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Journal

Microbiology Resource Announcements, 4(1)

ISSN

2576-098X

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Publication Date

2016-02-25

DOI

10.1128/genomea.00010-16

Peer reviewed

Draft Genome Sequences of Two *Pseudoalteromonas* Strains Isolated from Roots and Leaf Blades of the Seagrass *Zostera marina*

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Here, we present the draft genome sequences for *Pseudoalteromonas* sp. strain UCD-33C and *Pseudoalteromonas lipolytica* UCD-48B. *Pseudoalteromonas* sp. UCD-33C was isolated from *Zostera marina* roots and *P. lipolytica* UCD-48B from *Z. marina* leaf blades, both collected in Woods Hole, MA. These assemblies contain 4,479,285 bp and 4,592,435 bp, respectively.

Received 4 January 2016 Accepted 5 January 2016 Published 18 February 2016

Citation Alexiev A, Krusor ML, Jospin G, Lang JM, Eisen JA, Coil DA. 2016. Draft genome sequences of two *Pseudoalteromonas* strains isolated from roots and leaf blades of the seagrass *Zostera marina*. *Genome Announc* 4(1):e00010-16. doi:10.1128/genomeA.00010-16.

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Pseudoalteromonas lipolytica was first isolated from Yangtze River estuary seawater. It is Gram-negative, motile, and strictly aerobic, has rod-shaped cells, and produces exopolysaccharides (1). Some *Pseudoalteromonas* strains exhibit antimicrobial abilities that are inhibitory to cystic fibrosis-associated opportunistic pathogens (2). *Pseudoalteromonas* sp. strain UCD-33C and *P. lipolytica* UCD-48B were both isolated from seagrass (*Zostera marina*) collected in Woods Hole, MA. *Pseudoalteromonas* sp. UCD-33C came from roots, whereas strain UCD-48B was isolated from leaf blades. This culturing project was done as part of a collaboration between researchers at the University of California, Davis, CA, and University of Oregon, Eugene, OR, called The Seagrass Microbiome Project (<http://www.seagrassmicrobiome.org>). The project seeks to characterize and analyze the microbial communities living in and on seagrasses.

Bacterial isolates were grown and double-dilution struck on Luria broth (LB) agar (Difco), seawater agar (SWA), 10% diluted seawater agar (SW10), and *Azotobacter* isolation medium agar (NFM). The isolates were incubated at 25°C for 1 to 21 days. Scrapings were then frozen in 25% glycerol for long-term storage. The isolates were later thawed and grown in seawater nutrient agar medium (ATCC medium 2205, using Instant Ocean instead of synthetic seawater). DNA was subsequently extracted from a fresh overnight culture using the Wizard genomic DNA purification kit (Promega).

A paired-end library was produced using a Nextera DNA sample prep kit (Illumina) and sequenced on an Illumina HiSeq (250 bp paired-end reads). Sequencing of *Pseudoalteromonas* sp. UCD-33C resulted in 807,945 reads and approximately 90× coverage. The genome size was 4,479,285 bp, and the G+C content was 41.3%. Sequencing of *P. lipolytica* UCD-48B yielded 885,488 reads and approximately 96× coverage. Its genome size was 4,592,435 bp and had 41.4% G+C content. The sequences were processed using the A5-miseq assembly pipeline (3, 4), which automates error correction, data cleaning, contig assembly, scaffolding, and quality control. The completeness of the genome was assessed using PhyloSift (5), which utilizes a list of 37 highly con-

served single-copy marker genes (6). One copy of each marker gene was found in the sequences. Automated annotation was done using the RAST annotation server (7). A combination of BLAST and phylogenetic trees using the full-length assembled 16S rRNA sequences revealed strain UCD-48B to belong to *P. lipolytica*. However, the placement of the UCD-33C strain was ambiguous, falling into a polyphyletic and poorly resolved group, making it impossible to determine a species without further work.

Nucleotide sequence accession numbers. The genome sequence for *Pseudoalteromonas* sp. UCD-33C has been deposited at DDBJ/EMBL/GenBank under the accession no. [LJTB00000000](https://www.ncbi.nlm.nih.gov/nuclink/LJTB00000000). The version described in this paper is no. [LJTB00000000.1](https://www.ncbi.nlm.nih.gov/nuclink/LJTB00000000.1). The genome sequence for *P. lipolytica* UCD-48B has been deposited at DDBJ/EMBL/GenBank under the accession no. [LJTC00000000](https://www.ncbi.nlm.nih.gov/nuclink/LJTC00000000). The version described in this paper is no. [LJTC00000000.1](https://www.ncbi.nlm.nih.gov/nuclink/LJTC00000000.1).

ACKNOWLEDGMENTS

Bacterial strains were isolated as part of the 2014 Microbial Diversity course at the Marine Biological Laboratory in Woods Hole, MA. Illumina sequencing was performed at the DNA Technologies Core Facility in the Genome Center at University of California, Davis, CA. This work was funded by a grant from the Gordon and Betty Moore Foundation (grant GBMF333), “Investigating the co-evolutionary relationships between seagrasses and their microbial symbionts.”

We thank Colleen Cavanaugh for her assistance with seagrass sampling and John Zhang for help with library preparation.

FUNDING INFORMATION

The Gordon and Betty Moore Foundation provided funding to Jenna M. Lang and Jonathan Eisen.

REFERENCES

- Xu XW, Wu YH, Wang CS, Gao XH, Wang XG, Wu M. 2010. *Pseudoalteromonas lipolytica* sp. nov., isolated from the Yangtze River estuary. *Int J Syst Evol Microbiol* 60:2176–2181. <http://dx.doi.org/10.1099/ijs.0.017673-0>.
- Maida I, Bosi E, Fondi M, Perrin E, Orlandini V, Papaleo MC, Mengoni A, de Pascale D, Tutino ML, Michaud L, Lo Giudice A, Fani R. 2015. Antimicrobial activity of *Pseudoalteromonas* strains isolated from the Ross

- Sea (Antarctica) versus cystic fibrosis opportunistic pathogens. *Hydrobiologia* 761:443–457. <http://dx.doi.org/10.1007/s10750-015-2190-8>.
3. Tritt A, Eisen JA, Facciotti MT, Darling AE. 2012. An integrated pipeline for *de novo* assembly of microbial genomes. *PLoS One* 7:e42304. <http://dx.doi.org/10.1371/journal.pone.0042304>.
 4. Coil D, Jospin G, Darling AE. 2015. A5-miseq: an updated pipeline to assemble microbial genomes from Illumina MiSeq data. *Bioinformatics* 31:587–589. <http://dx.doi.org/10.1093/bioinformatics/btu661>.
 5. Darling AE, Jospin G, Lowe E, Matsen FA, Bik HM, Eisen JA. 2014. PhyloSift: phylogenetic analysis of genomes and metagenomes. *PeerJ* 2:e243. <http://dx.doi.org/10.7717/peerj.243>.
 6. Wu D, Jospin G, Eisen JA. 2013. Systematic identification of gene families for use as “markers” for phylogenetic and phylogeny-driven ecological studies of bacteria and archaea and their major subgroups. *PLoS One* 8:e77033. <http://dx.doi.org/10.1371/journal.pone.0077033>.
 7. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:. <http://dx.doi.org/10.1186/1471-2164-9-75>.