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Contributions of nearby agricultural insecticide applications to indoor residential exposures

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Abstract

Background: Pesticide exposure has been associated with adverse health effects. We evaluated relationships between proximity to agricultural insecticide applications and insecticides in household dust, accounting for land use and wind direction.

Methods: We measured concentrations (ng/g) of nine insecticides in carpet-dust samples collected from 598 California homes. Using a geographic information system (GIS), we integrated the California Pesticide Use Reporting (CPUR) database to estimate agricultural use within residential buffers with radii of 0.5 to 4km. We calculated the density of use (kg/km²) during 30-, 60-, 180-, and 365-day periods prior to dust collection and evaluated relationships between three density metrics (CPUR unit-based, agricultural land area adjusted, and average daily wind

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Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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direction adjusted) and dust concentrations. We modeled natural-log transformed concentrations using Tobit regression for carbaryl, chlorpyrifos, cypermethrin, diazinon, and permethrin. Odds of detection were modeled with logistic regression for azinphos-methyl, cyfluthrin, malathion, and phosmet. We adjusted for season, year, occupation, and home/garden uses.

Results: Chlorpyrifos use within 1–4km was associated with 1 to 2-times higher dust concentrations in both the 60- and 365-day periods. Carbaryl applications within 2–4km of homes 60-days prior to dust collection were associated with 3 to 7-times higher concentrations and the 4km trend was strongest using the wind-adjusted metric (p-trend=0.04). For diazinon, there were 2-times higher concentrations for the 60-day metrics in the 2km buffer and for the CPUR and wind-adjusted metrics within 4km. Cyfluthrin, phosmet, and azinphos-methyl applications within 4km in the prior 365-days were associated with 2-, 6-, and 3-fold higher odds of detection, respectively.

Conclusions: Agricultural use of six of the nine insecticides within 4km is an important determinant of indoor contamination. Our findings demonstrated that GIS-based metrics for quantifying potential exposure to fugitive emissions from agriculture should incorporate tailored distances and time periods and support wind-adjustment for some, but not all insecticides.

Keywords

Pesticides; Insecticides; Exposure assessment; Environmental epidemiology; Dust; Agriculture

1. Introduction

In the United States (U.S.) pesticides are frequently applied to crops to control weeds and insects to produce greater yields. Annually, 899 million pounds of active ingredient are typically applied to the approximately 20% of U.S. land used for agriculture (Atwood and Paisley-Jones 2017; Bigelow and Borchers 2017). Pesticides have the potential to travel beyond the treated area through wind drift and volatilization that causes deposition in adjacent agricultural and non-agricultural areas (Farha et al. 2016; Unsworth et al. 1999). As a result, individuals living in proximity to agricultural areas with pesticide applications can be exposed through inhalation, dermal, and ingestion pathways. Epidemiologic studies have indicated that exposures to agricultural pesticides through residential proximity to pesticide applications may be associated with childhood cancer (Booth et al. 2015; Park et al. 2020; Patel et al. 2020; Reynolds et al. 2005), adult cancer (Cockburn et al. 2011; Swartz et al. 2022; Tayour et al. 2019; VoPham et al. 2015), reproductive outcomes (Eskenazi et al. 2004; Gemmill et al. 2013; Rull et al. 2006), neurobehavioral outcomes in children (Coker et al. 2017; Gunier et al. 2017a; Gunier et al. 2017b; Hyland et al. 2021; Ongono et al. 2021; Rowe et al. 2016; Shelton et al. 2014; von Ehrenstein et al. 2019), depression in adults (Furlong et al. 2020), and Parkinson's disease (Brouwer et al. 2017; Costello et al. 2009; van der Mark et al. 2012; Wang et al. 2014).

Investigating the contribution of agricultural pesticide applications to residential exposures is challenging in large part due to the lack of detailed pesticide use reporting. In California, the largest agricultural state in the U.S. (United States Department of Agriculture 2019), the California Department of Pesticide Regulation has required reporting of all agricultural

pesticide applications since 1990 for public land survey sections (PLSS; approximately 1 square mile). The California Pesticide Use Reporting (CPUR) database (California Department of Pesticide Regulation 2020b) records identify the active ingredient, date of application, crop treated, and the PLSS section. In 2017, nearly one-quarter of the pesticides applied in the U.S. were used in California (California Department of Pesticide Regulation 2020a).

Using a geographic information system (GIS), CPUR and land use data can be used to estimate pesticide use around residences (Gunier et al. 2011; Nuckols et al. 2007; Rull and Ritz 2003). A study of 89 homes in agricultural areas of central and northern California found that pesticide use within 1,250 meters of the residence was a significant determinant of concentrations of pesticide active ingredients for two herbicides, one fungicide, and two of the four insecticides assessed (Gunier et al. 2011). Here, we build on this prior work by investigating a larger number of insecticides, using expanded buffer sizes, and incorporating agricultural land use and wind direction into the exposure metrics. Our aim was to evaluate agricultural insecticide use near residences and the importance of timing of insecticide applications, location of agricultural land, and wind direction as determinants of concentrations of insecticides in homes.

2. Methods

2.1 Study Population

2.1.1 California Childhood Leukemia Study (CCLS)—Our analysis includes 577 residences from the California Childhood Leukemia Study (CCLS). The CCLS is a population-based case-control study of childhood leukemia in 35 counties in the Central Valley San Francisco Bay area (Chang et al. 2006; Ma et al. 2004; Ma et al. 2005). In 2001 to 2007, CCLS participants who were < 8 years old at diagnosis and residentially stable were invited to participate in a second interview (i.e., living in the same home at the initial and second interviews) (Ward et al. 2009) in which a carpet dust sample was collected (Colt et al. 2008). The CCLS study protocol was approved by the institutional review boards at the University of California, Berkeley, the California Committee for Protection of Human Subjects, and the National Cancer Institute.

2.1.2 Fresno Agricultural Pesticide Study (FAPS)—The Fresno Agricultural Pesticide Study (FAPS) in Fresno County, California, was a complementary exposure study conducted within the same period using the same sampling and laboratory analysis protocol and similar questionnaires (Deziel et al. 2013; Gunier et al. 2011). Interviews in 21 homes took place from 2003 to 2005. The FAPS study protocol was approved by the institutional review boards at Colorado State and Fresno State Universities and the National Cancer Institute.

2.2 Interviews and Dust Sampling

In both studies, interviewers took a Global Positioning System (GPS) reading just outside of the home. Experienced interviewers asked participants about their pesticide use using standardized questions and visual aids that identified different types of pests. Home and

garden use was ascertained by asking about specific pest treatments in the previous twelve months including treatments for ants/cockroaches, bees/wasps/hornets, flea/ticks in the home and on pets, flies/mosquitos, indoor insects, lawn and garden weeds, and professional indoor and outdoor treatments (Deziel et al. 2015). Participants were also asked if they removed their shoes before entering the home, and if in the previous twelve months anyone in the home had worked as a 1) farmer/farm or ranch worker; 2) gardener, landscaper, nursery worker, or groundskeeper; 3) packer or agricultural worker; or 4) pesticide handler or mixer.

The methods for dust sampling have been previously described (Colt et al. 2008). In short, a carpet dust sample was collected using a high-volume surface sampler vacuum (HVS3; Cascade Stack Sampling System, Venice, FL) for all the FAPS and most of the CCLS participants; if the amount of HVS3 dust was insufficient, household vacuum dust was collected by removing the used bag or emptying the loose dust from the canister into a sealable polyethylene bag. When scheduling the interview, participants were instructed not to change their vacuum bag for at least a week before the scheduled interview date. In the FAPS, the sample room was selected from the rooms that were located on the side of the home facing crops. In the CCLS, parents were asked to identify the room in which their child spent most of their waking time in the year prior to the diagnosis or reference date. If the room had a carpet or area rug measuring at least 0.84 square meters (nine square feet) and it was present before the reference date, a dust sample was collected. In the final analytic sample 168 samples (28%) were from the household vacuum and 430 (72%) were from the HVS3.

2.3 Laboratory Analysis

We measured 13 insecticides that had agricultural use in the study area counties during the study period. Allethrin, azinphos-methyl, carbaryl, chlorpyrifos, cyfluthrin, cypermethrin, deltamethrin, diazinon, malathion, permethrin, phosmet, propoxur, and tetramethrin were detected in >5% of dust samples and were included in our analyses. Isomers were measured for allethrin (1 and 2), cyfluthrin (1–4), cypermethrin (1–4), tetramethrin (1 and 2), and permethrin (cis and trans). Dust samples were shipped to the Battelle Memorial Institute (Columbus, OH) where they were stored in –20° Celsius freezers until analysis. Methods describing the laboratory quantification methods (Colt et al. 2008), quality control procedures, and detection limits have been previously described (Madrigal et al. 2021). Briefly, the 13 insecticides were extracted using a hexane:acetone solution. Extracts were quantified using gas chromatography/mass spectrometry. Detection limits ranged from 2 ng/g dust for permethrin to 100 ng/g dust for azinphos-methyl (Table 1) and mean sample recoveries ranged from 73% for malathion, phosmet, diazinon and chlorpyrifos to 122% for azinphos-methyl. The mean percent differences for within batch duplicates ranged from 0.5% for tetramethrin 1 to 38% for carbaryl.

2.4 Pesticide Use Metrics

We obtained CPUR data for insecticide applications for years 1999–2007 to correspond with the 365-day period prior to the dust collection. Data included the insecticide applied, application date, pounds applied, and PLSS area. As described previously (Gunier et al.

2011; Nuckols et al. 2007), we used a geographic information system (GIS) to create a CPUR density metric for each insecticide. This metric is defined as the sum of area-weighted kilograms of insecticide active ingredient applied in a circular buffer around a participant's residence within a time period that intersected the PLSS, divided by the area of the buffer. We created metrics for buffers with radii of 0.5-, 1-, 2-, and 4-km (buffer areas: 0.79 km², 3.14 km², 12.57 km², and 50.27 km², respectively) for each of four time periods (30, 60, 180, 365 days before dust collection).

2.4.1 CPUR Unit-based Density Metric—The CPUR density metric was computed for each residence, insecticide active ingredient, and buffer as follows ("CPUR method";

Equation 1, Figure 1):
$$CPUR_k\left(\frac{kg}{km^2}\right) = \sum_{k=0}^{\infty} \frac{\left(\left(\frac{A_j}{T_j}\right) * X_j\right)}{\text{buffer area } (km)^2}$$
, where k is the insecticide active

ingredient used in n sections intersected by the buffer around the residence, A_j is the acreage of section j within the buffer, T_j is the total acreage of section j, and X_j is the kilograms of insecticide applied in the section, j, during specific time periods.

2.4.2 Crop Area-adjusted CPUR Metric—This metric refines estimates of pesticide use density by restricting the effective area of pesticide application to areas of agricultural land with reported use of our study insecticides by section and time period in the CPUR database. To identify areas of agricultural land, we used information on pasture/hay and row crops (e.g., vegetables, grains, orchards, and vineyards), termed agricultural land hereafter, from the National Land Cover Database (NLCD) (30 m resolution) versions 2001, 2004, and 2006 (Fry et al. 2011; Homer et al. 2007; Xian et al. 2009). We linked residence locations to the version of the database that most closely corresponded to the period prior to the dust collection (e.g., visits that took place in 2003 were linked to the 2001 NLCD). Areas of agricultural land that intersected the buffers within each PLSS were summed. We excluded CPUR applications for which no agricultural land was identified by NLCD within the buffer (31% and 7% of all CPUR reported applications of our study insecticides in the 0.5km and 4km buffers, respectively). We computed a crop-area density metric (CROP-A; Figure 2) by multiplying the proportion of agricultural land within the buffer by the amount of insecticide applied to the section, summing across all the sections within the buffer, and dividing by

the agricultural land area (Equation 2):
$$CROP - A_k \left(\frac{kg}{km^2}\right) = \sum^m \frac{\left(\left(\frac{A_{ij}}{T_{ij}}\right) * X_{ij}\right)}{\text{buffer area } (km)^2}$$
, where k is the

insecticide active ingredient used on crop type i in the n sections intersected by the buffer around the residence; m is the total number of crop types on which insecticide k was applied in section j, A_{ij} is the acreage of crop type i within section j within the buffer, T_{ij} is the total acreage of crop types i within section j, and X_{ij} is the kilograms of insecticide applied to crop type i in section j.

2.4.3 Wind-adjusted CPUR Metric—To account for potential insecticide drift due to wind conditions on the date of application and subsequent days until the date of residential dust collection, we created a metric that adjusted for the proportion of days the home was downwind of insecticide applications (Figure 3). Information on daily wind direction was derived from the North American Regional Reanalysis (NARR) database which provides meteorological data at a spatial resolution of 32km × 32km and temporal resolution of 1

day (Mesinger et al. 2006). Daily NARR wind raster data at the 32km scale were used to determine the daily wind direction at the PLSS centroids that were intersected by the buffers. We determined the direction between the centroid of each portion of the section within the buffer relative to the home and weighted insecticide use in each section according to the percentage of time the wind blew from those locations during the period between application and dust collection. We considered the contributing area for pesticide drift to be within a 90° downwind 'capture zone' wedge centered on the wind direction and anchored at the section centroid. The wind-adjusted CPUR density metric (W-CPUR) was computed for each

insecticide active ingredient as follows (Equation 3): $W - CPUR_k \left(\frac{\text{kg}}{\text{km}^2}\right) = \sum_{i=1}^n \frac{\left(\left(\frac{A_i}{T_j}\right) * X_j * W_j\right)}{\text{buffer area } (km)^2}$

where k is the insecticide active ingredient used in n sections intersected by the buffer around the residence, A_j is the acreage of section j within the buffer, T_j is the total acreage of section j, X_j is the kilograms of insecticide applied in the section j, and W_j is the proportion of days the home was within 90° downwind of the centroid of section j during the specific time period. Daily average wind direction estimated at the centroid of each PLSS was assumed constant over the entire PLSS.

2.5 Statistical Analyses

We computed detection frequencies and determined that the insecticide dust concentrations (ng/g) were not normally distributed; therefore, non-parametric tests were used to compare distributions. For insecticides that had at least a 40% detection rate in the dust (cypermethrin, propoxur, carbaryl, diazinon, chlorpyrifos, permethrin), we used a single imputation method (Lubin et al. 2004) to assign values for each sample where the insecticide was below the detection limit. We included isomers of each insecticide as covariates in the imputation model; all insecticide isomers were summed (cyfluthrin, cypermethrin, tetramethrin, permethrin) for analysis.

We examined the number of residences with agricultural insecticide applications within each of the 0.5km to 4km buffers. Four insecticides were excluded from further analysis due to their limited applications in the buffers surrounding homes in our study population (allethrin (zero homes), deltamethrin (1–6 homes within 0.5–4km), propoxur (zero homes), and tetramethrin (1 home within 0.5–4km). For the other nine insecticides, we calculated the median density (kg/km²) within each buffer for each metric (CPUR, CROP-A, and W-CPUR). We computed Spearman rank correlations to evaluate the associations between the three metrics for each insecticide, buffer, and time period. Then we used the 365-day CPUR metric to compare dust detection frequencies among participants with and without insecticide applications, and for the five insecticides detected in 40% of dust samples we compared median dust concentrations.

For active ingredients detected in <40% of dust samples (azinphos-methyl, malathion, cyfluthrin, phosmet), we used logistic regression to individually model the odds of each insecticide being detected in the dust (detect vs. non-detect; dependent variable). For active ingredients detected in >40% of dust samples (cypermethrin, carbaryl, diazinon, chlorpyrifos, permethrin), we used Tobit regression to individually model the natural log transformed dust concentrations (dependent variable) and exponentiated the β -coefficients

to show the associated unit increase in the predicted dust value. In both the logistic and Tobit regression models we assessed the CPUR, CROP-A, and W-CPUR metrics (independent variables) separately for each active ingredient as an indicator variable (any applications vs. none) for each buffer and time period. In addition, we categorized each metric as no use within the buffer, median, and > median for insecticides with 10-25% of homes with potential exposure, and as tertiles with a separate non-exposed category for insecticides with more than 25% of homes exposed. Median and tertile cutpoints were based on the distribution of the CPUR metric. We examined the following covariates using a forward selection approach: year (continuous variable from 2001 to 2006, centered by subtracting 2000) and season (winter, spring, summer, and fall) of dust collection, and binary variables for self-reported usual removal of shoes before entering the home, home and garden use of products for the control of specific pests (ants, bees, fleas/ticks, and flies) and weeds, and agricultural occupations. Any covariate with a p-value 0.1 was retained in the final multivariable model. For each insecticide, we examined associations for consistency between time periods, buffer sizes, and metrics, with presentation of the strongest statistically significant associations for each insecticide in the main results table. All analyses were conducted in SAS version 9.4 (Cary, NC).

3. Results

The insecticides with the lowest detection frequencies were allethrin (7.4%), azinphosmethyl (7.5%), and malathion (8.7%); whereas, detections were highest for diazinon (80.4%), chlorpyrifos (89.6%), and permethrin (99.8%) (Table 1). Among homes with these insecticides detected in the dust, median concentrations ranged from 15.9 ng/g for diazinon to 1069.5 ng/g for permethrin.

Agricultural insecticide applications in the 365 days before dust collection (Table 2) were most frequent for chlorpyrifos in each buffer (CPUR metric: 0.5km: 20%, 2km: 36%, and 4km: 48%); whereas azinphos-methyl was the least applied. For the 0.5 km buffer, the median density (CPUR metric) in exposed homes ranged from 0.1 kg/km² for cyfluthrin to 6.5 kg/km² for phosmet. For the 4km buffer, the densities ranged from 0.03 kg/km² for cyfluthrin to 3.4 kg/km² for chlorpyrifos. For the CROP-A metric, the proportion of exposed homes was smaller but most median densities were similar to those for the CPUR metric. The W-CPUR metric generally had the lowest density compared to the CPUR and CROP-A metrics, and the prevalence of exposure was generally lower than that of the CPUR metric, but higher than the CROP-A metric. As expected, the number of applications and densities decreased for 30, 60, and 180 days before dust collection (data not shown).

We evaluated the Spearman rank correlations between the pesticide use density metrics for each insecticide (Supplemental Table 1). Correlations for the 30-day period were similar to those for 60-days, and correlations for the 180-day period were similar to those for 365-days. Therefore, we present results only for 60-day and 365-day metrics. Correlations between CPUR and CROP-A were high (ρ_s 0.67) for all the insecticides except for 60-day carbaryl and 60- and 365-day diazinon, malathion, and permethrin in the 0.5km buffer. Likewise, correlations between CPUR and W-CPUR were high (ρ_s 0.67) except for 60-day and 365-day cypermethrin and phosmet metrics in the 0.5km buffers and 60-day azinphos-

methyl, carbaryl, cypermethrin, and malathion metrics in the 2km buffers. Correlations between CROP-A and CPUR-W were $\rho_s{<}0.67$ for carbaryl, chlorpyrifos, cypermethrin, diazinon, malathion, and phosmet (0.5km/60day metrics) and for azinphos-methyl, carbaryl, cypermethrin, and malathion (2km/60-day metrics). All correlations were high between all the metrics in the 4km buffer.

For the five insecticides with >40% detections, we compared concentrations in homes with and without nearby use based on the CPUR metric in the prior 365 days. Generally, median concentrations in exposed homes decreased slightly (carbaryl, chlorpyrifos, cypermethrin) or stayed the same (diazinon, permethrin) with increasing buffer size. In the 0.5km buffer, chlorpyrifos, cypermethrin, and diazinon concentrations were 2–4 times higher in homes with agricultural applications compared to homes without (Figure 4). Median carbaryl, chlorpyrifos, and diazinon concentrations were 1.5–2 times greater in homes with applications, whereas cypermethrin concentrations were three times greater in homes with agricultural applications within the 1–4km buffers. Permethrin concentrations did not differ among homes with and without nearby insecticide use (not shown).

When we evaluated 30-, 60- and 180-day time periods between application and dust for these insecticides, only carbaryl showed substantial differences from the 365 results. In the 30 and 60 days before dust collection, carbaryl concentrations were 2.5–11 times greater in homes with applications (Supplemental Table 2). The only other notable differences from the 365-day results were for cypermethrin and permethrin. Homes with cypermethrin applications within 180 days in the 4km buffer had 3-times higher median dust concentrations compared to homes without applications (p=0.04), but there were no differences for the other buffers and time periods (not shown). Permethrin concentrations in the 60-day period within 2 km and in the 30-day period within 2–4 km, were 2 times higher among unexposed homes (p<0.05; data not shown).

Results of multivariable models for carbaryl, chlorpyrifos, cypermethrin, and diazinon for 60-day CPUR, CROP-A, and W-CPUR metrics are shown in Table 3. All three metrics were associated with concentrations of carbaryl, diazinon, and chlorpyrifos dust concentrations. For carbaryl, the metrics were associated with 3–5 times significantly greater concentrations in the 4km buffers and the linear trend across density categories was only significant for the W-CPUR metric (p-trend=0.04; Table 3). In the 2km buffer, the 60-day metrics were associated with elevated concentrations. Year, season, and shoe removal were inversely associated and use of flea or tick products and treatments for lawn/garden weeds were positively associated with carbaryl concentrations. Associations for the 365-day metrics were positive but weaker than the 60-day metrics and none were significant (Supplemental Table 3).

For diazinon, we observed a 2- to 2.5-fold increase with significant linear trends for the CPUR and W-CPUR metrics in the 4km buffer and for all metrics in the 2 km buffer (Table 3). In the 1km buffer, the CROP-A metric was a predictor of dust concentrations, whereas the CPUR and the W-CPUR metrics were positive but not statistically significant. The 365-day metrics in the 1km buffer were predictors of dust concentrations but only the

W-CPUR metric had a significant linear trend (Supplemental Table 3). In the other buffers, we observed elevated concentrations for all metrics, but none of the trends were significant.

For chlorpyrifos, all three 4km metrics showed associations with concentrations that were of similar magnitude with significant trends across density tertiles (Table 3). In the 1km buffer, the CPUR and W-CPUR metrics were predictors compared to the CROP-A metric. The 365-day chlorpyrifos metrics showed similar patterns to the 60-day metrics and similar magnitudes of association with >median and the highest tertile categories (Supplemental Table 3).

For cypermethrin, none of the metrics predicted dust concentrations but household use of flea/tick products, and treatments for ants and flies were positively associated with concentrations (Table 3). Results for the 365-day metric were similar (Supplemental Table 3).

None of the CPUR metrics during any period or buffer size were predictors of permethrin concentrations (Supplemental Table 3). Year of dust collection, use of flea or tick products, and treatments for ants, bees, and flies were positively associated with permethrin dust concentrations.

For the four insecticides detected in <40% of homes, the percent detections in homes with and without CPUR applications in the prior 365 days are shown in Supplemental Table 4. Azinphos-methyl was detected more frequently in exposed homes compared to unexposed homes for the 2–4km buffers. Cyfluthrin detections were more frequent in exposed homes in the 0.5 and 1km buffers, but not the larger buffers. There was no difference in the proportion detected in exposed and unexposed homes for malathion for any buffer. For phosmet, exposed homes had higher detection frequencies than unexposed homes for all buffers. The patterns were similar for earlier time periods (not shown).

With these four insecticides, we did not have sufficient exposure data to model more than a yes/no agricultural exposure variable for the 60-day metrics, therefore we present the results of the 365-day metrics. For cyfluthrin, the 365-day 0.5km CPUR, CROP-A and W-CPUR metrics showed the strongest associations with the odds of detection of cyfluthrin, and the 4km metrics showed significant trends with increasing density except for the cyfluthrin W-CPUR metric (Table 4). For the 60-day metrics (Supplemental Table 5) odds of detection were about six-fold higher for the 0.5km CPUR and CROP-A metrics, but there was no association with the W-CPUR metric. Point estimates were non-significant for the other buffers for all of the metrics. Year of dust collection, use of flea or tick products, and treatments for lawn/garden weeds were positively associated with cyfluthrin dust detections.

For phosmet, the CPUR, CROP-A and W-CPUR 1, 2 and 4km metrics were each associated with 6–8 times higher odds of dust detection (all p-values for trend <0.0001; Table 4). Each of the 0.5km buffer metrics was associated with 4–5 times higher odds of detection. The 60-day metrics (Supplemental Table 5) showed similar associations with higher odds of detection compared to unexposed homes for all metrics, although ORs were somewhat stronger than the 365-day estimates. Year, season of dust collection, and occupation as a farmworker were positively associated with dust detection.

For azinphos-methyl, the 4km 365-day metrics were predictors of dust detections with significant trends across the median exposure categories (Table 4). There was no association with the 0.5km-1km metrics, but all metrics were predictors of dust detections in 2km buffer. We were limited in our ability to examine the metrics using the 60-day period due to the small number of homes with azinphos-methyl applications (0.5km: 2 exposed homes; 4km: 31 exposed homes; Supplemental Table 5). No covariates were predictors of azinphos-methyl concentrations.

There were no associations with any metric for malathion in the 365- (not shown) or 60-day time periods (Supplemental Table 5). Home use of products for the control of bees and lawn/garden weeds, as well as having someone in the home who identified as a farmworker were predictors of malathion detections.

4. Discussion

We estimated agricultural insecticide use around participant's homes and evaluated whether nearby agricultural applications were predictors of concentrations of insecticide active ingredients in house dust. Our analyses expand upon prior work by Gunier et al. (2011) in this study population by including additional insecticide active ingredients and using data from a greater number of homes (598 vs. 89). In addition to a CPUR metric, we created two additional metrics using information on agricultural land use and wind direction contemporaneous with the time between insecticide application and dust collection. We showed that agricultural applications of carbaryl, diazinon, chlorpyrifos, cyfluthrin, phosmet, and azinphos-methyl were associated with concentrations or detections in the home. Comparing the three different metrics across four time periods and four buffer sizes using multivariable models, we found that varying the time between agricultural use and dust collection and adjusting for wind direction was particularly important for carbaryl applications. Generally, wind-adjusted carbaryl applications within 2–4km of homes within 30- and 60-days of dust collection showed the strongest associations.

We evaluated insecticide applications during the 30-, 60-, 180-, and 365-day periods prior to dust collection. Varying the time period did not impact the associations with cypermethrin, permethrin, and malathion applications that were not associated with dust levels during any time period, while chlorpyrifos applications were associated during all time periods. Chlorpyrifos has a field dissipation half-life of around 40 days, which may be why we observed an association during any time period (USDA Agricultural Research Service 1995b). For carbaryl and diazinon, associations differed by the time period. Consistent with our prior work (Gunier et al. 2011), we observed no associations between carbaryl applications within 180 and 365-days of dust collection. When we examined shorter periods of 30- and 60-days between application and dust collection, the density of carbaryl use at all buffer sizes was a predictor of the dust concentrations. For diazinon, use in the prior 30- and 60-days was a more consistent predictor of dust concentrations. Both carbaryl and diazinon have short field dissipation halflives of 4–13 and 7 days (USDA Agricultural Research Service 1995a; c), respectively, which may be why we only observed associations for these active ingredients in the shorter time periods.

Numerous prior studies have shown higher levels of pesticides in dust samples taken from homes close to agricultural activities compared to non-proximal homes (Dereumeaux et al. 2020). In a meta-regression of published data, Deziel et al. showed that house dust pesticide concentrations decreased sharply and non-linearly with increasing distance from treated fields that was linear on a log-log scale (Deziel et al. 2017). Using data from 89 (15%) of the homes in our study, Gunier et al. examined a 365- and 180-day CPUR metric and a metric that accounted for the location of crops at the time of the dust sampling and found that chlorpyrifos and phosmet but not carbaryl or diazinon concentrations were higher in homes with agricultural insecticide use within a 1.25km buffer (Gunier et al. 2011). The null findings for carbaryl and diazinon may have been due to the longer time periods between application and dust collection used in the study; in our study we also did not observe associations with carbaryl in the 180- or 365-day time periods. For diazinon, the 60-day metrics were significant predictors of dust concentrations. Several exposure studies demonstrate findings generally consistent with ours, including a California study of 504 house dust samples found that agricultural applications of chlorpyrifos but not diazinon or permethrin within 3 miles (4.8km) of the home were associated with dust concentrations (Harnly et al. 2009). Also consistent with the results of our study is a study of 61 homes that showed those within 200 feet of a pesticide-treated fruit orchard had higher median chlorpyrifos, azinphos-methyl, and phosmet concentrations levels in their house dust compared to homes more than 200 feet from an orchard (Fenske et al. 2002; Lu et al. 2000; Simcox et al. 1995).

We explored associations using buffer sizes ranging from 0.5 to 4km. Numerous studies have shown that exposure to pesticides is greater at closer residential distances to agricultural fields (Dereumeaux et al. 2020; Nuckols et al. 2007; Obendorf et al. 2006; Ward et al. 2006), but little is known about the optimal distance to use for each insecticide. The distance from the source of the agricultural insecticide application may be an important determinant of the presence and concentration of active ingredients in the home environment, and may vary by insecticide, yet many studies that use spatial exposure metrics consider only one buffer size in their analyses. A recent review of residential exposure to agricultural pesticides found that only 25% of studies that used GIS-based exposure metrics explored more than one buffer size (Teysseire et al. 2020). Factors that influence each ingredient's ability to persist and be transported by wind or water flow from agricultural land include temperature, metabolism in plants and soil microorganisms, and the chemical properties of the active ingredient such as half-life in soil, vapor pressure, octanol/water partition coefficient, and water solubility (Bennett et al. 1998; Farha et al. 2016; Gavrilescu 2005; Juraske et al. 2008; Van Eerd et al. 2003). This indicates that an individualized modeling approach may need to be tailored to each pesticide to better understand the association between location of applications and residential exposure. This consideration is supported by our study demonstrating that agricultural applications predicted dust concentrations in the home at different distances and time periods for different insecticides. Our findings highlight the need to explore different exposure time periods and buffer sizes for each active ingredient to accurately characterize the potential for non-occupational human exposure.

In this study we attempted to enhance the exposure assessment based on CPUR use by incorporating data on agricultural land use and wind conditions. Although studies have shown that land use information such as crop field area and distance from the home to a crop field (Nuckols et al. 2007; Rull and Ritz 2003; Vannier et al. 2020) and wind direction (Gunier et al. 2018; Rowe et al. 2016; Sagiv et al. 2019) can be incorporated into exposure metrics for use in epidemiologic studies, few have compared different metrics to demonstrate the utility of incorporating additional information to refine the metric. Offtarget pesticide exposure has been linked to wind conditions (Pfleeger et al. 2006) and modelling studies suggests that pesticide dispersion is strongly affected by meteorological conditions (e.g., wind direction) during application (Costanzini et al. 2018), highlighting the need to evaluate the impact of incorporating wind direction into exposure metrics. In this study, incorporating agricultural land use categories led to similar estimates of insecticide density between the crop-area adjusted and the CPUR metric. Generally, the density estimates for the wind metric were smaller than the crop-adjusted and CPUR metrics, since the weighting we applied reduced the prevalence of exposure. When we assessed correlations between the three metrics, most of the insecticides showed moderate to high correlation between the three metrics across the different buffer sizes and time periods. The weakest correlations were observed with the CROP-A metric, which may be due to limitations in the land use data that was available for linkage. Despite differences in the density estimates among the three metrics, the resulting associations between each metric and dust concentrations were similar to each other with few exceptions. For example, for the 60-day 4km carbaryl estimates only the wind-adjusted metric shows a positive trend. The 60-day diazinon CPUR and wind-adjusted metrics were associated with dust concentrations in the 4km buffer, but the crop-area adjusted metric was not. We did not observe many other differences in the associations between insecticide applications and dust concentrations using the different metrics. This suggests that the simpler CPUR metric could provide a reasonable estimate of exposure for use in epidemiologic studies investigating links between agricultural insecticide use and health outcomes. However, it should be noted that the CPUR metric is based on a robust and mandatory pesticide use reporting program administered only in the State of California, USA. The results of our study underscore the need for similar data resources in all regions where cultivated and residential land uses are highly integrated.

Strengths of this study include the availability of a comprehensive database of agricultural insecticide use for the study area that included detailed information on the types and quantities of insecticides applied to crops including both spot spraying and broadcast spraying for insect control. We used data from a large number of homes, allowing us to investigate relationships with a diverse set of insecticides, including those that were less frequently applied. Further, our study included many homes from both urban, suburban, and rural areas and we were able to vary the time period between insecticide application and dust collection. We had self-reported information on home and garden use of insecticides and whether household members had occupational exposures, allowing us to evaluate additional factors as determinants of the insecticide concentrations in the house dust and control for these factors in multivariate models.

Our study has several limitations. Although we were able to link the pesticide application data with available land use and wind information to further refine our exposure metrics, the

accuracy and resolution of these data impacts the quality of the subsequent exposure metrics. Our dust collections spanned 2001–2007, but land use information was only available for 2001, 2004, and 2006. California agriculture is characterized by multiple crops per field within an annual cycle as well as relatively frequent replacement/interchange of orchards, berries, and grapes within a short time based on market demand. The land use data may not have accurately captured crop locations for crops that were rotated during months/ years without available land use data. Prior studies have accounted for potential downwind pesticide drift from application sites to homes by incorporating historical wind direction data from the nearest meteorological station to each residence (Gunier et al. 2018; Rowe et al. 2016; Sagiv et al. 2019). These studies used a 45° wedge, which is more conservative than our 90° wedge. In our study we modeled the wind from the source to the residence and ascertained wind direction from a low-resolution atmospheric and land surface hydrology dataset available at a 32km scale. Given the scale of our data, it is possible that data from the nearest meteorological station may have resulted in different estimates of the number of days downwind, but station data is also limited by the variation in distance between the station and the residence. We did not account for wind speed in our wind-adjusted metric, a factor that could also impact pesticide drift. Our study relates agricultural insecticide use to concentrations measured in house dust, and it is not clear how levels in house dust relate to levels measured in biological samples taken from residents of the home. Research is needed to relate dust levels in the home to exposures measured in biological samples. Our findings for some insecticides (e.g., cypermethrin, phosmet) were limited by sparse data due to infrequent applications during the shorter time periods and/or in smaller buffer sizes. For example, although cypermethrin was detected in 49% of dust samples, this level of detection is likely due to its use in products around the home as only 28 homes (5%) had applications in the 0.5km buffer within the 365 days before dust collection.

5. Conclusions

For six of the nine insecticides, agricultural use within 4km of a home was a significant determinant of indoor contamination. Dust concentrations and detections were higher in homes with nearby agricultural insecticide use compared to homes without use for carbaryl, diazinon, chlorpyrifos, cyfluthrin, phosmet, and azinphos-methyl. Our findings demonstrated the utility of GIS-based metrics for quantifying potential exposure to fugitive insecticide emissions from cultivated agriculture but indicated that associations with measured levels of insecticides in homes varies depending on buffer size (i.e., defined proximity) and the time elapsed between application and house dust collection. Our findings suggest inclusion of wind enhanced prediction for some, but not all insecticides studied. Taken together, our results imply that GIS-based exposure metrics used in epidemiologic studies should be tailored to the fate and transport characteristics of each insecticide.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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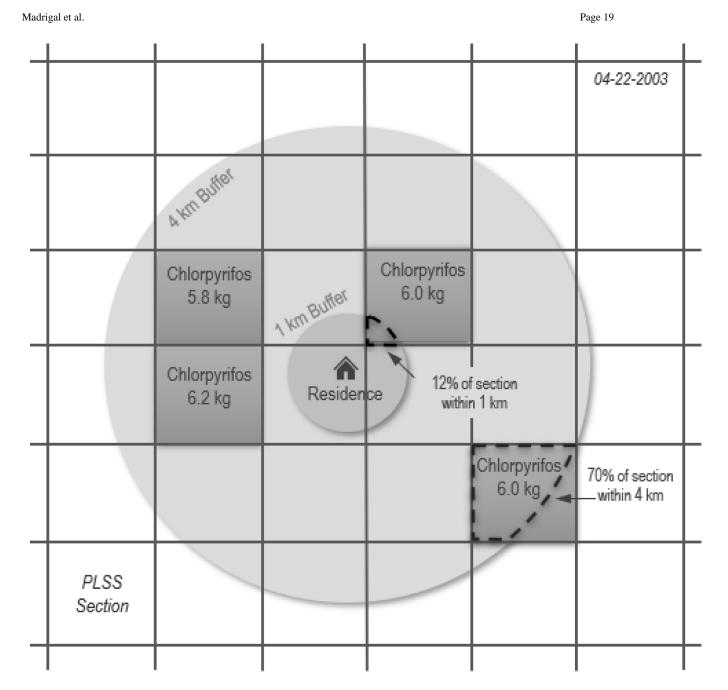


Figure 1. Illustration of the CPUR density metric for the 1km and 4km buffer around the residence on one day for four application sites. In the 1km buffer, one application of chlorpyrifos is included after weighting the 6.0 kg applied by 12% due to most of the public land survey section (PLSS) being outside of the buffer. In the 4km buffer, three applications of chlorpyrifos are included in entirety and one application is weighted by 70% due to 30% of the PLSS being outside of the buffer.

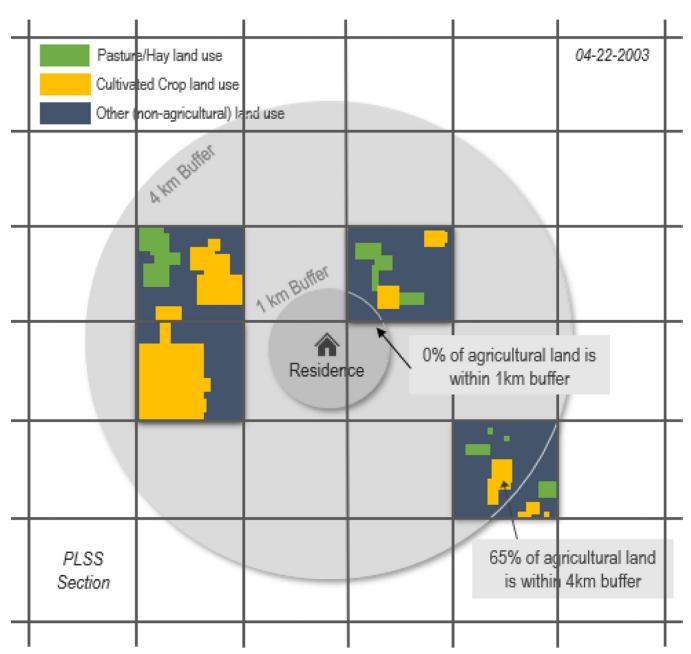


Figure 2. Illustration of the CROP-A metric for the 1km and 4km buffer around the residence on one day for four application sites. The applications of chlorpyrifos are matched to the acres of pasture/hay and cultivated crops identified in each public land survey section (PLSS) within the buffer. The metric is refined by weighting the amount of chlorpyrifos applied by the proportion of pasture/hay and cultivated crops (i.e., agricultural land) identified in each

PLSS within the buffer.

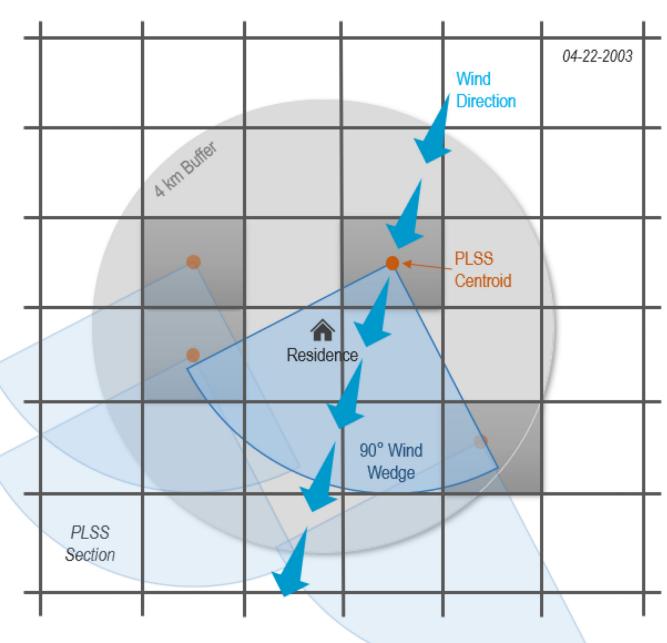


Figure 3.

Illustration of the W-CPUR metric within a 4km buffer around the residence on one day for four application sites. The wind direction is determined for each day between application and dust collection for each public land survey section (PLSS) centroid with a chlorpyrifos application within the buffer to calculate the proportion of days the home was downwind of the application site.

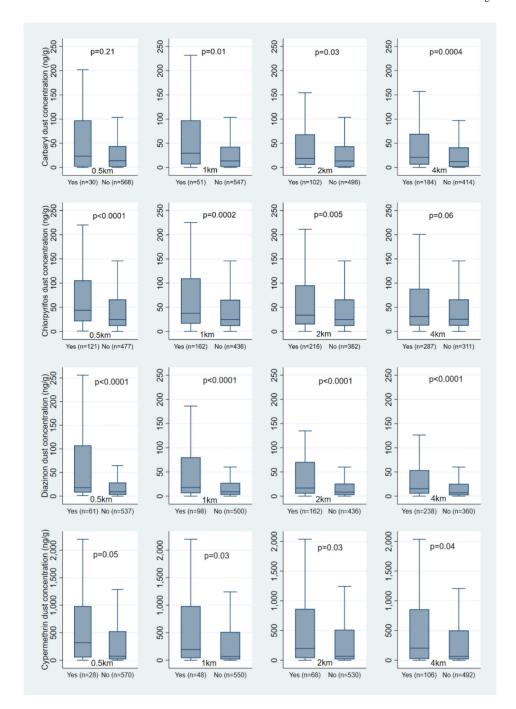


Figure 4.
Distributions of carbaryl, chlorpyrifos, diazinon, and cypermethrin concentrations in house dust of homes with (Yes) and without (No) nearby agricultural use in the prior 365 days Yes=Homes with CPUR applications (exposed); No=Homes without CPUR applications (unexposed); p-values from Wilcoxon rank sum test used to compare the distribution of the insecticide concentration in the dust among homes classified as exposed and those classified

as unexposed; dust concentrations with values 1.5 times the IQR above the upper quartile or below the lower quartile were excluded from the graphs.

Table 1.

Detection limits, percent detections, and concentrations for 13 agricultural insecticides measured in 598 dust samples from California homes

| Insecticide | Detection limit (ng/g) | Detected (%) | Median (interquartile range) concentration among homes with detection $({\rm ng/g})^d$ |
|------------------------|------------------------|--------------|--|
| Allethrin ^b | 20 | 7.4 | 251.8 (86.1–615.8) |
| Azinphos-methyl | 100 | 7.5 | 126.2 (80.3–219.7) |
| Malathion | 10 | 8.7 | 93.7 (58.0–192.1) |
| Deltamethrin | 50 | 11.9 | 452.1 (182.8–1046.3) |
| Tetramethrin b | 2 | 12.2 | 116.9 (39.8–350.3) |
| Cyfluthrin b | 20 | 26.1 | 468.8 (221.4–946.8) |
| Phosmet | 25 | 28.1 | 22.4 (11.1–72.9) |
| Cypermethrin b | 20 | 49.5 | 569.6 (251.4–1366.1) |
| Propoxur | 5 | 66.7 | 22.3 (9.2–53.8) |
| Carbaryl | 2 | 68.9 | 30.2 (13.3–91.9) |
| Diazinon | 2 | 80.4 | 15.9 (6.2–45.1) |
| Chlorpyrifos | 5 | 89.6 | 32.9 (15.8–87.7) |
| Permethrin b | 2 | 99.8 | 1069.5 (387.5–4296.0) |

 $^{^{\}it a}_{\rm Distribution}$ among residences with insecticide concentrations above the detection limit

 $b_{\text{Isomers have been summed}}$

Table 2.

Percent of homes with insecticide applications and density of insecticide as (kg/km²) within 0.5km, 2km, and 4km among homes with CPUR applications within 365 days before dust collection for three exposure metrics (CPUR^b, CROP-A^c, and W-CPUR^d)

| Insecticide | Metric | Residences within 0.5km, n (%) | Median (IQR) density of use 0.5km, kg/km² | Residences within 2km, n (%) | Median (IQR) density of use 2km, kg/km ² | Residences within 4km, n (%) | Median (IQR) density of use 4km, kg/km² |
|---------------------|--------|-----------------------------------|--|---------------------------------|---|---------------------------------|---|
| Azinphos- methyl | CPUR | 13 (2.2) | 1.8 (1.1–5.6) | 49 (8.2) | 0.7 (0.2–2.4) | 101 (16.9) | 0.6 (0.1–1.1) |
| | CROP-A | 13 (2.2) | 2.2 (0.2–8.0) | 43 (7.2) | 0.7 (0.3–2.0) | 97 (16.2) | 0.6 (0.1–1.1) |
| | W-CPUR | 12 (2.0) | 0.5 (0.2–0.9) | 41 (6.9) | 0.1 (0.01–0.5) | 91 (15.2) | 0.06 (0.01–0.3) |
| Carbaryl | CPUR | 30 (5.0) | 2.5 (0.9–11.4) | 102 (17.1) | 0.4 (0.1–2.2) | 184 (30.8) | 0.4 (0.05–1.5) |
| | CROP-A | 23 (3.8) | 2.0 (0.7–12.6) | 91 (15.2) | 0.4 (0.1–3.1) | 173 (28.9) | 0.4 (0.1–1.5) |
| | W-CPUR | 26 (4.3) | 0.7 (0.05–2.0) | 95 (15.9) | 0.1 (0.01–0.3) | 171 (28.6) | 0.1 (0.01–0.3) |
| Chlorpyrifos | CPUR | 121 (20.2) | 5.5 (1.2–14.0) | 216 (36.1) | 3.4 (0.7–12.4) | 287 (48.0) | 3.4 (0.3–10.4) |
| | CROP-A | 95 (15.9) | 5.6 (1.3–16.8) | 193 (32.3) | 4.6 (1.2–12.8) | 265 (44.3) | 3.9 (0.5–12.1) |
| | W-CPUR | 114 (19.1) | 1.0 (0.13–3.3) | 212 (35.4) | 0.7 (0.1–3.1) | 280 (46.8) | 0.7 (0.04–3.3) |
| Cyfluthrin | CPUR | 36 (6.0) | 0.1 (0.02–0.3) | 100 (16.7) | 0.03 (0.004–0.1) | 168 (28.1) | 0.03 (0.003–0.1) |
| | CROP-A | 29 (4.8) | 0.08 (0.03–0.3) | 91 (15.2) | 0.04 (0.004–0.1) | 154 (25.7) | 0.03 (0.004–0.1) |
| | W-CPUR | 31 (5.2) | 0.01 (0.001–0.05) | 95 (15.9) | 0.003 (0.001–0.01) | 162 (27.1) | 0.003 (0.0003–0.02) |
| Cypermethrin | CPUR | 28 (4.7) | 0.2 (0.07–1.0) | 68 (11.4) | 0.2 (0.05-0.7) | 106 (17.7) | 0.1 (0.02–0.4) |
| | CROP-A | 19 (3.2) | 0.2 (0.05–0.6) | 59 (9.9) | 0.2 (0.1-0.6) | 99 (16.6) | 0.1 (0.02–0.4) |
| | W-CPUR | 26 (4.3) | 0.04 (0.01–0.11) | 62 (10.4) | 0.02 (0.01-0.1) | 96 (16.0) | 0.02 (0.003–0.1) |
| Diazinon | CPUR | 61 (10.2) | 3.0 (0.4–7.3) | 162 (27.1) | 1.0 (0.1–4.2) | 238 (40.0) | 0.8 (0.05–3.5) |
| | CROP-A | 42 (7.0) | 3.5 (0.8–7.7) | 139 (23.2) | 1.2 (0.2–4.8) | 211 (35.3) | 1.0 (0.1–4.2) |
| | W-CPUR | 57 (9.5) | 0.4 (0.06–2.1) | 158 (26.4) | 0.1 (0.01–0.8) | 232 (38.8) | 0.1 (0.005–0.7) |
| Malathion | CPUR | 30 (5.0) | 1.9 (0.2–14.6) | 98 (16.4) | 0.7 (0.09–5.2) | 189 (31.6) | 0.3 (0.04–1.7) |
| | CROP-A | 24 (4.0) | 1.6 (0.2–13.4) | 87 (14.5) | 0.7 (0.1–5.1) | 170 (28.4) | 0.4 (0.1–2.0) |
| | W-CPUR | 29 (4.8) | 0.2 (0.03–3.3) | 95 (15.9) | 0.1 (0.01–0.9) | 175 (29.3) | 0.05 (0.004–0.4) |
| Permethrin | CPUR | 48 (8.0) | 0.6 (0.1–2.3) | 137 (22.9) | 0.1 (0.02–0.7) | 209 (35.0) | 0.1 (0.02–0.7) |

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| Insecticide | Metric | Residences within 0.5km, n (%) | Median (IQR) density of Residences within use 0.5km, kg/km ² 2 km, n (%) | Residences within 2km, n (%) | Median (IQR) density of Residences within use 2km, kg/km 2 4km, n (%) | Residences within 4km, n (%) | Median (IQR) density of use 4km, kg/km² |
|-------------|--------|--------------------------------|---|---------------------------------|--|---------------------------------|---|
| | CROP-A | 37 (6.2) | 0.5 (0.1–2.7) | 127 (21.2) | 0.1 (0.02–0.8) | 200 (33.4) | 0.1 (0.02–0.7) |
| | W-CPUR | 45 (7.5) | 0.07 (0.02–0.57) | 128 (21.4) | 0.04 (0.005–0.2) | 201 (33.6) | 0.03 (0.003–0.2) |
| Phosmet | CPUR | 48 (8.0) | 6.5 (0.8–17.7) | 112 (18.7) | 2.5 (0.5–11.7) | 178 (30.0) | 1.4 (0.2–7.5) |
| | CROP-A | 38 (6.3) | 7.1 (3.0–19.1) | 103 (17.2) | 3.6 (0.5–12.9) | 165 (27.6) | 1.8 (0.3–7.7) |
| | W-CPUR | 46 (7.7) | 1.2 (0.1–3.6) | 106 (17.7) | 0.3 (0.03–2.1) | 166 (27.8) | 0.4 (0.03–2.0) |

²Four insecticides were excluded from further analysis due to their limited applications in the buffers surrounding homes in our study population (allethrin (zero homes), deltamethrin (1–6 homes within 0.5–4km)

 b CPUR= CPUR Metric (kg/km 2); insecticide use density proportional to the area of the buffer

CROP-A= Crop area adjusted CPUR Metric (kg/km²): insecticide use density proportional to the area of cultivated crops and pasture/hay in the buffer

d/W-CPUR= Wind-adjusted CPUR Metric (kg/km²): insecticide use density proportional to the area of the buffer, weighted to account for the proportion of days the residence was downwind of the section in which the insecticide was applied during the period between application and dust collection

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Table 3.

Multivariable associations of 60-day insecticide density metrics (kg/km²) with concentrations of four insecticides detected in 40% of house-dust samples

| Insecticide | Buffer | 60-day CPUR ^b | | 60-day CROP-A ^C | s | 60-day W-CPUR ^d | p |
|-----------------------------------|--------|---------------------------------|--|---------------------------------|--|---------------------------------|---|
| | | Exposure density (kg/km²) vs. 0 | Exp \(\beta \) (95% CI) | Exposure density (kg/km²) vs. 0 | Exp \(\beta \) (95% CI) | Exposure density (kg/km²) vs. 0 | Exp \(\beta \) (95% CI) |
| | 0.5km | 9 | 0.8 (0.1–9.6) | 0< | 0.3 (0.0–8.3) | 0× | 4.9 (0.2–100.4) |
| | Ikm | 0× | 0.8 (0.1–5.5) | 0< | 1.6 (0.2–15.4) | 0× | 1.3 (0.1–14.4) |
| Cypermethrin ^e | 2km | 9 | 1.4 (0.3–6.5) | 0< | 1.5 (0.3–7.6) | 0× | 1.9 (0.3–10.9) |
| | 4km | 9 | 1.7 (0.5–5.8) | 0< | 2.3 (0.7–7.7) | 0× | 1.3 (0.3–5.5) |
| | 0.5km | 0× | 10.3 (0.7–143.0) | 0< | 10.3 (0.7–143.0) | 0 | 27.8 (0.8–913.1) |
| | Ikm | 9 | 6.4 (0.8–53.9) | 0< | 6.4 (0.8–53.9) | 0× | 8.5 (0.7–103.2) |
| $\operatorname{Carbaryl}^f$ | 2km | 9 | 6.3 (1.7–24.0) | 0< | 5.1 (1.2–21.1) | 0× | 6.9 (1.3–36.0) |
| | 4km | 0.11 > 0.11 | 4.2 (1.3–14.2) 3.4 (1.0–12.2) p-trend=0.05 | 0.13 | 5.0 (1.4-17.4) 3.1 (0.9-11.3) p-trend=0.08 | 0.04 > 0.04 | 1.8 (0.4–8.1) 4.7 (1.1–20.5) p-trend=0.04 |
| | 0.5km | 0× | 3.1 (1.3–7.6) | 0< | 2.5 (0.9–7.3) | 0× | 2.8 (1.1–7.5) |
| | Ikm | 0× | 1.7 (0.9–3.3) | 0< | 2.5 (1.2–5.2) | 0× | 1.3 (0.6–2.6) |
| Diazinon $^{\mathcal{G}}$ | 2km | 0.4 | 0.9 (0.4–1.9) 2.5 (1.2–4.9) p-trend=0.01 | 0.5 | 1.2 (0.6–2.4) 2.5 (1.2–5.1) p-trend=0.01 | 0.11 | 0.8 (0.4–1.9) 2.4 (1.1–5.3) p-trend=0.03 |
| | 4km | 0.3 | 1.4 (0.8–2.5) 2.0 (1.1–3.5) p-trend=0.03 | 0.4 | 1.8 (1.0–3.4) 1.8 (1.0–3.3) p-trend=0.07 | 0.05 | 1.4 (0.7–2.7) 2.2 (1.1–4.1) p-trend=0.02 |
| | 0.5km | 0< | 1.4 (0.9–2.2) | 0< | 1.2 (0.7–2.1) | 0< | 1.1 (0.6–1.9) |
| $\operatorname{Chlorpyrifos}^{h}$ | Ikm | 1.3 | 1.1 (0.7–1.9) 1.8 (1.0–3.0) p-trend=0.04 | 0.9 | 1.3 (0.7–2.4) 1.5 (0.9–2.7) p-trend=0.14 | 0.3 > 0.3 | 0.9 (0.5–1.6) 1.8 (1.0–3.4) p-trend=0.06 |

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| Insecticide | Buffer | $60\text{-}\mathrm{day}~\mathrm{CPUR}^{b}$ | | 60-day CROP-A ^C | 9 | 60-day W-CPUR ^d | p |
|-------------|--------|--|---|---------------------------------|--|--|--|
| | | Exposure density (kg/km²) vs. 0 | Exp \(\beta \) (95% CI) | Exposure density (kg/km²) vs. 0 | Exp \(\beta \) (95% CI) | Exposure density (kg/km²) vs. 0 Exp β (95% CI) Exposure density (kg/km²) vs. 0 Exp β (95% CI) Exposure density (kg/km²) vs. 0 Exp β (95% CI) | Exp \(\begin{align*} |
| | 2кт | 0.93 > 0.93 | 1.0 (0.7–1.5) 1.4 (0.9–2.2) p-trend=0.09 | == ^ | 1.2 (0.8–1.9) 1.5 (1.0–2.3) p-trend=0.08 | 0.3 | 1.0 (0.6–1.6) 1.3 (0.8–2.1) p-trend=0.25 |
| | 4km | 0.23 0.24–1.7 > 1.7 | 0.8 (0.5–1.2) 1.1 (0.7–1.7) 1.7 (1.1–2.6) p-trend=0.01 | 0.30 0.31–1.9 > 1.9 | 0.9 (0.6–1.4) 1.1 (0.7–1.8) 1.8 (1.2–2.9) p-trend=0.006 | 0.05 0.06-0.6 > 0.6 | 0.8 (0.5–1.3) 1.4 (0.9–2.3) 1.9 (1.2–2.9) p-trend=0.006 |

W-CPUR) comparing >0 or density categories to homes with no use. A forward approach was used to evaluate covariates. Any covariate with a p-value 0.1 was retained in the final multivariable model and insecticide, three separate Tobit regression models were used to model the natural log transformed dust concentrations (dependent variable) with each density metric (CPUR, CROP-A, and

 b CPUR= CPUR Metric (kg/km 2); insecticide use density proportional to the area of the buffer

CROP-A= Crop area adjusted CPUR Metric (kg/km²); insecticide use density proportional to the area of cultivated crops and pasture/hay with insecticide use in the buffer

d. CPUR= Wind-adjusted CPUR Metric (kg/km²): insecticide use density proportional to the area of the buffer, weighted to account for the proportion of days the residence was downwind of the section in which the insecticide was applied during the period between application and dust collection

e Adjusted for the use of flea or tick products, and treatments for ants and flies that were positively associated with cypermethrin concentrations.

f
Adjusted for year, season, and shoe removal that were inversely associated with carbaryl concentrations and use of flea or tick products, and treatments for lawn/garden weeds that were positively associated with carbaryl concentrations.

g Adjusted for year and season that were inversely associated with concentrations and use of flea or tick products and farm working occupation that were positively associated with concentrations.

 h Adjusted for year that was inversely associated with concentrations and farm working occupation that was positively associated

Table 4.

Multivariable associations ^a of 365-day insecticide metrics with detections of three insecticides detected in <40% of house-dust samples

| | | 365-6 | lay CPUR ^b | 365-da | ny CROP-A ^c | 365-da | y W-CPUR ^d |
|--------------------------------------|---------|--|--|--|--|--|--|
| Insecticide | Buff er | Exposure density (kg/ km²) vs. 0 | OR (95% CI) detect vs. non- detect | Exposure density (kg/ km²) vs. 0 | OR (95% CI) detect vs. non- detect | Exposure density (kg/ km²) vs. 0 | OR (95% CI) detect vs. non- detect |
| | 0.5km | >0 | 3.0 (1.5–6.0) | >0 | 2.5 (1.1–5.4) | >0 | 2.4 (1.1–5.0) |
| | 1km | >0 | 1.7 (1.0–3.1) | >0 | 1.6 (0.8–2.9) | >0 | 1.6 (0.8–2.9) |
| | 2km | 0.03 | 1.1 (0.6–2.2) | 0.04 | 1.3 (0.6–2.5) | 0.003 | 1.3 (0.6–2.4) |
| Cyfluthrin ^e | | > 0.03 | 1.9 (1.0-3.6) | > 0.04 | 1.8(0.9–3.4) | > 0.003 | 1.5(0.8–2.9) |
| Cynumrin | | | p-trend=0.04 | | p-trend=0.10 | | p-trend=0.22 |
| | 4km | 0.007 0.008-0.05 > 0.05 | 1.0 (0.5–1.9) 1.0 (0.5–1.9) 2.2 (1.2–4.0) | 0.011 0.12-0.06 > 0.06 | 1.1 (0.6–2.2) 0.9 (0.4–1.8) 2.4 (1.3–4.4) | <0.001 0.001-0.01 > 0.01 | 1.0 (0.5–2.0) 1.4 (0.7–2.6) 1.5 (0.8–2.8) |
| | | | p-trend=0.01 | | p-trend=0.01 | | p-trend=0.19 |
| | 0.5km | >0 | 4.9 (2.6–9.4) | >0 | 5.5 (2.7–11.3) | >0 | 4.5 (2.3–8.7) |
| | 1km | 3.4 | 1.9 (0.9–4.2) | 4.0 | 3.1 (1.4–7.0) | 0.7 | 2.0 (0.9–4.5) |
| | | > 3.4 | 7.7 (3.6–16.4) | > 4.0 | 7.5 (3.4–16.9) | > 0.7 | 6.4 (3.0–13.8) |
| | | | p-trend=<0.0001 | | p-trend=<0.0001 | | p-trend=<0.000 |
| Phosmet f | 2km | 2.4 | 3.2 (1.7–6.0) | 3.6 | 3.6 (1.9–6.9) | 0.3 | 3.0 (1.6–5.7) |
| | | > 2.4 | 7.0 (3.7–13.3) p-trend=<0.0001 | > 3.6 | 7.3 (3.7–14.2) p-trend=<0.0001 | > 0.3 | 7.6 (3.9–14.7) p-trend=<0.0003 |
| | 4km | 0.4 0.5–4.9 > 4.9 | 1.2 (0.6–2.4) 2.4 (1.3–4.6) 6.4 (3.4–11.9) | 0.4 0.5–5.4 > 5.4 | 1.2 (0.6–2.5) 3.2 (1.7–6.2) 6.6 (3.5–12.7) | < 0.07 0.07-1.1 > 1.1 | 0.6 (0.3–1.5) 3.6 (1.9–6.9) 6.3 (3.3–12.1) |
| | | | p-trend=<0.0001 | | p-trend=<0.0001 | | p-trend=<0.000 |
| | 0.5k | >0 | 1.0 (0.1–8.1) | >0 | 1.0 (0.1–8.1) | >0 | 1.1 (0.1–8.9) |
| | 1km | >0 | 2.0 (0.6–7.1) | >0 | 1.5 (0.3–6.6) | >0 | 2.3 (0.6–8.0) |
| Azinphos- methyl $^{\mathcal{G}}$ | 2km | >0 | 2.7 (1.2–6.2) | >0 | 3.2 (1.4–7.4) | >0 | 2.8 (1.2–6.8) |
| - | 4km | 0.6 > 0.6 | 2.5 (1.0–6.0) 3.0 (1.3–6.9) | 0.6 > 0.6 | 2.6 (1.1–6.3) 3.1 (1.4–7.3) | 0.06 > 0.06 | 2.2 (0.9–5.6) 2.7 (1.1–6.6) |
| | | | p-trend=0.01 | | p-trend=0.01 | | p-trend=0.03 |

^aFor each insecticide, three separate logistic regression models were used to model the odds of detection (dependent variable) with each density metrics (CPUR, CROP-A, and W-CPUR) comparing >0 or density categories to homes with no use. A forward approach was used to evaluate covariates. Any covariate with a p-value 0.1 was retained in the final multivariable model.

 $[^]b$ CPUR= CPUR Metric (kg/km 2): insecticide use density proportional to the area of the buffer

 $^{^{}c}$ CROP-A= Crop area adjusted CPUR Metric (kg/km 2): insecticide use density proportional to the area of cultivated crops and pasture/hay with insecticide use in the buffer

 $d_{W-CPUR=Wind-adjusted CPUR Metric (kg/km^2)}$: insecticide use density proportional to the area of the buffer, weighted to account for the proportion of days the residence was downwind of the section in which the insecticide was applied during the period between application and dust collection

 e Detected in 26% of samples. Adjusted for year of dust collection, use of flea or tick products, and treatments for lawn/garden weeds that were positively associated with detection.

f Detected in 28% of samples. Adjusted for year and season of dust collection, and farm working occupation that were positively associated with detection.

^gDetected in 7% of samples. None of the covariates were predictors of detections.