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TGF- β mediated DNA methylation in prostate cancer

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Abstract: Almost all tumors harbor a defective negative feedback loop of signaling by transforming growth factor- β (TGF- β). Epigenetic mechanisms of gene regulation, including DNA methylation, are fundamental to normal cellular function and also play a major role in carcinogenesis. Recent evidence demonstrated that TGF- β signaling mediates cancer development and progression. Many key events in TGF- β signaling in cancer included auto-induction of TGF- β 1 and increased expression of DNA methyltransferases (DNMTs), suggesting that DNA methylation plays a significant role in cancer development and progression. In this review, we performed an extensive survey of the literature linking TGF- β signaling to DNA methylation in prostate cancer. It appeared that almost all DNA methylated genes detected in prostate cancer are directly or indirectly related to TGF- β signaling. This knowledge has provided a basis for our future directions of prostate cancer research and strategies for prevention and therapy for prostate cancer.

Keywords: TGF- β ; DNA methylation; prostate cancer; DNMT; Erk activation; tumor development and progression



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Introduction

The underlying mechanism promoting tumor progression has been elusive. Almost all tumors harbor a defective negative feedback loop of signaling by transforming growth factor- β (TGF- β). TGF- β signaling consists of Smad and non-Smad pathways (1). In advanced cancer cells, the non-Smad pathways predominate and progress leading to deregulated signaling cascades (2). This deregulation creates a unique TGF- β mediated tumor microenvironment that sets off a vicious cycle and promotes many of the hallmarks of tumor progression, including sustained angiogenesis, immune system evasion, proliferation, loss of the apoptotic response, epithelial-to-mesenchymal transition (EMT) and metastasis. These combined effects lead to uncontrolled

tumor growth and spread, for which we coin the term "TGF- β mediated vicious cycle in tumor progression". Recent evidence demonstrated that TGF- β mediates aggressive cancer including auto-induction of TGF- β 1 and increased expression of DNA methyltransferases (DNMTs) (2,3). This latter observation suggests that the expression of these methylated genes may be an important event in TGF- β mediated tumor progression.

DNA methylation in cancer

Epigenetic changes are characteristic of nearly all malignancies and include changes in DNA methylation, histone modification and altered expression of microRNAs. DNA methylation plays a critical role in cancer development

and progression. Alteration of DNA methylation patterns leads to deregulation of gene expression, in the absence of mutation. In the past few years, there has been an explosion in the number of publications in DNA methylation in all types of cancers (900 papers as of March 2012), including representative publications in prostate cancer (4-7), bladder cancer (8), renal cell carcinoma (9), breast cancer (10), lung cancer (11), ovarian cancer (12), oral cancer (13), pancreatic cancer (14), and other cancers. All tumors that have been examined show changes in DNA methylation, suggesting that this may represent a basic element of cancer biology, which has a significant impact on tumor pathology. Readers are referred to many excellent reviews on the biology of DNA methylation (15-17). This increased interest in the study of DNA methylation has created an opportunity for us to query the relationship between TGF- β signaling and DNA methylation in cancer, which has not been appreciated to date.

Biology of TGF- β signaling

TGF- β is a potent pleiotropic cytokine that regulates mammalian development, differentiation, and homeostasis in essentially all cell types and tissues. Its signaling is mediated through Smad and non-Smad pathways to regulate transcription, translation, microRNA biogenesis, protein synthesis and post-translational modifications (1,18,19). TGF- β binds to the type II TGF- β receptor (T β RII) which recruits and transphosphorylates the type I TGF- β receptor (T β RI) (20). The activated T β RI then phosphorylates Smad2 and Smad3 at the c-terminus. Activated Smad2/3 forms heterooligomers with Smad4 and migrates to the nucleus to regulate transcription. The Smad complexes interact with a myriad of transcriptional co-regulators and other factors to mediate target gene expression or repression (21,22). Smad2/3 also interacts with and regulates microRNA processing. TGF- β also signals through a number of non-Smad pathways, including m-TOR, RhoA, Ras, MAPK, PI3K/AKT, PP2A/p70s6K, and JNK (1,23,24). Finally, a direct action of the activated T β RI can interact with eEF1A1 to block protein synthesis (19). Dysregulation of both Smad and non-Smad pathways is implicated in aberrant TGF- β signaling and its pro-tumorigenic events in advanced cancer (3).

TGF- β signaling and DNA methylation

TGF- β is a key regulator for DNA methylation through an increase in DNMTs expression, especially in cancer (3,12).

There exists a differential effect of TGF- β mediated DNMT activities between benign and malignant cells. In benign cells, TGF- β inhibits DNMT expression (25,26). In cancer cells, TGF- β stimulates DNMT expression (3,12). It should be noted that, in light of the importance of both TGF- β signaling and DNA methylation in tumor progression, the majority of the methylated genes in cancer are relevant to TGF- β signaling (12). This is consistent with our observation that over-expression of TGF- β and/or DNMTs is associated with aggressiveness and poor prognosis in prostate cancer (3,27).

Review of literature

In this review, we will focus our discussion in prostate cancer as an example, because the pattern of DNA methylation is organ specific. We surveyed the recent literature to identify the existing methylated genes in prostate cancer and attempt to determine which ones are mediated by TGF- β signaling. We have identified over 80 genes in which promoters are methylated in prostate cancer. This is a significant increase from 2006, when only 30 genes had been identified (28). Interestingly, the non-Smad pathways of known relevance to TGF- β are more often associated with de novo gene methylation (3,29). In contrast, the Smad-mediated pathways often lead to promoter de-methylation of genes (see below). In *Table 1*, we summarize the known TGF- β relevant genes in which the promoter becomes methylated in prostate cancer. We also identified those which have been known to be induced by TGF- β . Since, in advanced cancer cells, TGF- β induces the activation of Erk, JNK, AKT, and NF- κ B (1,3), the above methylated gene have been documented in the literature to be related with one of the above transcription factors, thus are considered as TGF- β relevant.

In addition, there are a few genes that are de-methylated and are mediated through Smad2/3 activation, such as α 2 [1] collagen (113), CD133 (26), and maspin (or SFN, 14-3-3 sigma) (41,59,67,114,115). However, a reversal of the methylation status in these genes can be observed in cancer cells when the TGF- β signaling events switched from the Smad pathways to the non-Smad pathways in cancer cells as in the case for maspin (116) and CD133 (117).

Table 2 lists genes that are not currently documented in the literature as TGF- β relevant. However, TGF- β mediates an over-expression of DNMTs in cancer cells, which is responsible for promoter methylation of these genes and. in non-cancer cells, TGF- β down-regulates the expression of DNMTs (25,26).

Table 1 Genes with known association with TGF- β that have DNA hypermethylation in prostate cancer

Name	Function	Reference
1. TBRI	TGF- β receptor type I	(30,31)
2. TBRIITGF- β	TGF- β receptor type II	(31,32)
3. cdh13herin	Adhesion molecule, tumor suppressor	(33,34)
4. TTP (tristetrapolin)	Loss of TTP stabilizes c-Myc mRNA	(35)
5. TGFBI (Betaig-h3)	TGF- β induced gene	(36-38)
6. IGFBP3	IGF binding protein 3	(39,40)
7. beta 4-integrin	Promotes focal adhesion	(34)
8. MAL	Promotes cell differentiation	(41,42)
9. SLIT2	Negative regulation of migration	(36,41,43)
10. Bcl2	Involved in apoptosis	(40,41)
11. Caspase 8	Pro-apoptotic gene	(44)
12. EPHA7	Tumor suppressor in prostate cancer	(45-47)
13. BTG3	Tumor suppressor	(48,49)
14. PTGS2	Pro-inflammatory enzyme	(50-52)
15. HIN1 (or SCGB3A1)	Tumor suppressor	(41,53)
16. RASSF1A	Tumor suppressor gene	(54-56)
17. CHD13	Adhesion molecule	(41,57,58)
18. p15, p16, p21, p27, p57	Cell cycle regulators	(57,59-61)
19. RASSF1A	Pro-apoptotic, negative Ras effector	(41,62)
20. TWIST1	Suppressor of E-cadherin	(41)
21. FHIT	Induces apoptosis through Bak	(63,64)
22. SOCS3	Negative regulator of cytokine	(65,66)
23. TIMP-2, TIMP-3	Inhibitors of metalloproteinase	(67-69)
24. PITX2	Activator of cyclin D2	(41,70-72)
25. DcR1, DcR2	Fail to induced apoptosis through TRAIL	(73,74)
26. GLIPR1 (or RTVP-1)	p53 target gene	(75,76)
27. MGMT	DNA repair gene	(77-81)
28. DKK3 (SFRP1)	Wnt antagonist	(82,83)
29. RUNX3	Tumor suppressor	(84-86)
30. CAV-1	Tumor suppressor	(87,88)
31. Clusterin	Apoptotic protein	(89-91)
32. TFPI2 (PP5, MSP1)	A potent inhibitor of matrix-metalloproteinases	(92,93)
33. SOX7	Suppressor of β -catenin	(94,95)
34. SLC5A8	Tumor suppressor	(96,97)
35. SLC18A2 (or VMAT2)	Affects apoptosis and migration	(98,99)
36. LPL	Tumor suppressor gene	(100,101)
37. HRK (or ATF-2)	Proapoptosis	(102,103)
38. INHBB	Inhibin betaB	(104,105)
39. ID4	Inhibitor of DNA binding	(41,106-108)
40. FYN	Promotes proliferation and motility	(109,110)
41. HPP1 (TMEFF2)	TGF- β signal pathway	(73,84)
42. RRAD	Ras-related GTPases	(111,112)
43. DRM/Gremlin	Down-regulated in Mos-transformed cells	(73,84)

Table 2 Methylated genes in prostate cancer whose regulation by TGF- β is not yet known

Name	Function	Reference
1. HLAa	HLA class-I antigen	(41)
2. ER β	Estrogen receptor	(67)
3. ER α	Estrogen receptor	(67)
4. AR	Androgen receptor	(67)
5. RAR β	Tumor suppressor	(67)
6. DAPK1	Regulate cell death	(118)
7. MDR1	Multi-drug resistant gene	(41,119)
8. APC	Antagonist of Wnt	(41,119-121)
9. CD44	Cell migration and adhesion	(52,57)
10. MCAM (MUC18, CD146)	In advanced PCa	(41,122)
11. TIG1	Retinoic acid receptor responder	(41,123)
12. THRB	Thyroid hormone receptor B	(41)
13. Laminin-5	Role in adhesion and motility	(124)
14. WIF1	Wnt inhibitory factor	(125-127)
15. TSLC1	Tumor suppressor	(128)
16. RIZ1	Rb-interacting zinc finger gene 1	(73,129)
17. Cyclin D2 (or CCND2)	Regulate cell cycle	(54,67,130)
18. GSTP1	Cell detoxification	(4,7,121,131)
19. PDLIM4	Actin binding protein, tumor suppressor	(41,132)
20. Sprouty1	negative regulators of MAPK/PI3K	(133)
21. ZNF331	Tumor suppressor	(134)
22. TMS1(ASC, PYCARD)	Induces apoptosis by caspase	(57,73,135)
23. GPX3	Anti-oxidant	(82,119)
24. NKX2.5	Repress calreticulin expression	(41)
25. NKX3.1	Promotes normal differentiation	(136)
26. DPYS	Sensitivity to 5-FU	(41,137)
27. ENDRB	Endothelin receptor type B	(5,41)
28. CADM2	Cell adhesion molecule	(138)
29. XAF1	Interference with caspase inhibition of XIAP	(139-141)
30. CRBP1	Cellular retinol binding protein, promotes apoptosis	(73,142)
31. FAS (TNFRSF6, APT1, CD95/Apo-1)	Induces apoptosis	(143)
32. RPRM	Inhibits Cdc2-cyclin b1 activity	(73,123)
33. GSTM1	Detoxification	(82)
34. EPB41L3	Erythrocyte membrane protein band 4.1-like 3	(28)
35. SCTR	Gene encoding the secretin receptor	(105)
36. SOCS1	Negative regulator of cytokine	(73,84)
37. HIC	Tumor suppressor	(79,81)

DNA methylation associated with tumor initiation and progression

A characteristic of DNA methylation in cancer is its heterogeneity. Despite of this variation, some trends can

be discerned. We rationalize that genes that are widely methylated are likely involved during early stages of tumor development, such as GSTP-1 (4), which may be used for the early detection of prostate cancer. Many investigators

have used specific methylation pattern for prediction of cancer progression. However, during progression of prostate cancer, the tumor becomes increasingly heterogeneous, it will be difficult to pinpoint which genes are methylated that can be used as a prognostic marker and such efforts have been met with mixed results (144). It is reasonable to assume that as tumors progress, there will be more genes that undergo promoter methylation and demethylation. Therefore, the development of a rapid analysis of DNA methylation profile make it possible to follow the methylation patterns which may be used as an indication of disease progression.

Conclusions and future directions

Based on the present review, it is apparent that TGF- β signaling and DNA methylation are two important events in prostate cancer development and progression. In tumor progression, the deregulated TGF- β signaling mediates an increase in the number of genes undergoing DNA hypermethylation. These genes are generally associated with prevention of apoptosis, promotion of proliferation, facilitation of cell migration and evasion of the immune surveillance, resulting in tumor progression. In the era of personalized medicine, it becomes more important that we clearly define which genes are affected by TGF- β signaling and which genes are promoter hypermethylated during prostate cancer progression. Recent reports point out that some dietary and lifestyle interventions in cancer patients are mainly mediated through a reduction in DNA methylation (125,145,146), while others may lead to both gains and losses (147). It is possible that these dietary and lifestyle factors may be mediated at least partly through a normalization of the vicious cycle of TGF- β signaling in cancer microenvironment (148).

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Footnote

Conflicts of Interest: M. McClelland and D. Mercola are cofounders Proveri Inc., which is engaged in translational

development of aspects of the subject matter. The other authors have no conflicts of interest to declare.

References

1. Mu Y, Gudey SK, Landström M. Non-Smad signaling pathways. *Cell Tissue Res* 2012;347:11-20.
2. Yu N, Kozlowski JM, Park II, et al. Over-expression of transforming growth factor 1 in malignant prostate cells is partly caused by a runaway of TGF-1 auto-induction mediated through a defective recruitment of protein phosphatase 2A by TGF-type I receptor. *Urology* 2010;76:1519.e8-13.
3. Zhang Q, Chen L, Helfand BT, et al. Transforming Growth Factor- β -induced DNA methyltransferase contributes to aggressive prostate cancer phenotypes and predicts biochemical recurrence after radical prostatectomy. *PLoS ONE* 2011;6:e25168.
4. Nelson WG, De Marzo AM, Yegnasubramanian S. Epigenetic alterations in human prostate cancers. *Endocrinology* 2009;150:3991-4002.
5. Phé V, Cussenot O, Rouprêt M. Methylated genes as potential biomarkers in prostate cancer. *BJU Int* 2010;105:1364-70.
6. Park JY. Promoter hypermethylation in prostate cancer. *Cancer Control* 2010;17:245-55.
7. Goering W, Kloth M, Schulz WA. DNA methylation changes in prostate cancer. *Methods Mol Biol* 2012;863:47-66.
8. Sánchez-Carbayo M. Hypermethylation in bladder cancer: biological pathways and translational applications. *Tumour Biol* 2012;33:347-61.
9. Morris MR, Maher ER. Epigenetics of renal cell carcinoma: the path towards new diagnostics and therapeutics. *Genome Med* 2010;2:59.
10. Huang Y, Nayak S, Jankowitz R, et al. Epigenetics in breast cancer: what's new? *Breast Cancer Res* 2011;13:225.
11. Rauch TA, Wang Z, Wu X, et al. DNA methylation biomarkers for lung cancer. *Tumour Biol* 2012;33:287-96.
12. Matsumura N, Huang Z, Mori S, et al. Epigenetic suppression of the TGF-beta pathway revealed by transcriptome profiling in ovarian cancer. *Genome Res* 2011;21:74-82.
13. González-Ramírez I, García-Cuellar C, Sánchez-Pérez Y, et al. DNA methylation in oral squamous cell carcinoma: molecular mechanisms and clinical implications. *Oral Dis* 2011;17:771-8.
14. Delpu Y, Hanoun N, Lulka H, et al. Genetic and

- epigenetic alterations in pancreatic carcinogenesis. *Curr Genomics* 2011;12:15-24.
15. Cedar H, Bergman Y. Programming of DNA Methylation Patterns. *Annu Rev Biochem* 2012;81:97-117.
 16. Chiam K, Ricciardelli C, Bianco-Miotto T. Epigenetic biomarkers in prostate cancer: Current and future uses. *Cancer Lett* 2014;342:248-56.
 17. Sandoval J, Esteller M. Cancer epigenomics: beyond genomics. *Curr Opin Genet Dev* 2012 22:50-5.
 18. Hata A, Davis BN. Control of microRNA biogenesis by TGFbeta signaling pathway-A novel role of Smads in the nucleus. *Cytokine Growth Factor Rev* 2009;20:517-21.
 19. Hussey GS, Chaudhury A, Dawson AE, et al. Identification of an mRNP complex regulating tumorigenesis at the translational elongation step. *Mol Cell* 2011;41:419-31.
 20. Shi Y, Massagué J. Mechanisms of TGF-beta Signaling from Cell Membrane to the Nucleus. *Cell* 2003;113:685-700.
 21. Sontag E, Sontag JM, Garcia A. Protein phosphatase 2A is a critical regulator of protein kinase C zeta signaling targeted by SV40 small t to promote cell growth and NF-kappaB activation. *EMBO J* 1997;16:5662-71.
 22. Vogelmann R, Nguyen-tat MD, Giehl K, et al. TGFbeta-induced downregulation of E-cadherin-based cell-cell adhesion depends on PI3-kinase and PTEN. *J Cell Sci* 2005;118:4901-12.
 23. Kang JS, Liu C, Derynck R. New regulatory mechanisms of TGF-beta receptor function. *Trends Cell Biol* 2009;19:385-94.
 24. Hong M, Wilkes MC, Penheiter SG, et al. Non-Smad transforming growth factor- signaling regulated by focal adhesion kinase binding the p85 subunit of phosphatidylinositol 3-kinase. *J Biol Chem* 2011;286:17841-50.
 25. Luo X, Zhang Q, Liu V, et al. Cutting Edge: TGF-Induced expression of Foxp3 in T cells is mediated through inactivation of ERK. *J Immunol* 2008;180:2757-61.
 26. You H, Ding W, Rountree CB. Epigenetic regulation of cancer stem cell marker CD133 by transforming growth factor-beta. *Hepatology* 2010;51:1635-44.
 27. Zhang Q, Helfand BT, Jang TL, et al. NF-kappaB-Mediated Transforming Growth Factor--Induced Expression of Vimentin is an Independent Predictor of Biochemical Recurrence After Radical Prostatectomy. *Clin Cancer Res* 2009;15:3557-67.
 28. Schulz WA, Ingenwerth M, Djuidje CE, et al. Changes in cortical cytoskeletal and extracellular matrix gene expression in prostate cancer are related to oncogenic ERG deregulation. *BMC Cancer* 2010;10:505.
 29. Lu R, Wang X, Chen ZF, et al. Inhibition of the extracellular signal-regulated kinase/mitogen-activated protein kinase pathway decreases DNA methylation in colon cancer cells. *J Biol Chem* 2007;282:12249-59.
 30. Kim IY, Ahn HJ, Zelner DJ, et al. Loss of expression of transforming growth factor type I and type II receptors correlates with tumor grade in human prostate cancer tissues. *Clin Cancer Res* 1996;2:1255-61.
 31. Zhang Q, Rubenstein JN, Jang TL, et al. Insensitivity to transforming growth factor-signaling is resulted from promoter methylation of cognate receptors in human prostate cancer cells (LNCaP). *Mol Endocrinol* 2005;19:2390-9.
 32. Yamashita S, Takahashi S, McDonnell N, et al. Methylation silencing of transforming growth factor-beta receptor type II in rat prostate cancers. *Cancer Res* 2008;68:2112-21.
 33. Dumont N, Wilson MB, Crawford YG, et al. Sustained induction of epithelial to mesenchymal transition activates DNA methylation of genes silenced in basal-like breast cancers. *Proc Natl Acad Sci USA* 2008;105:14867-72.
 34. Yang X, Pursell B, Lu S, et al. Regulation of beta 4-integrin expression by epigenetic modifications in the mammary gland and during the epithelial-to-mesenchymal transition. *J Cell Sci* 2009;122:2473-80.
 35. Sohn BH, Park IY, Lee JJ, et al. Functional switching of TGF-beta1 signaling in liver cancer via epigenetic modulation of a single CpG site in TTP promoter. *Gastroenterology* 2010;138:1898-908.
 36. Yu J, Cao Q, Yu J, et al. The neuronal repellent SLIT2 is a target for repression by EZH2 in prostate cancer. *Oncogene* 2010;29:5370-80.
 37. Skonier J, Neubauer M, Madisen L, et al. cDNA cloning and sequence analysis of beta ig-h3, a novel gene induced in a human adenocarcinoma cell line after treatment with transforming growth factor-beta. *DNA Cell Biol* 1992;11:511-22.
 38. Shah JN, Shao G, Hei TK, et al. Methylation screening of the TGFBI promoter in human lung and prostate cancer by methylation-specific PCR. *BMC Cancer* 2008;8:284.
 39. Kawasaki T, Nosho K, Ohnishi M, et al. IGFBP3 promoter methylation in colorectal cancer: relationship with microsatellite instability, CpG island methylator phenotype, and p53. *Neoplasia* 2007;9:1091-8.
 40. Yang YA, Zhang GM, Feigenbaum L, et al. Smad3 reduces susceptibility to hepatocarcinoma by sensitizing hepatocytes to apoptosis through downregulation of Bcl-2.

- Cancer Cell 2006;9:445-57.
41. Vasiljević N, Wu K, Brentnall AR, et al. Absolute quantitation of DNA methylation of 28 candidate genes in prostate cancer using pyrosequencing. *Dis Markers* 2011;30:151-61.
 42. Kojima T, Takasawa A, Kyuno D, et al. Downregulation of tight junction-associated MARVEL protein marvelD3 during epithelial-mesenchymal transition in human pancreatic cancer cells. *Exp Cell Res* 2011;317:2288-98.
 43. Zhou W, Yu W, Xie W, et al. The role of SLIT-ROBO signaling in proliferative diabetic retinopathy and retinal pigment epithelial cells. *Mol Vis* 2011;17:1526-36.
 44. Ying TH, Yang SF, Tsai SJ, et al. Fisetin induces apoptosis in human cervical cancer HeLa cells through ERK1/2-mediated activation of caspase-8/-caspase-3-dependent pathway. *Arch Toxicol* 2012;86:263-73.
 45. Battle E, Bacani J, Begthel H, et al. EphB receptor activity suppresses colorectal cancer progression. *Nature* 2005;435:1126-30.
 46. Nakanishi H, Nakamura T, Canaani E, et al. ALL1 fusion proteins induce deregulation of EphA7 and ERK phosphorylation in human acute leukemias. *Proc Natl Acad Sci USA* 2007;104:14442-7.
 47. Guan M, Xu C, Zhang F, et al. Aberrant methylation of EphA7 in human prostate cancer and its relation to clinicopathologic features. *Int J Cancer* 2009;124:88-94.
 48. Majid S, Dar AA, Shahryari V, et al. Genistein reverses hypermethylation and induces active histone modifications in tumor suppressor gene B-Cell translocation gene 3 in prostate cancer. *Cancer* 2010;116:66-76.
 49. Lin TY, Cheng YC, Yang HC, et al. Loss of the candidate tumor suppressor BTG3 triggers acute cellular senescence via the ERK-JMJD3-p16(INK4a) signaling axis. *Oncogene* 2012;31:3287-97.
 50. Fosslien E. Review: molecular pathology of cyclooxygenase-2 in cancer-induced angiogenesis. *Ann Clin Lab Sci* 2001;31:325-48.
 51. Bastian PJ, Ellinger J, Wellmann A, et al. Diagnostic and prognostic information in prostate cancer with the help of a small set of hypermethylated gene loci. *Clin Cancer Res* 2005;11:4097-106.
 52. Woodson K, O'Reilly KJ, Ward DE, et al. CD44 and PTGS2 methylation are independent prognostic markers for biochemical recurrence among prostate cancer patients with clinically localized disease. *Epigenetics* 2006;1:183-6.
 53. Krop I, Parker MT, Bloushtain-Qimron N, et al. HIN-1, an inhibitor of cell growth, invasion, and AKT activation. *Cancer Res* 2005;65:9659-69.
 54. Henrique R, Costa VL, Cerveira N, et al. Hypermethylation of Cyclin D2 is associated with loss of mRNA expression and tumor development in prostate cancer. *J Mol Med (Berl)* 2006;84:911-8.
 55. Hesson LB, Cooper WN, Latif F. The role of RASSF1A methylation in cancer. *Dis Markers* 2007;23:73-87.
 56. Lee SJ, Lee MH, Kim DW, et al. Cross-Regulation between Oncogenic BRAFV600E Kinase and the MST1 Pathway in Papillary Thyroid Carcinoma. *PLoS One* 2011;6:e16180.
 57. Alumkal JJ, Zhang Z, Humphreys EB, et al. Effect of DNA methylation on identification of aggressive prostate cancer. *Urology* 2008;72:1234-9.
 58. Rodrigues RF, Roque L, Krug T, et al. Poorly differentiated and anaplastic thyroid carcinomas: chromosomal and oligo-array profile of five new cell lines. *Br J Cancer* 2007;96:1237-45.
 59. Wang SE, Narasanna A, Whitell CW, et al. Convergence of p53 and transforming growth factor beta (TGFbeta) signaling on activating expression of the tumor suppressor gene maspin in mammary epithelial cells. *J Biol Chem* 2007;282:5661-9.
 60. Hinshelwood RA, Huschtscha LI, Melki J, et al. Concordant epigenetic silencing of transforming growth factor-beta signaling pathway genes occurs early in breast carcinogenesis. *Cancer Res* 2007;67:11517-27.
 61. Wang X, Sun DF, Lu R, et al. RAF may induce cell proliferation through hypermethylation of tumor suppressor gene promoter in gastric epithelial cells. *Cancer Sci* 2009;100:117-25.
 62. Bhaskaran N, Souchelnytskyi S. Systemic analysis of TGFbeta proteomics revealed involvement of Plag1/CNK1/RASSF1A/Src network in TGFbeta1-dependent activation of Erk1/2 and cell proliferation. *Proteomics* 2008;8:4507-20.
 63. Mishra DK, Chen Z, Wu Y, et al. Global methylation pattern of genes in androgen-sensitive and androgen-independent prostate cancer cells. *Mol Cancer Ther* 2010;9:33-45.
 64. Kelley K, Berberich SJ. FHIT gene expression is repressed by mitogenic signaling through the PI3K/AKT/FOXO pathway. *Am J Cancer Res* 2011;1:62-70.
 65. Qin H, Wang L, Feng T, et al. TGF-beta promotes Th17 cell development through inhibition of SOCS3. *J Immunol* 2009;183:97-105.
 66. Pierconti F, Martini M, Pinto F, et al. Epigenetic silencing of SOCS3 identifies a subset of prostate cancer with an aggressive behavior. *Prostate* 2011;71:318-25.

67. Diaw L, Woodson K, Gillespie JW. Prostate cancer epigenetics: a review on gene regulation. *Gene Regul Syst Bio* 2007;1:313-25.
68. Rao ZY, Cai MY, Yang GF, et al. EZH2 supports ovarian carcinoma cell invasion and/or metastasis via regulation of TGF-beta1 and is a predictor of outcome in ovarian carcinoma patients. *Carcinogenesis* 2010;31:1576-83.
69. Shin YJ, Kim JH. The role of EZH2 in the regulation of the activity of matrix metalloproteinases in prostate cancer cells. *PLoS ONE* 2012;7:e30393.
70. Bamforth SD, Bragança J, Farthing CR, et al. Cited2 controls left-right patterning and heart development through a Nodal-Pitx2c pathway. *Nat Genet* 2004;36:1189-96.
71. Bañez LL, Sun L, van Leenders GJ, et al. Multicenter clinical validation of PITX2 methylation as a prostate specific antigen recurrence predictor in patients with post-radical prostatectomy prostate cancer. *J Urol* 2010;184:149-156.
72. Vinarskaja A, Schulz WA, Ingenwerth M, et al. Association of PITX2 mRNA down-regulation in prostate cancer with promoter hypermethylation and poor prognosis. *Urol Oncol* 2013;31:622-7.
73. Suzuki M, Shigematsu H, Shivapurkar N, et al. Methylation of apoptosis related genes in the pathogenesis and prognosis of prostate cancer. *Cancer Lett* 2006;242:222-30.
74. Lunghi P, Giuliani N, Mazzerà L, et al. Targeting MEK/MAPK signal transduction module potentiates ATO-induced apoptosis in multiple myeloma cells through multiple signaling pathways. *Blood* 2008;112:2450-62.
75. Ren C, Li L, Yang G, et al. RTVP-1, a tumor suppressor inactivated by methylation in prostate cancer. *Cancer Res* 2004;64:969-76.
76. Hameetman L, Rozeman LB, Lombaerts M, et al. Peripheral chondrosarcoma progression is accompanied by decreased Indian Hedgehog signalling. *J Pathol* 2006;209:501-11.
77. Yamada H, Vijayachandra K, Penner C, et al. Increased sensitivity of transforming growth factor (TGF) beta 1 null cells to alkylating agents reveals a novel link between TGFbeta signaling and O(6)-methylguanine methyltransferase promoter hypermethylation. *J Biol Chem* 2001;276:19052-8.
78. Konishi N, Nakamura M, Kishi M, et al. DNA hypermethylation status of multiple genes in prostate adenocarcinomas. *Jpn J Cancer Res* 2002;93:767-73.
79. Yamanaka M, Watanabe M, Yamada Y, et al. Altered methylation of multiple genes in carcinogenesis of the prostate. *Int J Cancer* 2003;106:382-7.
80. Kang GH, Lee S, Lee HJ, et al. Aberrant CpG island hypermethylation of multiple genes in prostate cancer and prostatic intraepithelial neoplasia. *J Pathol* 2004;202:233-40.
81. Yegnasubramanian S, Kowalski J, Gonzalgo ML, et al. Hypermethylation of CpG islands in primary and metastatic human prostate cancer. *Cancer Res* 2004;64:1975-86.
82. Lodygin D, Epanchintsev A, Menssen A, et al. Functional epigenomics identifies genes frequently silenced in prostate cancer. *Cancer Res* 2005;65:4218-27.
83. Noordhuis MG, Fehrmann RS, Wisman GB, et al. Involvement of the TGF-beta and beta-catenin pathways in pelvic lymph node metastasis in early-stage cervical cancer. *Clin Cancer Res* 2011;17:1317-30.
84. Suzuki M, Shigematsu H, Shames DS, et al. DNA methylation-associated inactivation of TGFbeta-related genes DRM/Gremlin, RUNX3, and HPP1 in human cancers. *Br J Cancer* 2005;93:1029-37.
85. Hasegawa K, Yazumi S, Wada M, et al. Restoration of RUNX3 enhances transforming growth factor-beta-dependent p21 expression in a biliary tract cancer cell line. *Cancer Sci* 2007;98:838-43.
86. Richiardi L, Fiano V, Vizzini L, et al. Promoter methylation in APC, RUNX3, and GSTP1 and mortality in prostate cancer patients. *J Clin Oncol* 2009;27:3161-8.
87. Cui J, Rohr LR, Swanson G, et al. Hypermethylation of the caveolin-1 gene promoter in prostate cancer. *Prostate* 2001;46:249-56.
88. Razani B, Zhang XL, Bitzer M, et al. Caveolin-1 regulates transforming growth factor (TGF)-beta/SMAD signaling through an interaction with the TGF-beta type I receptor. *J Biol Chem* 2001;276:6727-38.
89. Rosemblyt N, Chen CL. Regulators for the rat clusterin gene: DNA methylation and cis-acting regulatory elements. *J Mol Endocrinol* 1994;13:69-76.
90. Rauhala HE, Porkka KP, Saramäki OR, et al. Clusterin is epigenetically regulated in prostate cancer. *Int J Cancer* 2008;123:1601-9.
91. Rizzi F, Bettuzzi S. Clusterin (CLU) and prostate cancer. *Adv Cancer Res* 2009;105:1-19.
92. Becker J, Volland S, Noskova I, et al. Keratoepithelin reverts the suppression of tissue factor pathway inhibitor 2 by MYCN in human neuroblastoma: a mechanism to inhibit invasion. *Int J Oncol* 2008;32:235-40.
93. Ribarska T, Ingenwerth M, Goering W, et al. Epigenetic

- inactivation of the placentally imprinted tumor suppressor gene TFPI2 in prostate carcinoma. *Cancer Genomics Proteomics* 2010;7:51-60.
94. Zhang C, Basta T, Fawcett SR, et al. SOX7 is an immediate-early target of VegT and regulates Nodal-related gene expression in *Xenopus*. *Dev Biol* 2005;278:526-41.
 95. Guo L, Zhong D, Lau S, et al. Sox7 Is an independent checkpoint for beta-catenin function in prostate and colon epithelial cells. *Mol Cancer Res* 2008;6:1421-30.
 96. Park JY, Zheng W, Kim D, et al. Candidate tumor suppressor gene SLC5A8 is frequently down-regulated by promoter hypermethylation in prostate tumor. *Cancer Detect Prev* 2007;31:359-65.
 97. Bennett KL, Romigh T, Eng C. Disruption of transforming growth factor-beta signaling by five frequently methylated genes leads to head and neck squamous cell carcinoma pathogenesis. *Cancer Res* 2009;69:9301-5.
 98. Sørensen KD, Wild PJ, Mortezaei A, et al. Genetic and epigenetic SLC18A2 silencing in prostate cancer is an independent adverse predictor of biochemical recurrence after radical prostatectomy. *Clin Cancer Res* 2009;15:1400-10.
 99. Bourque M, Liu B, Dluzen DE, et al. Sex differences in methamphetamine toxicity in mice: effect on brain dopamine signaling pathways. *Psychoneuroendocrinology* 2011;36:955-69.
 100. Irvine SA, Foka P, Rogers SA, et al. A critical role for the Sp1-binding sites in the transforming growth factor-beta-mediated inhibition of lipoprotein lipase gene expression in macrophages. *Nucleic Acids Res* 2005;33:1423-34.
 101. Kim JW, Cheng Y, Liu W, et al. Genetic and epigenetic inactivation of LPL gene in human prostate cancer. *Int J Cancer* 2009;124:734-8.
 102. Higuchi T, Nakamura M, Shimada K, et al. HRK inactivation associated with promoter methylation and LOH in prostate cancer. *Prostate* 2008;68:105-13.
 103. Ribas VT, Arruda-Carvalho M, Linden R, et al. Early c-Jun N-terminal kinase-dependent phosphorylation of activating transcription factor-2 is associated with degeneration of retinal ganglion cells. *Neuroscience* 2011;180:64-74.
 104. Dragovic RA, Ritter LJ, Schulz SJ, et al. Oocyte-secreted factor activation of SMAD 2/3 signaling enables initiation of mouse cumulus cell expansion. *Biol Reprod* 2007;76:848-57.
 105. Devaney J, Stirzaker C, Qu W, et al. Epigenetic deregulation across chromosome 2q14.2 differentiates normal from prostate cancer and provides a regional panel of novel DNA methylation cancer biomarkers. *Cancer Epidemiol Biomarkers Prev* 2011;20:148-59.
 106. Carey JP, Asirvatham AJ, Galm O, et al. Inhibitor of differentiation 4 (Id4) is a potential tumor suppressor in prostate cancer. *BMC Cancer* 2009;9:173.
 107. Hogg K, Etherington SL, Young JM, et al. Inhibitor of differentiation (Id) genes are expressed in the steroidogenic cells of the ovine ovary and are differentially regulated by members of the transforming growth factor-beta family. *Endocrinology* 2010;151:1247-56.
 108. Vinarskaja A, Goering W, Ingenwerth M, et al. ID4 is frequently downregulated and partially hypermethylated in prostate cancer. *World J Urol* 2012;30:319-25.
 109. Sørensen KD, Borre M, Ørntoft TF, et al. Chromosomal deletion, promoter hypermethylation and downregulation of FYN in prostate cancer. *Int J Cancer* 2008;122:509-19.
 110. Kim AN, Jeon WK, Lim KH, et al. Fyn mediates transforming growth factor-beta1-induced down-regulation of E-cadherin in human A549 lung cancer cells. *Biochem Biophys Res Commun* 2011;407:181-4.
 111. Kannangai R, Diehl AM, Sicklick J, et al. Hepatic angiomyolipoma and hepatic stellate cells share a similar gene expression profile. *Hum Pathol* 2005;36:341-7.
 112. Suzuki M, Shigematsu H, Shames DS, et al. Methylation and gene silencing of the Ras-related GTPase gene in lung and breast cancers. *Ann Surg Oncol* 2007;14:1397-404.
 113. Yamane K, Suzuki H, Ihn H, et al. Cell type-specific regulation of the TGF-beta-responsive alpha2(I) collagen gene by CpG methylation. *J Cell Physiol* 2005;202:822-30.
 114. Hurtubise A, Momparler RL. Evaluation of antineoplastic action of 5-aza-2'-deoxycytidine (Dacogen) and docetaxel (Taxotere) on human breast, lung and prostate carcinoma cell lines. *Anticancer Drugs* 2004;15:161-7.
 115. Li X, Kaplun A, Lonardo F, et al. HDAC1 inhibition by maspin abrogates epigenetic silencing of glutathione S-transferase pi in prostate carcinoma cells. *Mol Cancer Res* 2011;9:733-45.
 116. Rivenbark AG, Stolzenburg S, Beltran AS, et al. Epigenetic reprogramming of cancer cells via targeted DNA methylation. *Epigenetics* 2012;7:350-60.
 117. Pellacani D, Packer RJ, Frame FM, et al. Regulation of the stem cell marker CD133 is independent of promoter hypermethylation in human epithelial differentiation and cancer. *Mol Cancer* 2011;10:94.
 118. Liu WB, Ao L, Zhou ZY, et al. CpG island hypermethylation of multiple tumor suppressor genes

- associated with loss of their protein expression during rat lung carcinogenesis induced by 3-methylcholanthrene and diethylnitrosamine. *Biochem Biophys Res Commun* 2010;402:507-14.
119. Dobosy JR, Roberts JL, Fu VX, et al. The expanding role of epigenetics in the development, diagnosis and treatment of prostate cancer and benign prostatic hyperplasia. *J Urol* 2007;177:822-31.
 120. Henrique R, Ribeiro FR, Fonseca D, et al. High promoter methylation levels of APC predict poor prognosis in sextant biopsies from prostate cancer patients. *Clin Cancer Res* 2007;13:6122-9.
 121. Tilandová P, Kajo K, Kliment J, et al. Detection of DNA hypermethylation as a potential biomarker for prostate cancer. *Klin Onkol* 2010;23:293-9.
 122. Liu JW, Nagpal JK, Jeronimo C, et al. Hypermethylation of MCAM gene is associated with advanced tumor stage in prostate cancer. *Prostate* 2008;68:418-26.
 123. Ellinger J, Bastian PJ, Jurgan T, et al. CpG island hypermethylation at multiple gene sites in diagnosis and prognosis of prostate cancer. *Urology* 2008;71:161-7.
 124. Sathyanarayana UG, Padar A, Suzuki M, et al. Aberrant promoter methylation of laminin-5-encoding genes in prostate cancers and its relationship to clinicopathological features. *Clin Cancer Res* 2003;9:6395-400.
 125. Yee DS, Tang Y, Li X, et al. The Wnt inhibitory factor 1 restoration in prostate cancer cells was associated with reduced tumor growth, decreased capacity of cell migration and invasion and a reversal of epithelial to mesenchymal transition. *Mol Cancer* 2010;9:162.
 126. Wissmann C, Wild PJ, Kaiser S, et al. WIF1, a component of the Wnt pathway, is down-regulated in prostate, breast, lung, and bladder cancer. *J Pathol* 2003;201:204-12.
 127. Gudjonsson JE, Johnston A, Stoll SW, et al. Evidence for altered Wnt signaling in psoriatic skin. *J Invest Dermatol* 2010;130:1849-59.
 128. Fukuhara H, Kuramochi M, Fukami T, et al. Promoter methylation of TSLC1 and tumor suppression by its gene product in human prostate cancer. *Jpn J Cancer Res* 2002;93:605-9.
 129. Dong SW, Cui YT, Zhong RR, et al. Decreased expression of retinoblastoma protein-interacting zinc-finger gene 1 in human esophageal squamous cell cancer by DNA methylation. *Clin Lab* 2012;58:41-51.
 130. Padar A, Sathyanarayana UG, Suzuki M, et al. Inactivation of cyclin D2 gene in prostate cancers by aberrant promoter methylation. *Clin Cancer Res* 2003;9:4730-4.
 131. Meiers I, Shanks JH, Bostwick DG. Glutathione S-transferase pi (GSTP1) hypermethylation in prostate cancer: review 2007. *Pathology* 2007;39:299-304.
 132. Vanaja DK, Ballman KV, Morlan BW, et al. PDLIM4 repression by hypermethylation as a potential biomarker for prostate cancer. *Clin Cancer Res* 2006;12:1128-36.
 133. Kwabi-Addo B, Ren C, Ittmann M. DNA methylation and aberrant expression of Sprouty1 in human prostate cancer. *Epigenetics* 2009;4:54-61.
 134. Yu J, Liang QY, Wang J, et al. Zinc-finger protein 331, a novel putative tumor suppressor, suppresses growth and invasiveness of gastric cancer. *Oncogene* 2013;32:307-17.
 135. Das PM, Ramachandran K, Vanwert J, et al. Methylation mediated silencing of TMS1/ASC gene in prostate cancer. *Mol Cancer* 2006;5:28.
 136. Schulz WA, Hatina J. Epigenetics of prostate cancer: beyond DNA methylation. *J Cell Mol Med* 2006;10:100-25.
 137. Chung W, Kwabi-Addo B, Ittmann M, et al. Identification of novel tumor markers in prostate, colon and breast cancer by unbiased methylation profiling. *PLoS ONE* 2008;3:e2079.
 138. Chang G, Xu S, Dhir R, et al. Hypoexpression and epigenetic regulation of candidate tumor suppressor gene CADM-2 in human prostate cancer. *Clin Cancer Res* 2010;16:5390-401.
 139. Fang X, Liu Z, Fan Y, et al. Switch to full-length of XAF1 mRNA expression in prostate cancer cells by the DNA methylation inhibitor. *Int J Cancer* 2006;118:2485-9.
 140. Lee MG, Huh JS, Chung SK, et al. Promoter CpG hypermethylation and downregulation of XAF1 expression in human urogenital malignancies: implication for attenuated p53 response to apoptotic stresses. *Oncogene* 2006;25:5807-22.
 141. Murphy TM, Perry AS, Lawler M. The emergence of DNA methylation as a key modulator of aberrant cell death in prostate cancer. *Endocr Relat Cancer* 2008;15:11-25.
 142. Jerónimo C, Henrique R, Oliveira J, et al. Aberrant cellular retinol binding protein 1 (CRBP1) gene expression and promoter methylation in prostate cancer. *J Clin Pathol* 2004;57:872-6.
 143. Santourlidis S, Warskulat U, Florl AR, et al. Hypermethylation of the tumor necrosis factor receptor superfamily 6 (APT1, Fas, CD95/Apo-1) gene promoter at rel/nuclear factor kappaB sites in prostatic carcinoma. *Mol Carcinog* 2001;32:36-43.
 144. Jerónimo C, Bastian PJ, Bjartell A, et al. Epigenetics in prostate cancer: biologic and clinical relevance. *Eur Urol*

- 2011;60:753-66.
145. Ho E, Beaver LM, Williams DE, et al. Dietary factors and epigenetic regulation for prostate cancer prevention. *Adv Nutr* 2011;2:497-510.
146. Lim U, Song MA. Dietary and lifestyle factors of DNA methylation. *Methods Mol Biol* 2012;863:359-76.
147. Ornish D, Magbanua MJ, Weidner G, et al. Changes in

- prostate gene expression in men undergoing an intensive nutrition and lifestyle intervention. *Proc Natl Acad Sci USA* 2008;105:8369-74.
148. Lee C, Zhang Q, Kozlowski J, et al. Natural products and transforming growth factor-beta (TGF-) signaling in cancer development and progression. *Curr Cancer Drug Targets* 2013;13:500-5.

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