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A long-wavelength xanthene dye for photoacoustic imaging

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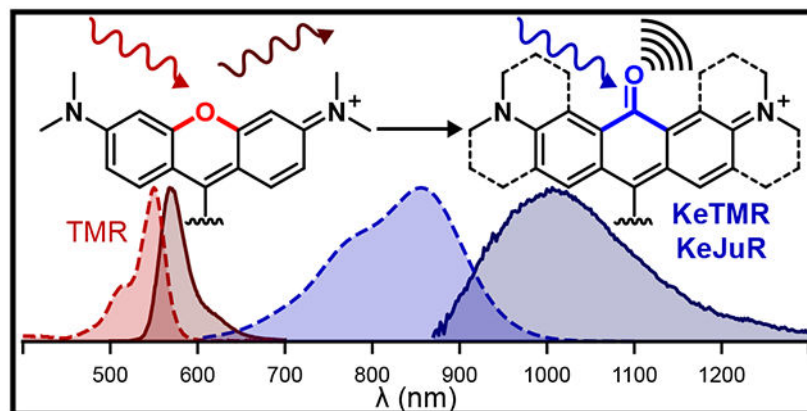
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Abstract

Photoacoustic (PA) imaging is a powerful biomedical imaging modality. We designed KeTMR and KeJuR, two xanthene-based dyes that readily obtained through a 2-step synthetic route. KeJuR has low molecular weight, good aqueous solubility, and superior chemical stability compared to KeTMR. KeJuR shows robust PA signal at 860 nm excitation and can be paired with traditional PA dyes for multiplex imaging in blood samples under tissue-mimicking environment.

Graphical Abstract



Photoacoustic (PA) tomography, a unique “light in, sound out” imaging technique, is a powerful biomedical imaging modality.¹ Commercially available PA instrumentations rely on excitation of PA contrast agents with a pulsed laser ranging from 680 – 980 nm.² Non-radiative decay, typically in the form of local heating (~0.1 K), leads to thermoelastic expansion, and the pulsed nature of excitation light results in fluctuating pressure waves that propagate through the samples as soundwaves with megahertz (MHz) frequencies.

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These soundwaves are detected at the surface by an ultrasound transducer to allow for the reconstruction of a 3D image of the sample. Since soundwaves scatter ~1000-fold less than light in tissue, PA offers high resolution imaging at deep tissue levels. The spatial resolution is about 1/200th of the imaging depth and could reach 350 μm at 7 cm depth.³ Importantly, unlike X-ray and positron emission tomography (PET) imaging techniques, PA does not require ionizing radiation and is therefore considered less hazardous.¹

A range of small molecule chromophores have been used to generate contrast for PA imaging: polymethines, indocyanine green (ICG), phthalans, xanthenes, squarans, and porphyrins can generate PA signals.⁴ The ideal PA contrast agent should absorb near infrared light (NIR, 650 – 1000 nm) strongly ($\epsilon > 10^4 \text{ M}^{-1}\cdot\text{cm}^{-1}$) to mitigate optical interferences like tissue scattering and interference from endogenous chromophores. Unlike fluorescence imaging, a low emissive quantum yield ($\Phi \approx 5\%$) is beneficial for PA imaging: excitation energy is dissipated as local heating to generate strong PA signal. Additionally, the agent should meet typical bioimaging criteria like low molecular weight, good aqueous solubility, and chemical stability.⁵ Achieving desirable PA qualities of long-wavelength absorption, high extinction coefficients, and low quantum yield while maintaining low molecular weight and good aqueous solubility remains an outstanding challenge.⁶

Historically, xanthene-based dyes like fluorescein and rhodamine have made significant contributions to fluorescence imaging owing to their visible-light excitation and emission profiles and high brightness.⁷ Traditional oxygen substitution at the 10' position limits the excitation and emission to visible or far-red wavelengths (Figure 1). More recently, xanthene dyes were pulled into NIR bioimaging window through a bridging atom switching method. Both the excitation and emission maximum of xanthene dyes can be red shifted by over 100 nm upon replacement of the bridging 10' oxygen with other, less electron rich groups like borinate,⁸ dimethyl silicon,⁹ phosphinate,¹⁰ phosphine oxide,¹¹ and sulfone.¹² All these modifications involved a sp^3 hybridized bridging atom with a tetrahedral geometry, which decouples their electronic orbitals from the xanthene's π -conjugated backbone. This low coupling results in little effect on the molar extinction coefficient (ϵ), quantum yield (Φ_{fl}), or shape of the excitation/emission spectra of the dye. This 10' atomic substitution method has been used to design bright, long wavelength fluorescent dyes for various purposes like protein labeling,^{13, 14} sensing,^{15, 16} and functional imaging.¹⁷⁻¹⁹

Xanthene dyes have been previously explored for PA applications.^{4, 20-22} The strategies generally involved forcing the excited state dye to decay non-radiatively through the twisted intramolecular charge transfer (TICT) pathway. However, this strategy often results in decreased water solubility, increased molecular weight, installation of polar sulfonates which complicate synthetic methods, and decreased chemical stability. We thought we could achieve PA imaging with a minimally disruptive modification to xanthene dyes, thereby retaining the low molecular weight, chemical stability, and good solubility profiles of xanthene dyes.

We hypothesized that an electron deficient group bearing a sp^2 hybridized bridging atom inserted at the bridging position would possess good π -overlap and integration into the molecular orbitals of the xanthene chromophore, thereby substantially red-shifting the

excitation and enhancing vibronic coupling. To test this hypothesis, we designed our xanthene-based PA agents by introducing a ketone functional group into this bridging position (Figure 1). According to the Dewar-Knott color rule^{23, 24} and computational calculations of other xanthene dyes,^{10, 12} the strong electron withdrawing nature of the bridging ketone group should lower the dye's LUMO energy level, providing a dramatic red-shift of both absorption and emission. Additionally, the sp² hybridization should provide effective vibronic coupling²⁵ to further boost PA signal. While we were preparing this manuscript for publication, an elegant synthesis and characterization of a ketone-containing tetramethyl rhodamine was reported, establishing the viability of the ketone-incorporation approach.²⁶

We first synthesized the ketone-bearing tetramethyl rhodamine (**KeTMR**) derivative (Figure 1). We elected to follow a different synthetic route^{8, 10} from the previously reported synthesis of **KeTMR**,²⁶ in an effort to generate a concise and generalizable synthesis by introducing the ketone functionality at a late stage (Figure 2). Similar to previously reported syntheses of Nebraska Red⁶ and CAFS⁴ dyes, we started from a brominate triarylmethane, **1**. Lithiation of **1** with *sec*-butyl lithium, followed by quenching with an electrophile would generate the ketone-containing dye core.

We screened several electrophiles, including diethyl carbonate, carbonyldiimidazole, and dimethylcarbonyl chloride; the use of *N*-methoxy-*N*-methylcarbonyl chloride gives the best yield (Table S1). The hemiaminal intermediate was then effectively converted to ketone under mild acidic condition. After a brief work-up the leuco-dye can be oxidized with iodine in non-polar solvent like diethyl ether to precipitate **KeTMR** as brown solid (Figure 2). All the side products were soluble and could be washed away. This column-free, 2-step synthetic approach gives the desired **KeTMR** in 83 % yield (Figure 2). While characterizing **KeTMR** compound by ¹H NMR, pure **KeTMR** was NMR silent (**Spectrum S1**). Addition of I₂ regenerated the ¹H NMR signal of **KeTMR** (**Spectrum S2**). EPR measurements reveal radical character in **KeTMR** without I₂; the EPR of **KeTMR** + I₂ shows no radical character (**Spectrum S3**). Together, these experiments establishes that **KeTMR** possesses radical character.

We characterized the optical properties of **KeTMR**. In aqueous solution under physiological conditions (phosphate-buffered saline, PBS, pH = 7.2, with 0.5 % DMSO), **KeTMR** absorbs strongly ($\epsilon = 26,900 \text{ M}^{-1} \text{ cm}^{-1}$) at 856 nm, with a wide full width at half maximum (FWHM, 2422 cm⁻¹, Figure 3a). The fluorescence emission peaks at 1006 nm after solvent re-absorption correction (see SI), with a FWHM of 1857 cm⁻¹ (Figure 3a). The quantum yield is low: 0.013 % (see SI). These values agree with previously reported absorbance, emission, and fluorescence quantum yield values (Table S4).²⁶ In comparison with xanthene-based fluorophores, **KeTMR** has a much broader FWHM of both absorption and emission, larger Stokes shift (150 nm or 1742 cm⁻¹), and lower quantum efficiencies (Figure S1 and Table S2). These parameters provide strong evidence that the ketone bridging group causes a significant vibronic coupling to the xanthene dye core.

The excitation and emission spectral profiles of **KeTMR** is influenced by solvent dielectric, on account of the direct conjugation of the highly polarizable ketone group with the rest of

the xanthene molecular orbital. Indeed, the emission maximum of **KeTMR** increased almost linearly from 837 nm in toluene to 1006 nm in water ($r^2 = 0.924$), indicating a strong solvent–dye interaction (Figure 3b, Table S3, Figure S2). The absorbance maximum does not vary strongly with solvent dielectric ($r^2 = 0.20$, Figure S3), suggesting the solvent-dye interaction is strongest in the excited state.

A plot of Stokes shifts vs. solvent orientation polarizability (Δf)²⁷ for **KeTMR** reveals a strong correlation between Stokes shift and Δf for **KeTMR** in protic solvents ($r^2 = 0.917$), but weak correlation in aprotic solvents ($r^2 = 0.135$, Figure S4a), indicating that hydrogen bonding between solvent and the ketone group directly influence the photophysical properties of **KeTMR**. The width of the emission peak of **KeTMR** varies linearly with the hydrogen bond donation (HBD) strength or α value,²⁸ of the solvent ($r^2 = 0.927$ Figure S4c), suggesting the degree of vibronic coupling could be tuned by the dye's local environment. This was also supported by the low quantum yield of fluorescence in water (0.013%) and the relatively higher quantum yield in CH_2Cl_2 (1.3%, Table S4).²⁶

Taken together, the photophysical properties of **KeTMR** provided strong evidence to support our design hypothesis that electron deficient sp^2 ketone bridged xanthene should have stronger vibronic coupling at higher wavelength, making them suitable for PA applications.

We next sought to improve the fluorescence quantum efficiency, so that the same molecule could be applied for both fluorescence and PA imaging modalities. We took advantage of our modular synthetic design to prepare the julolidine version of the ketone rhodamine, **KeJuR** (Figure 4). Rigidification of the aniline substituents by annulation should improve the fluorescence quantum yield, as the same method has been reported on various dyes before.^{10, 29} The synthesis of **KeJuR** was also through a 2-step route, beginning from **2**, available on gram scale in 40% yield from commercially available starting material. Using conditions identical to the synthesis provided **KeJuR** in 11% yield. The addition of lanthanum(III) salt to further activate the carbamoyl group³⁰ improved the overall yield to 30% (Figure 4).

KeJuR shows slightly red-shifted absorbance maximum, and blue-shifted emission maximum compared to **KeTMR** under the same solvent conditions (Figure 5, Table S3). In aqueous solution (PBS, pH 7.2, 0.5% DMSO), **KeJuR** has an absorbance maximum at 861 nm ($\epsilon = 21000 \text{ M}^{-1} \text{ cm}^{-1}$, $\text{FWHM} = 2080 \text{ cm}^{-1}$) and a broad emission at 988 nm ($\text{FWHM} = 1743 \text{ cm}^{-1}$). The fluorescence quantum yield of **KeJuR** is 0.032% in DPBS, and 2.3% in dichloromethane, slightly higher than the quantum yield of **KeTMR**.

KeTMR shows greater solvatochromism than **KeJuR**. First, a plot of emission vs. solvent dielectric for **KeJuR** also gives a near-linear fit ($r^2 = 0.911$), Figure 5b), but with a shallower slope compared to **KeTMR** (1.86 vs. 1.52). Like **KeTMR**, the Stokes shift of **KeJuR** depends on solvent orientation polarizability (Δf , Figure S3b). In aprotic solvents, the correlation is quite weak ($r^2 = 0.135$). In protic solvents **KeJuR** ($r^2 = 0.794$) shows a weaker correlation than **KeTMR** ($r^2 = 0.917$), consistent with the hypothesis that the carbonyl of **KeJuR** is more shielded from solvent than the carbonyl of **KeTMR**. Finally, in contrast to **KeTMR**, the FWHM of **KeJuR** emission shows only a weak correlation with solvent HBD

strength (α , $r^2 = 0.68$, Figure S3c), indicating that hydrogen bond donation from the solvent does not play as strong a role in altering the photophysical properties of **KeJuR**.

Both of the new ketone rhodamines give good PA signals in aqueous buffer, establishing that introduction of the carbonyl group at 10' is a viable strategy for generating PA contrast reagents from xanthene scaffolds (Figure 6a). **KeTMR** and **KeJuR** possess PA peaks at approximately 860 nm. **KeJuR** shows a slightly stronger PA signal than **KeTMR** (Figure 6a, magenta vs tan). Introduction of the highly conjugated, strongly electron withdrawing ketone group might decrease chemical stability of ketone xanthene dyes by increasing their susceptibility to nucleophiles. We find this is the case for **KeTMR**, but not **KeJuR**. After treatment with glutathione (GSH), 86% **KeJuR** survived the reaction while only 6% **KeTMR** did under the same conditions (Figure S5). As a result, **KeJuR** was used in subsequent PA imaging studies.

KeJuR shows good aqueous solubility. Long wavelength dyes used for PA often suffer from low aqueous solubilities on account of the extended conjugation needed to achieve wavelengths compatible with PA imaging. This often requires either a high percentage of organic co-solvent like DMSO or DMF (in the range of 10-50 %), or detergents like SDS or CTAB (typically 0.1 wt %) to achieve solubility. Without these solubilizing reagents, dyes would aggregate, and display altered absorption and emission profiles, appearing as deviations in the signal at higher dye concentrations. Since the **KeJuR** was designed with a minimum structural change compared to its parent rhodamine scaffold, we envisioned this dye should enjoy a good aqueous solubility and maintain its spectral shape even at higher concentrations. The shape of the PA spectrum of **KeJuR** does not change over a concentration range of 5 to 50 μM **KeJuR** (DPBS, pH 7.2, 1% DMSO) (Figure S6). A plot of PA signal (860 nm) vs concentration is linear up through 50 μM ($r^2 = 0.998$), indicating the good aqueous solubility of **KeJuR** (Figure 6b). **KeJuR** and **KeTMR** show good photostability compared to ICG (Figure S7).

With strong PA signal at 860 nm, good aqueous solubility, and high chemical stability, we wondered whether **KeJuR** could be paired with other PA dyes for multi-color imaging purposes. Since **KeJuR** gave a maximum PA signal at 860 nm, we picked the FDA-approved indocyanine green (ICG) as our partner dye.⁶

ICG gives strong PA signal at 680 nm due to strong H-aggregation,³¹ while **KeJuR** gives even higher signal at 860 nm (Figure 6a). **KeJuR** performs well in a model of blood and tissue. After spectral unmixing, we observe 10 \times higher PA signal from **KeJuR** compared to ICG when the dyes are imaged in defibrinated sheep blood within a tissue phantom (Figure 6c–e).³²

In conclusion, we offer a new strategy to re-purpose the well-known fluorescent xanthene scaffold for PA imaging by manipulating the bridging hybridization geometry. The trigonal planar, ketone-containing **KeTMR** and **KeJuR** can be synthesized through a straightforward 2-step approach. The photophysical properties measured in various solvents of these two dyes strongly supported our hypothesis, that installing an sp^2 hybridized ketone group at the 10' position of the xanthene core can red-shift the optical properties and decrease the

fluorescence quantum yield via enhanced vibronic coupling with the dye scaffold, making xanthenes more useful in PA applications.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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References

1. Wang LHV and Hu S, *Science*, 2012, 335, 1458–1462. [PubMed: 22442475]
2. Knox HJ and Chan J, *Accn Chem Res*, 2018, 51, 2897–2905.
3. Li H, Zhang P, Smaga LP, Hoffman RA and Chan J, *J Am Chem Soc*, 2015, 137, 15628–15631. [PubMed: 26652006]
4. Borg RE and Rochford J, *Photochem Photobiol*, 2018, 94, 1175–1209. [PubMed: 29953628]
5. Luo S, Zhang E, Su Y, Cheng T and Shi C, *Biomaterials*, 2011, 32, 7127–7138. [PubMed: 21724249]
6. Weber J, Beard PC and Bohndiek SE, *Nat Methods*, 2016, 13, 639–650. [PubMed: 27467727]
7. Lavis LD, *Annual review of biochemistry*, 2017, 86, 825–843.
8. Zhou XQ, Lesiak L, Lai R, Beck JR, Zhao J, Elowsky CG, Li H and Stains CI, *Angew Chem Int Edit*, 2017, 56, 4197–4200.
9. Fu MY, Xiao Y, Qian XH, Zhao DF and Xu YF, *Chem Commun*, 2008, DOI: 10.1039/b718544h, 1780–1782.
10. Zhou XQ, Lai R, Beck JR, Li H and Stains CI, *Chem Commun*, 2016, 52, 12290–12293.
11. Chai X, Cui X, Wang B, Yang F, Cai Y, Wu Q and Wang T, *Chemistry*, 2015, 21, 16754–16758. [PubMed: 26420515]
12. Liu J, Sung YQ, Zhang HX, Shi HP, Shi YW and Guo W, *Acs Appl Mater Inter*, 2016, 8, 22953–22962.
13. Grimm JB, Tkachuk AN, Xie LQ, Choi H, Mohar B, Falco N, Schaefer K, Patel R, Zheng QS, Liu Z, Lippincott-Schwartz J, Brown TA and Lavis LD, *Nat Methods*, 2020, 17, 815–+. [PubMed: 32719532]
14. Wang L, Tran M, D'Este E, Roberti J, Koch B, Xue L and Johnsson K, *Nat Chem*, 2020, 12, 165–+. [PubMed: 31792385]
15. Deo C, Sheu SH, Seo J, Clapham DE and Lavis LD, *J Am Chem Soc*, 2019, 141, 13734–13738. [PubMed: 31430138]
16. Mertes N, Busch M, Huppertz MC, Hacker CN, Wilhelm J, Gurth CM, Kuhn S, Hiblot J, Koch B and Johnsson K, *J Am Chem Soc*, 2022, 144, 6928–6935. [PubMed: 35380808]
17. Huang YL, Walker AS and Miller EW, *J Am Chem Soc*, 2015, 137, 10767–10776. [PubMed: 26237573]
18. Liu P and Miller EW, *Accn Chem Res*, 2020, 53, 11–19.
19. Gonzalez MA, Walker AS, Cao KJ, Lazzari-Dean JR, Settineri NS, Kong EJ, Kramer RH and Miller EW, *J Am Chem Soc*, 2021, 143, 2304–2314. [PubMed: 33501825]
20. Myochin T, Hanaoka K, Iwaki S, Ueno T, Komatsu T, Terai T, Nagano T and Urano Y, *J Am Chem Soc*, 2015, 137, 4759–4765. [PubMed: 25764154]
21. Ikeno T, Hanaoka K, Iwaki S, Myochin T, Murayama Y, Ohde H, Komatsu T, Ueno T, Nagano T and Urano Y, *Anal Chem*, 2019, 91, 9086–9092. [PubMed: 31265237]

22. Rathnamalala CSL, Pino NW, Herring BS, Hooper M, Gwaltney SR, Chan J and Scott CN, *Org Lett*, 2021, 23, 7640–7644. [PubMed: 34550707]
23. Dewar MJS, *Journal of the Chemical Society (Resumed)*, 1950, 2329–2334.
24. Knott EB, *Journal of the Chemical Society (Resumed)*, 1951, 1024–1028.
25. Butkevich AN, Sednev MV, Shojaei H, Belov VN and Hell SW, *Org Lett*, 2018, 20, 1261–1264. [PubMed: 29405716]
26. Daly HC, Matikonda SS, Steffens HC, Ruehle B, Resch-Genger U, Ivanic J and Schnermann MJ, *Photochem Photobiol*, 2022, 98, 325–333. [PubMed: 34676539]
27. Noboru M, Yozo K and Masao K, *Bull Chem Soc Jpn*, 1956, 29, 465–470.
28. Kamlet MJ, Abboud JLM, Abraham MH and Taft RW, *J Org Chem*, 1983, 48, 2877–2887.
29. Butkevich AN, Ta H, Ratz M, Stoldt S, Jakobs S, Belov VN and Hell SW, *ACS Chemical Biology*, 2018, 13, 475–480. [PubMed: 28933823]
30. Butkevich AN, *Org Lett*, 2021, 23, 2604–2609. [PubMed: 33720740]
31. Jung B, Vullev VI and Anvari B, *IEEE Journal of Selected Topics in Quantum Electronics*, 2014, 20, 149–157.
32. Gardner SH, Brady CJ, Keeton C, Yadav AK, Mallojjala SC, Lucero MY, Su S, Yu Z, Hirschi JS, Mirica LM and Chan J, *Angew Chem Int Edit*, 2021, 60, 18860–18866.

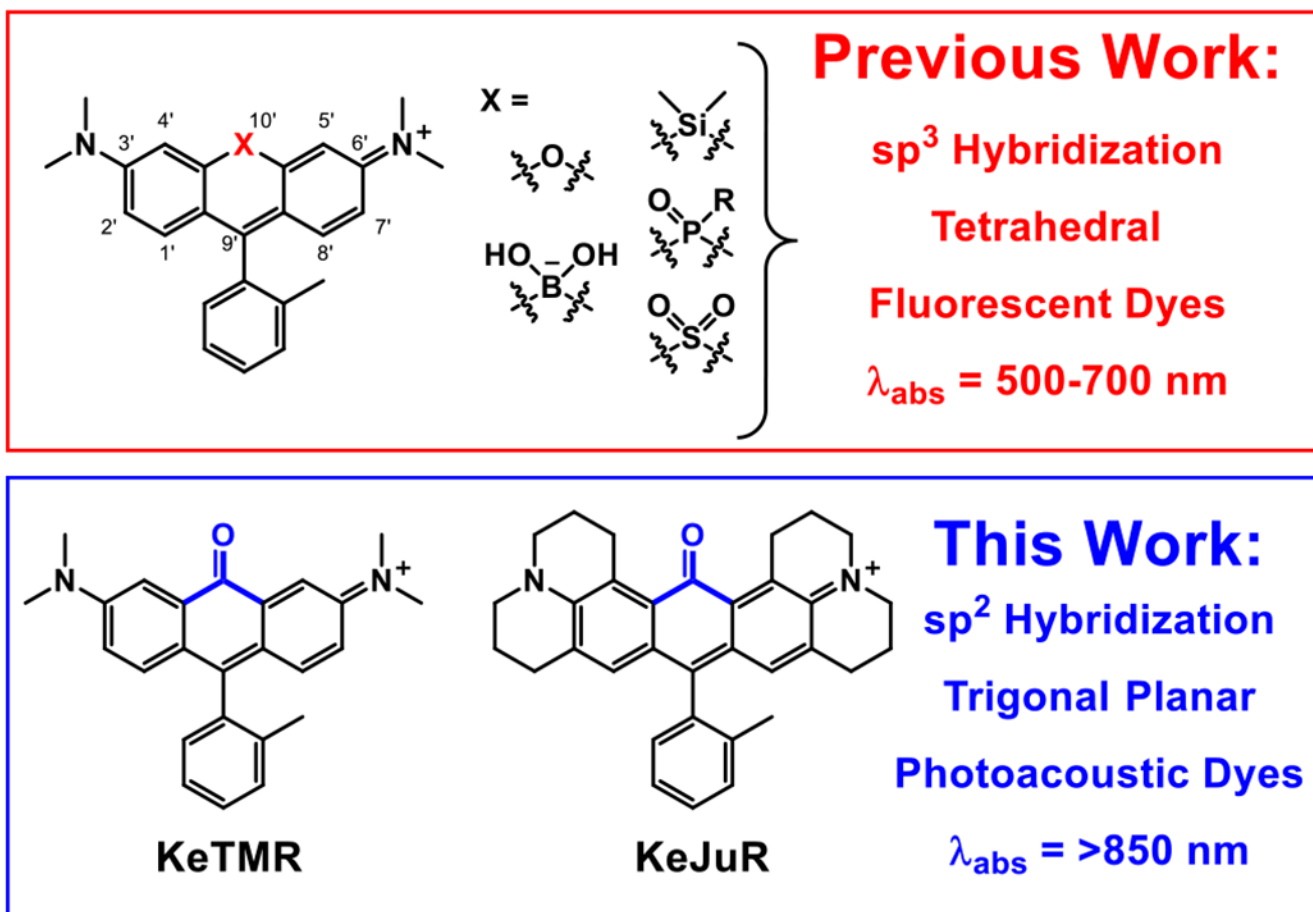


Figure 1. Structural comparison of previously reported xanthene based fluorescent dyes and the re-purposed PA dyes reported in this work.

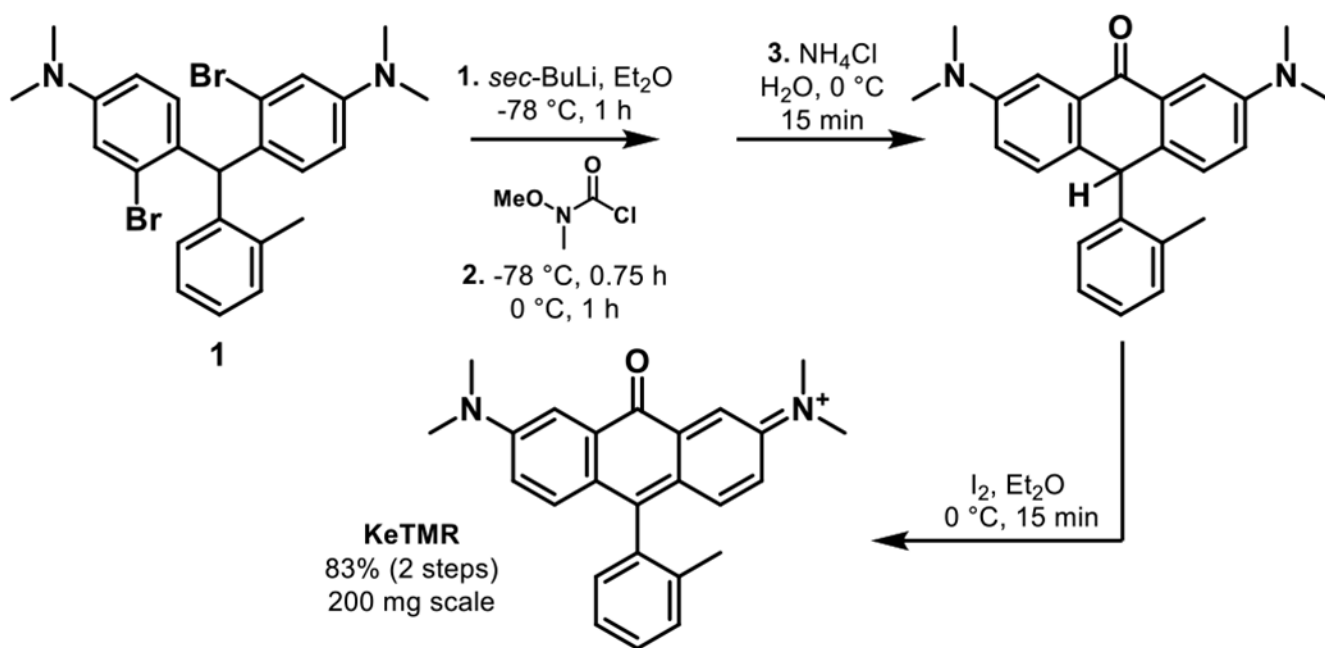


Figure 2.
Synthesis of KeTMR.

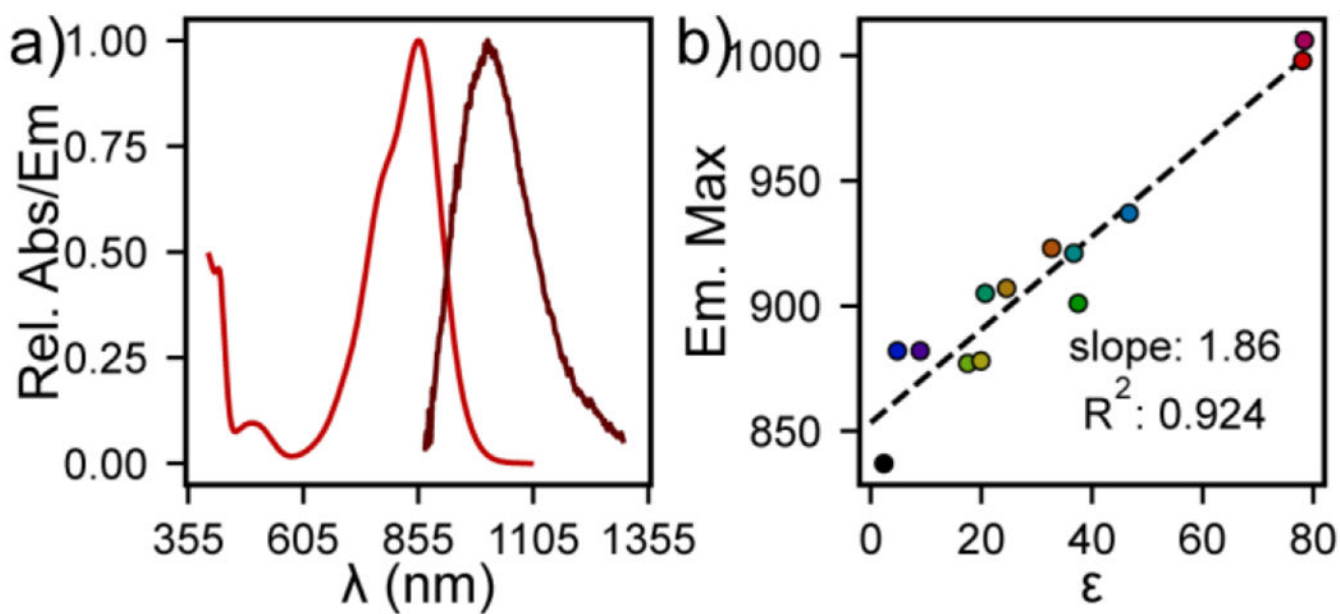


Figure 3.

Absorbance and Emission Spectra of KeTMR. **a)** Plot of relative absorbance (red) or emission intensity (dark red) for **KeTMR** (10 μM, DPBS). Emission intensity is corrected for solvent re-absorption (SI). **b)** Plot of emission maxima for **KeTMR** (10 μM) vs. dielectric constant (ϵ).

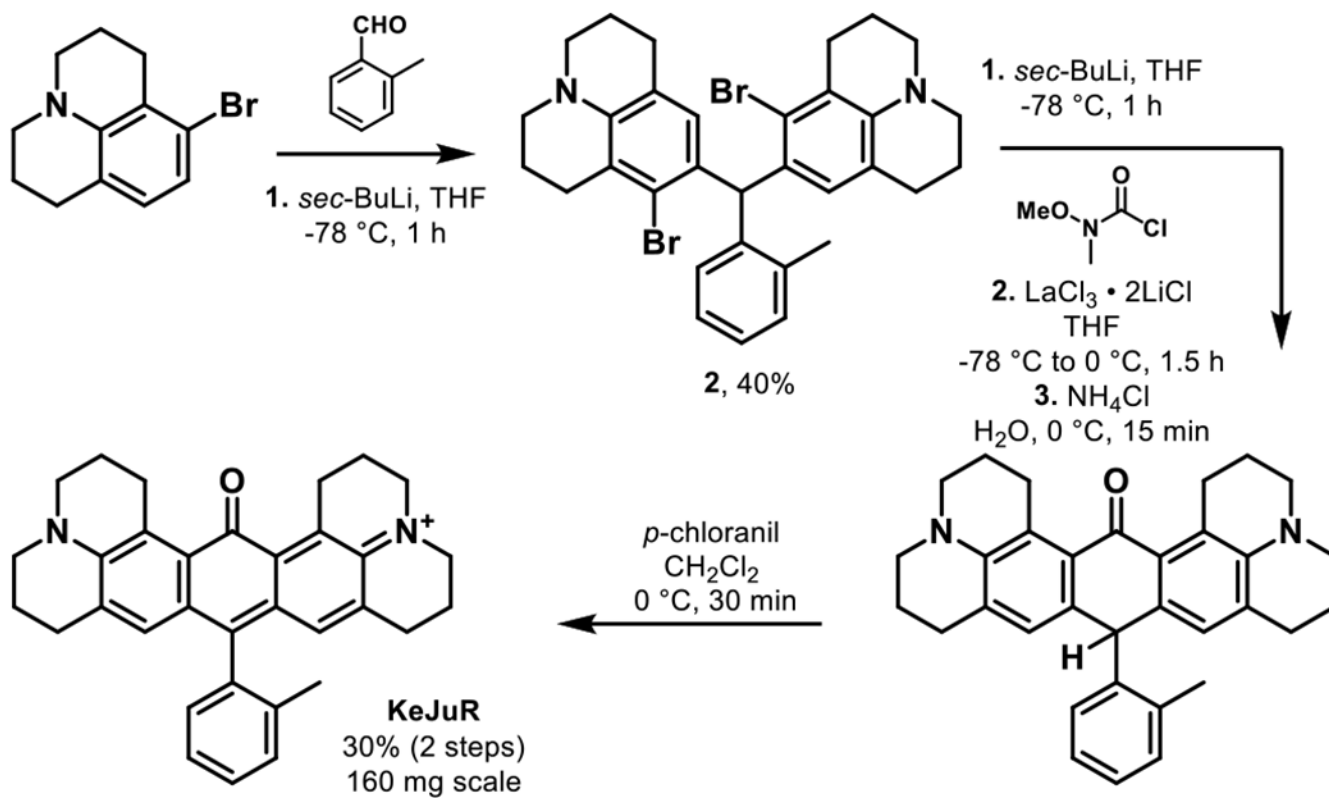


Figure 4.
Synthesis of KeJuR.

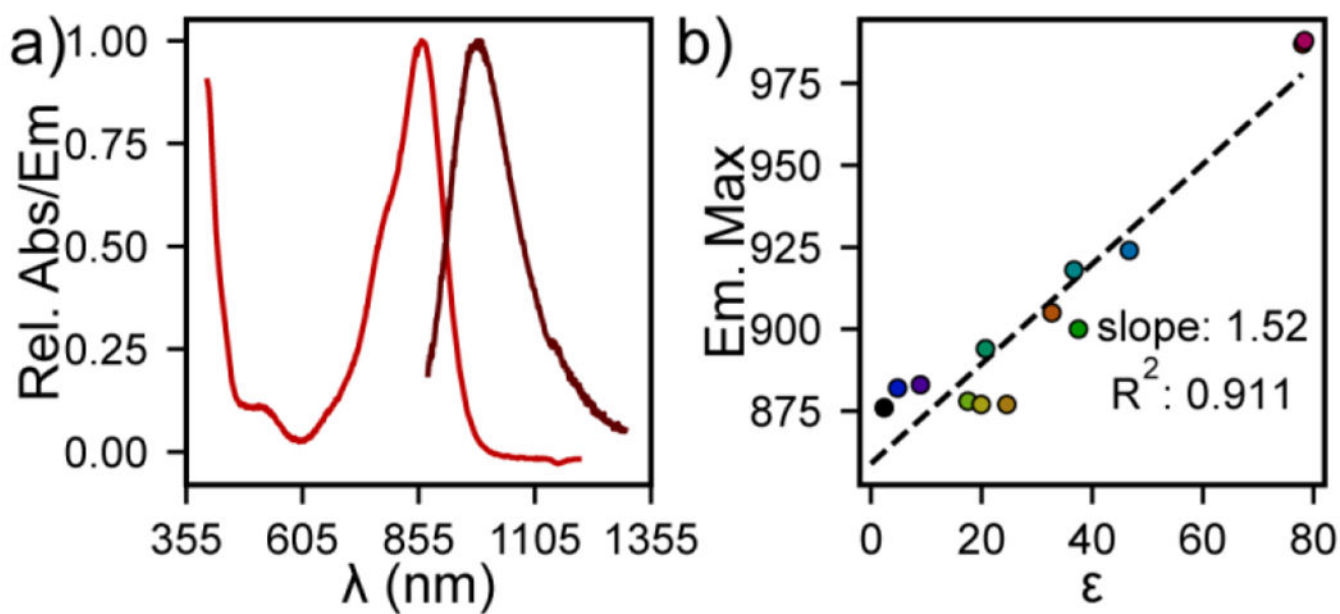


Figure 5. Absorbance and Emission Spectra of KeJuR. **a)** Plot of relative absorbance (red) or emission intensity (dark red) for **KeJuR** (10 μM, DPBS). Emission intensity is corrected for solvent re-absorption (SI). **b)** Plot of emission maxima for **KeJuR** (10 μM) vs. dielectric constant (ϵ).

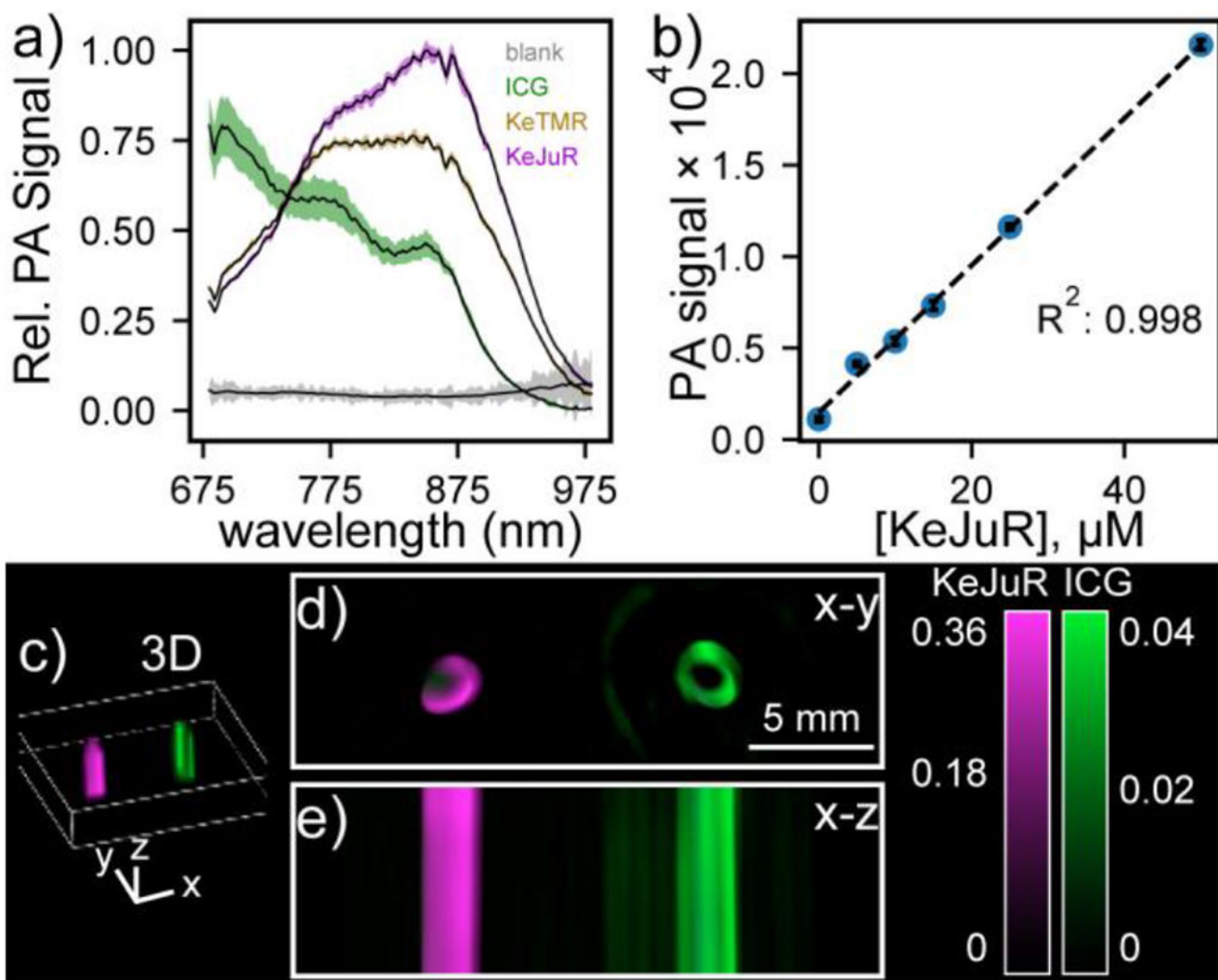


Figure 6. Photoacoustic imaging with Ke-xanthene dyes. **a)** Plot of relative PA intensity for 50 μM of the indicated dye in DBPS (pH 7.2, 1% DMSO). Data are mean ± S.D. for n = 7 different depths. **b)** Plot of PA signal (ex: 860 nm) vs. KeJuR concentration. **c)** 3D reconstruction of PA tomograph of KeJuR (magenta) or ICG (green) in sheep blood within a tissue phantom containing 60% milk. Cross - sections through **d)** x-y or **e)** x-z planes show strong PA signal for KeJuR.