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A Correlational Analysis of the Recovery from Sleep Deprivation across a Panel of Inbred Mouse Strains

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# UNIVERSITY OF CALIFORNIA

Los Angeles

A Correlational Analysis of Recovery from Sleep Deprivation across a Panel of Inbred

Mouse Strains

A thesis submitted in partial satisfaction of the requirements for the degree Master of Science in Physiological Science

by

Damion Lamar Trotter

### ABSTRACT OF THE THESIS

# A Correlational Analysis of Recovery from Sleep Deprivation across a Panel of Inbred Mouse Strains

by Damion Lamar Trotter

Master of Science in Physiological Science University of California, Los Angeles, 2022 Professor Ketema N. Paul, Chair

Sleep is known to be necessary for everyday life for humans. Although sleep is common across phylogeny, there still is much not known about sleep, including many of the genetic components that regulate it. In this study, we investigated 10 sleep phenotypes in 24 inbred mouse strains while looking at both spontaneous sleep conditions and recovery following six hours of sleep deprivation beginning at the light onset in a 24-hour light-dark cycle. We found that there were significant differences across strains in ten sleep phenotypes. We then tested the hypothesis that there are strain differences in the homeostatic ability to recover from sleep loss. Our findings suggested that there are strain differences in the recovery from sleep loss across different mouse strains. These findings suggest that sleep homeostasis is sensitive to the genetic differences across these inbred strains. This work furthers our understanding of the genetic mechanisms underlying sleep homeostasis.

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The thesis of Damion Lamar Trotter is approved.

Arthur P. Arnold Gina R. Poe Christopher S. Colwell Ketema N. Paul, Committee Chair

University of California, Los Angeles

Dedicated to my family and friends Who stood by me And encouraged me to never give up

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### **Chapter 1: Introduction**

### 1.1 What is Sleep?

Sleep is a period of dormancy exhibited by all known animals and is characterized by reduced responsiveness to external stimuli and reduced muscle activity. A key aspect of sleep is that it is easily reversible, unlike a coma or other disorders of consciousness. This prevalence across phylogeny denotes an evolutionary necessity for sleep, which is further substantiated by the fact that prolonged deprivation of sleep can lead to cognitive impairment (Alhola & Polo-Kantola, 2007), loss of bodily function (Goel et al., 2009), organ failure (Periasamy et al., 2015), and eventual death (Vaccaro et al., 2020). The impact of sleep loss on mortality has been reported in studies with dogs (Anderson et al., 2020), rats, mice, and flies (Vaccaro et al., 2020) as well as many other model organisms.

There are two major processes involved in sleep, Process C and Process S (Borbély et al., 2016). Process C, also known as the circadian process, is involved in the body's circadian regulation of internal biological processes and alertness. As a wake-promoting process, it is more active during the daytime and begins to dissipate during an animal's time for rest. Process C is critical also as it consolidates sleep and wake into distinct episodes. Process S is the process of sleep homeostasis. Sleep pressure builds up the longer and animal stays awake and dissipates when they enter a period of sleep.

Sleep is composed of two major stages: non-Rapid Eye Movement (NREM) and Rapid Eye Movement (REM). NREM sleep consists of three stages: N1, N2 and N3 (Patel et al., 2022). N1 sleep is the period of light sleep immediately transitioning from wakefulness and is characterized by slowing down of eye movement and muscle activity and lasts for about 5-10 minutes. N1 sleep is also when hypnic jerks occur,

when the body experiences false sensory phenomena such as a feeling of falling. N2 sleep involves periods of muscle contraction and relaxation. In N2 sleep, eye movement comes to an almost complete stop, the heart rate slows down, body temperature lowers, and the body begins to prepare for deep sleep. Following N2 sleep the body enters N3 sleep. N3 sleep is characterized by the presence of delta waves and there are no eye movements or muscle activity. REM sleep follows NREM sleep, and brain activity starts to increase similar to that of wakefulness. Dreams are formed and are present during this stage and REM begins around 90 minutes into sleep on average in humans. Both stages are important in different aspects. NREM is important for tissue regeneration (Eugene & Masiak, 2015) and strengthening of the immune system whereas REM sleep is important for memory consolidation (Peever et al., 2016) and both are important for learning (Tamaki et al., 2020).

#### 1.2 How is Sleep Studied?

There are many different paradigms and models that can be used to study sleep in both humans and animals. Standard tools to measure sleep include electroencephalographs (EEGs) which are used to measure the activity of electrical brainwaves (Campbell, 2019), and electromyographs (EMGs) which are used to measure muscle activity (Oishi et al., 2016). EEGs are especially helpful when it comes to distinguishing between NREM and REM sleep as delta waves (.5-4 Hz) become more prevalent during NREM sleep. EMGs are a helpful tool that serve as an additional check when it is difficult to distinguish between wake and REM sleep, as the EEGs can look similar, but there should be a lack of muscle movement during REM as opposed to being awake. The process of using EEG and EMG to determine sleep stage is called polysomnography and most sleep-wake data in animal models is analyzed by scoring

the data manually. A number of labs have looked at automatic scoring (Barger et al., 2019; Gao et al., 2016; Rytkönen et al., 2011) but currently, there is no consistent mechanism for reliable sleep scoring, and as such manual scoring is still used in many labs.

A popular model organism to study sleep is the mouse. Mice are used to study sleep because genetic tools are more advanced in mice than in other organisms and mice have clearly defined phenotypes, specifically bouts of sleep which tend to last about 2-4 minutes on average (Toth & Bhargava, 2013). These short bouts also tend to help when trying to use mice as a model for sleep disorders such as insomnia, with which mouse strains like DBA/2J could be used as a potential model (Toth & Bhargava, 2013).

#### 1.3 Sleep and Genetics

Sleep can differ between animal species, and even within the same species, but a major question that exists is the degree of genetic control of sleep homeostasis. Scientists have explored this in a myriad of ways. A popular purpose for studying sleep and genetics involves sleeping disorders and their potential heritability. A 2011 study (Sehgel & Mignot, 2011) conducted a review of sleep studies looking at genes and found that narcolepsy can be driven by genetic anomalies (Miyagawa & Tokunaga, 2019; Mignot, 1998) and may even be driven by ethnic components (Kornum et al., 2011). But the possible genetic contributions to sleep homeostatic mechanisms are not known.

### 1.4 Purpose

In this study, I compared the sleep/wake architecture of 22 different strains of inbred mice in both spontaneous and sleep deprivation conditions. Following this, I then analyzed how the sleep phenotypes of these strains differed between spontaneous and

recovery conditions. My central hypothesis is that there would be strain differences in sleep homeostatic mechanisms that would be reflected in underlying genetic differences between the strains.

#### **Chapter 2: Materials and Methods**

### 2.1 Animals

In order to assess the potential for a genetic component of sleep in adult male mice, 24 inbred mouse strains were analyzed. The strains studied were: 129/J (n = 7), A/J (n = 5), AKR/J (n = 7), BTBR T+ Itpr3tf/J (n = 11), BUB/BnJ (n = 7), C3H/HeJ (n = 4), C57BL/6J (n= 4), C57L/J (n = 7), C58/J (n = 7), CAST (n = 8), CBA/J (n = 6), CZECH/EiiJ (n = 3), DBA/2J (n = 5), FVB/NJ (n = 2), KK/HiJ (n = 8), MOLF/EIJ (n =5), NOD/ShiITj (n = 8), NZB (n = 2), PL/J (n = 5), PWD/PhJ (n = 4), SJL/J (n = 7), SM/J (n = 3), SWR/J (n = 3), and WSB/EiJ (n = 6) strains. All of the mice were maintained on a 12-hour light:12-hour dark schedule throughout the study. Food and water were provided ad-libitum and they were individually housed for two weeks prior to the experimental aspect of the study. All animals were purchased from Jackson Laboratories.

### 2.2 Phenotypes

For the purposes of this study, 10 sleep phenotypes were studied. These were NREM sleep, REM sleep, total sleep, number of bouts of NREM, duration of bouts of NREM, number of bouts of REM, duration of bouts of REM, number of bouts of total sleep, duration of bouts of total sleep, and number of bouts of arousal.

### 2.3 Surgical Methods

EEG and EMG electrodes were implanted in anesthetized mice. The heads were positioned using a head mount provided by Pinnacle Technologies and three electrodes

were implanted into the head [Figure 1]. The first electrode is implanted into the frontal area of the brain and the second (recording) and third (reference) electrodes are implanted into the intraparietal area. A fourth screw was used to serve as a ground.

### 2.4. EEG and EMG Recordings

Following the surgery, the mice are given a 14-day period to recover from surgery. Following that week, they were moved to the sleep recording suite and given time to accommodate to the new areas. They were then connected to the tether which is attached to the low-resistance commutator mounted over the cage [**Figure 1**] (provided by Pinnacle Technologies). Mice were able to freely roam their cage while becoming acclimated to their environment. Seven days after entering the sleep recording chambers, EEG and EMG began recording. The data was collected on a computer running Sirenia Acquisition software (Provided by Pinnacle Technologies). This software was developed solely for the purpose of recording sleep in animals, specifically rodents. The data was recorded for 24 hours in a baseline condition and subsequently 6 hours in a sleep deprivation condition followed by 18 hours of recovery.

#### 2.5 Sleep Deprivation

After 24 hours of baseline recording, sleep deprivation was conducted in the first 6 hours of the light phase [ZT 0-6] across all studied mouse lines. Gentle handling was used to keep the mice awake either by the introduction of a novel object into the cage, disturbing the bedding, tapping on the cage, and when necessary, the mice were touched delicately. [**Figure 2**] During this period access to food was still allowed ad-libitum.

#### 2.6 Sleep Scoring

Once the data was collected, it was classified by a trained observer. The classification process used data recorded by the EEG and EMG waves. Wake, NREM, and REM sleep were primarily determined by the frontal electrode and the EMG served to help differentiate between wake and REM sleep. This would be conducted in 10-second intervals. Artifacts (caused by movement, eating, drinking, and scratching) were excluded from the data set. Wake is denoted by a yellow color, NREM is denoted by blue, REM is denoted by red, and artifacts are denoted by white. [Figure 3] The baseline data and sleep recovery data were analyzed for each individual mouse across all the strains.

Most of the methods above were completed at the time that I started my project. I contributed to the manual analysis of the sleep records.

### 2.7 Statistics

Kruskal-Wallis ANOVAs were conducted to identify strain differences in phenotypes. A paired t-tests were conducted to observe within strain differences between the baseline conditions and the recovery conditions across all phenotypes. To compare the recovery from sleep loss across all the strains Pearson correlations were run (comparing the differences in baseline vs recovery sleep across strain). This approach has been used by prior researchers (Diessler et al., 2018).

#### **Chapter 3: Results**

I compiled, analyzed, and interpreted all of the data reported in this section.

#### 3.1 Sleep Phenotypes across Strains

Across 24 strains, all the data for the phenotypes were collected and segmented into

three 6-hour time periods, ZT 6-12, 12-18, and 18-24. ZT 0-6 was excluded from the

dataset since we are comparing sleep states between sleep-replete (spontaneous) and sleep-deprived mice, and ZT 0-6 was the period of sleep deprivation in the sleep-deprived group. In a 12-hr light:12-hr dark cycle, we examine light-phase phenotypes independently of dark-phase phenotypes. Since the light-phase phenotype in recovery is only six hours, we divided all phenotypes into six-hour time bins for ease of comparison. These kinds of time bins are standard in sleep research in animal models.

Kruskal-Wallis ANOVA revealed strain differences in all the phenotypes in each of the 6-hour studied windows in both the baseline condition and the recovery conditions **[Figure 4]** (NREM, REM, Total Sleep, number and duration of bouts of NREM, number and duration of bouts of REM, number and duration of bouts of total sleep, and number of bouts of arousal) with p < 0.0001 for every analysis, and for each phenotype there was a significant difference (p<.05) across strains in both baseline and recovery after analysis with a Kruskal-Wallis test. A Kruskal-Wallis test was performed because the number of animals in each strain were different.

#### 3.2 Between Strain Differences in Sleep Homeostasis

For this experiment, we conducted within-mouse comparisons of baseline and recovery. We used only 22 strains because in two of the strains (NZB and FVB/NJ) only two mice shared the within-mouse condition. Within the strains, all of the data was collected into three 6-hour time periods (6-12, 12-18, and 18-24). ZT 0-6 was excluded from the data pool because there is no corresponding period for comparison in sleep-deprived mice. Mice that were present in the baseline condition but not in the sleep deprivation condition were excluded from the data pool. In addition, individual mice that were present in the sleep deprivation condition but not present in the baseline condition

were removed from the data pool. This was done to reduce possible data confounds as well as making within-mouse comparisons easier.

To see if there was a strain difference in the effect of sleep homeostasis across phenotypes, we first had to normalize our data. In order to do so, we ran Pearson correlations across three-time domains. Our goal was to determine whether the degree of coincidence between the baseline and recovery values within each strain changed significantly as a function of time [Figure 5]. The reason we looked for the Pearson coefficient is that it is one of the most common ways to analyze the relationship between two variables and it was also run in previous sleep studies across inbred mouse strains (Diessler et al., 2018). The higher the positive correlation between baseline and recovery the less robust the homeostatic response to sleep loss and vice versa. In that regard, the correlation is a way of normalizing of sleep homeostasis across strains with significant differences in absolute sleep phenotypes. In this regard, the correlations are an indirect measurement of sleep homeostasis as a function of time. This helps to compensate for the limitation of not having a direct measurement of sleep homeostasis across the strains. The reason we chose three time bins is that since the sleep deprivation is six hours, we expressed recovery in six-hour bins to accommodate the light-dark transition (light phase recovery is only six hours) while keeping the time comparisons consistent. We used a one-way repeated-measures ANOVA (with R-value as the repeated measure) to find potential differences in the correlation of baseline and recovery between three time domains across all the strains combined. This comparison is to determine if there is a significant difference in the correlation between baseline and recovery between the three time domain. In this

comparison, our sample size is the number of strains (22), our independent variable is the time domain (three), and our dependent variable is the R-value. We found that the correlations between baseline and recovery changed across the time bins. We found significant differences in correlation coefficients across time bins between baseline and recovery for two phenotypes: Number of Bouts of NREM (p = 0.0181) and Duration of Bouts of NREM (p = 0.0156) [**Figure 5a**]. We didn't find differences between strains in the correlation coefficients for NREM (p = 0.1501), REM (p = 0.2781), Total Sleep (p = 0.2368), DB of NREM (p = 0.169), Number of Bouts of REM (p = 0.348) and Number of Bouts of Total Sleep (p = 0.348) and Number of Bouts of Arousal (p = 0.7582).

When comparing whether there were any significant differences between baseline and recovery there was quite a variation with some strains not showing any significant differences between baseline and recovery across all three-time points and other strains demonstrating a variety of significant differences in each of the 6-hour time bins **[Figure 5b]**. This finding is true across all of the 10 listed phenotypes, although DB REM had the fewest significant differences shown between baseline and recovery. *This variation in recovery from sleep deprivation across strains suggests that the homeostat may be regulated by genetic mechanisms*.

Since we initially compared these phenotypes using relative data, via the Pearson R values, we also examined whether there would be any differences in the phenotypes across the strains using absolute values. We examined sleep fragmentation, which is often used as a measurement of sleep homeostasis. We took the four sleep fragmentation phenotypes (NREM Bouts, REM Bouts, Total Sleep Bouts, and Number

of Bouts of Arousal) of each animal in each strain, and we subtracted the recovery values from the baseline values in each of the studied 6-hour intervals [Figure 6]. We then ran a Kruskal-Wallis ANOVA to examine strain differences regarding this change between baseline and recovery. The analysis yielded that there were significant differences in this change between the strains. With the number of bouts of NREM there was a significance difference between all strains in ZT 7-12 (p < 0.001), 13-18 (p < 0.001) (0.001) and (19-24) (p = (0.0212) [Figure 6a]. With the number of bouts of REM, a significant difference between strains was found in ZT 6-12 (p < 0.0001), 12-18 (p =0.0032) but surprisingly not for 18-24 (p = 0.1148) [Figure 6b]. With the number of bouts of Total Sleep, a significant difference between strains was found in ZT 6-12 (p < 0.0001), 12-18 (p = 0.0011), and 18-24 (p = 0.0286) [Figure 6c]. Looking at the number of bouts of arousal, a significant difference between strains was found in ZT 6-12 (p = 0.0018), 12-18 (p = 0.0014), and 19-24 (p = 0.0091) [Figure 6d]. These findings suggest that the homeostatic response to sleep loss is indeed genetically regulated, and that regulation is likely encoded by adjustments to sleep-wake fragmentation after sleep loss.

#### Discussion

The mechanisms that underlie sleep homeostasis are currently not fully understood. To better understand how genetic heterogeneity contributes to sleep homeostasis, we analyzed data to examine differences between strains across 10 different phenotypic conditions (both in baseline and in recovery). We also examined differences within strains (between baseline and recovery) across 10 different phenotypes and to see whether there was a difference in sleep phenotype response between baseline and recovery. All of the data was separated into three time periods: ZT 6-12, ZT 12-18 and ZT 18-24.

Differences in between-strain comparisons let us know which specific phenotypes have a high degree of genetic heterogeneity. The finding of strain differences would suggest that genetic differences (as opposed to environmental perturbations) underlie these variations in sleep phenotypes. In order to explore this, I conducted a between-strain analysis of variance across three time windows (ZT 6-12, 12-18 and 18-24) in their baseline condition. Subsequently, I conducted a between-strain analysis of variance across all three time windows in the recovery condition. In addition, I compared the sleep phenotypes across all strains in their baseline condition to their recovery condition across three different time bins to see if there was a phenotypic difference in the dissipation of sleep pressure. In order to do this, we ran Pearson's correlations to normalize the dataset for comparison.

# Ten sleep phenotypes exhibited significant differences across strains during baseline and recovery conditions

For the between-strain analyses, there was a significant difference between the strains for all phenotypes in the baseline condition. For the between-strain analysis in the recovery condition, we also found a significant difference in all of the strains in every phenotype. These differences are illustrated in the graphs [**Fig. 4**] which show several examples of how different the strains are from each other. Prior studies also substantiate this evidence as different strains are expected to spend different amounts of time sleeping and displaying sleep phenotypes such as NREM sleep (Hoekstra et al., 2019) and REM sleep (Niwa et al., 2018). *These data demonstrate robust strain* 

difference among all 10 sleep phenotypes in both the baseline and recovery conditions. They also indicate that several key sleep phenotypes are genetically influenced and suggest that future studies can interrogate these stains to look for specific genes that regulate sleep. However, it is important to note that the strain differences observed during baseline do not necessarily recapitulate the strain differences that occur during recovery.

In order to account for the effects of sleep loss on strain differences during recovery, we examined correlations between baseline and recovery phenotypes within strains. This provides a high throughput measurement of the ability of sleep deprivation to alter strain differences in these phenotypes. This also provides a novel and useful tool to examine sleep homeostasis, which is the ability to recover from sleep loss. This analysis will reveal whether or not there is a genetic component to sleep homeostasis across strains.

# Two sleep phenotypes exhibited significant correlations between baseline and recovery across three-time windows

We ran a Pearson's correlation for the phenotypes between baseline and recovery for each animal in three different 6-hour intervals. The R-values gained from the statistical analysis were then compared across all the time intervals for each of the strains. The reason that we used Pearson's correlation is that the different phenotypes are expressed in different units, and all have different responses to sleep loss. In order to perform a comprehensive analysis of the responses to sleep loss across all phenotypes, we chose to apply a normalization that avoids biasing any of the specific phenotypes. Analyzing the recovery from sleep over time is the most efficient method to

detect the effects of sleep deprivation across a panel of numerous different phenotypes with different measurement criteria (Diessler et al., 2018). This statistical analysis revealed significant differences in 2 of the 10 phenotypes: Number of Bouts of NREM and Duration of Bouts of Total Sleep. The remaining 8 phenotypes did not yield any significant differences, as is compatible with the findings in (Diessler et al., 2018) and (Jan et al., 2020). This finding suggests that the genetic mechanisms that are responsible for the homeostatic ability to recover from sleep loss are not reflected in the genetic variation of most of the phenotypes under the current analysis. However, with the number of bouts of NREM and duration of bouts of total sleep showing a significant difference across the strains, this is a promising result for QTL mapping or genetic mapping. To date, few homeostatic sleep phenotypes that account for the differences between baseline and recovery have yielded promising genetic targets (Diessler et al., 2018; Franken et al., 2001; Maret et al., 2007). The finding that these differences in correlations for the number of bouts of NREM and duration of bouts of total sleep suggest that these two specific phenotypes may yield valuable insights into the genes that regulate sleep homeostasis.

# Sleep-wake fragmentation is a promising measure of sleep homeostasis for genetic mapping studies

Few studies in mammals have examined sleep-wake fragmentation as a measure of sleep homeostasis, in order to conduct forward genetic approaches. When looking at the absolute values of sleep fragmentation, and the differences between baseline and recovery within each of the strains, we found significant differences in many of the strains in all of the phenotypes. This suggests that sleep deprivation produced

significant effects on sleep fragmentation in many inbred strains, but not all of them. However, when we examined recovery with respect to baseline sleep across strains, few phenotypes exhibited significant differences. This finding may provide insights into future studies that seek to explore the role of genetics and sleep homeostasis in mouse models.

We examined sleep phenotypes associated with sleep fragmentation as a measurement of sleep homeostasis. These phenotypes examined recovery relative to baseline. We found that among all homeostatic sleep fragmentation phenotypes, there were significant differences across strains (Fig. 6). In all the fragmentation phenotypes, there were more bouts (either in NREM, REM, Total Sleep or Arousal) in the baseline condition as opposed to the recovery condition but after ZT 12 the opposite becomes more apparent. This is the first inbred strain study focusing on sleep fragmentation as a measure of sleep homeostasis in mammals, and the glaring differences between strains is promising. The data suggests that sleep fragmentation may play a larger role in the genetic regulation of sleep homeostasis than previously thought and opens the door to future exploration of the potential genetic components underlying it.

The next step for this study utilizes a special mouse model that could help with trying to identify a potential genetic component of sleep homeostasis, namely the BxD mouse. An experiment in 2018 (Diessler et al., 2018) was run conducting QTL analysis on 33 BxD mice and they found that sleep traits in mice were heritable and could possibly even be identified in specific loci. The next step for this study is to record the BxD model in both the baseline and recovery conditions and add the BxD data with the rest of the strains. Then QTL analysis will be conducted between the 24 strains and an

investigation will be underway to find specific loci correlated with sleep traits. Other approaches to looking into strain differences, including the new data with the BxD model, could be looking into the Homer1a gene (Mackiewicz et al., 2008) which is associated with Fragile X syndrome or looking at a connection between sleep and Vitamin A signaling (Tafti, 2007).

There also has been several studies on sleep homeostasis which could provide the foundation for future research. Specifically, Dr. Liza Ashbrook (Ashbrook et al., 2020) found that genetic variations in humans could lead to a phenotype of short sleep (Familial natural short sleep) where people would only get 4.5-6 hours of sleep as a result of the DEC2 gene, P385R. Dr. Geraldine Mang and Dr. Paul Franken found that sleep homeostasis is likely to have a myriad of different pathways involved and that sleep homeostasis can vary by sex within specific animal strains (Mang, G. & Franken P., 2015). This data suggests that sleep homeostasis may be involved in other pathways regarding hormones. Dr. Maxime Jan found that the cortical expression of the core clock genes Npas2 and Clock seemed to be driven by sleep-wake homeostasis and didn't seem to have a circadian drive (Jan et al., 2020). Research into sleep homeostasis and its potential mechanisms has great promise and there are a number of different approaches to better understand it. Bettering our understanding of the genetic components of sleep has the potential to help us better understand how sleep disorders functions and possibly even assist in developing treatments for them.

### Figure Legends Figure 1: Head Mount and Commutator

Provided on the left is an image of how the mice look after a complete surgery with the head mount sticking upwards. On the right is an image of the commutator that connects to the head mount to relay EEG data to the computer.

### Figure 2: Sleep Deprivation Container and Example Novel Object

On the left is an image of the mouse cage during sleep deprivation. They would have bedding, food and water and would be kept awake by gently moving the cage, introducing a novel object (right image) or disturbing bedding. A novel object like a paintbrush would be of great assistance as it would allow for disturbing of the bedding without having to place my hand inside the cage.

## Figure 3: An Example of Scored Sleep-Wake Architecture

This image showcases how the sleep-wake architecture of a 30-second epoch is scored using both EEG and EMG. Wake is yellow, NREM is blue, and REM is red as color coding helps with identification when going through a file. The EMG is beneath the EEG and shows movement during the wake period.

Figure 4: Comparison of Strain Differences among all 12 Sleep Phenotypes in both Baseline and Recovery Conditions.

4a. Comparison of Strain Differences in NREM in the baseline and recovery Conditions.

A one way ANOVA was run across all strains in NREM ZT 6-12, 12-18, and 18-24 which found significant differences in NREM (p< 0.001) for all time points across both baseline and recovery.

# 4b. Comparison of Strain Differences in REM in both the Baseline and Recovery Conditions.

A one way ANOVA was run across all strains in ZT 6-12, 12-18, and 18-24 in baseline and recovery, yielding significance in in REM at all time points (p < 0.0001).

# 4c. Comparison of Strain Differences in Total Sleep in both the Baseline and Recovery Conditions.

A one-way ANOVA was run comparing all strains in baseline and recovery for ZT 6-12, 12-18 and 18-24. The test showed significant differences in Total Sleep amongst the strains (p < 0.001).

# 4d. Comparison of Strain Differences in Number of Bouts of NREM in both the Baseline and Recovery Conditions.

In order to see if there were strain differences in the number of bouts of NREM we performed a Kruskal-Wallis ANOVA in ZT 6-12 (p < 0.0001), 12-18 (p < 0.0001) and 18-24 (p < 0.0001) and found that the differences between strains were significant. This was also true in the recovery condition.

# 4e. Comparison of Strain Differences in Duration of Bouts of NREM in both the Baseline and Recovery Conditions.

A significant difference across strains during duration of bouts in baseline was found in ZT 6-12 (p < 0.0001), 12-18 (p < 0.0001) and 18-24 (p < 0.0001). A significant difference across strains during recovery was found in ZT 6-12 (p < 0.0001), 12-18 (p < 0.0001) and 18-24 (p < 0.0001).

# 4f. Comparison of Strain Differences in Number of Bouts of REM in both the Baseline and Recovery Conditions.

A significant difference across strains in number of bouts of REM during baseline was also found in ZT 6-12 (p < 0.0001), 12-18 (p < 0.0001) and 18-24 (p < 0.0001) according to a one way ANOVA. A significant difference across strains during recovery was found in ZT 6-12 (p < 0.0001), 12-18 (p < 0.0001) and 18-24 (p < 0.0001) according to a one way ANOVA.

# 4g. Comparison of Strain Differences in Duration of Bouts of REM in both the Baseline and Recovery Conditions.

A significant difference across strains in duration of Bouts of REM during baseline was also found in ZT 6-12 (p < 0.0001), 12-18 (p < 0.0001) and 18-24 (p < 0.0001) according to a one way ANOVA. A significant difference across strains during recovery was found in ZT 6-12 (p < 0.0001), 12-18 (p < 0.0001) and 18-24 (p < 0.0001) according to a one way ANOVA.

# 4h. Comparison of Strain Differences in Number of Bouts of Total Sleep in both the Baseline and Recovery Conditions.

A significant difference across strains in number of bouts of total sleep during baseline was also found in ZT 6-12 (p < 0.0001), 12-18 (p < 0.0001) and 18-24 (p < 0.0001)

according to a one way ANOVA. A significant difference across strains during recovery was found in ZT 6-12 (p < 0.0001), 12-18 (p < 0.0001) and 18-24 (p < 0.0001) according to a one way ANOVA.

# 4i. Comparison of Strain Differences in Duration of Bouts of Total Sleep in both the Baseline and Recovery Conditions.

A significant difference across strains in duration of bouts of total sleep during baseline was also found in ZT 6-12 (p < 0.0001), 12-18 (p < 0.0001) and 18-24 (p < 0.0001) according to a one way ANOVA. A significant difference across strains during recovery was found in ZT 6-12 (p < 0.0001), 12-18 (p < 0.0001) and 18-24 (p < 0.0001) according to a one way ANOVA.

# 4j. Comparison of Strain Differences in Number of Bouts of Arousal in both the Baseline and Recovery Conditions.

A significant difference across strains in number of bouts of arousal during baseline was also found in ZT 6-12 (p < 0.0001), 12-18 (p < 0.0001) and 18-24 (p < 0.0001) according to a one way ANOVA. A significant difference across strains during recovery was found in ZT 6-12 (p < 0.0001), 12-18 (p < 0.0001) and 18-24 (p < 0.0001) according to a one way ANOVA.

### Figure 5: Tables Comparing Strains across Three 6 hour Time Bins

### 5a: Tables Comparing the r-values of Strains across Three 6 hour Time Bins

These tables show the correlation between baseline and recovery for each strain of mouse in each of the six-hour time bins. They are organized (from left to right) as such:

NREM, REM, Total Sleep, Number of Bouts of NREM, Duration of Bouts of NREM, Number of Bouts of REM, Duration of Bouts of REM, Number of Bouts of Total Sleep, Duration of Bouts of Sleep and Number of Bouts of Arousal.

### 5b: Tables Comparing the p-values of Strains across Three 6 hour Time Bins

These tables show the correlation between the baseline and recovery for each of strain of mouse in each other the six hour time bins. They are organized similar to that of figure 6a. Significance is color-coded in these tables with green being significant at p < 0.05, yellow being significant at p < 0.01, and red being significant at p < 0.001.

Figure 6: Analysis of Baseline - Recovery in Sleep Fragmentation Phenotypes between Strains

6a. Analysis of Baseline – Recovery in Sleep Fragmentation between Strains in Number of Bouts of NREM

With the number of bouts of NREM there was a significance difference between all strains in ZT 7-12 (p < 0.001), 13-18 (p < 0.001) and 19-24 (p = 0.0212).

# 6b. Analysis of Baseline – Recovery in Sleep Fragmentation between Strains in Number of Bouts of REM

With the number of bouts of REM, a significant difference between the baseline and recovery of all strains was found in ZT 6-12 (p < 0.0001), 12-18 (p = 0.0032) but surprisingly not for 18-24 (p = 0.1148).

6c. Analysis of Baseline – Recovery in Sleep Fragmentation between Strains in Number of Bouts of Total Sleep

With the number of bouts of Total Sleep, a significant difference between baseline and recovery was found in ZT 6-12 (p < 0.0001), 12-18 (p = 0.0011), and 18-24 (p = 0.0286).

# 6d. Analysis of Baseline – Recovery in Sleep Fragmentation between Strains in Number of Bouts of Arousal

Looking at the number of bouts of arousal, a significant difference between baseline and recovery across the strains were found in ZT 6-12 (p = 0.0018), 12-18 (p = 0.0014), and 19-24 (p = 0.0091).

# Figures

Figure 1: Head Mount + Commutator



Figure 2: Sleep Deprivation Set Up



Figure 3: An Example of Scored Sleep-Wake Architecture



Figure 4: Comparison of Strain Differences among all 10 Sleep Phenotypes in both Baseline and Recovery Conditions

# 4a. NREM 6-12 NREM \*\*\* \*\*\*













## 4c. Total Sleep







### 4d. Number of Bouts of NREM







## 4e. Duration of Bouts of NREM











BSLN REC

# 4g. Duration of Bouts of REM















# 4i. Duration of Bouts of Total Sleep







## 4j. Number of Bouts of Arousal







## *Figure 5: Tables Comparing Strains across Three 6 Hour Time Bins*

Figure 5a: Tables of R Values Comparing Strains across Three 6 Hour Time Bins

NREM					REM							TS			
Strain 💽	(06-12) 💌	(12-18) 💌	(18-24) 💌		Strain 💌	(06-12	) - (1	12-18) 💌	(18-	24) 💌		Strain 💌	(06-12) 💌	(12-18) 💌	(18-24) 💌
129S1/Svlm.	0.39855	0.44489	-0.17498		129S1/Sv	0.375	597	0.10711	0.6	62694		129S1/Svl	0.36333	0.33898	-0.05642
A/J	0.1157	0.73019	-0.21365		A/J	0.645	547 -	0.66284	-0.4	48452		A/J	-0.01155	0.47096	-0.37448
AKR/J	-0.19972	0.64947	0.4864		AKR/J	0.527	796	0.18318	0.7	76821		AKR/J	-0.11496	0.62466	0.6715
BTBR T+ Itp	0.66189	0.58469	-0.08677		BTBR T+	0.063	385	0.15321	0.5	58721		BTBR T+ I	0.60277	0.5427	0.01372
BUB/BnJ	0.90738	0.56914	0.86361		BUB/BnJ	0.513	343	0.06216	0.7	79977		BUB/BnJ	0.89278	0.38093	0.86911
C3H/HeJ	-0.55311	0.75641	0.72412		C3H/HeJ	-0.529	915	0.86243	0.6	61978		C3H/HeJ	-0.71317	0.90671	0.67678
C57BL/6J	-0.88169	-0.53145	0.72806		C57BL/6	0.346	584	0.69573	0.8	87297		C57BL/6J	-0.74073	-0.37818	0.74346
C57L/J	0.37196	0.9458	0.70072		C57L/J	0.774	132	0.90457	0	.2466		C57L/J	0.26927	0.94394	0.66668
C58/J	0.04058	0.0676	0.9382		C58/J	0.605	581	0.7729	0.9	94982		C58/J	0.6282	0.86305	0.77192
CAST/EiJ	0.74539	0.45379	-0.04654		CAST/EiJ	0.624	195	0.56534	0.5	57103		CAST/EiJ	0.70273	0.43651	0.2783
CBA/J	0.81365	0.9285	0.64229		CBA/J	0.680	037	0.86499	0	.6436		CBA/J	0.78043	0.94459	0.67194
CZECHII/Ei.	0.63601	-0.5241	0.83014		CZECHII/	E 0.956	694 -	0.92447	0.5	55492		CZECHII/E	0.62968	-0.60752	0.94209
DBA/2J	0.61514	0.75479	0.93144		DBA/2J	0.982	285	0.75235	0.8	83623		DBA/2J	0.80435	0.76565	0.92198
KK/HIJ	0.27386	0.70157	0.2779		KK/HIJ	0.797	783	0.69286	0.3	32487		KK/HIJ	0.44679	0.71525	0.28507
MOLF/EiJ	0.00744	0.99327	0.4939		MOLF/Ei	0.89	952	0.92384	0.6	61384		MOLF/EiJ	0.26748	0.99029	0.51167
NOD/ShiLtJ	0.25591	0.7772	0.45127		NOD/ShiL	-0.153	318	0.69825	0.8	80674		NOD/ShiL	-0.0123	0.77702	0.53785
PL/J	0.94318	0.76887	0.89819		PL/J	0.960	071 -	0.43265	0.7	75736		PL/J	0.90044	0.73986	0.89021
PWD/PhJ	0.86913	0.59117	0.88		PWD/PhJ	0.23	376	0.79143	0.6	68414		PWD/PhJ	-0.93867	0.13228	0.85123
SJL/J	0.41503	-0.10833	-0.2484		SJL/J	-0.378	347	0.84201	0.0	09236		SJL/J	0.23181	-0.02762	-0.38949
SM/J	0.89728	0.90733	0.08299		SM/J	-0.319	985	0.99978	-0.7	75939		SM/J	0.92064	0.93769	-0.60345
SWR/J	0.81706	0.96533	0.73506		SWR/J	-0.951	122	0.28947	-0	.8508		SWR/J	0.8041	0.97602	0.76741
WSB/EiJ	0.94876	0.78514	0.8646		WSB/EiJ	0.833	389 -	0.16607	0.4	45474		WSB/EiJ	0.91587	0.76508	0.83022
NB NREM				DB NREM					P	NB REM				DB RE	M
Strain 💌 (06	5-12) - (12-	18) 💌 (18-24	*	Strain 💌	(06-12) • (	12-18) 💌	(18-24	1) 👻	S	Strain 💌	(06-12) -	(12-18) - (1	8-24) 💌	Strain	· (06-12) ·
129S1/Svlm0	0.31645 0.3	29301 -0.31	585	129S1/Svl	-0.05278	0.536276	-0.	219	1	129S1/Svli	0.365399	0.395926 0	.434836	12951	/Svli -0.25358

Strain 💌	(06-12) -	(12-18) 💌	(18-24) 💌	Strain 💌	(06-12) 💌	(12-18) 💌	(18-24) 💌		Strain 💌	(06-12) 💌	(12-18) -	(18-24) 💌	Strain 💌	(06-12) 💌	(12-18) 💌	(18-24) -
129S1/Svlm.	-0.31645	0.29301	-0.31585	129S1/Svli	-0.05278	0.536276	-0.219		129S1/Svli	0.365399	0.395926	0.434836	129S1/Svli	-0.25358	-0.07301	0.165311
A/J	0.309759	0.448425	0.928879	A/J	0.811568	-0.05647	0.319427		A/J	0.521951	-0.18798	-0.89348	A/J	0.760644	0.811263	-0.00261
AKR/J	0.373728	0.298311	0.077278	AKR/J	-0.05465	-0.00733	-0.11056		AKR/J	0.44664	0.429838	0.760891	AKR/J	0.186465	-0.09294	0.526389
BTBR T+ Itp	0.25045	0.569608	-0.42569	BTBR T+ I	-0.17853	-0.27171	-0.28176		BTBR T+ I	0.229524	0.609523	0.119323	BTBR T+ I	0.322632	0.781001	0.453637
BUB/BnJ	0.701481	0.42222	0.596672	BUB/BnJ	0.93063	0.395392	0.613869		BUB/BnJ	0.565708	0.571718	0.363069	BUB/BnJ	0.545954	0.358376	0.74312
C3H/HeJ	0.931925	0.812535	0.681135	C3H/HeJ	0.455358	0.603989	0.703244		C3H/HeJ	0.993399	0.994039	-0.3143	C3H/HeJ	-0.10439	0.887977	0.281144
C57BL/6J	0.728309	0.652377	0.371032	C57BL/6J	0.941619	-0.09038	0.874788		C57BL/6J	0.885134	0.242065	0.824131	C57BL/6J	0.737542	0.282104	0.470843
C57L/J	0.908843	0.886705	0.819313	C57L/J	0.425395	0.810639	0.021482		C57L/J	0.37368	0.893728	0.292588	C57L/J	-0.04239	0.473331	0.118235
C58/J	0.711417	0.536075	0.929525	C58/J	0.619238	0.304463	-0.15618		C58/J	0.300369	0.563463	0.920747	C58/J	0.368457	-0.14595	0.733407
CAST/EiJ	0.717776	0.563641	0.382106	CAST/EiJ	0.617477	0.426902	0.176294		CAST/EiJ	0.452818	0.580977	0.481942	CAST/EiJ	0.096222	0.0248	0.704827
CBA/J	0.978289	0.793793	0.796424	CBA/J	0.923639	0.849693	0.829141		CBA/J	0.594053	0.975227	-0.07297	CBA/J	0.043367	0.805072	0.414105
CZECHII/EiJ	0.914318	-0.24019	0.993134	CZECHII/E	0.996268	0.918459	-0.04812		CZECHII/E	-0.14286	-0.86603	0.755929	CZECHII/E	0.983046	-0.98381	0.804818
DBA/2J	-0.19242	0.988509	0.154453	DBA/2J	-0.13352	0.425685	0.650417		DBA/2J	0.432549	0.86267	0.516025	DBA/2J	0.616514	0.18207	0.184585
KK/HIJ	0.795752	0.847166	0.004025	KK/HIJ	0.35315	0.513164	0.445149		KK/HIJ	0.582532	0.520389	0.252965	KK/HIJ	0.617553	0.784777	0.680785
MOLF/EiJ	-0.26041	0.939263	0.63876	MOLF/EiJ	0.069386	0.845389	0.396553		MOLF/EiJ	0.892178	0.762161	0.010183	MOLF/EiJ	0.423039	0.889889	0.64248
NOD/ShiLtJ	0.850537	0.765571	0.860623	NOD/ShiLt	0.758003	0.43672	0.719282	1	NOD/ShiLt	0.132167	0.765332	0.590561	NOD/ShiLt	0.517399	0.082968	0.16938
PL/J	0.639957	0.371993	0.90798	PL/J	0.495705	0.832864	0.97316		PL/J	0.717607	0.774913	0.75585	PL/J	0.909526	0.8784	0.085929
PWD/PhJ	0.765987	0.955114	0.996655	PWD/PhJ	0.612382	0.96736	0.545335		PWD/PhJ	0.279553	0.839109	0.60553	PWD/PhJ	0.316442	0.185966	0.61993
SJL/J	0.089669	-0.01908	-0.12394	SJL/J	0.360571	0.634926	-0.32138		SJL/J	0.098943	0.839516	0.859449	SJL/J	0.301519	0.467953	0.606898
SM/J	-0.9519	0.732331	-0.82603	SM/J	0.999566	0.062221	0.306849		SM/J	0.802955	0.995567	-0.05241	SM/J	0.523797	-0.41324	0.893161
SWR/J	-0.16733	0.991442	0.907831	SWR/J	-0.6202	-0.3651	0.876782		SWR/J	-0.67193	0.06647	-0.77771	SWR/J	0.680885	0.750487	-0.88396
WSB/EiJ	0.8955	0.59254	0.816575	WSB/EiJ	0.942881	0.914601	-0.45242		WSB/EiJ	0.505514	0.160645	0.475247	WSB/EiJ	0.746109	-0.23389	0.347767

NB TS				DB TS				N B Ar			
Strain 🛛 💌	(06-12) 💌	(12-18) 💌	(18-24)	Strain 💌	(06-12) 💌	(12-18) 💌	(18-24)	Strain 💌	(06-12) 💌	(12-18) 💌	(18-24) 💌
129S1/Svlm	-0.05493	0.08042	-0.15321	129S1/Svli	-0.09438	0.408294	-0.31313	129S1/Svli	-0.17176	0.538473	0.305233
A/J	0.134066	0.683366	0.914969	A/J	0.788485	0.253697	0.428191	A/J	0.020758	0.709306	0.120681
AKR/J	0.42278	0.590358	0.708354	AKR/J	-0.11411	-0.10446	0.407348	AKR/J	0.653031	0.379946	-0.24014
BTBR T+ Itp	0.341582	0.688705	-0.25333	BTBR T+ I	-0.17303	-0.0931	-0.20148	BTBR T+ I	0.395141	0.70395	0.424995
BUB/BnJ	0.612038	0.451449	0.506327	BUB/BnJ	0.896229	0.452446	0.46375	BUB/BnJ	0.72008	0.203719	0.695123
C3H/HeJ	0.961814	0.863965	0.761728	C3H/HeJ	0.539686	0.703629	0.776882	C3H/HeJ	0.832284	0.725555	0.412068
C57BL/6J	0.817491	0.518407	0.475546	C57BL/6J	0.975786	0.862461	0.899368	C57BL/6J	0.259481	0.933368	0.720438
C57L/J	0.926081	0.892491	0.738533	C57L/J	0.693472	-0.09644	0.338499	C57L/J	0.743346	0.691489	0.866181
C58/J	0.839883	0.51344	0.937485	C58/J	0.673533	0.326001	0.357128	C58/J	0.926905	0.27073	0.673547
CAST/EiJ	0.776596	0.489126	0.385752	CAST/EiJ	0.561595	0.514387	0.177783	CAST/EiJ	0.821531	0.101482	0.44549
CBA/J	0.954273	0.793706	0.571544	CBA/J	0.941229	0.906875	0.626347	CBA/J	0.896396	0.864594	0.768369
CZECHII/EiJ	0.980481	-0.29785	0.978983	CZECHII/E	0.996106	0.526144	-0.4632	CZECHII/E	0.762875	-0.73241	0.863365
DBA/2J	-0.00917	0.972643	0.180925	DBA/2J	0.648088	0.031858	0.617934	DBA/2J	0.073229	0.98555	0.141162
KK/HIJ	0.684169	0.775705	0.095601	KK/HIJ	0.188649	0.257975	0.28667	KK/HIJ	0.920158	0.880374	0.359976
MOLF/EiJ	-0.52479	0.925644	0.438493	MOLF/EiJ	-0.38089	0.872057	0.505435	MOLF/EiJ	0.573945	0.845378	0.929546
NOD/ShiLtJ	0.726233	0.898004	0.783001	NOD/ShiLt	0.74959	0.246124	0.652459	NOD/ShiLt	0.139931	0.80591	0.832738
PL/J	0.597719	0.555523	0.886612	PL/J	0.551671	0.926661	0.929652	PL/J	0.65028	-0.57971	0.679971
PWD/PhJ	0.735344	0.936959	0.047377	PWD/PhJ	0.633279	0.76707	0.987522	PWD/PhJ	-0.6787	0.996838	0.981348
SJL/J	0.038928	0.169428	0.404779	SJL/J	0.299704	0.661183	-0.0617	SJL/J	-0.21008	0.006796	0.234903
SM/J	0.453921	0.903799	-0.97073	SM/J	0.980693	0.519853	0.471471	SM/J	0.831157	-0.82199	-0.06102
SWR/J	-0.15435	0.783382	0.047377	SWR/J	-0.64442	-0.4678	0.647828	SWR/J	0.256257	-0.88385	0.936244
WSB/EiJ	0.613746	0.525765	0.907463	WSB/EiJ	0.928068	0.690323	0.038251	WSB/EiJ	0.41787	0.713984	0.715679

	Figure 5b: Tab	les of P Values	Comparing I	Recovery-Baselii	ne in Strains acro	oss Three 6 Hour	<sup>.</sup> Time Bins
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NREM				REM				TS			
Strain	<b>•</b> (6-12) <b>•</b>	(12-18) 🔻	(18-24) 💌	Strain 💽	(6-12) 💌	(12-18) 🔻	(18-24) 💌	Strain	• (6-12) •	(12-18) 💌	(18-24) 🔻
129S1/Svl	mJ 0.3758	0.3172	0.7075	129S1/Svlm	nJ 0.4059	0.8192	0.1319	129S1/Svlr	nJ 0.4231	0.457	0.9044
A/J	0.853	0.1613	0.7301	A/J	0.2395	0.2227	0.4082	A/J	0.9853	0.4233	0.5346
AKR/J	0.6677	0.1144	0.2684	AKR/J	0.2232	0.6942	0.0437	AKR/J	0.8061	0.1337	0.0986
BTBR T+ It	pr 0.0265	0.0589	0.7997	BTBR T+ Itp	or 0.8521	0.6529	0.0575	BTBR T+ It	pr 0.0497	0.0845	0.9681
BUB/BnJ	0.0048	0.1824	0.0122	BUB/BnJ	0.2386	0.8947	0.0308	BUB/BnJ	0.0068	0.3992	0.0111
C3H/HeJ	0.4469	0.2436	0.2759	C3H/HeJ	0.4708	0.1376	0.3802	C3H/HeJ	0.2868	0.0933	0.3232
C57BL/6J	0.1183	0.4685	0.2719	C57BL/6J	0.6532	0.3043	0.127	C57BL/6J	0.2593	0.6218	0.2565
C57L/J	0.4113	0.0013	0.0795	C57L/J	0.041	0.0051	0.594	C57L/J	0.5593	0.0014	0.1019
C58/J	0.0722	0.0153	0.0753	C58/J	0.1494	0.0416	0.0011	C58/J	0.1308	0.0124	0.042
CAST/EiJ	0.0338	0.2587	0.9129	CAST/EiJ	0.0976	0.1442	0.1393	CAST/EiJ	0.0782	0.3275	0.5456
CBA/J	0.0579	0.0075	0.169	CBA/J	0.1369	0.0261	0.1679	CBA/J	0.067	0.0045	0.1438
CZECHII/E	iJ 0.5612	0.6488	0.3765	CZECHII/EiJ	0.1875	0.249	0.6255	CZECHII/Ei	J 0.5664	0.5843	0.2177
DBA/2J	0.2694	0.1403	0.0213	DBA/2J	0.0027	0.1423	0.0776	DBA/2J	0.1008	0.1313	0.0259
кк/ніј	0.5116	0.0525	0.5052	кк/ніј	0.0177	0.0568	0.4324	кк/нIJ	0.2671	0.0461	0.4937
MOLF/EiJ	0.9905	0.0007	0.3977	MOLF/EiJ	0.0401	0.0249	0.2708	MOLF/EiJ	0.6635	0.0011	0.3782
NOD/ShiL	tJ 0.5407	0.0232	0.2617	NOD/ShiLtJ	0.7172	0.0541	0.0155	NOD/ShiLt	J 0.9769	0.0233	0.1691
PL/J	0.0161	0.1287	0.0384	PL/J	0.0093	0.4668	0.1381	PL/J	0.0371	0.1529	0.0429
PWD/PhJ	0.1309	0.4088	0.12	PWD/PhJ	0.7624	0.2086	0.3159	PWD/PhJ	0.0613	0.8677	0.1488
SJL/J	0.3545	0.8172	0.5912	SJL/J	0.4025	0.0175	0.8439	SJL/J	0.617	0.9531	0.3878
SM/J	0.2911	0.2762	0.9471	SM/J	0.7927	0.0133	0.451	SM/J	0.2553	0.2259	0.5876
SWR/J	0.3912	0.1681	0.4743	SWR/J	0.1997	0.813	0.3522	SWR/J	0.4053	0.1397	0.4431
WSB/EiJ	0.0039	0.0643	0.0263	WSB/EiJ	0.0391	0.7532	0.3649	WSB/EiJ	0.0103	0.0763	0.0408

NB NREM				DB NREM				NB REM				DB REM			
Strain 🔤	(6-12) 💌	(12-18) 💌	(18-24) 💌	Strain	<b>• (6-12)</b>	(12-18) 💌	(18-24) 💌	Strain	<b>•</b> (6-12)	· (12-18) 💌	(18-24) 💌	Strain 💌	(6-12) 💌	(12-18) 💌	(18-24) 💌
129S1/Svlm	nJ 0.4893	0.5236	0.4901	129S1/Svl	m. 0.9105	0.2146	0.6371	12951/5	ivlmJ 0.420	2 0.3793	0.3296	129S1/Svlm	0.5832	0.8764	0.7232
A/J	0.612	0.4488	0.0225	A/J	0.0954	0.9281	0.6003	A/J	0.36	7 0.7621	0.0411	A/J	0.1354	0.0956	0.9967
AKR/J	0.4089	0.5158	0.8692	AKR/J	0.9074	0.9876	0.8135	AKR/J	0.315	1 0.3358	0.047	AKR/J	0.6889	0.8429	0.2248
BTBR T+ Itp	or 0.4576	0.0674	0.1918	BTBR T+ It	pr 0.5994	0.419	0.4012	BTBR T+	ltpr 0.497	0.0465	0.7268	BTBR T+ Itp	r 0.3332	0.0045	0.1611
BUB/BnJ	0.079	0.3454	0.1573	BUB/BnJ	0.0023	0.38	0.1426	BUB/Bn	J 0.185	6 0.1799	0.4234	BUB/BnJ	0.2049	0.4299	0.0556
C3H/HeJ	0.0681	0.1875	0.3189	C3H/HeJ	0.5446	0.396	0.2968	C3H/He	J 0.006	6 0.006	0.6857	C3H/HeJ	0.8956	0.112	0.7189
C57BL/6J	0.2717	0.3476	0.629	C57BL/6J	0.0584	0.9096	0.1252	C57BL/6	5J 0.114	9 0.7579	0.1759	C57BL/6J	0.2625	0.7179	0.5292
C57L/J	0.0046	0.0078	0.0241	C57L/J	0.3414	0.027	0.9635	C57L/J	0.40	9 0.0067	0.5243	C57L/J	0.9281	0.2834	0.8007
C58/J	0.073	0.2148	0.0024	C58/J	0.1381	0.5067	0.7381	C58/J	0.512	8 0.1878	0.0033	C58/J	0.4161	0.7549	0.0607
CAST/EiJ	0.045	0.1457	0.3502	CAST/EiJ	0.1029	0.2915	0.6762	CAST/E	J 0.259	9 0.131	0.2265	CAST/EiJ	0.8207	0.9535	0.0509
CBA/J	0.0007	0.0594	0.0579	CBA/J	0.0085	0.0322	0.0413	CBA/J	0.213	7 0.0009	0.8907	CBA/J	0.935	0.0533	0.4143
CZECHII/Ei.	J 0.2655	0.8456	0.0746	CZECHII/E	iJ 0.055	0.2589	0.9694	CZECHI	/EiJ 0.908	7 0.3333	0.4544	CZECHII/EiJ	0.1174	0.1147	0.4045
DBA/2J	0.7565	0.0015	0.8041	DBA/2J	0.8305	0.4748	0.2347	DBA/2J	0.46	7 0.0598	0.3734	DBA/2J	0.2681	0.7695	0.7663
кк/нIJ	0.0182	0.0079	0.9925	кк/ніј	0.3908	0.1934	0.2691	KK/HIJ	0.129	7 0.1861	0.5455	KK/HIJ	0.1028	0.0211	0.0631
MOLF/EiJ	0.6722	0.0178	0.246	MOLF/EiJ	0.9117	0.0713	0.5087	MOLF/E	iJ 0.041	8 0.1342	0.987	MOLF/EiJ	0.4779	0.0431	0.2424
NOD/ShiLt	0.0074	0.0268	0.0061	NOD/ShiL	U 0.0293	0.2793	0.0443	NOD/Sh	iLtJ 0.755	0.0269	0.1232	NOD/ShiLtJ	0.1891	0.8451	0.6884
PL/J	0.2448	0.5375	0.033	PL/J	0.3957	0.0799	0.0053	PL/J	0.172	3 0.1238	0.1394	PL/J	0.0322	0.05	0.8907
PWD/PhJ	0.234	0.0449	0.0033	PWD/PhJ	0.3876	0.0326	0.4547	PWD/P	nJ 0.720	4 0.1609	0.3945	PWD/PhJ	0.6836	0.814	0.3801
SJL/J	0.8484	0.9676	0.7912	SJL/J	0.4269	0.1255	0.4821	SJL/J	0.832	8 0.0181	0.0132	SJL/J	0.5111	0.2896	0.1484
SM/J	0.1983	0.4769	0.3812	SM/J	0.0188	0.9604	0.8015	SM/J	0.406	5 0.06	0.9666	SM/J	0.649	0.7288	0.297
SWR/J	0.893	0.0833	0.2755	SWR/J	0.5741	0.7621	0.3194	SWR/J	0.530	9 0.9577	0.4328	SWR/J	0.5232	0.4596	0.3097
WSB/EiJ	0.0158	0.2152	0.0474	WSB/EiJ	0.0048	0.0106	0.3677	WSB/Ei	0.306	3 0.7611	0.3408	WSB/EiJ	0.0885	0.6556	0.4994

NB TS				DB TS				N B Ar			
Strain 🗾	(06-12) 💌	(12-18) 💌	(18-24) 💌	Strain 🗾 💌	(06-12) 💌	(12-18) 💌	(18-24) 💌	Strain 🗾 💌	(06-12) 💌	(12-18) 💌	(1
129S1/SvlmJ	0.9069	0.8639	0.743	129S1/Svlm.	0.8405	0.3632	0.4941	129S1/SvlmJ	0.7127	0.2124	
A/J	0.8298	0.2034	0.0294	A/J	0.113	0.6805	0.472	A/J	0.9736	0.1797	
AKR/J	0.3447	0.1629	0.0748	AKR/J	0.8075	0.8236	0.3644	AKR/J	0.1118	0.4005	
BTBR T+ Itpr	0.3039	0.0191	0.4523	BTBR T+ Itpr	0.6109	0.7854	0.5525	BTBR T+ Itpr	0.2291	0.0156	
BUB/BnJ	0.1441	0.3092	0.2462	BUB/BnJ	0.0063	0.308	0.2946	BUB/BnJ	0.068	0.6613	
C3H/HeJ	0.0382	0.136	0.2383	C3H/HeJ	0.4603	0.2964	0.2231	C3H/HeJ	0.1677	0.2744	
C57BL/6J	0.1825	0.4816	0.5245	C57BL/6J	0.0242	0.1375	0.1006	C57BL/6J	0.7405	0.0666	
C57L/J	0.0027	0.0069	0.058	C57L/J	0.084	0.8558	0.4577	C57L/J	0.0555	0.0853	(
C58/J	0.018	0.2386	0.0018	C58/J	0.0972	0.4755	0.4316	C58/J	0.0027	0.5571	
CAST/EiJ	0.0234	0.2187	0.3453	CAST/EiJ	0.1475	0.1921	0.6736	CAST/EiJ	0.0124	0.811	
CBA/J	0.0031	0.0594	0.236	CBA/J	0.0051	0.0126	0.1833	CBA/J	0.0155	0.0263	
CZECHII/EiJ	0.126	0.8075	0.1308	CZECHII/EiJ	0.0562	0.6473	0.6934	CZECHII/EiJ	0.4476	0.4768	
DBA/2J	0.9883	0.0054	0.7709	DBA/2J	0.2369	0.9594	0.2666	DBA/2J	0.9068	0.0021	
KK/HIJ	0.0613	0.0237	0.8218	KK/HIJ	0.6546	0.5373	0.4912	KK/HIJ	0.0012	0.0039	
MOLF/EiJ	0.3639	0.0241	0.4601	MOLF/EiJ	0.527	0.0539	0.385	MOLF/EiJ	0.3116	0.0713	
NOD/ShiLtJ	0.0413	0.0025	0.0216	NOD/ShiLtJ	0.0323	0.5568	0.0795	NOD/ShiLtJ	0.741	0.0157	
PL/J	0.2871	0.331	0.045	PL/J	0.3351	0.0236	0.0222	PL/J	0.2348	0.3056	
PWD/PhJ	0.2647	0.063	0.9698	PWD/PhJ	0.3667	0.2329	0.0125	PWD/PhJ	0.3213	0.0032	
SJL/J	0.934	0.7165	0.3677	SJL/J	0.5137	0.1058	0.8954	SJL/J	0.6512	0.9885	
SM/J	0.7	0.2815	0.1544	SM/J	0.1253	0.652	0.6874	SM/J	0.3754	0.3857	
SWR/J	0.9013	0.427	0.9698	SWR/J	0.5542	0.6901	0.5514	SWR/J	0.835	0.3099	
WSB/EiJ	0.195	0.284	0.0124	WSB/EiJ	0.0229	0.3097	0.9513	WSB/EiJ	0.4097	0.111	

## 6a. Number of Bouts of NREM





Strain



# 6b. Number of Bouts of REM









## 6c. Number of Bouts of Total Sleep



TS Bout Diff 13-18







N Bout Arousal Diff 13-18







Strain

### References

 National Center for Chronic Disease Prevention and Health Promotion, Division of Population Health. (2017, May 2). CDC - Data and Statistics - Sleep and Sleep Disorders. Retrieved October 22, 2020, from

https://www.cdc.gov/sleep/data\_statistics.html

- 2. Alhola, P., & Polo-Kantola, P. (2007). Sleep deprivation: Impact on cognitive performance. *Neuropsychiatric disease and treatment*, *3*(5), 553–567.
- Goel, N., Rao, H., Durmer, J. S., & Dinges, D. F. (2009). Neurocognitive consequences of sleep deprivation. *Seminars in neurology*, *29*(4), 320–339. <u>https://doi.org/10.1055/s-0029-1237117</u>
- Periasamy, S., Hsu, D. Z., Fu, Y. H., & Liu, M. Y. (2015). Sleep deprivationinduced multi-organ injury: role of oxidative stress and inflammation. *EXCLI journal*, *14*, 672–683. https://doi.org/10.17179/excli2015-245
- Vaccaro, A., Kaplan Dor, Y., Nambara, K., Pollina, E. A., Lin, C., Greenberg, M. E., & Rogulja, D. (2020). Sleep Loss Can Cause Death through Accumulation of Reactive Oxygen Species in the Gut. *Cell*, *181*(6), 1307–1328.e15. https://doi.org/10.1016/j.cell.2020.04.049
- Andersen M. L. (2020). A brief report on early sleep studies. Sleep science (Sao Paulo, Brazil), 13(1), 1–2. <u>https://doi.org/10.5935/1984-0063.20190144</u>
- Borbély, A. A., Daan, S., Wirz-Justice, A., & Deboer, T. (2016). The two-process model of sleep regulation: a reappraisal. *Journal of sleep research*, *25*(2), 131– 143. <u>https://doi.org/10.1111/jsr.12371</u>

- Basner, M., & Dinges, D. F. (2011). Maximizing sensitivity of the psychomotor vigilance test (PVT) to sleep loss. *Sleep*, *34*(5), 581–591. https://doi.org/10.1093/sleep/34.5.581
- Patel, A. K., Reddy, V., & Araujo, J. F. (2022). Physiology, Sleep Stages.
   In StatPearls. StatPearls Publishing.
- Eugene, A. R., & Masiak, J. (2015). The Neuroprotective Aspects of Sleep. *MEDtube science*, 3(1), 35–40.
- Peever, J., & Fuller, P. M. (2016). Neuroscience: A Distributed Neural Network Controls REM Sleep. *Current biology: CB*, 26(1), R34–R35.

https://doi.org/10.1016/j.cub.2015.11.011

- 12. Tamaki, M., Wang, Z., Barnes-Diana, T., Guo, D., Berard, A. V., Walsh, E., Watanabe, T., & Sasaki, Y. (2020). Complementary contributions of non-REM and REM sleep to visual learning. *Nature neuroscience*, 23(9), 1150–1156. <u>https://doi.org/10.1038/s41593-020-0666-y</u>
- 13. Campbell I. G. (2009). EEG recording and analysis for sleep research. *Current protocols in neuroscience, Chapter 10*, Unit10.2.

https://doi.org/10.1002/0471142301.ns1002s49

- 14. Oishi, Y., Takata, Y., Taguchi, Y., Kohtoh, S., Urade, Y., & Lazarus, M. (2016).
  Polygraphic Recording Procedure for Measuring Sleep in Mice. *Journal of visualized experiments: JoVE*, (107), e53678. <u>https://doi.org/10.3791/53678</u>
- Blagrove, M., Fouquet, N. C., Henley-Einion, J. A., Pace-Schott, E. F., Davies, A.
   C., Neuschaffer, J. L., & Turnbull, O. H. (2011). Assessing the dream-lag effect

for REM and NREM stage 2 dreams. PloS one, 6(10), e26708.

https://doi.org/10.1371/journal.pone.0026708

16. Barger, Z., Frye, C. G., Liu, D., Dan, Y., & Bouchard, K. E. (2019). Robust, automated sleep scoring by a compact neural network with distributional shift correction. *PloS one*, *14*(12), e0224642.

https://doi.org/10.1371/journal.pone.0224642

- 17. Gao, V., Turek, F., & Vitaterna, M. (2016). Multiple classifier systems for automatic sleep scoring in mice. *Journal of neuroscience methods*, 264, 33–39. <u>https://doi.org/10.1016/j.jneumeth.2016.02.016</u>
- Rytkönen, K. M., Zitting, J., & Porkka-Heiskanen, T. (2011). Automated sleep scoring in rats and mice using the naive Bayes classifier. *Journal of neuroscience methods*, 202(1), 60–64. <u>https://doi.org/10.1016/j.jneumeth.2011.08.023</u>
- Toth, L. A., & Bhargava, P. (2013). Animal models of sleep disorders.
   Comparative medicine, 63(2), 91–104.
- 20. Sehgal, A., & Mignot, E. (2011). Genetics of sleep and sleep disorders. *Cell*, *146*(2), 194–207. https://doi.org/10.1016/j.cell.2011.07.004
- 21. Miyagawa, T., & Tokunaga, K. (2019). Genetics of narcolepsy. Human genome variation, 6, 4. <u>https://doi.org/10.1038/s41439-018-0033-7</u>
- Mignot E. (1998). Genetic and familial aspects of narcolepsy. *Neurology*, *50*(2
   Suppl 1), S16–S22. <u>https://doi.org/10.1212/wnl.50.2\_suppl\_1.s16</u>
- 23. Kornum BR, Kawashima M, Faraco J, Lin L, Rico TJ, Hesselson S, Axtell RC, Kuipers H, Weiner K, Hamacher A, et al. Common variants in P2RY11 are associated with narcolepsy. *Nat Genet.* 2011; 43:66–71.

- 24. Summa, K. C., & Turek, F. W. (2011). The Genetics of Sleep: Insight from Rodent Models. *Sleep medicine clinics*, *6*(2), 141–154.
  https://doi.org/10.1016/j.jsmc.2011.04.004
- 25. Hoekstra, M. M., Emmenegger, Y., Hubbard, J., & Franken, P. (2019). Coldinducible RNA-binding protein (CIRBP) adjusts clock-gene expression and REMsleep recovery following sleep deprivation. *eLife*, *8*, e43400.

https://doi.org/10.7554/eLife.43400

26. Niwa, Y., Kanda, G. N., Yamada, R. G., Shi, S., Sunagawa, G. A., Ukai-Tadenuma, M., Fujishima, H., Matsumoto, N., Masumoto, K. H., Nagano, M., Kasukawa, T., Galloway, J., Perrin, D., Shigeyoshi, Y., Ukai, H., Kiyonari, H., Sumiyama, K., & Ueda, H. R. (2018). Muscarinic Acetylcholine Receptors Chrm1 and Chrm3 Are Essential for REM Sleep. *Cell reports*, *24*(9), 2231–2247.e7. https://doi.org/10.1016/j.celrep.2018.07.082

 Diessler, S., Jan, M., Emmenegger, Y., Guex, N., Middleton, B., Skene, D. J., Ibberson, M., Burdet, F., Götz, L., Pagni, M., Sankar, M., Liechti, R., Hor, C. N., Xenarios, I., & Franken, P. (2018). A systems genetics resource and analysis of sleep regulation in the mouse. *PLoS biology*, *16*(8), e2005750. <u>https://doi.org/10.1371/journal.pbio.2005750</u>

28. Mackiewicz, M., Paigen, B., Naidoo, N., & Pack, A. I. (2008). Analysis of the QTL for sleep homeostasis in mice: Homer1a is a likely candidate. *Physiological genomics*, 33(1), 91–99. <u>https://doi.org/10.1152/physiolgenomics.00189.2007</u>

29. Tafti M. (2007). Quantitative genetics of sleep in inbred mice. *Dialogues in clinical neuroscience*, *9*(3), 273–278.

https://doi.org/10.31887/DCNS.2007.9.3/mtafti

- 30. Ashbrook, L. H., Krystal, A. D., Fu, Y. H., & Ptáček, L. J. (2020). Genetics of the human circadian clock and sleep homeostat. *Neuropsychopharmacology: official publication of the American College of Neuropsychopharmacology*, 45(1), 45–54. <u>https://doi.org/10.1038/s41386-019-0476-7</u>
- 31. Mang, G. M., & Franken, P. (2015). Genetic dissection of sleep homeostasis. *Current topics in behavioral neurosciences*, 25, 25–63. <u>https://doi.org/10.1007/7854\_2013\_270</u>
- 32. Jan, M., O'Hara, B. F., & Franken, P. (2020). Recent advances in understanding the genetics of sleep. *F1000Research*, *9*, F1000 Faculty Rev-214. https://doi.org/10.12688/f1000research.22028.1
- 33. Franken, P., Chollet, D., & Tafti, M. (2001). The homeostatic regulation of sleep need is under genetic control. *The Journal of neuroscience: the official journal of the Society for Neuroscience*, *21*(8), 2610–2621. <u>https://doi.org/10.1523/JNEUROSCI.21-08-02610.2001</u>
- Maret, S., Dorsaz, S., Gurcel, L., Pradervand, S., Petit, B., Pfister, C., Hagenbuchle, O., O'Hara, B. F., Franken, P., & Tafti, M. (2007). Homer1a is a core brain molecular correlate of sleep loss. Proceedings of the National Academy of Sciences of the United States of America, 104(50), 20090–20095. https://doi.org/10.1073/pnas.0710131104