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### The *ACVRL1* c.314-35A>G Polymorphism is Associated with Organ Vascular Malformations in Hereditary Hemorrhagic Telangiectasia Patients with *ENG* Mutations, but not in Patients with *ACVRL1* Mutations

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#### Abstract

Hereditary hemorrhagic telangiectasia (HHT) is characterized by the presence of vascular malformations (VMs) and caused by mutations in TGFβ/BMP9 pathway genes, most commonly *ENG* or *ACVRL1*. Patients with HHT have diverse phenotypes related to skin and mucosal telangiectases and organ VMs, including arteriovenous malformations (AVM). The clinical heterogeneity of HHT suggests a potential role for genetic modifier effects. We hypothesized that the common polymorphisms *ACVRL1* c.314-35A>G and *ENG* c.207G>A, previously associated with sporadic brain AVM, are also associated with organ VM in HHT. We genotyped *ACVRL1* c. 314-35A>G and *ENG* c.207G>A in 716 patients with HHT recruited by the Brain Vascular Malformation Consortium and evaluated association of genotype with presence of any organ VM, and specifically with brain VM, liver VM and pulmonary AVM, by multivariate logistic

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regression analyses stratified by HHT mutation. Among all patients with HHT, neither polymorphism was significantly associated with presence of any organ VM; *ACVRL1* c. 314-35A>G showed a trend toward association with pulmonary AVM (OR=1.48, p=0.062). *ACVRL1* c.314-35A>G was significantly associated with any VM among patients with HHT with *ENG* (OR=2.66, p=0.022), but not *ACVRL1* (OR=0.79, p=0.52) mutations. *ACVRL1* c.314-35A>G was also significantly associated with pulmonary AVM and liver VM among *ENG* mutation carriers. There were no significant associations between *ENG* c.207G>A and any VM phenotype. These results suggest that common polymorphisms in HHT genes other than the mutated gene modulate phenotype severity of HHT disease, specifically presence of organ VM.

#### Keywords

hereditary hemorrhagic telangiectasia; vascular malformation; arteriovenous malformation; phenotype; genetic modifier

#### INTRODUCTION

Hereditary hemorrhagic telangiectasia (HHT) is an autosomal dominant Mendelian disorder caused by mutations in several genes encoding proteins in the TGF $\beta$ /BMP9 signaling pathway [McDonald et al., 2011]. Approximately 85% of patients with HHT have a mutation in *ENG* [McAllister et al., 1994] or *ACVRL1* [Johnson et al., 1996] and 3% have a mutation in *SMAD4* [Gallione et al., 2004]. *BMPR2* [Rigelsky et al., 2008] and *BMP9* [Wooderchak-Donahue et al., 2013] mutations have been reported in syndromes with phenotypic overlap. Patients with HHT have skin and mucosal telangiectases and vascular malformations (VM), including arteriovenous malformations (AVM), in visceral organs including brain, liver and lung, which can lead to chronic bleeding, anemia, life-threatening hemorrhage, stroke and heart failure[Faughnan et al., 2011; McDonald et al., 2011]. Pulmonary AVM and brain VM are more common in patients with *ENG* mutations, while liver VM are more common in patients with *ACVRL1* mutations [Bayrak-Toydemir et al., 2006]. The considerable clinical heterogeneity of HHT phenotypes, even among patients with the same mutated HHT gene and within families, suggests a possible role for genetic modifier effects.

Two common polymorphisms in HHT genes, *ACVRL1* c.314-35A>G (*ACVRL1* IVS3-35A>G) and *ENG* c.207G>A, have been reported to be associated with sporadic brain AVM [Pawlikowska et al., 2005]; the *ACVRL1* c.314-35A>G association has been replicated in 2 independent cohorts [Simon et al., 2006]. *ENG* c.207G>A was also associated with surgical outcomes in sporadic brain AVM [Shen et al., 2014]. We hypothesized that these polymorphisms are also associated with organ VM in HHT and tested this hypothesis in an HHT cohort recruited by the Brain Vascular Malformation Consortium (BVMC) [Akers et al., 2013].

#### METHODS

#### **Cohort Recruitment**

Patients with a confirmed clinical HHT diagnosis by the Curaçao criteria [Faughnan et al., 2011] were enrolled by the BVMC HHT Project as previously described (http:// rarediseasesnetwork.epi.usf.edu/BVMC) [Akers et al., 2013]. We analyzed 716 patients enrolled at 11 sites between June 2010 and December 2013. All patients provided written informed consent, including for genetic studies. The study protocol was approved by each institutional review board. Data collected included age, sex, family relationships, genetic testing results (mutated gene: ACVRL1, ENG, SMAD4, unknown), clinical presentation and symptoms. Patients were screened for organ VM and other clinical features according to standard clinical practice and International HHT Guidelines [Faughnan et al., 2011], including: comprehensive history, physical, routine blood tests, screening for pulmonary AVM by contrast echocardiography, brain VM screening by magnetic resonance imaging, clinical screening for liver VM (chronic right upper quadrant pain, portal hypertension, high-output heart failure, liver bruit on examination, abnormal liver function tests) and clinical screening for recurrent spontaneous epistaxis and HHT-related gastrointestinal bleeding. If screening was positive for pulmonary AVM or brain VM, patients underwent further diagnostic imaging and treatment, where appropriate. If clinical assessment was suggestive of symptomatic liver VM, diagnostic imaging was recommended and therapy where appropriate. The BVMC HHT cohort targets 25% brain VM-positive patients; other cohort characteristics are similar to other cohorts [Letteboer et al., 2006; Nishida et al., 2012]. The majority of the cohort is Caucasian (96%) reflecting patient populations at participating centers.

Patients provided blood or saliva (Oragene, DNA Genotek, Ontario, Canada) samples, which were sent for DNA extraction and banking to the NINDS Repository at Coriell Institute (http://ccr.coriell.org/Sections/Collections/NINDS), or to the University of California, San Francisco (UCSF) (saliva).

#### Genotyping

Genotyping was performed by staff blinded to clinical phenotype. *ACVRL1* c.314-35A>G (formerly *ACVRL1* IVS3-35A>G, rs2071219) and *ENG* c.207G>A (rs11545664) were genotyped using Taqman<sup>TM</sup> assays (C\_15868502\_10 and C\_25592400\_10, Applied Biosystems, Foster City, CA). Both SNPs had genotype call rates >98% and were in Hardy Weinberg equilibrium among Caucasians (p>0.05).

#### Statistical Analysis

Phenotype frequencies were compared by Fisher's exact test. Genotypes were collapsed for analysis into risk genotype carriers (*ACVRL1* c.314-35A>G: AA+AG, *ENG* c.207G>A: GG) vs. non-carriers [Pawlikowska et al., 2005]. Phenotypes evaluated included pulmonary AVM, liver VM, brain VM and the composite phenotype "any organ VM", defined as presence of any organ VM or AVM. Association of genotype with phenotype was evaluated by logistic regression analyses adjusted for age, gender and clustering within known families and further stratified by HHT mutation status (*ACVRL1* or *ENG*) for the two most frequently

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mutated genes. Since liver VM analysis restricted to *ENG* subjects produced an infinite estimated odds ratio (OR) for *ACVRL1* c.314-35A>G, we calculated a one-sided 95% confidence interval (CI) based on the profile likelihood and derived a p-value from the likelihood ratio test comparing models with and without the polymorphism. To ensure familial clustering could be disregarded for this specific analysis, we tested the effect of family with a mixed-effects logistic regression model. The statistical significance threshold was set at p=0.025 after Bonferroni correction for 2 polymorphisms. We also evaluated associations using a co-dominant genetic model testing each genotype compared to the lowest-risk genotype [Pawlikowska et al., 2005], and restricted to Caucasians.

#### RESULTS

Of 716 patients with HHT studied, 93.7% were Caucasian and 59% were female; 71% had at least one VM of any type, 50% had pulmonary AVM, 20% had liver VM and 20% had brain VM (Table I). Known family relationships were recorded for 177 patients (2–4 patients per family, except for 1 family of 17 distant relatives). Among the 436 (61%) patients with mutation data, 48% had *ENG* mutations, 43% had *ACVRL1* mutations and 4% had *SMAD4* mutations (Table I). As reported for other cohorts [Bayrak-Toydemir et al., 2006; Letteboer et al., 2006], patients with *ENG* mutations were more likely to have any organ VM, brain VM and pulmonary AVM (all p<0.0001), while patients with *ACVRL1* mutations were more likely to have liver VM (p<0.0001) and anemia (p=0.003) (Table I). Compared to patients without genetic testing results, patients with *ACVRL1* or *ENG* mutations were more likely to have any organ VM (p=0.002), pulmonary AVM (p<0.0001) and anemia (p=0.001), likely due to more extensive clinical evaluation of patients from multiplex families.

Among 689 patients with HHT with VM data, neither polymorphism was significantly associated with organ VM, although *ACVRL1* c.314-35A>G showed a trend toward association with pulmonary AVM (OR=1.48, 95%CI=0.90–2.22, p=0.062) and liver VM (OR=1.46, 95%CI=0.85–2.50, p=0.17) (Table II).

When stratifying by HHT mutation, *ACVRL1* c.314-35A>G was significantly associated with organ VM among the 211 *ENG* mutation carriers (overall: OR=2.66, 95%CI=1.15–6.13, p=0.022; Caucasians: OR=2.83, 95%CI=1.18–6.76, p=0.020), but not among the 189 *ACVRL1* mutation carriers (OR=0.79, CI=0.38–1.63, p=0.52). In *ENG* mutation carriers, *ACVRL1* c.314-35A>G was also significantly associated with pulmonary AVM (OR=2.45, p=0.016) and liver VM (p=0.001, all 21 liver VM-positive patients carried the A risk allele), but not with brain VM (Table II). When restricted to Caucasians, all significant associations were consistent (data not shown).

There were no statistically significant associations between *ENG* c.207G>A and VM of any type. However, among *ACVRL1* mutation carriers only, the effect direction was consistent with association of *ENG* c.207G>A with VM overall (OR=1.29, p=0.5) and with brain VM (OR=3.09, p=0.16) (Table II). Again, no associations with *ENG* c.207G>A were observed among *ENG* mutation carriers.

#### DISCUSSION

We report here the first evidence that common polymorphisms in HHT genes other than the mutated gene are associated with differences in HHT phenotype severity, and specifically presence of organ VM. In a well-characterized cohort of patients with HHT, the common polymorphism *ACVRL1* c.314-35A>G, previously reported as associated with sporadic brain AVM [Pawlikowska et al., 2005; Simon et al., 2006] was also associated with pulmonary AVM, liver VM and overall, with presence of any organ VM in patients with *ENG* mutations, but not in patients with *ACVRL1* mutations. Conversely, there was also a pattern, although not statistically significant, for association of the *ENG* c.207G>A with any VM and brain VM in patients with *ACVRL1*, but not *ENG* mutations. These findings suggest that common genetic variation in HHT genes other than the mutated gene may modify HHT phenotype severity.

Common polymorphisms acting as genetic modifiers have been reported in other Mendelian diseases, such as cystic fibrosis [Wright et al., 2011] and there are other reports of phenotype-modifying polymorphisms near the gene that when mutated causes disease: a common polymorphism in *KCNQ1*, rs2074238, is associated with symptomatic status in patients with long-QT syndrome, who are heterozygous for *KCNQ1* or *KCNH2* (disease causative) mutations [Duchatelet et al., 2013]; and 2) a common polymorphism in *TERT* is associated with survival and with disease recurrence in bladder cancer caused by somatic mutations in the *TERT* promoter [Rachakonda et al., 2013]. In patients with HHT, common variants in *PTPN14* and *ADAM17*, genes in loci originally mapped as phenotype modifiers in TGF $\beta$  knockout mice, have been reported to be associated with pulmonary VM [Benzinou et al., 2012; Kawasaki et al., 2014]. These associations have not yet been replicated in independent cohorts.

The association with the common polymorphism *ACVRL1* c.314-35A>G was originally reported for sporadic brain AVM in two independent cohorts [Pawlikowska et al., 2005; Simon et al., 2006] and for dural arteriovenous fistulae in one cohort [Simon et al., 2006]. Extension of these associations to organ VMs in HHT suggests common mechanisms of VM development in different organs (brain, lung, liver) in both sporadic and HHT disease.

The functional effects of *ACVRL1* c.314-35A>G are unknown. It is located 35 base-pairs from an intron-exon junction and is hypothesized to affect splicing [Pawlikowska et al., 2005]. As with sporadic brain AVM disease, the observed HHT VM association is with the major A allele carried by the majority of patients (GG genotype carriers have fewer VMs). ENG c.207A>G is a synonymous exonic variant, however it is also a cis eQTL linked to ENG mRNA expression in monocytes [Zeller et al., 2010]. Thus both polymorphisms could result in reduced expression or expression of an abnormal protein, further impairing TGF $\beta$ /BMP9 pathway signaling.

We acknowledge several limitations of our study. The genetic association results require replication in an independent HHT cohort and functional studies to determine their molecular mechanism. Family relationships (known and cryptic) within the cohort are a confounder; however, the statistical adjustment we used for known relatedness is

conservative, and cryptic relationships should be rare. The findings are currently limited to Caucasians, as there are not enough patients of other ethnicities in the BVMC cohort to evaluate the effect in non-Caucasians. The *ENG* c.207A>G associations did not reach statistical significance and require a larger cohort to confirm or rule out. Unlike for sporadic brain AVM, we found no strong evidence for *ACVRL1* c.314-35A>G association with brain VM in HHT, however the number of brain VM-positive patients is small resulting in low statistical power. The prevalence of brain VM in the BVMC cohort is targeted by design (25% planned, 20% observed), but similar (top end of the reported range) to frequencies observed in other HHT cohorts [Letteboer et al., 2006; Nishida et al., 2012]. Screening for liver VM was based on clinical assessment, and therefore the liver VM-positive group includes patients with more severe, clinically evident liver VM, consistent with the overall focus on organ VM involvement as a marker of more severe HHT disease. The strengths of our study include the large size of the HHT cohort and the comprehensive phenotype information collected.

In conclusion, we report for the first time that common polymorphisms in HHT genes other than the mutated gene may modify the HHT phenotype, and that genetic associations previously reported in sporadic brain AVM extend to the Mendelian form of the disease. Our results suggest a multiple-hit mechanism, where diverse genetic hits to multiple HHT genes may predispose to the most severe phenotypic manifestations. The findings provide insights into potential mechanisms underlying the marked clinical heterogeneity of HHT, where some patients suffer from severe VM organ involvement with devastating complications, while others have only skin telangiectases. A better understanding of which patients with HHT are at highest risk of complications would aid in clinical management, risk stratification and therapy development.

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**TABLE I** 

Demographic and Clinical Characteristics of HHT Subjects

haracteristic	All Subjects	ACVRLI	ENG	$\mathbf{P}^{*}$
emale sex	425 / 716 (59%)	107/189 (57%)	127/211 (60%)	0.48
ge at baseline (y)	$46.1 \pm 19.1$	$45.2\pm21.3$	$43.1\pm18.8$	0.30
aucasian	671 / 700 (96%)	180/184 (98%)	201/208 (97%)	0.52
HT mutation				
ACVRLI	189 / 436 (43%)	189/189 (100%)		
ENG	211 / 436 (48%)	·	211/211 (100%)	
SMAD4	16 / 436 (4%)	ı		
All negative	20 / 436 (5%)	I		
И	490 / 689 (71%)	90/181 (50%)	172/209 (82%)	<0.001
Brain VM	140 / 716 (20%)	21/189 (11%)	66/211 (31%)	<0.001
Pulmonary AVM	346 / 690 (50%)	34/183 (19%)	140/208 (67%)	<0.001
Liver VM	138/ 677 (20%)	47/182 (26%)	21/203 (10%)	<0.001
pistaxis	678 / 700 (97%)	177/187 (95%)	201/208 (97%)	0.46
[ Bleeding	115 / 682 (17%)	22/183 (12%)	34/205 (17%)	0.25
nemia	331 / 675 (49%)	92/180 (51%)	74/206 (36%)	0.003
<i>CVRL1</i> c.314-35A>G gei	notype			0.68
66	141 / 716 (20%)	37/189 (20%)	46/211 (22%)	
AG	337 / 716 (47%)	82/189 (43%)	97/211 (46%)	
AA	238 / 716 (33%)	69/189 (37%)	68/211 (32%)	
VG c.207G>A genotype				0.27
AA	9 / 716 (2%)	6/189 (3%)	3/211 (1%)	
AG	121 / 716 (17%)	34/189 (18%)	30/211 (14%)	
66	581 / 716 (81%)	149/189 (79%)	178/211 (84%)	

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Values are the number observed over the total number of non-missing (and the percent) or mean  $\pm$  standard deviation.

\* P-values are from comparison of subjects with known ACVRLI and ENG mutations with Fisher's exact test or a two-sample t-test. HHT, hereditary hemorrhagic telangiectasia; VM, any HHT organ vascular malformation including: brain VM, pulmonary arteriovenous malformation (AVM) or liver VM; GI, gastro-intestinal. Author Manuscript

# **TABLE II**

Association of ACVRL1 c.314-35A>G and ENG c.207G>A with VM Phenotypes in Patients with HHT

			Α	ll Subjects		Subje	ects wit	h <i>ACVRLI</i> Mı	ıtation	Su	bjects v	vith <i>ENG</i> Mut	ation
Outcome	SNP	=	OR	95% CI	d	u	OR	95% CI	d	п	OR	95% CI	ď
MV	ACVRL1 c.314-35A>G	689	1.34	(0.87, 2.07)	0.178	181	0.79	(0.38, 1.63)	0.520	209	2.66	(1.15, 6.13)	0.022
	ENG c.207G>A	689	1.10	(0.70, 1.72)	0.682	181	1.29	(0.57, 2.93)	0.537	209	0.41	(0.12, 1.40)	0.155
Brain VM	ACVRL1 c.314-35A>G	716	1.11	(0.68, 1.80)	0.675	189	1.03	(0.33, 3.16)	0960	211	0.74	(0.36, 1.52)	0.411
	ENG c.207G>A	716	1.19	(0.73, 1.93)	0.482	189	3.09	(0.63, 15.1)	0.163	211	0.62	(0.29, 1.32)	0.215
Pulmonary AVM	<i>ACVRL1</i> c.314-35A>G	690	1.48	(0.90, 2.22)	0.062	183	0.57	(0.25, 1.33)	0.192	208	2.45	(1.18, 5.06)	0.016
	ENG c.207G>A	690	1.29	(0.86, 1.93)	0.220	183	0.91	(0.36, 2.27)	0.835	208	1.80	(0.86, 3.77)	0.122
Liver VM	<i>ACVRL1</i> c.314-35A>G	677	1.46	(0.85, 2.50)	0.170	182	1.57	(0.65, 3.81)	0.320	203	n/a*	$(3.10,\infty)^*$	$0.001^{*}$
	ENG c.207G>A	677	1.05	(0.62, 1.79)	0.856	182	1.17	(0.44, 3.10)	0.758	203	0.55	(0.16, 1.85)	0.332
Results from multiva significance threshold	riable logistic regression and 1 (0.025) are in bold.	lysis fo	r carrie	rs of the risk ge	enotypes	adjuste	d for ag	e, sex and fam	ily cluster	ring (ex	cept*, s	ee below) are	shown. P va

is meeting the statistical

no estimated family effect; hence, the model was not adjusted for familial clustering. VM, any HHT organ vascular malformation including: brain VM, pulmonary arteriovenous malformation (AVM), liver confidence interval (CI) based on the profile likelihood and a p-value from the likelihood ratio test comparing models with and without the polymorphism. A mixed-effects logistic regression model found \* The odds ratio (OR) cannot be determined, because 100% (21/21) of liver VM-positive patients carry the A risk allele, compared to 75% of liver VM-negative patients. We show a one-sided 95% VM; SNP, single nucleotide polymorphism.