10^{-2} L d⁻¹). A survey of California USTs found fuel release rates between 7.6×10^{-4} and 1.5 L d⁻¹ (Young and Golding, 2002), and our CH₄ injection rate assumed that ethanol was the only fuel constituent broken down methanogenically, meaning that the rates estimated in this experiment may underestimate CH₄ generation at many small-rate fuel release sites. Higher ethanol blends of fuel, such as E85, are becoming more common and could also potentially increase CH₄ generation. However, further studies adding E10 or E85 fuel blends to the vadose zone are needed to more accurately estimate actual rates of CH₄ generation expected from shallow small-rate fuel releases and further elucidate the risks of small-rate shallow fuel releases.

Acknowledgments

Funding from the American Petroleum Institute (Contract no. 2014-108690-3), Chevron Technology Ventures (Contract no. 201500561-45), USDA–NRCS (Grant no. 69-3A75-12-249), and USDA National Institute of Food and Agriculture (Hatch Project CA-2122-H and multistate regional project W-2082). Approvals and other support from Sue Fields (UCD Environmental Health and Safety), Andrew Fulks (UC Davis Putah Creek Riparian Reserve), Brian Newman (California Regional Water Quality Control Board), Gary Lemon (California Central Valley Flood Protection Board), and Josuwa Bernardo and Anthony Endow (Solano County Department of Resource Management). Field assistance by Jason Emmons, Nicole Spadone, Mirann Tsumura, and Leah Zivalic.

References

- Abichou, T., T. Johnson, K. Mahieu, J.P. Chanton, M. Romdhane, and I. Mansouri. 2010. Developing a design approach to reduce methane emissions from California landfills. In: D. Fratta et al., editors, *GeoFlorida 2010: Advances in Analysis, Modeling & Design*, Orlando, FL. 20–24 Feb. 2010. *Geotech. Spec. Publ.* 199. Am. Soc. Civ. Eng., Reston, VA. p. 2878–2887.
- Abreu, L.D.V., and P.C. Johnson. 2006. Simulating the effect of aerobic biodegradation on soil vapor intrusion into buildings: Influence of degradation rate, source concentration, and depth. *Environ. Sci. Technol.* 40:2304–2315. doi:10.1021/es051335p
- Aronson, E., S. Allison, and B. Helliker. 2013. Environmental impacts on the diversity of methane-cycling microbes and their resultant function. *Front. Microbiol.* 4:225. doi:10.3389/fmicb.2013.00225
- Bender, M., and R. Conrad. 1995. Effect of CH₄ concentrations and soil conditions on the induction of CH₄ oxidation activity. *Soil Biol. Biochem.* 27:1517–1527. doi:10.1016/0038-0717(95)00104-M
- Boeckx, P., O. Van Cleemput, and I. Villaralvo. 1997. Methane oxidation in soils with different textures and land use. *Nutr. Cycl. Agroecosyst.* 49:91–95. doi:10.1023/A:1009706324386
- Börjesson, G., J. Samuelsson, and J. Chanton. 2007. Methane oxidation in Swedish landfills quantified with the stable carbon isotope technique in combination with an optical method for emitted methane. *Environ. Sci. Technol.* 41:6684–6690. doi:10.1021/es062735v
- Chan, A.S.K., and T.B. Parkin. 2000. Evaluation of potential inhibitors of methanogenesis and methane oxidation in a landfill cover soil. *Soil Biol. Biochem.* 32:1581–1590. doi:10.1016/S0038-0717(00)00071-7
- Chanton, J., T. Abichou, C. Langford, K. Spokas, G. Hater, R. Green, et al. 2011. Observations on the methane oxidation capacity of landfill soils. *Waste Manage.* 31:914–925. doi:10.1016/j.wasman.2010.08.028
- Chanton, J.P., D.K. Powelson, and R.B. Green. 2009. Methane oxidation in landfill cover soils: Is a 10% default value reasonable? *J. Environ. Qual.* 38:654–663. doi:10.2134/jeq2008.0221
- Chen, H., Q. Zhu, C. Peng, N. Wu, Y. Wang, X. Fang, et al. 2013. Methane emissions from rice paddies natural wetlands, lakes in China: Synthesis new estimate. *Global Change Biol.* 19:19–32. doi:10.1111/gcb.12034
- Czepiel, P.M., B. Mosher, R.C. Harriss, J.H. Shorter, J.B. McManus, C.E. Kolb, et al. 1996. Landfill methane emissions measured by enclosure and atmospheric tracer methods. *J. Geophys. Res.* 101(D11):16711–16719. doi:10.1029/96JD00864
- Dörr, H., L. Katruff, and I. Levin. 1993. Soil texture parameterization of the methane uptake in aerated soils. *Chemosphere* 26:697–713. doi:10.1016/0045-6535(93)90454-D
- Etiopo, G., and R.W. Klusman. 2010. Microseepage in drylands: Flux and implications in the global atmospheric source/sink budget of methane. *Global Planet. Change* 72:265–274. doi:10.1016/j.gloplacha.2010.01.002
- Gebert, J., A. Groenroeft, and E.-M. Pfeiffer. 2011. Relevance of soil physical properties for the microbial oxidation of methane in landfill covers. *Soil Biol. Biochem.* 43:1759–1767. doi:10.1016/j.soilbio.2010.07.004
- Gulledge, J., Y. Hrywna, C. Cavanaugh, and P.A. Steudler. 2004. Effects of long-term nitrogen fertilization on the uptake kinetics of atmospheric methane in temperate forest soils. *FEMS Microbiol. Ecol.* 49:389–400. doi:10.1016/j.femsec.2004.04.013
- Gulledge, J., and J.P. Schimel. 1998. Moisture control over atmospheric CH₄ consumption and CO₂ production in diverse Alaskan soils. *Soil Biol. Biochem.* 30:1127–1132. doi:10.1016/S0038-0717(97)00209-5
- Hütsch, B. 1998. Tillage and land use effects on methane oxidation rates and their vertical profiles in soil. *Biol. Fertil. Soils* 27:284–292. doi:10.1007/s003740050435
- IPCC. 2013. *Climate change 2013: The physical science basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change.* Cambridge Univ. Press, Cambridge, UK. doi:10.1017/CBO9781107415324
- Jewell, K.P., and J.T. Wilson. 2011. A new screening method for methane in soil gas using existing groundwater monitoring wells. *Groundwater Monit. Rem.* 31:82–94. doi:10.1111/j.1745-6592.2011.01345.x
- Johnson, P., P. Lundegard, and Z. Liu. 2006. Source zone natural attenuation at petroleum hydrocarbon spill sites: I. Site-specific assessment approach. *Ground Water Monit. Rem.* 26:82–92. doi:10.1111/j.1745-6592.2006.00114.x
- Kammann, C., L. Grunhage, H.J. Jager, and G. Wachinger. 2001. Methane fluxes from differentially managed grassland study plots: The important role of CH₄ oxidation in grassland with a high potential for CH₄ production. *Environ. Pollut.* 115:261–273. doi:10.1016/S0269-7491(01)00103-8
- Klute, A. 1986. Water retention: Laboratory methods. In: A. Klute, editor, *Methods of soil analysis. Part 1. Physical and mineralogical methods.* SSSA Book Ser. 5. SSSA and ASA, Madison, WI. p. 635–662. doi:10.2136/sssabookser5.1.2ed.c26
- Le Mer, J., and P. Roger. 2001. Production, oxidation, emission and consumption of methane by soils: A review. *Eur. J. Soil Biol.* 37:25–50. doi:10.1016/S1164-5563(01)01067-6
- Lundegard, P.D., and P.C. Johnson. 2006. Source zone natural attenuation at petroleum hydrocarbon spill sites: II. Application to a former oil field. *Ground Water Monit. Rem.* 26:93–106. doi:10.1111/j.1745-6592.2006.00115.x
- Ma, J., H. Luo, G.E. DeVaul, W.G. Rixey, and P.J.J. Alvarez. 2014. Numerical model investigation for potential methane explosion and benzene vapor intrusion associated with high-ethanol blend releases. *Environ. Sci. Technol.* 48:474–481. doi:10.1021/es403926k
- Ma, J., W.G. Rixey, and P.J.J. Alvarez. 2013. Microbial processes influencing the transport, fate and groundwater impacts of fuel ethanol releases. *Curr. Opin. Biotechnol.* 24:457–466. doi:10.1016/j.copbio.2012.09.005

- Ma, J., W.G. Rixey, G.E. Devaull, B.P. Stafford, and P.J.J. Alvarez. 2012. Methane bioattenuation and implications for explosion risk reduction along the groundwater to soil surface pathway above a plume of dissolved ethanol. *Environ. Sci. Technol.* 46:6013–6019. doi:10.1021/es300715f
- Maxfield, P.J., E.R.C. Hornibrook, and R.P. Evershed. 2008. Acute impact of agriculture on high-affinity methanotrophic bacterial populations. *Environ. Microbiol.* 10:1917–1924. doi:10.1111/j.1462-2920.2008.01587.x
- Miller, L.G., C. Sasson, and R.S. Oremland. 1998. Difluoromethane, a new and improved inhibitor of methanotrophy. *Appl. Environ. Microbiol.* 64:4357–4362.
- Mønster, J., J. Samuelsson, P. Kjeldsen, and C. Scheutz. 2015. Quantification of methane emissions from 15 Danish landfills using the mobile tracer dispersion method. *Waste Manage.* 35:177–186. doi:10.1016/j.wasman.2014.09.006
- Nelson, D.K., T.M. LaPara, and P.J. Novak. 2010. Effects of ethanol-based fuel contamination: Microbial community changes, production of regulated compounds, and methane generation. *Environ. Sci. Technol.* 44:4525–4530. doi:10.1021/es903940q
- Nelson, D.W., and L.E. Sommers. 1996. Total carbon, organic carbon, and organic matter. In: D.L. Sparks, editor, *Methods of soil analysis. Part 3. Chemical methods.* SSSA Book Ser. 5. SSSA and ASA, Madison, WI. p. 961–1010. doi:10.2136/sssabookser5.3.c34
- Okamoto, H., and Y. Gomi. 2011. Empirical research on diffusion behavior of leaked gas in the ground. *J. Loss Prev. Process Ind.* 24:531–540. doi:10.1016/j.jlp.2011.01.007
- Poulsen, T.G., M. Christophersen, P. Moldrup, and P. Kjeldsen. 2001. Modeling lateral gas transport in soil adjacent to old landfill. *J. Environ. Eng.* 127:145–153. doi:10.1061/(ASCE)0733-9372(2001)127:2(145)
- Powers, S.E., C.S. Hunt, S.E. Heermann, H.X. Corseuil, D. Rice, and P.J.J. Alvarez. 2001. The transport and fate of ethanol and BTEX in groundwater contaminated by gasohol. *Crit. Rev. Environ. Sci. Technol.* 31:79–123. doi:10.1080/20016491089181
- Raich, J.W., and W.H. Schlesinger. 1992. The global carbon dioxide flux in soil respiration and its relationship to vegetation and climate. *Tellus B* 44:81–99. doi:10.1034/j.1600-0889.1992.t01-1-00001.x
- Rasa, E., B.A. Bekins, D.M. Mackay, N.R. de Sieyes, J.T. Wilson, K.P. Feris, et al. 2013. Impacts of an ethanol-blended fuel release on groundwater and fate of produced methane: Simulation of field observations. *Water Resour. Res.* 49:4907–4926. doi:10.1002/wrcr.20382
- Rong, Y., L. Ma, and D.A. Johnson. 2015. Methane uptake by four land-use types in the agro-pastoral region of northern China. *Atmos. Environ.* 116:12–21. doi:10.1016/j.atmosenv.2015.06.003
- Sass, R., F. Fisher, Y. Wang, F. Turner, and M. Jund. 1992. Methane emission from rice fields: The effect of floodwater management. *Global Biogeochem. Cycles* 6:249–262. doi:10.1029/92GB01674
- Schnell, S., and G.M. King. 1996. Responses of methanotrophic activity in soils and cultures to water stress. *Appl. Environ. Microbiol.* 62:3203–3209.
- Seghers, D., E.M. Top, D. Reheul, R. Bulcke, P. Boeckx, W. Verstraete, and S.D. Siciliano. 2003. Long-term effects of mineral versus organic fertilizers on activity and structure of the methanotrophic community in agricultural soils. *Environ. Microbiol.* 5:867–877. doi:10.1046/j.1462-2920.2003.00477.x
- Sihota, N.J., K.U. Mayer, M.A. Toso, and J.F. Atwater. 2013. Methane emissions and contaminant degradation rates at sites affected by accidental releases of denatured fuel-grade ethanol. *J. Contam. Hydrol.* 151:1–15. doi:10.1016/j.jconhyd.2013.03.008
- Sihota, N.J., J.J. Trost, B. Bekins, A. Berg, G.N. Delin, B. Mason, et al. 2016. Seasonal variability in vadose zone biodegradation at a crude oil pipeline rupture site. *Vadose Zone J.* 15(5). doi:10.2136/vzj2015.09.0125
- Soil Survey Staff. 2017. Web soil survey. *Natl. Soil Surv. Ctr., Lincoln, NE.* <https://websoilsurvey.sc.egov.usda.gov/> (accessed 31 May 2017).
- Spalding, R.F., M.A. Toso, M.E. Exner, G. Hattan, T.M. Higgins, A.C. Sekely, and S.D. Jensen. 2011. Long-term groundwater monitoring results at large, sudden denatured ethanol releases. *Groundwater Monit. Rem.* 31:69–81. doi:10.1111/j.1745-6592.2011.01336.x
- Tate, K.R., A.S. Walcroft, and C. Pratt. 2012. Varying atmospheric methane concentrations affect soil methane oxidation rates and methanotroph populations in pasture, an adjacent pine forest, and a landfill. *Soil Biol. Biochem.* 52:75–81. doi:10.1016/j.soilbio.2012.04.011
- van den Pol-van Dasselaar, A., M.L. van Beusichem, and O. Oenema. 1998. Effects of soil moisture content and temperature on methane uptake by grasslands on sandy soils. *Plant Soil* 204:213–222. doi:10.1023/A:1004371309361
- Whalen, S.C., and W.S. Reebergh. 1996. Moisture and temperature sensitivity of CH₄ oxidation in boreal soils. *Soil Biol. Biochem.* 28:1271–1281. doi:10.1016/S0038-0717(96)00139-3
- Wilson, J., M. Toso, D. Mackay, N. de Sieyes, and G. DeVaul. 2012. What's the deal with methane at LUST spill sites? Part 1. In: *L.U.S.T. Line Bulletin 71.* New England Interstate Water Pollution Control Commission, Lowell, MA. p. 6–8 and 13.
- Xu, X., and K. Inubushi. 2009. Ethylene oxidation, atmospheric methane consumption, and ammonium oxidation in temperate volcanic forest soils. *Biol. Fertil. Soils* 45:265–271. doi:10.1007/s00374-008-0324-0
- Yan, Y., X. Dong, and J. Li. 2015. Experimental study of methane diffusion in soil for an underground gas pipe leak. *J. Nat. Gas Sci. Eng.* 27:82–89. doi:10.1016/j.jngse.2015.08.039
- Young, T.M., and R.D. Golding. 2002. Underground storage tank system field-based research project report. California State Water Resources Control Board, Sacramento.
- Zechmeister-Boltenstern, S., and K.A. Smith. 1998. Ethylene production and decomposition in soils. *Biol. Fertil. Soils* 26:354–361. doi:10.1007/s003740050388