UCSF

UC San Francisco Previously Published Works

Title

The BIN1 rs744373 SNP is associated with increased tau-PET levels and impaired memory

Permalink <https://escholarship.org/uc/item/4bx0r1tm>

Journal Nature Communications, 10(1)

ISSN

2041-1723

Authors

Franzmeier, Nicolai Rubinski, Anna Neitzel, Julia [et al.](https://escholarship.org/uc/item/4bx0r1tm#author)

Publication Date

2019

DOI

10.1038/s41467-019-09564-5

Peer reviewed

ARTICLE

https://doi.org/10.1038/s41467-019-09564-5 **OPEN**

The BIN1 rs744373 SNP is associated with increased tau-PET levels and impaired memory

Nicolai Franzmeier¹, Anna Rubinski¹, Julia Neitzel¹, Michael Ewers¹ & the Alzheimer's Disease Neuroimaging Initiative (ADNI)

The single nucleotide polymorphism (SNP) rs744373 in the bridging integrator-1 gene (BIN1) is a risk factor for Alzheimer's disease (AD). In the brain, BIN1 is involved in endocytosis and sustaining cytoskeleton integrity. Post-mortem and in vitro studies suggest that BIN1 associated AD risk is mediated by increased tau pathology but whether rs744373 is associated with increased tau pathology in vivo is unknown. Here we find in 89 older individuals without dementia, that BIN1 rs744373 risk-allele carriers show higher AV1451 tau-PET across brain regions corresponding to Braak stages II–VI. In contrast, the BIN1 rs744373 SNP was not associated with AV45 amyloid-PET uptake. Furthermore, the rs744373 risk-allele was associated with worse memory performance, mediated by increased global tau levels. Together, our findings suggest that the BIN1 rs744373 SNP is associated with increased tau but not beta-amyloid pathology, suggesting that alterations in BIN1 may contribute to memory deficits via increased tau pathology.

¹ Institute for Stroke and Dementia Research, Klinikum der Universität München, Ludwig-Maximilians-Universität LMU, Feodor-Lynen Straße 17, 81377 Munich, Germany. A full list of consortium members appears at the end of the paper. Correspondence and requests for materials should be addressed to M.E. (email: [michael.ewers@med.uni-muenchen.de\)](mailto:michael.ewers@med.uni-muenchen.de)

[A](#page-9-0)lzheimer's disease (AD) is the most common cause of late
onset dementia and is characterized by the development
of pathological amyloid plaques and tau tangles in the
brain¹ While late onset AD is an age-related disea onset dementia and is characterized by the development brain¹. While late onset AD is an age-related disease, results from twin and family studies have emphasized that ~50% of phenotypic variance in AD can be explained by genetic variations[2.](#page-9-0) Recent genome-wide association studies (GWAS) have identified several loci that are associated with increased risk of AD, among which the single nucleotide polymorphisms (SNPs) in the bridging integrator 1 (BIN1) gene show the second highest oddsratios for sporadic AD, superseded only by apolipoprotein E $(APOE)$ variants^{3-[7](#page-9-0)}. Specifically, the most frequently reported BIN1 AD risk variant is the SNP rs744373 which shows a global allele frequency of 37% and is associated with an increase in AD risk by an odds-ratio of $1.17-1.19^{5,7-10}$ $1.17-1.19^{5,7-10}$ $1.17-1.19^{5,7-10}$. Thus, understanding the mechanisms by which BIN1 and in particular the rs744373 SNP contributes to AD risk will lead to a better understanding of the pathomechanisms of AD and help uncover novel therapeutic targets.

The BIN1 gene encodes the nucleocytoplasmic adaptor protein BIN1 also known as amphiphysin-2, a membrane deforming protein that is most highly expressed in muscle and brain tissue in an isoform dependent way^{11,12}. In the brain, BIN1 subserves multiple functions such as regulating endocytosis, cytoskeleton integrity, and apoptosis 13 . A key hypothesis for the role of BIN1 in AD is the aggravation of tau pathology, i.e. a key primary brain pathology associated with cognitive impairment in $AD^{14,15}$ $AD^{14,15}$ $AD^{14,15}$ $AD^{14,15}$ $AD^{14,15}$. In post-mortem studies in AD, higher brain BIN1 expression was found to be associated with the presence of neurofibrillary tau tangles^{[16](#page-9-0)}. Importantly, previous studies have described higher BIN1 mRNA expression in brain tissue of BIN1 risk SNP carriers with and without $AD^{15,17}$. Further, AD patients carrying a $BIN1$ risk SNP showed higher post-mortem tau pathology but not Aβ when compared to non-carrier AD patients¹⁵. Together these findings suggest that BIN1 SNP-associated alterations in BIN1 expression contribute to the development of tau pathology. Although the mechanisms that link BIN1 to tau pathology are only poorly understood, recent findings suggest that alterations in the protein level of a neuron-specific BIN1 isoform that binds exclusively to clathrin^{[18](#page-9-0)}, enhance transmission of tau between neurons via enhanced endocytosis of tau^{19} tau^{19} tau^{19} . Thus, alterations in BIN1 may promote prion-like spreading of tau within the brain. Alternatively, alterations in BIN1 may be associated with betaamyloid $(A\beta_{1-42})^{20}$. Post-mortem analyses in AD have shown that BIN1 accumulates in the vicinity of amyloid plaques^{[21](#page-9-0)}. In primary neuronal cultures, BIN1 was found to regulate the intraneuronal cleavage of the amyloid precursor protein (APP) by β-secretase[22](#page-9-0). However, the BIN1-mediated increase in the intraneuronal pool of $A\beta_{1-42}$ only weakly translated into higher extraneuronal levels of $A\beta_{1-42}$, i.e., a core feature of AD, and several histochemical brain autopsy and cell culture studies suggested that alterations in BIN1 expression were associated with stronger tau pathology rather than $A\beta^{15,16}$ $A\beta^{15,16}$ $A\beta^{15,16}$ $A\beta^{15,16}$ $A\beta^{15,16}$. Together, these findings from preclinical studies suggest that the risk conferred by BIN1 genetic variants may be exerted via promoting tau pathology rather than Aβ in the brain.

Up to now, the translation of these findings to patients with AD using in vivo biomarkers of tau pathology has been difficult. In mild cognitive impairment (MCI) and AD dementia, the BIN1 rs744373 SNP was reported to be associated with higher cerebrospinal fluid (CSF) levels of total and phospho-tau but not with $A\beta^{23}$ $A\beta^{23}$ $A\beta^{23}$, whereas others could not detect an association between the BIN1 rs744373 SNP and CSF tau or phospho-tau levels²⁴. However, CSF-phospho-tau levels are only moderately associated with neurofibrillary tangles in the brain as assessed post-mortem^{[25](#page-9-0)} or by tau-PET imaging^{26,27}. Furthermore, CSF tau

levels may reflect differences in tau production rather than the amount of pathological tau deposits in the brain^{[28](#page-9-0)}. A previous MRI neuroimaging study showed that a BIN1 SNP was associated with decreased cortical thickness in the entorhinal cortex and temporal pole, i.e., sites of early tau pathology²⁹. However, that study did not assess tau pathology itself. Thus, the question remains, whether the BIN1 rs744373 SNP is associated with increased tau pathology in subjects with AD.

The introduction of AV1451 tau-PET imaging allows to assess fibrillary tangles in the living brain^{[30](#page-10-0)}. Here we employ $AV1451$ PET imaging in elderly subjects in order to translate previous preclinical and post-mortem findings on the association BIN1 and primary AD pathology. We assess whether carriers of the BIN1 rs744373 SNP show elevated regional levels of AV1451 tau-PET in those brain regions that are known to show increased susceptibility to tau pathology as defined by the post-mortem established Braak staging³¹. We test the associations of the $BIN1$ rs744373 SNP in non-demented subjects to understand whether the BIN1 SNP is associated with tau in the early stages of the development of tau pathology. In addition, given previous evidence of a potential involvement of BIN1 rs744373 in Aβ pathology^{[21](#page-9-0),[23](#page-9-0)}, we assess whether the BIN1 rs744373 SNP is associated with higher regional Aβ deposition as assessed by AV45-PET in the same subjects. We hypothesize that carriage of the BIN1 rs744373 risk-allele selectively enhances tau pathology. Since BIN1 genetic variants were previously associated with faster cognitive decline, we lastly test whether alterations in AV1451 tau-PET levels mediate the association between the BIN1 rs744373 SNP and worse memory performance.

Results

Sample characteristics. For the current study, we analyzed data from 89 participants of the ADNI cohort, including 49 cognitively normal (CN) and 40 mild cognitively impaired (MCI) subjects (see Table 1 for sample characteristics). All subjects underwent AV1451 tau-PET, AV45-amyloid PET, structural MRI and cognitive testing at the same study visit of ADNI phase 3. The genotype of the BIN1 rs744373 SNP was extracted from ADNI GWAS data provided by the ADNI genetics core, where we found 22 CN subjects and 18 MCI subjects to carry at least one copy of the BIN1 rs744373 G-allele which confers higher risk of AD dementia as shown by GWAS⁴. Henceforth, these subjects will be referred to as BIN1 rs744373 risk-allele carriers. BIN1 rs744373 allele distribution (GG/GA/AA = 8/32/49) did not deviate from Hardy–Weinberg equilibrium ($p = 0.422$, Chisquared test). There were no differences in baseline demographics (age, gender, education) between CN vs. MCI or between BIN1 rs744373 risk-allele vs. normal-allele carriers. In total, 48 subjects (24 CN &, 24 MCI) showed abnormally elevated Aβ-levels as determined via AV45 PET (i.e., global standardized uptake value ratio (SUVR) > 1.11).

BIN1 rs744373 is associated with higher tau-PET uptake. In a first step, we tested the hypothesis that BIN1 rs744373 risk-allele carriers show higher tau pathology (i.e., global or for regions corresponding to Braak stages I–VI, Fig. [1a](#page-3-0)) than carriers of the normal BIN1 rs744373 allele. For global tau, we quantified the overall tau load as global AV1451 tau-PET uptake using an established Freesurfer-based protocol (i.e., standardized volume uptake ratio normalized to the inferior cerebellar $gray)^{32}$ $gray)^{32}$ $gray)^{32}$. When testing via ANCOVA whether the BIN1 rs744373 SNP had an effect on global AV1451 tau-PET uptake, we found risk-allele carriers to show elevated global tau levels with a Cohens d of 0.562, controlling for age, gender, education, ApoE ε4 status, diagnosis, and gray matter (GM) volume of the global tau ROI

Fig. 1 Staging systems for tau- and amyloid-PET. Spatial mapping of Braak- (a) and amyloid stage-specific ROIs (b) that were used to determine regional AV1451 tau- and AV45 amyloid-PET uptake within the sample of $n = 89$ subjects

 $(F(81,7) = 7.694, p = 0.007,$ see Fig. [2](#page-4-0)a). To test whether the effects of BIN1 rs744373 on AV1451 tau-PET uptake were independent of amyloid pathology, we further included overall amyloid load as a covariate, which was quantified as the global AV45 amyloid-PET uptake. Importantly, inclusion of global AV45 amyloid-PET uptake as a covariate did not alter the association between BIN1 rs744373 risk-allele carriage and elevated global AV1451 tau-PET uptake $(F(80,8) = 7.658, p = 0.007,$ ANCOVA, see model 2 in Table [2](#page-5-0) for statistics). Results remained unchanged when alternatively including Aβ-status as a covariate. This suggests that the associations between BIN1 rs744373 and increased global AV1451 tau-PET uptake are independent of diagnostic group or Aβ-status. Results for the main analysis (i.e., SNP effect on global AV1451 tau-PET) were consistent when repeated for the previously reported BIN1 rs7561528 risk var i ant¹⁴, which was also available from ADNI GWAS data (48 riskallele carriers vs. 41 normal-allele carriers, $F(80,8) = 3.217$, $p =$ 0.045 , ANCOVA $)$ ^{[14](#page-9-0)}.

We next tested, whether the effects of BIN1 rs744373 on tau pathology showed regional differences. To this end, we assessed the AV1451 tau-PET SUVR within brain regions corresponding to Braak stages I–VI (Fig. 1a) that recapitulate the spatial tauspreading pattern from early-to-late-stage tau pathology across the cortex³³. Here, we could consistently detect significantly (p < 0.05, ANCOVA) elevated tau load in BIN1 rs744373 risk-allele carriers across regions corresponding to Braak stages 2–6, with effect sizes ranging between 0.430 and 0.594. This suggests that the BIN1 rs744373 risk allele is associated with general brain-wide increases in tau. These analyses are summarized in Fig. [2a](#page-4-0) and Table [2](#page-5-0). Despite the highly consistent effects of the BIN1 rs744373 SNP on tau, only the associations for global and Braak stage 5 AV1451 tau-PET SUVR remained significant after applying a Bonferroni-corrected α -threshold of 0.0071 (i.e., $\alpha =$ 0.05 adjusted for 7 tests). Again, these results remained fully consistent when additionally controlling for amyloid levels (i.e., global AV45 amyloid-PET SUVR) as summarized in Table [2](#page-5-0) (shown as model 2). Results remained also unchanged when controlling for Aβ status. These findings suggest that the effects of BIN1 rs744373 on AV1451 tau-PET uptake are independent of Aβ or diagnosis.

When testing, whether the BIN1 rs744373 SNP had an effect on amyloid load, we did not detect any differences between risk-allele and normal-allele carriers for global AV45 amyloid-PET SUVR, controlling for age, gender, education, ApoE ε4 status, diagnosis, and GM volume of the respective amyloid ROI (F(81,7) = 0.148, $p = 0.701$, ANCOVA). To address potential effects of BIN1 rs744373 on regional amyloid levels, we quantified amyloid SUVRs within four distinct early- to late-amyloid stage ROIs (Fig. 1b) that have been previously shown to recapitulate amyloid spread^{[34](#page-10-0)}. In line with the results for global $AV45$ amyloid-PET uptake, we found no significant effect of the BIN1 rs744373 SNP on AV45 amyloid-PET uptake within the different amyloid stage ROIs (all $p > 0.05$, ANCOVAs, see Fig. [2b](#page-4-0) & Table [3](#page-5-0) for statistics), when controlling for age, gender, education, ApoE ε4 status, diagnosis, and GM volume of the, respective, amyloid-stage ROIs. All results remained consistent when additionally including global AV1451 tau-PET SUVRs as a covariate (see Table [3](#page-5-0)). Again, there was no interaction between diagnosis and BIN1 rs744373 or between global AV45 tau-PET uptake and BIN1 rs744373 on AV45 amyloid-PET uptake.

BIN1 rs744373 effects on tau are independent of amyloid. Next, we tested whether BIN1 rs744373 risk-allele carriage was associated with higher tau independent of the level of amyloid. Using linear regression controlling for age, gender diagnosis and ApoE ε4 status, we found that both global AV45 amyloid-PET SUVR $(t(81) = 2.506, \ \beta = 0.248, \ \ p = 0.014) \text{ and } BHNI \text{ rs}744373$ (t(81) = 2.882, β = 0.280, p = 0.005) had independent main effects on global AV1451 tau-PET SUVR. No interaction between Aβ status (binary) or global AV45 amyloid-PET SUVR (continuous) and BIN1 rs744373 on AV1451 tau-PET was found. To illustrate the main effect of BIN1 rs744373 effects on AV1451 tau-PET that was independent of AV45 amyloid-PET, we have plotted the association between BIN1 rs744373 and global AV1451 tau-PET uptake at each quartile of global AV45 amyloid-PET (Fig. [3\)](#page-6-0).

Fig. 2 Effects of BIN1 rs744373 on tau- and amyloid-PET uptake. a Boxplots showing the differences in global or regional AV1451 tau-PET SUVRs between BIN1 rs744373 risk allele ($n = 40$, yellow) vs. normal-allele carriers ($n = 49$, blue). P-values are based on ANCOVA models controlled for age, gender, education, diagnosis, memory performance (i.e., ADNI-MEM) and gray matter volume of the respective ROI. **b** Differences in global or regional AV45 amyloid-PET uptake between BIN1 rs744373 risk allele ($n = 40$, yellow) vs. normal-allele carriers ($n = 49$, blue). Boxplots are displayed as median (center line) ±interquartile range (box boundaries) with whiskers including observations falling within the 1.5 interquartile range. P-values are again derived from ANCOVA models controlled for age, gender, education, diagnosis, ApoE ε4 carrier status, and gray matter volume of the respective ROI. $p < 0.05$ (uncorrected); $p' =$ significant after Bonferroni correction (p < 0.0071)

Spatial match of BIN1 mRNA expression and tau pathology. Our finding of the association between BIN1 rs744373 and higher AV1451 tau-PET levels, plus previous reports of altered BIN1 mRNA levels in AD¹⁵ suggest that alterations in cerebral BIN1 expression are related to tau pathology. In order to further substantiate this hypothesis, we argued that regional expression patterns of BIN1 in the brain should overlap with those brain regions of increased vulnerability to tau pathology, given the previously reported association between BIN1 mRNA expression and neurofibrillary tangles in post-mortem brains^{15,16}. In order to

test this hypothesis, we obtained whole-brain mRNA expression levels of BIN1 from the Allen Brain Atlas, which is based on postmortem brain-wide microarray assessments of six healthy brain donors without any history of psychiatric or neurological disorders^{35,36}. Specifically, we used the median of microarraybased $log₂$ mRNA expression of *BIN1* across the six donors that have been recently summarized for each region of the Freesurferbased Desikan–Killiany atlas 3^7 . For the same atlas regions, we determined the group-median AV1451 tau-PET SUVR across all subjects in the current study. We restricted the analysis to the left hemisphere since full mRNA expression data of all six brain donors is only available for this hemisphere. Surface renderings of both group average left hemispheric tau and BIN1 mRNA expression are depicted in the upper panel of Fig. [4](#page-6-0). We used spatial regression on 10,000 bootstrapped samples based on which group-median tau levels were iteratively determined to assess the association between group-median AV1451 tau-PET SUVR and BIN1 mRNA expression patterns. Here, we found a significant average positive Pearson-Moment correlation ($r =$ 0.374, see Fig. [5\)](#page-7-0) with a 95% CI of $[0.370:0.379]$ ($p < 0.001$). These results suggest that regions with higher BIN1 mRNA expression have a higher likelihood of showing elevated tau levels. In order to assess the spatial overlap of regions with high BIN1 expression or high AV1451 tau-PET uptake, we thresholded both the groupmedian tau PET and the BIN1 mRNA expression maps at a percentile threshold of 75%, (Fig. [4](#page-6-0) lower panel). Visual inspection suggests that mesio- and inferior temporal brain regions corresponding to early Braak stages show both high BIN1 mRNA expression as well as high AV1451 tau-PET uptake.

Tau mediates BIN1 rs744373 effects on memory impairment. To assess whether the BIN1 rs744373 SNP is detrimental for cognition via increasing tau pathology, we tested whether riskallele carriage is associated with worse memory and whether these effects are mediated via increased tau pathology. To this end, we applied causal mediation analysis with 10 000 bootstrapping iterations controlling for age, gender, education, diagnosis, global AV45 amyloid-PET SUVR, and ApoE ε4 status. Memory performance was assessed based on ADNI-MEM, which is an established composite score developed by the ADNI core that summarizes the performance on multiple memory tests^{[38](#page-10-0)}. Supporting our hypothesis, we found that the BIN1 rs744373 riskallele was significantly associated with worse ADNI-MEM scores $(\beta = -0.25, p = 0.030)$, where this association was mediated via global AV1451 tau-PET uptake (bootstrapped average causal mediation effect: $β = −0.083$ [−0.180;0.00], $p = 0.016$). The effect was considered a full mediation, since the direct effect of the BIN1 rs744373 risk allele on ADNI-MEM was no longer significant $(\beta = -0.17, p = 0.15)$ in the presence of the mediator (i.e., global AV1451 tau-PET uptake). A path model of this mediation analysis is shown in Fig. [6](#page-7-0).

Discussion

The major finding of the current study was that the BIN1 rs744373 risk allele was associated elevated AV1451 tau-PET uptake. In contrast, we detected no association between BIN1 rs744373 and regional Aβ assessed by AV45-PET, suggesting that BIN1 is associated with higher tau pathology rather than Aβ. In addition, we found BIN1 rs744373 to be associated with worse memory performance, where this effect was mediated by BIN1 rs744373-associated elevation of global AV1451 tau-PET. Together, our findings support the hypothesis that the BIN1 rs744373 risk-allele is associated with elevated cerebral tau pathology, thereby worsening memory decline. Our findings represent an important contribution to the understanding of the role of BIN1

CN Cognitively Normal, MCI Mild Cognitive Impairment, M male, f female, MMSE Mini-Mental State Exam, ADNI-MEM Alzheimer's Disease Neuroimaging Initiative-Memory composite a sig. (p < 0.05) different from CN-BIN1 normal

b_{sig}. different from CN-BIN1 risk

c sig. different from MCI-BIN1 normal sig. different from MCI-BIN1 risk

Model 1 Covariates: age, gender, education, diagnosis, ROI gray matter, ApoE e4. Model 2 Covariates: Global AV45 amyloid-PET, age, gender, education, diagnosis, ROI gray matter, ApoE e4
'Significant at p < 0.05 (uncorrecte

^aSignificant after Bonferroni correction for 7 tests ($p < 0.0071$)

in AD, as we demonstrate in living non-demented elderly subjects an association of BIN1 rs744373 and regional elevation of tau pathology, i.e., a key AD pathology associated with cognitive impairment.

For our first finding, the association between BIN1 rs744373 and higher AV1451 tau-PET ROI values but not AV45 amyloid-PET, suggests a selective association between the BIN1 rs744373 SNP and PET-assessed tau pathology. Of note, we did not find an interaction between BIN1 rs744373 and AV45 amyloid-PET on AV1451 tau-PET levels, suggesting that the association between BIN1 and tau does not depend on the presence of Aβ pathology. Supporting this notion, effects of BIN1 rs744373 on AV1451 tau-PET levels remained consistent when controlled for Aβ-status or continuous AV45 amyloid-PET levels. These findings are in general agreement with several previous findings suggesting that BIN1 is linked to tau pathology rather than amyloid pathology. Here, it has been previously reported that brain BIN1 protein levels are correlated

with neurofibrillary tangle pathology but not with diffuse or neuritic amyloid plaques in AD brains¹⁶. In a similar vein, *BIN1* risk variants have been previously shown to correlate with post-mortem assessed brain levels of AT8 positive tau pathology but not with $A\beta$ ¹⁵. In contrast, one previous study showed that BIN1 becomes insoluble and accumulates in the vicinity of amyloid plaques in a mouse model of AD and in brain sections from AD patients³⁹. Still, it is unclear whether BIN1 alterations are a cause or a consequence of amyloid pathology. Our current results suggest that the BIN1 rs744373 risk allele is associated primarily with PET-assessed tau rather than amyloid levels.

The current results are also in agreement with previous reports of BIN1 rs744373 risk-allele carriage being associated with increased CSF-phospho-tau and CSF-total tau levels but not with CSF-A β levels and amyloid PET^{[23](#page-9-0)}. We caution however that our current finding of absence of an association between BIN1 rs744373 and AV45 amyloid-PET may partially be due to the

Fig. 3 BIN1 rs744373 effects on tau across the amyloid spectrum. Boxplot showing the association between BIN1 rs744373 (risk-allele $n = 40$, yellow; normal-allele $n = 49$, blue) and global AV1451 tau-PET uptake across amyloid quartiles. The statistical main effects presented in the upper left corner of the graph are derived from linear regression, with the BIN1 rs7443733 SNP and global AV45 amyloid-PET SUVR as predictors, controlling for age, gender, diagnosis, and ApoE ε4 status. Cohens d effects sizes displayed in the plot were derived for each quartile. Note, that no interaction between BIN1 rs744373 and global AV45 amyloid-PET uptake was found, suggesting that the effects of BIN1 rs744373 on tau are similar between high- and low-amyloid groups. Boxplots are displayed as median (center line) ±interquartile range (box boundaries) with whiskers including observations falling within the 1.5 interquartile range

fact that in many subjects amyloid may have reached already plateau levels 40 , leading to reduced variability in AV45 amyloid-PET levels. Thus, it is still possible that BIN1 is associated with Aβ pathology especially at the early presymptomatic phase before a plateau is reached.

The molecular mechanisms underlying the association between BIN1 genetic variants and increased AD risk are not known. For the link between BIN1 and tau pathology, in vitro studies showed that BIN1 binds to tau via a proline rich SH3 domain^{15,[41](#page-10-0)–43}, possibly reducing the integrity of the cytoskeleton $41,42$. Our findings of a spatial match between the BIN1 mRNA expression pattern in the brain and regions of increased AV1451 tau-PET uptake support the notion that BIN1 SNP-related alterations of BIN1 expression are associated with tau pathology. It is unclear, however, whether the binding of BIN1 to tau entails the development of pathological fibrillary tau¹⁵. Alternatively, BIN1 may enhance spreading of pathological tau across connected neurons⁴⁴ via endocytosis, i.e., a pathway that has recently been suggested to lead to prion-like spreading of tau in the brain^{45,46}. The neuron-specific BIN1 isoform 1 interacts with clathrin, thereby attenuating the post-synaptic endocytotic uptake of tau^{[19](#page-9-0),42}. Recent histological brain-autopsy studies showed that while BIN1 protein expression is overall increased in AD, the BIN1 isoform 1 is decreased in AD[16.](#page-9-0) Together, these results

Fig. 4 Spatial patterns of tau-PET and BIN1 mRNA expression. Spatial mapping of median BIN1 mRNA expression (i.e., log₂) derived from the Allen Brain Atlas and group-median AV1451 tau-PET SUVR (derived from n = 89 subjects), either for all ROIs (upper panel) or restricted to regions falling above the 75th percentile of either BIN1 mRNA expression or groupmedian AV1451 tau-PET uptake (lower panel). Color scales represent SUVR scores for AV1451 tau-PET and log₂ mRNA expression for BIN1 mRNA

suggest that reduced BIN1 isoform 1 levels may enhance the endocytosis-mediated spreading of tau pathology in AD. However, it is unknown whether the BIN1 genetic variants are associated with reduced BIN1 isoform 1 expression and future studies need to clarify the exact pathomechanisms of BIN1 alterations.

Our second major finding was that the association between BIN1 rs744373 and memory impairment was mediated via elevated global tau levels. This suggests that BIN1 rs744373 contributes to the development of tau pathology in at-risk subjects, resulting in stronger cognitive impairment. These results are in agreement with previous findings of BIN1 rs744373 being associated with faster decline in global cognition 47 and episodic memory⁴⁸. Our results are also consistent with previous studies showing a close association between tau PET and cognitive decline^{33,49}. Our mediation analysis suggests that the $BIN1$ rs744373 SNP is linked to memory impairment due to the increase in tau pathology in the BIN1 rs744373 risk-allele carriers. We caution that the study design is correlational in nature and thus a causative interpretation should not be drawn. However, our findings provide a putative pathomechanistic link between the BIN1 rs744373 SNP and the increase in dementia risk as reported by $GWAS³$.

We point out several caveats that should be considered when interpreting the results of the current study. Firstly, there are several BIN1 genetic variants associated with an increased risk of AD^{9,14,15}.

Fig. 5 Association between tau-PET and BIN1 mRNA expression. Scatterplot showing the positive association between ROI-based BIN1 mRNA expression obtained from the Allen Brain Atlas and group-median AV1451 tau-PET uptake (Number of ROIs = 34; Association illustrated as leastsquares regression line ±95% confidence interval). P-values and 95% confidence intervals of r-values are derived from bootstrapped spatial regression with 1000 iterations

The different SNPs on the BIN1 gene may be in disequilibrium and may add independently from each other to the risk of $AD¹⁴$. Here, we focused on rs744373 because this SNP is most frequently reported to be associated with AD across different GWAS studies^{3,7} (see also AlzGene database at <http://www.alzgene.org/>[\)50](#page-10-0). Although rs744373 is the primary BIN1 SNP associated with increased AD risk, a previous study suggested that the Indel rs59335482 is associated with increased BIN1 mRNA in post-mortem analyzed brains from AD patients, suggesting that rs59335482 is the functionally effective BIN1 genetic variant associated with AD risk. However, the Indel rs59335482, which was not available in the current GWAS, is in almost complete linkage equilibrium with $rs744373^{15}$, suggesting both SNPs share redundant predictive value. Furthermore, control analysis using an alternative SNP rs7561528 that was available from the GWAS analysis, i.e., another frequently reported BIN1 SNP as a risk factor of $AD¹⁴$, confirmed the association between the BIN1 SNP and increased regional AV1451 tau-PET, suggesting that the current findings were not specific to rs744373 as a tau-related genetic variant of BIN1.

Secondly, we caution that even though the current findings suggests that BIN1 rs744373 is associated with tau pathology, other pathomechanisms of BIN1 may contribute to the increased risk of $AD⁵¹$ $AD⁵¹$ $AD⁵¹$. The BIN1 protein is highly expressed in the white matter (WM) and oligodendrocytes^{[52](#page-10-0)} and BIN1 is a key driver of a oligodendrocyte-associated genetic co-expression network that is dysregulated in AD^{53} . These results suggest that BIN1 is associated with altered oligodendrocyte integrity that may contribute to white-matter alterations that are a core part of AD-related pathological changes^{[54](#page-10-0),55}. Also, BIN1 has been shown to interact with physiological tau in vitro, i.e., a key constituent of microtubules^{[15](#page-9-0)}. It is thus possible, that alterations of BIN1 in the white matter increase the development of pathological tau, which manifests distantly as neurofibrillary tau tangles in the soma. The current results are not in conflict with those alternative pathomechanisms, as the ubiquitously expressed BIN1 protein subserves multiple diverse functions in the brain^{12,14}, and may thus be involved in multiple AD-related pathological pathways.

Thirdly, the AV1451 tau-PET tracer has previously shown offtarget binding in the meninges, basal ganglia, and choroid plexus, which may confound the assessment of tau pathology in cortical and subcortical brain regions⁵⁶. To address this, we excluded ROIs covering the basal ganglia in our analysis to avoid known off-target

Fig. 6 Tau mediates effects of BIN1 rs744373 on memory impairment. Path diagram of the mediation model (assessed on the full sample of $n=$ 89 subjects), showing that associations between BIN1 rs744373 risk-allele carriage and worse memory are mediated via global AV1451 tau-PET uptake. Path-weights are displayed as beta values with standard errors in brackets. All paths are controlled for age, gender, education, diagnosis, global AV45 amyloid-PET uptake, and ApoE ε4 carrier status. Asterisks indicate p-values below 0.05. Significance of the indirect effect was determined using bootstrapping with 10,000 iterations

binding sites to confound our analyses. However, in the current study the AV1451 signal within the hippocampus may be affected by off-target binding in the choroid plexus, hence these results await replication using second generation tau tracers with a better off-target binding profile. Importantly, however, the BIN1 rs744373 association with higher tau was not limited to the hippocampal AV1451 tau-PET, but was widespread within the brain, rendering it highly unlikely that spill-off from regions of unspecific AV1451 tracer binding accounted for the current results.

In conclusion, our results show an association between BIN1 rs744373 risk-allele carriage and in vivo assessed tau pathology in elderly subjects with and without amyloid pathology. The current findings suggest that tau pathology provides a key link that underlies the association between BIN1 genetic variants and cognitive impairment. Future studies may further test whether BIN1 risk variants are associated with higher rates of increases in pathological tau and faster cognitive decline and conversion to dementia. It is currently unclear whether BIN1 genetic variants are associated with tauopathies other than AD. A recent study reported no association between BIN rs744373 and fronto-temporal dementia^{[57](#page-10-0)}, however, the association between BIN1 and tau pathology in conditions other than AD remains to be tested in the future. From a clinical point of view, our findings encourage future research in targeting BIN1 protein modification as a potential therapeutic approach to reduce levels of tau pathology. Preclinical studies showed that knock-down of BIN1 reduced tau-related toxicity in a drosophila model of neurodegeneration¹⁵. Understanding the exact mechanisms of the association between BIN1 and tau pathology are of pivotal clinical importance, since tau is the best predictor of clinical severity in contrast to $\text{A}\beta$ deposition⁵⁸. Neuroimaging of tau PET may be a marker to monitor treatment effects of disease modifying therapies targeting BIN1.

Methods

Study design. We included 89 participants from ADNI phase 3 (ClinicalTrials.gov ID: NCT02854033) in whom 18F-AV1451 tau-PET was obtained. 18F-AV1451 PET was added only in phase 3 of ADNI, and is thus only available in a smaller subset of the large ADNI cohort. The current set of 89 subjects resulted from inclusion criteria of the availability of T1-weighted MRI, ¹⁸F-AV45 amyloid-PET, cognitive and GWAS data in addition to 18F-AV1451 tau-PET. All imaging modalities had to be obtained at the same study visit. Selection bias was tested against the entire ADNI cohort of 1784 subjects. Here, we found no differences in gender or education between our selected sample and the entire ADNI sample, however, the mean age of the selected sample (\sim 78.7 years) was significantly (p < 0.05) higher than the mean age of the entire ADNI cohort (~73.7 years). Subjects were clinically classified by ADNI centers as cognitively normal (CN, $MMSE > 24$, $CDR = 0$, nondepressed) or mild cognitively impaired (MCI; MMSE > 24, CDR = 0.5, objective memory-loss on the education adjusted Wechsler Memory Scale II, preserved activities of daily living)^{[59](#page-10-0)}. Subjects with AD dementia were excluded from the analysis due to the small number of cases that met our inclusion criteria ($n = 3$). The BIN1 rs744373 genotype was extracted from GWAS data provided by the ADNI genetics core, where whole-genome sequencing was conducted using the Ilumina Omni 2.5 M Bead Chip. For a detailed description of the whole-genome sequencing methods, please refer to a previous publication by the ADNI genetics core^{[60](#page-10-0)}. Subjects were assigned to the *BIN1* rs744373 risk-group ($n = 40$) when carrying at least one G-allele¹⁴. Ethical approval was obtained by the ADNI investigators at each study site, all participants provided written informed consent and all work complied with ethical regulations for work with human participants.

Image acquisition. All imaging data was downloaded from the ADNI loni image archive [\(https://ida.loni.usc.edu](https://ida.loni.usc.edu)). Structural MRI in ADNI3 was recorded using a 3D T1-weighted MPRAGE sequence with 1 mm isotropic voxel-space and a TR = 2300 ms. Detailed sequence parameters can be found online at [\(http://adni.loni.usc.](http://adni.loni.usc.edu/wp-content/uploads/2017/07/ADNI3-MRI-protocols.pdf) [edu/wp-content/uploads/2017/07/ADNI3-MRI-protocols.pdf](http://adni.loni.usc.edu/wp-content/uploads/2017/07/ADNI3-MRI-protocols.pdf)).

18F-AV1451 tau-PET was acquired 75–105 min post-injection of 18F-AV1451, in 6×5 minute time frames. ¹⁸F-AV-45 Florbetapir amyloid PET scans were obtained during 4 × 5 min time frames measured 50–70 min post-injection of 18F-AV45 [\(http://](http://adni.loni.usc.edu/wp-content/uploads/2010/05/ADNI2_PET_Tech_Manual_0142011.pdf) [adni.loni.usc.edu/wp-content/uploads/2010/05/ADNI2_PET_Tech_Manual_0142011.](http://adni.loni.usc.edu/wp-content/uploads/2010/05/ADNI2_PET_Tech_Manual_0142011.pdf) [pdf](http://adni.loni.usc.edu/wp-content/uploads/2010/05/ADNI2_PET_Tech_Manual_0142011.pdf)).

For both AV1451 tau- and AV45 amyloid-PET we downloaded partially preprocessed data where dynamically acquired image frames are first registered to an

AC-PC orientation and standard voxel image grid and subsequently averaged to obtain a single image for each PET modality. Having PET images in a standard image matrix facilitates the combination of PET images from different scanners. For further details please see refer to the ADNI website ([http://adni.loni.usc.edu/methods/pet-analysis](http://adni.loni.usc.edu/methods/pet-analysis-method/pet-analysis/)[method/pet-analysis/](http://adni.loni.usc.edu/methods/pet-analysis-method/pet-analysis/)).

Image preprocessing. All MRI and PET images were inspected for artifacts prior to preprocessing. We applied two processing pipelines to PET data, following preestablished protocols to evaluate regional and global AV1451 tau- and AV45 amyloid-PET uptake. First, we used a SPM12-based pipeline to obtain stagespecific AV45 amyloid-PET SUVR scores³⁴. Second, we applied a Freesurfer-based pipeline to obtain global AV45 amyloid-^{[61](#page-10-0)} as well as global and regional AV1451 tau-PET SUVRs (i.e., Braak stage ROIs)³³.

For the pre-established SPM12-based pipeline^{[62](#page-10-0)-[67](#page-10-0)}, native-space structural MRI images were first segmented into gray matter (GM), white matter (WM) and CSF maps using SPMs new segment approach. Using SPMs high-dimensional DARTEL warping algorithm, we estimated subject-specific flow-fields to nonlinearly transform all GM, WM, and CSF maps to a sample-specific template that was determined in an iterative procedure. Using affine transformation, this sample-specific template was subsequently normalized to Montreal Neurological Institute (MNI) standard space. Next, subject-specific AV45 amyloid-PET images were co-registered to the corresponding high-resolution T1 image and subsequently DARTEL warped to MNI standard space. We did not spatially smooth the images to avoid spill over between adjacent regions during ROI-based analyses.

For the Freesurfer-based pipeline (Version 5.3), we applied volumetric segmentation to the high-resolution native-space structural MRI images, where subcortical and cortical areas are segmented automatically using the probabilistic Desikan–Killiany Atlas⁶⁸. The segmented anatomical ROIs from the highresolution structural MRI images were then applied to the co-registered AV45 amyloid- and AV1451 tau-PET images to extract ROI-based values. To obtain SUVR scores, all ROI values were normalized to the mean uptake of the whole cerebellum for AV45 data, and to the mean uptake of the inferior cerebellar gray for AV1451 data, following previous recommendations^{32,61}.

Amyloid staging. For AV45 amyloid-PET, we assessed global amyloid-PET levels that are commonly used for subject stratification into Aβ-positive/negative plus a more fine-grained anatomical amyloid staging system that was introduced recentl[y34](#page-10-0). For global amyloid load we computed global AV45 amyloid-PET SUVRs using an established Freesurfer pipeline. In brief, we averaged Freesurferdefined SUVR (normalized to the whole cerebellum) scores across lateral and medial frontal, anterior, and posterior cingulate, lateral parietal and lateral temporal regions[61.](#page-10-0) Based on these scores, Aβ-positivity was defined as a global AV45 amyloid-PET SUVR > 1.11⁶⁹. Summary statistics on amyloid status can be found in table 1.

We further assessed local amyloid levels using a 4-stage model, that suggests amyloid deposition to initiate in the temporobasal and mediofrontal areas with subsequent affection of the associative neocortex, primary sensorimotor areas and lastly the basal ganglia 34 . To this end, we used the MNI normalized AV45 amyloid-PET images from our SPM12 pipeline where we determined the mean scores within the four amyloid stage ROIs (shown in Fig. [1](#page-3-0)b) that were built using the MNI-space based Harvard–Oxford brain atlas following a previously described protocol^{[34](#page-10-0)}. Again, these mean values were intensity normalized to Freesurfer derived whole-cerebellar AV45 uptake to obtain SUVR scores.

Tau staging. For tau, we also obtained global as well as stage-specific AV1451 tau-PET SUVR scores.

For global tau, we averaged the size-weighted Freesurfer-ROI SUVRs across all Desikan–Killiany atlas regions, excluding the cerebellum, thalamus and basal ganglia (i.e., typical regions of AV1451 off-target binding) following a previously described approach³². For stage-specific AV1451 tau-PET uptake, we applied a recently described Braak-ROI staging system that allows application of the post-
mortem established tau staging system to tau PET imaging^{[33](#page-10-0)}. Here, we obtained size-weighted Freesurfer-ROI SUVRs for each Braak stage ROI, from Braak stage I (i.e., entorhinal cortex) to Braak stage VI (i.e., primary sensorimotor & primary visual cortex). A list of ROIs that are included within each Braak stage ROI can be found online [\(https://adni.bitbucket.io/reference/docs/UCBERKELEYAV1451/](https://adni.bitbucket.io/reference/docs/UCBERKELEYAV1451/UCBERKELEYAV1451_Methods_FINAL.pdf) [UCBERKELEYAV1451_Methods_FINAL.pdf](https://adni.bitbucket.io/reference/docs/UCBERKELEYAV1451/UCBERKELEYAV1451_Methods_FINAL.pdf)). A surface rendering of the Braak ROIs is shown in Fig. [1a](#page-3-0). Note that we excluded the thalamus or basal ganglia ROIs for all AV1451 tau-PET analyses, due to known off-target binding of the AV1451 tracer in these regions.

mRNA expression levels of BIN1. Regional gene expression was obtained from publicly available microarray measurements of regional mRNA expression based on post-mortem data from the Allen Brain Atlas [\(http://human.brain](http://human.brain-map.org/)[map.org/](http://human.brain-map.org/); RRID: SCR007416). The Allen Brain atlas includes more than 60,000 microarray probes collected from 3700 autopsy-based brain tissue samples from a total of 6 subjects aged 24–57 with no known history of neurological or psychiatric conditions^{[35](#page-10-0),36}. Microarray-based log₂ expression values of

20,737 genes within each of the 3700 samples were mapped back into MNI standard space by the Allen Brain Institute using stereotactic coordinates of the examined probes. The whole gene expression data has been recently mapped to the Freesurfer-based Desikan–Killiany atlas as median gene expression for probes falling within each of the 68 atlas ROIs³⁷. Here, we specifically extracted median expression of BIN1 mRNA within these Desikan–Killiany ROIs, to test associations between BIN1 expression and AV1451 tau-PET uptake. Since microarray assessments and thus BIN1 mRNA expression of all 6 Allen brain atlas subjects were available only for the left hemisphere (vs. 2 subjects for the right hemisphere), we restricted the analysis of BIN1 mRNA expression data to the more robust estimates of the left hemisphere in line with previous studies^{[70](#page-10-0)-72}.

Statistical analysis. Group demographics and baseline characteristics were compared between groups (i.e., diagnosis & BIN1 rs744373 status) using ANOVAs for continuous measures and Chi-squared tests for categorical measures. Global and regional AV45 amyloid- and AV1451 tau-PET SUVR scores were logtransformed prior to analysis to approximate a normal distribution.

For our main analysis, we tested whether BIN1 rs744373 risk-allele carriage was associated with increased AV1451 tau-PET uptake. To this end, we applied ANCOVAs to test whether presence of the BIN1 rs744373 risk allele had an effect on global or regional (i.e., Braak stage ROIs) AV1451 tau-PET SUVRs, controlling for age, gender, education, diagnosis, and ApoE ε4 carrier status and GM volume of the respective tau ROI. To assess any effects of the BIN1 rs744373 SNP on amyloid, we tested the same models this time using global or regional (i.e., amyloid-stage ROIs) AV45 amyloid-PET SUVRs as the dependent variable. Lastly, we tested whether BIN1 rs744373 risk-allele carriage was associated with higher AV1451 tau-PET SUVR independent of amyloid. To this end, we conducted linear regression with global AV1451 tau-PET SUVR as a dependent variable and global AV45 amyloid-PET SUVR and BIN1 rs744373 status as independent variables controlling for age, gender, education, ApoE ε4 carrier status, and diagnosis. Using the same covariates as described for the previous model, we further tested the interaction between BIN1 rs744373 and global AV45 amyloid-PET or Aβ status (Aβ− or Aβ+).

Next, we tested whether higher local BIN1 mRNA expression levels were associated with an increased likelihood of developing abnormal tau. To this end, we determined BIN1 mRNA expression using the Allen brain atlas data in Desikan–Killiany atlas space and determined the group-median regional tau PET SUVRs for corresponding anatomical regions. We then surface-mapped both group-median tau PET SUVRs and BIN1 mRNA expression and tested the Pearson–Moment correlation between regional BIN1 mRNA expression and tau load, applying a two-tailed alpha threshold of 0.05. We repeated this analysis based on 1000 randomly drawn bootstrapped samples to determine the 95% CI of the correlation coefficient between $B\tilde{N1}$ mRNA expression and AV1451 tau-PET uptake.

Lastly, we assessed whether BIN1 rs744373 risk-allele carriage was associated with worse memory performance, and whether this association was mediated by tau pathology. To test this, we conducted mediation analysis, testing whether the association between BIN1 rs744373 and ADNI-MEM was mediated via global AV1451 tau-PET uptake. Significance of the mediation effect was determined using 10,000 bootstrapped iterations, where each path of the model was controlled for global AV45 amyloid-PET SUVR, age, gender, education, and diagnosis and ApoE ε4 carrier status.

All statistical analyses were conducted with R statistical software. P-values were considered significant when meeting a two-tailed alpha threshold of 0.05. When a single hypothesis was tested multiple times, we also report Bonferroni-corrected p-values in case of significant uncorrected p-values.

Reporting summary. Further information on experimental design is available in the Nature Research Reporting Summary linked to this article.

Data availability

The data that support the findings of this study were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) and are available from the ADNI database ([adni.loni.usc.edu\)](http://adni.loni.usc.edu) upon registration and compliance with the data use agreement. A list including the anonymized participant identifiers of the currently used sample and the source file can be downloaded from the ADNI database [\(http://adni.loni.usc.edu/](http://adni.loni.usc.edu/)). The R-script used for the current study can be obtained from the first author upon request.

Received: 12 November 2018 Accepted: 14 March 2019 Published online: 16 April 2019

References

1. Selkoe, D. J. & Hardy, J. The amyloid hypothesis of Alzheimer's disease at 25 years. EMBO Mol. Med. 8, 595–608 (2016).

- 2. Ridge, P. G. et al. Assessment of the genetic variance of late-onset Alzheimer's disease. Neurobiol. Aging 41, 200 e213–200 e220 (2016).
- 3. Seshadri, S. et al. Genome-wide analysis of genetic loci associated with Alzheimer disease. JAMA 303, 1832–1840 (2010).
- 4. Zhu, R., Liu, X. & He, Z. The bridging integrator 1 gene polymorphism rs744373 and the risk of Alzheimer's disease in caucasian and asian populations: an updated meta-analysis. Mol. Neurobiol. 54, 1419–1428 (2017).
- 5. Hollingworth, P. et al. Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with Alzheimer's disease. Nat. Genet. 43, 429–435 (2011).
- 6. Wijsman, E. M. et al. Genome-wide association of familial late-onset Alzheimer's disease replicates BIN1 and CLU and nominates CUGBP2 in interaction with APOE. PLoS Genet. 7, e1001308 (2011).
- 7. Almeida, J. F. F., Dos Santos, L. R., Trancozo, M. & de Paula, F. Updated meta-analysis of BIN1, CR1, MS4A6A, CLU, and ABCA7 variants in Alzheimer's disease. J. Mol. Neurosci. 64, 471–477 (2018).
- 8. Antunez, C. et al. The membrane-spanning 4-domains, subfamily A (MS4A) gene cluster contains a common variant associated with Alzheimer's disease. Genome Med. 3, 33 (2011).
- 9. Hu, X. et al. Meta-analysis for genome-wide association study identifies multiple variants at the BIN1 locus associated with late-onset Alzheimer's disease. PLoS ONE 6, e16616 (2011).
- 10. AlzGene. Meta-Analysis of all published AD association studies (case-control only) rs744373. Alzgene.org retreived on 23 August 2018.
- 11. Sakamuro, D., Elliott, K. J., Wechsler-Reya, R. & Prendergast, G. C. BIN1 is a novel MYC-interacting protein with features of a tumour suppressor. Nat. Genet. 14, 69–77 (1996).
- 12. Butler, M. H. et al. Amphiphysin II (SH3P9; BIN1), a member of the amphiphysin/Rvs family, is concentrated in the cortical cytomatrix of axon initial segments and nodes of ranvier in brain and around T tubules in skeletal muscle. J. Cell. Biol. 137, 1355–1367 (1997).
- 13. Prokic, I., Cowling, B. S. & Laporte, J. Amphiphysin 2 (BIN1) in physiology and diseases. J. Mol. Med (Berl) 92, 453–463 (2014).
- 14. Tan, M. S., Yu, J. T. & Tan, L. Bridging integrator 1 (BIN1): form, function, and Alzheimer's disease. Trends Mol. Med. 19, 594–603 (2013).
- 15. Chapuis, J. et al. Increased expression of BIN1 mediates Alzheimer genetic risk by modulating tau pathology. Mol. Psychiatry 18, 1225–1234 (2013).
- 16. Holler, C. J. et al. Bridging integrator 1 (BIN1) protein expression increases in the Alzheimer's disease brain and correlates with neurofibrillary tangle pathology. J. Alzheimers Dis. 42, 1221–1227 (2014).
- 17. Bungenberg, J. et al. Gene expression variance in hippocampal tissue of temporal lobe epilepsy patients corresponds to differential memory performance. Neurobiol. Dis. 86, 121–130 (2016).
- 18. Wu, J. W. et al. Small misfolded Tau species are internalized via bulk endocytosis and anterogradely and retrogradely transported in neurons. J. Biol. Chem. 288, 1856–1870 (2013).
- 19. Calafate, S., Flavin, W., Verstreken, P. & Moechars, D. Loss of Bin1 promotes the propagation of Tau pathology. Cell Rep. 17, 931–940 (2016).
- 20. Yu, L. et al. Association of Brain DNA methylation in SORL1, ABCA7, HLA-DRB5, SLC24A4, and BIN1 with pathological diagnosis of Alzheimer disease. JAMA Neurol. 72, 15–24 (2015).
- 21. De Rossi, P. et al. Aberrant accrual of BIN1 near Alzheimer's disease amyloid deposits in transgenic models. Brain Pathology 0, 1–17 [https://doi.org/](https://doi.org/10.1111/bpa.12687) [10.1111/bpa.12687](https://doi.org/10.1111/bpa.12687) (2018).
- 22. Ubelmann, F. et al. Bin1 and CD2AP polarise the endocytic generation of beta-amyloid. EMBO Rep. 18, 102–122 (2017).
- 23. Wang, H. F. et al. Bridging Integrator 1 (BIN1) genotypes mediate Alzheimer's disease risk by altering neuronal degeneration. J. Alzheimer's Dis. 52, 179–190 (2016).
- 24. Kauwe, J. S. et al. Fine mapping of genetic variants in BIN1, CLU, CR1 and PICALM for association with cerebrospinal fluid biomarkers for Alzheimer's disease. PLoS ONE 6, e15918 (2011).
- 25. Buerger, K. et al. CSF phosphorylated tau protein correlates with neocortical neurofibrillary pathology in Alzheimer's disease. Brain 129, 3035–3041 (2006).
- 26. La Joie, R. et al. Associations between [(18)F]AV1451 tau PET and CSF measures of tau pathology in a clinical sample. Neurology 90, e282–e290 (2018).
- 27. Mattsson, N. et al. 18)F-AV-1451 and CSF T-tau and P-tau as biomarkers in Alzheimer's disease. EMBO Mol. Med. 9, 1212–1223 (2017).
- 28. Sato, C. et al. Tau kinetics in neurons and the human central nervous system. Neuron 98, 861–864 (2018).
- 29. Biffi, A. et al. Genetic variation and neuroimaging measures in Alzheimer disease. Arch. Neurol. 67, 677–685 (2010).

- 30. Marquie, M. et al. Validating novel tau positron emission tomography tracer [F-18]-AV-1451 (T807) on postmortem brain tissue. Ann. Neurol. 78, 787–800 (2015).
- 31. Braak, H. & Braak, E. Neuropathological stageing of Alzheimer-related changes. Acta Neuropathol. 82, 239–259 (1991).
- 32. Maass, A. et al. Comparison of multiple tau-PET measures as biomarkers in aging and Alzheimer's disease. Neuroimage 157, 448–463 (2017).
- 33. Scholl, M. et al. PET imaging of Tau deposition in the aging human brain. Neuron 89, 971–982 (2016).
- 34. Grothe, M. J. et al. In vivo staging of regional amyloid deposition. Neurology 89, 2031–2038 (2017).
- 35. Hawrylycz, M. et al. Canonical genetic signatures of the adult human brain. Nat. Neurosci. 18, 1832–1844 (2015).
- 36. Hawrylycz, M. J. et al. An anatomically comprehensive atlas of the adult human brain transcriptome. Nature 489, 391–399 (2012).
- 37. French, L. & Paus, T. A FreeSurfer view of the cortical transcriptome generated from the Allen Human Brain Atlas. Front. Neurosci. 9, 323 (2015).
- 38. Crane, P. K. et al. Development and assessment of a composite score for memory in the Alzheimer's Disease Neuroimaging Initiative (ADNI). Brain. Imaging Behav. 6, 502–516 (2012).
- 39. De Rossi, P. et al. BIN1 localization is distinct from Tau tangles in Alzheimer's disease. Matters (Zur) 1–7 [https://doi.org/10.19185/](https://doi.org/10.19185/matters.201611000018) [matters.201611000018](https://doi.org/10.19185/matters.201611000018) (2017).
- 40. Villemagne, V. L. et al. Amyloid β deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study. Lancet Neurol. 12, 357–367 (2013).
- 41. Sottejeau, Y. et al. Tau phosphorylation regulates the interaction between BIN1's SH3 domain and Tau's proline-rich domain. Acta Neuropathol. Commun. 3, 58 (2015).
- 42. Malki, I. et al. Regulation of the interaction between the neuronal BIN1 isoform 1 and Tau proteins—role of the SH3 domain. Febs. J. 284, 3218–3229 (2017).
- 43. Lasorsa, A. et al. Structural basis of Tau interaction with BIN1 and regulation by Tau phosphorylation. Front. Mol. Neurosci. 11, 421 (2018).
- 44. Franzmeier, N. et al. Functional connectivity associated with tau levels in ageing, Alzheimer's, and small vessel disease. Brain 0, 1–15 [https://doi.org/](https://doi.org/10.1093/brain/awz026) [10.1093/brain/awz026](https://doi.org/10.1093/brain/awz026) (2019).
- 45. de Calignon, A. et al. Propagation of tau pathology in a model of early Alzheimer's disease. Neuron 73, 685–697 (2012).
- Takeda, S. et al. Seed-competent high-molecular-weight tau species accumulates in the cerebrospinal fluid of Alzheimer's disease mouse model and human patients. Ann. Neurol. 80, 355–367 (2016).
- 47. Vivot, A. et al. Association of Alzheimer's related genotypes with cognitive decline in multiple domains: results from the Three-City Dijon study. Mol. Psychiatry 20, 1173–1178 (2015).
- 48. Barral, S. et al. Genotype patterns at PICALM, CR1, BIN1, CLU, and APOE genes are associated with episodic memory. Neurology 78, 1464–1471 (2012).
- 49. Maass, A. et al. Entorhinal Tau pathology, episodic memory decline, and neurodegeneration in aging. J. Neurosci. 38, 530–543 (2018).
- 50. Bertram, L., McQueen, M. B., Mullin, K., Blacker, D. & Tanzi, R. E. Systematic meta-analyses of Alzheimer disease genetic association studies: the AlzGene database. Nat. Genet. 39, 17–23 (2007).
- 51. Karch, C. M. & Goate, A. M. Alzheimer's disease risk genes and mechanisms of disease pathogenesis. Biol. Psychiatry 77, 43–51 (2015).
- 52. De Rossi, P. et al. Predominant expression of Alzheimer's disease-associated BIN1 in mature oligodendrocytes and localization to white matter tracts. Mol. Neurodegener. 11, 59 (2016).
- 53. McKenzie, A. T. et al. Multiscale network modeling of oligodendrocytes reveals molecular components of myelin dysregulation in Alzheimer's disease. Mol. Neurodegener. 12, 82 (2017).
- 54. Mito, R. et al. Fibre-specific white matter reductions in Alzheimer's disease and mild cognitive impairment. Brain 141, 888–902 (2018).
- 55. Araque Caballero, M. A. et al. White matter diffusion alterations precede symptom onset in autosomal dominant Alzheimer's disease. Brain 141, 3065–3080 (2018).
- 56. Marquie, M. et al. Lessons learned about [F-18]-AV-1451 off-target binding from an autopsy-confirmed Parkinson's case. Acta Neuropathol. Commun. 5, 75 (2017).
- 57. Ferrari, R. et al. A genome-wide screening and SNPs-to-genes approach to identify novel genetic risk factors associated with frontotemporal dementia. Neurobiol. Aging 36, 2904 e2913–2904 e2926 (2015).
- 58. Aschenbrenner, A. J., Gordon, B. A., Benzinger, T. L. S., Morris, J. C. & Hassenstab, J. J. Influence of tau PET, amyloid PET, and hippocampal

volume on cognition in Alzheimer disease. Neurology 91, e859–e866 (2018)

- 59. Petersen, R. C. et al. Alzheimer's Disease Neuroimaging Initiative (ADNI): clinical characterization. Neurology 74, 201–209 (2010).
- 60. Saykin, A. J. et al. Genetic studies of quantitative MCI and AD phenotypes in ADNI: progress, opportunities, and plans. Alzheimers Dement. 11, 792–814 (2015).
- 61. Landau, S. M. et al. Amyloid deposition, hypometabolism, and longitudinal cognitive decline. Ann. Neurol. 72, 578–586 (2012).
- 62. Franzmeier, N. et al. Resting-state connectivity of the left frontal cortex to the default mode and dorsal attention network supports reserve in mild cognitive impairment. Front. Aging Neurosci. 9, 264 (2017).
- 63. Franzmeier, N. et al. Left frontal cortex connectivity underlies cognitive reserve in prodromal Alzheimer disease. Neurology 88, 1054–1061 (2017).
- 64. Franzmeier, N. et al. Resting-state global functional connectivity as a biomarker of cognitive reserve in mild cognitive impairment. Brain. Imaging Behav. 11, 368–382 (2017).
- 65. Franzmeier, N. et al. Cognitive reserve moderates the association between functional network anti-correlations and memory in MCI. Neurobiol. Aging 50, 152–162 (2017).
- 66. Franzmeier, N. et al. Left frontal hub connectivity delays cognitive impairment in autosomal-dominant and sporadic Alzheimer's disease. Brain 141, 1186–1200 (2018).
- 67. Franzmeier, N. et al. The left frontal cortex supports reserve in aging by enhancing functional network efficiency. Alzheimers Res. Ther. 10, 28 (2018).
- 68. Desikan, R. S. et al. An automated labeling system for subdividing the human cerebral cortex on MRI scans into gyral based regions of interest. Neuroimage 31, 968–980 (2006).
- 69. Landau, S. M. et al. Amyloid-beta imaging with Pittsburgh compound B and florbetapir: comparing radiotracers and quantification methods. J. Nucl. Med. 54, 70–77 (2013).
- 70. Rittman, T. et al. Regional expression of the MAPT gene is associated with loss of hubs in brain networks and cognitive impairment in Parkinson disease and progressive supranuclear palsy. Neurobiol. Aging 48, 153–160 (2016).
- 71. Romme, I. A., de Reus, M. A., Ophoff, R. A., Kahn, R. S. & van den Heuvel, M. P. Connectome disconnectivity and cortical gene expression in patients with schizophrenia. Biol. Psychiatry 81, 495–502 (2017).
- 72. Grothe, M. J. et al. Molecular properties underlying regional vulnerability to Alzheimer's disease pathology. Brain 141, 2755–2771 (2018).

Acknowledgements

Data used in preparation of this manuscript were obtained from the ADNI database (adni. loni.usc.edu). As such, the investigators within the ADNI study contributed to the design and implementation of ADNI and/or provided data but did not participate in analysis or writing of this paper. The study was funded by grants from the Alzheimer Forschung Initiative (AFI, Grant 15035 to ME) and European Commission (Grant 334259 to ME). ADNI data collection and sharing for this project was funded by the ADNI (National Institutes of Health Grant U01 AG024904) and DOD ADNI (Department of Defense award number W81XWH-12-2-0012). ADNI is funded by the National Institute on Aging, the National Institute of Biomedical Imaging, and Bioengineering, and through contributions from the following: AbbVie, Alzheimer's Association; Alzheimer's Drug Discovery Foundation; Araclon Biotech; BioClinica, Inc.; Biogen; Bristol-Myers Squibb Company; CereSpir, Inc.; Cogstate; Eisai Inc.; Elan Phar- maceuticals, Inc.; Eli Lilly and Company; EuroImmun; F. Hoffmann-La Roche Ltd and its affiliated company Genentech, Inc.; Fujirebio; GE Healthcare; IXICO Ltd.; Janssen Alzheimer Immunotherapy Research & Development, LLC.; Johnson & Johnson Pharmaceutical Research & Development LLC.; Lumosity; Lundbeck; Merck & Co., Inc.; Meso Scale Diagnostics, LLC.; NeuroRx Research; Neurotrack Technologies; Novartis Pharmaceuticals Corporation; Pfizer Inc.; Piramal Imaging; Servier; Takeda Pharmaceutical Company; and Transition Therapeutics. The Canadian Institutes of Health Research is providing funds to support ADNI clinical sites in Canada. Private sector contributions are facilitated by the Foundation for the National Institutes of Health ([www.fnih.org\)](http://www.fnih.org).

Author contributions

N.F.: study concept and design, data processing, statistical analysis, interpretation of the results, and writing the manuscript. A.R.: data processing and critical revision of the manuscript. J.N.: data processing and critical revision of the manuscript. M.E.: study concept and design, interpretation of the results, and writing the manuscript. ADNI provided all data used for this study.

Additional information

Supplementary Information accompanies this paper at [https://doi.org/10.1038/s41467-](https://doi.org/10.1038/s41467-019-09564-5) [019-09564-5.](https://doi.org/10.1038/s41467-019-09564-5)

Competing interests: The authors declare no competing interests.

Reprints and permission information is available online at [http://npg.nature.com/](http://npg.nature.com/reprintsandpermissions/) [reprintsandpermissions/](http://npg.nature.com/reprintsandpermissions/)

Journal peer review information: Nature Communications thanks Dieder Moechars, Michael Murphy and the other anonymous reviewer(s) for their contribution to the peer review of this work. Peer reviewer reports are available.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons \bigcirc $\left($ cc Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit [http://creativecommons.org/](http://creativecommons.org/licenses/by/4.0/) [licenses/by/4.0/](http://creativecommons.org/licenses/by/4.0/).

© The Author(s) 2019

The Alzheimer's Disease Neuroimaging Initiative (ADNI)

Michael W. Weiner², Paul Aisen³, Ronald Petersen⁴, Clifford R. Jack⁴, William Jagust⁵, John Q. Trojanowski⁶, Arthur W. Toga⁷, Laurel Beckett⁸, Robert C. Green⁹, Andrew J. Saykin¹⁰, John Morris¹¹, Leslie M. Shaw⁶, Zaven Khachaturian^{8,12}, Greg Sorensen¹³, Lew Kuller¹⁴, Marcus Raichle¹¹, Steven Paul¹⁵, Peter Davies¹⁶, Howard Fillit¹⁷, Franz Hefti¹⁸, David Holtzman¹¹, Marek M. Mesulam¹⁹, William Potter²⁰, Peter Snyder²¹, Adam Schwartz²², Tom Montine²³, Ronald G. Thomas²³, Michael Donohue²³, Sarah Walter²³, Devon Gessert²³, Tamie Sather²³, Gus Jiminez²³, Danielle Harvey⁸, Matthew Bernstein⁴, Paul Thompson²⁴, Norbert Schuff^{2,8}, Bret Borowski⁴, Jeff Gunter⁴, Matt Senjem⁴, Prashanthi Vemuri⁴, David Jones⁴, Kejal Kantarci⁴, Chad Ward⁴, Robert A. Koeppe²⁵, Norm Foster²⁶, Eric M. Reiman²⁷, Kewei Chen²⁷, Chet Mathis¹⁴, Susan Landau⁵, Nigel J. Cairns¹¹, Erin Householder¹¹, Lisa Taylor-Reinwald¹¹, Virginia Lee⁶, Magdalena Korecka⁶, Michal Figurski⁶, Karen Crawford⁷, Scott Neu⁷, Tatiana M. Foroud¹⁰, Steven G. Potkin²⁸, Li Shen¹⁰, Kelley Faber¹⁰, Sungeun Kim¹⁰, Kwangsik Nho¹⁰, Leon Thal³, Neil Buckholtz²⁹, Marylyn Albert³⁰, Richard Frank³¹, John Hsiao²⁹, Jeffrey Kaye³², Joseph Quinn³², Betty Lind³², Raina Carter³², Sara Dolen³², Lon S. Schneider⁷, Sonia Pawluczyk⁷, Mauricio Beccera⁷, Liberty Teodoro⁷, Bryan M. Spann⁷, James Brewer³, Helen Vanderswag³, Adam Fleisher^{3,27}, Judith L. Heidebrink²⁵, Joanne L. Lord²⁵, Sara S. Mason⁴, Colleen S. Albers⁴, David Knopman⁴, Kris Johnson⁴, Rachelle S. Doody³³, Javier Villanueva-Meyer³³, Munir Chowdhury³³, Susan Rountree³³, Mimi Dang³³, Yaakov Stern³³, Lawrence S. Honig³³, Karen L. Bell³³, Beau Ances¹¹, Maria Carroll¹¹, Sue Leon¹¹, Mark A. Mintun¹¹, Stacy Schneider¹¹, Angela Oliver¹¹, Daniel Marson³⁴, Randall Griffith³⁴, David Clark³⁴, David Geldmacher³⁴, John Brockington³⁴, Erik Roberson³⁴, Hillel Grossman³⁵, Effie Mitsis³⁵, Leyla de Toledo-Morrell³⁶, Raj C. Shah³⁶, Ranjan Duara³⁷, Daniel Varon³⁷, Maria T. Greig³⁷, Peggy Roberts³⁷, Chiadi Onyike³⁰, Daniel D'Agostino³⁰, Stephanie Kielb³⁰, James E. Galvin³⁸, Brittany Cerbone³⁸, Christina A. Michel³⁸, Henry Rusinek³⁸, Mony J. de Leon³⁸, Lidia Glodzik³⁸, Susan De Santi³⁸, P Murali Doraiswamy³⁹, Jeffrey R. Petrella³⁹ Terence Z. Wong³⁹, Steven E. Arnold⁶, Jason H. Karlawish⁶, David Wolk⁶, Charles D. Smith⁴⁰, Greg Jicha⁴⁰, Peter Hardy⁴⁰, Partha Sinha⁴⁰, Elizabeth Oates⁴⁰, Gary Conrad⁴⁰, Oscar L. Lopez¹⁴, MaryAnn Oakley¹⁴, Donna M. Simpson³⁰, Anton P. Porsteinsson⁴¹, Bonnie S. Goldstein⁴¹, Kim Martin⁴¹, Kelly M. Makino⁴¹, M Saleem Ismail⁴¹, Connie Brand⁴¹, Ruth A. Mulnard²⁸, Gaby Thai²⁸, Catherine McAdams-Ortiz²⁸, Kyle Womack⁴², Dana Mathews⁴², Mary Quiceno⁴², Ramon Diaz-Arrastia⁴², Richard King⁴², Myron Weiner⁴², Kristen Martin-Cook⁴², Michael DeVous⁴², Allan I Levey⁴³, James J. Lah⁴³, Janet S. Cellar⁴³, Jeffrey M. Burns⁴⁴, Heather S. Anderson⁴⁴, Russell H. Swerdlow⁴⁴, Liana Apostolova²⁴, Kathleen Tingus²⁴, Ellen Woo²⁴, Daniel H.S. Silverman²⁴, Po H. Lu²⁴, George Bartzokis²⁴, Neill R. Graff-Radford⁴⁵, Francine Parfitt⁴⁵, Tracy Kendall⁴⁵, Heather Johnson⁴⁵, Martin R. Farlow¹⁰, Ann Marie Hake¹⁰, Brandy R. Matthews¹⁰,

Scott Herring¹⁰, Cynthia Hunt¹⁰, Christopher H. van Dyck⁴⁶, Richard E. Carson⁴⁶, Martha G. MacAvoy⁴⁶, Howard Chertkow⁴⁷, Howard Bergman⁴⁷, Chris Hosein⁴⁷, Ging-Yuek Robin Hsiung⁴⁸, Howard Feldman⁴⁸, Benita Mudge⁴⁸, Michele Assaly⁴⁸, Charles Bernick⁴⁹, Donna Munic⁴⁹, Andrew Kertesz⁵⁰, John Rogers⁵⁰, Dick Trost⁵⁰, Diana Kerwin¹⁹, Kristine Lipowski¹⁹, Chuang-Kuo Wu¹⁹, Nancy Johnson¹⁹, Carl Sadowsky⁵¹, Walter Martinez⁵¹, Teresa Villena⁵¹, Raymond Scott Turner⁵², Kathleen Johnson⁵², Brigid Reynolds⁵², Reisa A. Sperling⁹, Keith A. Johnson⁹, Gad Marshall⁹, Meghan Frey⁹, Barton Lane⁹, Allyson Rosen⁹, Jared Tinklenberg⁹, Marwan N. Sabbagh⁵³, Christine M. Belden⁵³, Sandra A. Jacobson⁵³, Sherye A. Sirrel⁵³, Neil Kowall⁵³, Ronald Killiany⁵⁴, Andrew E. Budson⁵⁴, Alexander Norbash⁵⁴, Patricia Lynn Johnson⁵⁴. Joanne Allard⁵⁵, Alan Lerner⁵⁶, Paula Ogrocki⁵⁶, Leon Hudson⁵⁶, Evan Fletcher⁸, Owen Carmichael⁸, John Olichney⁸, Charles DeCarli⁸, Smita Kittur⁵⁷, Michael Borrie⁵⁸, T-Y. Lee⁵⁸, Rob Bartha⁵⁸, Sterling Johnson⁵⁹, Sanjay Asthana⁵⁹, Cynthia M. Carlsson⁵⁹, Adrian Preda²⁴, Dana Nguyen²⁴, Pierre Tariot²⁶, Stephanie Reeder²⁶, Vernice Bates⁶⁰, Horacio Capote⁶⁰, Michelle Rainka⁶⁰, Douglas W. Scharre⁶¹, Maria Kataki⁶¹, Anahita Adeli⁶¹, Earl A. Zimmerman⁶², Dzintra Celmins⁶², Alice D. Brown⁶², Godfrey D. Pearlson⁶³, Karen Blank⁶³, Karen Anderson⁶³, Robert B. Santulli⁶⁴, Tamar J. Kitzmiller⁶⁴, Eben S. Schwartz⁶⁴, Kaycee M. Sink⁶⁵, Jeff D. Williamson⁶⁵, Pradeep Garg⁶⁵, Franklin Watkins⁶⁵, Brian R. Ott⁶⁶, Henry Querfurth⁶⁶, Geoffrey Tremont⁶⁶, Stephen Salloway⁶⁷, Paul Malloy⁶⁷, Stephen Correia⁶⁷, Howard J. Rosen², Bruce L. Miller², Jacobo Mintzer⁶⁸, Kenneth Spicer⁶⁸, David Bachman⁶⁸, Stephen Pasternak⁵⁰, Irina Rachinsky⁵⁰, Dick Drost⁵⁰, Nunzio Pomara⁶⁹, Raymundo Hernando⁶⁹, Antero Sarrael⁶⁹, Susan K. Schultz⁷⁰, Laura L. Boles Ponto⁷⁰, Hyungsub Shim⁷⁰, Karen Elizabeth Smith⁷⁰, Norman Relkin¹⁵, Gloria Chaing¹⁵, Lisa Raudin^{12,15}, Amanda Smith⁷¹, Kristin Fargher⁷¹, Balebail Ashok Raj⁷¹, Thomas Neylan², Jordan Grafman¹⁹, Melissa Davis³, Rosemary Morrison³, Jacqueline Hayes², Shannon Finley², Karl Friedl⁷², Debra Fleischman³⁶, Konstantinos Arfanakis³⁶, Olga James³⁹, Dino Massoglia⁶⁸, J Jay Fruehling⁵⁹, Sandra Harding⁵⁹, Elaine R. Peskind²³, Eric C. Petrie⁶¹, Gail Li⁶¹, Jerome A. Yesavage⁷³, Joy L. Taylor⁷³ & Ansgar J. Furst⁷³

²UC San Francisco, San Francisco, CA 94143, USA. ³UC San Diego, San Diego, CA 92093, USA. ⁴Mayo Clinic, Rochester, NY 14603, USA. ⁵UC Berkeley, Berkeley, CA 94720, USA. ⁶UPenn, Philadelphia, PA 9104, USA. ⁷USC, Los Angeles, CA 90089, USA. ⁸ Berkeley, Berkeley, CA 94720, USA. ⁶UPenn, Philadelphia, PA 9104, USA. ′USC, Los Angeles, CA 90089, USA. ⁸UC Davis, Davis, CA 95616, USA.
⁹Brigham and Women's Hospital/Harvard Medical School, Boston, MA 02115, USA. 80333, Germany. ¹⁴University of Pittsburgh, Pittsburgh, PA 15260, USA. ¹⁵Weill Cornell Medical College, Cornell University, New York City, NY 10065, USA. ¹⁶Albert Einstein College of Medicine of Yeshiva University, Bronx, NY 10461, USA. ¹⁷AD Drug Discovery Foundation, New York City,
NY 10019, USA. ¹⁸Acumen Pharmaceuticals, Livermore, CA 94551, USA. ¹⁹Nor ²⁰National Institute of Mental Health, Rockville, MD 20852, USA. ²¹Brown University, Providence, RI 02912, USA. ²²Eli Lilly, Indianapolis, IN 46225, USA. ²³University of Washington, Seattle, WA 98195, USA. ²⁴UCLA, Los Angeles, CA 90095, USA. ²⁵University of Michigan, Ann Arbor, MI 48109, USA. ²⁶University of Utah, Salt Lake City, UT 84112, USA. ²⁷Banner Alzheimer's Institute, Phoenix, AZ 85006, USA. ²⁸UC Irvine, Irvine, CA 92697, USA. ²⁹National Institute on Aging, Bethesda, MD 20892, USA. ³⁰Johns Hopkins University, Baltimore, MD 21218, USA. ³¹Richard Frank Consulting, Washington, DC 20001, USA. 32Oregon Health and Science University, Portland, OR 97239, USA. 33Baylor College of Medicine, Houston, TX 77030, USA. ³⁴University of Alabama, Birmingham, AL 35233, USA. ³⁵Mount Sinai School of Medicine, New York City, NY 10029, USA. ³⁶Rush University Medical Center, Chicago, IL 60612, USA. ³⁷Wien Center, Miami, FL 33140, USA. ³⁸New York University, New York City, NY 10003, USA. ³⁹Duke University Medical Center, Durham, NC 27710, USA. ⁴⁰University of Kentucky, Lexington, KY 0506, USA. ⁴¹University of Rochester Medical Center, Rochester, NY 14642, USA. ⁴²University of Texas Southwestern Medical School, Dallas, TX 75390, USA. ⁴³Emory University, Atlanta, GA 30322, USA. ⁴⁴Medical Center, University of Kansas, Kansas City, KS 66103, USA. ⁴⁵Mayo Clinic, Jacksonville, FL 32224, USA. 46Yale University School of Medicine, New Haven, CT 06510, USA. 47McGill University/Montreal-Jewish General Hospital, Montreal, QC H3T 1E2, Canada. ⁴⁸University of British Columbia Clinic for AD & Related Disorders, Vancouver, BC V6T 1Z3, Canada. ⁴⁹Cleveland Clinic Lou Ruvo Center for Brain Health, Las Vegas, NV 89106, USA. ⁵⁰St Joseph's Health Care, London, ON N6A 4V2, Canada. ⁵¹Premiere Research Institute, Palm Beach Neurology, Miami, FL 33407, USA. ⁵²Georgetown University Medical Center, Washington, DC 20007, USA. ⁵³Banner Sun Health Research Institute, Sun City, AZ 85351, USA. ⁵⁴Boston University, Boston, MA 02215, USA. ⁵⁵Howard University, Washington, DC 20059, USA. ⁵⁶Case Western Reserve University, Cleveland, OH 20002, USA. ⁵⁷Neurological Care of CNY, Liverpool, NY 13088, USA. ⁵⁸Parkwood Hospital, London, ON N6C 0A7, Canada. ⁵⁹University of Wisconsin, Madison, WI 53706, USA. ⁶⁰Dent Neurologic Institute, Amherst, NY 14226, USA. ⁶¹Ohio State University, Columbus, OH 43210, USA. ⁶²Albany Medical College, Albany, NY 12208, USA. ⁶³Hartford Hospital, Olin Neuropsychiatry Research Center, Hartford, CT 06114, USA. ⁶⁴Dartmouth- Hitchcock Medical Center, Lebanon, NH 03766, USA. ⁶⁵Wake Forest University Health Sciences, Winston-Salem, NC 27157, USA. ⁶⁶Rhode Island Hospital, Providence, RI 02903, USA. ⁶⁷Butler Hospital, Providence, RI 02906, USA. ⁶⁸Medical University South Carolina, Charleston, SC 29425, USA. ⁶⁹Nathan Kline Institute, Orangeburg, NY 10962, USA. ⁷⁰University of Iowa College of Medicine, Iowa City, IA 52242, USA. ⁷¹University of South Florida: USF Health Byrd Alzheimer's Institute, Tampa, FL 33613, USA. ⁷²Department of Defense, Arlington, VA 22350, USA. ⁷³Stanford University, Stanford, CA 94305, USA