UC Irvine

UC Irvine Previously Published Works

Title

Are changes in HLA Ags responsible for leukemia relapse after HLA-matched allogeneic hematopoietic SCT?

Permalink https://escholarship.org/uc/item/3wg150kj

Journal Bone Marrow Transplantation, 50(3)

Authors

Hamdi, A Cao, K Poon, L <u>et al.</u>

Publication Date 2015-03-01

DOI 10.1038/bmt.2014.285

Peer reviewed



HHS Public Access

Bone Marrow Transplant. Author manuscript; available in PMC 2016 March 24.

Published in final edited form as:

Author manuscript

Bone Marrow Transplant. 2015 March ; 50(3): 411-413. doi:10.1038/bmt.2014.285.

Are changes in HLA Ags responsible for leukemia relapse after HLA-matched allogeneic hematopoietic SCT?

A Hamdi¹, K Cao², LM Poon¹, F Aung², S Kornblau³, MA Fernandez Vina⁴, RE Champlin¹, and SO Ciurea¹

¹Department of Stem Cell Transplantation and Cellular Therapy, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

²Department of Laboratory Medicine, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

³Department of Leukemia, The University of Texas MD Anderson Cancer Center, Houston, TX, USA

⁴Department of Pathology, Stanford University, Stanford, CA, USA

Abstract

Loss of heterozygosity (LOH) has been shown to be associated with leukemia relapse after haploidentical transplantation. Whether such changes are an important cause of relapse after HLA-matched transplantation remains unclear. We retrospectively HLA-typed leukemic blasts for 71 patients with AML/myelodysplastic syndrome obtained from stored samples, and the results were compared with those obtained at diagnosis and/or before the transplant. No LOH or any other changes in HLA Ag were found in any of the samples tested post transplant as compared with pretransplant specimens. One patient had LOH in HLA class I Ag (HLA-A,-B and -C); however, these changes were present in the pretransplant sample indicating that they occurred before the transplant. We concluded that, in contrast with haploidentical transplantation, HLA loss does not have a major role as a mechanism of relapse after allogeneic transplantation with a closely HLA-matched donor.

INTRODUCTION

Disease relapse after allogeneic hematopoietic SCT (allo-HSCT) is the most important cause of treatment failure. The mechanisms of leukemia relapse after allo-SCT remain elusive.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

AUTHOR CONTRIBUTIONS

Correspondence: Dr SO Ciurea, Department of Stem Cell Transplantation and Cellular Therapy, The University of Texas MD Anderson Cancer Center, 1515 Holcombe Boulevard, Unit 423, Houston, TX 77030, USA. sociurea@mdanderson.org. This study was presented at the 2013 BMT Tandem Meetings, Salt Lake City, UT, USA.

AH collected data and wrote the paper; MAFV and KC contributed to study design, data collection and interpreted results; LMP and FA contributed to data collection and interpretation of results; SK performed the blast concentration; REC contributed with interpretation of the results and manuscript writing; SOC contributed with study design, data collection, interpretation of the results and manuscript writing. All authors critically reviewed and approved the manuscript.

Loss of HLA class I Ags has been associated with disease relapse, tumor progression and metastasis in solid tumors.^{1–5} In addition, loss of heterozygosity (LOH) has been shown to be a common cause of disease relapse after T-cell-depleted haploidentical transplantation.^{6,7} It is believed that these changes are a direct consequence of selective immune pressure mediated by T cells.^{8–10} Here we aimed to determine if changes in HLA Ags occur after closely HLA-matched transplants and can be responsible for leukemia relapse post transplant.

MATERIALS AND METHODS

Patients and transplantation procedure

We included all patients with AML or myelodysplastic syndrome who received an HLAmatched related or a closely HLA-matched unrelated donor transplant at our institution between March 1997 and August 2009, who relapsed after HSCT and had samples stored in the institutional sample bank. HLA typing was performed for all patients' samples at the time of the initial diagnosis, pretransplant and post relapse, either prospectively or on archived cryopreserved samples and for all donor samples. We compared the HLA typing for the post-relapse specimens with the specimens HLA typed at diagnosis and/or pretransplant and aimed to evaluate whether LOH occurred in the leukemic blasts at the time of relapse. This study was approved by Institutional Review Board, and a waiver of informed consent was obtained to analyze the archived samples. Written informed consent was obtained for treatment at our institution from all patients in accordance with the Declaration of Helsinki.

Sample collection and cryopreservation

Blood and BM aspirates underwent ficoll separation to remove RBC and neutrophils, leaving a mononuclear cell population along with B and T cells. We then performed a magnetic depletion of the B and T cells using beads for CD3+ and CD19+ cells using a MACS device (Miltenyi, Auburn, CA, USA), to yield a leukemia-enriched blast population. Cells were cryopreserved in Roswell Park Memorial Institute media supplemented with 20% FCS +10% DMSO and frozen using a programmed cell freezer. Contaminating B and T cells were <2% after the depletion. A validated methodology which detected leukemic cells >90% of the remaining material.

DNA isolation and HLA typing

Genomic DNA from malignant cells was extracted utilizing the QIAsymphony DNA Investigator Kit (QIAGEN, Valencia, CA, USA). HLA typing was performed for HLA-A, -B, -C, -DRB1 and -DQB1 loci by PCR amplification and oligonucleotide hybridization using molecular methods of the LABType SSO kit from One Lambda (Canoga Park, CA, USA), this is a reverse sequence-specific oligonucleotide hybridization in which the DNA is amplified by PCR and then hybridized with the bead probe array. The array platform uses microspheres as a solid support to immobilize oligonucleotide probes. Hybridization to each fluorescent bead is detected in a Luminex 100/200 System (Luminex Corporation, Austin, TX, USA).

HLA typing was performed for all samples at intermediate and high resolution for HLA-A, -B, -C, -DRB1 and -DQB1 loci by PCR amplification and oligonucleotide hybridization using molecular methods and commercial kits from Invitrogen (Carlsbad, CA, USA), ELPHA (Dreieich, Germany) and/or One Lambda (Canoga Park, CA, USA) that achieved intermediate resolution. The patients and donors were also typed prospectively for these loci by high-resolution methods (PCR amplification and nucleotide sequencing) using SEQR Sequence Based Typing Kits (Abbott Park, IL, USA).

RESULTS AND DISCUSSION

We identified 71 patients with a median age of 51 years (range 18–67) who met the inclusion criteria. Forty-eight percent of the patients were males. The majority of patients had AML/myelodysplastic syndrome relapse after a 10/10 allele matched HSCT (N = 57, 80%). Of 14 patients relapsed after mismatched HSCT, 12 had one allele mismatched at HLA class I or II loci (9/10, 17%) and two had two alleles mismatched (8/10, 3%). The patients' characteristics are summarized in Table 1. Conditioning consisted of a myeloablative regimen in 46 patients (65%) and reduce-intensity regimen in 25 patients (35%). Twenty-six patients (36%) were in remission at the time of transplant, whereas the rest had active disease. Forty-seven (66%) had peripheral blood and 24 patients (34%) had BM as the stem cell source. Out of the 71 patients tested, LOH at HLA class I loci (HLA-A, -B and -C) in relapsed blasts was present in only one case after transplant (Figure 1). In this patient who was a 58-year-old female with myelodysplastic syndrome who underwent 9/10 matched unrelated donor transplant there was loss of HLA-A02:05, B14:02:01 and C08:02 and her AML blasts became homozygous at these loci (Figure 1). However, these changes were also observed in the blasts collected in a pretransplant specimen, suggesting that the LOH happened before transplantation. No other HLA loss was observed in any other sample of studied patients.

The rate of HLA loss is low in hematologic malignancies compared with solid tumors.^{11–13} Previous studies have suggested that loss or downregulation of HLA class I may have an important role in relapsed leukemia and may be involved in relapse after HSCT or failure of DLI. The report by Masuda et al.¹⁴ showed that the loss or downregulation of HLA class I in acute leukemia blasts occurs more frequently in relapse compared with the initial diagnosis (2/5, 40% vs 3/39, 7.7%). However, they did not examine and compare the expression of HLA class I alleles at the initial diagnosis and relapse. Other investigators described pretransplant loss or downregulation of HLA class I in hematologic malignancies patients.^{15–20} Dubois et al.²¹ reported six patients with LOH in HLA region due to uniparental disomy before HSCT. In a study of 600 HSCT patients with myeloid malignancies, Pereira et al.¹⁹ showed pretransplant loss of HLA class I in three cases. Also, loss of HLA class I through acquired uniparental disomy has been described in relapsed leukemic blasts after haploidentical HSCT.^{6,7} Smith et al.²⁰ reported somatic mutations in HLA gene of 15 patients who underwent HSCT for hematological malignancies. Thirteen of these patients were found to have the mutation in pretransplant samples, whereas in 2 patients, mutations were detected in post-transplant relapse specimens. One was an ALL patient who received 1 Ag-mismatch unrelated donor transplant and the other one was a CML patient who received a haploidentical transplant. There is only one case report in the

literature of HLA loss in a relapsed AML after matched unrelated donor HSCT.²² It is, however, unclear if this was present immediately before transplant or occurred after the transplant, as we were fortunate to determine in our case.

Several mechanisms have been proposed for such HLA class I alterations including: absence of β 2-microglobulin⁴ or peptide transporter expression^{23–25} LOH, large deletions or mitotic recombination in chromosome 6^{25–28} transcriptional downregulation²⁹ or point mutation, partial deletion or somatic recombination.^{25,30,31}

In a series of 77 patients with relapse after mismatched transplantation for high-risk myeloid malignancies, no HLA loss among 11 mismatched unrelated donor transplants was identified.³² Also, in a series of nine pediatric patients with acute leukemia, Villalobos *at al.*⁷ found no HLA loss in post-HSCT leukemic blasts of six patients with matched HSCT. Our findings are consistent with these results and, although our methodology was not able to detect downregulation in HLA Ags, we showed that loss of HLA Ags may not have a major role in disease recurrence after a closely HLA-matched transplant. Further studies evaluating the alteration in HLA Ags post transplant are needed to determine if these changes are the result of immunologic pressure related to transplant or occurred just before transplantation and were identified only at the time of relapse.

In conclusion, in contrast to frequent finding of LOH in leukemic blasts after haploidentical transplantation and/or donor T-cell infusion, LOH was not identified in leukemic blasts at the time of relapse in T-cell-replete matched related or unrelated donor transplants. These results suggest that LOH could be a particular mechanism for leukemia relapse after haploidentical transplantation, whereas it does not appear to be a mechanism for leukemia relapse after allogeneic transplantation with a closely HLA-matched donor.

Acknowledgments

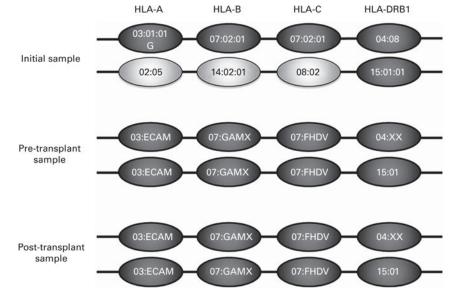
This paper was supported in part by an MD Anderson Cancer Center Institutional Research Grant to SOC.

References

- Garrido F, Ruiz-Cabello F, Cabrera T, Perez-Villar JJ, Lopez-Botet M, Duggan-Keen M, et al. Implications for immunosurveillance of altered HLA class I phenotypes in human tumours. Immunol Today. 1997; 18:89–95. [PubMed: 9057360]
- Algarra I, Collado A, Garrido F. Altered MHC class I antigens in tumors. Int J Clin Lab Res. 1997; 27:95–102. [PubMed: 9266279]
- Cabrera T, Angustias Fernandez M, Sierra A, Garrido A, Herruzo A, Escobedo A, et al. High frequency of altered HLA class I phenotypes in invasive breast carcinomas. Hum Immunol. 1996; 50:127–134. [PubMed: 8891736]
- Hicklin DJ, Wang Z, Arienti F, Rivoltini L, Parmiani G, Ferrone S. beta2-Microglobulin mutations, HLA class I antigen loss, and tumor progression in melanoma. J Clin Invest. 1998; 101:2720–2729. [PubMed: 9637706]
- Marincola FM, Jaffee EM, Hicklin DJ, Ferrone S. Escape of human solid tumors from T-cell recognition: molecular mechanisms and functional significance. Adv Immunol. 2000; 74:181–273. [PubMed: 10605607]
- Vago L, Perna SK, Zanussi M, Mazzi B, Barlassina C, Stanghellini MT, et al. Loss of mismatched HLA in leukemia after stem-cell transplantation. N Engl J Med. 2009; 361:478–488. [PubMed: 19641204]

- Villalobos IB, Takahashi Y, Akatsuka Y, Muramatsu H, Nishio N, Hama A, et al. Relapse of leukemia with loss of mismatched HLA resulting from uniparental disomy after haploidentical hematopoietic stem cell transplantation. Blood. 2010; 115:3158–3161. [PubMed: 20124217]
- Zheng P, Sarma S, Guo Y, Liu Y. Two mechanisms for tumor evasion of preexisting cytotoxic Tcell responses: lessons from recurrent tumors. Cancer Res. 1999; 59:3461–3467. [PubMed: 10416611]
- Garcia-Lora A, Algarra I, Gaforio JJ, Ruiz-Cabello F, Garrido F. Immunoselection by T lymphocytes generates repeated MHC class I-deficient metastatic tumor variants. Int J Cancer. 2001; 91:109–119. [PubMed: 11149409]
- Garcia-Lora A, Martinez M, Algarra I, Gaforio JJ, Garrido F. MHC class I-deficient metastatic tumor variants immunoselected by T lymphocytes originate from the coordinated downregulation of APM components. Int J Cancer. 2003; 106:521–527. [PubMed: 12845647]
- Gupta M, Raghavan M, Gale RE, Chelala C, Allen C, Molloy G, et al. Novel regions of acquired uniparental disomy discovered in acute myeloid leukemia. Genes Chromosomes Cancer. 2008; 47:729–739. [PubMed: 18506749]
- Bullinger L, Kronke J, Schon C, Radtke I, Urlbauer K, Botzenhardt U, et al. Identification of acquired copy number alterations and uniparental disomies in cytogenetically normal acute myeloid leukemia using high-resolution single-nucleotide polymorphism analysis. Leukemia. 2010; 24:438–449. [PubMed: 20016533]
- Tuna M, Knuutila S, Mills GB. Uniparental disomy in cancer. Trends Mol Med. 2009; 15:120– 128. [PubMed: 19246245]
- Masuda K, Hiraki A, Fujii N, Watanabe T, Tanaka M, Matsue K, et al. Loss or downregulation of HLA class I expression at the allelic level in freshly isolated leukemic blasts. Cancer Sci. 2007; 98:102–108. [PubMed: 17083564]
- Sayer DC, Smith LK, Krueger R, Chrisitansen FT. DNA sequencing-based HLA typing detects a B-cell ALL blast-specific mutation in HLA-A(*)2402 resulting in loss of HLA allele expression. Leukemia. 2004; 18:174–176. [PubMed: 14523467]
- Gaidulis L, Sun J-Y, Senitzer D. The case of a homozygous heterozygote. Hum Immunol. 2005; 66:24.
- Gaidulis L, Sun J, Senitzer D. Loss of HLA and STR alleles observed in a patient during blast crisis. Hum Immunol. 2007; 68:S130–S130.
- Fernandez-Vina M, Garcia-Manero G, Cortes J, Abruzzo L, Cano P. Typing the malignant clone vs the patient's tissue. Hum Immunol. 2007; 68:S131–S131.
- Pereira S, Vayntrub T, Hiraki DD, Cherry AM, Arai S, Dvorak CC, et al. Short tandem repeat and human leukocyte antigen mutations or losses confound engraftment and typing analysis in hematopoietic stem cell transplants. Hum Immunol. 2011; 72:503–509. [PubMed: 21463659]
- Smith AG, Fan W, Regen L, Warnock S, Sprague M, Williams R, et al. Somatic mutations in the HLA genes of patients with hematological malignancy. Tissue Antigens. 2012; 79:359–366. [PubMed: 22489945]
- Dubois V, Sloan-Bena F, Cesbron A, Hepkema BG, Gagne K, Gimelli S, et al. Pretransplant HLA mistyping in diagnostic samples of acute myeloid leukemia patients due to acquired uniparental disomy. Leukemia. 2012; 26:2079–2085. [PubMed: 22488219]
- Toffalori C, Cavattoni I, Deola S, Mastaglio S, Giglio F, Mazzi B, et al. Genomic loss of patientspecific HLA in acute myeloid leukemia relapse after well-matched unrelated donor HSCT. Blood. 2012; 119:4813–4815. [PubMed: 22596173]
- D'Urso CM, Wang ZG, Cao Y, Tatake R, Zeff RA, Ferrone S. Lack of HLA class I antigen expression by cultured melanoma cells FO-1 due to a defect in B2m gene expression. J Clin Invest. 1991; 87:284–292. [PubMed: 1898655]
- 24. Spies T, DeMars R. Restored expression of major histocompatibility class I molecules by gene transfer of a putative peptide transporter. Nature. 1991; 351:323–324. [PubMed: 2034277]
- Browning M, Petronzelli F, Bicknell D, Krausa P, Rowan A, Tonks S, et al. Mechanisms of loss of HLA class I expression on colorectal tumor cells. Tissue Antigens. 1996; 47:364–371. [PubMed: 8795136]

- Koopman LA, Mulder A, Corver WE, Anholts JD, Giphart MJ, Claas FH, et al. HLA class I phenotype and genotype alterations in cervical carcinomas and derivative cell lines. Tissue Antigens. 1998; 51:623–636. [PubMed: 9694355]
- 27. Torres MJ, Ruiz-Cabello F, Skoudy A, Berrozpe G, Jimenez P, Serrano A, et al. Loss of an HLA haplotype in pancreas cancer tissue and its corresponding tumor derived cell line. Tissue Antigens. 1996; 47:372–381. [PubMed: 8795137]
- Ramal LM, Maleno I, Cabrera T, Collado A, Ferron A, Lopez-Nevot MA, et al. Molecular strategies to define HLA haplotype loss in microdissected tumor cells. Hum Immunol. 2000; 61:1001–1012. [PubMed: 11082513]
- 29. Soong TW, Hui KM. Locus-specific transcriptional control of HLA genes. J Immunol. 1992; 149:2008–2020. [PubMed: 1517566]
- Browning MJ, Krausa P, Rowan A, Hill AB, Bicknell DC, Bodmer JG, et al. Loss of human leukocyte antigen expression on colorectal tumor cell lines: implications for anti-tumor immunity and immunotherapy. J Immunother Emphasis Tumor Immunol. 1993; 14:163–168. [PubMed: 8297898]
- Koopman LA, van Der Slik AR, Giphart MJ, Fleuren GJ. Human leukocyte antigen class I gene mutations in cervical cancer. J Natl Cancer Inst. 1999; 91:1669–1677. [PubMed: 10511595]
- 32. Crucitti L, Crocchiolo R, Toffalori C, Stanghellini MTL, Assanelli A, Carrabba M, et al. Incidence, risk factors and clinical outcome of leukemia relapses due to loss of the mismatched hla haplotype after partially-incompatible hematopoietic stem cell transplantation. Blood. 2013; 122:918–918.



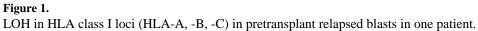


Table 1

Patients and transplants characteristics

Characteristic	Number (%)
Age, median range	51, 18–67
Sex	
Male	34 (48%)
Female	37 (52%)
Status at SCT	
CR1	11 (15%)
Primary induction failure	17 (24%)
CR2	15 (21%)
Active disease	28 (39%)
Cytogenetic risk group	
Good	7 (10%)
Intermediate	35 (49%)
Poor	27 (38%)
Unknown	2 (3%)
Donor type	
Matched	57 (80%)
1 Ag mismatched	12 (17%)
HLA-A	7
HLA-C	1
HLA-DRB1	1
HLA-DQB1	3
2 Ag mismatched	2 (3%)
HLA-A/HLA-C	1
HLA-A/HLA-DRB1	1
Donor relation	
Related	36 (51%)
Unrelated	35 (49%)
Stem cell source	
PB	47 (66%)
BM	24 (34%)
Conditioning regimen	
Myeloblative	46 (65%)
Reduced intensity	25 (35%)

Abbreviation: PB = peripheral blood.