UCSF UC San Francisco Previously Published Works

Title

Risk stratification of smoldering multiple myeloma: predictive value of free light chains and group-based trajectory modeling.

Permalink

https://escholarship.org/uc/item/40n403x5

Journal Blood Advances, 2(12)

Authors

Wu, Vernon Moshier, Erin Leng, Siyang <u>et al.</u>

Publication Date

2018-06-26

DOI

10.1182/bloodadvances.2018016998

Peer reviewed

Risk stratification of smoldering multiple myeloma: predictive value of free light chains and group-based trajectory modeling

Vernon Wu,¹ Erin Moshier,² Siyang Leng,³ Bart Barlogie,² Hearn Jay Cho,² Sundar Jagannath,² Deepu Madduri,² Madhu Mazumdar,² Samir Parekh,² and Ajai Chari²

¹Department of Medicine, Stony Brook University School of Medicine, Stony Brook, NY; ²Tisch Cancer Institute, Icahn School of Medicine at Mount Sinai, New York, NY; and ³Division of Hematology/Oncology, Columbia University Medical Center, New York, NY

Key Points

- FLCr ≥100 and BMPC ≥60% identify high-risk SMM, although with more modest median TTP and 2-year PD than previously published.
- Baseline immunoparesis, eMP, eHb, and edFLC can help identify an ultra-high-risk SMM cohort.

We investigated the predictive role for serum free light chain ratio (FLCr) \geq 100, bone marrow plasma cell (BMPC) \geq 60%, and evolving biomarkers through group-based trajectory modeling (GBTM) as high-risk defining events in 273 smoldering multiple myeloma (SMM) patients with a median follow-up of 74 months. FLCr \geq 100 was confirmed as a marker for high-risk progression with a median time to progression (TTP) of 40 months with a 44% risk of progression of disease (PD) at 2 years; however, 44% of FLCr \geq 100 also did not progress during follow-up. For patients with BMPC \geq 60% by core biopsy, the median TTP was 31 months with a 2-year PD of 41%. GBTM established high-risk trajectories for evolving hemoglobin (eHb; characterized as a 1.57 g/dL decrease in hemoglobin), evolving m-protein (eMP; 64% increase in m-protein), and evolving differences in FLC (edFLC; 169% increase in dFLC) within 1 year of diagnosis associated with a decreased median TTP and an increased 2 year rate of PD. Of all the variables examined, we identify a model where immunoparesis, eHb, eMP, and edFLC were significant predictors for ultra-high-risk progression with a median TTP of only 13 months with 3 or more variables present. Our results not only confirm a more modest 2 year PD associated with FLCr \geq 100 and BMPC \geq 60 but also suggest that eHb, eMP, and edFLC may help identify an ultra-high-risk SMM group.

Introduction

Smoldering multiple myeloma (SMM) is associated with 10% rate of progression to active MM annually for the first 5 years and 3% annually for the next 3 years compared with 1% annually for monoclonal gammopathy of undetermined significance.¹ In 2014, the International Myeloma Working Group (IMWG) updated its criteria for diagnosing MM from end-organ damage to include bone marrow plasma cell (BMPC) \geq 60%, serum involved: uninvolved serum free light chain ratio (FLCr) \geq 100, and magnetic resonance imaging/positron emission tomography-computed tomography/computed tomography (MRI/PET-CT/CT) with >1 focal lesion of 5 mm or greater. The goal of these updates was to identify a group of SMM patients at high risk of developing end-organ damage and to consider early therapy for these patients.^{2,3} The consensus was that biomarkers associated with 80% probability of progression to end-organ damage within 2 years or a median time to progression (TTP) of 12 months should be regarded as myeloma defining events and thus patients should be offered therapy.

Although 2 retrospective studies supported these updates, with median time to progression ranging from 13 to 15 months and 2-year progression rates as high as 98% for SMM patients with FLCr $\geq 100^{4,5}$ another study found a rate of only 53%.⁶ Most important, the only prospective registry published found a rate of progression of only 30%.⁷ Also, although several studies have found BMPC $\geq 60\%$ to be predictive of progression, each had very few such patients, with a cumulative sample size of

Submitted 5 February 2018; accepted 18 May 2018. DOI 10.1182/ bloodadvances.2018016998.

only 45 patients across all published studies.^{5,8-10} Interestingly, 1 study compared BMPC determined by core biopsy (BMB) vs aspirate (BMA) and found that BMB \geq 60% was not a predictor of rapid progression and only BMA \geq 60% by aspirate was high risk.¹⁰

The importance of the kinetics of SMM evolution have been previously described for evolving changes in m-protein and hemoglobin within the first 12 months of SMM diagnoses with identification of an ultra-high-risk group of patients at >80% risk of progression at 2 years.¹¹ More recently, another study also examined evolving changes in *m*-spike and found increases in 10% of m-protein by 12 months (if baseline m-protein \geq 3 g/dL) or by 3 years (if baseline m-protein < 3 g/dL) was associated with 71% risk of progression by 3 years independent of other standard risk factors of progression (m-protein \geq 3 g/dL, BMPC \geq 20%, or presence of immunoparesis).¹² Early intervention in SMM patients requires not only evidence that early treatment can improve these patients' long-term outcomes, but at minimum, reproducible criteria for the definition of high-risk SMM. The discrepancies in the predictive value of FLCr as well as the identification of evolving risk factors warrant further investigation. In this study, we survey characteristics and outcomes of SMM patients seen at our institution; we used a group-based trajectory modeling (GBTM) to compare evolving biomarkers, including the never previously investigated evolving FLC and evolving dFLC, as risk factors to identify a group of ultra-high-risk SMM.

Methods

Approval of this retrospective study was obtained from the institutional review board of the Mount Sinai School of Medicine, and the study was done in accordance with the Declaration of Helsinki. Patients were identified by query of the electronic medical record for consecutive patients with the diagnosis of myeloma seen at Mount Sinai Hospital between 2010 and 2015. Eligibility criteria included a diagnosis of SMM as defined by the 2003 IMWG definition: ≥10% BMPC or serum m-protein \geq 3 and the absence of hypercalcemia (Ca \geq 11.5 mg/dL), renal insufficiency (Cr \geq 2 mg/dL or creatinine clearance <40 mL/minute), anemia (hemoglobin [Hb] \leq 10 g/dL), and bone lesions (lytic lesions or diffuse osteopenia). Exclusion criteria were the absence of BMPC within 3 months of the time of SMM diagnosis and <1 year of follow-up. For analysis of evolving biomarkers, patients were required to have 2 or more laboratory tests within 12 months of SMM diagnosis in addition to their baseline value. Patients who participated in clinical trials involving agents known to have no single-agent activity were included in the analysis but also analyzed separately.

FLCs were measured using Freelite assay. FLCr was computed as a ratio of involved:uninvolved light chain. dFLC was computed as the difference between the involved and uninvolved light chain. BMPC is reported as the highest BMPC as determined by BMB or BMA unless otherwise stated. Immunoparesis is defined as reduction below the lower limit of normal in 2 uninvolved immunoglobulins (ie, IgG <700 mg/dL, IgA <70 mg/dL, Ig M <40 mg/dL). Progression to active disease was determined clinically by calcium elevation, renal dysfunction, anemia, and bone disease–related organ impairment, infection, or amyloidosis. Patients were censored if they were started on MM therapy in the absence of these events. Risk stratification for progression was performed according to the previously published Mayo baseline or evolving model.^{11,13}

Statistical analyses were performed using SAS, version 9.4 (SAS Institute Inc., Cary, NC). A "3-step approach" was adopted to estimate the relationship between latent class trajectories (described here) and TTP. All hypothesis testing was 2-sided with the type 1 error rate fixed at 5% for determination of factors associated with TTP.

GBTM was performed using an SAS macro named PROC TRAJ. This method identified distinct clusters of patients with similar patterns of evolving biomarkers within the first 12 months of SMM diagnosis: evolving Hb (eHb), evolving m-protein (eMP), evolving FLCr (eFLCr), and evolving differences in FLC (edFLC). Trajectory parameters were derived by latent class analysis using maximum likelihood estimation. In particular, the distinctive trajectories of biomarkers were derived by modeling the change (for Hb) and percent change (for m-protein and FLC), in each biomarker, from baseline, as a function of time since SMM diagnosis. Quadratic curves were used to model trends in Hb and FLC over time and cubic curves were used to model trends in m-protein over time. The optimal number of trajectories was determined using previously suggested guidelines¹⁴ and clinical relevance determined by research team. The output of PROC TRAJ included the equations for the different trajectories along with the assignment of each patient to 1 of the trajectory groups.

Once trajectory groups were estimated for each biomarker, the method of Kaplan-Meier was used to estimate median TTP for each group with high-risk patients defined as those with the shortest TTP. Time-dependent receiver operator characteristic (ROC) curves for right censored survival data were estimated using the inverse probability of censoring weighting method previously described.¹⁵ Sensitivities, specificities, and accuracies from the 2-year ROC curves for progression of disease (PD) were calculated for high-/low-risk baseline (at SMM diagnosis) and evolving biomarker groups. Accuracy was defined as the proportion of true positives and true negatives in all evaluated patients. Areas under the 2-year ROC curves (AUCs) as well as integrated time-dependent AUCs (IAUC - average of all time-dependent AUC), were provided as metrics of model discrimination. Multivariable Cox proportional hazards regression was used to identify prognostic risk factors for TTP. A complete case analysis was performed, with patients missing at least 1 variable excluded from the analysis. A covariate selection algorithm was used to identify the "best" subset of predictors in multivariable Cox modeling.¹⁶

Results

Baseline characteristics

A total of 273 patients with SMM met eligibility criteria; of these, 183 patients had FLCr within 3 months of diagnosis. Only 21 patients had to be censored at the time they began treatment of non-calcium elevation, renal dysfunction, anemia, and bone disease indications including infection (1), amyloid (2), rapid increase in paraprotein (4), BMPC \geq 60 or FLCr \geq 100 (7), other (4), and unknown (3). Of note, median (range) time to treatment from SMM diagnosis in these patients was 24.1 (6.3-74.6) months; median (range) time to treatment for BMPC and FLCr was 26.1 (7.4-72.3) months.

Patient characteristics are shown stratified by FLCr and BMPC (Table 1). The median age of the 273 patients at time of diagnosis was 60 years, with 52% males. The median BMPC at time of diagnosis was 20% with 92% of patients with BMPC <60%.

Table 1. Patient characteristics at SMM	diagnosis stratified by FLCr ≥100 or	<100 and BMPC ≥60 or <60
---	--------------------------------------	--------------------------

Variables	All patients	FLCr <100	FLCr >100	Р	BMPC <60	BMPC >60	Р
Patients, n	273	156	27	—	251	22	_
Median age (range), y	60 (20-90)	60 (25-82)	62 (33-75)	.93	60 (20-90)	60 (36-77)	.82
Male sex	143/273 (52)	80/156 (51)	16/27 (59)	.77	130/251 (52)	13/22 (59)	.51
ВМРС							
Median (range), %	20 (10-95)	20 (10-80)	30 (12-90)	.004	20 (10-50)	70 (60-95)	<.0001
Median %, if <60%	20	20	22	.02	—	_	_
10%-60%	251/273 (92)	147/156 (94)	22/27 (81)	.02	—	—	
Isotype							
lgG	196/273 (72)	103/156 (66)	21/27 (78)	.10	181/251 (72)	15/22 (68)	.79
IgA	57/273 (21)	41/156 (26)	3/27 (11)		51/251 (20)	6/22 (27)	
LC only	13/273 (5)	6/156 (4)	3/27 (11)		12/251 (5)	1/22 (5)	
Other (IgM, IgD, biclonal)	7/273 (3)	6/156 (4)	0/27 (0)		7/251 (3)	0/22 (0)	
Involved light chain							
к	192/273 (70)	108/156 (69)	21/27 (78)	.64	178/251 (71)	14/22 (64)	.68
Λ	79/273 (29)	47/156 (30)	6/27 (22)		71/251 (28)	8/22 (36)	
Biclonal	2/273 (1)	1/156 (1)	0/27 (0)		2/251 (1)	0/22 (0)	
Serum monoclonal protein							
Median m-protein (range), g/dL	1.9 (0-5.3)	1.7 (0.5-3)	2.2 (0-4.2)	.23	1.8 (0-5.3)	2.8 (0.4-43)	.003
<3 g/dL	189/220 (86)	124/156 (79)	20/24 (83)	.84	180/204 (8)	9/16 (56)	.004
≥3 g/dL	31/220 (14)	22/156 (21)	4/24 (17)		24/204 (12)	7/16 (44)	
Free light chain							
FLCr <100	156/183 (85)	_	—	—	147/169 (87)	9/14 (64)	.02
FLCr >100	27/183 (15)	—	—		22/169 (13)	5/14 (36)	
Median light chain concentration, mg/L							
Median difference in involved and uninvolved FLC (range)	61 (0-2838)	45 (0-2418)	532 (28-2838)	<.0001	57 (0-2838)	129 (22-765)	.10
Median κ of involved (range)	59 (3-2862)	46 (3-1288)	633 (28-2862)	<.0001	56 (3-2862)	107 (25-528)	.38
Median Λ of involved (range)	105 (2-2500)	80 (2-2499)	462 (117-1500)	.011	85 (2-2500)	154 (36-769)	.23
FISH risk							
Available FISH, n	149	96	18		138	11	
Normal	61/149 (41)	42/96 (43)	4/18 (22)	.33	60/138 (43)	1/11 (9)	.10
Other risk	30/149 (20)	15/96 (16)	4/18 (22)		27/138 (20)	3/11 (27)	
Hyperdiploidy	33/149 (22)	20/96 (21)	4/18 (22)		28/138 (20)	5/11 (46)	
t(4;14), del17p, +1q21†	25/149 (17)	19/96 (20)	6/18 (34)		23/138 (17)	2/11 (18)	
t(4;14)	14 (47)	11 (5)	3 (30)		13 (50)	1 (25)	
del17p	2 (6)	1 (5)	1 (10)		O (O)	2 (50)	
+ 1q21	14 (47)	8 (40)	6 (60)		13 (50)	1 (25)	

Values are n (%) unless otherwise indicated.

*Free light chain values at baseline were only available for 183 patients for free light chain ratio subgroup analysis.

The sum of all high-risk events is greater than the number of patients with high risk as patients may have met multiple high-risk criteria.

The predominant isotype was IgG (72%) and IgA (21%) with κ (70%) and λ (29%) light chain distribution. The median m-protein was 1.9 g/dL (range, 0-5.3). Fluorescence in situ hybridization (FISH) was available for 149 patients, with 17% patients at high risk defined as t(4;14), del 17p, or +1q21.17,18 Initial imaging performed to rule out MM included bone surveys (52%), PET-CT (24%), and MRI (24%) scans. Bisphosphonate use was noted in 22% of patients for osteoporosis.

There were no significant differences in baseline characteristics noted between all patients and the 183 patients with FLCr data available, with the exception of the year of diagnosis. When comparing patients diagnosed before and after 2001, the year of free light chain approval, as expected, more patients (100%) were missing FLC before this date vs 32% after this date.

FLCr was \geq 100 in 27 (15%) patients. When compared with patients with FLCr <100, no significant differences were noted in median m-protein, isotype, or FISH risk. As expected, the FLCr ≥100 group had higher median differences between involved and uninvolved FLC (532 vs 45 mg/L, P < .0001), involved κ

Table 2. Progression-free survival of SMM and symptoms at time of progression stratified by FLCr ≥100 or <100 and BMPC percentage ≥60 or <60

	All patients	FLCR <100	FLCR ≥100	Р	BMPC <60	BMPC ≥60	P
Patients, n	273	156	27		251	22	
Progressed, n (%)	123 (45)	56 (36)	15 (56)		107 (43)	16 (73)	
Progression event, n (%)*							
Hypercalcemia	4 (3)	2 (4)	0 (0)	1.0000	4 (4)	0 (0)	1.0000
Renal insufficiency	11 (9)	2 (4)	3 (20)	.0599	11 (10)	0 (0)	.3560
Anemia	62 (50)	33 (59)	10 (67)	.7677	52 (49)	10 (63)	.2996
Bone disease	64 (52)	25 (45)	8 (53)	.5742	55 (51)	9 (56)	.7173

*Total number of progression events exceeds the number of patients who progressed as patients may have progressed by multiple events.

concentration (633 vs 46 mg/L, P < .0001), and involved λ concentration (462 vs 80 mg/L, P = .011) when compared with the FLCr <100 group. There was also a significant difference in risk stratification by the Mayo baseline risk model with only intermediate- and high-risk patients in FLCr \geq 100 compared with the FLCr <100 group (P < .001). However, there were no differences in risk stratification by the Mayo evolving risk model between the FLCr subgroups (P = .42).

When comparing those with BMPC \geq 60 to <60 the median m-protein was 2.8 vs 1.8, respectively (P = .003). More high-risk patients were noted with BMPC \geq 60 when stratified by the Mayo baseline (P = .001) but not the evolving (P = .13) risk models.

Progression outcomes

With a median follow-up of 67 months, the median TTP for all patients was 74 months, predominantly from bone disease (52%) and anemia (50%) and to a lesser extent renal insufficiency (9%) and hypercalcemia (3%) (Table 2). No significant differences were noted in progression by bone disease vs other causes when comparing patients who had initial bone survey, PET-CT, or MRI scans to rule out active myeloma. However, of the patients who progressed by bone disease, the median TTP was 35 months (initial assessment by skeletal survey), 20 months (initial PET-CT scan), and 22 months (initial MRI scan), although it is important to note that the selection of imaging modality was based on the judgment of the treating physician.

Progression was analyzed along FLCr and BMPC stratifications. Similar to the overall population, the symptoms of progression in the FLCr and BMPC groups were predominately anemia and bone disease (Table 2). There was a trend toward increased renal insufficiency in FLCr \geq 100 (P = .06), although there were very few patients (n = 5) with adequate data for analysis. Patients with baseline Bence Jones proteinuria (BJP) were twice as likely to have FLCr \geq 100 at baseline as patients without BJP (P = .01); however, there was no significantly increased risk of renal progression if baseline BJP was present (P = .57).

FLCr \geq 100 had a specificity of 90% and sensitivity of 28% for predicting progression at 24 months (Table 3). BMPC \geq 60 had a specificity of 94% and sensitivity of 15% for predicting progression at 24 months. Therefore, similar to previously published studies,

FLCr \geq 100 and BMPC \geq 60 did predict for higher rates of MM progression than those without these features. However, the median TTP and 2-year PD rates were lower than reported initially: 40 months/44%, respectively, for FLCr \geq 100 (Figure 1A) and 31 months/41%, respectively, for BMPC \geq 60% (Figure 1B). Also, interestingly, 44% of the patients with FLCr \geq 100 and 27% with BMPC \geq 60% did not progress during the follow-up period. We also investigated baseline dFLC \geq 100 mg/L as a subgroup because FLCr is very sensitive to small changes to the uninvolved free light chain when FLCr \geq 100. dFLC \geq 100 identified a high-risk cohort with a median TTP of 71-month and 2-year PD 36% (Table 3).

Given the previously inconsistent predictive value of BMPC, the predictive values of BMB (n = 254) and BMA (n = 146) were also analyzed. A cutoff of BMB \geq 60 (n = 21) had a median TTP and 2-year PD of 25 months and 43%, respectively. A BMA \geq 60 cutoff appeared to correlate more with PD, with a median TTP and 2-year PD of 22 months and 67%, respectively, although there were only 3 such patients. Both BMB and BMA \geq 60 identified high-risk groups with shorter median TTP and higher 2-year PD than their respective counterparts.

GBTM identified high-risk groups of patients based on 1 year post-SMM diagnosis trajectories of their Hb, m-protein, FLCr, and dFLC (Figure 2). The high-risk eHb group experienced on average a decrease of 1.57 g/dL (95% confidence interval [CI], 1.29-1.84) in Hb at 1 year. High-risk eMP patients experienced an average increase of 64% (95% Cl, 44-84) in m-protein at 1 year. High-risk eFLCr patients experienced an average increase of 188% (95% Cl, 183-193) in FLCr at 1 year. High-risk edFLC patients experienced on average an increase of 169% (95% Cl, 143-195) in dFLC at 1 year. The GBTM-identified high-risk groups had significantly shorter median TTP and higher 2-year PD rates of 26 months/43% (eHb), 40 months/36% (eMP), 37 months/32% (eFLCr), and 45 months/30% (edFLC) relative to their low-risk counterparts (Table 3). Of note, eHb and eMP, as defined by the Mayo evolving model, individually had a median TTP/2-year PD of 42 months/29% and 35 months/33%, respectively, in our SMM population.

Baseline and GBTM evolving characteristics were available for 90 patients and were analyzed for their contribution to predicting PD using previously identified high-risk factors.^{11,13} The median (range) of the number of repeated tests are as follows: Hb 3 (2-12),

Table 3. SMM outcomes risk stratified by baseline variables and evolving variables over 1 y

	n (%)	Median TTP, mo	Log-rank P	2-y PD, %	Overall PD, %	Specificity, %	Sensitivity, %	Accuracy, %	2-y AUC	Integrated time-dependent AUC
Baseline Factors										
dFLC										
dFLC <100 mg/L	111 (41)	92.6	.0454	14	3					
$dFLC \geq 100 \text{ mg/L}$	72 (26)	71.3		36	44	68	61	66	0.64	0.55
NE	90 (33)									
FLCr										
FLCr <8	81 (30)	96.1	.0325	12	31					
FLCr ≥8	102 (37)	71.3		31	45	51	76	56	0.63	0.55
NE	90 (33)									
FLCr <100	156 (57)	92.6	.0019	19	36					
FLCr ≥100	27 (10)	40.2		44	56	90	28	75	0.59	0.59
NE	90 (33)									
m-protein										
<3 g/dL	189 (69)	115.2	<.0001	20	38					
≥3 g/dL	31 (11)	21.8		45	74	91	29	75	0.60	0.47
NE	53 (19)									
FISH*										
Normal	91 (33)	73.0	.1199	21	40					
Hyperploidy	33 (12)	74.2		27	45					
High risk	25 (9)	36.1		32	52	85	21	58	0.53	0.58
NE	124 (45)									
BMPC										
<60	251 (92)	79.2	.0006	20	43					
≥60	22 (8)	30.6		41	73	94	15	77	0.55	0.42
NE	0 (0)									
Mayo evolving factors										
eHB										
No eHb	94 (34)	115.2	.0002	12	30					
eHb	119 (44)	42.2		29	51	51	77	55	0.64	0.61
NE	60 (22)									
eMP										
No eMP	95 (35)	96.1	.0042	17	41					
eMP	106 (39)	35.1		33	53	68	56	57	0.62	0.46
NE	72 (26)									
GBTM factors										
eHB										
No eHb	180 (66)	115.2	<.0001	14	35					
eHb	35 (13)	26.3		43	66	89	36	79	0.62	0.43
NE	58 (21)									
eMP										
No eMP	112 (41)	115.2	.0230	14	38					
eMP	33 (12)	39.8		36	58	82	42	74	0.62	0.58
NE	128 (47)									

Baseline factors are dFLC, FLCr, or BMPC at the time of SMM diagnosis. Mayo evolving factors are defined as evolving m-protein/lg or evolving Hb over the first 12 mo of diagnosis as defined by Ravi et al. GBTM factors are determined using statistical software from evolving Hb, evolving m-protein, evolving FLCr, or evolving dFLC over the first 12 mo of diagnosis.

NE, not evaluable; NR, not reached.

*For calculation of 2-y diagnostic statistics (specificity, sensitivity, accuracy, AUC), normal/hyperdiploidy/other FISH cytogenetic groups were combined.

Table 3. (continued)

	n (%)	Median TTP, mo	Log-rank P	2-y PD, %	Overall PD, %	Specificity, %	Sensitivity, %	Accuracy, %	2-y AUC	Integrated time-dependent AUC
eFLCr										
No eFLCr	108 (40)	NR	.0028	14	31					
eFLCr	19 (7)	37.2		32	63	88	28	78	0.58	0.62
NE	146 (53)									
edFLC										
No edFLC	104 (38)	115.2	.0586	13	33					
edFLC	23 (9)	45.3		30	48	85	34	76	0.60	0.58
NE	146 (53)									

Baseline factors are dFLC, FLCr, or BMPC at the time of SMM diagnosis. Mayo evolving factors are defined as evolving m-protein/lg or evolving Hb over the first 12 mo of diagnosis as defined by Ravi et al. GBTM factors are determined using statistical software from evolving Hb, evolving m-protein, evolving FLCr, or evolving dFLC over the first 12 mo of diagnosis. NE. not evaluable: NR. not reached.

*For calculation of 2-y diagnostic statistics (specificity, sensitivity, accuracy, AUC), normal/hyperdiploidy/other FISH cytogenetic groups were combined.

M-protein 4 (2-12) and FLC 3 (2-10). BMPC \geq 20%, m-protein \geq 3 g/dL, immunoparesis, eHb, eMP, eFLCr, and edFLC were identified as significant contributors in univariable analyses models (Table 4). However, when analyzed in a multivariable model, immunoparesis, eMP, eHb, and edFLC were identified as the 4 most significant predictors of 2-year PD.

Ultra-high-risk patients by our multivariable model (\geq 3 risk factors) had a median TTP of 13 months and 2-year PD of 88% (Figure 3). The results of our 4-factor GBTM multivariable model were compared against previous risk stratification models applied to our patients (Table 5). The specificity of our model was 99% compared with 93% (Mayo baseline) and 85% (Mayo evolving).^{11,13}

Of the 35 SMM patients enrolled in clinical trials involving agents with no single-agent activity, there were 4 and 19 in the FLCr \geq 100 and <100 groups, respectively, and 5 and 24 in the BMPC \geq 60 and <60 groups, respectively. Compared with the nonclinical trial population, there were no significant differences in base-line characteristics or distribution according to Mayo baseline or

evolving risk models. Their removal slightly increased the median TTP but did not affect the GBTM modeling.

Discussion

Although there is great interest in treating high-risk SMM population to avoid end-organ damage, this must be counterbalanced by early and potentially indefinite therapy of some patients who may not necessarily progress. In this study, we examined the outcomes of 273 SMM patients followed at our institution.

Similar to other studies, progression events in our study predominantly resulting from anemia and bone lesions.^{4,5} Of the patients who progressed by bone disease, ~50% in our study were detected either by PET-CT or MRI scans, illustrating the importance of sensitive bone imaging. The progression by renal failure was also comparable with 9% in our patients vs 11% to 18% in other studies.^{4,5} Among those with FLCr \geq 100, the rate of renal PD was 20% vs 27%, as previously reported.⁴ Although those with baseline BJP were twice as likely to have FLCr \geq 100 at baseline, there was no associated increased risk of renal PD. There were insufficient



Figure 1. TTP to symptomatic myeloma stratified by free light chain ratio and BMPC percentage. (A) Median time to progression was 40 months in FLCr \geq 100 compared with 93 months in FLCr <100 (P = .0019). (B) Median time to progression was 31 months in BMPC \geq 60 compared with 79 months in BMPC <60 (P = .0006).



Figure 2. Evolving changes in hemoglobin, m-protein, free light chain ratio, and differences in free light chain identified through GBTM 1 year after diagnosis of SMM. (A) Changes in Hb are in g/dL. Changes in m-protein (B), free light chain ratio (C), and differences in free light chain ratio (D) are % from baseline. The trajectories of high risk evolving GBTM groups are indicated.

patients to analyze evolving BJP in our GBTM or multivariable model; this warrants further study.

Although FLCr \geq 100 and BMPC \geq 60 were confirmed as high-risk groups in our study, the groups' prognosis was better than initial studies reported. The median TTP in FLCr \geq 100 was 40 months, with 2-year progression at 44% and ultimate progression at 56%. When compared with previously published studies (Table 6), all previous studies have shown shorter median TTP for FLCr \geq 100 with the exception of the Danish study, in which median TTP was not even reached. Furthermore, our 2-year progression rate falls between previously reported values that range from 30% to 98%. The 56% overall rate of progression is substantially lower in our study than previously reported in 2 other studies.

The marked heterogeneity in median TTP and progression rates for FLCr \geq 100 may be due to several reasons. First of all, FLC measurements can vary depending on reagents, Freelite (the Binding Site Group Ltd.) a monoclonal-based antisera vs N Latex FLC assay (Siemens Healthcare) a polyclonal-based antisera, immunonephelometric vs immunoturbidimetric technique, and the choice of the analyzer used. In fact, most patients with FLCr of 100 by Freelite would be <100 by N Latex FLC.^{19,20} Single-institution

studies may also suffer from either increased referral from local oncologists or self-referral of patients with perceived higher of risk of progression. Indeed, the Danish study, the only prospective registry published to date, has the lowest rate of PD. Another possibility is the period of investigation, with some studies dating back to the 1970s; improvements in diagnosis (in particular, greater use of whole-body low-dose CT, PET-CT, and/or MRI scans to rule out active myeloma) may have appropriately excluded active myeloma patients in recent studies. Finally, the requirement for an FLCr within 3 months from SMM diagnosis is not clearly stated in all studies and the absence of this inclusion criterion might skew results as well. It is worth noting that the lack of standardized baseline and following up testing time points for both serologies as well as appropriate radiologic studies is an important limitation of all SMM retrospective studies, including ours, and should give pause when prospective/registry studies are yielding discrepant results.

In patients with BMPC \geq 60, the median TTP was 31 months with 41%/73% 2-year/overall PD rates, which are lower than previously reported median TTP ranging from 7 to 15 months, and nearly all patients progressed to active myeloma. BMB tend to have larger numbers of plasma cells than BMA and, in the United States, BMB are routine, whereas in Europe (eg, the GIMEMA study), all patients

Table 4. Multivariable model predicting SMM risk for PD

	Univariabl	e	Multivariable			
n = 90*	HR (95% CI)	Р	HR (95% CI)	Р		
Age	1.002 (0.97-1.03)	.9007				
Male	0.88 (0.47-1.65)	.6824				
BMPC ≥20%	3.29 (1.45-7.49)	.0046				
$BMPC \geq \!\! 60\%$	0.98 (0.30-3.25)	.9790				
m-protein ≥3 g/dL	3.59 (1.80-7.17)	.0003				
IgA SMM	0.72 (0.30-1.73)	.4645				
Immunoparesis	2.90 (1.46-5.77)	.0025	3.90 (1.80-8.44)	.0006		
$\begin{array}{l} FLCr \geq \! 100 \text{ and} \\ dFLC \geq \! 100 \end{array}$	1.53 (0.59-3.99)	.3827				
$dFLC \geq \! 100 \text{ mg/L}$	1.36 (0.70-2.64)	.3658				
eMP	3.64 (1.89-6.99)	.0001	3.98 (1.80-8.44)	<.0001		
eHb	4.54 (2.22-9.29)	<.0001	8.05 (3.53-18.35)	<.0001		
eFLCr	2.09 (1.04-4.21)	.0395				
edFLC	3.02 (1.45-6.27)	.0031	2.84 (1.28-6.29)	.0100		

Only 6 patients had BMPC $\geq\!60\%$, thereby limiting detection of statistical differences in progression curves at this cutoff.

HR, hazard ratio.

*Patients were required to have all listed variables to be eligible for complete case analysis.

had a BMA but only ~50% also had a BMB.^{21,22} Of the 127 patients who had both procedures, 7 had a BMB \ge 60%, but the only early PD occurred in the patient with concordant BMA \ge 60%. Given the limited number of patients studied to date (a combined global sample size of 45) and sampling issues (not only BMB vs BMA, but also heterogeneous MM involvement of the marrow), the optimal threshold for BMPC % in predicting a high-risk SMM cohort by BMB warrants further study.

FLCr and BMPC at the time of SMM diagnosis share the same limitation in predicting MM progression: baseline disease burden and characteristics do not take into account disease biology and

evolution of the disease. It is now known that the majority of cytogenetic changes in MM can also be detected in monoclonal gammopathy of undetermined significance and that the progression to MM likely involves the expansion of preexisting clones regulated by growth-permissive and restrictive signals from immune cells, bone cells, and other cells in the bone marrow niche.²³ The lack of progression by more than one-third of our FLCr ≥100 patients led us to an investigation of evolving biomarkers to identify an ultra-highrisk group for progression. Through GBTM, we identified groups at high risk for progression based on the evolving trajectories of eHb, eMP, eFLCr, and edFLC rather than through arbitrary percent changes. Although no single biomarker, neither baseline nor evolving, predicted 80% 2-year PD, multivariable analysis identified a model in which immunoparesis, eHb, eMP, and edFLC were significant predictors for MM progression, with a median TTP and 2-year PD of 13 months and 88% when \geq 3 were present. Of note, only edFLC (and not eFLCr) maintained significance in multivariable analysis, perhaps because of immunoparesis capturing the suppression of the uninvolved FLC that affects FLCr. Although requiring validation, these results suggest that, similar to the use of FLCr and dFLC in symptomatic MM and although baseline FLCr may be more significant in predicting SMM progression, evolving changes in dFLC may be more suitable for SMM monitoring.

The findings in this study support previous findings that FLCr \geq 100 and BMPC \geq 60 identify high-risk SMM patients, but with a more modest progression to MM than previously published. Further investigation into whether the predictive value of these biomarkers alone warrant treatment of otherwise asymptomatic patients is needed. This is particularly relevant for BMPC \geq 60 given the few patients with this finding in all published studies combined. Furthermore, the GBTM identified high-risk variables need to be further validated in other large data sets. Taken together, these suggest that a meta-analysis with pooled multi-institutional data from previously published studies, as is being proposed by the IMWG, would be of great benefit. This would allow a global, standardized risk stratification, avoid the inadvertent initiation of treatment of patients who may remain asymptomatic for long



Figure 3. TTP to symptomatic myeloma stratified based on risk factors (immunoparesis, eHb GBTM, eMP GBTM, and edFLC GBTM). The median times to progression for 0, 1, 2, or \geq 3 risk factors were not reached: 77, 26, and 13 months, respectively (*P* <.0001).

Table 5. Comparison of high-risk smoldering multiple myeloma

n = 90*	n (%)	Median TTP, mo	Log-rank P	2-y PD, %	Overall PD, %	Specificity, %	Sensitivity, %	Diagnostic accuracy	2-y AUC	Integrated time-dependent AUC
Mayo Clinic baseline score†										
1	37 (41)	96.1	.0218	8	32					
2	43 (48)	48.1		26	44					
3	10 (11)	24.7		50	80	93	25	79	0.70	0.59
Mayo Clinic evolving score‡										
0	10 (11)	115.2	<.0001	0	20					
1	34 (38)	96.1		9	35					
2	24 (27)	NR		21	33					
3	22 (24)	21.8		50	77	85	58	79	0.78	0.68
Mount Sinai multivariable score§										
0	29 (32)	NR	<.0001	3	14					
1	36 (40)	77.3		11	42					
2	17 (19)	26.3		41	71	86	74			
3 or 4	8 (9)	13.4		88	100	99	35	86	0.85	0.74

*Patients were required to have all listed variables described for the complete case analysis.

tBMPC ≥10%, m-protein ≥3 g/dL, FLCr ≥8.

\$BMPC ≥20%, eMP (≥10% increase in m-protein and/or immunoglobulin within the first 6 mo and/or ≥25% increase in m-protein and/or immunoglobulin within the first 12 mo), eHb (≥0.5 g/dL decrease within the first 12 mo).

§Immunoparesis, eHb GBTM, eMP GBTM, edFLC GBTM.

periods, and ensure appropriately powered prospective therapeutic studies because of a better understanding of the expected rates of progression without intervention. Most important, reproducible identification of a clinically defined ultra-high-risk group is essential to guide translational research efforts in identifying the best targets for early intervention.

Table 6. Comparison of high-risk SMM at various institutions

	Mayo Clinic ⁴	University of Athens ⁵	University of Pennsylvania ^{6,8}	Denmark ⁷	MM GIMEMA-Latium Working Group ¹⁰	Mount Sinai
No. of centers	Single	—	Single	Multi	Multi	Single
Inclusion criteria*	Yes	—	—	Yes	Yes	Yes
FLCR						
Years of SMM diagnosis	1970-2010	—	2008-2012†	2005-2013	_	2003-2015
n	586	96	118	209	_	185
Median TTP, mo	40	66	82	+	—	78
FLCR ≥100, n (%)	90 (15)	—	11 (9)	23 (11)	_	27 (15)
Median TTP, mo	15	13	20	+	—	40
2-y progression, %	72	98§	64	30	_	44
Overall progression, %	98	100	—	_	—	56
BMPC						
Years of SMM diagnosis	1996-2010	_	2008-2012†	_	1980-2010	1992-2015
n	655	96	121	_	397	273
Median TTP, mo	40	66	82	_	_	74
BMPC ≥60, n (%)	21 (3.2)	8 (8)	6 (5)	_	10 (2.5)	22 (8)
Median TTP, mo	7	15	1.2	_	_	31
2-y progression, %	95	95.5¶	100	_	100	41
Overall progression, %	_	100	100	_	100	73

*Stipulation that laboratory data be obtained within 3 mo of diagnosis.

†Years of investigation, not diagnosis, was provided in methodology.

#Not enough patients progressed to calculate median TTP.

§Data at 14 mo.

||n = 7 by bone marrow biopsy core; n = 10 by bone marrow aspirate. ¶Data at 18 mo.

Acknowledgments

This work was supported by grants from the Biostatistics Shared Resource Facility, Icahn School of Medicine at Mount Sinai, and National Institutes of Health, National Cancer Institute Cancer Center Support Grant P30 CA196521-01.

Authorship

Contribution: V.W., E.M., S.L., and A.C. contributed to data acquisition, interpretation, and analysis; and all authors drafted and reviewed the manuscript, approved the final version, decided to publish this report, and have vouched for data accuracy and completeness.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

Correspondence: Ajai Chari, Tisch Cancer Institute, Icahn School of Medicine at Mount Sinai, 1470 Madison Ave, New York NY 10029; e-mail: ajai.chari@mountsinai.org.

References

- 1. Kyle RA, Remstein ED, Therneau TM, et al. Clinical course and prognosis of smoldering (asymptomatic) multiple myeloma. N Engl J Med. 2007;356(25): 2582-2590.
- 2. Rajkumar SV, Dimopoulos MA, Palumbo A, et al. International Myeloma Working Group updated criteria for the diagnosis of multiple myeloma. *Lancet* Oncol. 2014;15(12):e538-e548.
- 3. Rajkumar SV, Kyle RA. Haematological cancer: treatment of smoldering multiple myeloma. Nat Rev Clin Oncol. 2013;10(10):554-555.
- Larsen JT, Kumar SK, Dispenzieri A, Kyle RA, Katzmann JA, Rajkumar SV. Serum free light chain ratio as a biomarker for high-risk smoldering multiple myeloma. Leukemia. 2013;27(4):941-946.
- 5. Kastritis E, Terpos E, Moulopoulos L, et al. Extensive bone marrow infiltration and abnormal free light chain ratio identifies patients with asymptomatic myeloma at high risk for progression to symptomatic disease. *Leukemia*. 2013;27(4):947-953.
- 6. Waxman AJ, Mick R, Garfall AL, et al. Classifying ultra-high-risk smoldering myeloma. Leukemia. 2015;29(3):751-753.
- 7. Sørrig R, Klausen TW, Salomo M, et al; Danish Myeloma Study Group. Smoldering multiple myeloma risk factors for progression: a Danish populationbased cohort study. *Eur J Haematol.* 2016;97(3):303-309.
- 8. Waxman AJ, Mick R, Garfall AL, et al. Modeling the risk of progression in smoldering multiple myeloma. Leukemia. 2015;29(3):751-753.
- 9. Rajkumar SV, Larson D, Kyle RA. Diagnosis of smoldering multiple myeloma. N Engl J Med. 2011;365(5):474-475.
- 10. Rago A, Grammatico S, Za T, et al; Multiple Myeloma GIMEMA-Latium Region Working Group. Prognostic factors associated with progression of smoldering multiple myeloma to symptomatic form. *Cancer.* 2012;118(22):5544-5549.
- 11. Ravi P, Kumar S, Larsen JT, et al. Evolving changes in disease biomarkers and risk of early progression in smoldering multiple myeloma. *Blood Cancer J*. 2016;6(7):e454.
- 12. Fernández de Larrea C, Isola I, Pereira A, et al. Evolving M-protein pattern in patients with smoldering multiple myeloma: impact on early progression. Leukemia. 2018;32(6):1427-1434.
- Dispenzieri A, Kyle RA, Katzmann JA, et al. Immunoglobulin free light chain ratio is an independent risk factor for progression of smoldering (asymptomatic) multiple myeloma. *Blood*. 2008;111(2):785-789.
- 14. Jones BL, Nagin DS, Roeder K. A SAS procedure based on mixture models for estimating developmental trajectories. Sociol Methods Res. 2001;29(3): 374-393.
- 15. Uno H, Cai T, Tian L, Wei LJ. Evaluating prediction rules for t-year survivors with censored regression models. J Am Stat Assoc. 2007;102:527-537.
- 16. Shtatland E, Kleinman K, Cain EM. Model Building in PROC PHREG with automatic variable selection and information. Paper presented at Thirtieth Annual SAS Users Group International Conference. 10-13 April, 2005. Philadelphia, PA.
- 17. Rajkumar SV, Gupta V, Fonseca R, et al. Impact of primary molecular cytogenetic abnormalities and risk of progression in smoldering multiple myeloma. Leukemia. 2013;27(8):1738-1744.
- Neben K, Jauch A, Hielscher T, et al. Progression in smoldering myeloma is independently determined by the chromosomal abnormalities del(17p), t(4; 14), gain 1q, hyperdiploidy, and tumor load. J Clin Oncol. 2013;31(34):4325-4332.
- 19. Jacobs JF, Tate JR, Merlini G. Is accuracy of serum free light chain measurement achievable? Clin Chem Lab Med. 2016;54(6):1021-1030.
- 20. Tate JR, Graziani MS, Mollee P, Merlini G. Protein electrophoresis and serum free light chains in the diagnosis and monitoring of plasma cell disorders: laboratory testing and current controversies. *Clin Chem Lab Med.* 2016;54(6):899-905.
- 21. Terpstra WE, Lokhorst HM, Blomjous F, Meuwissen OJ, Dekker AW. Comparison of plasma cell infiltration in bone marrow biopsies and aspirates in patients with multiple myeloma. *Br J Haematol.* 1992;82(1):46-49.
- 22. Chilosi M, Adami F, Lestani M, et al. CD138/syndecan-1: a useful immunohistochemical marker of normal and neoplastic plasma cells on routine trephine bone marrow biopsies. Mod Pathology. 1999;12(12):1101-1106.
- 23. Dhodapkar MV. MGUS to myeloma: a mysterious gammopathy of underexplored significance. Blood. 2016;128(23):2599-2606.