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RESEARCH ARTICLE

Mouse liver injury induces hepatic macrophage FGF23 production

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Abstract

Fibroblast growth factor 23 (FGF23) is a bone marrow cell produced hormone that functions in the intestine and kidney to regulate phosphate homeostasis. Increased serum FGF23 is a well-established predictor of mortality in renal disease, but recent findings linking increased levels to hepatic and cardiac diseases have suggested that other organs are sources of FGF23 or targets of its effects. The potential ability of the liver to produce FGF23 in response to hepatocellular injury was therefore examined. Very low levels of *Fgf23* mRNA and FGF23 protein were detected in normal mouse liver, but the amounts increased markedly during acute liver injury from the hepatotoxin carbon tetrachloride. Serum levels of intact FGF23 were elevated during liver injury from carbon tetrachloride. Chronic liver injury induced by a high fat diet or elevated bile acids also increased hepatic FGF23 levels. Stimulation of toll-like receptor (TLR) 4-driven inflammation by gut-derived lipopolysaccharide (LPS) underlies many forms of liver injury, and LPS induced *Fgf23* in the liver as well as in other organs. The LPS-inducible cytokines IL-1 β and TNF increased hepatic *Fgf23* expression as did a TLR2 agonist Pam2CSK3. Analysis of *Fgf23* expression and FGF23 secretion in different hepatic cell types involved in liver injury identified the resident liver macrophage or Kupffer cell as a source of hepatic FGF23. LPS and cytokines selectively induced the hormone in these cells but not in hepatocytes or hepatic stellate cells. FGF23 failed to exert any autocrine effect on the inflammatory state of Kupffer cells but did trigger proinflammatory activation of hepatocytes. During liver injury inflammatory factors induce Kupffer cell production of FGF23 that may have a paracrine proinflammatory effect on hepatocytes. Liver-produced FGF23 may have systemic hormonal effects as well that influence diseases in other organs.

Introduction

Fibroblast growth factor 23 (FGF23) is a bone marrow cell produced hormone that regulates phosphate homeostasis through modulation of intestinal absorption and renal excretion. FGF23 circulates as both a full-length intact bioactive protein (iFGF23) and inactive cleaved

fragments (cFGF23) [1,2]. Serum FGF23 levels are increased in chronic kidney disease in direct proportion to the loss of renal function. In addition to causing hyperphosphatemia, elevated FGF23 is associated with chronic kidney disease progression and mortality [3,4]. Risk from increased FGF23 is out of proportion to the resultant alterations in bone-mineral metabolism, suggesting that high levels of circulating FGF23 have pathophysiological effects through mechanisms other than the dysregulation of phosphate homeostasis [5]. Although the primary focus on FGF23's effects have been in the kidney, elevated FGF23 has also been linked to disease outcomes in other organs such as the liver, heart and lung [6–10]. Defining mechanisms of excessive FGF23 production and its pathophysiological effects are of importance to a variety of tissues and the relationship of renal disease to diseases in other organs.

In addition to the standard mineral-related inducers of FGF23 which include phosphate, calcium, parathyroid hormone, and vitamin D, recent studies have implicated tissue inflammation as a cause of elevated FGF23 [11]. This inflammatory effect has been attributed to increased cleavage of iFGF23 to cFGF23 rather than from upregulation of iFGF23 production [11]. FGF23 has also been linked to inflammation by findings that the hormone may promote inflammation through stimulatory effects on immune cells [12,13].

Inflammation is critical to the outcome of many diseases including those in the liver. Most forms of liver injury are due in large part to an excessive inflammatory response [14]. Self-limited inflammation after liver injury performs the beneficial function of tissue repair, but sustained inflammation leads to hepatocyte injury and death and further stimulation of the immune response. Critical to the development of hepatic inflammation is the initiation of an innate immune response by pattern recognition receptor stimulation from gut-derived lipopolysaccharide (LPS) [15]. It is the effects of LPS that promote liver injury from hepatotoxins such as carbon tetrachloride rather than the toxin itself or its metabolites [16]. LPS-activated hepatic macrophages release proinflammatory cytokines such as tumor necrosis factor (TNF), interleukin-1 β (IL-1 β), and interferon- γ (IFN γ) that induce hepatocyte injury and death and further amplify inflammation [17,18]. Serum FGF23 has been shown to be increased *in vivo* by bacteria or bacterial products such as LPS [13,19–21]. The organ sources of FGF23 are unclear but have been attributed to spleen and bone [13,19–21] and not the liver [20].

Liver injury may lead to disease in other organs. The most prevalent chronic liver disease, nonalcoholic fatty liver disease (NAFLD), has been linked to diseases in many other organs [22]. NAFLD has been implicated in promoting both cardiovascular disease and chronic renal failure. The mechanisms by which hepatocellular injury could trigger disease in the heart and kidney are unknown but likely involve the release of serum factors such as the products of inflammation.

To determine whether the liver could be a source of FGF23 during inflammation, the study examined whether hepatic expression of FGF23 is induced by liver injury. In response to acute hepatotoxic liver injury and chronic diet-induced fatty liver or cholestatic injury, FGF23 hepatic gene expression and protein content and serum iFGF23 increase markedly. Analysis of specific hepatic cell types identified Kupffer cells, the resident liver macrophage, as a source of hepatic FGF23. These findings establish that liver injury increases hepatic and systemic levels of FGF23 as the result of the stimulation of Kupffer cells by inflammatory factors.

Materials and methods

Animals

C57BL/6J mice (#000664; Jackson Laboratory, Bar Harbor, ME) were maintained in static cages with ¼ inch corncob bedding and cotton nestlets, automatic water feeding and unlimited access to a normal chow diet (5% fat; PicoLab[®] Rodent Diet 20 #5053; LabDiet, St. Louis,

MO) under 12 h light/dark cycles. Male and female mice 10–16 weeks of age were treated with 1 ml/kg of carbon tetrachloride (CCl₄) diluted 1:10 in mineral oil (MilliporeSigma, St. Louis, MO), as previously performed [23]. Results were identical in both sexes and combined in all analyses. After weaning male mice 8 weeks of age were fed normal diet or high-fat diet (HFD) (60% fat; Research Diets D12492; New Brunswick, NJ) for 16 weeks [24]. Male and female extracellular signal-regulated kinase 1/2 (*Erk1/2*) knockout mice were generated and oil- or tamoxifen-treated, as previously described [25]. Male C57BL/6J mice aged 10–14 weeks were administered the following dissolved in phosphate-buffered saline: intraperitoneal injections of 7.5 mg/kg of LPS (*E. coli* 0111:B4; MilliporeSigma) [26]; intravenous injections of 0.25 ug of TNF, 2.5 ug of IL-1 β and/or 0.25 ug of IFN γ (PeproTech, Rocky Hill, NJ) [17,18]; and intravenous injections of 25 ug of Pam2CSK4 (InvivoGen, San Diego, CA). Mice were placed under deep anesthesia by isoflurane anesthesia prior to blood drawing and sacrifice by cervical dislocation. All animal studies were approved by the Animal Care and Use Committee of the Emory University School of Medicine (Protocol #PROTO201800139 and #PROTO201800146) and followed National Institutes of Health guidelines for animal care.

Quantitative real-time reverse transcription polymerase chain reaction (qRT-PCR)

RNA extraction from whole liver or cultured cells, reverse transcription, and qRT-PCR using the primers (Integrated DNA Technologies, Coralville, IA) in Table 1 were performed as previously described [27]. Data were analyzed by the $2^{-\Delta\Delta CT}$ method for relative quantification and normalized to glyceraldehyde 3-phosphate dehydrogenase.

Fgf23 knockdown in JS1 cells

A stable lentiviral knockdown of *Fgf23* was established in the cell line JS1 (kindly provided by Scott L. Friedman, Icahn School of Medicine at Mount Sinai, New York, NY) [28]. A plasmid encoding for an shRNA to *Fgf23* (Cat. #TRCN0000372259; sequence: CCGGTCCCTCAGAGCC

Table 1. PCR primer sequences.

Gene	Forward primers (5'-3')	Reverse primers (5'-3')
<i>Ccl2</i>	CAGCCAGATGCAGTTAACGCCCA	TGGGGTCAGCACAGACCTCTCTC
<i>Cox2</i>	TGCACTATGGTTACAAAAGCTGG	TCAGGAAGCTCCTTATTTCCCTT
<i>Crp</i>	TTCCAAGGAGTCAGATACTTCC	TCAGAGCAGTGTAGAAATGGAGA
<i>Fgb</i>	CACCTGCCTCATCTTGAGCG	GCATTGACTCTGATGTCTCTCCA
<i>Fgf23</i>	CACTGCTAGAGCCTATCC	CACTGTAGATGGTCTGATGG
<i>Gapdh</i>	AGGTCGGTGTGAACGGATTG	TGTAGACCATGTAGTTGAGGTCA
<i>Fgfr1</i>	ACTCTGGGCTGGTTGAAAAAT	GGTGGCATAGCGAACCTTGTA
<i>Fgfr2</i>	GCTATAAGGTACGAAACCAGCAC	GGTTGATGGACCCGTATTCATTC
<i>Fgfr3</i>	GCCTGCGTGCTAGTGTCT	TACCATCCTTAGCCAGACCG
<i>Fgfr4</i>	TGAAGAGTACCTTGACCTCCG	TCATGTCTGCTGCGAGTCAG
<i>Il1b</i>	GCAACTGTTCTGAACTCAACT	ATCTTTTGGGGTCCGTCAACT
<i>Il6</i>	CACATGTTCTCTGGAAATCGTGGA	TCTCTCTGAAGACTCTGGCTTTGT
<i>Ifng</i>	ATGAACGCTACACACTGCATC	CCATCCTTTTGCCAGTTCCTC
<i>Kl</i>	TGTATGTGACAGCAATGGAATCG	GAATACGCAAAGTAGCCACAAAGG
<i>Klb</i>	ACGACCCGACGAGGGCTGTT	GGAGGAGACCGTAAACTCGGGCTTA
<i>Nos2</i>	GTTCTCAGCCCAACAATACAAGA	GTGGACGGGTCGATGTCAC
<i>Tnf</i>	CCCTCACACTCAGATCATCTTCT	GCTACGACGTGGGCTACAG

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TATCCCAATGCTCGAGCATTGGGATAGGCTCTGAGGATTTTGG) was purchased from MilliporeSigma. To generate lentiviral particles, HEK293T cells were transfected with empty vector or the shRNA target plasmid, packaging plasmids (pRSV-Rev, pMDLg/pRRE) and envelope plasmid (pMD2.VSVG) using Fugene[®]-HD transfection reagent (Promega, Madison, WI). Viral particles were isolated and JS1 cells infected with concentrated lentiviral particles in the presence of polybrene. Stably infected polyclonal cells were selected in puromycin dihydrochloride (10 µg/ml) and maintained in puromycin (2 µg/ml) for experiments.

Protein isolation and western blotting

Total liver or cell protein was isolated for western blotting, as previously described [29,30]. The antibodies employed were FGF23 (#MBS854462; MyBioSource, San Diego, CA) and tubulin (#2148; Cell Signaling, Danvers, MA).

FGF23 ELISA

The concentration of iFGF23 in serum and cell culture supernatants was detected by ELISA (Immutopics Inc, San Clemente, CA) following the manufacturer's instructions.

Primary hepatic cell isolation

Primary Kupffer cells and hepatocytes were isolated from perfusions of mouse livers with Liberase (Roche, Indianapolis, IN), and cultured as previously described [24,26]. Hepatic stellate cells were isolated from mice by sequential perfusion with pronase (Roche) and collagenase (Crescent Chemicals, Islandia, NY). The liver was manually digested, diluted in DMEM-F12 containing 10 µg/mL DNase, and then mixed with an equal volume of DMEM-F12 containing pronase (0.2 mg/mL) and collagenase (0.5 mg/mL). The mixture was incubated in a shaking water bath at 250 rpm, 37°C for 10 min, strained through sterile gauze, washed with Gey's Balanced Salt Solution (GBSS) containing 10 µg/mL DNase and pelleted. The pellet was resuspended in GBSS with 10 µg/mL DNase and layered atop a 9–10% Accudenz (Accurate Chemical Co., Tempe, AZ) gradient. After centrifugation at 1,400 g for 17 minutes, and hepatic stellate cells recovered at the GBSS-Accudenz interface. Cultured cells were treated with 10 ng/ml LPS, 25 ng/ml TNF, 25 ng/ml IL-1β, 100 ng/ml IFNγ (R&D Systems, Minneapolis, MN), 100 ng/ml Pam4CSK2, or 100 ng/ml FGF23 (R&D Systems) with 10 µg/ml heparin (MilliporeSigma).

Statistical analysis

Numerical results are reported as means ± S.E. from three or more independent experiments. Statistical significance among control and treated groups was determined by one way analysis of variance (ANOVA) with Tukey post hoc corrections. Statistical significance was defined as $P < 0.05$.

Results

Hepatic FGF23 expression is induced by toxic liver injury from CCl₄

To identify pathophysiological events that trigger hepatic FGF23 expression in the liver, *Fgf23* mRNA expression was assessed by qRT-PCR during acute mouse hepatic injury from the hepatotoxin CCl₄. Levels of *Fgf23* mRNA expression were very low in normal liver but increased significantly within 12 h after CCl₄ administration (Fig 1A). The increase in *Fgf23* gene expression was sustained throughout the period of maximal liver injury (36–48 h), and levels returned towards normal during the time of histological recovery (60–72 h) (Fig 1A).

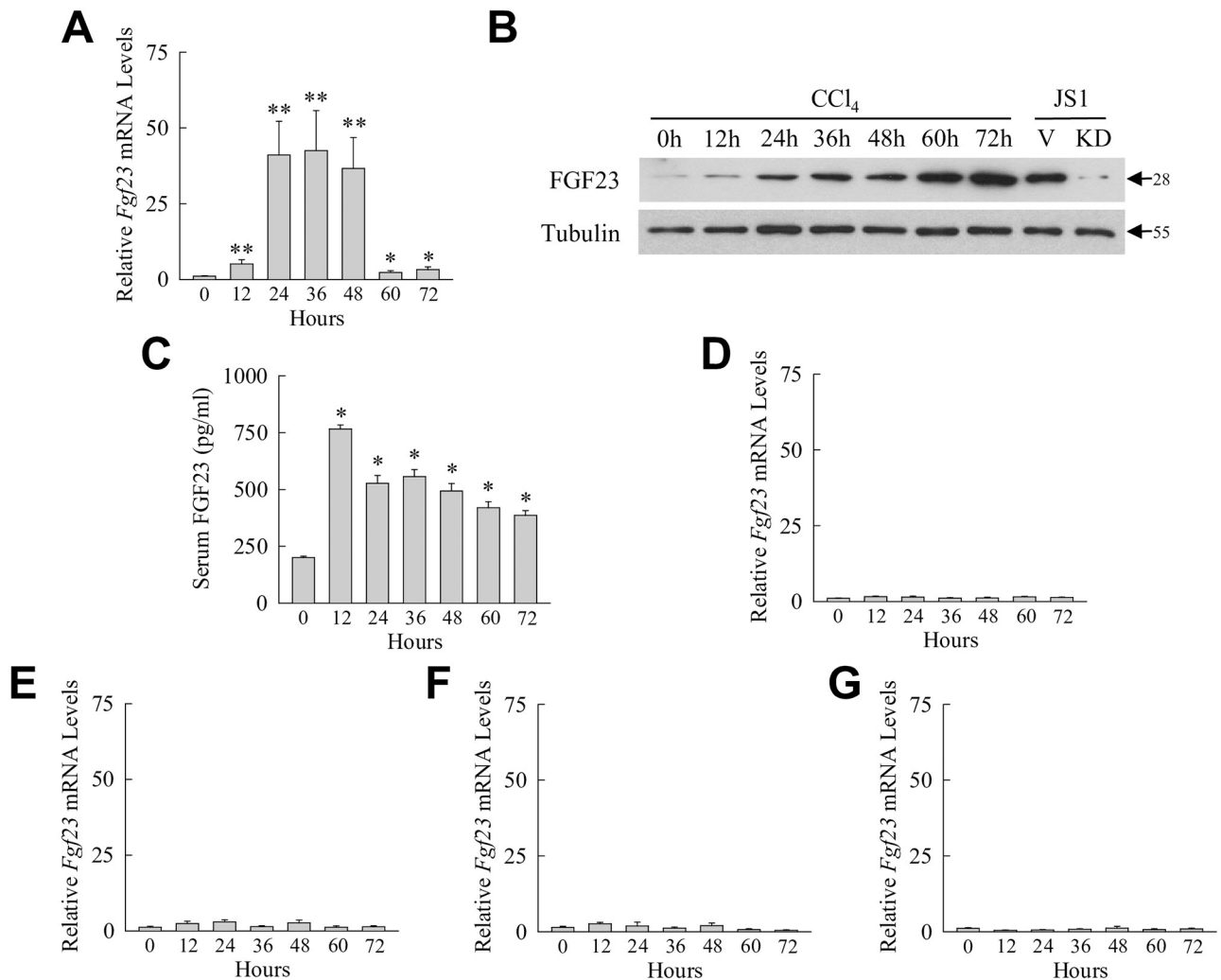


Fig 1. CCl₄-induced acute liver injury results in induction of hepatic and serum FGF23. (A) Liver *Fgf23* mRNA levels in mice untreated or treated with CCl₄ for the indicated number of hours (* $P < 0.05$, ** $P < 0.003$ compared to untreated mice; $n = 10-15$). (B) Immunoblots of total liver protein from the same mice and total protein from JS1 cells infected with empty vector (V) or an shRNA to *Fgf23* (KD). Arrows indicate molecular weights in kilodaltons. Images are representative of 4 independent experiments. (C) Serum levels of iFGF23 after the indicated hours of CCl₄ administration (* $P < 0.00001$ compared to untreated mice; $n = 6$). (D-G) *Fgf23* mRNA levels in bone marrow (D), kidney (E), lung (F), and spleen (G) at the indicated times after CCl₄ administration ($n = 8-9$).

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Fgf23 gene induction by CCl₄ resulted in increased hepatic FGF23 protein content. By immunoblotting, FGF23 was detected in low levels in control livers and increased markedly over time after CCl₄ treatment (Fig 1B). FGF23 antibody specificity was confirmed by the demonstration of an identical size band in the FGF23-producing cell line JS1 which was decreased in cells with a lentiviral knockdown of *Fgf23* (Fig 1B).

To determine whether systemic FGF23 levels are affected by hepatic *Fgf23* induction during CCl₄-induced liver injury, levels of iFGF23 were measured in mouse serum by ELISA. FGF23 was detectable in the serum of control mice and increased as much as 3.8-fold over 12–72 h after CCl₄ administration (Fig 1C). Hepatotoxic liver injury resulted in hepatic FGF23 production that could have autocrine and paracrine liver effects as well as a systemic increase in circulating iFGF23 that could have additional physiological effects on extra-hepatic organs.

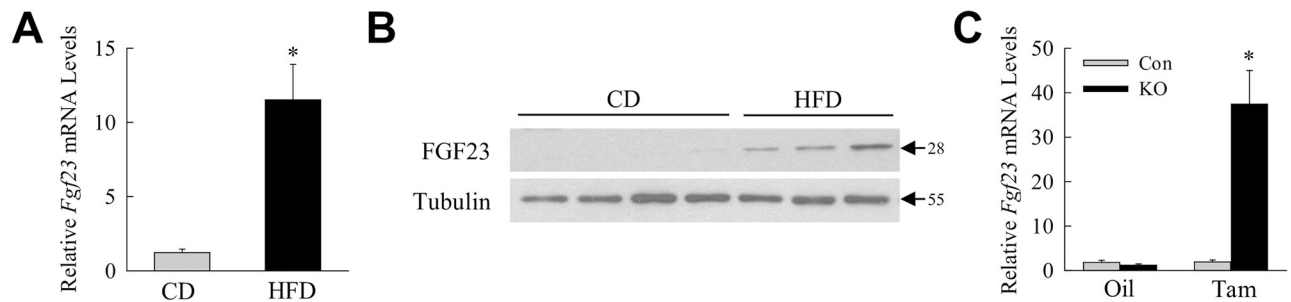


Fig 2. FGF23 expression is increased in chronic diet-induced fatty liver and in cholestatic liver disease. (A) *Fgf23* mRNA levels in the livers of chow diet (CD) and high fat diet (HFD) fed mice (* $P < 0.002$ compared to chow diet; $n = 7$). (B) Immunoblots of total liver protein from the same mice probed for FGF23 and tubulin. Molecular weights in kilodaltons are indicated by arrows. Images are representative of 3 independent experiments. (C) *Fgf23* mRNA levels in littermate control (Con) and *Erk1/2-knockout* (KO) mice treated with oil or tamoxifen (Tam) ($P < 0.0003$ compared to all other groups; $n = 11-13$).

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Increased serum FGF23 from CCl_4 treatment may have resulted from *Fgf23* induction in organs other than liver. To examine for this possibility, *Fgf23* mRNA levels were determined in other tissues. In bone marrow, the site of the primary cell source of FGF23 production [31], *Fgf23* mRNA levels were unchanged after CCl_4 administration (Fig 1D). Kidney, which is also a source of FGF23 [32–34], and a target of CCl_4 toxicity [35], similarly had no significant increase in *Fgf23* mRNA (Fig 1E). Induction of *Fgf23* mRNA also failed to occur in the lung (Fig 1F), another tissue affected by the toxic effects of CCl_4 [35], and the spleen, a reticular endothelium organ like the liver (Fig 1G). These findings indicate that injured liver is the sole source of the sustained elevation in serum iFGF23 during CCl_4 -induced hepatic injury.

Chronic fatty and cholestatic liver injury are associated with increased hepatic FGF23

To determine whether hepatic FGF23 production is a general response to liver injury, FGF23 levels were examined in two murine models of chronic liver injury distinct from that of acute CCl_4 administration. In an HFD-induced mouse model of NAFLD, *Fgf23* mRNA expression was induced greater than 10-fold in the livers of HFD-fed mice (Fig 2A). Elevated levels of FGF23 protein were also detected in the livers of HFD-fed mice by immunoblotting (Fig 2B). *Fgf23* expression was increased 37-fold in the livers of tamoxifen-injected *Erk1/2-knockout* mice that develop chronic bile acid-induced cholestatic liver injury and fibrosis (Fig 2C) [25]. Hepatic FGF23 production is induced in the liver in response to diverse injurious stimuli and in both acute and chronic liver injury.

LPS induces sustained hepatic *Fgf23* expression

Most forms of liver injury, including that from CCl_4 and HFD feeding, are dependent on hepatic effects of gut-derived bacterial products including LPS [16,36]. This fact, together with reports in nonhepatic cell types that *Fgf23* can be induced by LPS and other proinflammatory factors [11], suggested that LPS and/or its downstream effector cytokines may stimulate hepatic *Fgf23* expression. The effect of systemic LPS on *Fgf23* expression in the liver was therefore determined and compared to other organs. Liver *Fgf23* levels rose significantly by 2 h, peaked within 6 h after LPS administration and remained markedly elevated over 24 h (Fig 3A). In bone marrow there was an increase over 2–6 h with a reduction by 12 h and a return to baseline expression at 24 h (Fig 3B). Kidney demonstrated a rapid 2 h peak in expression and

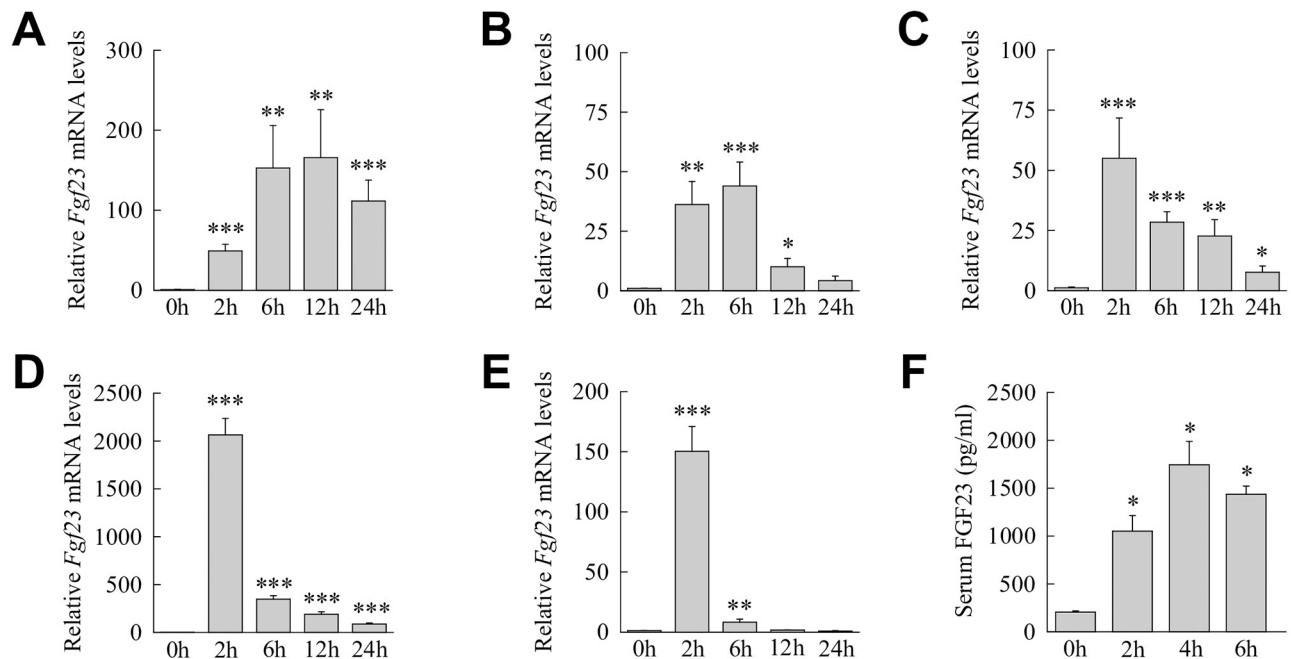


Fig 3. *Fgf23* is induced in the liver and other tissues in response to LPS. (A-E) *Fgf23* mRNA levels in untreated mice and mice treated for the indicated hours with LPS in liver (A), bone marrow (B), kidney (C), lung (D) and spleen (E) (* $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ compared to untreated mice; $n = 8-11$). (F) Serum iFGF23 levels in the mice (* $P < 0.001$ compared to untreated mice; $n = 5$).

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then *Fgf23* expression decreased although levels remained significantly elevated for 24 h (Fig 3C). Lung and spleen had similarly marked *Fgf23* induction at 2 h and levels that decreased dramatically but remained elevated at 6 h (Fig 3D and 3E). LPS led to FGF23 protein production as serum levels of iFGF23 were significantly increased after LPS administration (Fig 3F). Thus, the liver is one of multiple organs in which *Fgf23* induction occurs in response to the inflammatory stimulus of LPS and one in which expression remains sustained at high levels relative to other organs.

Liver *Fgf23* is induced by proinflammatory factors

To identify the downstream mediators of *Fgf23* induction by LPS-initiated toll-like receptor (TLR) 4 signaling, the ability of LPS-inducible proinflammatory cytokines to increase *Fgf23* expression in the liver was examined. Mice were injected with cytokine concentrations previously determined to be biologically active in the liver [17,18]. Administration of either of the two principal LPS-inducible cytokines, TNF and IL-1 β , failed individually to increase liver *Fgf23* mRNA expression (S1 Fig). However, combined treatment with TNF and IL-1 β led to a marked induction of hepatic *Fgf23* gene expression (Fig 4A). In contrast, the LPS-inducible proinflammatory cytokine IFN γ failed to significantly affect *Fgf23* levels (Fig 4B).

Induction of *Fgf23* by TLR4 may be selective for this pattern recognition receptor or other TLRs may stimulate *Fgf23* expression during liver injury. In addition to the involvement of TLR4 in liver disease, TLR2 signaling contributes to the pathogenesis of liver injury [37]. Administration of the TLR2 agonist Pam2CSK4 induced liver *Fgf23* gene expression (Fig 4C). Hepatic *Fgf23* expression is therefore under the regulation of redundant proinflammatory TLR signaling pathways that can activate this gene during liver injury.

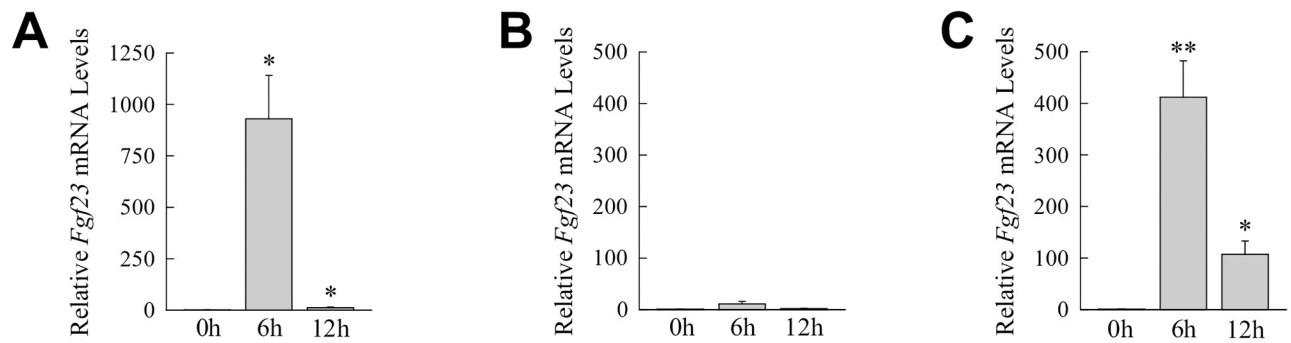


Fig 4. Liver *Fgf23* is induced by TNF/IL-1 β . Hepatic *Fgf23* mRNA levels in untreated mice and mice 6 h and 12 h after injection with TNF/IL-1 β (A), IFN γ (B) or Pam2CSK4 (C) (* P <0.01, ** P <0.0001 compared to untreated mice; n = 5–9).

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Kupffer cells but not hepatocytes or hepatic stellate cells express FGF23

To define the hepatic cell type(s) that produces FGF23 during liver injury, the three principal cell types involved in liver injury, hepatocytes, Kupffer cells and hepatic stellate cells, were isolated and examined *in vitro* for *Fgf23* mRNA baseline expression and induction by LPS. All three hepatic cell types had very low constitutive *Fgf23* mRNA levels. In mouse hepatocytes LPS induced only a late and modest 2.5-fold increase in *Fgf23* mRNA levels at 24 h (Fig 5A). In contrast, Kupffer cells had a sustained LPS induction of *Fgf23* over 24 h that peaked at 75-fold

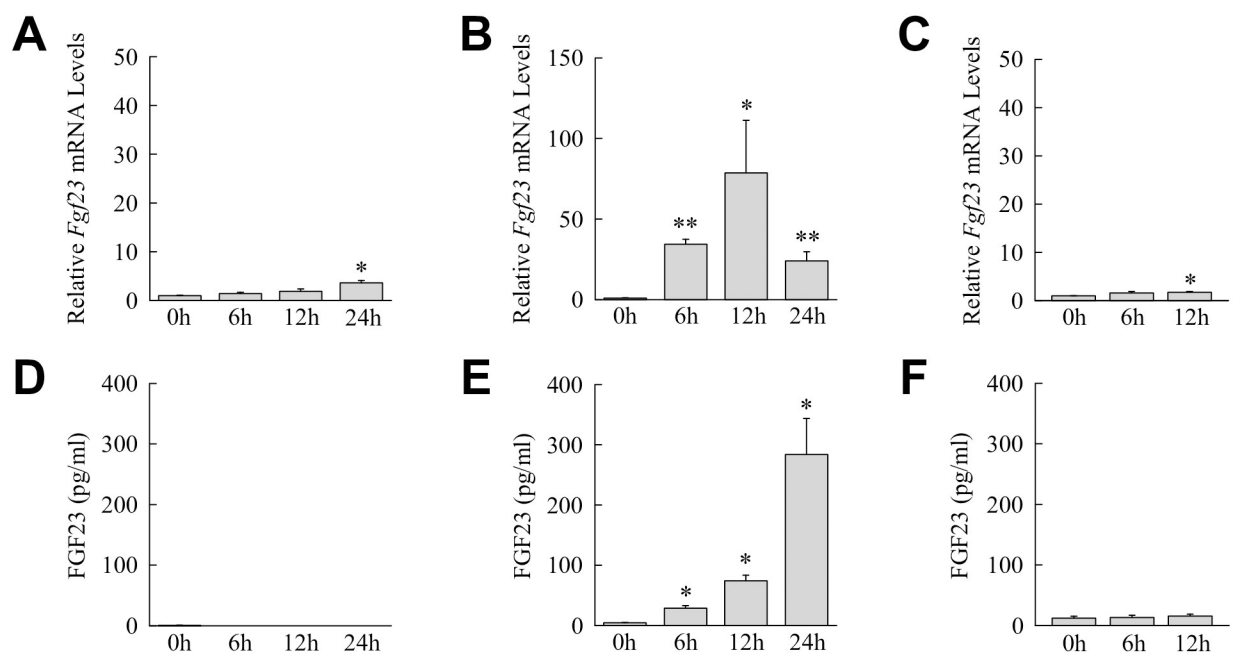


Fig 5. Kupffer cells are a source of hepatic FGF23. (A) Relative *Fgf23* mRNA levels in cultured mouse hepatocytes untreated or treated with LPS for the indicated hours (* P <0.0001 compared to untreated cells; n = 12). (B) *Fgf23* mRNA levels in LPS-treated Kupffer cells (* P <0.01, ** P <0.0001 compared to untreated cells; n = 9). (C) *Fgf23* mRNA content in LPS-treated hepatic stellate cells (* P <0.01 compared to untreated cells; n = 5–6). (D) iFGF23 protein levels by ELISA in the culture medium of mouse hepatocytes after LPS treatment (n = 4–6). (E) iFGF23 protein in the medium of Kupffer cells (* P <0.001 compared to untreated cells; n = 8–9). (F) iFGF23 levels in hepatic stellate cell medium (n = 5).

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at 12 h (Fig 5B). Primary hepatic stellate cells had a minor 2-fold increase over their low basal levels of *Fgf23* expression at 12 h after LPS treatment (Fig 5C). Significant LPS induction of *Fgf23* gene expression is therefore confined to Kupffer cells among these three hepatic cell types.

To further validate that Kupffer cells are a source of liver FGF23, medium from the three cell types untreated and LPS-treated was assayed for levels of iFGF23 by ELISA. No iFGF23 was detected in the medium of hepatocytes untreated or LPS-stimulated (Fig 5D). Kupffer cells constitutively secreted low amounts of iFGF23 that increased 100-fold with LPS treatment (Fig 5E). Primary hepatic stellate cells produced low amounts of iFGF23 that were unaffected by LPS stimulation (Fig 5F). These low levels of iFGF23 production in hepatic stellate cells are likely secondary to the Kupffer cell contamination present in these cultures.

Kupffer cells were further examined for their ability to mimic the hepatic induction of *Fgf23* by proinflammatory cytokines and TLR2 stimulation. Cultured cells had a significant induction in *Fgf23* mRNA levels (Fig 6A) and iFGF23 protein production (Fig 6B) with TNF/IL-1 β treatment. Similarly, mRNA and protein induction occurred in response to the TLR2 agonist Pam2CSK4 (Fig 6C and 6D). Identical to findings in mouse liver, *Fgf23* was not induced by IFN γ in Kupffer cells (Fig 6E and 6F). Kupffer cells therefore mimic the FGF23 induction pattern of mouse liver to inflammatory factors providing further evidence that these cells are a source of FGF23 during toxic liver injury.

To determine whether TLR4/TLR2 induction of *Fgf23* was specific to Kupffer cells or a property of all macrophages, *Fgf23* expression in response to TLR stimulation was examined in bone marrow-derived macrophages. *Fgf23* mRNA levels increased in bone marrow-derived

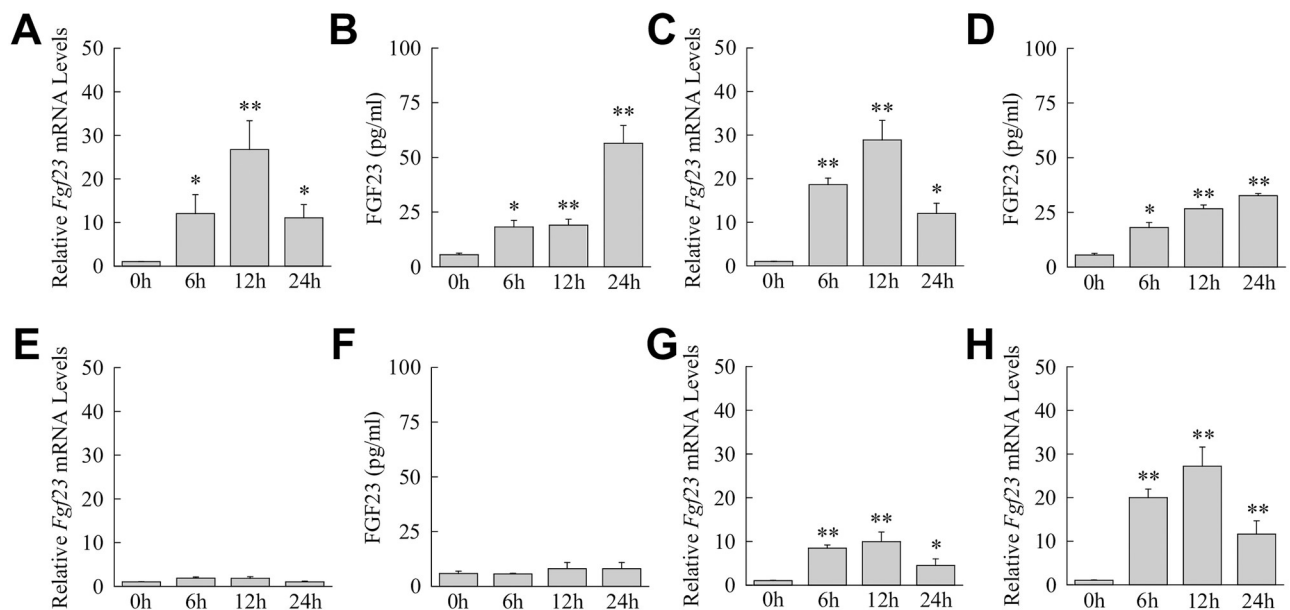


Fig 6. *Fgf23* is induced in Kupffer cells and bone marrow-derived macrophages in response to proinflammatory factors. (A) Relative *Fgf23* mRNA levels in Kupffer cells untreated or treated with TNF/IL-1 β for the indicated hours (* P <0.01, ** P <0.001 compared to untreated cells; n = 9–10). (B) iFGF23 levels in the medium of the same TNF/IL-1 β -treated cells (* P <0.01, ** P <0.001 compared to untreated cells; n = 6–7). (C) *Fgf23* mRNA levels in Kupffer cells treated with Pam2CSK4 (* P <0.001, ** P <0.0001 compared to untreated cells; n = 8–9). (D) Medium protein levels from the same Pam2CSK4-treated cells (* P <0.01, ** P <0.000001 compared to untreated cells; n = 6). (E) *Fgf23* mRNA levels with IFN γ treatment (n = 5–7). (F) iFGF23 levels in the medium with IFN γ treatment (n = 5). (G–H) *Fgf23* mRNA levels in bone marrow-derived macrophages treated with TNF/IL-1 β (G) or Pam2CSK4 (H) (* P <0.02, ** P <0.001 compared to untreated cells; n = 8–10).

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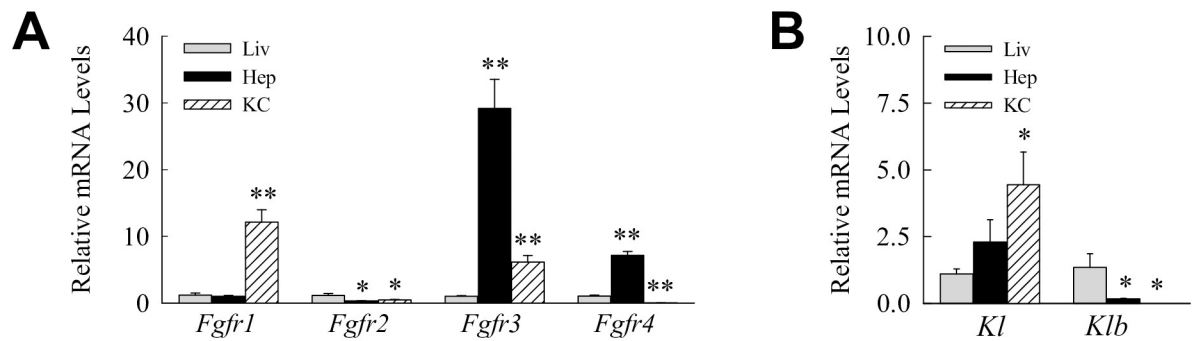


Fig 7. Relative levels of *Fgfr* and *Klotho* gene expression. (A) mRNA levels for the *Fgfr* genes in total liver (Liv), and isolated hepatocytes (Hep) and Kupffer cells (KC) (* $P < 0.03$, ** $P < 0.001$ compared to liver; $n = 6-7$). (B) Expression levels for α - (*Kl*) and β -Klotho (*Klb*) in the same samples (* $P < 0.03$ compared to liver; $n = 6-7$).

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macrophages in response to TNF/IL-1 β (Fig 6G) and Pam2CSK4 (Fig 6H). TLR4/2-dependent induction of *Fgf23* is therefore common to macrophages and not Kupffer cell specific.

FGF23 regulates the proinflammatory phenotype of hepatocytes but not Kupffer cells

Cellular responsiveness to FGF23 is mediated by the family of FGF receptors (FGFRs) and a coreceptor Klotho [31]. Of the four FGFRs, FGFR1, FGFR3 and FGFR4 have been implicated in the transduction of FGF23 signaling in different cell types [38,39]. In some tissues signal initiation requires α - or β -Klotho as a coreceptor [40]. To define potential FGF23 target cells in the liver, the level of gene expression of these receptors/coreceptors was examined in hepatocytes and Kupffer cells relative to whole liver. Differential receptor expression exists in hepatocytes and Kupffer cells. Mouse hepatocytes have higher expression of *Fgfr3* and to a lesser extent of *Fgfr4* with low amounts of *Fgfr2* (Fig 7A). In contrast, Kupffer cells express higher levels of *Fgfr1*, lower levels of *Fgfr3*, and no *Fgfr4* (Fig 7A). α -Klotho mRNA is barely detectable in liver and levels are modestly higher in Kupffer cells (Fig 7B). β -Klotho expression is similarly very low in liver as well as in both hepatocytes and Kupffer cells (Fig 7B).

The induction of hepatic FGF23 by inflammatory mediators, the ability of this factor to regulate inflammation in nonhepatic immune cells, and the presence of FGFRs in Kupffer cells suggested that FGF23 may have an autocrine effect on the inflammatory state of these cells. FGF23 treatment had no effect on basal Kupffer cell expression of the proinflammatory cytokine genes *Tnf*, *Il1b*, *Il6* and *Ifng* (Fig 8A). FGF23 also failed to alter the induction of these genes by LPS (Fig 8A). FGF23 similarly failed to affect expression of chemokine (C-C motif) ligand 2 (*Ccl2*) (Fig 7B), or the proinflammatory genes nitric oxide synthase (*Nos2*) and cyclooxygenase-2 (*Cox2*) (Fig 8C). FGF23 had no effect on basal or LPS-stimulated inflammatory Kupffer cell activation.

Although Kupffer cells are the main cellular regulator of the hepatic immune response, hepatocytes also contribute to liver inflammation [41]. FGF23 has been reported to induce IL-6, a cytokine with mixed proinflammatory and anti-inflammatory hepatic effects, in hepatocytes [42]. Kupffer cell generated FGF23 may therefore regulate hepatic inflammation through a paracrine hepatocyte effect. FGF23 induced hepatocyte gene expression for the cytokines *Il6* and *Il1b* (Fig 9A), the chemokine *Ccl2* (Fig 9B), and the proinflammatory genes *Nos2* and *Cox2* (Fig 9C). In contrast, FGF23 failed to induce expression for the acute phase reactants *c-*

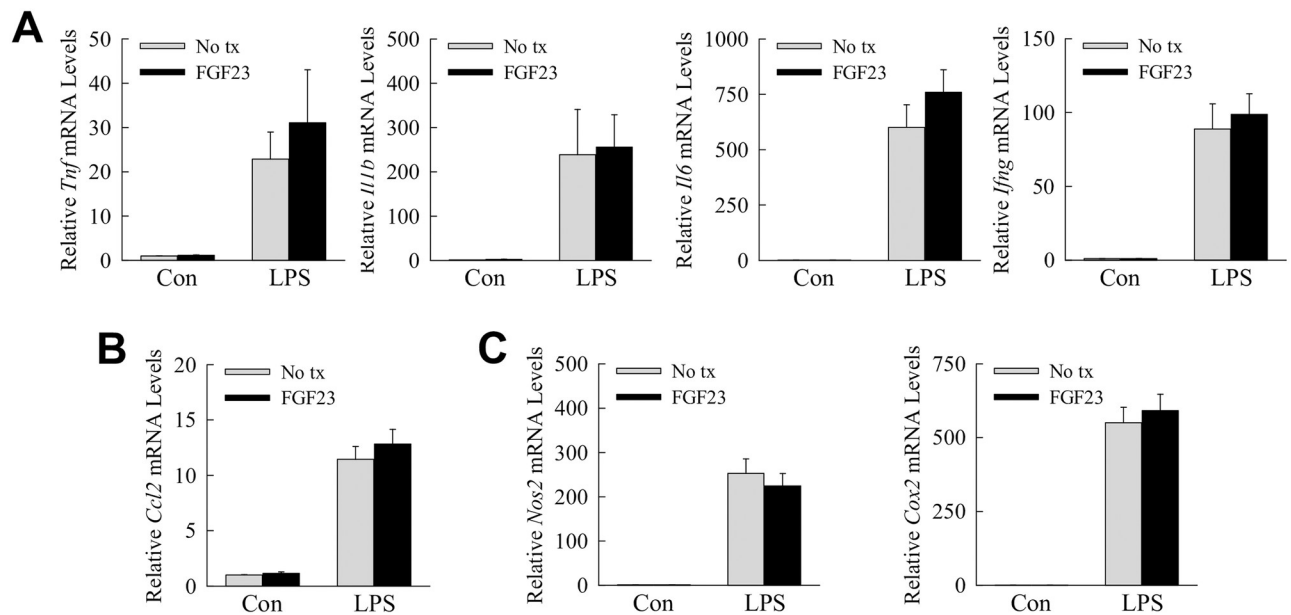


Fig 8. The inflammatory state of Kupffer cells untreated or LPS-treated is unaffected by FGF23. (A-C) Primary mouse Kupffer cells were untreated (No tx) or pre-treated with FGF23, and 1 h later some cells were untreated (Con) or treated with LPS. Cells were harvested 6 h after LPS treatment. Relative mRNA levels for the indicated genes are shown (n = 8–10).

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reactive protein and *fibrinogen* (Fig 9D). FGF23 therefore induced a specific proinflammatory activation in primary hepatocytes.

Discussion

FGF23 is produced primarily by bone for the function of regulating total body phosphorous homeostasis. There is a long-standing and well-established relationship of diseases of the kidney, the critical site for phosphate excretion, with elevated serum FGF23 levels. However, increases in FGF23 have more recently also been linked to diseases and their outcomes in a number of organs outside the bone-kidney axis of mineral regulation including liver, lung and heart [6–8,10,43–45]. Whether these organs are a source of FGF23 and/or are responsive to this hormone are unclear. This study demonstrates that acute and chronic liver injury induces hepatic production of FGF23 through the stimulation of the resident liver macrophages or Kupffer cells by injury-associated inflammatory factors.

Our findings demonstrate that the liver produces FGF23 in response to the diverse injuries of a hepatotoxin CCl₄, diet-induced fatty liver, and bile acid-induced cholestatic liver disease. Hepatic FGF23 was generated rapidly by acute liver injury from CCl₄, and its production was sustained in chronic NAFLD and cholestatic injury as demonstrated by both increased gene expression and protein content in the liver. FGF23 generation by acute liver injury from CCl₄ was sufficient to elevate serum iFGF23 to levels comparable to that achieved by systemic LPS administration. Findings that *Fgf23* induction from CCl₄ occurred only in the liver, and not in other tissues known to produce FGF23, indicate that the injured liver accounted for the increase in serum iFGF23 and can be a major source of excessive systemic FGF23. Increased serum FGF23 has been reported in human NAFLD [7], and in patients with chronic liver disease awaiting liver transplant [6]. Whether liver was the source of FGF23 in these conditions

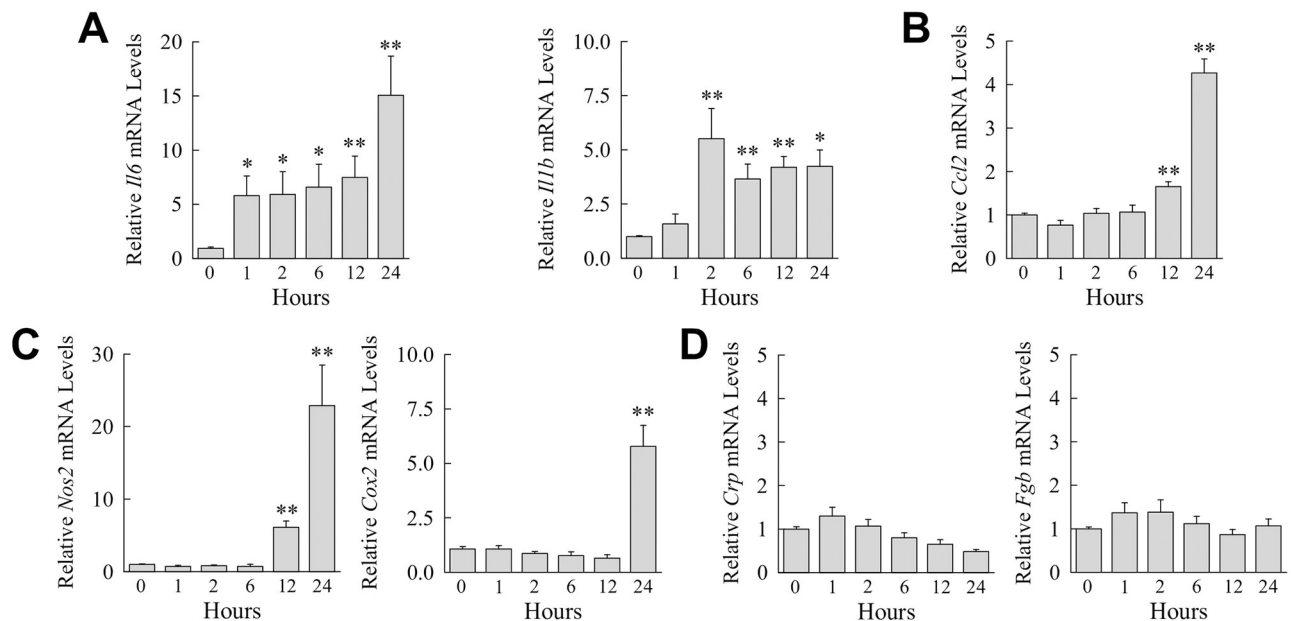


Fig 9. FGF23 induces a proinflammatory hepatocyte phenotype. (A–C) Primary mouse hepatocytes were untreated or administered FGF23 for the number of hours shown. Relative mRNA levels for the indicated genes including *c-reactive protein (Crp)* and *fibrinogen (Fgb)* are indicated (* $P < 0.05$, ** $P < 0.01$ compared to untreated cells; $n = 6$).

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was not examined, but our mouse studies suggest that the liver itself could be the tissue responsible for FGF23 production in human liver diseases.

Critical to the outcome from liver injury is the extent of the associated hepatic innate immune response stimulated by intestinal bacterial products including LPS and regulated by cytokines. In contrast to several prior findings of increased serum cFGF23 but not iFGF23 during endotoxemia or bacteremia [19,21], our study demonstrates that systemic LPS administration increased serum iFGF23 levels consistent with the findings of Masuda et al. [13]. The results demonstrate that the liver is a source of FGF23 during endotoxemia whereas others have concluded that principally spleen, and to a lesser extent other tissues including bone and thymus, are the organs that produce FGF23 in response to circulating LPS [13,19,21]. Interestingly, liver was the one organ with sustained *Fgf23* induction for 24 h after LPS administration whereas mRNA levels in all other tissues dropped precipitously within 6–12 h. Liver may therefore be a major source of FGF23 during systemic inflammation. *Fgf23* induction occurred in nonhepatic tissues with LPS but not CCl_4 injection even though CCl_4 -induced liver injury is LPS dependent [16]. This difference could reflect the fact that with CCl_4 LPS delivery is restricted largely to the liver as the source is endogenous intestinal LPS that travels by the portal circulation directly to the liver in contrast to the systemic uptake from an intraperitoneal injection of exogenous LPS. Hepatic FGF23 expression is induced by TLR2 as well as LPS-induced TLR4 stimulation and by the proinflammatory cytokines TNF/IL1- β which may be the downstream effectors of LPS/TLR4 signaling. The fact that multiple inducers of hepatic FGF23 are proinflammatory stimuli clearly link FGF23 to the liver inflammatory response in keeping with recent evidence that this hormone may regulate inflammation.

The principal cell source of FGF23 is the bone osteoblast consistent with this protein's known involvement with phosphorous homeostasis [31]. FGF23 expression has also been demonstrated in normal brain [46], and in renal tubule cells in a variety of forms of kidney

injury [32–34]. Both the absence and presence of FGF23 expression has been reported in vascular smooth muscle cells [34,47]. Macrophages and dendritic cells have also been demonstrated to produce FGF23 [12,13]. Our findings extend the known cellular sources of FGF23 to hepatic Kupffer cells further strengthening the relationship between inflammation and this hormone.

Similar to findings with renal diseases, increases in systemic FGF23 have been linked to a poor prognosis in liver disease. Increased human FGF23 levels have been shown to predict mortality in patients waiting for liver transplant [6], and are associated with progression of hepatocellular carcinoma [48]. How FGF23 may modulate liver disease and its outcome are unclear. Autocrine/paracrine actions of FGF23 in the liver are possible as the studies demonstrate differential *Fgfr* isoform expression in hepatocytes and Kupffer cells. Given the growing association of FGF23 with inflammation, and the presence of FGFRs in these two hepatic cell types, a function for FGF23 in the regulation of the inflammatory state of these cells was examined. Although proinflammatory activation of the macrophage cell line RAW264.7 by FGF23 has been reported [12], no pro- or anti-inflammatory effect of FGF23 was detected in Kupffer cells. Hepatic FGF23 does not function in an autocrine fashion to modulate the hepatic macrophage inflammatory response. Prior studies have indicated that FGF23 induces c-reactive protein and IL-6, a cytokine with pro- and anti-inflammatory functions in the liver, in cultured hepatocytes [42]. Our findings in hepatocytes indicate that FGF23 causes a proinflammatory activation of these cells with an induction of cytokines and chemokines, but no increase in acute phase reactants such as c-reactive protein was detected. One function of FGF23 in liver injury may therefore be the promotion of the hepatocyte inflammatory response. A determination of additional effects of liver-produced FGF23 during liver injury will require further study.

Hepatic diseases are often linked to effects in other organ but the mechanisms of crosstalk between the injured liver and other tissues are largely unknown. A prime example of the effects of injury in the liver on other organs is the association of NAFLD with cardiovascular and renal disease [22]. We have demonstrated *Fgf23* induction in a mouse model of NAFLD and increased serum FGF23 levels have been found in human NAFLD [7,43]. Liver-produced FGF23 may therefore have endocrine effects in these extrahepatic diseases. Patients with NAFLD have an increased prevalence of cardiovascular disease which is the major cause of their increased mortality [49]. Increased FGF23 levels have been linked to cardiovascular disease and mortality [45]. Hepatic FGF23 generated in the setting of a fatty liver may therefore be a mechanism of cardiovascular disease in NAFLD. Our findings of hepatic FGF23 production therefore suggest a novel mechanism by which injury in the liver may adversely affect other organs. In addition, the promotion of inflammation in nonhepatic organs by hepatic FGF23 may result in the release of other proinflammatory factors from these organs which in turn trigger the progression of liver disease. The interesting possibility of the hormonal effects of hepatic FGF23 mediating the crosstalk between liver and other organs in hepatic diseases requires further study.

The investigations demonstrate that increased serum FGF23 levels do not simply reflect renal dysfunction or phosphate retention but may result from hepatic pathophysiology. Acute or chronic liver injury leads to hepatic production of FGF23 through the effects of proinflammatory factors such as LPS and cytokines on hepatic macrophages. The studies identify FGF23 as a potential new autocrine or paracrine regulator of liver disease. The findings also uncover a new possible hormonal mediator of crosstalk between an injured liver and extrahepatic organs that may be a mechanism of disease in distal tissues or explain the ability of these other organs to promote liver injury.

Supporting information

S1 Fig. TNF and IL-1 β fail to induce hepatic *Fgf23* expression. Hepatic *Fgf23* mRNA levels in untreated mice and mice 6 h and 12 h after injection with TNF (A), or IL-1 β (B) (n = 4–5). (PDF)

S2 Fig. Unaltered immunoblot images. Unaltered radiographic images for Figs 1B and 2B. (PDF)

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References

1. Farrow EG, Yu X, Summers LJ, Davis SI, Fleet JC, Allen MR, et al. Iron deficiency drives an autosomal dominant hypophosphatemic rickets (ADHR) phenotype in fibroblast growth factor-23 (*Fgf23*) knock-in mice. *Proc Natl Acad Sci U S A*. 2011; 108:E1146–E1155. <https://doi.org/10.1073/pnas.1110905108> PMID: 22006328
2. Imel EA, Peacock M, Gray AK, Padgett LR, Hui SL, Econs MJ. Iron modifies plasma FGF23 differently in autosomal dominant hypophosphatemic rickets and healthy humans. *J Clin Endocrinol Metab*. 2011; 96:3541–3549. <https://doi.org/10.1210/jc.2011-1239> PMID: 21880793
3. Fliser D, Kollerits B, Neyer U, Ankerst DP, Lhotta K, Lingenhel A, et al. Fibroblast growth factor 23 (*FGF23*) predicts progression of chronic kidney disease: the Mild to Moderate Kidney Disease (MMKD) Study. *J Am Soc Nephrol*. 2007; 18:2600–2608. <https://doi.org/10.1681/ASN.2006080936> PMID: 17656479
4. Isakova T, Xie H, Yang W, Xie D, Anderson AH, Scialla J, et al. Fibroblast growth factor 23 and risks of mortality and end-stage renal disease in patients with chronic kidney disease. *JAMA*. 2011; 305:2432–2439. <https://doi.org/10.1001/jama.2011.826> PMID: 21673295
5. Gutierrez OM, Mannstadt M, Isakova T, Rauh-Hain JA, Tamez H, Shah A, et al. Fibroblast growth factor 23 and mortality among patients undergoing hemodialysis. *N Engl J Med*. 2008; 359:584–592. <https://doi.org/10.1056/NEJMoa0706130> PMID: 18687639
6. Prie D, Forand A, Francoz C, Elie C, Cohen I, Courbebaisse M, et al. Plasma fibroblast growth factor 23 concentration is increased and predicts mortality in patients on the liver-transplant waiting list. *PLoS One*. 2013; 8:e66182. <https://doi.org/10.1371/journal.pone.0066182> PMID: 23825530
7. He X, Shen Y, Ma X, Ying L, Peng J, Pan X, et al. The association of serum FGF23 and non-alcoholic fatty liver disease is independent of vitamin D in type 2 diabetes patients. *Clin Exp Pharmacol Physiol*. 2018; 45:668–674. <https://doi.org/10.1111/1440-1681.12933> PMID: 29574933
8. Haring R, Enserro D, Xanthakis V, Mitchell GF, Benjamin EJ, Hamburg NM, et al. Plasma fibroblast growth factor 23: clinical correlates and association with cardiovascular disease and mortality in the

- Framingham Heart Study. *J Am Heart Assoc.* 2016; 5:e003486. <https://doi.org/10.1161/JAHA.116.003486> PMID: 27385427
9. Lutsey PL, Alonso A, Selvin E, Pankow JS, Michos ED, Agarwal SK, et al. Fibroblast growth factor-23 and incident coronary heart disease, heart failure, and cardiovascular mortality: the Atherosclerosis Risk in Communities study. *J Am Heart Assoc.* 2014; 3:e000936. <https://doi.org/10.1161/JAHA.114.000936> PMID: 24922628
 10. Gulati S, Wells JM, Urdaneta GP, Balestrini K, Vital I, Tovar K, et al. Fibroblast growth factor 23 is associated with a frequent exacerbator phenotype in COPD: a cross-sectional pilot study. *Int J Mol Sci.* 2019; 20:2292. <https://doi.org/10.3390/ijms20092292> PMID: 31075857
 11. David V, Francis C, Babitt JL. Ironing out the cross talk between FGF23 and inflammation. *Am J Physiol Renal Physiol.* 2017; 312:F1–F8. <https://doi.org/10.1152/ajprenal.00359.2016> PMID: 27582104
 12. Han X, Li L, Yang J, King G, Xiao Z, Quarles LD. Counter-regulatory paracrine actions of FGF-23 and 1,25(OH)₂D in macrophages. *FEBS Lett.* 2016; 590:53–67. <https://doi.org/10.1002/1873-3468.12040> PMID: 26762170
 13. Masuda Y, Ohta H, Morita Y, Nakayama Y, Miyake A, Itoh N, et al. Expression of Fgf23 in activated dendritic cells and macrophages in response to immunological stimuli in mice. *Biol Pharm Bull.* 2015; 38:687–693. <https://doi.org/10.1248/bpb.b14-00276> PMID: 25739891
 14. Brenner C, Galluzzi L, Kepp O, Kroemer G. Decoding cell death signals in liver inflammation. *J Hepatol.* 2013; 59:583–594. <https://doi.org/10.1016/j.jhep.2013.03.033> PMID: 23567086
 15. Zannetti C, Roblot G, Charrier E, Ainouze M, Tout I, Briat F, et al. Characterization of the inflammasome in human Kupffer cells in response to synthetic agonists and pathogens. *J Immunol.* 2016; 197:356–367. <https://doi.org/10.4049/jimmunol.1502301> PMID: 27226092
 16. Czaja MJ, Xu J, Ju Y, Alt E, Schmiedeberg P. Lipopolysaccharide-neutralizing antibody reduces hepatocyte injury from acute hepatotoxin administration. *Hepatology.* 1994; 19:1282–1289. PMID: 8175153
 17. Shen Y, Cingolani F, Malik SA, Wen J, Liu Y, Czaja MJ. Sex-specific regulation of interferon-gamma cytotoxicity in mouse liver by autophagy. *Hepatology.* 2021; 74:2745–2758. <https://doi.org/10.1002/hep.32010> PMID: 34118081
 18. Shen Y, Malik SA, Amir M, Kumar P, Cingolani F, Wen J, et al. Decreased hepatocyte autophagy leads to synergistic IL-1 β and TNF mouse liver injury and inflammation. *Hepatology.* 2020; 72:595–608. <https://doi.org/10.1002/hep.31209> PMID: 32108953
 19. Bansal S, Friedrichs WE, Velagapudi C, Fellers D, Khazim K, Horn D, et al. Spleen contributes significantly to increased circulating levels of fibroblast growth factor 23 in response to lipopolysaccharide-induced inflammation. *Nephrol Dial Transplant.* 2017; 32:960–968. <https://doi.org/10.1093/ndt/gfw376> PMID: 27836924
 20. Bayer J, Vaghela R, Drechsler S, Osuchowski MF, Erben RG, Andrukhova O. The bone is the major source of high circulating intact fibroblast growth factor-23 in acute murine polymicrobial sepsis induced by cecum ligation puncture. *PLoS One.* 2021; 16:e0251317. <https://doi.org/10.1371/journal.pone.0251317> PMID: 33989306
 21. David V, Martin A, Isakova T, Spaulding C, Qi L, Ramirez V, et al. Inflammation and functional iron deficiency regulate fibroblast growth factor 23 production. *Kidney Int.* 2016; 89:135–146. <https://doi.org/10.1038/ki.2015.290> PMID: 26535997
 22. Wijarnpreecha K, Aby ES, Ahmed A, Kim D. Evaluation and management of extrahepatic manifestations of nonalcoholic fatty liver disease. *Clin Mol Hepatol.* 2021; 27:221–235. <https://doi.org/10.3350/cmh.2020.0239> PMID: 33317243
 23. Schmiedeberg P, Biempica L, Czaja MJ. Timing of protooncogene expression varies in toxin-induced liver regeneration. *J Cell Physiol.* 1993; 154:294–300. <https://doi.org/10.1002/jcp.1041540212> PMID: 8425910
 24. Liu K, Zhao E, Ilyas G, Lalazar G, Lin Y, Haseeb M, et al. Impaired macrophage autophagy increases the immune response in obese mice by promoting proinflammatory macrophage polarization. *Autophagy.* 2015; 11:271–284. <https://doi.org/10.1080/15548627.2015.1009787> PMID: 25650776
 25. Cingolani F, Liu Y, Shen Y, Wen J, Farris AB, Czaja MJ. Redundant functions of ERK1 and ERK2 maintain mouse liver homeostasis through down-regulation of bile acid synthesis. 2021 Dec 22. <https://doi.org/10.1002/hep4.1867> Online ahead of print. PMID: 34936222
 26. Lalazar G, Ilyas G, Malik SA, Liu K, Zhao E, Amir M, et al. Autophagy confers resistance to lipopolysaccharide-induced mouse hepatocyte injury. *Am J Physiol Gastrointest Liver Physiol.* 2016; 311:G377–G386. <https://doi.org/10.1152/ajpgi.00124.2016> PMID: 27469366
 27. Fontana L, Zhao E, Amir M, Dong H, Tanaka K, Czaja MJ. Aging promotes the development of diet-induced murine steatohepatitis but not steatosis. *Hepatology.* 2013; 57:995–1004. <https://doi.org/10.1002/hep.26099> PMID: 23081825

28. Guo J, Loke J, Zheng F, Hong F, Yea S, Fukata M, et al. Functional linkage of cirrhosis-predictive single nucleotide polymorphisms of Toll-like receptor 4 to hepatic stellate cell responses. *Hepatology*. 2009; 49:960–968. <https://doi.org/10.1002/hep.22697> PMID: 19085953
29. Schattenberg JM, Wang Y, Rigoli RM, Koop DR, Czaja MJ. CYP2E1 overexpression alters hepatocyte death from menadione and fatty acids by activation of ERK1/2 signaling. *Hepatology*. 2004; 39:444–455. <https://doi.org/10.1002/hep.20067> PMID: 14767997
30. Wang Y, Singh R, Lefkowitz JH, Rigoli RM, Czaja MJ. Tumor necrosis factor-induced toxic liver injury results from JNK2-dependent activation of caspase-8 and the mitochondrial death pathway. *J Biol Chem*. 2006; 281:15258–15267. <https://doi.org/10.1074/jbc.M512953200> PMID: 16571730
31. Vervloet M. Renal and extrarenal effects of fibroblast growth factor 23. *Nat Rev Nephrol*. 2019; 15:109–120. <https://doi.org/10.1038/s41581-018-0087-2> PMID: 30514976
32. Spichtig D, Zhang H, Mohebbi N, Pavik I, Petzold K, Stange G, et al. Renal expression of FGF23 and peripheral resistance to elevated FGF23 in rodent models of polycystic kidney disease. *Kidney Int*. 2014; 85:1340–1350. <https://doi.org/10.1038/ki.2013.526> PMID: 24402093
33. Zanchi C, Locatelli M, Benigni A, Corna D, Tomasoni S, Rottoli D, et al. Renal expression of FGF23 in progressive renal disease of diabetes and the effect of ACE inhibitor. *PLoS One*. 2013; 8:e70775. <https://doi.org/10.1371/journal.pone.0070775> PMID: 23967103
34. Sugiura H, Matsushita A, Futaya M, Teraoka A, Akiyama KI, Usui N, et al. Fibroblast growth factor 23 is upregulated in the kidney in a chronic kidney disease rat model. *PLoS One*. 2018; 13:e0191706. <https://doi.org/10.1371/journal.pone.0191706> PMID: 29518087
35. Abraham P, Wilfred G, Cathrine SP. Oxidative damage to the lipids and proteins of the lungs, testis and kidney of rats during carbon tetrachloride intoxication. *Clin Chim Acta*. 1999; 289:177–179. [https://doi.org/10.1016/s0009-8981\(99\)00140-0](https://doi.org/10.1016/s0009-8981(99)00140-0) PMID: 10636716
36. Carpino G, Del Ben M, Pastori D, Carnevale R, Baratta F, Overi D, et al. Increased liver localization of lipopolysaccharides in human and experimental NAFLD. *Hepatology*. 2020; 72:470–485. <https://doi.org/10.1002/hep.31056> PMID: 31808577
37. Miura K, Yang L, van Rooijen N, Brenner DA, Ohnishi H, Seki E. Toll-like receptor 2 and palmitic acid cooperatively contribute to the development of nonalcoholic steatohepatitis through inflammatory activation in mice. *Hepatology*. 2013; 57:577–589. <https://doi.org/10.1002/hep.26081> PMID: 22987396
38. Gattineni J, Bates C, Twombly K, Dwarakanath V, Robinson ML, Goetz R, et al. FGF23 decreases renal NaPi-2a and NaPi-2c expression and induces hypophosphatemia in vivo predominantly via FGF receptor 1. *Am J Physiol Renal Physiol*. 2009; 297:F282–F291. <https://doi.org/10.1152/ajprenal.90742.2008> PMID: 19515808
39. Gattineni J, Twombly K, Goetz R, Mohammadi M, Baum M. Regulation of serum 1,25(OH)₂ vitamin D₃ levels by fibroblast growth factor 23 is mediated by FGF receptors 3 and 4. *Am J Physiol Renal Physiol*. 2011; 301:F371–F377. <https://doi.org/10.1152/ajprenal.00740.2010> PMID: 21561999
40. Kovesdy CP, Quarles LD. FGF23 from bench to bedside. *Am J Physiol Renal Physiol*. 2016; 310:F1168–F1174. <https://doi.org/10.1152/ajprenal.00606.2015> PMID: 26864938
41. Jia L, Vianna CR, Fukuda M, Berglund ED, Liu C, Tao C, et al. Hepatocyte Toll-like receptor 4 regulates obesity-induced inflammation and insulin resistance. *Nat Commun*. 2014; 5:3878. <https://doi.org/10.1038/ncomms4878> PMID: 24815961
42. Singh S, Grabner A, Yanucil C, Schramm K, Czaya B, Krick S, et al. Fibroblast growth factor 23 directly targets hepatocytes to promote inflammation in chronic kidney disease. *Kidney Int*. 2016; 90:985–996. <https://doi.org/10.1016/j.kint.2016.05.019> PMID: 27457912
43. Hu X, Yang L, Yu W, Pan W, Chen X, Li Q, et al. Association of serum fibroblast growth factor 23 levels with the presence and severity of hepatic steatosis is independent of sleep duration in patients with diabetes. *Diabetes Metab Syndr Obes*. 2020; 13:1171–1178. <https://doi.org/10.2147/DMSO.S241348> PMID: 32368110
44. Krick S, Grabner A, Baumlin N, Yanucil C, Helton S, Grosche A, et al. Fibroblast growth factor 23 and Klotho contribute to airway inflammation. *Eur Respir J*. 2018; 52. <https://doi.org/10.1183/13993003.00236-2018> PMID: 29748308
45. Qin Z, Liu X, Song M, Zhou Q, Yu J, Zhou B, et al. Fibroblast growth factor 23 as a predictor of cardiovascular and all-cause mortality in prospective studies. *Atherosclerosis*. 2017; 261:1–11. <https://doi.org/10.1016/j.atherosclerosis.2017.03.042> PMID: 28411494
46. Yamashita T, Yoshioka M, Itoh N. Identification of a novel fibroblast growth factor, FGF-23, preferentially expressed in the ventrolateral thalamic nucleus of the brain. *Biochem Biophys Res Commun*. 2000; 277:494–498. <https://doi.org/10.1006/bbrc.2000.3696> PMID: 11032749

47. Zhu D, Mackenzie NC, Millan JL, Farquharson C, MacRae VE. A protective role for FGF-23 in local defence against disrupted arterial wall integrity? *Mol Cell Endocrinol.* 2013; 372:1–11. <https://doi.org/10.1016/j.mce.2013.03.008> PMID: 23523568
48. Zou B, Liu X, Gong Y, Cai C, Li P, Xing S, et al. A novel 12-marker panel of cancer-associated fibroblasts involved in progression of hepatocellular carcinoma. *Cancer Manag Res.* 2018; 10:5303–5311. <https://doi.org/10.2147/CMAR.S176152> PMID: 30464627
49. Targher G, Day CP, Bonora E. Risk of cardiovascular disease in patients with nonalcoholic fatty liver disease. *N Engl J Med.* 2010; 363:1341–1350. <https://doi.org/10.1056/NEJMra0912063> PMID: 20879883