UC Berkeley UC Berkeley Previously Published Works

Title

Ecological network structure in response to community assembly processes over evolutionary time

Permalink https://escholarship.org/uc/item/40k0h6hq

Author Graham, Natalie Rose

Publication Date 2023

DOI 10.1111/mec.16873

Data Availability

The data associated with this publication are available at: <u>https://doi.org/10.6078/D1DX4T</u>

Peer reviewed

1	Ecological network s	tructure in response t	o community assembl	bly processes over evolutionar
---	----------------------	------------------------	---------------------	--------------------------------

2 time

- 3 Ecological networks and community assembly
- 4 Natalie R. Graham^{1*}, Henrik Krehenwinkel², Jun Ying Lim³, Phillip Staniczenko⁴, Jackson Callaghan⁵,
- 5 Jeremy C. Andersen⁶, Daniel S. Gruner⁷, Rosemary G. Gillespie¹
- 6 ¹ Department of Environmental Sciences Policy and Management, University of California Berkeley,
- 7 Mulford Hall, Berkeley, California, USA
- 8 ² Department of Biogeography, Trier University, Faculty of Regional and Environmental Sciences, Trier
- 9 54286, Germany
- ³ Department of Biological Sciences, National University of Singapore, Singapore
- ⁴ Department of Biology, Brooklyn College, City University of New York, USA
- ⁵ Department of Integrative, Structural and Computational Biology, The Scripps Research Institute, San
- 13 Diego, California, USA
- ⁶ Department of Environmental Conservation, University of Massachusetts Amherst, Amherst,
- 15 Massachusetts, USA
- ⁷ Department of Entomology, University of Maryland, College Park, Maryland, USA
- 17
- 18 *Corresponding author
- 19 Email: n.graham@berkeley.edu
- 20
- 21 Abstract
- 22

23 The dynamical structure of ecological communities results from interactions among taxa that change with

- 24 shifts in species composition in space and time. However, our ability to study the interplay of ecological
- and evolutionary processes on community assembly remains relatively unexplored due to the difficulty of
- 26 measuring community structure over long temporal scales. Here, we made use of a geological
- 27 chronosequence across the Hawaiian Islands, representing 50 years to 4.15 million years of ecosystem

28 development, to sample 11 communities of arthropods and their associated plant taxa using semi-29 quantitative DNA metabarcoding. We then examined how ecological communities changed with 30 community age by calculating quantitative network statistics for bipartite networks of arthropod-plant 31 associations. The average number of interactions per species (linkage density), ratio of plant to arthropod 32 species (vulnerability), and uniformity of energy flow (interaction evenness) increased significantly in 33 concert with community age. The index of specialization H_2 ' has a curvilinear relationship with community 34 age. Our analyses suggest that younger communities are characterized by fewer but stronger 35 interactions, while biotic associations become more even and diverse as communities mature. These 36 shifts in structure became especially prominent on East Maui (~0.5 my) and older volcanos, after enough 37 time had elapsed for adaptation and specialization to act on populations in situ. Such natural progression 38 of specialization during community assembly is likely impeded by the rapid infiltration of non-native 39 species, with special risk to younger or more recently disturbed communities that are composed of fewer 40 specialized relationships.

41

42 **Keywords** community assembly, ecological networks, DNA metabarcoding, oceanic islands

43

44 Introduction

45

46 Biodiversity is organized into complex ecological networks of interacting species that change through time 47 in response to ecological and evolutionary processes. Understanding these changes is important for 48 predicting the impacts of global change on higher multispecies organization (Dell et al., 2019; Smith-49 Ramesh et al., 2017; Staniczenko et al., 2017). A suite of analytical tools (Delmas et al., 2019) exist to 50 quantify changing community structure in response to a variety of perturbations (Aizen et al., 2008; Fricke 51 et al., 2017; Vacher et al., 2010). A major challenge remaining is to understand the configuration of 52 ecological networks in a predictive context over long spatiotemporal scales (Poisot et al., 2015; 53 Trøjelsgaard & Olesen, 2016; Yeakel et al., 2014). Consequently the effect of community assembly 54 processes on the structure of interaction networks describing ecological communities remains poorly 55 understood (Ponisio et al., 2019; Rominger et al., 2016).

56 Early research on community assembly often ignored ecological interactions due to their complexity. 57 Notably, neutral models for community assembly are even agnostic to organismal identity (Hubbell, 2001; 58 Rosindell et al., 2011). As species identity and interactions began to be incorporated into models, the 59 initial 'assembly rules' of Diamond (1975) highlighted 'forbidden species combinations' and nonrandom 60 patterns of co-occurrence. A growing recent theme focuses on the effect of abiotic and biotic filters on a 61 regional species pool (Münkemüller et al., 2020) with varying temporal and spatial filters dictating network 62 structure (Peralta et al., 2019). However, much of this work ignores the role of evolution in shaping 63 interactions through time. The extent of adaptation, and possible speciation, in shaping interactions as 64 communities assemble, depends on the isolation of the community from the source pool (Gillespie et al., 65 2020; Rosindell & Phillimore, 2011). At the extreme, evolution will have shaped interactions among every 66 member of a community and the effects of filtering from a regional species pool might thus appear 67 relatively weak (Ponisio et al., 2019). While most communities will include the role of both ecological 68 filtering and evolutionary adaptation, our ability to thread complex ecological questions of network 69 structure into an evolutionary framework has presented a major obstacle.

71 Recognizing this impediment, recent work has examined avenues to approach the problem. In particular, 72 models of trait evolution on phylogenies provide a means to understand how eco-evolutionary feedbacks 73 shape interactions as communities assemble (Segar et al., 2020). Likewise, based on theory showing 74 how change across short time scales affects longer-term evolutionary dynamics, clade-level phylogenetic 75 comparative approaches can be applied to community data to understand the dynamics of network 76 structure (Weber et al., 2017). Both these approaches focus on the lineages that make up communities, 77 asking how interacting sets of lineages affect each other. However, another approach is to focus explicitly 78 on the community rather than individual lineages, connecting large-scale understanding of community 79 interactions at a given time period in a spatially variable environment with the understanding of how the 80 integrated structure of biodiversity changes through time. Such an approach attempts to address a major 81 gap in the field by bridging macroecology and macroevolution (McGill et al., 2019) and hence showing 82 how network structure changes across scales of space and time within a whole-community context 83 (Weber et al., 2017).

84

85 While theory indicates a clear role for biotic interactions leading to individual and community 86 specialization over long-term community development, empirical evaluation has been challenging. One 87 difficulty is in obtaining measures of community composition and interactions at relevant spatial scales, 88 and another obstacle is the vast timeframe over which evolutionary phenomena occur. With their short 89 generation times that are amenable to laboratory studies, microbial systems provide exceptional cases 90 that document community assembly over evolutionary time scales (Boon et al., 2014; Koskella et al., 91 2017; Koskella & Brockhurst, 2014; Venturelli et al., 2018). In particular, studies of the plant phyllosphere 92 showed a more prominent role of nonneutral selection over time and an increase in the strength of biotic 93 interactions and community cohesion (Morella et al., 2020). However, to infer the role of interactions in 94 community assembly of longer-lived macroorganisms requires very particular systems. Here, we make 95 use of two sets of circumstances that, together, provide an extraordinary opportunity to assess the nexus 96 of ecological and evolutionary community assembly in the context of interaction networks.

98 First, we use the system provided by the Hawaiian Islands. Islands in general provide discrete 99 communities that can be used for natural experiments in interaction dynamics (Brodie, 2017; Castro-Urgal 100 & Traveset, 2014; Olesen et al., 2002). In particular, oceanic archipelagos formed in situ over millions of 101 years offer the opportunity to study species interactions over evolutionary timescales (Hembry et al., 102 2018; Ponisio et al., 2019; Rominger et al., 2016; Trøjelsgaard et al., 2013). Moreover, the geological 103 series of islands in the Hawaiian archipelago represents a chronosequence (Vitousek, 2002; Walker et 104 al., 2010); each substrate age represents communities of different ages, ranging from \sim 50 years to \sim 5 105 Myr (Shaw & Gillespie, 2016). Notably, the native montane forest of Hawaii is dominated by just two 106 canopy tree species (Metrosideros polymorpha and Acacia koa), making it relatively simple ecologically 107 and hence more amenable to capturing and characterizing whole communities.

108

109 Second, we make use of the emerging field of DNA metabarcoding (Krehenwinkel, Wolf, et al., 2017; Yu 110 et al., 2012), which makes a comprehensive analysis of taxonomic composition possible, offering the 111 opportunity to simultaneously assess thousands of species rapidly, offering enormous potential for 112 reconstructing complex ecological networks (Clare, 2014; Hrček & Godfray, 2015; Vacher et al., 2016). 113 Relative sequence abundances offer a proxy for interaction strength (Lim et al., 2021), providing greater 114 reliability for co-occurrence studies to measure biotic associations (Bálint et al., 2018; Mata et al., 2021). 115 Combining high-throughput sequencing with theoretical approaches, such as statistical modeling (Faust & 116 Raes, 2012; Newman & Girvan, 2004) and machine learning (Bohan et al., 2011), shows considerable 117 promise in helping to close the gap on the historical impediments for comprehensive quantification of 118 interactions in ecological communities.

119

Here we use semi-quantitative DNA metabarcoding to build networks of arthropod-plant associations at 11 sites across the Hawaiian chronosequence, using the substrate age at a site as measure of community age, and then use those networks to test a range of expectations on how ecological and evolutionary processes shape community structure over long timescales (e.g., Table 1). We expect network size - both the number of nodes and number of links - will increase with community age, but disproportionately, as younger communities gain taxa through colonization only and older communities 126 assemble through colonization and evolutionary processes. This allows tests of the following hypotheses 127 for evolutionary assembly of networks (Figure 1): (H1) Starting from bare lava, early successional 128 communities offer low resource diversity yet are necessarily composed of assemblages from nearby 129 species pools. Therefore, younger communities will have a high proportion of generalists - a subset of 130 the nearby species pool most likely to persist without particular interaction partners - but greater 131 interaction frequency on fewer interaction pathways because of resource heterogeneity. (H2) The set of 132 biotic interactions that a given taxon will experience will decrease with community age, and the evenness 133 of the interactions among resources will increase, resulting in greater network specialization (Ponisio et 134 al., 2019; Rominger et al., 2016). One ideal component of this study is the large temporal span of time for 135 community assembly. We can assume that younger communities will gain taxa only through colonization 136 given that they are not established long enough for in situ speciation to take place. Of course, older 137 communities will assemble through colonization and evolutionary processes. We cannot tease apart the 138 effects of both processes at the oldest sites, but we can compare the youngest to oldest sites and their 139 related ecological networks for signatures of assembly after evolutionary processes have taken effect. 140 With an increasing number of taxa that have evolved together in a community it follows there will also be 141 an increase in the specialization of the interactions among these species that may be detectible at the 142 network architecture level.

143 Materials and Methods

144 Site selection methods

145 The Hawaiian Islands are formed as the Pacific plate moves northwestward across a stationary volcanic hotspot, therefore the archipelago represents a chronosequence of geological age from the youngest 146 147 island (Hawaii, ~0–0.5 Myr), to the oldest high island of Kauai (~5 Myr), (Clague, 1996). Discrete 148 volcanoes within islands present additional contrasts in geological age, and the underlying substrate age 149 has been mapped in fine detail (Wolfe & Morris, 1996). Metrosideros polymorpha (Myrtaceae) is the 150 dominant canopy tree in these forests across islands, with patches of sub-dominant Acacia koa (Fabaceae) and numerous associated understory trees, shrubs, herbs, and ferns (Gagne & Cuddihy, 151 152 1990).

153

We selected 14 sites of varying geologic age, ranging from 50 to 4.15 x 10⁶ years old, across four islands
of the archipelago: Hawaii, Maui, Molokai, Kauai (SI Figure 1, SI Table 1). To control for climatic
differences and disturbance across sites, sites were constrained to ranges of elevation (1000-1300 m)
and precipitation (average annual precipitation 2500-3000 mm) and within accessible protected forest
lands (*Gap Analysis Project* | *U.S. Geological Survey*, 2019; Giambelluca et al., 2013).

159

160 For each potential site, spatial polygons were created based on the intersections of these layers and 161 initial field reconnaissance to confirm remotely sensed data and feasibility of access. Within these 162 potential site polygons, airborne high-resolution laser scanning from the Global Airborne Observatory 163 (GAO; formerly named Carnegie Airborne Observatory; Asner et al. (2012)) was used to generate forest 164 canopy height profiles using a physical model described in Asner et al. (2008). The ground digital 165 elevation model was also generated using the method of Asner et al. (2007). The data were collected at 166 4 laser shots per square meter, processed to height profiles at 5 m resolution and then averaged at a grid 167 cell spacing of 30 m (SI Figure 2 and SI Table 2).

168

169 20 randomized candidate plots were generated for each site, with the intention of ultimately selecting six, 170 15 m-radius plots. These 20 randomized candidate plots were constrained to be a minimum distance of 171 200 m from all other plots and to be within the top 40% quantile of LiDAR-estimated canopy heights. 172 Candidate plot generation was achieved with custom scripts in the R programming language (R Core 173 Development Team, 2019) using a simple rejection sampling algorithm: random sets of spatial locations 174 are generated within pixels of sufficient canopy height until a set of locations is found which meet the 175 requirement of being 200 m distant. The minimum distance of 200 m was a constraint to maximize 176 independence among sampling areas while capturing more spatial heterogeneity within sites.

177

At each site, each of the 20 candidate plots were ground-truthed to confirm the plot was dominated by
native vegetation and minimally impacted by human use and/or invasive vertebrates. This ground-truthing
process eliminated a variable number of the initial 20 candidate plots. If fewer than six final plots

remained after ground-truthing, another set of candidate plots were generated and ground-truthed to find
a final set of six plots. If more than six plots remained after ground-truthing, the final six plots were
selected by randomly selecting from the ground-truthed plots.

184

185 Collection protocol

186 We collected arthropods using vegetation beating at six 15 m radius plots per site during May 2015 187 through January 2016, with plots sampled randomly to avoid seasonality effects on arthropod 188 composition. To ensure equal sampling effort across sites, sampling was limited to a total of 420 seconds 189 in each plot. If after all arthropod collection processing steps (described below) the total vegetation 190 beating time for a plot was not within one standard deviation of 420 seconds sampling effort then that plot 191 was dropped from further analyses, resulting in a total of 50 plots from 11 sites (SI Table 1). As we were 192 interested in characterizing plant-arthropod associations, we sampled plant genera in each plot 193 proportional to their relative abundance. Percentage cover of each understory plant genus was estimated 194 visually. Where plants could not be identified to the genus-level, we grouped them into morphotaxa and 195 sampled them accordingly. Vegetation beating was performed by placing 1 m x 1 m white beating sheets 196 under individual plants and gently agitating the foliage using a one meter length pvc pole for timed second 197 intervals. Arthropods dislodged by the agitation which drop onto the beating sheet are aspirated into a vial 198 containing 95% ethanol. Each plant associated arthropod community sample was transferred to one or 199 more 2 ml vials containing fresh 95% ethanol, labeled, and transported to the lab where it was stored at -200 20 °C.

201

202 Specimen sorting and DNA extraction

203

To reduce bias due to differently sized individuals contributing disproportionate amounts of DNA (Elbrecht & Leese, 2015) specimens were sorted following procedures described in Lim et al. (2021). Each plant beating sample was sorted in petri dishes on 1 mm graph paper under a stereoscope into four size categories (0-2 mm, 2-4 mm, 4-7 mm, 7 mm and up) based on the body size distribution found in a common Hawaiian ecosystem. Individuals in each size category were counted and placed with fresh ethanol into a single well in a 96-well plate. Thus, all individuals from a particular plant genus at a
particular plot have their DNA extracted together and are prepared together using a dual-indexing
strategy described below into NGS amplicon libraries for sequencing. The Collembola had considerably
higher abundance than the remaining arthropods in the small size categories, therefore Collembola were
separated into 1.5 ml Eppendorf tubes and processed for DNA extraction and sequencing parallel to the
remaining arthropod community samples.

215

Specimens from public and private collections were also used to generate a DNA barcode reference
library for 57 species. We used whole bodies of species from private collections where available because
these were easiest to generate sequences from preserved material (86% barcode generation success).
Genomic DNA extraction of size sorted arthropod-plant community samples was performed in 600 µl
volumes using the Tissue protocol described in the Qiagen Puregene kit modified for automation (Lim et
al., 2021). DNA was eluted in 50 ul DNA Hydration Solution.

222

223 Sequence analysis

224 Each size sorted sample and a PCR negative for each 96 well plate (containing no template DNA) was amplified with a primer combination (ArF1/ Fol-degen-rev) (Gibson et al., 2014; Yu et al., 2012) that 225 226 targets a 418 bp fragment in the barcode region of the Cytochrome Oxidase I (COI) gene. This primer 227 pair has been suggested as the most appropriate for capturing arthropod diversity in DNA metabarcoding 228 studies (Elbrecht & Leese, 2015) and has been shown to reliably amplify the Hawaiian arthropod 229 community (de Kerdrel et al., 2020). PCRs were run in 10 µl volumes using the Qiagen Multiplex PCR kit 230 at an annealing temperature of 46 °C, with 1 µl of DNA and 0.5 µl of each 10 µM primer. A first round of 231 PCR consisted of 32 cycles using tailed primers; each primer additionally had a unique 6 bp inline 232 barcode so that multiple plates of the same primer can be pooled together. PCR products were cleaned 233 of residual primer using a 1x ratio of SPRI beads (Sera-Mag™) and pooled together based on band 234 intensity (i.e., DNA concentration) on an agarose gel relative to a DNA ladder (NEB) and using the Gel 235 Doc XR System with the Quantity One software (Bio-Rad). A second indexing PCR of 6 cycles was 236 performed with the pooled amplicons to introduce dual indexes and Illumina® TruSeq sequencing

adapters to 5'-tails of the locus-specific PCR primers (Lange et al., 2014), with a final 5'-3' layout as
Illumina adapter, 6 bp inline barcode, PCR primer as described in de Kerdrel et al. (2020). The indexed
products were cleaned again with SPRI beads, quantified by electrophoresis, and then pooled in equal
amounts into a single tube. The samples were then sequenced on an Illumina® MiSeq using V3 (600
cycles) chemistry according to the manufacturer's protocol (Illumina, San Diego, USA). We aimed for a
total of 30,000 reads per sample. Each PCR negative was sequenced with each plate of specimen
libraries regardless of the absence of detectable PCR product on a gel.

244

245 We generated 2276 metabarcode libraries with each library representing the total arthropods collected for 246 each plant genus for each plot (a sampling event), sorted into one of four size categories (a sequencing 247 pool). Sequences were demultiplexed on Illumina® BaseSpace by sample well based on the two 8-bp 248 indexes with no mismatches allowed. We merged paired reads using PEAR (Zhang et al., 2014) with a 249 minimum overlap of 50 bp and a minimum quality of Q20. Merged reads were quality filtered (≥90% of 250 bases ≥Q30) and transformed into fasta files using the FastX Toolkit (Gordon & Hannon, 2010). The 251 resulting fasta files were demultiplexed by PCR primer and 6 bp inline barcode combination, using the 252 forward and reverse primer sequences as indices with the grep command in UNIX, and the primer 253 sequences were then trimmed using the UNIX stream editor.

254

255 Rarefaction and pseudogene removal

We rarefied each sample using a custom unix command that drew from the total reads of the metabarcoding analysis a number of reads that was equivalent to the numerical abundance of individual arthropods counted into each well of the 96-well plate, repeating the draw of sequences 100 X with replacement. The process of rarifying by repeated random draw based on the expected individual specimen abundance should correct the disproportionate abundance of sequences that accumulate for larger specimens compared to smaller specimens, due to the amplification bias that is inherently caused by differential starting tissue amounts (Lim et al., 2021).

264 We generated zero-radius OTUs (zOTUs), from the rarefied raw reads with the unoise3 command (Edgar, 265 2016) following the recommended protocols in the USEARCH v11 pipeline (Edgar, 2010). Specifically, 266 the quality trimmed reads were dereplicated and clustered into zOTUs using the unoise3 command in 267 USEARCH. Chimera were removed *de novo* in USEARCH. The resulting zOTUs were compared against 268 the NCBI Genbank database and our custom-made DNA reference library for Hawaiian taxa using 269 BLASTn with a maximum of 10 target sequences. All non-arthropod zOTUs were removed after which 270 5.046 zOTUs remained. We aligned these 5046 zOTUs using default settings in Clustal Omega (Sievers 271 et al., 2011). To remove putative pseudogenes from the zOTU dataset we ran metaMATE with default 272 specifications and the example specifications file to detail how per-zOTU read frequencies should be 273 assessed (Andujar et al., 2021). Using the output of metaMATE we applied the least stringent Numt 274 removal strategy so that we could retain as many putatively true zOTUs as possible (Graham et al., 275 2021), this reduced the number of zOTUs from 5046 to 4330.

276

277 Taxonomic matching and abundance estimates

278 About a quarter of the zOTUs (n = 901) were matched to the Blast or voucher DNA reference library with 279 less than 85 percent similarity. To validate the taxonomic identification for each zOTU at higher taxonomic 280 levels (e.g. order, family) we compared the top 10 blast and reference library hits with phylogenetic 281 clustering from a ML tree. A ML tree with bootstop autoMRE bootstrap support was generated by running 282 RAxML-HPC v.8 on XSEDE on the Cipres science gateway (Miller et al., 2010) under the GTR model with 283 a gamma distribution plus invariant sites. For 28 zOTUs taxonomic order could not be determined via 284 sequence similarity to databases or phylogenetic clustering and were thus removed from downstream 285 analysis. Taxonomic assignment was considered trustworthy if the percent similarity of the metabarcoding 286 sequence to the NCBI GenBank or DNA reference voucher was: between 88-94% for family, between 287 94%-98% for genus and greater than 98% percent similarity for species, while matches below 88% 288 similarity were made only to order. These threshold values were arbitrarily chosen based on previous 289 investigations using mock communities or photo voucher integrative taxonomy of select taxa from the 290 same high-elevation wet forest communities of Hawaiian arthropods and amplified using the same COI 291 marker (Krehenwinkel et al. 2017, de Kerdrel et al. 2020).

292

293

294

295

To create a table with OTU abundances for community analyses we mapped a query set of raw reads to the filtered and taxonomically identified search database of zOTUs in USEARCH v11 (Edgar, 2010) using the otutab command with the default 97% percent similarity mapping threshold. After OTU mapping and

read removal based on the PCR negative control sequencing pool the number of unique sequences wasreduced by 133 zOTUs to 4197 OTUs.

298

299 In order to use relative sequence abundance of arthropod OTUs as an approximation of arthropod-plant 300 associations, we adopted a semi-quantitative processing pipeline (Lim et al., 2021) to ameliorate 301 differences in sampling effort, body size of specimens, and genomic procedures. To review, (1) for each 302 site (community) there was six plots, (2) within each plot each plant taxon was sampled by seconds of 303 time corresponding to its relative abundance in the plot, (3) we sorted bulk arthropod samples by size and 304 counted the individuals, (4) sequences were generated using a DNA region with demonstrated success 305 for Hawaiian arthropod taxa (de Kerdrel et al., 2020) and false reads (pseudogenes) were removed 306 (Graham et al., 2021), (5) we randomly sampled the sequencing reads based on the count of individuals 307 in each size class, and finally, (6) sequence reads were summed across size classes and plots (Figure 308 1b).

309

310 Calculation of quantitative ecological network metrics

311

312 Using bipartite networks of arthropod-plant associations at each community age, we tested our hypotheses by calculating quantitative (weighted) and qualitative (binary) network metrics (Table 2) 313 314 expected to occur in the transition from younger communities to older communities (Table 1). Data 315 processing and statistical analyses were performed in R version 4.0.2. To distinguish between taxa that 316 have colonized the archipelago historically or more recently, we characterized the probable native and 317 non-native composition for each aged community based solely on sequence characteristics as outlined in 318 Andersen et al. (2019). The approach considers both the evolutionary distances between species and the 319 genetic diversity within species. Sequence characteristics of OTUs show a higher amount of neutral (or

320 otherwise) sequence variation among endemic taxa, as they have evolved from a common ancestor on 321 the islands, when compared to non-native taxa that evolved elsewhere and have no close relatives. The 322 approach was implemented into a machine learning strategy using random forests in sklearn and 323 packaged with multiple utilities and a graphical user interface in NIClassify 324 (https://github.com/tokebe/niclassify). By annotating the nativeness status for sequences which are 325 identifiable to species level (98% or above match to databases), NIClassify can accurately assign status 326 for the remaining sequences. As part of the NIClassify classifications an output of accuracy is obtained by 327 withholding species with known statuses during the training, and then comparing the results for those 328 samples based on the classifications. These samples are randomly selected by the program, so biases in

329 regards to well- versus under-sampled taxa are not expected to influence the training.

330 We aggregated the sequence abundance for each arthropod OTU according to its association with a 331 particular plant genus within a site. For example, we found the sum of the sequence abundances for OTU 332 'X', a Hemiptera from genus Nesodyne, that was associated with (i.e., collected on) plants in the genus 333 Coprosma. We configured the arthropod-plant abundance data as a matrix with arthropods as columns 334 and plants as rows; there were 11 matrices, one for each site of different substrate age. As such, we 335 measure the strength of an interaction as the sequence abundance of the arthropod that was collected on 336 a particular plant species, as it is an aggregated assessment of the arthropod-plant association across 337 multiple plants and multiple plots within a site. We also graphed quantitative and qualitative metrics for 338 matrices of arthropod-plant interactions at each plot within a site. However, we constrain our discussion to 339 the aggregated network data because our confidence in the network statistics increases with the size of 340 the networks. This was a particular issue for some plots at the youngest and most depauperate sites, 341 where < 4 plant species were sampled within the plot radius and networks would be small.

We plotted the ecological network matrix for each community age using the 'plotweb' command in the R package bipartite. For each network, lower bars represent plant abundance based on sampling time and upper bars represent arthropod abundance based on OTU frequency. For visual simplicity, we grouped upper bars by arthropod order. As described above, link width represents relative read abundance of arthropod OTUs collected on each plant taxon (Alberdi et al., 2019); in other words, link width
 corresponds to the relative frequency of each association.

348 The information contained in ecological networks can be summarized in various ways. Qualitative 349 properties used to describe networks, which treat all interactions as equal irrespective of their magnitude 350 or frequency, tend to be highly sensitive to variation in sampling effort (Goldwasser & Roughgarden, 351 1997; Martinez et al., 1999). Quantitative metrics that weight each taxon by the total amount of its 352 incoming and outgoing biomass flows (Bersier et al., 2002) are more robust to sampling differences 353 (Banašek-Richter et al., 2004). Using the 'networklevel' commands in the R package bipartite (Dormann 354 et al., 2008) we calculated six quantitative indices for our bipartite networks of arthropods and associated 355 plants: (1) linkage density, (2) connectance, (3) generality, (4) vulnerability, (5) interaction evenness, and 356 (6) the index of specialization H_2 ' (Table 2), that we reasoned would be associated with network 357 specialization (Table 1). We converted each matrix to a binary presence-absence matrix and calculated 358 the qualitative equivalent of: (1) linkage density, (2) connectance, (3) generality, and (4) vulnerability. We 359 additionally calculated the ratio of resource species to consumers for the qualitative matrices, which is the 360 ratio of plant genera to arthropod OTUs. These metrics represent the most fundamental biological and 361 ecological properties of a community. We reasoned that the simplest metrics are a reasonable starting 362 point given the limited understanding of how evolution shapes network structure, which would be 363 necessary to justify the application of more involved network metrics. Further, these metrics have values 364 that are interpretable with respect to their effect on specialization over time.

365 Tests of network metric significance and correlation between network properties

We used null models (Vázquez & Aizen, 2006) to test the statistical significance of empirical network metric values for the weighted data. For each weighted empirical network, we generated 1000 synthetic networks so that the total number of interactions and the identity of interaction partners is maintained while the weight associated with each interaction is shuffled (Staniczenko et al., 2013). With this simple quantitative null model, the distribution of interaction weights is conserved, along with the pattern of binary interactions, but not the identities of which interaction partners are associated with which weights. In terms of biological reasoning, the null model assumes that the identities of any two species involved in a non-forbidden interaction are unimportant for explaining network metrics. We calculated pvalues and z-scores for each combination of empirical network and metric by comparing the observed
metric value calculated from the empirical network to the distribution of metric values calculated from
synthetic matrices generated by the null model, i.e., the p-value quantifies how unlikely the observed,
empirical metric value is to have been generated by the null model.

378 To compare the effect of community assembly on network size, arthropod diversity, and network metrics, 379 we regressed the dependent variables by mean substrate age for each collection site. The untransformed 380 substrate age data departed significantly from normality, so comparisons were performed using regressions on natural log-transformed substrate age data (Cowie, 1995; Gruner, 2007). We tested the 381 382 significance of the correlation between network size and community age, each network metric and 383 community age, and each network metric and network size, using Spearman's correlation tests. 384 Additionally, we fit a second-degree polynomial equation for the curvilinear relationship between the index 385 of specialization H₂' and community age.

386 Results

387 Composition of communities

388 Sites were selected using climatic and lidar data to restrict abiotic and biotic variation between sites so 389 the effect of community age on ecological network structure could best be explored (SI Table 1 and SI 390 Table 2). There was some variation in forest structure as would be expected with sites during primary 391 succession (e.g., forest height and density changes) (SI Figure 2). Our ecological networks document 34 392 plant genera and 3517 arthropod OTUs, distributed across six classes: Entognatha, Crustacea 393 (Amphipods and Isopods), Insecta, Arachnida, Chilopoda, and Diplopoda. The arthropod-plant 394 associations in our networks represent many kinds of trophic and non-trophic biotic interactions that 395 capture functional differences among species of the understory of the Hawaiian native forest. The 396 barcode reference library increased taxonomic assignment from low taxonomic resolution to genus or 397 species for 401 OTUs. Confident assignment was accomplished for a percentage of OTUs at each taxonomic level: Order 99.9%, Family 67.3%, Genus 38.1% and Species 24.9% (SI Table 3). 398

400 There were 2747 OTUs classified as native and 770 classified as non-native using NIClassify. The overall 401 accuracy for our dataset predictions of nativeness using NIClassify was 99.9%. Of the native OTUs, 402 Hemiptera were the dominant order (652 OTUs), followed by Araneae (467 OTUs), Diptera (327 OTUs), 403 and Coleoptera (266 OTUs). We found a highly significant (SI Table 4) increase in network size with 404 community age for both nodes and links, with a disproportionate increase in the number of links 405 (interactions) after several hundred years of community development (Figure 2a). The number of native 406 arthropod species increases dramatically over both ecological and evolutionary time while the number of 407 non-native arthropod species remains relatively steady (Figure 2b). The abundance of native and non-408 native arthropods peaks in the middle-aged communities but the proportion of non-native taxon 409 abundance is highest in younger communities (Figure 2c). Plant diversity increased with community age 410 (Figure 2d). 411 412 Arthropod-plant association networks 413 414 Arthropod OTU richness, plant diversity and number of interactions increased with the geologic age of the 415 site. Bipartite networks of younger communities contain linkage widths between the few dominant taxa 416 (e.g., Hemiptera and Metrosideros) while older communities contain smaller linkage widths representative 417 of the many more associations distributed among the greater diversity of both higher and lower level taxa 418 (Figure 3, SI Figure 3). 419 420 For the null model analyses of the weighted matrices, some observed network metric values were not

421 significantly different (p<0.05) from metric values produced from the synthetic matrices (SI Table 5, SI422 Figure 6).

423

Results of the Spearman's correlation tests show linkage density (average number of interactions per
species), network vulnerability (a measure of the ratio of plant generic richness to arthropod OTU
richness) and interaction evenness (a measure of the uniformity of energy flows along different pathways)
increased significantly with community age (Figure 4, Table 3). Generality (a measure of the ratio of

428 arthropod OTU richness to plant generic richness) and the index of specialization H₂' both increased with 429 community age but were not significantly positively correlated. The index of specialization H2' has a 430 curvilinear relationship with community age, first decreasing then increasing. A second-degree polynomial 431 provides the best approximation of the relationship between the H₂' and community age (F = 6.85, R^2 = 432 0.5392, p < 0.05). By beat sampling and sequencing all plant-associated arthropods, our sampling of 433 arthropod taxa is at finer taxonomic resolution than plants. As a result, generality (links/arthropods) is very 434 close arithmetically to linkage density (links/arthropods + plants) in our dataset because the number of 435 arthropod OTUs is many times greater than the number of plant genera for all communities. Connectance 436 (proportion of realized interactions) was not significantly correlated with increasing community age, but 437 instead is highest at the youngest site, and relatively constant for the remainder of sites. 438 439 For the gualitative metrics calculated from the binary matrices, linkage density (links/species), 440 connectance links/(arthropods*plants), and generality (links/arthropods) were significantly correlated with 441 community age, while vulnerability (links/plants) and the ratio of resource species to consumers 442 (plants/arthropods) was not (SI Figure 4, SI Table 4). The results from plot level analysis are consistent 443 with the site level data and the variance among plots at the same sites is minimal (SI Figure 7). These 444 results help corroborate the trend of increasing specialization over time. 445 446 For the regressions of network metrics against network size, with the exception of generality, quantitative 447 network metrics were not significantly correlated with network size (SI Figure 5A, SI Table 4). By contrast,

448 qualitative metrics were significantly correlated with network size with the exception of the ratio of

resource species to consumers, and vulnerability (SI Figure 5B, SI Table 4).

450

451 Discussion

Using a dataset of biotic associations during the course of community assembly here we present strong evidence of increasing specialization within arthropod communities through evolutionary time. Our DNA metabarcoding data have allowed us to collect a large sample of the arthropods from the understory of Hawaiian forests, representing a broad swath of trophic and non-trophic arthropod-plant associations. As 456 expected, the qualitative metrics were strongly biased by network size (Banašek-Richter et al., 2004; 457 Goldwasser & Roughgarden, 1997) and showed higher linkage density, generality, vulnerability, 458 interaction evenness, and lower connectance in older communities, because the diversity of plants and 459 arthropods was higher in these communities (SI Figure 4, SI Table 4). Our null model analysis helped to 460 demonstrate that the distribution of link weights was itself an important feature of the observed network 461 structure (i.e., not which species they are between). We present clear signatures of change in 462 quantitative, weighted network metrics with community age (Figure 4, Table 3) that resulted from 463 changing community composition and ecological dynamics.

464

465 Ecological processes dominate younger communities

466

467 Theory suggests that the composition of the youngest communities is shaped through colonization from a 468 regional species pool. This expectation is supported by our results, with the younger communities having 469 significantly lower linkage density, vulnerability, and interaction evenness (Figure 4). These results 470 indicate that species in younger communities are interacting with greater frequency along less uniform 471 interaction pathways, compared to species assemblages at older sites. However, notably the very 472 youngest site, the 1973 lava flow, is an outlier. At the 1973 lava flow, linkage density is high (LD = 24.2), 473 likely reflecting strong environmental filtering and an opportunistic community of generalist species 474 (Bufford et al., 2020; Kortsch et al., 2015) suited for survival during primary succession. At other young 475 sites, linkage density is low, from < 10 (Tree Planting Rd.) whereas it peaks and levels off at Maui (LD = 476 38.6) and Kauai (LD = 38.9), respectively. Thus, linkage density was low at young sites with low resource 477 diversity while the diversity of interactions increased over evolutionary time in step with increasing 478 community complexity.

479

Interaction evenness was low, as expected, on the youngest sites, again with the exception of the 1973 flow. As a measure of the uniformity of energy flows along different pathways, we expected interaction evenness to be low in young communities because some interaction partners would dominate the associations in the network. For example, a large proportion of interactions on the youngest sites (< 300</p> 484 years old) belong to the associations of Hemiptera and Collembola species with early successional plant 485 species, Metrosideros polymorpha and Dicranopteris linearis (Figure 3, SI Figure 3). Low interaction 486 evenness has also been demonstrated among bees and wasps and their associated natural enemies 487 (e.g. parasitoids) under conditions of intensive management (Staniczenko et al., 2017; Tylianakis et al., 488 2007). The early successional communities in Hawaii are ecologically similar to highly modified sites, due 489 to the recent disturbance from lava and the paucity of resource diversity. We suggest that the higher 490 interaction evenness at the 1973 lava flow is due to the extremely limited resources (plants) on the 491 sparsely vegetated lava substrate. At this site colonists may be joined by a relatively large representation 492 of transient arthropods, which may be less host-specific and appear randomly associated with the 493 available plants, increasing interaction evenness. Connectance also peaked in the youngest community 494 (1973 lava flow) likely due to the greater representation of generalists within this network (Kortsch et al., 495 2015; Ponisio et al., 2019).

496

497 The network metric values are less consistent among the youngest sites compared to the older sites 498 (Figure 4). This is likely due to the relatively rapid changes in community composition in early primary 499 succession (Atkinson, 1970; Roderick et al., 2012) compared to older established sites. An alternative 500 explanation is that change in the composition of the understory plants (Figure 3 and SI Figure 3) and 501 canopy structure (SI Figure 2 and SI Table 2) results in the network metric variation at the youngest sites. 502 The higher variation among network values at the youngest sites may also point to the different rates of 503 specialization and adaptation among different lineages of arthropods. Among functional groups of beetles 504 (e.g., xylophages, fungivores, predators), community composition and network specialization changed 505 differently during early succession (Wende et al., 2017).

506

507 Specialization increases through evolutionary time

508

509 For a given taxon on average, the number of biotic interactions it is involved in decreases with community 510 age, resulting in greater network specialization. This is reflected in the increased linkage density with 511 community age, as early colonizing species gave way to a greater diversity of associations (Figure 4). 512 However, weighted connectance stabilized at around the same level for the remainder of the communities 513 after the 1973 lava flow. This may be explained by the 'constant connectance' hypothesis (Martinez, 514 1992) that posits that species are linked to a fixed fraction of species in a network, independent of the 515 number of species in a community. A similar pattern of constant connectance and community age was 516 found in arthropods recolonizing defaunated mangrove islands (Piechnik et al., 2008). For the Hawaiian 517 Islands, several factors likely produce constant connectance over long-term community development. 518 First, resource availability limits specialists at early stages; e.g., Escape Road (~ 300 yrs) is dominated by 519 a single species of fern. Next, over evolutionary time, the Hawaiian fauna is characterized by a 520 remarkably high rate of lineage diversification (Gillespie, 2016; Gillespie et al., 2020; Zimmerman, 1970) 521 that has added novel species and associations. Finally, at more recent timescales (after human arrival) 522 immigration of non-natives has been sufficiently high so as to add generalist taxa across all stages of 523 community development (Figure 2b).

524

525 A previous study which used an island chronosequence to examine how pollinator interactions change 526 through extended time (Trøjelsgaard et al 2013) also found connectance was poorly explained by age. 527 However, contrary to our results, the Canary Islands study showed hump-shaped relationships of 528 interaction richness and specialization with island age. One reason for the different results is that we used 529 a natural log scale for the skewness of island age. For the Hawaiian islands, values for linkage density, 530 vulnerability, interaction evenness, and index of specialization H₂' were especially high on the volcano of 531 East Maui. The islands of Maui Nui are also where richness peaks for many native arthropod lineages 532 (Gillespie & Baldwin, 2009; Gruner, 2007). However, unlike the Canary Island pollinators, our values of 533 linkage density were highest on the oldest island, and values for interaction evenness, vulnerability, and 534 index of specialization H_2 ' were nearly as high, indicating that the overall changes in network structure 535 were more linear than hump-shaped. An alternative explanation for the difference in the results is that the 536 older islands of the Canary archipelago have environments that are very different from the younger 537 islands. Although the Canary Island study focused on communities that were characterized by the plant 538 species Euphorbia balsamifera, the abiotic environment changes significantly across their 539 chronosequence, with the older islands much lower and drier (Juan et al., 2000). Thus, the finding of a

540 hump-shaped relationship in the Canary Islands is associated with the combined effects of time, island 541 geomorphological transitions, and associated change in climate regimes. In contrast, the current study in 542 the Hawaiian Islands aimed to standardize environments (elevation, precipitation, and forest cover, with 543 sampling from standardized plots across the islands). Therefore, any confounding environmental 544 differences were minimized and changes in network properties should largely reflect the influence of 545 community age.

546

547 Vulnerability and generality both show positive correlations with community age (Figure 4, Table 3), thus 548 the average number of arthropods per plant species (vulnerability) and the average number of plant 549 species per arthropod (generality) are increasing over time. This is consistent with our expectation that 550 specialization increases resource overlap when a reduction in antagonistic interactions leads to some 551 level of resource redundancy and an increase in diversity of beneficial interactions leads to greater 552 resource complementarity (Table 1). In other words, over evolutionary time, if two species are in direct 553 competition for resources, they can evolve traits that allow them to coexist. One result of trait matching 554 between interaction partners is decreasing niche breadth (i.e., decreasing diversity of resources used). 555 Thus, our results are consistent with decreasing niche breadth with island age found previously from 556 literature for herbivores (Ponisio et al., 2019). Moreover, although the rate of specialization and 557 adaptation, such as occurs through trait matching and decreasing niche breadth, can vary among 558 functional groups in a community, our data show that community specialization is happening at the 559 network level, averaging over the high variation in rates of specialization.

560

The network-level specialization index H2' is largely unaffected by network size, network architecture or total number of interactions for a fixed matrix size (Blüthgen et al. 2006), making it an ideal metric compare between different networks for understanding specialization over time. We find that the index of specialization increases over time but is better fit by a second-degree polynomial equation. In early stage communities from 50 to 575 years the index of specialization is decreasing. This drop in specialization in the first several hundred years, is followed by an increase over the next tens of thousands of years. For random associations H₂' is usually close to zero. On Maui it reaches a value of 0.6 then levels out to 0.5 on Molokai and Kauai. This pattern is consistent with other metrics in our analysis suggesting that very
young communities are organized by assembly rules making them appear specialized. After ecological
sorting and the impact of *in situ* evolution in later stage communities we see organization at a secondary,
evolutionary stage of development.

572

573 Resilience of communities increases through time

574

While communities sampled from the youngest sites are composed primarily of native species from the regional pool (Figure 2b), younger communities have proportionally higher abundances of non-native species infiltrating the system (Figure 2c). Thus, it appears that young communities are more invasible, which is consistent with previous studies showing that communities composed of endemic generalist taxa are more vulnerable to infiltration by non-natives (Olesen et al., 2002). By increasing connectance and lowering network specialization, higher numbers of alien species may in turn facilitate increasing numbers and impacts of invasions (Simberloff, 2006; Simberloff & Von Holle, 1999).

582

583 However, this result runs counter to work suggesting higher-connectance food webs tend to host fewer 584 invaders and exert stronger biotic resistance compared to low-connectance webs (Smith-Ramesh et al., 585 2017). Further, community resistance to invasion is known to increase with native species diversity 586 (Gallien & Carboni, 2017) and network complexity (Wei et al., 2015). Considering the results from our 587 study within the context of this previous work, older communities, which are characterized by low 588 connectance and high specialization, may be more resistant to invasion; however, individual taxa may be 589 more susceptible to extinction. From an individual species level, because all species are linked together 590 either directly or indirectly (Montoya et al., 2006), individual species with high specialization and low 591 connectance are susceptible to extinction because of secondary extinctions occurring when specialized 592 consumers lose their only prey (Dunne et al., 2002; Staniczenko et al., 2010). From a network level, as 593 communities age, several species may be associated with the same resource (resource redundancy) or 594 utilize a single resource more effectively (resource complementarity), minimizing variability in the

functioning of an ecosystem, for example when some consumer species decline in number (Peralta et al.,2014).

597

598 Although ecological processes, such as interspecific interactions or disturbance, are often attributed to 599 the geographical differences in exotic species richness (Lockwood et al., 2013) an alternative explanation 600 for the apparent reduced biotic resistance to invasion of younger communities may be that they 601 experience increased propagule pressure (Lockwood et al., 2005). The younger sites on Kilauea volcano 602 are accessed more frequently by tourists compared to the older sites, which require greater on foot 603 distances to reach or special access permits. Furthermore, while our study directly assesses arthropod-604 plant associations, it only indirectly measures the effect of higher trophic associations. Differential top-605 down pressure (e.g., predator turnover) during community assembly likely also changes biotic resistance 606 to invasion, for example, generalist insectivorous birds reduced infiltration of an invasive species of spider 607 at the 133 years old Tree Planting Rd. community (Gruner, 2005).

608

609 Conclusions and outlook

610

611 Our study uses whole-community DNA metabarcoding data to assess the biotic associations of 612 thousands of arthropod OTUs on plants across a geological chronosequence. By including relative 613 abundance data, we achieve a signature of interaction strength (Popovic et al., 2019) not captured for co-614 occurrences with presence-absence observations (Blanchet et al., 2020). Although DNA metabarcoding 615 can be used for observation of trophic interactions (Alberdi et al., 2019; Krehenwinkel, Kennedy, et al., 2017), our analysis instead includes all biotic associations between arthropod-plant communities, 616 617 including those that can be difficult to detect (e.g. involving cryptic species, new non-natives, endangered 618 species, juveniles). Thus, we are able to include complex community interactions including substrates 619 chosen for acoustic signaling (Mullet et al., 2017), predator avoidance (Lindstedt et al., 2019; Stachowicz 620 & Hay, 1999) and gregarious plant-feeding insects (Hunter, 2000) that are often overlooked in traditional 621 network studies. Compared to the limitations of small, unweighted early food web studies (Cohen et al.,

622 1993; Hall & Raffaelli, 1991) DNA metabarcoding offers exciting avenues forward for capturing623 community complexity.

624

625 This research revealed a strong association between the network structure of ecological communities and 626 community development over evolutionary time. Quantitative network metrics demonstrate that younger 627 communities are composed of more generalist species that interact with greater frequency along fewer 628 interaction pathways, with individual and network specialization increasing with community age. Our data 629 highlight the utility of DNA metabarcoding for understanding longstanding questions of ecology and 630 evolutionary biology that remain time consuming (e.g., keying out morphological species) or impossible 631 (e.g., identification of juveniles) to assess with traditional methods. From a conservation perspective, our 632 results indicate that habitat disturbance erodes a complex web of biotic associations, far greater than the 633 sum of the community metrics of richness and abundance, that have evolved in situ over thousands to 634 millions of years.

635

636 Acknowledgements

637

638 We would like to thank the field collection team including Curtis Ewing, Andy Rominger, Elske Tielens, 639 Thomas Fezza, Kylle Roy, Christa Nicols, Madeline Stark, Brendan Cote, Kalena Shiroma, Loreto 640 Villegas-Villeza, and Monica Sheffer as well as the many undergraduate students at UH Hilo and UC 641 Berkeley that helped to identify arthropod specimens and sort them to size. Airborne laser scanning data 642 collection and processing was provided by Greg Asner of Arizona State University's Global Airborne 643 Observatory. The GAO is made possible by support from private foundations, visionary individuals, and 644 Arizona State University. We are indebted to many scientists and land managers in Hawaii who have 645 provided access to the lands: Pat Bily (The Nature Conservancy of Hawaii), Melissa Dean, Christian 646 Giardina, and Tabetha Block (Hawaii Experimental Tropical Forests), the late Betsy Gagne (Natural Area 647 Reserve System), Lisa Hadway, Steve Bergfeld, and Joey Mello (Department of Forestry and Wildlife 648 Hilo), Cynthia King and Charmian Dang (Department of Land and Natural Resources) and Rhonda Loh 649 (Hawaii Volcanoes National Park). N.R.G. was supported by the Robert van den Bosch Memorial

- 650 Scholarship and Philomathia Environmental Science Fellowship. The research was supported by an NSF
- Dimensions in Biodiversity grant to R.G.G. (DEB 1241253) and D.S.G. (DEB-1240774).

- 653 References
- Aizen, M. A., Morales, C. L., & Morales, J. M. (2008). Invasive mutualists erode native pollination webs.
 PLoS Biology, 6(2), e31.
- Alberdi, A., Aizpurua, O., Bohmann, K., Gopalakrishnan, S., Lynggaard, C., Nielsen, M., & Gilbert, M. T. P.
 (2019). Promises and pitfalls of using high-throughput sequencing for diet analysis. *Molecular Ecology Resources*, 19(2), 327–348. https://doi.org/10.1111/1755-0998.12960
- Andersen, J. C., Oboyski, P., Davies, N., Charlat, S., Ewing, C., Meyer, C., Krehenwinkel, H., Lim, J. Y.,
 Noriyuki, S., Ramage, T., Gillespie, R. G., & Roderick, G. K. (2019). Categorization of species as
 native or nonnative using DNA sequence signatures without a complete reference library.
 Ecological Applications, 29(5), e01914. https://doi.org/10.1002/eap.1914
- Andujar, C., Creedy, T. J., Arribas, P., López, H., Salces-Castellano, A., Pérez-Delgado, A. J., Vogler, A. P., &
 Emerson, B. C. (2021). Validated removal of nuclear pseudogenes and sequencing artefacts from
 mitochondrial metabarcode data. *Molecular Ecology Resources*, *21*(6), 1772–1787.
- Asner, G. P., Hughes, R. F., Vitousek, P. M., Knapp, D. E., Kennedy-Bowdoin, T., Boardman, J., Martin, R.
 E., Eastwood, M., & Green, R. O. (2008). Invasive plants transform the three-dimensional
 structure of rain forests. *Proceedings of the National Academy of Sciences*, 105(11), 4519–4523.
 https://doi.org/10.1073/pnas.0710811105
- Asner, G. P., Knapp, D. E., Boardman, J., Green, R. O., Kennedy-Bowdoin, T., Eastwood, M., Martin, R. E.,
 Anderson, C., & Field, C. B. (2012). Carnegie Airborne Observatory-2: Increasing science data
 dimensionality via high-fidelity multi-sensor fusion. *Remote Sensing of Environment*, *124*, 454–
 465.
- Asner, G. P., Knapp, D. E., Kennedy-Bowdoin, T., Jones, M. O., Martin, R. E., Boardman, J. W., & Field, C.
 B. (2007). Carnegie Airborne Observatory: In-flight fusion of hyperspectral imaging and
 waveform light detection and ranging for three-dimensional studies of ecosystems. *Journal of Applied Remote Sensing*, 1(1), 013536. https://doi.org/10.1117/1.2794018
- Atkinson, A. E. (1970). Successional Trends in the Coastal and Lowland Forest of Mauna Loa and Kilauea
 Volcanoes, Hawaii! *PACIFIC SCIENCE*, *24*, 14.
- Bálint, M., Pfenninger, M., Grossart, H.-P., Taberlet, P., Vellend, M., Leibold, M. A., Englund, G., &
 Bowler, D. (2018). Environmental DNA time series in ecology. *Trends in Ecology & Evolution*,
 33(12), 945–957.
- Banašek-Richter, C., Cattin, M.-F., & Bersier, L.-F. (2004). Sampling effects and the robustness of
 quantitative and qualitative food-web descriptors. *Journal of Theoretical Biology*, *226*(1), 23–32.
 https://doi.org/10.1016/S0022-5193(03)00305-9
- Bersier, L.-F., Banašek-Richter, C., & Cattin, M.-F. (2002). Quantitative Descriptors of Food-Web
 Matrices. *Ecology*, *83*(9), 2394–2407.
- Blanchet, F. G., Cazelles, K., & Gravel, D. (2020). Co-occurrence is not evidence of ecological interactions.
 Ecology Letters, 23(7), 1050–1063.
- Blüthgen, N., Menzel, F. & Blüthgen, N. (2006). Measuring specialization in species interaction networks.
 BMC Ecol 6, 9. https://doi.org/10.1186/1472-6785-6-9

- Bohan, D. A., Caron-Lormier, G., Muggleton, S., Raybould, A., & Tamaddoni-Nezhad, A. (2011).
 Automated discovery of food webs from ecological data using logic-based machine learning.
 PLoS One, 6(12), e29028.
- Boon, E., Meehan, C. J., Whidden, C., Wong, D. H.-J., Langille, M. G., & Beiko, R. G. (2014). Interactions in
 the microbiome: Communities of organisms and communities of genes. *FEMS Microbiology Reviews*, 38(1), 90–118.
- Brodie, J. F. (2017). Evolutionary cascades induced by large frugivores. *Proceedings of the National Academy of Sciences*, *114*(45), 11998–12002.
- Bufford, J. L., Hulme, P. E., Sikes, B. A., Cooper, J. A., Johnston, P. R., & Duncan, R. P. (2020). Novel
 interactions between alien pathogens and native plants increase plant–pathogen network
 connectance and decrease specialization. *Journal of Ecology*, *108*(2), 750–760.
- Castro-Urgal, R., & Traveset, A. (2014). Differences in flower visitation networks between an oceanic and
 a continental island. *Botanical Journal of the Linnean Society*, *174*(3), 478–488.
 https://doi.org/10.1111/boj.12134
- Chamberlain, S. A., Cartar, R. V., Worley, A. C., Semmler, S. J., Gielens, G., Elwell, S., Evans, M. E., Vamosi,
 J. C., & Elle, E. (2014). Traits and phylogenetic history contribute to network structure across
 Canadian plant–pollinator communities. *Oecologia*, *176*(2), 545–556.
- Clague, D. A. (1996). The growth and subsidence of the Hawaiian-Emperor volcanic chain. *The Origin and Evolution of Pacific Island Biotas, New Guinea to Eastern Polynesia: Patterns and Processes,* 35–
 50.
- Clare, E. L. (2014). Molecular detection of trophic interactions: Emerging trends, distinct advantages,
 significant considerations and conservation applications. *Evolutionary Applications*, 7(9), 1144–
 1157.
- Cohen, J. E., Beaver, R. A., Cousins, S. H., DeAngelis, D. L., Goldwasser, L., Heong, K. L., Holt, R. D., Kohn,
 A. J., Lawton, J. H., & Martinez, N. (1993). Improving food webs. *Ecology*, 74(1), 252–258.
- Coux, C., Rader, R., Bartomeus, I., & Tylianakis, J. M. (2016). Linking species functional roles to their
 network roles. *Ecology Letters*, *19*(7), 762–770. https://doi.org/10.1111/ele.12612
- Cowie, R. H. (1995). Variation in Species Diversity and Shell Shape in Hawaiian Land Snails: In Situ
 Speciation and Ecological Relationships. *Evolution*, 49(6), 1191–1202.
 https://doi.org/10.1111/j.1558-5646.1995.tb04446.x
- de Kerdrel, G. A., Andersen, J. C., Kennedy, S. R., Gillespie, R., & Krehenwinkel, H. (2020). Rapid and cost effective generation of single specimen multilocus barcoding data from whole arthropod
 communities by multiple levels of multiplexing. *Scientific Reports*, *10*(1), 1–12.
- Dell, J. E., Salcido, D. M., Lumpkin, W., Richards, L. A., Pokswinski, S. M., Loudermilk, E. L., O'Brien, J. J., &
 Dyer, L. A. (2019). Interaction diversity maintains resiliency in a frequently disturbed ecosystem.
 Frontiers in Ecology and Evolution, *7*, 145.
- Delmas, E., Besson, M., Brice, M.-H., Burkle, L. A., Dalla Riva, G. V., Fortin, M.-J., Gravel, D., Guimarães
 Jr., P. R., Hembry, D. H., Newman, E. A., Olesen, J. M., Pires, M. M., Yeakel, J. D., & Poisot, T.
 (2019). Analysing ecological networks of species interactions. *Biological Reviews*, *94*(1), 16–36.
 https://doi.org/10.1111/brv.12433
- Dormann, C. F., Gruber, B., & Fründ, J. (2008). Introducing the bipartite package: Analysing ecological
 networks. *Interaction*, 1(0.2413793).
- Dunne, J. A., & Williams, R. J. (2009). Cascading extinctions and community collapse in model food webs.
 Philosophical Transactions of the Royal Society B: Biological Sciences, 364(1524), 1711–1723.
- Dunne, J. A., Williams, R. J., & Martinez, N. D. (2002). Network structure and biodiversity loss in food
 webs: Robustness increases with connectance. *Ecology Letters*, 5(4), 558–567.
- Edgar, R. C. (2010). Search and clustering orders of magnitude faster than BLAST. *Bioinformatics*, 26(19),
 2460–2461.

- Edgar, R. C. (2016). UNOISE2: Improved error-correction for Illumina 16S and ITS amplicon sequencing.
 BioRxiv, 081257.
- Elbrecht, V., & Leese, F. (2015). Can DNA-based ecosystem assessments quantify species abundance?
 Testing primer bias and biomass—sequence relationships with an innovative metabarcoding
 protocol. *PloS One*, *10*(7), e0130324.
- Faust, K., & Raes, J. (2012). Microbial interactions: From networks to models. *Nature Reviews Microbiology*, *10*(8), 538–550. https://doi.org/10.1038/nrmicro2832
- Fricke, E. C., Tewksbury, J. J., Wandrag, E. M., & Rogers, H. S. (2017). Mutualistic strategies minimize
 coextinction in plant–disperser networks. *Proceedings of the Royal Society B: Biological Sciences*,
 284(1854), 20162302. https://doi.org/10.1098/rspb.2016.2302
- Gagne, W. C., & Cuddihy, L. W. (1990). *Vegetation. Manual of the flowering plants of Hawaii*. University
 of Hawaii Press, Honolulu.
- Gallien, L., & Carboni, M. (2017). The community ecology of invasive species: Where are we and what's
 next? *Ecography*, 40(2), 335–352. https://doi.org/10.1111/ecog.02446
- Gap Analysis Project | U.S. Geological Survey. (2019, July 29). https://www.usgs.gov/programs/gap analysis-project
- Giambelluca, T. W., Chen, Q., Frazier, A. G., Price, J. P., Chen, Y.-L., Chu, P.-S., Eischeid, J. K., & Delparte,
 D. M. (2013). Online rainfall atlas of Hawai 'i. *Bulletin of the American Meteorological Society*,
 94(3), 313–316.
- Gibson, J., Shokralla, S., Porter, T. M., King, I., Konynenburg, S. van, Janzen, D. H., Hallwachs, W., &
 Hajibabaei, M. (2014). Simultaneous assessment of the macrobiome and microbiome in a bulk
 sample of tropical arthropods through DNA metasystematics. *Proceedings of the National Academy of Sciences*, *111*(22), 8007–8012. https://doi.org/10.1073/pnas.1406468111
- Gillespie. (2016). Island time and the interplay between ecology and evolution in species diversification.
 Evolutionary Applications, 9(1), 53–73. https://doi.org/10.1111/eva.12302
- Gillespie, R. G., & Baldwin, B. G. (2009). Island biogeography of remote archipelagoes. In J. B. Losos & R.
 E. Ricklefs (Eds.), *The Theory of Island Biogeography Revisited* (pp. 358–387). Princeton
 University Press.
- Gillespie, R. G., Bennett, G. M., De Meester, L., Feder, J. L., Fleischer, R. C., Harmon, L. J., Hendry, A. P.,
 Knope, M. L., Mallet, J., & Martin, C. (2020). Comparing adaptive radiations across space, time,
 and taxa. *Journal of Heredity*, *111*(1), 1–20.
- Goldwasser, L., & Roughgarden, J. (1997). Sampling Effects and the Estimation of Food-Web Properties.
 Ecology, 78(1), 41–54. https://doi.org/10.1890/0012-9658(1997)078[0041:SEATEO]2.0.CO;2
- Gordon, A., & Hannon, G. J. (2010). Fastx-toolkit. FASTQ/A Short-Reads Preprocessing Tools
 (Unpublished) Http://Hannonlab. Cshl. Edu/Fastx toolkit, 5.
- Graham, N. R., Gillespie, R. G., & Krehenwinkel, H. (2021). Towards Eradicating the Nuisance of Numts
 and Noise in Molecular Biodiversity Assessment. *Molecular Ecology Resources*, n/a(n/a).
 https://doi.org/10.1111/1755-0998.13414
- Gruner, D. S. (2005). Biotic resistance to an invasive spider conferred by generalist insectivorous birds on
 Hawai'i Island. *Biological Invasions*, 7(3), 541–546. https://doi.org/10.1007/s10530-004-2509-2
- Gruner, D. S. (2007). Geological age, ecosystem development, and local resource constraints on
 arthropod community structure in the Hawaiian Islands. *Biological Journal of the Linnean Society*, 90(3), 551–570.
- Hall, S. J., & Raffaelli, D. (1991). Food-web patterns: Lessons from a species-rich web. *The Journal of Animal Ecology*, 823–841.
- Hembry, D. H., Raimundo, R. L., Newman, E. A., Atkinson, L., Guo, C., Guimarães Jr, P. R., & Gillespie, R.
 G. (2018). Does biological intimacy shape ecological network structure? A test using a brood

787 pollination mutualism on continental and oceanic islands. Journal of Animal Ecology, 87(4), 788 1160-1171. 789 Hrček, J., & Godfray, H. C. J. (2015). What do molecular methods bring to host-parasitoid food webs? 790 Trends in Parasitology, 31(1), 30–35. 791 Hubbell, S. P. (2001). The Unified Neutral Theory of Species Abundance and Diversity. Princeton 792 University Press. 793 Hunter, A. F. (2000). Gregariousness and repellent defences in the survival of phytophagous insects. 794 Oikos, 91(2), 213-224. https://doi.org/10.1034/j.1600-0706.2000.910202.x 795 Juan, C., Emerson, B. C., Oromí, P., & Hewitt, G. M. (2000). Colonization and diversification: Towards a 796 phylogeographic synthesis for the Canary Islands. Trends in Ecology & Evolution, 15(3), 104–109. 797 Kortsch, S., Primicerio, R., Fossheim, M., Dolgov, A. V., & Aschan, M. (2015). Climate change alters the 798 structure of arctic marine food webs due to poleward shifts of boreal generalists. Proceedings of 799 the Royal Society B: Biological Sciences, 282(1814), 20151546. 800 https://doi.org/10.1098/rspb.2015.1546 801 Koskella, B., & Brockhurst, M. A. (2014). Bacteria-phage coevolution as a driver of ecological and 802 evolutionary processes in microbial communities. FEMS Microbiology Reviews, 38(5), 916–931. 803 Koskella, B., Hall, L. J., & Metcalf, C. J. E. (2017). The microbiome beyond the horizon of ecological and 804 evolutionary theory. *Nature Ecology & Evolution*, 1(11), 1606–1615. 805 Krehenwinkel, H., Kennedy, S., Pekár, S., & Gillespie, R. G. (2017). A cost-efficient and simple protocol to 806 enrich prey DNA from extractions of predatory arthropods for large-scale gut content analysis by 807 Illumina sequencing. *Methods in Ecology and Evolution*, 8(1), 126–134. 808 Krehenwinkel, H., Wolf, M., Lim, J. Y., Rominger, A. J., Simison, W. B., & Gillespie, R. G. (2017). Estimating 809 and mitigating amplification bias in qualitative and quantitative arthropod metabarcoding. 810 Scientific Reports, 7(1), 1–12. Lange, V., Böhme, I., Hofmann, J., Lang, K., Sauter, J., Schöne, B., Paul, P., Albrecht, V., Andreas, J. M., & 811 812 Baier, D. M. (2014). Cost-efficient high-throughput HLA typing by MiSeq amplicon sequencing. 813 BMC Genomics, 15(1), 1–11. 814 Lim, J. Y., Patiño, J., Noriyuki, S., Cayetano, L., Gillespie, R. G., & Krehenwinkel, H. (2021). Semi-815 quantitative metabarcoding reveals how climate shapes arthropod community assembly along 816 elevation gradients on Hawaii Island. Molecular Ecology. 817 Lindstedt, C., Murphy, L., & Mappes, J. (2019). Antipredator strategies of pupae: How to avoid predation 818 in an immobile life stage? Philosophical Transactions of the Royal Society B, 374(1783), 819 20190069. 820 Lockwood, J. L., Cassey, P., & Blackburn, T. (2005). The role of propagule pressure in explaining species 821 invasions. Trends in Ecology & Evolution, 20(5), 223–228. 822 https://doi.org/10.1016/j.tree.2005.02.004 823 Lockwood, J. L., Hoopes, M. F., & Marchetti, M. P. (2013). Invasion Ecology. John Wiley & Sons. 824 Martinez, N. D., Hawkins, B. A., Dawah, H. A., & Feifarek, B. P. (1999). Effects of Sampling Effort on 825 Characterization of Food-Web Structure. Ecology, 80(3), 1044–1055. 826 Mata, V. A., da Silva, L. P., Veríssimo, J., Horta, P., Raposeira, H., McCracken, G. F., Rebelo, H., & Beja, P. 827 (2021). Combining DNA metabarcoding and ecological networks to inform conservation 828 biocontrol by small vertebrate predators. Ecological Applications, 31(8), e02457. 829 McGill, B. J., Chase, J. M., Hortal, J., Overcast, I., Rominger, A. J., Rosindell, J., Borges, P. A. V., Emerson, 830 B. C., Etienne, R. S., Hickerson, M. J., Mahler, D. L., Massol, F., McGaughran, A., Neves, P., 831 Parent, C., Patiño, J., Ruffley, M., Wagner, C. E., & Gillespie, R. (2019). Unifying macroecology 832 and macroevolution to answer fundamental questions about biodiversity. Global Ecology and 833 *Biogeography*, 28(12), 1925–1936. https://doi.org/10.1111/geb.13020

834 Miller, Pfeiffer, W., & Schwartz, T. (2010). Creating the CIPRES Science Gateway for inference of large 835 phylogenetic trees. 2010 Gateway Computing Environments Workshop (GCE), 1–8. 836 https://doi.org/10.1109/GCE.2010.5676129 837 Montoya, J. M., Pimm, S. L., & Solé, R. V. (2006). Ecological networks and their fragility. Nature, 838 442(7100), 259-264. 839 Morella, N. M., Weng, F. C.-H., Joubert, P. M., Metcalf, C. J. E., Lindow, S., & Koskella, B. (2020). 840 Successive passaging of a plant-associated microbiome reveals robust habitat and host 841 genotype-dependent selection. Proceedings of the National Academy of Sciences, 117(2), 1148-842 1159. 843 Mullet, T. C., Farina, A., & Gage, S. H. (2017). The Acoustic Habitat Hypothesis: An Ecoacoustics 844 Perspective on Species Habitat Selection. *Biosemiotics*, 10(3), 319–336. 845 https://doi.org/10.1007/s12304-017-9288-5 Münkemüller, T., Gallien, L., Pollock, L. J., Barros, C., Carboni, M., Chalmandrier, L., Mazel, F., Mokany, 846 847 K., Roquet, C., & Smyčka, J. (2020). Dos and don'ts when inferring assembly rules from diversity 848 patterns. Global Ecology and Biogeography, 29(7), 1212–1229. 849 Newman, M. E. J., & Girvan, M. (2004). Finding and evaluating community structure in networks. 850 Physical Review E, 69(2), 026113. https://doi.org/10.1103/PhysRevE.69.026113 851 Oksanen, J., Kindt, R., Legendre, P., O'Hara, B., Stevens, M. H. H., Oksanen, M. J., & Suggests, M. (2007). 852 The vegan package. Community Ecology Package, 10(631–637), 719. 853 Olesen, J. M., Eskildsen, L. I., & Venkatasamy, S. (2002). Invasion of pollination networks on oceanic 854 islands: Importance of invader complexes and endemic super generalists. Diversity and 855 *Distributions*, *8*(3), 181–192. 856 Peralta, A. M., Sánchez, A. M., Luzuriaga, A. L., de Bello, F., & Escudero, A. (2019). Evidence of functional 857 species sorting by rainfall and biotic interactions: A community monolith experimental 858 approach. Journal of Ecology, 107(6), 2772–2788. 859 Peralta, Frost, C. M., Rand, T. A., Didham, R. K., & Tylianakis, J. M. (2014). Complementarity and 860 redundancy of interactions enhance attack rates and spatial stability in host-parasitoid food 861 webs. Ecology, 95(7), 1888–1896. 862 Piechnik, D. A., Lawler, S. P., & Martinez, N. D. (2008). Food-web assembly during a classic biogeographic 863 study: Species'"trophic breadth" corresponds to colonization order. Oikos, 117(5), 665–674. 864 https://doi.org/10.1111/j.0030-1299.2008.15915.x 865 Poisot, T., Stouffer, D. B., & Gravel, D. (2015). Beyond species: Why ecological interaction networks vary through space and time. Oikos, 124(3), 243–251. https://doi.org/10.1111/oik.01719 866 867 Ponisio, L. C., Valdovinos, F. S., Allhoff, K. T., Gaiarsa, M. P., Barner, A., Guimarães Jr, P. R., Hembry, D. 868 H., Morrison, B., & Gillespie, R. (2019). A network perspective for community assembly. 869 Frontiers in Ecology and Evolution, 7, 103. 870 Popovic, G. C., Warton, D. I., Thomson, F. J., Hui, F. K., & Moles, A. T. (2019). Untangling direct species 871 associations from indirect mediator species effects with graphical models. Methods in Ecology 872 and Evolution, 10(9), 1571–1583. 873 R Core Development Team. (2019). R: A language and environment for statistical computing. 874 Roderick, G. K., Croucher, P. J. P., Vandergast, A. G., & Gillespie, R. G. (2012). Species Differentiation on a 875 Dynamic Landscape: Shifts in Metapopulation Genetic Structure Using the Chronology of the 876 Hawaiian Archipelago. Evolutionary Biology, 39(2), 192–206. https://doi.org/10.1007/s11692-877 012-9184-5 878 Rominger, A. J., Goodman, K. R., Lim, J. Y., Armstrong, E. E., Becking, L. E., Bennett, G. M., Brewer, M. S., 879 Cotoras, D. D., Ewing, C. P., Harte, J., Martinez, N. D., O'Grady, P. M., Percy, D. M., Price, D. K., 880 Roderick, G. K., Shaw, K. L., Valdovinos, F. S., Gruner, D. S., & Gillespie, R. G. (2016). Community

881 assembly on isolated islands: Macroecology meets evolution. Global Ecology and Biogeography, 882 25(7), 769–780. https://doi.org/doi:10.1111/geb.12341 883 Rosindell, J., Hubbell, S. P., & Etienne, R. S. (2011). The Unified Neutral Theory of Biodiversity and 884 Biogeography at Age Ten. Trends in Ecology & Evolution, 26(7), 340–348. 885 https://doi.org/10.1016/j.tree.2011.03.024 886 Rosindell, J., & Phillimore, A. B. (2011). A unified model of island biogeography sheds light on the zone of 887 radiation. Ecology Letters, 14(6), 552-560. https://doi.org/10.1111/j.1461-0248.2011.01617.x 888 Segar, S. T., Fayle, T. M., Srivastava, D. S., Lewinsohn, T. M., Lewis, O. T., Novotny, V., Kitching, R. L., & 889 Maunsell, S. C. (2020). The role of evolution in shaping ecological networks. Trends in Ecology & 890 Evolution, 35(5), 454-466. 891 Shaw, & Gillespie, R. G. (2016). Comparative phylogeography of oceanic archipelagos: Hotspots for 892 inferences of evolutionary process. Proceedings of the National Academy of Sciences, 113(29), 893 7986–7993. https://doi.org/10.1073/pnas.1601078113 894 Sievers, F., Wilm, A., Dineen, D., Gibson, T. J., Karplus, K., Li, W., Lopez, R., McWilliam, H., Remmert, M., 895 & Söding, J. (2011). Fast, scalable generation of high-quality protein multiple sequence 896 alignments using Clustal Omega. *Molecular Systems Biology*, 7(1), 539. 897 Simberloff. (2006). Invasional meltdown 6 years later: Important phenomenon, unfortunate metaphor, 898 or both? *Ecology Letters*, 9(8), 912–919. 899 Simberloff, D. S., & Von Holle, B. (1999). Positive interactions of nonindigenous species: Invasional 900 meltdown? *Biological Invasions*, 1(1), 21–32. Smith-Ramesh, L. M., Moore, A. C., & Schmitz, O. J. (2017). Global synthesis suggests that food web 901 902 connectance correlates to invasion resistance. Global Change Biology, 23(2), 465–473. 903 https://doi.org/10.1111/gcb.13460 904 Stachowicz, J. J., & Hay, M. E. (1999). Reducing predation through chemically mediated camouflage: 905 Indirect effects of plant defenses on herbivores. Ecology, 80(2), 495–509. 906 Staniczenko, Lewis, O. T., Tylianakis, J. M., Albrecht, M., Coudrain, V., Klein, A.-M., & Reed-Tsochas, F. 907 (2017). Predicting the effect of habitat modification on networks of interacting species. Nature 908 Communications, 8(1), 1–10. 909 Staniczenko, P. P., Kopp, J. C., & Allesina, S. (2013). The ghost of nestedness in ecological networks. 910 *Nature Communications*, 4(1), 1–6. 911 Staniczenko, P. P., Lewis, O. T., Jones, N. S., & Reed-Tsochas, F. (2010). Structural dynamics and 912 robustness of food webs. *Ecology Letters*, 13(7), 891–899. 913 Trøjelsgaard, K., Báez, M., Espadaler, X., Nogales, M., Oromí, P., Roche, F. L., & Olesen, J. M. (2013). 914 Island biogeography of mutualistic interaction networks. Journal of Biogeography, 40(11), 2020-915 2031. 916 Trøjelsgaard, K., & Olesen, J. M. (2016). Ecological networks in motion: Micro- and macroscopic 917 variability across scales. Functional Ecology, 30(12), 1926–1935. https://doi.org/10.1111/1365-918 2435.12710 919 Tylianakis, J. M., Tscharntke, T., & Lewis, O. T. (2007). Habitat modification alters the structure of 920 tropical host-parasitoid food webs. Nature, 445(7124), 202-205. 921 https://doi.org/10.1038/nature05429 922 Vacher, C., Daudin, J.-J., Piou, D., & Desprez-Loustau, M.-L. (2010). Ecological integration of alien species 923 into a tree-parasitic fungus network. *Biological Invasions*, 12(9), 3249–3259. 924 Vacher, C., Tamaddoni-Nezhad, A., Kamenova, S., Peyrard, N., Moalic, Y., Sabbadin, R., Schwaller, L., 925 Chiquet, J., Smith, M. A., & Vallance, J. (2016). Learning ecological networks from next-926 generation sequencing data. Advances in Ecological Research, 54, 1–39.

- Vázquez, D. P., & Aizen, M. A. (2006). Community-wide patterns of specialization in plant–pollinator
 interactions revealed by null models. *Plant–Pollinator Interactions: From Specialization to Generalization*, 200–219.
- Vázquez, D. P., Melián, C. J., Williams, N. M., Blüthgen, N., Krasnov, B. R., & Poulin, R. (2007). Species
 abundance and asymmetric interaction strength in ecological networks. *Oikos*, *116*(7), 1120–
 1127. https://doi.org/10.1111/j.0030-1299.2007.15828.x
- Venturelli, O. S., Carr, A. V., Fisher, G., Hsu, R. H., Lau, R., Bowen, B. P., Hromada, S., Northen, T., &
 Arkin, A. P. (2018). Deciphering microbial interactions in synthetic human gut microbiome
 communities. *Molecular Systems Biology*, *14*(6), e8157.
- Vitousek, P. M. (2002). Oceanic islands as model systems for ecological studies. *Journal of Biogeography*,
 29(5–6), 573–582.
- Walker, L. R., Wardle, D. A., Bardgett, R. D., & Clarkson, B. D. (2010). The use of chronosequences in
 studies of ecological succession and soil development. *Journal of Ecology*, *98*(4), 725–736.
- Weber, M. G., Wagner, C. E., Best, R. J., Harmon, L. J., & Matthews, B. (2017). Evolution in a Community
 Context: On Integrating Ecological Interactions and Macroevolution. *Trends in Ecology & Evolution*, *32*(4), 291–304. https://doi.org/10.1016/j.tree.2017.01.003
- Wei, Z., Yang, T., Friman, V.-P., Xu, Y., Shen, Q., & Jousset, A. (2015). Trophic network architecture of
 root-associated bacterial communities determines pathogen invasion and plant health. *Nature Communications*, 6(1), 8413. https://doi.org/10.1038/ncomms9413
- 946 Wolfe, E. W., & Morris, J. (1996). *Geologic map of the Island of Hawaii*. US Geological Survey.
- Yeakel, J. D., Pires, M. M., Rudolf, L., Dominy, N. J., Koch, P. L., Guimarães, P. R., & Gross, T. (2014).
 Collapse of an ecological network in Ancient Egypt. *Proceedings of the National Academy of Sciences*. https://doi.org/10.1073/pnas.1408471111
- Yu, D. W., Ji, Y., Emerson, B. C., Wang, X., Ye, C., Yang, C., & Ding, Z. (2012). Biodiversity soup:
 Metabarcoding of arthropods for rapid biodiversity assessment and biomonitoring. *Methods in Ecology and Evolution*, 3(4), 613–623.
- Zhang, Kobert, K., Flouri, T., & Stamatakis, A. (2014). PEAR: A fast and accurate Illumina Paired-End reAd
 mergeR. *Bioinformatics*, *30*(5), 614–620.
- Zimmerman, E. C. (1970). Adaptive Radiation in Hawaii with Special Reference to Insects. *Biotropica*,
 2(1), 32–38. https://doi.org/10.2307/2989786
- 957 958

959 Data Accessibility

- 960 Data are deposited in a Dryad data repository at https://doi.org/10.6078/D1DX4T and code is hosted on
- 261 Zenodo at https://zenodo.org/record/7349067 including: 1) DNA sequence data (fasta files) from whole
- 962 organism community metabarcoding and the DNA barcode reference library, and 2) processed data used
- 963 in these analyses including the OTU table, phylogenetic species ids, NIClassify predictions, and
- 964 geographic information.

965

966 Author Contributions

- 968 processing of the samples, N.R.G. and P.S. analyzed the data, J.C. and J.C.A. contributed new analytical
- tools, N.R.G. and R.G.G. wrote the manuscript with input and comments from all coauthors.

971 Tables and Figures

972 Table 1: Some expectations for network structure in response to ecological and evolutionary

factors influencing community assembly through time. Specialization should change at the network
level to depict the changes at the taxon level of more evolved associations. Since the associations of
each species are becoming more specialized there is an expectation that an individual taxon will have
fewer interactions and the makeup of the whole community will have more interactions overall. [1]
(Bufford et al., 2020) [2] (Vázquez et al., 2007) [3] (Tylianakis et al., 2007) [4] (Peralta et al., 2014) [5]
(Coux et al., 2016) [6] (Ponisio et al., 2019) [7] (Segar et al., 2020) [8] (Chamberlain et al., 2014) [9]

979 (Blüthgen et al. 2006) [10] (Dunne & Williams, 2009)

<u>~</u> ~

Process	Richness	Specialization 3	e.g., metric change	Expected change in metric through extended period of community assembly
Immigration	Ţ	-	Connectance	Connectance should increase in response to immigration when generalist colonists take advantage of weak resource defenses, increasing the number of associations that form out of all possible associations [1]
Environmental filtering	Ļ	¢	Linkage Density	Linkage density should be higher in novel (young or recently disturbed) networks due to generalists with a greater number of interactions per species [1]
Increasing abundance	↑↓	-	Interaction Evenness	Interaction evenness should decrease with dynamics in taxon abundance because skewed frequency distributions (i.e., a few species with many links and many species with few links) are largely driven by species abundance [2]
Antagonistic interactions	¢↓	ţ	Vulnerability	Vulnerability is high in highly modified systems [3] because the average number of consumers per resource is high. Thus, we expect vulnerability to be high in the very youngest community forming on bare lava and go down. However, we expect vulnerability will increase over long temporal scales with reduced competition, predation, parasitism, etc. leading to some level of resource redundancy, for example several species feed on the same resource.
Beneficial interactions	Ť	¢	Generality	Generality should increase with increasing pollination, frugivory, camouflage, etc. because species can share the same resources in different locations or times resulting in resource complementarity. [4]
Increasing niches	Ţ	Ţ	Generality, Vulnerability	The narrowing of interactions to a subset of resources increases specialization. [5] As niches proliferate the fraction of species associations per interaction partner will go down, e.g., niche breadth of herbivores decreased with island age. [6]
Trait diversification	ţ	¢	Linkage Density, H_2 '	Evolution and network structure are linked [7]. For example, traits with a phylogenetic signal (such as flower symmetry and pollinator size) can accurately predict interaction partners. [8] If species traits diversify together (co-evolve) then more one-to-one relationships will result in lower linkage density and higher specialization (H_2 ').
Speciation	¢	ſ	Interaction Evenness	Where food webs are dominated by a single link interaction evenness is lowest. [3] With the addition of species through <i>in situ</i> speciation, the number of links will increase with greater interaction evenness.
Extinction	Ļ	Ļ	Connectance	A specialist species may go extinct from the loss of an essential resource. Species interaction networks composed of many specialist species should have low connectance, and primary extinctions are expected to propagate quickly. [9]

983 Table 2. Binary and Weighted network summary statistics. Metrics calculated from binary (i.e. unweighted, presence-absence) matrices are easily interpretable but sensitive to sampling differences 984 (Banašek-Richter et al., 2004). Quantitative versions based on information theory are more conservative 985 when comparing differences among sites. Each metric incorporates the diversity of individuals comprising 986 987 the resource (H_N , the diversity of inflows) and of that going to the consumers (H_P , the diversity of outflows) 988 for each species k. The quantitive metrics are then based on the reciprocals of these Shannon Entropy 989 values ($n_{N,k}$, and $n_{p,k}$, respectively). The notation q is applied to the quantitive version of that metric. All equations and notations reference [1] (Bersier et al., 2002) [2] (Tylianakis et al., 2007) [3] (Blüthgen et al. 990 991 2006).

992

Summary statistic		Equation or notation	Description
Number of nodes	В	S = R+C	Total number of species (S) or 'nodes' is equal to the number of prey or resource species (R; lower-level) plus the number of consumer species (C; upper-level)
Number of links	В	L	Total number of interactions or 'links'
Ratio resource: consumers	В	R/C [1]	Average number of resource species per consumer species
Diversity of inflows	W	H _{N,k} , (5) [1]	Shannon entropy of weights for a given consumer sp.
Diversity of outflows	W	H _{P,k} , (6) [1]	Shannon entropy of weights for a given resource sp.
Log-reciprocal of (5)	W	n _{N,k} , (7) [1]	Effective number of resource spp. for a given consumer sp.
Log-reciprocal of (6)	W	n _{p,k} , (8) [1]	Effective number of consumer spp. for a given resource sp.
Link density	В	LD = L/S [1]	Average number of interactions per species
	W	LD _q , (14) [1]	weighted version
Connectance	В	Conn = L/(RxC) [1]	Proportion of realized links
	W	Conn _q = LD _q /S [1]	weighted version
Generality	В	G = L/C [1]	Average number of resource sp. per consumer sp.
	W	G _q , (25) [1]	weighted version
Vulnerability	В	V = L/R [1]	Average number of consumer sp. per resource sp.
	W	V _q , (27) [1]	weighted version
Interaction evenness	W	I.E. [2]	Shannon entropy of interaction weights
Index of specialization	W	H2' [3]	Ranges between 0 and 1.0 for extreme generalization and specialization, respectively

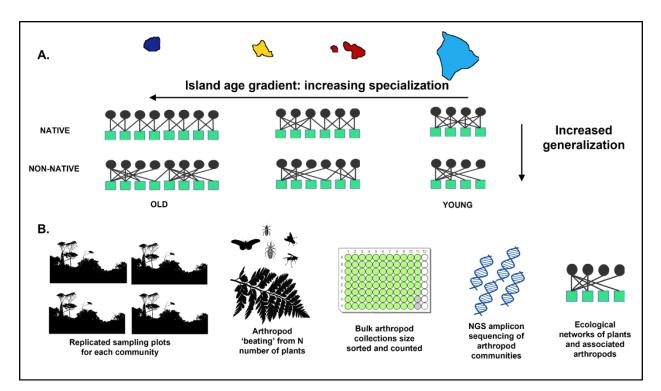
993

Table 3. Spearman's correlation tests for network metrics and community age.

Spearman's correlation tests were used to determine the significance of the relationship between each
quantitative network metric value and In substrate age (community age). Graphs of regressions in Figure
4.

	S	p- value	Spearman's rho
Quantitative (weighted) x community age			
linkage density	76	0.033	0.65
weighted connectance	154	0.371	0.30
generality	124	0.183	0.44
vulnerability	80	0.040	0.64
interaction evenness	70	0.025	0.68
index of specialization H ₂ '	134	0.237	0.39





1001 1002

1003 Figure 1. Study overview. A) Study aims. As communities assemble over time species will be added 1004 through ecological and evolutionary processes. Network size will increase over time. There will be a trend 1005 towards greater specialization as relationships among species are modified through ecological fitting and 1006 evolutionary adaptation over extended time, young to old sites (top panel). Recently introduced species 1007 (i.e., non-natives) evolved elsewhere and have not adapted in place to biotic and abiotic factors, thus 1008 limiting their specialization within communities at all stages of development (bottom panel). B) Study 1009 design. Within multiple 15 m radius plots at 11 communities from ages 50 y to 4.15 Myr plant species were sampled for associated arthropods by vegetation beating according to their relative abundance. 1010 Each sample of plant-associated arthropods was size sorted, counted, and placed into a well of a 96-well 1011 plate, such that well 'A1' contained sample 1, size category 0-2 mm, and well 'A2' contained sample 1, 1012 size category 2-4mm, and so on. DNA extraction and PCR amplification with dual-indexing was used to 1013 1014 prepare the size-sorted samples into amplicon libraries which were sequenced on an Illumina Miseq® for 1015 the cytochrome oxidase I locus. Ecological networks were constructed from the arthropod-plant 1016 associations for each community age.

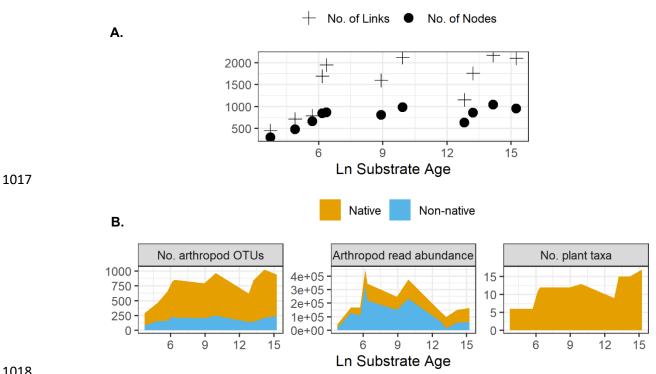
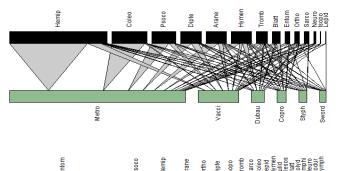
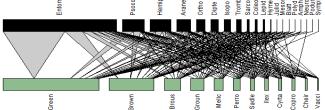


Figure 2. Effect of community assembly over evolutionary time on network size and diversity of native and non-native taxa. A) The number of nodes (arthropod and plant richness) and the number of links (arthropod-plant associations) significantly increase in concert with community age. Spearman's correlation test values SI Table 4. B) Native arthropod richness increases, while non-native richness does not increase, with community age. C) Abundance of native and non-native arthropod species peaks at middle-aged communities but the abundance of non-native taxa is proportionately higher in the youngest communities. D) Native plant richness increases with community age.

1



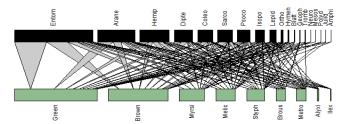


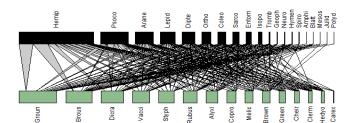


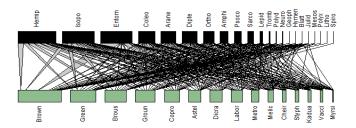
Kilauea: 1973 lava flow 50 yo

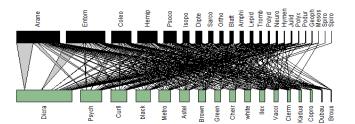
* **

Mauna Loa: Olaa 7,500 yo











Kohala: Kohala 365,000 yo

Maui: Waikamoi 545,000 yo



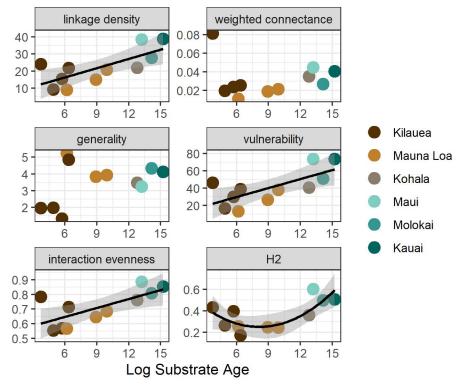
Molokai: Kamakou 1.4 myo



Kauai: Kokee 4.15 myo

1028 Figure 3. Quantitative arthropod-plant networks along a gradient of increasing community 1029 assembly (top to bottom). For each network, lower bars represent plant abundance based on sampling time and upper bars represent arthropod abundance based on OTU frequency. Each network is plotted in 1030 order of the most abundant taxa from left to right so that the turnover in arthropod-plant association can 1031 1032 be seen for each community. Linkage width indicates the frequency of each association as measured 1033 using arthropod read abundance. As a summary, the networks show interaction data pooled across all 1034 plots for each community age with OTUs pooled by arthropod order, but analyses were performed at the OTU per plant genus level. The bipartite graphs from each of the 11 sampled sites are in SI Figure 3. 1035 1036 Arthropod and plant id codes are given in SI Table 3. 1037

1038



1039



1041 Statistical measures of network architecture indicating changes in arthropod-plant associations in concert

1042 with community age. Each network was weighted with the read abundance of the arthropod OTU

1043 associated with the plant genus it was collected from, across all plots for a community age. Three metrics

show significant relationships with community assembly, increasing over time: linkage density,

1045 vulnerability, interaction evenness. Spearman's correlation test values Table 3. Results of the null model

1046 analysis for the quantitative ecological networks metrics are presented in SI Figure 6 and SI Table 5. A 1047 graph of the results when analyzed for each of the sampled plots within a community age site is

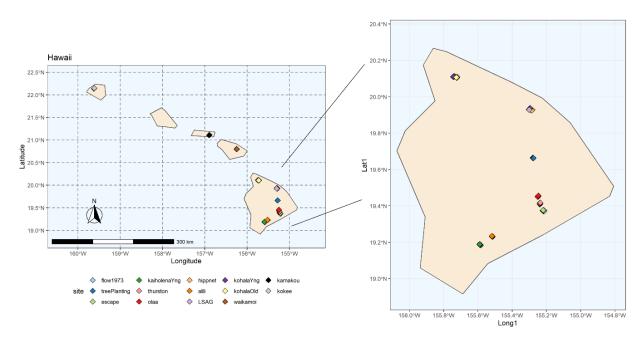
1048 presented in SI Figure 7.



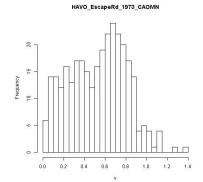
Supplemental Information for:

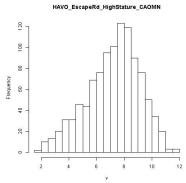
Ecological network structure in response to community assembly processes over evolutionary time

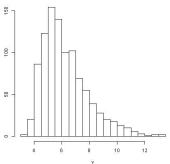
Natalie R. Graham, Henrik Krehenwinkel, Jun Ying Lim, Phillip Staniczenko, Jackson Callaghan, Jeremy C. Andersen, Daniel S. Gruner, Rosemary G. Gillespie



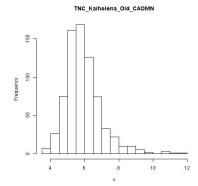
SI Figure 1. Map of sampling sites. 14 sites of varying geologic age were selected, ranging from 50 to 4.15×10^6 years old, across four islands of the archipelago: Hawaii, Maui, Molokai, Kauai. To control for climatic differences and disturbance across sites, sites were constrained to ranges of elevation (1000-1300 m) and precipitation (average annual precipitation 2500-3000 mm) and within accessible protected forest lands. See SI Table 2 for substrate age and other site characteristics.

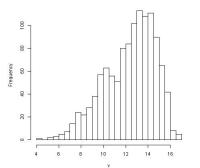




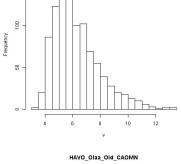


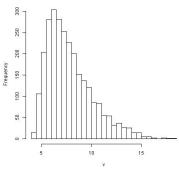
TNC_Kaiholena_Young_CAOMN



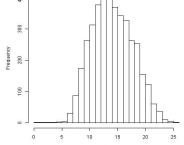


HAVO_Thurston_2polygons_CAOMN

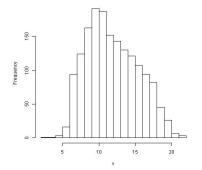




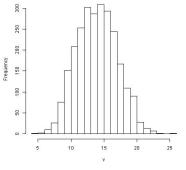


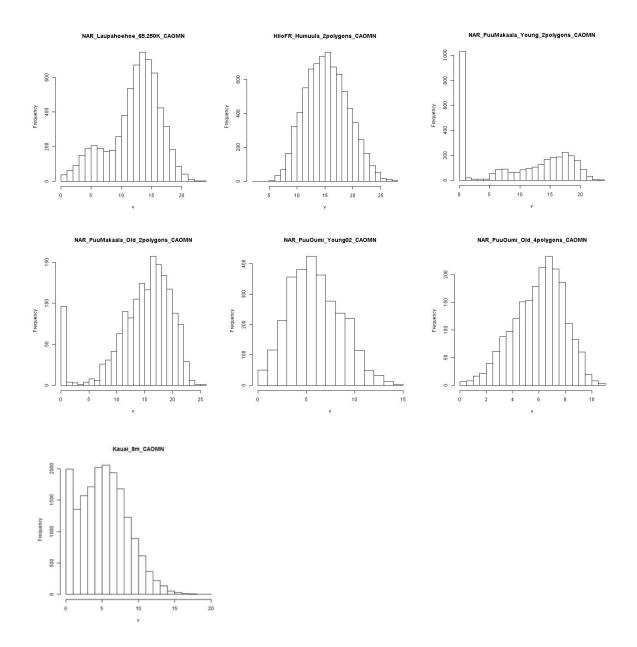




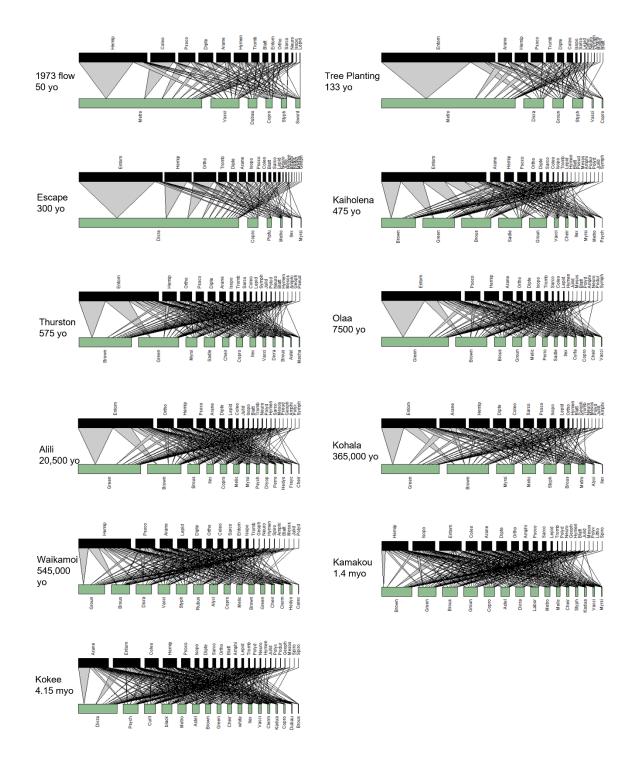


HiloFR_Laupahoehoe_Old_CAOMN



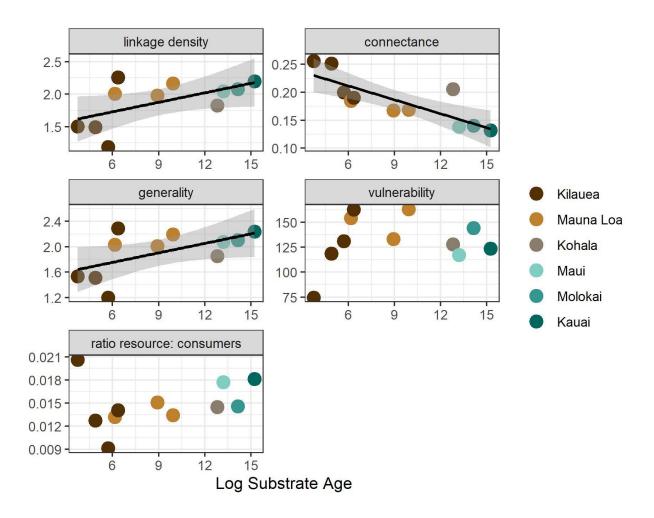


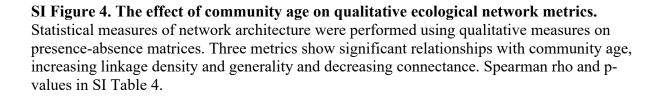
SI Figure 2. Histograms of height profiles from lidar flyover data. Each of the potential sites for the study was assessed for site suitability including these lidar data to evaluate the forest structure. Grain size for lidar was 20 m for all sites except Kauai for which it was 8 m grain size. We used 40% quantile for site comparison because the tails of the distribution of heights was long for some sites, and it would have been difficult to find sites with overlapping height profiles with a more restrictive choice.



SI Figure 3. Additional bipartite network plots. For each network, lower bars represent plant abundance based on sampling time and upper bars represent arthropod abundance based on OTU

frequency. Each network is plotted in order of the most abundant taxa from left to right so that the turnover in arthropod-plant association can be seen for each community. Linkage width indicates the frequency of each association as measured using arthropod read abundance. As a summary, the networks show interaction data pooled across all plots for each community age with OTUs pooled by arthropod order. However, analyses were performed at the OTU per plant genus level. Arthropod and plant id codes are given in Supplementary Table 1.



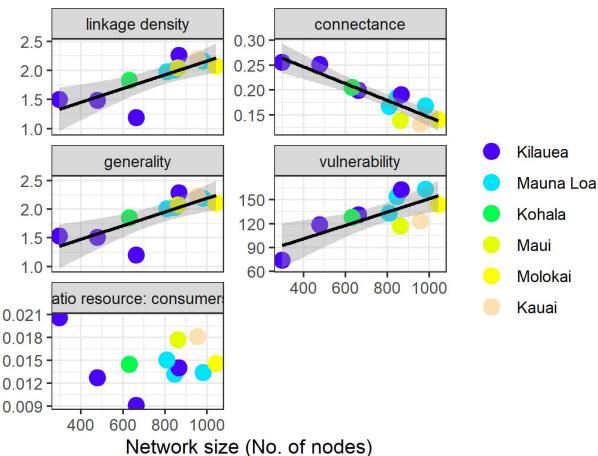




weighted connectance linkage density 40 80.0 30 -0.06 0.04 -20 . 0.02 10 -Kilauea vulnerability generality Mauna Loa 60 · 4. Kohala 40· 2 Maui 20 0 Molokai H2 interaction evenness Kauai 0.9 0.6 -0.5 0.8 0.4 0.7 0.3 0.6 0.2 -600 800 1000 400 600 800 1000 400 Network size (No. of nodes)

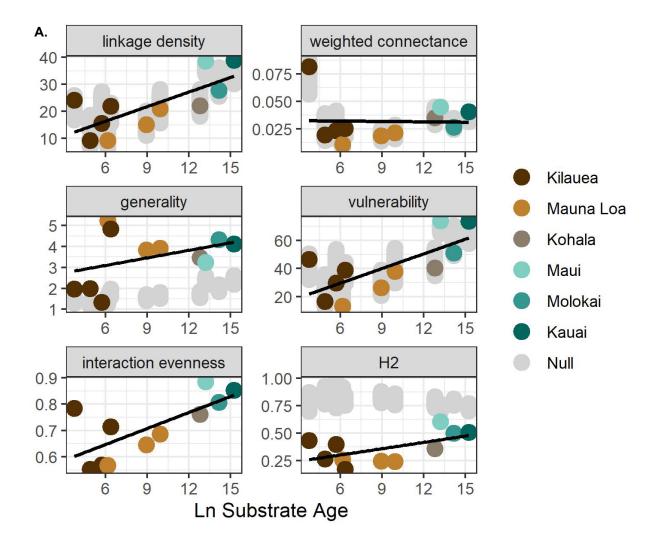
A. The effect of network size on quantitative food web metrics

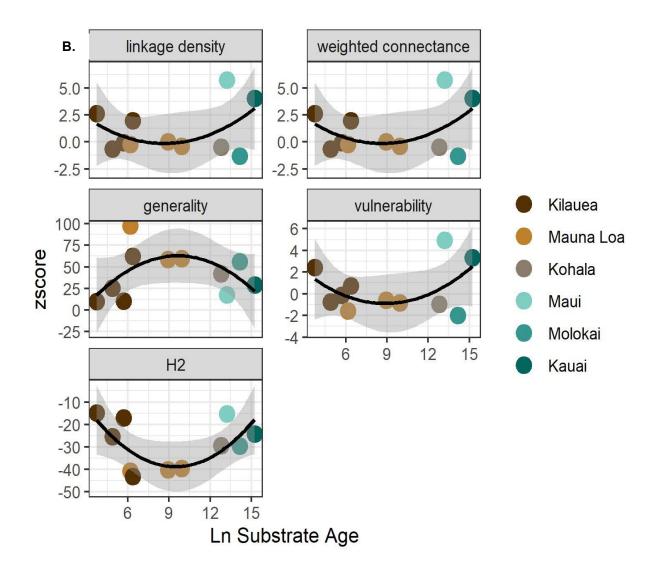




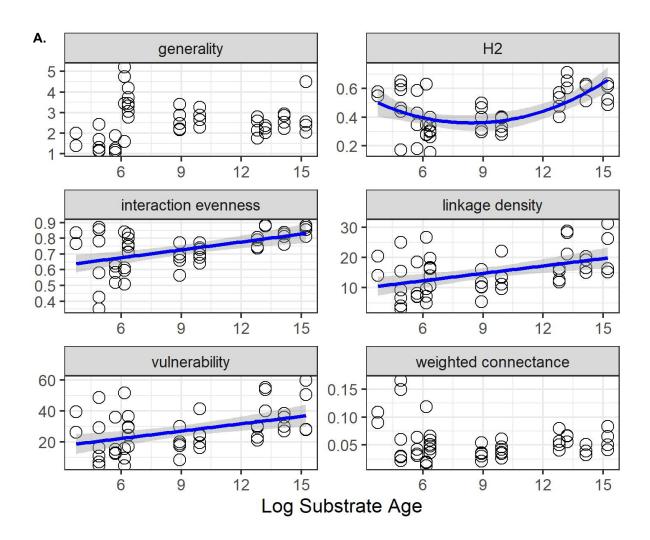
в. The effect of network size on qualitative food web metrics

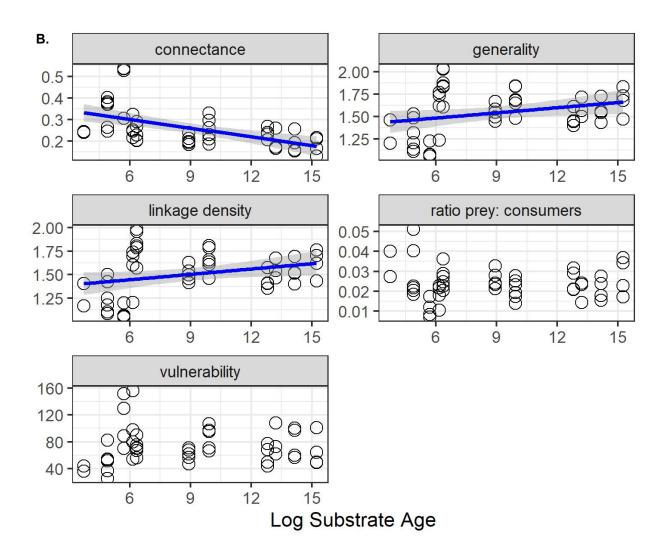
SI Figure 5. The effect of network size on ecological network metrics. Each quantitative and qualitative metrics was regressed against the number of nodes (representing network size). A) Quantitative metrics were not significantly correlated with network size with the exception of generality. B) Qualitative metrics were significantly correlated with network size, with the exception of the ratio of resource:consumers. Spearman rho and p-values in SI Table 4.





SI Figure 6. Null model analysis for weighted networks. A. Graphs of all 1000 null matrices metric values (gray) and observed values (color) by community age. B. Z-scores from null model analysis of weighted metrics are plotted by community age. There are no null values or z-score for interaction evenness because the null model we used wasn't appropriate given that interaction evenness is based on the empirical distribution of weights, regardless of which species those weights are between.





SI Figure 7. Plot level. A. Quantitative and B. Qualitative metrics shown for matrices of arthropod-plant interactions at each plot within a site. In the main text, one network was created per site because our confidence in the network statistics increases with the size of the networks. This was a particular issue for some plots at the youngest and most depauperate sites, where < 4 plant species were sampled within the plot radius and networks would be small. The results from plot level analysis are consistent with the site level data. The variance among plots at the same sites is minimal. These results help corroborate the trend of increasing specialization over time.

SI Table 1. Site information. Information for the sampling plots within each age community. Six plots were sampled in fourteen sites across the island. Plots with greater or less than 1 standard deviation of sample beating time were dropped from analysis. This left a total of 11 sites and 50 plots (highlighted).

Island	Site	Plot	Elev m	AAP m	Lat	Long	Flow age min	Flow age max	Flow age mean
Hawaii	flow1973	flow1973_079	975	2661	19.37	-155.21	42	42	42
Hawaii	flow1973	flow1973_109	975	2613	19.37	-155.22	42	42	42
Hawaii	flow1973	flow1973_084	975	2756	19.37	-155.21	42	42	42
Hawaii	flow1973	flow1973_081	975	2661	19.37	-155.22	42	42	42
Hawaii	flow1973	flow1973_107	975	2570	19.37	-155.22	42	42	42
Hawaii	flow1973	flow1973_110	975	2657	19.37	-155.21	42	42	42
Hawaii	treePlanting	treePlanting_01	1209	4515	19.66	-155.28	133	133	133
Hawaii	treePlanting	treePlanting_03	1204	4495	19.66	-155.28	133	133	133
Hawaii	treePlanting	treePlanting_04	1196	4719	19.66	-155.28	133	133	133
Hawaii	treePlanting	treePlanting_05	1198	4700	19.66	-155.28	133	133	133
Hawaii	treePlanting	treePlanting_02	1198	4515	19.66	-155.28	133	133	133
Hawaii	treePlanting	treePlanting_06	1192	4700	19.66	-155.28	133	133	133
Hawaii	escape	escape_02	997	2658	19.38	-155.22	200	400	300
Hawaii	escape	escape_04	1012	2565	19.37	-155.22	200	400	300
Hawaii	escape	escape_06	1007	2610	19.37	-155.22	200	400	300
Hawaii	escape	escape_87	1004	2748	19.38	-155.22	200	400	300
Hawaii	escape	escape_90	1013	2647	19.38	-155.22	200	400	300
Hawaii	escape	escape_89	1003	2748	19.38	-155.21	200	400	300
Hawaii	kaiholenaYng	kaiholenaYng_02	966	2888	19.18	-155.58	200	750	475
Hawaii	kaiholenaYng	kaiholenaYng_08	1003	2947	19.19	-155.59	200	750	475
Hawaii	kaiholenaYng	kaiholenaYng_09	1045	2937	19.19	-155.59	200	750	475
Hawaii	kaiholenaYng	kaiholenaYng_04	1012	2909	19.19	-155.59	200	750	475
Hawaii	kaiholenaYng	kaiholenaYng_10	1029	2932	19.19	-155.59	200	750	475
Hawaii	kaiholenaYng	kaiholenaYng_11	1045	2926	19.19	-155.59	200	750	475
Hawaii	thurston	thurston_01	1177	2610	19.41	-155.24	400	750	575
Hawaii	thurston	thurston_04	1181	2880	19.41	-155.24	400	750	575
Hawaii	thurston	thurston_05	1189	2838	19.41	-155.24	400	750	575
Hawaii	thurston	thurston_06	1195	2709	19.42	-155.24	400	750	575
Hawaii	thurston	thurston_08	1195	2797	19.42	-155.24	400	750	575
Hawaii	thurston	thurston_12	1189	2850	19.42	-155.24	400	750	575
Hawaii	olaa	olaa_01	1170	3011	19.45	-155.25	5000	10000	7500
Hawaii	olaa	olaa_03	1169	3071	19.45	-155.25	5000	10000	7500
Hawaii	olaa	olaa_06	1177	2957	19.45	-155.25	5000	10000	7500
Hawaii	olaa	olaa_12	1183	2908	19.46	-155.25	5000	10000	7500

Hawaii	olaa	olaa_02	1163	3022	19.45	-155.25	5000	10000	7500
Hawaii	olaa	olaa_10	1183	2915	19.45	-155.25	5000	10000	7500
Hawaii	hippnet	hippnet_05	1131	3538	19.93	-155.28	4000	14000	9000
Hawaii	hippnet	hippnet_06	1152	3506	19.93	-155.29	4000	14000	9000
Hawaii	hippnet	hippnet_08	1126	3654	19.93	-155.28	4000	14000	9000
Hawaii	hippnet	hippnet_09	1212	3318	19.93	-155.29	4000	14000	9000
Hawaii	hippnet	hippnet_11	1231	3295	19.93	-155.29	4000	14000	9000
Hawaii	hippnet	hippnet_13	1177	3479	19.93	-155.29	4000	14000	9000
Hawaii	alili	alili_01	917	2460	19.23	-155.51	11000	30000	20500
Hawaii	alili	alili_04	960	2573	19.23	-155.52	11000	30000	20500
Hawaii	alili	alili_05	917	2504	19.23	-155.52	11000	30000	20500
Hawaii	alili	alili_06	942	2541	19.23	-155.51	11000	30000	20500
Hawaii	alili	alili_14	969	2597	19.23	-155.52	11000	30000	20500
Hawaii	alili	alili_15	985	2583	19.23	-155.52	11000	30000	20500
Hawaii	laupLSAG	laupLSAG_01	1102	3433	19.94	-155.3	14000	65000	39500
Hawaii	laupLSAG	laupLSAG_04	1158	3235	19.94	-155.3	14000	65000	39500
Hawaii	laupLSAG	laupLSAG_07	1155	3235	19.93	-155.3	14000	65000	39500
Hawaii	laupLSAG	laupLSAG_10	1181	3291	19.93	-155.3	14000	65000	39500
Hawaii	laupLSAG	laupLSAG_12	1251	3080	19.93	-155.3	14000	65000	39500
Hawaii	laupLSAG	laupLSAG_15	1242	3112	19.93	-155.3	14000	65000	39500
Hawaii	kohalaYng	kohalaYng_03	1285	2779	20.11	-155.74	120000	230000	175000
Hawaii	kohalaYng	kohalaYng_04	1303	2863	20.11	-155.74	120000	230000	175000
Hawaii	kohalaYng	kohalaYng_07	1274	3004	20.11	-155.74	120000	230000	175000
Hawaii	kohalaYng	kohalaYng_08	1266	3177	20.11	-155.73	120000	230000	175000
Hawaii	kohalaYng	kohalaYng_10	1305	2779	20.11	-155.74	120000	230000	175000
Hawaii	kohalaYng	kohalaYng_20	1281	2918	20.11	-155.74	120000	230000	175000
Hawaii	kohalaOld	kohalaOld_01	1294	3568	20.11	-155.72	230000	500000	365000
Hawaii	kohalaOld	kohalaOld_03	1273	3704	20.11	-155.72	230000	500000	365000
Hawaii	kohalaOld	kohalaOld_08	1313	3402	20.11	-155.73	230000	500000	365000
Hawaii	kohalaOld	kohalaOld_05	1318	3501	20.11	-155.72	230000	500000	365000
Hawaii	kohalaOld	kohalaOld_06	1309	3501	20.11	-155.72	230000	500000	365000
Hawaii	kohalaOld	kohalaOld_07	1335	3419	20.1	-155.72	230000	500000	365000
Maui	waikamoi	waikamoi_07	1591	2518	20.8	-156.25	140000	950000	545000
Maui	waikamoi	waikamoi_16	1545	3040	20.8	-156.24	140000	950000	545000
Maui	waikamoi	waikamoi_36	1564	2765	20.8	-156.25	140000	950000	545000
Maui	waikamoi	waikamoi_06	1547	2834	20.8	-156.25	140000	950000	545000
Maui	waikamoi	waikamoi_18	1552	3107	20.8	-156.24	140000	950000	545000
Maui	waikamoi	waikamoi_32	1524	2598	20.8	-156.25	140000	950000	545000
Molokai	kamakou	kamakou_07	1191	2718	21.11	-156.9	1300000	1500000	1400000
Molokai	kamakou	kamakou_09	1223	3007	21.11	-156.9	1300000	1500000	1400000
Molokai	kamakou	kamakou_17	1213	3007	21.11	-156.89	1300000	1500000	1400000
Molokai	kamakou	kamakou_39	1182	2800	21.11	-156.89	1300000	1500000	1400000

Molokai	kamakou	kamakou_15	1245	3276	21.11	-156.89	1300000	1500000	1400000
Molokai	kamakou	kamakou_41	1159	2532	21.11	-156.9	1300000	1500000	1400000
Kauai	kokee	kokee_16	1116	2251	22.14	-159.62	4000000	4300000	4150000
Kauai	kokee	kokee_21	1200	2860	22.15	-159.62	4000000	4300000	4150000
Kauai	kokee	kokee_23	1210	2877	22.15	-159.61	4000000	4300000	4150000
Kauai	kokee	kokee_11	1147	2487	22.14	-159.62	4000000	4300000	4150000
Kauai	kokee	kokee_09	1172	2413	22.14	-159.62	4000000	4300000	4150000
Kauai	kokee	kokee_22	1202	2724	22.15	-159.62	4000000	4300000	4150000

SI Table 2. Summary statistics from lidar data during site selection. Here we show the basic forest structure characteristics for each of the sites that was characterized as a possible area for arthropod community assessment, The highlighted sites were included for arthropod community assessment using DNA metabarcoding. The sites with NA we did not have lidar data availability.

Site	MEAN	MEDIAN	MIN	MAX	SD	Q40	Kurtosis	KurtSE	Skewness	SkewSE
HAVO_EscapeRd_1973	0.530168	0.563037	0.0099	1.364975	0.274771	0.631655	-0.61762	0.271608	0.044372	0.137784
Tree Planting Road	NA									
HAVO_EscapeRd_HighStature	7.21131	7.457712	1.78265	11.91813	1.905806	7.93115	-0.25263	0.14969	-0.42313	0.075164
TNC_Kaiholena_Young	6.254615	5.85335	3.378	13.40897	1.664489	6.35712	1.333989	0.154296	1.105489	0.077498
TNC_Kaiholena_Old	5.928797	5.7883	3.774975	11.6454	1.070454	5.99883	3.697232	0.180073	1.37507	0.090596
HAVO_Thurston	12.37064	12.76153	4.408525	16.77472	2.261814	13.35568	-0.1841	0.141771	-0.61303	0.071156
HAVO_Olaa_Old	7.830925	7.329275	4.076075	18.0546	2.272184	7.9105	0.922662	0.093658	1.041491	0.046906
NAR_Laupahoehoe_HIPPNET	14.05824	13.8389	0.030125	25.2793	3.653928	14.90676	-0.47155	0.077027	0.153058	0.038557
KauFR_Alili	11.86238	11.4214	2.585675	21.81295	3.492847	12.57993	-0.73541	0.118676	0.311218	0.059496
HiloFR_Laupahoehoe_Old	13.94185	13.91757	4.52675	25.16357	3.045798	14.75297	-0.37097	0.096224	0.125453	0.048196
NAR_Laupahoehoe_65-250K	12.23722	13.00572	0.1083	23.588	4.243211	13.84491	-0.04895	0.061824	-0.66387	0.030934
HiloFR_Humuula	15.33543	15.18775	2.23905	27.6691	3.801326	16.22402	-0.40367	0.056136	0.17102	0.028085
NAR_PuuMakaala_Young	5.951452	5.722412	0.14485	14.82202	2.678494	6.496805	-0.37364	0.091493	0.30729	0.045819
NAR_PuuMakaala_Old	14.71206	15.72586	0	25.28247	5.365967	16.84449	1.253179	0.123676	-1.16721	0.062017
NAR_PuuOumi_Young02	5.951452	5.722412	0.14485	14.82202	2.678494	6.496805	-0.37364	0.091493	0.30729	0.045819
NAR_PuuOumi_Old	5.999913	6.243275	0.3082	10.97857	1.918523	6.687105	-0.25813	0.10721	-0.3717	0.053721
Kamakou Molokai	NA									
Waikamoi Maui	NA									
Kauai_8m	5.248667	5.121489	0.010311	19.3347	3.227693	6.003569	-0.26206	0.036622	0.38	0.018316

SI Table 3. Taxonomic coverage. Taxonomic coverage of arthropod OTUs as determined by percent match to GenBank accessions and a 57 species reference collection. Taxonomy of plants as determined by morphological identification in the field. Plants that were unidentified were grouped by morphotaxa. The bipartite web id is the 5-letter code to match the bipartite plots in Figure 3 and SI Figure 3.

arthropod molecular taxonomyAmphipoda2911AmphiAraneae5395898AraneBlattodea1221BlattColeoptera42118412ColeoDiptera45427819DipteEntomobryomorpha295831EntomGeophilomorpha9GeophGeophHemiptera77437119HemipHymenoptera981235HymenIsopoda59221IsopoJulida431JulidLepidoptera290181615LepidLithobiomorpha91MesosNeuroptera0rthoptera197422OrthopPoduromorpha711PolydPolydesnida11PolydPolycenida11PolydPolycenida11PolydPolycenida11PolyxPseudoscorpiones41PsecoSarcooptiformes26212SarcoSpirobolida11Spiro	Arthropod Order or Plant	- 4		~ ~ ~ ~ ~ ~ ~	families	bipartite web id
Amphipoda 29 11AmphiAraneae 539 58 9 8 AraneBlattodea 12 2 1BlattColeoptera 421 18 4 12 ColeoDiptera 454 27 8 19 DipteEntomobryomorpha 295 8 3 1EntomGeophilomorpha 9 GeophGeophHemiptera 774 37 11 9 HemipHymenoptera 98 12 3 5 HymenIsopoda 59 2 2 1 IsopoJulida 4 3 1 JulidLepidoptera 290 18 16 15 LepidLithobiomorpha 9 1 MesosNeuroptera 30 2 1 2 NeuroOrthoptera 197 4 2 2 OrthoPolydPolydPolydPolydPolydesmida 1 1 PolydPolydPolydPolydPolydPolydPolyxPseudoscorpiones 4 1 2 SarcoSpiroSarcoSpiro	Genus/Morphotaxon	otus	species	genus	lamines	web id
Araneae5395898AraneBlattodea1221BlattColeoptera42118412ColeoDiptera45427819DipteEntomobryomorpha295831EntomGeophilomorpha9GeophGeophHemiptera77437119HemipHymenoptera981235HymenIsopoda59221IsopoJulida431JulidLepidoptera290181615LepidLithobiomorpha91MesosNeuroptera30212NeuroOrthoptera197422OrthoPoduromorpha711PodyrPolydesmida11PolydPolyxenida11PolyxPseudoscorpiones41PsocoSarcoptiformes26641PsocoSarcoptiformes26212SarcoSpirobolida11Spiro	arthropod molecular taxonomy					
Blattodea1221BlattColeoptera42118412ColeoDiptera45427819DipteEntomobryomorpha295831EntomGeophilomorpha9GeophGeophHemiptera77437119HemipHymenoptera981235HymenIsopoda59221IsopoJulida431JulidLepidoptera290181615LepidLithobiomorpha1KesosNeuroptera30212NeuroOrthoptera197422OrthoPolydPolydesmida11PolydPolyxPseudoscorpiones41PolyxPseudoscorpiones411PolyxPseudoscorpiones26641PsocoSarcoptiformes26212SarcoSpiroSpiroSpiro	Amphipoda	29		1	1	Amphi
Coleoptera42118412ColeoDiptera45427819DipteEntomobryomorpha295831EntomGeophilomorpha9GeophHemiptera77437119HemipHymenoptera981235HymenIsopoda59221IsopoJulida431JulidLepidoptera290181615LepidLithobiomorpha1LithoMesosNeuroptera30212NeuroOrthoptera197422OrthoPoduromorpha711PodurPolydesmida11PolydPolyxenida11PolydPoscoptera26641PsocoSarcoptiformes26212SarcoSpirobolida11Spiro	Araneae	539	58	9	8	Arane
Diptera 454 27 8 19 DipteEntomobryomorpha 295 8 3 1 EntomGeophilomorpha 9 GeophHemiptera 774 37 11 9 HemipHymenoptera 98 12 3 5 HymenIsopoda 59 2 2 1 IsopoJulida 4 3 1 JulidLepidoptera 290 18 16 15 LepidLithobiomorpha 1 LithoMesosNeuroptera 30 2 1 2 NeuroOrthoptera 197 4 2 2 OrthoPoduromorpha 7 1 1 PodurPolydesmida 1 1 PolydPolyxenida 1 1 PolydPolyxenida 266 4 1 PsecoSarcoptiformes 262 1 2 SarcoSpirobolida 1 1 Spiro	Blattodea	12	2	1		Blatt
L295831EntomGeophilomorpha9GeophHemiptera77437119HemipHymenoptera981235HymenIsopoda59221IsopoJulida431JulidLepidoptera290181615LepidLithobiomorpha1LithoMesosNeuroptera30212NeuroOrthoptera197422OrthoPolydPolydPolydPolydPolydPolydPolyxPseudoscorpiones411PolyxPseudPseudSarcoSarcoSpirobolida11Spiro	Coleoptera	421	18	4	12	Coleo
Geophilomorpha9GeophHemiptera 774 37 11 9HemipHymenoptera98 12 3 5HymenIsopoda 59 2 2 1 IsopoJulida4 3 1 JulidLepidoptera 290 18 16 15 LepidLithobiomorpha1LithoLithoMesostigmata9 1 MesosNeuroptera 30 2 1 2 Orthoptera 197 4 2 2 Orthoptera 197 4 2 2 Polydesmida 1 1 PolydPolycenida 1 1 PolyxPseudoscorpiones 4 1 PseudPsocoptera 266 4 1 PsocoSarcoptiformes 262 1 2 SarcoSpirobolida 1 1 Spiro	Diptera	454	27	8	19	Dipte
Hemiptera 774 37 11 9 HemipHymenoptera 98 12 3 5 HymenIsopoda 59 2 2 1 IsopoJulida 4 3 1 JulidLepidoptera 290 18 16 15 LepidLithobiomorpha 1 LithoLithoMesostigmata 9 1 MesosNeuroptera 30 2 1 2 Orthoptera 197 4 2 2 Orthoptera 197 4 2 2 Polydesmida 1 1 PolydPolyxenida 1 1 PolyxPseudoscorpiones 4 7 PseudPsocoptera 266 4 1 PsocoSarcoptiformes 262 1 2 SarcoSpirobolida 1 1 Spiro	Entomobryomorpha	295	8	3	1	Entom
Hymenoptera981235HymenIsopoda59221IsopoJulida431JulidLepidoptera290181615LepidLithobiomorpha1LithoLithoMesostigmata91MesosNeuroptera30212NeuroOrthoptera197422OrthoPoduromorpha711PodurPolydesmida11PolydPolyxenida11PolydPseudoscorpiones4PseudPsocoptera26641PsocoSarcoptiformes26212SarcoSpirobolida11SpiroSpiro	Geophilomorpha	9				Geoph
Isopoda59221IsopoJulida431JulidLepidoptera290181615LepidLithobiomorpha1LithoLithoMesostigmata91MesosNeuroptera30212Orthoptera197422Orthoptera197422Polydesmida11PolydPolydesmida11PolydPseudoscorpiones4PseudPsocoptera26641Psocoptiones26212Spirobolida11Spiro	Hemiptera	774	37	11	9	Hemip
Julida431JulidLepidoptera290181615LepidLithobiomorpha1LithoLithoMesostigmata91MesosNeuroptera30212NeuroOrthoptera197422OrthoPoduromorpha711PodurPolydesmida11PolydPolyxenida11PolyxPseudoscorpiones4PseudPsocoptera26641Spirobolida12SarcoSpirobolida15piro	Hymenoptera	98	12	3	5	Hymen
Lepidoptera290181615LepidLithobiomorpha1LithoMesostigmata91MesosNeuroptera30212NeuroOrthoptera197422OrthoPoduromorpha711PodurPolydesmida11PolydPolyxenida11PolydPseudoscorpiones4PseudPsocoptera26641Spirobolida12SarcoSpirobolida11Spiro	Isopoda	59	2	2	1	Isopo
Lithobiomorpha1LithoMesostigmata91MesosNeuroptera30212NeuroOrthoptera197422OrthoPoduromorpha711PodurPolydesmida11PolydPolyxenida11PolydPseudoscorpiones4PseudPsocoptera26641Sarcoptiformes26212Spirobolida1Spiro	Julida	4	3		1	Julid
Mesostigmata91MesosNeuroptera30212NeuroOrthoptera197422OrthoPoduromorpha711PodurPolydesmida11PolydPolyxenida11PolydPseudoscorpiones4PseudPsocoptera26641Sarcoptiformes26212SarcoSpirobolida11Spiro	Lepidoptera	290	18	16	15	Lepid
Neuroptera30212NeuroOrthoptera197422OrthoPoduromorpha711PodurPolydesmida11PolydPolyxenida11PolydPseudoscorpiones4PseudPsocoptera26641Sarcoptiformes26212Spirobolida1Spiro	Lithobiomorpha	1				Litho
Orthoptera197422OrthoPoduromorpha711PodurPolydesmida111PolydPolyxenida11PolyxPseudoscorpiones4PseudPsocoptera26641Sarcoptiformes26212Spirobolida1Spiro	Mesostigmata	9	1			Mesos
Poduromorpha711PodurPolydesmida111PolydPolyxenida11PolyxPseudoscorpiones4PseudPsocoptera26641Sarcoptiformes26212Spirobolida1Spiro	Neuroptera	30	2	1	2	Neuro
Polydesmida11PolydPolyxenida11PolyxPseudoscorpiones4PseudPsocoptera26641Sarcoptiformes26212Spirobolida1Spiro	Orthoptera	197	4	2	2	Ortho
Polyxenida1PolyxPseudoscorpiones4PseudPsocoptera26641Sarcoptiformes26212Spirobolida1Spiro	Poduromorpha	7		1	1	Podur
Pseudoscorpiones4PseudPsocoptera26641PsocoSarcoptiformes26212SarcoSpirobolida1Spiro	Polydesmida	1	1			Polyd
Psocoptera26641PsocoSarcoptiformes26212SarcoSpirobolida1Spiro	Polyxenida	1				Polyx
Sarcoptiformes26212SarcoSpirobolida1Spiro	Pseudoscorpiones	4				Pseud
Spirobolida 1 Spiro	Psocoptera	266	4	1		Psoco
• •	Sarcoptiformes	262	1	2		Sarco
	Spirobolida	1				Spiro
Spirostreptida 4 Spiro	Spirostreptida	4				Spiro
Symphypleona 4 1 1 Symph	Symphypleona	4	1	1		Symph
Trombidiformes 14 1 1 Tromb	Trombidiformes	14	1		1	Tromb

plant morphological taxonomy

Metrosideros	Metro
Styphelia	Styph
Vaccinium	Vacci

Swordfern	Sword
Coprosma	Copro
Dubautia	Dubau
Dicranopteris	Dicra
Groundfern	Groun
Myrsine	Myrsi
Pipturus	Piptu
Ilex	Ilex
Broussaisia	Brous
BrownCibotium	Brown
GreenCibotium	Green
Sadleria	Sadle
Cheirodendron	Cheir
Psychotria	Psych
Astelia	Astel
Machaerina	Macha
Melicope	Melic
Perrottetia	Perro
Cyrtandra	Cyrta
Dryopteris	Dryop
Freycinetia	Freyc
Hedyotis	Hedyo
Alyxia	Alyxi
Rubus	Rubus
Clermontia	Clerm
Carex	Carex
Kadua	Kadua
Labordia	Labor
Curlleaf	Curll
whitemid	white
blackvelvet	black

SI Table 4. Spearman's correlation tests. Spearman's correlation tests were used to determine significance for the: A) regression against log substrate age (community age) of qualitative network metric values calculated from binary matrices at each site. B) regression against number of nodes (community size) of quantitative network metric values calculated from weighted matrices at each site C) regression against number of nodes (community size) of qualitative network metric values calculated from binary matrices at each site and D) regression of size of network (number of nodes, number of links) by log flow age (community age).

	G	p-	Spearman's
	S	value	rho
<i>Qualitative (binary) x community age</i>			
linkage density	78	0.037	0.65
connectance	404	0.003	-0.84
generality	78	0.037	0.65
vulnerability	188	0.670	0.15
ratio prey:consumers	132	0.225	0.40
Quantitative (weighted) x number of n	odes		
linkage density	140	0.273	0.36
weighted connectance	226	0.946	-0.03
generality	68	0.023	0.69
vulnerability	144	0.300	0.35
interaction evenness	134	0.237	0.39
index of specialization H ₂ '	218	0.989	0.01
Qualitative (binary) x number of			
nodes			
linkage density	34	0.002	0.85
connectance	396	0.005	-0.80
generality	34	0.002	0.61
vulnerability	86	0.051	0.61
ratio prey:consumers	202	0.818	0.08
number of nodes x log flow age	3741	0.000	0.74
number of links x log flow age	3096	0.000	0.78

SI Table 5. p-value table weighted metrics null model analysis. The network metrics of 1000 null matrices were compared to the network metrics of the empirical matrix. The null hypothesis is said to be true if the empirical values are within the null model distribution. This is a two tailed distribution. A "1" indicates that every null model value that we compared to our observed was greater than or equal to the observed value. A "0" indicates that no null model value that we compared to our observed was greater than or equal to the observed value. A value of greater than 0 and less than 1 is simply the number values that the null metric was greater than the empirical metric.

site	pval.Connectance	pval.LD	pval.Gen	pval.Vul	pval.H2
flow1973	0.001	0.001	0	0.006	1
treePlanting	0.518	0.518	0	0.553	1
escape	0.283	0.283	0.002	0.293	1
kaiholenaYng	0.571	0.571	0	0.96	1
thurston	0.031	0.031	0	0.225	1
olaa	0.492	0.492	0	0.645	1
alili	0.596	0.596	0	0.749	1
kohalaOld	0.653	0.653	0	0.825	1
waikamoi	0	0	0	0	1
kamakou	0.889	0.889	0	0.973	1
kokee	0	0	0	0	1