

UC Davis

UC Davis Previously Published Works

Title

Genome Sequence of the Sulfate-Reducing Thermophilic Bacterium
Thermodesulfovibrio yellowstonii Strain DSM 11347T (Phylum Nitrospirae)

Permalink

<https://escholarship.org/uc/item/45h7g5nc>

Journal

Microbiology Resource Announcements, 3(1)

ISSN

2576-098X

Authors

Bhatnagar, Srijak
Badger, Jonathan H
Madupu, Ramana
et al.

Publication Date

2015-02-26

DOI

10.1128/genomea.01489-14

Peer reviewed

Genome Sequence of the Sulfate-Reducing Thermophilic Bacterium *Thermodesulfovibrio yellowstonii* Strain DSM 11347^T (Phylum *Nitrospirae*)

Srijak Bhatnagar,^a Jonathan H. Badger,^b Ramana Madupu,^c Hoda M. Khouri,^{c*} Elizabeth M. O'Connor,^d Frank T. Robb,^d Naomi L. Ward,^e Jonathan A. Eisen^f

Microbiology Graduate Group, University of California Davis, Davis, California, USA^a; J. Craig Venter Institute, La Jolla, California, USA^b; J. Craig Venter Institute, Rockville, Maryland, USA^c; Institute of Marine and Environmental Technology, and Department of Microbiology and Immunology, University of Maryland, Baltimore, Maryland, USA^d; Department of Molecular Biology, University of Wyoming, Laramie, Wyoming, USA^e; Department of Evolution and Ecology, UC Davis Genome Center, Department of Medical Microbiology and Immunology, University of California Davis, Davis, California, USA^f

* Present address: Hoda M. Khouri, Independent Consultant, Bethesda, Maryland, USA.

Here, we present the complete 2,003,803-bp genome of a sulfate-reducing thermophilic bacterium, *Thermodesulfovibrio yellowstonii* strain DSM 11347^T.

Received 10 December 2014 Accepted 15 December 2014 Published 29 January 2015

Citation Bhatnagar S, Badger JH, Madupu R, Khouri HM, O'Connor EM, Robb FT, Ward NL, Eisen JA. 2015. Genome sequence of the sulfate-reducing thermophilic bacterium *Thermodesulfovibrio yellowstonii* strain DSM 11347^T (phylum *Nitrospirae*). *Genome Announc* 3(1):e01489-14. doi:10.1128/genomeA.01489-14.

Copyright © 2015 Bhatnagar et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Jonathan A. Eisen, jaeisen@ucdavis.edu.

Thermodesulfovibrio yellowstonii is a sulfate-reducing, strictly anaerobic bacterium first isolated from the Sedge Bay of Yellowstone Lake in Wyoming, USA. It is a Gram-negative bacterium with curved rod-shaped cells averaging 1.5 μm in length and 0.3 μm in width. It is a motile organism, propelled by a single polar flagellum. *T. yellowstonii* can use sulfate, thiosulfate, and sulfite as terminal electron acceptors (1). It grows between the temperature range of 40°C and 70°C, with optimal growth at 65°C. The genome of *T. yellowstonii* was sequenced as part of an “Assembling the Tree of Life” project at the Institute for Genomic Research (TIGR). At the time that the project started (2002), there were no genomes available from the phylum *Nitrospirae*, of which *T. yellowstonii* is a member.

The type strain of *T. yellowstonii* (DSM 11347^T) was obtained from the Leibniz Institute DSMZ-German Collection of Microorganisms and Cell Cultures and grown anaerobically at 65°C using DSMZ medium 749. DNA was obtained by solubilizing cells with *N*-lauryl sulfate and sodium dodecyl sulfate, followed by incubation with proteinase K. The lysate was extracted with tris-EDTA-saturated phenol and chloroform/isoamyl alcohol and was precipitated from the aqueous phase with 95% ethanol. It was resolubilized, incubated with DNase-free RNase and further purified by cesium-chloride gradient centrifugation, and visualized using 365 nm UV light (2). Pulse-field gel electrophoresis was used to confirm the size and uniformity of the DNA preparation. Genome sequencing was performed in the following way: small (2 to 3 kb), medium (4 to 5 kb), and large (8 to 10 kb) insert libraries were made and Sanger sequenced, and assemblies were generated as previously described (3–5); assemblies were edited and gaps were closed by clone walking and targeted PCR and sequencing. Finishing was completed by generating additional coverage in low-coverage regions, verification of repeats, and resolution of

ambiguities (6). The final assembly had $\sim 9\times$ coverage for the 2,003,803-bp genome and a GC content of 34.13%.

The origin of replication was identified using GC skew and colocalization of origin-associated genes (7). All the universal single-copy bacterial marker genes (8) were found in the sequenced genome using PhyloSift (9). The genome was annotated as previously described (10). Of the 2,084 putative genes that were identified in the genome, 2,029 were putative protein-coding sequences (CDS) and 54 were putative noncoding RNA genes (3 noncoding RNAs, 3 rRNAs, 1 transfer-messenger RNA, and 47 tRNAs). Additionally, CRISPRFinder (11) identified five CRISPR repeats in the genome.

Nucleotide sequence accession numbers. The genome sequence has been deposited at GenBank under the accession number CP001147 and has been curated by NCBI staff under the accession number NC_011296. The version NC_011296.1 described in this paper was last modified on 20 March 2014.

ACKNOWLEDGMENTS

Sanger sequencing was performed at the Institute for Genomic Research (TIGR), in Rockville, Maryland, USA. We thank many others who contributed to this project, including the IT, sequencing, finishing, and informatics groups at TIGR; Claire Fraser for general support; and the current members of the Eisen lab. We thank Shannon Smith, Grace Pai, and Nadia B. Fedorova for their contributions.

This work was funded by the National Science Foundation’s “Assembling the Tree of Life” grant (no. 0228651), which was overseen by Jonathan A. Eisen and Naomi L. Ward.

REFERENCES

1. Henry EA, Devereux R, Maki JS, Gilmour CC, Woese CR, Mandelco L, Schauder R, Remsen CC, Mitchell R. 1994. Characterization of a new thermophilic sulfate-reducing bacterium. *Arch Microbiol* 161:62–69. <http://dx.doi.org/10.1007/BF00248894>.

2. Charbonnier F, Forterre P. 1995. Purification of plasmids from thermophilic and hyperthermophilic archaea, p 87–90. In Robb FT, Place AR (ed), *Archaea: a laboratory manual—thermophiles*. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
3. Wu D, Daugherty SC, Van Aken SE, Pai GH, Watkins KL, Khouri H, Tallon LJ, Zaborsky JM, Dunbar HE, Tran PL, Moran NA, Eisen JA. 2006. Metabolic complementarity and genomics of the dual bacterial symbiosis of sharpshooters. *PLoS Biol* 4:e188. <http://dx.doi.org/10.1371/journal.pbio.0040188>.
4. Heidelberg JF, Seshadri R, Haveman SA, Hemme CL, Paulsen IT, Kolonay JF, Eisen JA, Ward N, Methe B, Brinkac LM, Daugherty SC, Deboy RT, Dodson RJ, Durkin AS, Madupu R, Nelson WC, Sullivan SA, Fouts D, Haft DH, Selengut J, Peterson JD, Davidsen TM, Zafar N, Zhou L, Radune D, Dimitrov G, Hance M, Tran K, Khouri H, Gill J, Utterback TR, Feldblyum TV, Wall JD, Voordouw G, Fraser CM. 2004. The genome sequence of the anaerobic, sulfate-reducing bacterium *Desulfovibrio vulgaris* Hildenborough. *Nat Biotechnol* 22:554–559. <http://dx.doi.org/10.1038/nbt959>.
5. Heidelberg JF, Paulsen IT, Nelson KE, Gaidos EJ, Nelson WC, Read TD, Eisen JA, Seshadri R, Ward N, Methe B, Clayton RA, Meyer T, Tsapin A, Scott J, Beanan M, Brinkac L, Daugherty S, DeBoy RT, Dodson RJ, Durkin AS, Haft DH, Kolonay JF, Madupu R, Peterson JD, Umayam LA, White O, Wolf AM, Vamathevan J, Weidman J, Impraim M, Lee K, Berry K, Lee C, Mueller J, Khouri H, Gill J, Utterback TR, McDonald LA, Feldblyum TV, Smith HO, Venter JC, Nealon KH, Fraser CM. 2002. Genome sequence of the dissimilatory metal ion-reducing bacterium *Shewanella oneidensis*. *Nat Biotechnol* 20:1118–1123. <http://dx.doi.org/10.1038/nbt749>.
6. Tettelin H, Radune D, Kasif S, Khouri H, Salzberg SL. 1999. Optimized multiplex PCR: efficiently closing a whole-genome shotgun sequencing project. *Genomics* 62:500–507. <http://dx.doi.org/10.1006/geno.1999.6048>.
7. Lobry JR. 1996. Asymmetric substitution patterns in the two DNA strands of bacteria. *Mol Biol Evol* 13:660–665. <http://dx.doi.org/10.1093/oxfordjournals.molbev.a025626>.
8. Wu D, Jospin G, Eisen JA. 2013. Systematic identification of gene families for use as “markers” for phylogenetic and phylogeny-driven ecological studies of bacteria and archaea and their major subgroups. *PLoS One* 8:e77033. <http://dx.doi.org/10.1371/journal.pone.0077033>.
9. Darling AE, Jospin G, Lowe E, Matsen FA, Bik HM, Eisen JA. 2014. PhyloSift: phylogenetic analysis of genomes and metagenomes. *PeerJ* 2:e243. <http://dx.doi.org/10.7717/peerj.243>.
10. Coil DA, Badger JH, Forberger HC, Riggs F, Madupu R, Fedorova N, Ward N, Robb FT, Eisen JA. 2014. Complete genome sequence of the extreme thermophile *Dictyoglomus thermophilum* H-6-12. *Genome Announc* 2(1):e00109-00114. <http://dx.doi.org/10.1128/genomeA.00109-14>.
11. Grissa I, Vergnaud G, Pourcel C. 2007. CRISPRFinder: a Web tool to identify clustered regularly interspaced short palindromic repeats. *Nucleic Acids Res* 35:W52–W57. <http://dx.doi.org/10.1093/nar/gkm360>.