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Title

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Permalink https://escholarship.org/uc/item/4599155k

Journal Expert Opinion on Pharmacotherapy, 16(5)

ISSN 1465-6566

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Publication Date 2015-03-24

DOI

10.1517/14656566.2015.998650

Peer reviewed



HHS Public Access

Expert Opin Pharmacother. Author manuscript; available in PMC 2015 October 02.

Published in final edited form as:

Author manuscript

Expert Opin Pharmacother. 2015 April; 16(5): 633-643. doi:10.1517/14656566.2015.998650.

Advances in pharmacotherapy for primary biliary cirrhosis

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Abstract

Introduction—Primary Biliary Cirrhosis (PBC) is a chronic autoimmune liver disease mostly seen in middle aged women characterized by progressive non-suppurative destruction of small bile ducts resulting in intrahepatic cholestasis, parenchymal injury, and ultimately end stage liver disease. Despite major breakthroughs in our understanding of PBC, there remains only one FDA-approved agent for treatment: ursodeoxycholic acid (UDCA) to which one third of patients are unresponsive.

Areas covered—Biochemical response to treatment with UDCA is associated with excellent survival rates in PBC patients. However, there is a need for alternative treatments for non-responders. Results from human epidemiological and genetic studies as well as preclinical studies in PBC animal models have provided a strong impetus for the development of new therapeutic agents. In this review, we discuss the recent advances in translational research in PBC focusing on promising therapeutic approaches, namely immune-based targeted therapies and agents targeting the synthesis and circulation of bile acids.

Expert opinion—We are in a new era for the development of novel therapies for PBC. Data on fibrates, budesonide, and obsticholic acid offer encouragement for non-responders to UDCA.

Keywords

Primary Biliary Cirrhosis; ursodeoxycholic acid; FXR agonists; biologics

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The contribution represents original work that has not been previously published or simultaneously submitted for publication elsewhere.

The manuscript has been read and approved by all the authors, and all the conditions as previously stated by the ICMJE have been met.

The body providing explicit ethical approval of the work reported is stated.

The authors' preference for publication is US spellings.

The authors have enrolled patients in some cited trials (NCT01473524, NCT00746486, NCT01389973, NCT01430429)

1.0 INTRODUCTION

In 1851, Addison and Gull described three patients with light skin lesions, jaundice and enlarged livers; one of these patients later developed yellowish plaques around her eyes and finger joints. A century later, Ahrens and his colleagues coined the term Primary Biliary Cirrhosis (PBC) to describe jaundice, hepatomegaly and pruritus affecting middle aged women, in the absence of apparent obstruction of the large bile ducts [1, 2].

Since then, our understanding of PBC has grown considerably: anti-mitochondrial antibodies (AMA), which are present in 80% or more of PBC patients have been identified and the autoantigens recognized by them have been cloned [3, 4]. The major targets of AMAs, the E2 component of mitochondrial dehydrogenases, particularly the pyruvate dehydrogenase complex (PDC-E2), have been rigorously defined at the molecular level [3, 5, 6]. Moreover, autoreactive CD4+ and CD8+ T cell responses present in human peripheral blood and liver have been dissected [7–10]. Epidemiological and genetic studies, including Genome Wide Association Studies (GWAS), have delineated several crucial environmental factors and cellular pathways involved in the pathophysiology of the disease [11, 12].

There have been enormous advances in understanding the pathophysiology of PBC, including cloning of the major mitochondrial autoantigen, development of animal models, and refinement in dissection of effector pathways [13–18].

Despite the great progress made in understanding the disease, a major breakthrough in treatment has yet to be realized [19]. The only FDA-approved agent for the treatment of PBC is still ursodeoxycholic acid (UDCA), which was introduced in 1987. A complete biochemical response to UDCA is associated with both improved survival and liver histology [20-22]. However, while UDCA therapy has a marked impact on clinical outcomes in PBC, up to 40% of patients have an insufficient response to UDCA and accordingly have a significantly increased risk of developing an adverse outcome, such as liver transplantation or death [21, 23]. Several studies have proposed various criteria as predictors of treatment success with UDCA [20, 21, 23-27]. Recent studies of large patient cohorts from France and the United Kingdom have demonstrated that reduction of the alkaline phosphatase (ALP) and aspartate transaminase (AST) to < 1.5 times the upper limit of normal (ULN) and a normal total bilirubin after 1 year of UDCA therapy (Paris II criteria) is associated with significantly better transplant-free survival [28]. Numerous therapies including newer biologics have been studied in the one third of PBC patients who are incomplete responders to UDCA with little success. Importantly, an increased dose of UDCA in patients who had incomplete response to UDCA was not found to be of any benefit in this subpopulation of patients [21].

In this review we will summarize recent advances in therapeutics and potential imminent breakthroughs that may accelerate the development of therapies in PBC. We will focus both on bile acid-based therapies and the new therapeutic approaches originated from the discoveries of immune pathways in PBC. Indeed, while in the incipient stages of the disease, the immunological breach of tolerance is most significant, in later stages, cholestasis plays an important role in the recurring hepatic injury where the prime targets for novel

therapeutics focus on limiting the cytotoxic effects of bile acids at the level of the liver parenchyma [19, 29, 30].

2.0 PAST AND PRESENT: BILE ACID SIGNALING

In cholestatic conditions including PBC, the hydrophobic bile acids accumulate within the liver parenchyma contributing to hepatic injury. Multiple mechanisms of bile acid injury include: direct cytotoxic effects due to cell membrane solubilization, bile-induced reactive oxygen species (ROS), recruitment of mononuclear cells through the release of chemoattractant cytokines, upregulation of the expression of MHC class I on hepatocytes (rendering them more vulnerable to immune destruction), and induction of hepatocyte apoptosis (by direct activation of Fas). When given at therapeutic doses, UDCA, a hydrophilic bile acid occurring naturally in humans normally constituting less than 5% of the bile acid pool changes the composition of this pool by increasing the fraction UDCA. Experimental evidence suggests that UDCA protects cholangiocytes against cytotoxicity of hydrophobic bile acids, resulting from changes in the phospholipid-rich micelles, reduction of bile acid cytotoxicity of bile and decrease of the concentration of hydrophobic bile acids [31]. In addition, UDCA upregulates the anion exchanger 2 (AE2) transporter involved in biliary bicarbonate secretion resulting in less toxic bile [32]. Further, UDCA helps to reinstate antioxidant defenses mediated by glutathione-related upregulation of y-glutamyl cysteine synthetase [33]. Other possible mechanisms of action of UDCA include its inhibition of apoptosis [34] and reported immunomodulatory effects that can be partially accounted for by its effect on the glucocorticoid receptor [35].

UDCA is currently the only drug approved for the treatment of patients with PBC. Several clinical studies support the assertion that UDCA not only improves biochemical indices, but also delays histologic progression and improves survival without transplantation. In fact, the transplant-free survival of UDCA treated PBC patients who respond to UDCA is comparable to that of the general population [21, 36]. Indeed both AASLD and EASL guidelines recommend the use of UDCA in PBC patients [37, 38]. Yet, one third of PBC patients do not respond to UDCA [21] and the efficacy of UDCA in terms of overall survival and progression to end stage liver disease is still unclear [39].

2.1 Bile acids as nuclear receptor ligands in the liver cell signaling

In the last two decades, endogenous bile acids have been identified as natural ligands for previously identified nuclear receptors. These receptors include the farnesoid X receptor (FXR), the pregnane X receptor (PXR), the constitutive androstane receptor (CAR) and the vitamin D3 receptor (VDR). As nuclear receptors, they translocate to the cell nucleus upon activation, bind to hormone response elements (HSR) and control the expression of certain genes. Once activated by bile acids, FXR translocates to the cell membrane, dimerizes with the retinoid X receptor (RXR) and induces the expression of small heterodimer partner (SHP) which regulates *de novo* bile acid synthesis and transport by inhibiting the expression of the CYP7A1 gene [40] and increasing the expression of multidrug resistance-associated protein 3 (MDR-3) [41]. Eventually, FXR is involved in the regulation of hepatic bile acid production and flow and the modulation of hepatic inflammation, fibrosis and regeneration (Figure 1). FXR is also an important regulator of the enterohepatic circulation of bile acids

at the level of hepatocytes and enterocytes [42]. FXR activation in hepatocytes downregulates bile acid uptake [43] and indirectly stimulates bile salt export pump (BSEP, also known as ABCB11) [44]. In enterocytes, FXR induces the release of FGF-19, an ileal hormone that is released into the portal circulation and ultimately regulates bile acid synthesis, acting through the FGFR4/Klotho- β receptor complexes in the liver to inhibit CYP7A1 [45]. FXR is also an important inhibitor of hepatic inflammation, as evident by the spontaneous liver injury, inflammation and increased sensitivity to NF- κ B activation seen in in FXR-deficient mice (Figure 1) [46].

Another recently discovered bile acid receptor is the transmembrane G protein–coupled receptor (TGR-5) expressed on macrophages, biliary epithelial cells (BEC), sinusoidal endothelial cells, gallbladder epithelium, and in brown adipose tissue and muscle. Activation of TGR-5 by bile acids result in inhibition of the NF- κ B-mediated expression of proinflammatory genes in macrophages and Kupffer cells in hepatic tissue [47]. Some evidence also suggests a protective role against liver carcinogenesis through the negative regulation of STAT3 [48]. The favorable effects of bile acid receptor activation have made these receptors attractive targets for drug development over the last decade.

2.2 Bile Acid Pharmacotherapies

Obeticholic acid (OCA) (6-alpha-ethyl-chenodeoxycholic acid) (INT-747) is a semisynthetic analogue of chenodeoxycholic acid with a 100-fold higher affinity for FXR [49]. OCA is notable for decreasing bile synthesis by directly suppressing CYP7A1 and inducing FGF-19, FGF-15 in mice, release to promote bile excretion by acting on various bile transporters [50]. OCA also may ameliorate portal hypertension through an increase in eNOS production, modulate liver regeneration and have anti-fibrotic effects [51]. FXR activation by OCA also has important anti-inflammatory effects evident by reduced cytokine production [52]. Two Phase 2 studies of OCA in PBC have been completed and the 1-year double blind, placebo controlled phase of a Phase 3 trial (POISE) evaluating OCA for the treatment of PBC patients with an incomplete biochemical response to UDCA has concluded recently (NCT01473524). Response rates (intended as alkaline phosphatase reduction, a surrogate marker of damage progression [27]) to OCA or placebo given in addition to UDCA in the Phase 3 trial were 10% with placebo compared to 47% with 10 mg OCA and 46% with 5 mg OCA with titration to 10 mg OCA (both OCA groups p < 0.0001vs. placebo). The placebo group experienced a mean decrease in ALP from baseline of 5%, compared to a mean decrease of 39% in the 10 mg OCA dose group and 33% in the 5–10 mg OCA titration group (both OCA groups p < 0.0001 vs. placebo). Pruritus is the most severe and common side effect reported during the trial (up to 70% of patients). It is dosedependent and has led to therapy discontinuation at high-dosage of OCA, but seems to be well tolerated at the lowest effective dosage. The POISE study is currently in a long-term safety extension phase [53].

Another steroidal semi-synthetic bile acid analogue INT-767 is a dual FXR and TGR5 agonist [54]. It has been shown to modulate the activity of monocytes and macrophages [55], to reduce inflammation though the inactivation of NF- κ B via a protein kinase A dependent manner, and to reduce liver injury by promoting biliary bicarbonate excretion

[56]. A TGR-5 selective agonist, INT-777 has been shown to increase bile flow in animal models [57]. Regulating bile acid synthesis is also being attempted through a recombinant variant of FGF-19 (NGM-282). As noted above, FGF-19 plays an important role in suppressing bile synthesis and promoting hepatocyte proliferation [58]. A phase 2 clinical trial to evaluate the use of NGM-282 in PBC patients is currently ongoing (NCT02026401).

Finally, inhibition of the Ileal Bile Acid Transporter (IBAT, or ASBT) may result in increased excretion of bile. Indeed, ASBT inhibitors have been developed but, as far as we know, they have not yet been tested in cholestatic liver diseases (Table 1).

2.3 Fibrates

Fibrates are carboxylic acids that have been used as hypolipidemic agents for decades. They exert their effect by acting on nuclear transcription factors of the peroxisome proliferatoractivated receptors (PPARs) family. Several mechanisms of action suggest that fibrates might be beneficial in PBC. Mediated via PPARa, fibrates upregulate MDR-3 and FXR, therefore may inhibit bile salt synthesis, reduce IL-1-induced C-reactive protein expression on hepatocytes, and inhibit NF- κ B through the induction of I κ B α expression. Through their activity on PPAR δ , fibrates activate PGC-1 α , which increases FXR activity. Fibrates also up-regulate bile acid efflux transporters and the ileal bile acid binding protein (I-BABP). Finally, it has recently been suggested that fibrates may act as a dual PPAR and PXR agonist. In the last two decades, several clinical trials have shown that fibrates have anticholestatic effects accompanied by decreases in inflammatory markers and relief from itching when combined with UDCA in PBC patients who were refractory to UDCA therapy [59-61]. Two different fibrates have been tested in PBC, Bezafibrate, a pan-PPAR isoform agonist, and fenofibrate which is a PPARa specific agonist; whereas Bezafibrate is approved in Europe and Japan, only fenofibrate is available in the USA. Although the results of these studies seem consistent, most of them included only a small number of patients [62]. The results of phase III studies are currently awaited.

3.0 NEAR FUTURE: THE IMMUNE NETWORK

The serologic hallmark of PBC is the presence of antibodies to mitochondria, i.e. AMA, especially to PDC-E2. PBC is considered a model autoimmune disease because of the homogeneity between patients and the high specificity of AMAs. However, over the past decade there has been great progress on defining the multi-lineage response to PDC-E2, including immunological definition of the antigenic epitopes, the nature of reactive autoantibodies, the characterization of T-cell responses [7–9], and the crucial role of the innate immune system [29]. Moreover pathway-based analysis of GWAS on PBC has confirmed that key immune mechanisms such as TNF signaling and antigen processing and presentation may underlie the genetic predisposition known to exist in PBC [63]. The effector mechanisms of PBC are indeed a multi-orchestrated response [64], and a better understanding of effector mechanisms will suggest new approaches to clinical intervention.

3.1 IL-12 and IL-23 axis

Interleukin-12 (IL-12) is a heterodimer composed of two subunits: IL-12p35 and IL-12p40 subunits. It is produced by antigen presenting cells (APCs) and drives the differentiation of T lymphocytes into a pro-inflammatory T helper (Th1) phenotype [30]. The importance of the IL-12/Th1 pathway in PBC has been suggested by a mouse model of PBC [65] and human genetic studies, which have identified predisposing polymorphism associated with genes downstream of the IL-12/Th1 signaling cascade [66, 67]. Interleukin-23 (IL-23) is composed of an IL-12p40 subunit that is shared with IL-12 and an IL-23p19 subunit. IL-23 drives the development of Th17 cells [68, 69] and is implicated in a number of autoimmune diseases, including PBC [70, 71]. While both Th1 and Th17 play cardinal roles in PBC pathogenesis, advanced disease stages are characterized by Th17 skewing emphasizing the potential importance of targeting the IL-23/Th17 axis [17].

The monoclonal antibody ustekinumab targets the IL-12p40 and therefore exerts its effect on both the IL-12/Th1 as well as the IL-23/Th17 axes. Ustekinumab is of therapeutic benefit in psoriasis and in trials for Crohn's disease. A phase II trial, aiming to test the efficacy and safety of ustekinumab in PBC patients (NCT01389973) showed a modest decrease in ALP, enhanced liver fibrosis (ELF) score and bile concentration, none of the patients achieved the predefined primary endpoint of ALP reduction from baseline of >40% or ALP normalization [72]. Given the strong evidence of an essential role for IL12 and IL23 in PBC [30], studies with agents specifically targeting either IL-12 or IL-23 may have different outcomes.

3.2 CTLA-4 Agents

Under physiological conditions, T-cell activation requires not only engagement of the T cell receptor with it cognate antigen presented by an MHC, but also a second signal from costimulatory molecules, including the cytotoxic T lymphocyte antigen 4 (CTLA-4) on T cells. In addition to MHC-peptide complexes, APCs express membrane proteins (CD80 and CD86) that bind to the CD28 co-receptor expressed on T-cells and produce a co-stimulatory signal further enhancing their activation. When CD80/86 interact with CTLA-4 on T cells instead of CD28, they convey an inhibitory signal to the T-cell. Importantly, CTLA-4 binds to CD80/86 with greater affinity than CD28 limiting T cell activation. As a consequence, cytokine production is reduced, cell cycle progression inhibited, TCR signaling down-modulated, and activation of B cells and macrophages decreased. Moreover, CTLA-4 is crucial for the function of Foxp3⁺ T regulatory cells (Tregs), which mediate immune tolerance.

These properties of CTLA-4 as well as preclinical studies in mouse models of PBC [73] suggest a strong rationale for CTLA4-based therapy in PBC. Two chimeric CTLA-4-Ig proteins have been approved in recent years; abatacept for the treatment of rheumatoid arthritis and belatacept for the prevention of acute rejection in kidney transplant patients. This has triggered [73]a Phase 2 study to evaluate the use of abatacept in PBC patients with an incomplete biochemical response to UDCA (NCT02078882).

3.3 CD40-CD40L Signaling

CD40 is expressed on all APCs, binds to its natural ligand CD40L, which is expressed primarily on activated CD4+ T cells, and after cell activation is up-regulated [74]. Moreover, CD40 is constitutively expressed by B cells and its interaction with CD40L is critical for immunoglobulin (Ig) class-switching [75]. Dysregulation of CD40-CD40L has been documented in various autoimmune diseases [76]. In the context of PBC, CD40 activation induces Fas/FasL-mediated apoptosis of biliary epithelial cells (BEC) [77] and recent findings suggest that epigenetic changes in the CD40L could account for a defect in class switching resulting in the high titers of IgM seen in PBC patients [78]. Agents against CD40L are a promising means to target autoreactive T-cells [79], but, have only been tested in animal models. Interestingly, anti-CD40L has been shown to decrease activated CD8+ T-cells and hepatic NK-T cells and to ameliorate inflammation and bile duct destruction in an animal model of autoimmune cholangitis [80].

3.4 CXCL-10: T-Cell Recruitment

The CXC motif chemokine 10 (CXCL10), or Interferon-γ-inducible protein-10 (IP-10), plays a role in the recruitment of T-cells during biliary injury [81] and demonstrated to be involved in the pathogenesis of PBC [82]. Unfortunately, a trial targeting CXCL-10 with anti-CXCL10 (NI-0801) in PBC patients was not able to demonstrate a clinical efficacy (NCT01430429).

3.5 Regulatory T-Cell Activity

Regulatory T cells (Tregs) have been long known to mediate tolerance and different subtypes of Tregs have been implicated in the pathogenesis of PBC, including CD8+ CD28– Tregs [83] and the more studied CD4+ CD25+ Tregs [84]. Moreover, CD25 (IL-2R α) deficient mice develop PBC-like features [85] and a child with congenital IL-2R α (CD25) deficiency presented PBC-like liver disease and AMAs [86]. Low dose IL-2 therapy might have beneficial effects on the Treg population in a variety of conditions [87, 88] and successful induction of remission in a patient with systemic lupus erythematosus has been reported. A clinical trial is currently underway to study the efficacy of low-dose IL-2 therapy in a host of autoimmune diseases (NCT01988506). Cell therapy is an alternative approach to reinstitute Treg activity. In the child with IL-2R α deficiency, allogenic stem cell transplantation resolved his PBC [86]. In addition, adoptive transfer of wild type Tregs can reduce inflammatory cytokine production and prevent disease in a mouse model of PBC [89, 90].

3.6 B-cells

Although T-lymphocytes are considered the main mediators of tissue damage in PBC, the role of B cells is strongly suggested by the presence of highly specific AMA in more than 90% of patients, and the high titers of IgM that are frequently seen in the sera of patients [2, 78]. However, it is important to highlight that while the production of antibodies has an important diagnostic value, it does not correlate with disease severity [91]. However, in addition to antibody production, antigen presentation by B-cells plays an important role in T-cell mediated autoimmunity. Moreover, B-cells secret cytokines including interferon- γ

and IL-4 and have a suppressive role on Tregs. Indeed, the beneficial effect of B-cell depletion in T cell mediated autoimmune diseases has been attributed to the attenuated T-cell activation, and the effect this depletion has on Treg cells.

Depletion of B-cells with anti-CD20 therapy has yielded some positive results in murine models [92] as well as in PBC patients, using Rituximab, with suboptimal response to UDCA [93] with significant improvement seen in both biochemical and immunologic markers (NCT00364819). Yet, depletion of B-cells can be a double-edged sword. Pretreatment of mice with anti-CD20 prior to induction of a PBC-like illness with xenobiotic immunization exacerbates the disease [92]. In addition, in a genetic model of PBC expressing a dominant negative TGF- β receptor II transgene, B-cell deficiency through genetic manipulation leads to a more severe liver inflammation [94]. A plausible explanation for these seemingly contradictory results may lay in the different subsets of B-cells, i.e. reactive B cells versus regulatory B cells, and in the role that each of these subsets play at different stages of disease progression. While B effector cells seem to contribute to disease progression, regulatory B cells may be protective against disease initiation, including IL-10 producing B-cells [95].

3.7 Budesonide And Immunosuppressive Agents

Although combination of UDCA with a glucocorticoid might have more favorable results compared to UDCA monotherapy [96], initial trials with prednisolone resulted in a high incidence of adverse effects, especially loss of bone density, that precluded its use [97, 98]. As a result, clinical trials of budesonide, a glucocorticoid with a higher receptor affinity and a higher first-pass metabolism, have been conducted and shown to improve both biochemical and histological markers [99, 100]. Yet, in late stage PBC budesonide results in high serum levels that are associated with serious adverse effects [101]. A phase 3 trial is currently underway for the evaluation of budesonide in UDCA-refractory PBC patients (ClinicalTrials.gov Identifier: NCT00746486).

Other immunosuppressive agents assessed for PBC treatment including methotrexate [102] and azathioprine, which is currently recommended only in the overlap syndrome of PBC with autoimmune hepatitis [103, 104]. Mycophenolate mofetil, an inhibitor of T- and B- cells proliferation, has also been evaluated with conflicting results [105]. The cumulative body of evidence is currently insufficient to support the use of any of these agents in PBC patients.

Several other potential therapies are not discussed herein as the data is too preliminary and/or confidential. We should note that Moexipril, an angiotensin-converting enzyme inhibitor, has been studied in conjunction with UDCA but demonstrated no beneficial effect [106]. Combivir, an antiretroviral agent, is under investigation in PBC, but its rationale and any conclusive data remain elusive [107]. Colchicine and methotrexate have a long history in the treatment of PBC [108–110], but their mechanisms of action and their roles, if any, are unclear. Finally, other drugs like tetrathiomolybdate [111], and the use of mesenchymal stem cells have been studied but again require more detailed analyses and there is insufficient data for discussion [112]. It is also important to note that PBC is associated with

systemic symptoms, including pruritus, fatigue, and bone density impairment, all of whom still lack specific treatments [113].

4.0 EXPERT OPINION

Since the FDA approval of UDCA over twenty years ago, there was little to be offered to PBC patients beyond the conventional UDCA therapy. For those patients that do not respond to UDCA there are currently no options to delay the progression of PBC to liver failure, death, or liver transplant. Interestingly, although PBC has a strong autoimmune pathogenesis, immunosuppressive drugs have not shown a beneficial effect. We hypothesize that such drugs would need to be utilized during the earliest phases of disease development, at the time when tolerance is broken. However, patients present with clinical symptoms several or more years after detection of autoantibodies and thus it has been difficult to treat patients during the first phases of disease. On the other hand, and during the last decade, there have been significant advances in our understanding of the pathogenesis and immunobiology of PBC. Epidemiological studies, GWAS and preclinical testing of new agents using PBC animal models have led to the development of new agents and a great number of clinical trials in PBC over recent years. Indeed the clinical trials presented in this review herald a new era of specifically targeted therapies that are based on sound understanding of the immunological and bile acid-related mechanisms that drive disease initiation and progression (Table 1). Clearly, the challenge is the design of high quality clinical trials, with molecules chosen on the basis of structure function and mechanisms of action if we are to develop successful new therapeutics. It is no trivial matter to obtain approval for an orphan disease like PBC and particularly a disease which progresses over years to decades. There is required persistence and a balance between the ideal and the practical.

Acknowledgments

Supported in part by National Institutes of Health grant DK39588.

References

- Ahrens EH Jr, Payne MA, Kunkel HG, Eisenmenger WJ, Blondheim SH. Primary biliary cirrhosis. Medicine (Baltimore). 1950 Dec; 29(4):299–364. [PubMed: 14796348]
- 2*. Kaplan MM, Gershwin ME. Primary biliary cirrhosis. The New England journal of medicine. 2005 Sep 22; 353(12):1261–73. Structural description of PBC. [PubMed: 16177252]
- Dubel L, Tanaka A, Leung PS, Van de Water J, Coppel R, Roche T, et al. Autoepitope mapping and reactivity of autoantibodies to the dihydrolipoamide dehydrogenase-binding protein (E3BP) and the glycine cleavage proteins in primary biliary cirrhosis. Hepatology. 1999 Apr; 29(4):1013–8. [PubMed: 10094940]
- 4. Van de Water J, Fregeau D, Davis P, Ansari A, Danner D, Leung P, et al. Autoantibodies of primary biliary cirrhosis recognize dihydrolipoamide acetyltransferase and inhibit enzyme function. J Immunol. 1988 Oct 1; 141(7):2321–4. [PubMed: 3049806]
- Leung PS, Chuang DT, Wynn RM, Cha S, Danner DJ, Ansari A, et al. Autoantibodies to BCOADC-E2 in patients with primary biliary cirrhosis recognize a conformational epitope. Hepatology. 1995 Aug; 22(2):505–13. [PubMed: 7543435]

- Ichiki Y, Selmi C, Shimoda S, Ishibashi H, Gordon SC, Gershwin ME. Mitochondrial antigens as targets of cellular and humoral auto-immunity in primary biliary cirrhosis. Clin Rev Allergy Immunol. 2005 Apr; 28(2):83–91. [PubMed: 15879615]
- Kita H, Matsumura S, He XS, Ansari AA, Lian ZX, Van de Water J, et al. Quantitative and functional analysis of PDC-E2-specific autoreactive cytotoxic T lymphocytes in primary biliary cirrhosis. J Clin Invest. 2002 May; 109(9):1231–40. [PubMed: 11994412]
- Kita H, Naidenko OV, Kronenberg M, Ansari AA, Rogers P, He XS, et al. Quantitation and phenotypic analysis of natural killer T cells in primary biliary cirrhosis using a human CD1d tetramer. Gastroenterology. 2002 Oct; 123(4):1031–43. [PubMed: 12360465]
- Shimoda S, Van de Water J, Ansari A, Nakamura M, Ishibashi H, Coppel RL, et al. Identification and precursor frequency analysis of a common T cell epitope motif in mitochondrial autoantigens in primary biliary cirrhosis. J Clin Invest. 1998 Nov 15; 102(10):1831–40. [PubMed: 9819369]
- Shimoda S, Harada K, Niiro H, Yoshizumi T, Soejima Y, Taketomi A, et al. Biliary epithelial cells and primary biliary cirrhosis: the role of liver-infiltrating mononuclear cells. Hepatology. 2008 Mar; 47(3):958–65. [PubMed: 18181218]
- Podda M, Selmi C, Lleo A, Moroni L, Invernizzi P. The limitations and hidden gems of the epidemiology of primary biliary cirrhosis. J Autoimmun. 2013 Oct.46:81–7. [PubMed: 23871640]
- Gershwin ME, Selmi C, Worman HJ, Gold EB, Watnik M, Utts J, et al. Risk factors and comorbidities in primary biliary cirrhosis: a controlled interview-based study of 1032 patients. Hepatology. 2005 Nov; 42(5):1194–202. [PubMed: 16250040]
- Gershwin ME, Mackay IR, Sturgess A, Coppel RL. Identification and specificity of a cDNA encoding the 70 kd mitochondrial antigen recognized in primary biliary cirrhosis. J Immunol. 1987 May 15; 138(10):3525–31. [PubMed: 3571977]
- 14. Zhang J, Zhang W, Leung PS, Bowlus CL, Dhaliwal S, Coppel RL, et al. Ongoing activation of autoantigen-specific B cells in primary biliary cirrhosis. Hepatology. 2014 Jul 12.
- Lleo A, Zhang W, McDonald WH, Seeley EH, Leung PS, Coppel RL, et al. Shotgun proteomics: Identification of unique protein profiles of apoptotic bodies from biliary epithelial cells. Hepatology. 2014 Oct; 60(4):1314–23. [PubMed: 24841946]
- Huang W, Kachapati K, Adams D, Wu Y, Leung PS, Yang GX, et al. Murine autoimmune cholangitis requires two hits: cytotoxic KLRG1(+) CD8 effector cells and defective T regulatory cells. J Autoimmun. 2014 May.50:123–34. [PubMed: 24556277]
- Yang CY, Ma X, Tsuneyama K, Huang S, Takahashi T, Chalasani NP, et al. IL-12/Th1 and IL-23/ Th17 biliary microenvironment in primary biliary cirrhosis: implications for therapy. Hepatology. 2014 May; 59(5):1944–53. [PubMed: 24375552]
- Hudspeth K, Pontarini E, Tentorio P, Cimino M, Donadon M, Torzilli G, et al. The role of natural killer cells in autoimmune liver disease: a comprehensive review. J Autoimmun. 2013 Oct.46:55– 65. [PubMed: 23880068]
- 19**. Gershwin ME, Mackay IR. The causes of primary biliary cirrhosis: Convenient and inconvenient truths. Hepatology. 2008 Feb; 47(2):737–45. an interesting summary of critical aspects of PBC pathogenesis. [PubMed: 18098322]
- 20. Kuiper EM, Hansen BE, de Vries RA, den Ouden-Muller JW, van Ditzhuijsen TJ, Haagsma EB, et al. Improved prognosis of patients with primary biliary cirrhosis that have a biochemical response to ursodeoxycholic acid. Gastroenterology. 2009 Apr; 136(4):1281–7. [PubMed: 19208346]
- 21**. Pares A, Caballeria L, Rodes J. Excellent long-term survival in patients with primary biliary cirrhosis and biochemical response to ursodeoxycholic Acid. Gastroenterology. 2006 Mar; 130(3):715–20. this paper demonstrated that UDCA responders have an excellent prognosis. [PubMed: 16530513]
- 22. Zein CO, Angulo P, Lindor KD. When is liver biopsy needed in the diagnosis of primary biliary cirrhosis? Clin Gastroenterol Hepatol. 2003 Mar; 1(2):89–95. [PubMed: 15017500]
- Corpechot C, Abenavoli L, Rabahi N, Chretien Y, Andreani T, Johanet C, et al. Biochemical response to ursodeoxycholic acid and long-term prognosis in primary biliary cirrhosis. Hepatology. 2008 Sep; 48(3):871–7. [PubMed: 18752324]

- Kumagi T, Guindi M, Fischer SE, Arenovich T, Abdalian R, Coltescu C, et al. Baseline ductopenia and treatment response predict long-term histological progression in primary biliary cirrhosis. Am J Gastroenterol. 2010 Oct; 105(10):2186–94. [PubMed: 20502446]
- 25. Azemoto N, Kumagi T, Abe M, Konishi I, Matsuura B, Hiasa Y, et al. Biochemical response to ursodeoxycholic acid predicts long-term outcome in Japanese patients with primary biliary cirrhosis. Hepatol Res. 2011 Apr; 41(4):310–7. [PubMed: 21426448]
- Zhang LN, Shi TY, Shi XH, Wang L, Yang YJ, Liu B, et al. Early biochemical response to ursodeoxycholic acid and long-term prognosis of primary biliary cirrhosis: results of a 14-year cohort study. Hepatology. 2013 Jul; 58(1):264–72. [PubMed: 23408380]
- 27**. Lammers WJ, van Buuren HR, Hirschfield GM, Janssen HL, Invernizzi P, Mason AL, et al. Levels of Alkaline Phosphatase and Bilirubin Are Surrogate End Points of Outcomes of Patients With Primary Biliary Cirrhosis: An International Follow-up Study. Gastroenterology. 2014 Aug 23. Alkaline phosphatase levels are a good surrogate marker of outcome in PBC.
- 28. Carbone M, Mells GF, Pells G, Dawwas MF, Newton JL, Heneghan MA, et al. Sex and age are determinants of the clinical phenotype of primary biliary cirrhosis and response to ursodeoxycholic acid. Gastroenterology. 2013 Mar; 144(3):560–69. e7. quiz e13–4. [PubMed: 23246637]
- Lleo A, Bowlus CL, Yang GX, Invernizzi P, Podda M, Van de Water J, et al. Biliary apotopes and anti-mitochondrial antibodies activate innate immune responses in primary biliary cirrhosis. Hepatology. 2010 Sep; 52(3):987–98. [PubMed: 20568301]
- 30*. Lleo A, Gershwin ME, Mantovani A, Invernizzi P. Towards common denominators in primary biliary cirrhosis: the role of IL-12. Journal of hepatology. 2012 Mar; 56(3):731–3. This paper helps to understand the role of IL-12 pathways in PBC. [PubMed: 22005588]
- 31. Paumgartner G, Beuers U. Ursodeoxycholic acid in cholestatic liver disease: mechanisms of action and therapeutic use revisited. Hepatology. 2002 Sep; 36(3):525–31. [PubMed: 12198643]
- Arenas F, Hervias I, Uriz M, Joplin R, Prieto J, Medina JF. Combination of ursodeoxycholic acid and glucocorticoids upregulates the AE2 alternate promoter in human liver cells. J Clin Invest. 2008 Feb; 118(2):695–709. [PubMed: 18188457]
- Serviddio G, Pereda J, Pallardo FV, Carretero J, Borras C, Cutrin J, et al. Ursodeoxycholic acid protects against secondary biliary cirrhosis in rats by preventing mitochondrial oxidative stress. Hepatology. 2004 Mar; 39(3):711–20. [PubMed: 14999689]
- Rodrigues CM, Fan G, Ma X, Kren BT, Steer CJ. A novel role for ursodeoxycholic acid in inhibiting apoptosis by modulating mitochondrial membrane perturbation. J Clin Invest. 1998 Jun 15; 101(12):2790–9. [PubMed: 9637713]
- 35. Miura T, Ouchida R, Yoshikawa N, Okamoto K, Makino Y, Nakamura T, et al. Functional modulation of the glucocorticoid receptor and suppression of NF-kappaB-dependent transcription by ursodeoxycholic acid. The Journal of biological chemistry. 2001 Dec 14; 276(50):47371–8. [PubMed: 11577102]
- 36. Invernizzi P, Setchell KD, Crosignani A, Battezzati PM, Larghi A, O'Connell NC, et al. Differences in the metabolism and disposition of ursodeoxycholic acid and of its taurineconjugated species in patients with primary biliary cirrhosis. Hepatology. 1999 Feb; 29(2):320–7. [PubMed: 9918905]
- Lindor KD, Gershwin ME, Poupon R, Kaplan M, Bergasa NV, Heathcote EJ, et al. Primary biliary cirrhosis. Hepatology. 2009 Jul; 50(1):291–308. [PubMed: 19554543]
- European Association for the Study of the L. EASL Clinical Practice Guidelines: management of cholestatic liver diseases. Journal of hepatology. 2009 Aug; 51(2):237–67. [PubMed: 19501929]
- Gong Y, Huang Z, Christensen E, Gluud C. Ursodeoxycholic acid for patients with primary biliary cirrhosis: an updated systematic review and meta-analysis of randomized clinical trials using Bayesian approach as sensitivity analyses. Am J Gastroenterol. 2007 Aug; 102(8):1799–807. [PubMed: 17459023]
- Goodwin B, Jones SA, Price RR, Watson MA, McKee DD, Moore LB, et al. A regulatory cascade of the nuclear receptors FXR, SHP-1, and LRH-1 represses bile acid biosynthesis. Mol Cell. 2000 Sep; 6(3):517–26. [PubMed: 11030332]

- 41. Huang L, Zhao A, Lew JL, Zhang T, Hrywna Y, Thompson JR, et al. Farnesoid X receptor activates transcription of the phospholipid pump MDR3. The Journal of biological chemistry. 2003 Dec 19; 278(51):51085–90. [PubMed: 14527955]
- Boyer JL, Trauner M, Mennone A, Soroka CJ, Cai SY, Moustafa T, et al. Upregulation of a basolateral FXR-dependent bile acid efflux transporter OSTalpha-OSTbeta in cholestasis in humans and rodents. Am J Physiol Gastrointest Liver Physiol. 2006 Jun; 290(6):G1124–30. [PubMed: 16423920]
- Denson LA, Sturm E, Echevarria W, Zimmerman TL, Makishima M, Mangelsdorf DJ, et al. The orphan nuclear receptor, shp, mediates bile acid-induced inhibition of the rat bile acid transporter, ntcp. Gastroenterology. 2001 Jul; 121(1):140–7. [PubMed: 11438503]
- 44. Ananthanarayanan M, Balasubramanian N, Makishima M, Mangelsdorf DJ, Suchy FJ. Human bile salt export pump promoter is transactivated by the farnesoid X receptor/bile acid receptor. The Journal of biological chemistry. 2001 Aug 3; 276(31):28857–65. [PubMed: 11387316]
- 45. Kurosu H, Choi M, Ogawa Y, Dickson AS, Goetz R, Eliseenkova AV, et al. Tissue-specific expression of betaKlotho and fibroblast growth factor (FGF) receptor isoforms determines metabolic activity of FGF19 and FGF21. The Journal of biological chemistry. 2007 Sep 14; 282(37):26687–95. [PubMed: 17623664]
- 46. Wang YD, Chen WD, Wang M, Yu D, Forman BM, Huang W. Farnesoid X receptor antagonizes nuclear factor kappaB in hepatic inflammatory response. Hepatology. 2008 Nov; 48(5):1632–43. [PubMed: 18972444]
- 47. Wang YD, Chen WD, Yu D, Forman BM, Huang W. The G-protein-coupled bile acid receptor, Gpbar1 (TGR5), negatively regulates hepatic inflammatory response through antagonizing nuclear factor kappa light-chain enhancer of activated B cells (NF-kappaB) in mice. Hepatology. 2011 Oct; 54(4):1421–32. [PubMed: 21735468]
- Chen WD, Yu D, Forman BM, Huang W, Wang YD. Deficiency of G-protein-coupled bile acid receptor Gpbar1 (TGR5) enhances chemically induced liver carcinogenesis. Hepatology. 2013 Feb; 57(2):656–66. [PubMed: 22911633]
- 49. Pellicciari R, Fiorucci S, Camaioni E, Clerici C, Costantino G, Maloney PR, et al. 6alpha-ethylchenodeoxycholic acid (6-ECDCA), a potent and selective FXR agonist endowed with anticholestatic activity. J Med Chem. 2002 Aug 15; 45(17):3569–72. [PubMed: 12166927]
- Song KH, Li T, Owsley E, Strom S, Chiang JY. Bile acids activate fibroblast growth factor 19 signaling in human hepatocytes to inhibit cholesterol 7alpha-hydroxylase gene expression. Hepatology. 2009 Jan; 49(1):297–305. [PubMed: 19085950]
- 51*. Fiorucci S, Distrutti E, Ricci P, Giuliano V, Donini A, Baldelli F. Targeting FXR in cholestasis: hype or hope. Expert Opin Ther Targets. 2014 Sep.9:1–11. This paper clarifes FXR role in cholestatic diseases.
- 52. Gadaleta RM, van Erpecum KJ, Oldenburg B, Willemsen EC, Renooij W, Murzilli S, et al. Farnesoid X receptor activation inhibits inflammation and preserves the intestinal barrier in inflammatory bowel disease. Gut. 2011 Apr; 60(4):463–72. [PubMed: 21242261]
- Beuers U. Drug insight: Mechanisms and sites of action of ursodeoxycholic acid in cholestasis. Nat Clin Pract Gastroenterol Hepatol. 2006 Jun; 3(6):318–28. [PubMed: 16741551]
- 54. Rizzo G, Passeri D, De Franco F, Ciaccioli G, Donadio L, Rizzo G, et al. Functional characterization of the semisynthetic bile acid derivative INT-767, a dual farnesoid X receptor and TGR5 agonist. Mol Pharmacol. 2010 Oct; 78(4):617–30. [PubMed: 20631053]
- 55. Miyazaki-Anzai S, Masuda M, Levi M, Keenan AL, Miyazaki M. Dual Activation of the Bile Acid Nuclear Receptor FXR and G-Protein-Coupled Receptor TGR5 Protects Mice against Atherosclerosis. PLoS One. 2014; 9(9):e108270. [PubMed: 25237811]
- 56. Baghdasaryan A, Claudel T, Gumhold J, Silbert D, Adorini L, Roda A, et al. Dual farnesoid X receptor/TGR5 agonist INT-767 reduces liver injury in the Mdr2–/– (Abcb4–/–) mouse cholangiopathy model by promoting biliary HCO(–)(3) output. Hepatology. 2011 Oct; 54(4): 1303–12. [PubMed: 22006858]
- 57. Pellicciari R, Gioiello A, Macchiarulo A, Thomas C, Rosatelli E, Natalini B, et al. Discovery of 6alpha-ethyl-23(S)-methylcholic acid (S-EMCA, INT-777) as a potent and selective agonist for

the TGR5 receptor, a novel target for diabesity. J Med Chem. 2009 Dec 24; 52(24):7958–61. [PubMed: 20014870]

- Wu X, Ge H, Lemon B, Vonderfecht S, Weiszmann J, Hecht R, et al. FGF19-induced hepatocyte proliferation is mediated through FGFR4 activation. The Journal of biological chemistry. 2010 Feb 19; 285(8):5165–70. [PubMed: 20018895]
- Honda A, Ikegami T, Nakamuta M, Miyazaki T, Iwamoto J, Hirayama T, et al. Anticholestatic effects of bezafibrate in patients with primary biliary cirrhosis treated with ursodeoxycholic acid. Hepatology. 2013 May; 57(5):1931–41. [PubMed: 22911624]
- Lens S, Leoz M, Nazal L, Bruguera M, Pares A. Bezafibrate normalizes alkaline phosphatase in primary biliary cirrhosis patients with incomplete response to ursodeoxycholic acid. Liver Int. 2014 Feb; 34(2):197–203. [PubMed: 23998489]
- 61. Levy C, Peter JA, Nelson DR, Keach J, Petz J, Cabrera R, et al. Pilot study: fenofibrate for patients with primary biliary cirrhosis and an incomplete response to ursodeoxycholic acid. Aliment Pharmacol Ther. 2011 Jan; 33(2):235–42. [PubMed: 21083674]
- 62*. Ghonem NS, Boyer JL. Fibrates as adjuvant therapy for chronic cholestatic liver disease: its time has come. Hepatology. 2013 May; 57(5):1691–3. An interesting comment on the perspectives offered by fibrates. [PubMed: 23174993]
- Kar SP, Seldin MF, Chen W, Lu E, Hirschfield GM, Invernizzi P, et al. Pathway-based analysis of primary biliary cirrhosis genome-wide association studies. Genes Immun. 2013 Apr; 14(3):179– 86. [PubMed: 23392275]
- 64. Gershwin ME, Ansari AA, Mackay IR, Nakanuma Y, Nishio A, Rowley MJ, et al. Primary biliary cirrhosis: an orchestrated immune response against epithelial cells. Immunol Rev. 2000 Apr. 174:210–25. [PubMed: 10807518]
- 65. Yoshida K, Yang GX, Zhang W, Tsuda M, Tsuneyama K, Moritoki Y, et al. Deletion of interleukin-12p40 suppresses autoimmune cholangitis in dominant negative transforming growth factor beta receptor type II mice. Hepatology. 2009 Nov; 50(5):1494–500. [PubMed: 19676134]
- 66**. Hirschfield GM, Liu X, Xu C, Lu Y, Xie G, Lu Y, et al. Primary biliary cirrhosis associated with HLA, IL12A, and IL12RB2 variants. The New England journal of medicine. 2009 Jun 11; 360(24):2544–55. GWAS study in PBC patients. [PubMed: 19458352]
- 67*. Liu X, Invernizzi P, Lu Y, Kosoy R, Lu Y, Bianchi I, et al. Genome-wide meta-analyses identify three loci associated with primary biliary cirrhosis. Nat Genet. 2010 Aug; 42(8):658–60. GWAS study in PBC patients. [PubMed: 20639880]
- Cua DJ, Sherlock J, Chen Y, Murphy CA, Joyce B, Seymour B, et al. Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain. Nature. 2003 Feb 13; 421(6924):744–8. [PubMed: 12610626]
- 69. Aggarwal S, Ghilardi N, Xie MH, de Sauvage FJ, Gurney AL. Interleukin-23 promotes a distinct CD4 T cell activation state characterized by the production of interleukin-17. The Journal of biological chemistry. 2003 Jan 17; 278(3):1910–4. [PubMed: 12417590]
- Lan RY, Salunga TL, Tsuneyama K, Lian ZX, Yang GX, Hsu W, et al. Hepatic IL-17 responses in human and murine primary biliary cirrhosis. J Autoimmun. 2009 Feb; 32(1):43–51. [PubMed: 19101114]
- 71. Rong G, Zhou Y, Xiong Y, Zhou L, Geng H, Jiang T, et al. Imbalance between T helper type 17 and T regulatory cells in patients with primary biliary cirrhosis: the serum cytokine profile and peripheral cell population. Clin Exp Immunol. 2009 May; 156(2):217–25. [PubMed: 19302244]
- 72. Hirschfield GM. P367 phase 2 study evaluating the efficacy and safety of Ustekinumab in patients with primary biliary cirrhosis who had an inadequate response to ursodeoxycholic acid. Journal of hepatology. 2014; 60(1):S189–S90.
- Dhirapong A, Yang GX, Nadler S, Zhang W, Tsuneyama K, Leung P, et al. Therapeutic effect of cytotoxic T lymphocyte antigen 4/immunoglobulin on a murine model of primary biliary cirrhosis. Hepatology. 2013 Feb; 57(2):708–15. [PubMed: 22996325]
- 74. Schoenberger SP, Toes RE, van der Voort EI, Offringa R, Melief CJ. T-cell help for cytotoxic T lymphocytes is mediated by CD40-CD40L interactions. Nature. 1998 Jun 4; 393(6684):480–3. [PubMed: 9624005]

- Pessach IM, Notarangelo LD. X-linked primary immunodeficiencies as a bridge to better understanding X-chromosome related autoimmunity. J Autoimmun. 2009 Aug; 33(1):17–24. [PubMed: 19361956]
- 76. Goules A, Tzioufas AG, Manousakis MN, Kirou KA, Crow MK, Routsias JG. Elevated levels of soluble CD40 ligand (sCD40L) in serum of patients with systemic autoimmune diseases. J Autoimmun. 2006 May; 26(3):165–71. [PubMed: 16621447]
- 77. Afford SC, Ahmed-Choudhury J, Randhawa S, Russell C, Youster J, Crosby HA, et al. CD40 activation-induced, Fas-dependent apoptosis and NF-kappaB/AP-1 signaling in human intrahepatic biliary epithelial cells. FASEB J. 2001 Nov; 15(13):2345–54. [PubMed: 11689460]
- Lleo A, Liao J, Invernizzi P, Zhao M, Bernuzzi F, Ma L, et al. Immunoglobulin M levels inversely correlate with CD40 ligand promoter methylation in patients with primary biliary cirrhosis. Hepatology. 2012 Jan; 55(1):153–60. [PubMed: 21898485]
- 79. Law CL, Grewal IS. Therapeutic interventions targeting CD40L (CD154) and CD40: the opportunities and challenges. Adv Exp Med Biol. 2009; 647:8–36. [PubMed: 19760064]
- Tanaka H, Yang GX, Iwakoshi N, Knechtle SJ, Kawata K, Tsuneyama K, et al. Anti-CD40 ligand monoclonal antibody delays the progression of murine autoimmune cholangitis. Clin Exp Immunol. 2013 Dec; 174(3):364–71. [PubMed: 23981074]
- Borchers AT, Shimoda S, Bowlus C, Keen CL, Gershwin ME. Lymphocyte recruitment and homing to the liver in primary biliary cirrhosis and primary sclerosing cholangitis. Semin Immunopathol. 2009 Sep; 31(3):309–22. [PubMed: 19533132]
- Chuang YH, Lian ZX, Cheng CM, Lan RY, Yang GX, Moritoki Y, et al. Increased levels of chemokine receptor CXCR3 and chemokines IP-10 and MIG in patients with primary biliary cirrhosis and their first degree relatives. J Autoimmun. 2005 Sep; 25(2):126–32. [PubMed: 16243485]
- Bernuzzi F, Fenoglio D, Battaglia F, Fravega M, Gershwin ME, Indiveri F, et al. Phenotypical and functional alterations of CD8 regulatory T cells in primary biliary cirrhosis. J Autoimmun. 2010 Nov; 35(3):176–80. [PubMed: 20638239]
- Lan RY, Cheng C, Lian ZX, Tsuneyama K, Yang GX, Moritoki Y, et al. Liver-targeted and peripheral blood alterations of regulatory T cells in primary biliary cirrhosis. Hepatology. 2006 Apr; 43(4):729–37. [PubMed: 16557534]
- Wakabayashi K, Lian ZX, Moritoki Y, Lan RY, Tsuneyama K, Chuang YH, et al. IL-2 receptor alpha(-/-) mice and the development of primary biliary cirrhosis. Hepatology. 2006 Nov; 44(5): 1240–9. [PubMed: 17058261]
- 86. Aoki CA, Roifman CM, Lian ZX, Bowlus CL, Norman GL, Shoenfeld Y, et al. IL-2 receptor alpha deficiency and features of primary biliary cirrhosis. J Autoimmun. 2006 Aug; 27(1):50–3. [PubMed: 16904870]
- Saadoun D, Rosenzwajg M, Joly F, Six A, Carrat F, Thibault V, et al. Regulatory T-cell responses to low-dose interleukin-2 in HCV-induced vasculitis. The New England journal of medicine. 2011 Dec 1; 365(22):2067–77. [PubMed: 22129253]
- Matsuoka K, Koreth J, Kim HT, Bascug G, McDonough S, Kawano Y, et al. Low-dose interleukin-2 therapy restores regulatory T cell homeostasis in patients with chronic graft-versushost disease. Sci Transl Med. 2013 Apr 3.5(179):179ra43.
- Tanaka H, Zhang W, Yang GX, Ando Y, Tomiyama T, Tsuneyama K, et al. Successful immunotherapy of autoimmune cholangitis by adoptive transfer of forkhead box protein 3(+) regulatory T cells. Clin Exp Immunol. 2014 Nov; 178(2):253–61. [PubMed: 25041369]
- Jeffery LE, Burke F, Mura M, Zheng Y, Qureshi OS, Hewison M, et al. 1,25-Dihydroxyvitamin D3 and IL-2 combine to inhibit T cell production of inflammatory cytokines and promote development of regulatory T cells expressing CTLA-4 and FoxP3. J Immunol. 2009 Nov 1; 183(9):5458–67. [PubMed: 19843932]
- Invernizzi P, Crosignani A, Battezzati PM, Covini G, De Valle G, Larghi A, et al. Comparison of the clinical features and clinical course of antimitochondrial antibody-positive and -negative primary biliary cirrhosis. Hepatology. 1997 May; 25(5):1090–5. [PubMed: 9141422]

- 92. Dhirapong A, Lleo A, Yang GX, Tsuneyama K, Dunn R, Kehry M, et al. B cell depletion therapy exacerbates murine primary biliary cirrhosis. Hepatology. 2011 Feb; 53(2):527–35. [PubMed: 21274873]
- 93. Tsuda M, Moritoki Y, Lian ZX, Zhang W, Yoshida K, Wakabayashi K, et al. Biochemical and immunologic effects of rituximab in patients with primary biliary cirrhosis and an incomplete response to ursodeoxycholic acid. Hepatology. 2012 Feb; 55(2):512–21. [PubMed: 22006563]
- 94. Moritoki Y, Lian ZX, Lindor K, Tuscano J, Tsuneyama K, Zhang W, et al. B-cell depletion with anti-CD20 ameliorates autoimmune cholangitis but exacerbates colitis in transforming growth factor-beta receptor II dominant negative mice. Hepatology. 2009 Dec; 50(6):1893–903. [PubMed: 19877182]
- 95. Wang H, Shin DM, Abbasi S, Jain S, Kovalchuk AL, Beaty N, et al. Expression of plasma cell alloantigen 1 defines layered development of B-1a B-cell subsets with distinct innate-like functions. Proc Natl Acad Sci U S A. 2012 Dec 4; 109(49):20077–82. [PubMed: 23169635]
- 96. Leuschner M, Guldutuna S, You T, Hubner K, Bhatti S, Leuschner U. Ursodeoxycholic acid and prednisolone versus ursodeoxycholic acid and placebo in the treatment of early stages of primary biliary cirrhosis. Journal of hepatology. 1996 Jul; 25(1):49–57. [PubMed: 8836901]
- 97. Mitchison HC, Bassendine MF, Malcolm AJ, Watson AJ, Record CO, James OF. A pilot, doubleblind, controlled 1-year trial of prednisolone treatment in primary biliary cirrhosis: hepatic improvement but greater bone loss. Hepatology. 1989 Oct; 10(4):420–9. [PubMed: 2777203]
- Wolfhagen FH, van Hoogstraten HJ, van Buuren HR, van Berge-Henegouwen GP, ten Kate FJ, Hop WC, et al. Triple therapy with ursodeoxycholic acid, prednisone and azathioprine in primary biliary cirrhosis: a 1-year randomized, placebo-controlled study. Journal of hepatology. 1998 Nov; 29(5):736–42. [PubMed: 9833911]
- Leuschner M, Maier KP, Schlichting J, Strahl S, Herrmann G, Dahm HH, et al. Oral budesonide and ursodeoxycholic acid for treatment of primary biliary cirrhosis: results of a prospective double-blind trial. Gastroenterology. 1999 Oct; 117(4):918–25. [PubMed: 10500075]
- 100. Rautiainen H, Karkkainen P, Karvonen AL, Nurmi H, Pikkarainen P, Nuutinen H, et al. Budesonide combined with UDCA to improve liver histology in primary biliary cirrhosis: a three-year randomized trial. Hepatology. 2005 Apr; 41(4):747–52. [PubMed: 15754377]
- 101. Hempfling W, Grunhage F, Dilger K, Reichel C, Beuers U, Sauerbruch T. Pharmacokinetics and pharmacodynamic action of budesonide in early- and late-stage primary biliary cirrhosis. Hepatology. 2003 Jul; 38(1):196–202. [PubMed: 12830002]
- 102. Combes B, Emerson SS, Flye NL, Munoz SJ, Luketic VA, Mayo MJ, et al. Methotrexate (MTX) plus ursodeoxycholic acid (UDCA) in the treatment of primary biliary cirrhosis. Hepatology. 2005 Nov; 42(5):1184–93. [PubMed: 16250039]
- 103. EASL Clinical Practice Guidelines: management of cholestatic liver diseases. Journal of hepatology. 2009 Aug; 51(2):237–67. [PubMed: 19501929]
- 104. Christensen E, Neuberger J, Crowe J, Altman DG, Popper H, Portmann B, et al. Beneficial effect of azathioprine and prediction of prognosis in primary biliary cirrhosis. Final results of an international trial. Gastroenterology. 1985 Nov; 89(5):1084–91. [PubMed: 3899841]
- 105. Treiber G, Malfertheiner P. Mycophenolate mofetil for the treatment of primary biliary cirrhosis in patients with an incomplete response to ursodeoxycholic acid. J Clin Gastroenterol. 2005 Oct; 39(9):837–8. author reply 38. [PubMed: 16145353]
- 106. Charatcharoenwitthaya P, Talwalkar JA, Angulo P, Gossard AA, Keach JC, Petz JL, et al. Moexipril for treatment of primary biliary cirrhosis in patients with an incomplete response to ursodeoxycholic acid. Digestive diseases and sciences. 2010 Feb; 55(2):476–83. [PubMed: 19255851]
- 107. Mason AL, Wasilenko ST. Other potential medical therapies: the use of antiviral agents to investigate and treat primary ciliary cirrhosis. Clinics in liver disease. 2008 May; 12(2):445–60. xi. [PubMed: 18456190]
- 108. Kaplan MM, Alling DW, Zimmerman HJ, Wolfe HJ, Sepersky RA, Hirsch GS, et al. A prospective trial of colchicine for primary biliary cirrhosis. The New England journal of medicine. 1986 Dec 4; 315(23):1448–54. [PubMed: 3537784]

- 109. Kaplan MM, Schmid C, Provenzale D, Sharma A, Dickstein G, McKusick A. A prospective trial of colchicine and methotrexate in the treatment of primary biliary cirrhosis. Gastroenterology. 1999 Nov; 117(5):1173–80. [PubMed: 10535881]
- Bonis PA, Kaplan M. Methotrexate improves biochemical tests in patients with primary biliary cirrhosis who respond incompletely to ursodiol. Gastroenterology. 1999 Aug; 117(2):395–9. [PubMed: 10419921]
- 111. Askari F, Innis D, Dick RB, Hou G, Marrero J, Greenson J, et al. Treatment of primary biliary cirrhosis with tetrathiomolybdate: results of a double-blind trial. Translational research: the journal of laboratory and clinical medicine. 2010 Mar; 155(3):123–30. [PubMed: 20171597]
- 112. Wang L, Han Q, Chen H, Wang K, Shan GL, Kong F, et al. Allogeneic bone marrow mesenchymal stem cell transplantation in patients with UDCA-resistant primary biliary cirrhosis. Stem cells and development. 2014 Oct 15; 23(20):2482–9. [PubMed: 24835895]
- 113. Eaton JE, Talwalkar JA, Lazaridis KN, Gores GJ, Lindor KD. Pathogenesis of primary sclerosing cholangitis and advances in diagnosis and management. Gastroenterology. 2013 Sep; 145(3): 521–36. [PubMed: 23827861]

HIGHLIGHTS BOX

- For decades UDCA has been the only approved therapy for PBC, and although responders to UDCA have a good prognosis and survival rate, 30% of patients are still in need of therapy.
- Epidemiological studies, GWAS have identify specific molecular pathways that are potential therapeutic targets; preclinical testing of new agents using PBC animal models have led to the development of new agents and clinical trials in PBC. We are in a new era for the therapy of this chronic liver disease.
- Obeticholic acid (OCA) is a semi-synthetic analogue of chenodeoxycholic acid with a strong affinity for farnesoid X receptor (FXR). Response rates to OCA, given in addition to UDCA, in a Phase 3 trial seem are promising.
- Several mechanisms of action suggest that fibrates might be beneficial in PBC. However most studies include only a small number of patients and more data are needed.
- The effector mechanisms of PBC are a multi-orchestrated response involving both innate and adaptive immunity. A number of biologics targeting specific immune pathways are being tested in PBC, aimed at regulating pathogenic mechanisms involved in disease perpetuation.

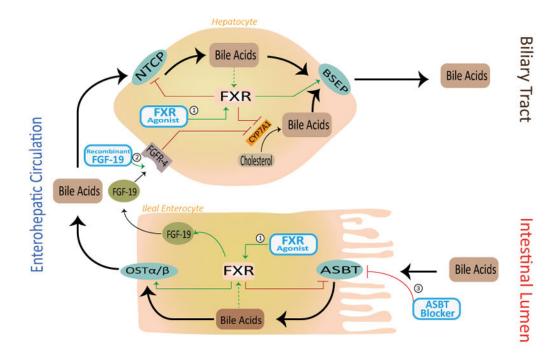


Figure 1. Bile acid based new therapeutics for PBC

(1) FXR-agonists exert their effect both on enterocytes and hepatocytes, inhibiting de novo synthesis of bile acids and promoting their excretion. (2) Recombinant FGF-19 interacts with the FGFR-4 receptor expressed on hepatocytes to inhibit bile acid synthesis. (3) ASBT blocker interferes with the reabsorption of bile acids from the intestinal lumen. Dashed green line: not all bile acids can activate the FXR receptor (see text). ASBT: Apical Sodium-dependent Bile acid Transporter. Osta/ β : Organic Solute Transporter α and β . NTCP: Na+ - Taurocholate Cotransporting Polypeptide. BSEP: Bile Salt Export Pump. FXR: Farnesoid X Receptor. FGF-19: Fibroblast Growth Factor-19. FGFR-4: Fibroblast Growth Factor Receptor-4.

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Beyond UDCA – F	Beyond UDCA – Future Therapeutic Approaches for PBC	BC			
Treatment Approach	Mechanism	Class of molecule	Rationale	Typical Molecule(s)	References
	Targeting B-cells	Anti-CD20	The role of B-cells in PBC is not reduced to antibody production. B-cells can also act as antigen presenting cells, secrete important cytokines (including IL-4 and $\mathrm{IFN}\gamma$) and suppress Tregs.	Rituximab	NCT00364819*
		Anti-IL.12/IL23	Th1/IL-12 and Th17/IL-23 axis involvement in PBC has been confirmed by GWAS. Later stages are marked by Th17 skewing.	Ustekinumab	NCT01389973*
	Targeting Autoreactive T-Cells	Chimeric CTLA4	Chimeric CTLA4 proteins bind to the the B7 molecules on T-cells and prevent their interaction with CD28; thus blocking the second stimulus.	Abatacept Betalecept	NCT02078882*
Immunological		Anti-CD40	CD40 activation induce Fas/FasL-mediated apoptosis of biliary epithelial cells.	FFP104	NCT02193360*
		Anti-CXCL-10	CXCL-10 is important for the recruitment of T- cells during biliary injury.	NI-0801	NCT01430429*
	Enhancing Regulatory T-Cells	Low dose IL-2	Low dose IL-2 therapy might enhance Treg population. A clinical trial is currently underway to check the efficacy of low-dose IL-2 therapy in other autoimmune diseases (including Sclerosing Cholangitis). Therapeutic applicability to PBC is yet to be cleared.	Aldesleukin	NCT01988506*
		Cellular Therapy	Reinstitution of the Treg population might be possible with allogenic stem cell transplantation or adoptive transfer of wild type Tregs.		
		FXR-Agonists	Nuclear Receptor Agonists (of which the most studied are FXR-agonists) feed on the regulatory mechanisms for bile acid synthesis and circulation, decreasing de novo synthesis and promoting excretion.	OCA, INT767, GW4064, WAY-362450, GSK2324, Fexaramine, PX-102	NCT01473524*
	Activating Bile Acid Regulatory Mechanisms	Agonists of Other Nuclear Receptor	Activation of other nuclear receptors might have similar effects to those of FXR activation.	Rifampin, Hypricum, Vitamin D3	
Bile Acid Based		TGR5 agonists	Activation of the transmembrane TGR5 has been shown to increase bile flow in different animal models.	INT767, INT777	
		Recombinant FGF-19	Feeding on the FGF19/FGFR4 regulatory axis inhibits bile acid synthesis	NGM-282	NCT02135536* NCT02026401*
	Direct bile acid transporter blockers	ASBT Inhibitor	Inhibition of the IIeal Bile Acid Transporter (IBAT, or ASBT) may result in increased excretion of bile.	A4250	
Other	Activation of Key Nuclear Transcription Factor (PPAR)	Fibrates	Fibrates upregulate the expression of MDR-3, have anti-inflammatory effects, may increase FXR	Bezafibrate, Fenofibrate	NCT01654731*

Treatment Approach	Mechanism	Class of molecule	Rationale	Typical Molecule(s)	References
			activity and have shown to upregulate different bile acid transporters. activity and have shown to upregulate different bile acid transporters.	bid transporters. Bid transporters.	
	Steroidal Anti-inflammatory Agents	Glucocorticoids	Budesonide is a potent anti-inflammatory agent and at the same time has high first pass metabolism.	Budesonide	NCT00746486*

* These can be looked up at: *ClinicalTrials.gov*