UC Irvine UC Irvine Previously Published Works

Title Translating Extranuclear Steroid Receptor Signaling to Clinical Medicine

Permalink https://escholarship.org/uc/item/4jz865wv

Journal Discover Oncology, 5(3)

ISSN 1868-8497

Author Levin, Ellis R

Publication Date 2014-06-01

DOI 10.1007/s12672-014-0179-9

Peer reviewed



NIH Public Access

Author Manuscript

Horm Cancer. Author manuscript; available in PMC 2015 June 01.

Published in final edited form as:

Horm Cancer. 2014 June ; 5(3): 140-145. doi:10.1007/s12672-014-0179-9.

Translating Extra-Nuclear Steroid Receptor Signaling to Clinical Medicine

Ellis R Levin

Division of Endocrinology, Departments of Medicine, University of California, Irvine, Irvine CA, 92717, USA, Department of Veterans Affairs Medical Center, Long Beach, Long Beach, CA, 90822, USA

Abstract

The existence and function of extra-nuclear steroid receptors (SR) to rapidly modulate signal transduction is now acknowledged as present in cells and organs throughout the body. Work over the past 15 years has defined key mechanisms that are required for sex steroid receptors to traffic to the plasma membrane, but mechanisms of localization in other cell organelles such as mitochondria is still unclear. Signaling by membrane-localized SR has now been reported to impact many aspects of adult organ functions, while the roles in organ development are under investigation. In hormone-responsive cancers, both extra-nuclear and nuclear sex steroid receptors appear to collaborate in the regulation of some key genes that promote malignancy. Here I review what is understood about the impact of extra-nuclear steroid receptor signaling to mitigate or promote disease processes.

Introduction

Steroid receptors outside the nucleus have now been identified in many organs and cells including various hormone-responsive malignancies [1]. These receptors transduce multiple rapid signals that impact the functional biology of target tissues. Rapid signaling occurs in concert with steroid hormone action in the nucleus and is sometimes required for regulation of specific gene transcription [2]. However, membrane initiated steroid signaling (MISS) can also affect important functions in a non-genomic fashion. The latter most often occurs from plasma membrane-localized receptors signaling to the post-translational modification of existing proteins such as enzymes [3]. In this way the body can rapidly adapt to environmental or other stresses, impacting fundamental processes for cardiovascular regulation, bone health, reproductive tract functions, and cell survival. Importantly we previously showed that it is the membrane and not the nuclear estrogen receptor (ER) that is responsible for the ability of the sex steroid to activate rapid signal transduction [4,5]. In this review, I provide examples of how MISS affects in-vivo animal models of disease and human functions.

^{*}Correspondence to: Ellis R. Levin M.D., Medical Service (111-I), Long Beach VA Medical Center, 5901 E. 7th St., Long Beach, CA 90822, Tel: 562-826-5748, Fax: 562-926-5515, ellis.levin@va.gov.

Conflicts of Interest

Ellis Levin discloses no apparent conflicts.

Hormone Responsive Cancers

Breast

The breast cancer field has focused on how tyrosine kinase growth factor receptors for insulin, insulin-like growth factor I (IGF1) epidermal growth factor (EGF) and other ErbB receptor family members signal to the pathogenesis and progression of this malignancy [6–8]. Important pathways downstream of these membrane receptors include PI3K-AKT-mTOR and MEK-ERK signaling that contribute through innumerable ways to breast cancer ontogenesis. Such signaling also underlies some forms of resistance to endocrine therapies in ER/progesterone receptor (PR) positive tumors [9,10]. Interestingly the ability of *membrane* ER or PR to signal through the same multiple pathways in breast cancer epithelial cells partially occurs when membrane-localized sex steroid receptors form complexes with the oncogenic tyrosine kinase Src and are subsequently activated by either estrogen or progesterone [11,12]. Rapid signaling trans activates the EGFR, and IGFR1 proteins [13,14], resulting in activation of multiple downstream kinase cascades that are essential to the proliferation and survival of malignant cells.

Recently, Poulard et al. reported that in ER/PR+ human breast cancer tissues, ~55% of 175 samples show ER/PR in complex with Src just under/at the plasma membrane [15]. High amounts of complexes seen in the tissue sections correlated to multiple clinical factors of poor prognosis, including lymph node involvement, HER2 overexpression and higher tumor grade, and decreased disease free survival [15]. Key oncogenes in this malignancy such as cyclin D1 and c-myc [12,16–18] are up-regulated at the mRNA/protein levels by sex steroids in part from various rapid signals that include the wnt pathway (19). Cyclin D1 also acts as a nuclear co-activator, collaborating with PR in breast cancer cells to regulate genes that promote proliferation and may contribute to endocrine therapy resistance (20). In addition, tumor suppressors such as p53 are functionally inhibited from post-translational protein modifications that result from MISS [21].

Identical rapid signaling from membrane ER, PR, or growth factor tyrosine kinase receptors modifies transcription factor abundance and recruitment to DNA [2], stimulates co-activator phosphorylation that promotes recruitment to gene promoters [20,22], and modulates the epigenome. An example of the latter is that estradiol (E2) activates PI3K-AKT in breast cancer cells, phosphorylating the EZH2 histone methyltransferase at an inhibitory site, serine 21. As a result, the repressive histone 3 lysine 27 tri-methyl (H3K27me3) mark is lifted at the promoter of key genes such as PR [23,24]. This allows for chromatin remodeling that results in E2 and AKT-dependent transactivation of the PR gene, a target of great importance for the ability of hormone replacement after menopause to cause an increased incidence of breast cancer [25]. Bredfelt et al. also showed this epigenetic modification occurs in response to diethylstilbestrol, an estrogenic compound given to women to stabilize their pregnancy that unfortunately caused a high incidence of a previously rare vaginal cancer in the female offspring of the women who used this drug [23]. This suggests that xenoestrogens and perhaps environmental steroid hormone mimetics/disruptors could activate similar pathways to produce toxic effects.

Although it is not yet clear as to how pervasively important MISS is to gene transcription, work from Miguel Beato and his colleagues suggests there is strong impact. His group reported that membrane PR signaling through ERK and the MSK1 kinase results in important chromatin modifications that promote gene transcription by nuclear PR in MCF7 cells [26]. These investigators showed that 28% of genes that were up-regulated by progesterone in these cells specifically required ERK signaling [26] and ERK/MSK-1 inhibition prevented E2 or progesterone-induced proliferation of breast tumor xenografts [27]. The multiple mechanisms by which MISS contributes to transcription suggests that many important genes will ultimately be shown to require this input for regulation, working in conjunction with nuclear steroid receptor functions.

In addition, rapid signaling by estrogen may impact tumor metabolism by non-genomic mechanisms. Depending on glucose substrate, membrane ER signaling through AKT promotes glycolysis in breast cancer cells, or shifts the cells into the Kreb's cycle and oxidative phosphorylation when glucose is reduced [3]. E2/ER promotes the latter compensation as a survival mechanism, stimulating AMP kinase-induced phosphorylation and activation of the pyruvate dehydrogenase enzyme, thereby allowing utilization of the glucose metabolite pyruvate for entry into the Kreb's cycle. Lowering glucose and inhibiting this compensatory pathway greatly enhanced γ -radiation induced apoptosis of the tumor cells [3].

Prostate

Compared to breast cancer, there has been comparatively little work on the importance of membrane-localized androgen receptors (AR) in prostate cancer. Peterziel et al described MEK-ERK, PI3K, and protein kinase C activation by androgens in prostate cancer cells [28]. Although it was unclear whether this action originated from extra-nuclear AR, such signaling was described to correlate to relative androgen independence in aggressive prostate cancer cells [29]. Similar to ER and PR, AR acts as a G protein-coupled receptor to initiate such signaling [30]. Recent work in both cell lines [31] and xenograft models [32] indicates that the paxilin protein is phosphorylated by either androgen or EGF-induced ERK signaling, facilitating signal transduction and resulting in paxillin translocation to the nucleus. In the nucleus, paxillin serves as a co-activator for AR-induced up-regulation of various oncogenes such as *CFOS* and *CCND1*, linking MISS and nuclear AR actions [32].

Osteoporosis

From utilization of membrane ER-engaging estrogenic compounds, strong evidence indicates that signaling from this pool of ER α may be sufficient to prevent osteoclast development, mitigating osteoporosis in mouse models [33]. Such prevention may specifically involve kinase-mediated signaling by estrogens to the up-regulation of expression and phosphorylation of important transcription factors [34]. Membrane ER α signaling also impacts survival of osteoblast progenitor cells and stimulates osteoclast apoptosis in cortical bone [35].

Mitigation of Cardiac Hypertrophy and Progression

Abundant data now exists from many in-vivo and in-vitro models that rapid signaling by membrane ER β prevents hypertension, resulting cardiac hypertrophy, fibrosis, and progression to heart failure [36–38]. These actions occur from the membrane ER β pool blocking the actions of hypertrophic stimuli that stimulate ERK and AKT activity, and protein phosphatase 2B activation. As a result, hypertrophic transcription factors (TFs) are retained in cytoplasm (e.g.-NFATs), or are kept inactive in the nucleus by inhibitory phosphorylation (e.g.-GATA4) [39,40]. Recently, estrogen acting at membrane ER β was shown to repress pro-hypertrophic histone deacetylase (HDAC) expression and activity, while also simulating the production and nuclear localization of anti-hypertrophic HDACs. This resulted in prevention of hypertrophic gene expression [41]. E2 and ER β also suppress cardiac fibrosis; this occurs when membrane ER^β blocks TGF^β production, the transition of fibroblasts to myofibroblasts, and TGF β signaling to pro-fibrotic gene expression [42]. Using an ER β specific agonist that avoids the uterine and breast proliferation induced by E2, strong benefit was shown in all these aspects in wild type but not $ER\beta$ deleted mice [42]. These findings also extend to $ER\beta$ preventing or even reversing pulmonary hypertension and resulting right ventricular hypertrophy in rat models [43]. Several types of acute vascular endothelial injury in mice have also been show to be prevented by administration of an estrogen-dendrimeric compound that only binds to membrane and not nuclear ER in blood vessel cells [44].

Mitochondrial Estrogen Receptors

Although steroid action in mitochondria was originally identified many years ago, the nature of the proteins that mediated steroid action was poorly defined. Recent work has identified classical glucocorticoid and estrogen receptors in mitochondria of various cells. Regarding ER, both alpha and beta isoforms have been identified in mitochondria of cardiomyocytes, neurons, and breast cancer [45,46]. Interestingly, ER α and ER β in breast cancer cells have been shown by electron microscopy to be localized primarily to the mitochondrial matrix (interior) [47]. Some data suggests that mitochondrial gene regulation occurs in part from E2 binding to ER in this organelle in breast cancer cell lines [47]. Other data indicates that mitochondrial ER^β mediates cyto-protection of MCF7 breast cancer cells upon estrogen binding this receptor pool as an agonist [48]. In response to radiation of such cells, strong oxidant stress triggers the intrinsic apoptotic program, leading to cell death. Estrogen mitigates ROS formation by up-regulating manganese superoxide dismutase (MnSOD) activity, preventing superoxide-induced apoptosis. Thus, conceptually, women with this malignancy that are receiving adjuvant therapies and are undergoing symptoms of estrogen deprivation can't take hormone replacement because it opposes the fundamental mechanism of adjuvant therapy action.

In addition, endocrine therapy sensitivity or resistance in breast cancer might be governed by mitochondrial ER β acting as an ROS rheostat [49]. From in-vitro and xenograft mouse models of tamoxifen (TAM) resistance, the SERM is effective cytotoxic therapy when it generates large amounts of superoxide. In part this is due to TAM initiating nitric oxide (NO) generation from binding ER β in mitochondria of breast cancer cells. Nitric oxide

causes nitrosylation of tyrosine 34 of MnSOD thereby inactivating this enzyme. As a result superoxide levels are very high and subsequently activate apoptosis (Fig.1). In contrast, TAM resistant tumors that grow in the presence of a TAM pellet under the skin of mouse xenograft models show high amounts of MnSOD activity due to lack of NO generation. Targeting the in-vivo tumors with lenti-virus shRNA to MnSOD completely restored the cytotoxic response to TAM, causing massive apoptosis and tumor involution in 3 weeks. Current strategies include using nanosphere-siRNA to MnSOD for expression in TAM-resistant breast tumors in-vivo and appear promising [50]. Additional studies in lung cancer implicate mitochondrial ER β as stimulating Bcl2 gene expression and moderating oxidant stress, thus promoting the survival of this malignancy [51].

Therapeutic Targets Arise from Understanding Membrane Steroid Receptor Signaling

The realization that rapid but often sustained signaling occurs when steroid ligands bind to receptor isoforms outside the nucleus suggests that selective engagement of these receptors could either prevent or treat existing diseases. Engagement of membrane ERa alone prevents osteoporosis [33] or acute vascular damage [44] in female mouse models. Membrane ERa also signals through AMPK to the suppression of all lipid synthesis in the in-vivo liver. These actions do not involve nuclear ER and potentially impact cholesterol homeostasis and related disorders [5]. Thus, compounds such as EDC that do not enter the cell and only bind to membrane ER point to an approach that could prove fruitful, avoiding toxicities that also require engagement of nuclear ER.

In some cells, steroid receptor isoforms such as ER β exist mainly outside the nucleus, and agonists for this receptor isoform could prevent or treat multiple aspects of cardiovascular disease [40]. ER antagonists that only bind to membrane receptors could also prove therapeutically useful, limiting toxicities that arise from nuclear ER binding. In the first regard, favorable brain remodeling from rapid signaling by estrogen/membrane ERa to MEK-ERK involves dendritic spine and synapse formation [52,53]. This mechanism along with reduction of oxidant stress may underlie the ability of estrogen to prevent the death of dopamine-secreting neurons that underlies the development of Parkinson's Disease [54]. MEK-ERK activation by E2 in the brain cortex limits the extent of stroke, although the latter benefit is largely confined to younger mammals as supported by animal models [55]. From understanding the key signals generated to mediate the effects of extra-nuclear steroid receptors, these pathways can also be targeted by existing kinase, calcium, or other directed inhibitors. Many therapies for disease require a multi-factorial approach, such as in hormone responsive cancers. Novel extra-nuclear steroid receptors agonists or antagonist molecules that specifically affect extra-nuclear steroid receptor functions could prove valuable in these regards.

Acknowledgments

The studies were supported by a Merit Review Award from the Department of Veterans Affairs and grant 2R01CA100366 from the National Institutes of Health.

References

- Hammes, Stephen; Levin, Ellis R. Recent advances in extra-nuclear steroid receptor actions. Endocrinology. 2011; 152:4489–4495. [PubMed: 22028449]
- Bjornstrom, Linda; Sjoberg, Maria. Mechanisms of estrogen receptor signaling: convergence of genomic and nongenomic actions on target genes. Mol Endocrinol. 2005; 19:833–842. [PubMed: 15695368]
- O'Mahony, Fiona; Pedram, Ali; Razandi, Mahnaz; Levin, Ellis R. Estrogen modulates metabolic pathway adaptation to available glucose in breast cancer cells. Mol Endocrinol. 2012; 26:2058– 2070. [PubMed: 23028062]
- Pedram, Ali; Razandi, Mahnaz; Kim, Jin K.; O'Mahony, Fiona; Lee, Eva; Luderer, Ulrike; Levin, Ellis R. Developmental phenotype of a membrane only estrogen receptor; (MOER) mouse. J Biol Chem. 2009; 284:3488–3495. [PubMed: 19054762]
- Pedram, Ali; Razandi, Mahnaz; O'Mahony, Fiona; Harvey, Harry; Harvey, Brian J.; Levin, Ellis R. Estrogen reduces lipid content in the liver exclusively from membrane receptor signaling. Science Signaling. 2013; 6:RA36. [PubMed: 23695162]
- Werner, Haim. Tumor suppressors govern insulin-like growth factor signaling pathways: implications in metabolism and cancer. Oncogene. 2012; 31:2703–2714. [PubMed: 21963847]
- Masuda, Hiroko; Zhang, Dangwei; Bartholomeusz, Chandra; Doihara, Hiroshi; Hortobagyi, Gabriel Naoto; Ueno, NT. Role of epidermal growth factor receptor in breast cancer. Breast Cancer Res Treat. 2012; 136:331–345. [PubMed: 23073759]
- Nielsen, Dorte L.; Kümler, Iben; Palshof, Jesper A.; Andersson, Michael. Efficacy of HER2targeted therapy in metastatic breast cancer. Monoclonal antibodies and tyrosine kinase inhibitors. Breast. 2013; 22:1–12. [PubMed: 23084121]
- Osborne, C Kent; Schiff, Rachel. Mechanisms of endocrine resistance in breast cancer. Annu Rev Med. 2011; 62:233–247. [PubMed: 20887199]
- Hurtado, Antoni; Holmes, Kelly A.; Geistlinger, Timothy R.; Hutcheson, Iain R.; Nicholson, Robert I.; Brown, Myles; Jiang, Jie; Howat, William J.; Ali, Simak; Carroll, Jason S. Regulation of ERBB2 by oestrogen receptor-PAX2 determines response to tamoxifen. Nature. 2008; 456:663– 666. [PubMed: 19005469]
- Migliaccio, Antimo; Di Domenico, Marina; Castoria, Gabriella; de Falco, Antonio; Bontempo, Paolo; Nola, Enrico; Auricchio, Ferdinando. Tyrosine kinase/p21ras/MAP-kinase pathway activation by estradiol-receptor complex in MCF-7 cells. EMBO J. 1996; 15:1292–1300. [PubMed: 8635462]
- Skildum, Andrew; Faivre, Emily; Lange, Carol A. Progesterone receptors induce cell cycle progression via activation of mitogen-activated protein kinases. Mol Endocrinol. 2005; 19:327– 339. [PubMed: 15486045]
- 13. Razandi, Mahnaz; Pedram, Ali; Parks, Steven T.; Levin, Ellis R. Proximal events in ER signaling from the plasma membrane. J Biol Chem. 2003; 278:2701–2712. [PubMed: 12421825]
- 14. Song, Robert X.; Zhang, Zhenguo; Chen, Yucai; Bao, Yongde; Santen, Richard J. Estrogen signaling via a linear pathway involving insulin-like growth factor I receptor, matrix metalloproteinases, and epidermal growth factor receptor to activate mitogen-activated protein kinase in MCF-7 breast cancer cells. Endocrinology. 2007; 148:4091–4101. [PubMed: 17525128]
- Poulard, Coralie; Treilleux, Isabelle; Lavergne, Emilie; Bouchekioua-Bouzaghou, Katia; Goddard-Leon, Sophie; Chabaud, Sylvie; Tredan, Olivier; Corbo, Laura; Le Romancer, Muriel. Activation of rapid estrogen signaling in aggressive human breast cancers. EMBO Mol Med. 2012; 4:1200–1213. [PubMed: 23065768]
- Razandi, Mahnaz; Pedram, Ali; Rosen, Eliot M.; Levin, Ellis R. BRCA1 inhibits membrane estrogen and growth factor receptor signaling to cell proliferation in breast cancer. Mol Cell Biol. 2004; 24:5900–5913. [PubMed: 15199145]
- Marino, Maria; Acconcia, Filippo; Bresciani, Francesco; Weisz, Alessandro; Trentalance, Anna. Distinct nongenomic signal transduction pathways controlled by 17β-estradiol regulate DNA synthesis and cyclin D1 gene transcription in HepG2 cells. Mol Biol Cell. 2002; 13:3720–3729. [PubMed: 12388769]

- 18. Kim, Hogyoung; Abd Elmageed, Zakaria Y.; Ju, Jihang; Naura, Amarjit S.; Abdel-Mageed, Asim B.; Varughese, Shibu; Paul, Dennis; Alahari, Suresh; Catling, Andrew; Kim, Jong G.; Boulares, A Hamid. PDZK1 is a novel factor in breast cancer that is indirectly regulated by estrogen through IGF-1R and promotes estrogen-mediated growth. Mol Med. 2013; 19:253–262. [PubMed: 23821363]
- Faivre, Emily J.; Lange, Carol A. Progesterone receptors upregulate Wnt-1 to induce epidermal growth factor receptor transactivation and c-Src-dependent sustained activation of Erk1/2 mitogenactivated protein kinase in breast cancer cells. Mol Cell Biol. 2007; 27:466–480. [PubMed: 17074804]
- Dressing, Gwen E.; Knutson, Todd P.; Schiewer, Matthew J.; Daniel, Andrea R.; Hagan, Christy R.; Diep, Caroline H.; Knudsen, Karen E.; Lange, Carol A. Progesterone Receptor-Cyclin D1 Complexes Induce Cell Cycle-Dependent Transcriptional Programs In Breast Cancer Cells. Mol Endocrinol. 2014 Feb.25 me20131196.
- Pedram, Ali; Razandi, Mahnaz; Evinger, Albert J.; Lee, Eva; Levin, Ellis R. Estrogen inhibits ATR signaling to cell cycle checkpoints and DNA repair. Mol Biol Cell. 2009; 20:3374–3389. [PubMed: 19477925]
- 22. York, Brian; Yu, Chundong; Sagen, Jørn V.; Liu, Zhaliang; Nikolai, Bryan C.; Wu, Ray-Chang; Finegold, Milton; Xu, Jianming; O'Malley, Bert W. Reprogramming the posttranslational code of SRC-3 confers a switch in mammalian systems biology. Proc Natl Acad Sci. 2010; 107:11122– 11127. [PubMed: 20534466]
- Bredfeldt, Tiffany G.; Greathouse, K Leigh; Safe, Stephen H.; Hung, Mien-Chie; Bedford, Mark T.; Walker, Cheryl L. Xenoestrogen-induced regulation of EZH2 and histone methylation via estrogen receptor signaling to PI3K/AKT. Mol Endocrinol. 2010; 24:993–1006. [PubMed: 20351197]
- Pedram, Ali; Razandi, Mahnaz; Deschenes, Robert; Levin, Ellis R. DHHC 7 and 21 are palmitoylacyltranferases for sex steroid receptors. Mol Biol Cell. 2012; 23:188–199. [PubMed: 22031296]
- 25. Chlebowski, Rowan T.; Hendrix, Susan L.; Langer, Robert D.; Stefanick, Marcia L.; Gass, Margery; Lane, Dorothy; Rodabough, Rebecca J.; Gilligan, Mary Ann; Cyr, Michele G.; Thomson, Cynthia A.; Khandekar, Janardan; Petrocich, Helen; McTiernan, Anne. WHI Investigators. Influence of estrogen plus progestin on breast cancer and mammography in healthy postmenopausal women: the Women's Health Initiative Randomized Trial. JAMA. 2003; 289:3243–3253. [PubMed: 12824205]
- Vicent, Guillermo P.; Ballaré, Cecilia; Nacht, A Silvina; Clausell, Jaime; Subtil-Rodriguez, Alicia; Quiles, Ignacio; Jordan, Albert; Beato, Miguel. Induction of progesterone target genes requires activation of Erk and Msk kinases and phosphorylation of histone H3. Mol Cell. 2006; 24:367– 381. [PubMed: 17081988]
- 27. Reyes, Diego; Ballaré, Carlos; Castellano, Giuliano; Soronellas, Daniel; Bagó, Julie R.; Blanco, Jose; Beato, Miguel. Activation of mitogen-and stress-activated kinase 1 is required for proliferation of breast cancer cells in response to estrogens or progestins. Oncogene. 2013
- Peterziel, Heike; Mink, Sigrun; Schonert, Annette; Becker, Mathias; Klocker, Helmut; Cato, Andrew C. Rapid signaling by androgen receptor in prostate cancer cells. Oncogene. 1999; 18:6322–6329. [PubMed: 10597231]
- Unni, Emmanual; Sun, Shihua; Nan, Bicheng; McPhaul, Michael J.; Cheskis, Boris; Mancini, Michael A.; Marcelli, Marco. Changes in androgen receptor nongenotropic signaling correlate with transition of LNCaP cells to androgen independence. Cancer Res. 2004; 64:7156–7168. [PubMed: 15466214]
- Sen, Aritro; Prizant, Hen; Hammes, Stephen R. Understanding extranuclear (nongenomic) androgen signaling: what a frog oocyte can tell us about human biology. Steroids. 2011; 76:822– 828. [PubMed: 21354434]
- 31. Sen, Aritro; O'Malley, Katherine; Wang, Zhou; Raj, Ganesh V.; Defranco, Donald B.; Hammes, Stephen R. Paxillin regulates androgen-and epidermal growth factor-induced MAPK signaling and cell proliferation in prostate cancer cells. J Biol Chem. 2010; 285:28787–28795. [PubMed: 20628053]

- 32. Sen, Aritro; De Castro, Ismary; Defranco, Donald B.; Deng, Fang-Ming; Melamed, Jonathan; Kapur, Payel; Raj, Ganesh V.; Rossi, Randall; Hammes, Stephen R. Paxillin mediates extranuclear and intranuclear signaling in prostate cancer proliferation. J Clin Invest. 2012; 122:2469–2481. [PubMed: 22684108]
- 33. Kousteni, Stavroula; Chen, JR.; Bellido, Teresita; Han, Li; Ali, Alan A.; O'Brien, Christine A.; Plotkin, Lilian; Fu, Quen; Mancino, AndrewT; Wen, Yin; Vertino, Antonio M.; Powers, Christopher C.; Stewart, Steven A.; Ebert, Robert; Parfitt, A Michael; Weinstein, Robert S.; Jilka, Robert L.; Manolagas, Stavros C. Reversal of bone loss in mice by nongenotropic signaling of sex steroids. Science. 2002; 298:843–846. [PubMed: 12399595]
- Kousteni, Stavroula; Han, Li; Chen, Jin-Ran; Almeida, Maria; Plotkin, Lilian I.; Bellido, Teresita; Manolagas, Stavros C. Kinase-mediated regulation of common transcription factors accounts for the bone-protective effects of sex steroids. J Clin Invest. 2003; 111:1651–1664. [PubMed: 12782668]
- 35. Bartell, Shashana M.; Han, Li; Kim, Ha-neui; Kim, Sung H.; Katzenellenbogen, John A.; Katzenellenbogen, Benita S.; Chambliss, Ken L.; Shaul, Philip W.; Roberson, Paula K.; Weinstein, Robert S.; Jilka, Robert L.; Almeida, Maria; Manolagas, Stavros C. Non-nuclearinitiated actions of the estrogen receptor protect cortical bone mass. Mol Endocrinol. 2013; 27:649–656. [PubMed: 23443267]
- 36. Cavasin, Maria A.; Sankey, Steadman S.; Yu, Ai-Li; Menon, Shreevidya; Yang, Xiao-Ping. Estrogen and testosterone have opposing effects on chronic cardiac remodeling and function in mice with myocardial infarction. Am J Physiol Heart Circ Physiol. 2003; 284:H1560–H1569. [PubMed: 12560213]
- 37. Satoh, Minora; Matter, Christian M.; Ogita, Hisakuzu; Takeshita, Kyosuke; Wang, Chao-Yung; Dorn, Gerald W., II; Liao, James K. Inhibition of apoptosis-regulated signaling kinase-1 and prevention of congestive heart failure by estrogen. Circulation. 2007; 115:3197–3204. [PubMed: 17562954]
- 38. Jazbutyte, Virginija; Arias-Loza, Paula A.; Hu, Kai; Widder, Julian; Govindaraj, Vijayakumar; von Poser-Klein, Christine; Bauersachs, Johann; Fritzemeier, Karl-Heinrich; Hegele-Hartung, Christa; Neyses, Ludwig; Ertl, Georg; Pelzer, Theo. Ligand-dependent activation of ER lowers blood pressure and attenuates cardiac hypertrophy in ovariectomized SHR. Cardiovasc Res. 2008; 77:774–781. [PubMed: 18056768]
- Pedram, Ali; Razandi, Mahnaz; Lubahn, Dennis; Liu, Jinghua; Vannan, Mani; Levin, Ellis R. Estrogen inhibits cardiac hypertrophy: Role of estrogen receptor beta to inhibit calcineurin. Endocrinology. 2008; 149:3361–3369. [PubMed: 18372323]
- Pedram, Ali; Razandi, Mahnaz; Korach, Kenneth; Narayanan, Ramesh; Dalton, James; Levin, Ellis R. ERb selective agonist inhibits angiotensin-induced cardiovascular pathology in female mice. Endocrinology. 2013; 154:4352–4364. [PubMed: 23970786]
- Pedram, Ali; Razandi, Mahnaz; Narayanan, Ramesh; Dalton, James; McKinsey, Timothy A.; Levin, Ellis R. Estrogen regulates histone deacetylases to prevent cardiac hypertrophy. Mol Biol Cell. 2013; 24:3805–3818. [PubMed: 24152730]
- Pedram, Ali; Razandi, Mahnaz; O'Mahony, Fiona; Lubahn, Dennis; Levin, Ellis R. Estrogen receptor beta prevents cardiac fibrosis. Mol Endocrinol. 2010; 24:2152–2165. [PubMed: 20810711]
- Nadadur, Rangarajan D.; Umar, Soban; Wong, Gabriel; Eghbali, Mansour; Iorga, Andrea; Matori, Humann; Partow-Navid, Rod; Eghbali, Mansoureh. Reverse right ventricular structural and extracellular matrix remodeling by estrogen in severe pulmonary hypertension. J Appl Physiol. 2012; 113:149–158. [PubMed: 22628376]
- 44. Chambliss, Ken L.; Wu, Qian; Oltmann, Sarah; Konaniah, Eddy S.; Umetani, Michihisa; Korach, Kenneth S.; Thomas, Gail D.; Mineo, Chieko; Yuhanna, Ivan S.; Kim, Sung H.; Madak-Erdogan, Zeynep; Maggi, Adriana; Dineen, Sean P.; Roland, Christina L.; Hui, David Y.; Brekken, Rolf A.; Katzenellenbogen, John A.; Katzenellenbogen, Benita S.; Shaul, Philip W. Non-nuclear estrogen receptor a signaling promotes cardiovascular protection but not uterine or breast cancer growth in mice. J Clin Invest. 2010; 120:2319–2330. [PubMed: 20577047]
- 45. Yang, Shao-Hun; Liu, Ran; Perez, Evelyn J.; Wen, Yi; Stevens, Stanley M., Jr; Valencia, Thomas; Brun-Zinkernagel, Anne-Marie; Prokai, Laszlo; Will, Yvonne; Dykens, James; Koulen, Peter;

Simpkins, James W. Mitochondrial localization of estrogen receptor beta. Proc Natl Acad Sci U S A. 2004; 101:4130–4135. [PubMed: 15024130]

- 46. Chen, Jin-Qiang; Delannoy, Michael; Cooke, Carol; Yager, James D. Mitochondrial localization of ERα and ERβ in human MCF-7 cells. Am J Physiol Endocrinol Metab. 2004; 28:E1011–E1022. [PubMed: 14736707]
- 47. Chen, Jin-Qiang; Yager, James D.; Russo, Jose. Regulation of mitochondrial respiratory chain structure and function by estrogens/estrogen receptors and potential physiological/ pathophysiological implications. Biochim Biophys Acta. 2005; 1746:1–17. [PubMed: 16169101]
- Pedram, Ali; Razandi, Mahnaz; Wallace, Douglas C.; Levin, Ellis R. Functional estrogen receptors in the mitochondria of breast cancer cells. Mol Biol Cell. 2006; 17:2125–2137. [PubMed: 16495339]
- Razandi, Mahnaz; Pedram, Ali; Jordan, V Craig; Fuqua, Suzanne; Levin, Ellis R. Tamoxifen regulates cell fate through mitochondrial estrogen receptor beta in breast cancer. Oncogene. 2013; 32:3274–3285. [PubMed: 22907432]
- 50. Cho, Soo K.; Pedram, Ali; Levin, Ellis R.; Kwon, Young J. Acid-degradable core-shell nanoparticles for reversed Tamoxifen-reistance in breast cancer by silencing manganese superoxide dismutase (MnSOD). Biomaterials. 2013; 34:10228–10237. [PubMed: 24055523]
- Zhang, GuangFeng; Yanamala, Naveena; Lathrop, Kira L.; Zhang, Lin; Klein-Seetharaman, Judith; Srinivas, Harish. Ligand-independent antiapoptotic function of estrogen receptor-beta in lung cancer cells. Mol Endocrinol. 2010; 24:1737–1747. [PubMed: 20660297]
- Tabatadze, Nino; Smejkalova, Tereza; Woolley, Catherine S. Distribution and posttranslational modification of synaptic ERα in the adult female rat hippocampus. Endocrinology. 2013; 154:819–830. [PubMed: 23183182]
- Woolley, Catherine S. Acute effects of estrogen on neuronal physiology. Annu Rev Pharmacol Toxicol. 2007; 47:657–680. [PubMed: 16918306]
- 54. Rodriguez-Perez, Ana I.; Dominguez-Meijide, Antonio; Lanciego, Jose L.; Guerra, Maria J.; Labandeira-Garcia, Jose L. Inhibition of Rho kinase mediates the neuroprotective effects of estrogen in the MPTP model of Parkinson's disease. Neurobiol Dis. 2013; 58:209–219. [PubMed: 23774254]
- Koellhoffer, Edward C.; McCullough, Louise D. The effects of estrogen in ischemic stroke. Transl Stroke Res. 2013; 4:390–401. [PubMed: 24323337]

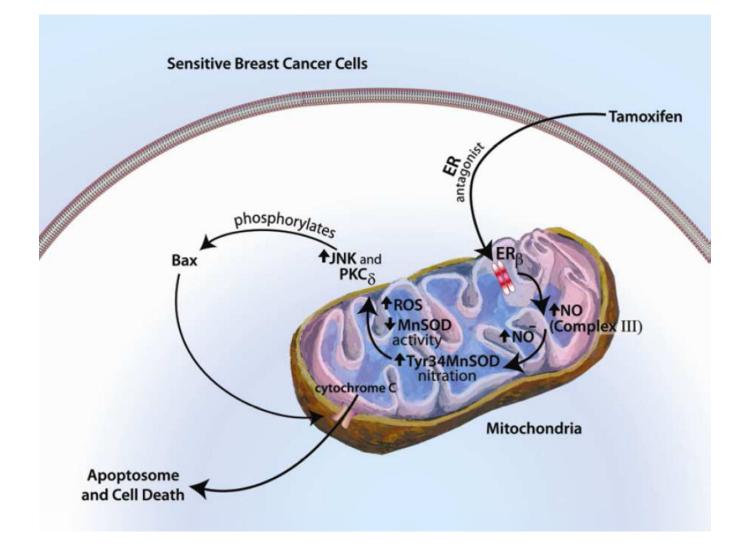


Fig. 1.

Cytotoxic effects of Tamoxifen in responsive breast cancer cells and tumors. Tamoxifen binds mitochondrial ER β as an antagonist, stimulating nitric oxide generation from Complex III of the electron transport chain, nitrosylating MnSOD to reduce its activity that results in high amounts of superoxide. Oxidative stress induces the intrinsic apoptotic program. In Tamoxifen resistant cells, this selective estrogen receptor modulator binds ER β as an agonist and does not reduce MnSOD activity, thus promoting tumor survival.