

# **UC Berkeley**

## **UC Berkeley Previously Published Works**

### **Title**

VLDL receptor gene therapy for reducing atherogenic lipoproteins.

### **Permalink**

<https://escholarship.org/uc/item/4np5r1pg>

### **Authors**

Krauss, Ronald M

Lu, Jonathan T

Higgins, Joseph J

et al.

### **Publication Date**

2023-03-01

### **DOI**

10.1016/j.molmet.2023.101685

Peer reviewed

# VLDL receptor gene therapy for reducing atherogenic lipoproteins



Ronald M. Krauss<sup>1,\*</sup>, Jonathan T. Lu<sup>2</sup>, Joseph J. Higgins<sup>2</sup>, Cathryn M. Clary<sup>2</sup>, Ray Tabibiazar<sup>2</sup>

## ABSTRACT

Over the past 40 years, there has been considerable research into the management and treatment of atherogenic lipid disorders. Although the majority of treatments and management strategies for cardiovascular disease (CVD) center around targeting low-density lipoprotein cholesterol (LDL-C), there is mounting evidence for the residual CVD risk attributed to high triglyceride (TG) and lipoprotein(a) (Lp(a)) levels despite the presence of lowered LDL-C levels. Among the biological mechanisms for clearing TG-rich lipoproteins, the VLDL receptor (VLDLR) plays a key role in the trafficking and metabolism of lipoprotein particles in multiple tissues, but it is not ordinarily expressed in the liver. Since VLDLR is capable of binding and internalizing apoE-containing TG-rich lipoproteins as well as Lp(a), hepatic VLDLR expression has the potential for promoting clearance of these atherogenic particles from the circulation and managing the residual CVD risk not addressed by current lipid lowering therapies. This review provides an overview of VLDLR function and the potential for developing a genetic medicine based on liver-targeted VLDLR gene expression.

© 2023 The Authors. Published by Elsevier GmbH. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

**Keywords** Lipid disorders; VLDL; VLDL receptor; Triglycerides; lipoprotein(a); Gene therapy

## 1. INTRODUCTION

Lipid disorders affect over 105 million people in the US alone [1]. Although many patients are asymptomatic, left untreated, chronic elevations in atherogenic lipoproteins can lead to clinical cardiovascular disease (CVD) events, namely myocardial infarction and stroke [2]. In addition, extreme elevations of plasma triglyceride (TG) levels can result in acute pancreatitis [3], and as with CVD, a potentially fatal outcome. There has been extensive research devoted to the study of this diverse class of disorders and, in particular, to improving treatments that are aimed at controlling and reducing pathologic lipoprotein levels.

Multiple strategies have been pursued for managing atherogenic lipid disorders, most notably low-density lipoprotein cholesterol (LDL-C) lowering, which exhibits the highest level of evidence for preventing atherosclerotic CVD [4–6]. For this reason, current treatment guidelines for CVD risk reduction center around the reduction of LDL-C, with the intensity of lowering dependent on the overall assessment of CVD risk [7]. There is abundant evidence for the atherogenic properties of TG-rich lipoproteins, which include very low-density lipoprotein (VLDL) and intermediate-density lipoprotein (IDL), as well as lipolytic remnants of VLDL and chylomicrons [8,9]. While genetic evidence has pointed to plasma triglyceride (TG) as an independent CVD risk factor [10], there has yet been no consensus on targeting elevated levels of these lipoproteins for CVD prevention. However, TG reduction is recommended for pancreatitis protection in individuals with severe hypertriglyceridemia (TG > 500 mg/dL) [11].

## 2. CURRENT GAPS IN THE MANAGEMENT OF HIGH-RISK LEVELS OF TG-RICH LIPOPROTEINS AND LP(A)

### 2.1. TG-associated residual risk with statin treatment

There have been several studies that highlight the importance of TG lowering for meaningful CVD risk reduction with statins, the most widely used drug class for LDL-C reduction and CVD prevention. Subgroup analyses in statin clinical trials have shown that patients with high TG and low high-density lipoprotein cholesterol (HDL-C) have worse CVD outcomes than those with isolated elevation of LDL-C. For example, an analysis of the Scandinavian Simvastatin Survival Study (4 S) trial found that the CVD event rate in patients with hypercholesterolemia was highest in the subgroup with high TG and low HDL-C and that this group had a more favorable effect with simvastatin treatment than those with isolated elevated LDL-C levels [12]. Likewise, the Pravastatin or Atorvastatin Evaluation and Infection Therapy-Thrombolysis in Myocardial Infarction 22 trial (PROVE IT-TIMI 22), which assessed the impact of on-treatment TG levels on coronary heart disease (CHD) risk after an acute coronary syndrome, found that on-treatment TG < 150 mg/dl was independently associated with a lower risk of recurrent CHD events [13]. These data suggest that reducing TG can be a means of treating residual risk and further reducing CVD events beyond LDL-C lowering.

### 2.2. CVD risk in trials of TG-lowering drugs

Clinical trials of TG-lowering with fibrate drugs, in particular fenofibrate, have had limited success in reducing CVD risk, though subgroup

<sup>1</sup>University of California, San Francisco, 5700 Martin Luther King, Jr. Way, Oakland CA 94609, USA <sup>2</sup>SalioGen Therapeutics, Lexington, MA, USA

\*Corresponding author. E-mail: [ronald.krauss@ucsf.edu](mailto:ronald.krauss@ucsf.edu) (R.M. Krauss).

Received November 18, 2022 • Revision received January 16, 2023 • Accepted January 29, 2023 • Available online 4 February 2023

<https://doi.org/10.1016/j.molmet.2023.101685>

**Abbreviations:**

AAV	adeno-associated virus	HDL-C	high-density lipoprotein cholesterol
apo(a)	apolipoprotein a	IDL	intermediate-density lipoprotein
apoB	apolipoprotein B	LDL	low-density lipoprotein
apoB48	apolipoprotein B48	LDLR	low-density lipoprotein receptor
apoB100	apolipoprotein B100	LDL-C	low-density lipoprotein cholesterol
apoC3	apolipoprotein C-III	LNP	lipid nanoparticle
apoE	apolipoprotein E	Lp(a)	lipoprotein (a)
CHD	coronary heart disease	LRP-1	low-density lipoprotein receptor-related protein 1
CRISPR	clustered regularly interspaced short palindromic repeats	MHC	major histocompatibility complex
CVD	cardiovascular disease	NAFLD	nonalcoholic fatty liver disease
DHA	docosahexaenoic acid	PPAR $\alpha$	peroxisome proliferator-activated receptor alpha
DSBs	double-stranded breaks	PPAR $\beta/\delta$	peroxisome proliferator-activated receptor beta delta
EPA	eicosapentanoic acid	PCSK9	proprotein convertase subtilisin/kexin type 9
ER	endoplasmic reticulum	siRNA	small interfering ribonucleic acid
FGF21	fibroblast growth factor 21	T2DM	type-II diabetes mellitus
GalNAc	N-acetyl-D-galactosamine	TG	triglyceride
		VLDL	very low-density lipoprotein
		VLDLR	very low-density lipoprotein receptor

analyses have suggested CVD reducing benefit in subgroups with high TG and low HDL-C [14–16]. However, the recent PROMINENT trial of pemetreotide, a more potent fibrate drug, in patients with type 2 diabetes mellitus, mild-to-moderate hypertriglyceridemia, and low levels of HDL-C while treated with statins, found no effect on the primary CVD endpoint despite significant reductions of TG, VLDL cholesterol, remnant cholesterol, and apolipoprotein C-III [17]. There was however no reduction of plasma apoB-100, a measure representing levels of all atherogenic particles, and it was suggested in an accompanying editorial that to be effective for CVD risk reduction, a TG-lowering therapy should have a mechanism for increasing clearance of VLDL remnant particles, rather than, as appeared to be the case for pemetreotide, converting them to LDL [18]. Notably, induction of hepatic VLDL expression by fenofibrate was found to be required for its TG-lowering effect in mouse models [19], although this study did not assess fenofibrate's effect on VLDL remnant clearance or plasma apoB levels. Trials of the omega-3 fatty acids eicosapentanoic acid (EPA) and docosahexaenoic acid, which can reduce plasma TG levels, have also not consistently been found to reduce CVD risk [20]. While the Japan EPA Lipid Intervention Study (JELIS) found that EPA (1800 mg/day) in addition to treatment with a statin reduced CVD events in the statin-treated patients [21], the more recent STRENGTH trial of Epanova—a mixture of EPA and DHA—was stopped early because no significant reduction in a composite outcome of major adverse CVD events was seen [22]. In contrast, the REDUCE-IT trial of icosapent ethyl, an EPA-derived drug, showed overall reduction in myocardial infarction in statin-treated patients, but the effect was not found to be related to plasma lipid levels [23,24] and the interpretation of the results has been questioned due to the use of mineral oil as a control [25–28]. While treatment with an anti-sense inhibitor of ANGPTL3 (Vupanorsen) has been shown to lower TG levels [29], it is only currently approved for LDL lowering in homozygous familial hypercholesterolemia [30]. Furthermore, another TG-lowering treatment currently in development is the inhibition of apoC3 synthesis with an anti-sense oligonucleotide or siRNA [31–34], but as yet there are no studies of CVD outcomes with these agents. An overall consideration with regard to assessing the cardiovascular benefit of TG-lowering drugs is that TG levels may need to be lower than those achieved in study trials. For example, it has been reported that even for individuals with low to moderate CVD

risk and normal LDL-C levels, TG levels  $\geq 150$  mg/dl were significantly associated with the presence of arterial inflammation, a significant contributor to development of atherosclerotic CVD [35].

### 2.3. Reduction of elevated Lp(a)

Another important factor contributing to residual CVD risk in statin-treated patients is elevated lipoprotein (a) (Lp(a)), an atherogenic lipoprotein particle whose levels are not lowered by statin treatment, as discussed further below [36]. Modest reductions of Lp(a) can be achieved with high dose nicotinic acid [37] or PCSK9 inhibitors [38], but these effects have not been shown to normalize high-risk Lp(a) levels, and there are no studies to date that have assessed the effects of Lp(a) lowering on CVD outcomes [39,40]. There are newer treatments for Lp(a) currently in development that involve inhibiting apo(a) synthesis with antisense oligonucleotides [41] and siRNA [42], but their effects on CVD risk remain to be tested.

## 3. HEPATIC EXPRESSION OF THE VLDL RECEPTOR AS A MEANS OF REDUCING LEVELS OF ATHEROGENIC LIPOPROTEINS AND TREATING SEVERE HYPERTRIGLYCERIDEMIA

As reviewed above, there is a strong rationale for reducing residual CVD risk in statin-treated patients by achieving maximal reduction of atherogenic TG-rich lipoproteins and Lp(a). While currently available drugs and those in development have this potential to varying degrees, other therapeutic possibilities should be considered, including the use of a genetic medicine that would preclude the compliance issues that significantly affect the efficacy of lipid lowering drugs in clinical practice [43,44].

An attractive possibility in this regard is a targeted gene therapy for achieving hepatic expression of the VLDL receptor (VLDLR). The VLDLR has been shown to mediate binding and cellular internalization of both TG-rich lipoproteins and Lp(a). VLDLR is not ordinarily expressed in hepatocytes, where lipoprotein particles are cleared from plasma and degraded. In the sub-sections below, we review key features of the VLDLR that provide a rationale for ectopically introducing the *VLDLR* gene into the liver as a means of achieving sustained reductions of pathologic lipoprotein levels and lowering the residual CVD risk resulting from conventional lipid lowering drug therapies.

### 3.1. Structure of VLDLR

VLDLR is a multifunctional receptor that shares structural homology with members of the LDL receptor (LDLR) gene family [45]. VLDLR has five protein domains: extracellular N-terminal ligand-binding domain with eight cysteine-rich repeats, epidermal growth factor domain, O-linked glycosylation sugar domain, single transmembrane domain, and a cytoplasmic domain with the NPxY motif for signal transduction. Isolation and characterization of cDNAs encoding human *VLDLR* show two forms of the receptor: one full length that resembles the low-density lipoprotein receptor (LDLR), with the exception that LDLR has five cysteine-rich repeats, and a variant form that lacks the O-linked sugar domain [46]. In addition, alternative splicing of *VLDLR* generates multiple transcript variants encoding distinct isoforms, though their protein expression and precise functions have not been established [47].

### 3.2. Physiological functions of VLDLR

VLDLR is ubiquitously expressed in the heart, skeletal muscle, adipose tissue, endothelium, and brain, as well as in macrophages [45,48]. While not natively present in the liver, hepatic expression can be induced by specific conditions including endoplasmic reticulum stress and treatment with fenofibrate, a PPAR $\alpha$  agonist. VLDLR expression is insulin dependent [49] and unlike LDLR, VLDLR expression is not regulated by cellular cholesterol content [50].

VLDLR plays a variety of roles in different tissues. A key function of greatest relevance to this review is its role in lipoprotein uptake, by which it promotes fatty acid  $\beta$ -oxidation in heart and muscle and storage of TGs in adipose tissue. VLDLR also has a role in cellular signaling, notably in the Reelin pathway, which is responsible for the migration of neurons to their proper locations during brain development [51].

While apoE is a ligand for both VLDLR and LDLR binding of lipoproteins, VLDLR differs from LDLR in that it does not bind apoB, the major structural protein of VLDL and LDL particles [52]. Hence it promotes binding and endocytosis of apoE-containing TG-rich lipoproteins. VLDLR up-regulates lipoprotein lipase (LPL)-mediated TG hydrolysis along with the direct uptake of TG-rich lipoproteins in endothelial cell [53]. TG-rich lipoprotein uptake is further increased by the addition of apoE and inactivated LPL [54,55].

### 3.3. Atherogenic lipoproteins recognized by VLDLR

#### 3.3.1. VLDL

VLDLs are primarily responsible for the transport of endogenously synthesized TG and cholesterol from the liver into the bloodstream and to other areas of the body. The main structural features of VLDL consist of a hydrophobic lipid core (TG and cholesteryl ester) coated by a hydrophilic monolayer composed of phospholipids, free cholesterol, and multiple apolipoproteins, with apoB100 as the primary structural component [56]. In contrast to the accepted model that VLDL is a spherical emulsion-like particle, cryo-electron microscopy-derived 3D structural reconstructions of VLDL reveal a polyhedral shape [57]. This finding suggests that the flat polyhedral surfaces contribute to its binding affinity for VLDLR.

Following the secretion of VLDL into the circulation, TG is hydrolyzed by LPL in peripheral tissues [58], resulting in uptake of free fatty acids and the formation of apoE-enriched remnant lipoproteins. The hepatic uptake of these apoE-enriched remnant lipoproteins is mediated by the binding of apoE to three types of receptors, LDLR, LRP-1, and syndecan, and inhibited by apoC3 [59]. Further intravascular metabolism

of these remnants leads to the formation of LDL. The elevated levels of VLDL and VLDL remnant lipoproteins, like LDL, result in an increased risk for cardiovascular disease [60].

#### 3.3.2. Chylomicrons

Chylomicrons are responsible for the transport of exogenous (dietary) TG and cholesterol from the intestines into the circulation [61]. They are larger than VLDL, with a higher TG content, but structurally resemble VLDL except that the primary protein component in chylomicrons is ApoB48 rather than ApoB100. Chylomicron metabolism is similar to that of VLDL, except that the remnants are cleared rapidly by the liver without the intermediate formation of LDL. Importantly, elevated levels of chylomicron remnants, like VLDL remnants, result in an increased risk for atherosclerotic cardiovascular disease. On the other hand, extreme elevations of chylomicron levels as in patients with LPL deficiency and other genetic traits, result in severe hypertriglyceridemia with the concomitant risk for acute pancreatitis, a potentially fatal condition [62–64].

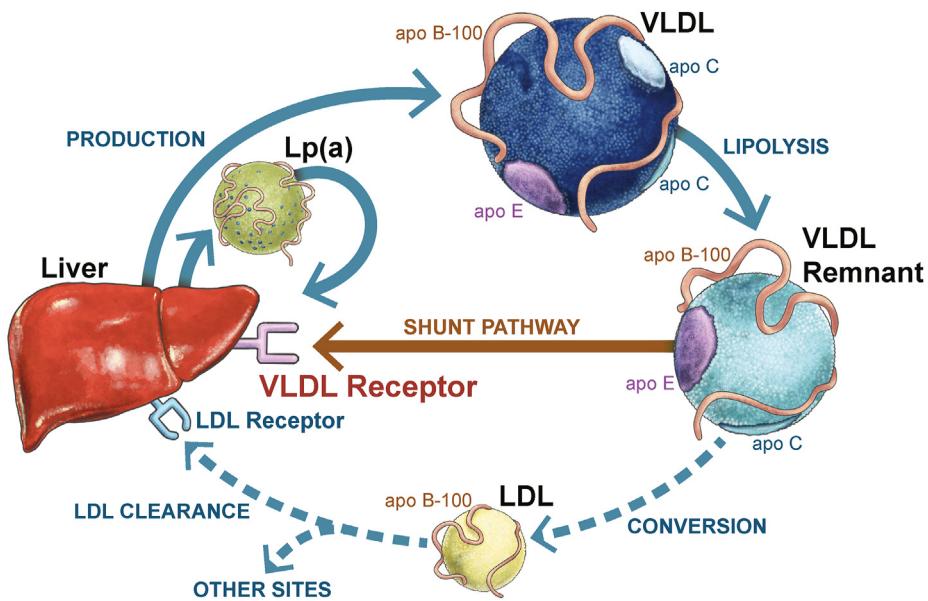
#### 3.3.3. Lp(a)

Lp(a) is an atherogenic particle that consists of an LDL particle covalently attached to apo(a), a protein with high homology to plasminogen by virtue of multiple repeats resembling plasminogen's kringle domain [65,66]. Lp(a) is a major plasma transporter of oxidized phospholipids that, together with its other compositional features, plays a key role in its atherogenesis, including inflammation and thrombosis [67–69]. Allelic variations within the apo(a) gene locus strongly determine plasma levels of Lp(a) [70]. Lp(a) genetic heterogeneity is also manifested by the wide variation in the number of its plasminogen kringle-like repeats. High levels of Lp(a) are associated with increased risk of CHD and stroke, as well as calcific aortic stenosis [71]. Population studies support that Lp(a) levels in the upper tertile are associated with significantly increased CVD risk [72]. Hepatic synthesis and secretion are major determinants of plasma Lp(a) levels, since Lp(a) fractional plasma clearance is low. This is likely related to its relatively low affinity for LDLR, which accounts for the failure of statin therapy to significantly lower Lp(a) levels [73]. VLDLR has a high affinity for Lp(a), and is capable of mediating its cellular uptake, by a mechanism other than the binding to apoE [74]. Thus, the endocytosis of Lp(a) by macrophage-laden VLDLR promotes the formation of lipid-enriched foam cells [74,75]. This mechanism along with VLDLR-mediated uptake of TG-rich particles, contributes to the atherogenic effect of macrophage VLDLR expression. To date, however, the potential for specifically expressing VLDLR in the liver for lowering plasma levels of Lp(a) by promoting its clearance has not been examined.

## 4. VLDLR AS A THERAPEUTIC TARGET FOR LIPID DISORDERS

**Figure 1** illustrates the potential for genetically mediated hepatic VLDLR expression to reduce CVD risk by promoting plasma clearance of atherogenic VLDL remnant and Lp(a) particles.

A number of studies in mouse models have shown reductions in plasma lipid levels and atherosclerosis with liver-directed VLDLR gene therapy via adeno-associated virus (AAV) or helper-dependent adenoviral vector delivery [76–78], though as described below, there have been limitations of this approach in terms of efficacy and safety, and thus, more recently, the use of other gene delivery systems has been explored [79]. However, despite its promise, VLDLR has not been further pursued as a therapeutic target, and the reason appears to be two-fold.



**Figure 1: Role of hepatic VLDL receptor expression in the clearance of atherogenic particles.** Overview of the pathways by which hepatic VLDL receptor expression may increase plasma clearance of atherogenic VLDL remnant and Lp(a) particles. The VLDL receptor can promote hepatic uptake of VLDL remnants via binding to apoE, and this shunt pathway can limit further remnant processing, resulting in reduced LDL production. Lp(a) has also been shown to be a ligand for the VLDL receptor, and thus hepatic VLDL receptor expression would be expected to increase plasma clearance and reduce concentrations of Lp(a) particles. VLDL: very low-density lipoprotein. LDL: low-density lipoprotein. Lp(a): lipoprotein a. apoC: apolipoprotein C. apoB-100: apolipoprotein B-100. apoE: apolipoprotein E.

First, as reviewed by Rader, there are pitfalls to AAV-mediated gene therapy including a lack of long term expression and safe viral delivery vector systems [80]. Other challenges for use of such a system include: 1) the immunogenicity of the AAV capsid that limits the amount of viral particles dosed; 2) an inability to repeat and titrate treatment to achieve optimal levels of lipids and lipoproteins; and 3) waning durability and clinical efficacy since the AAV transduced gene exists in hepatocytes as an episome, and gene expression declines as the hepatocytes undergo division [81].

Second, previous studies reported that the overexpression of ectopic *VLDLR* in hepatocytes may be pro-inflammatory; thus increasing ER stress and the potential for hepatic steatosis [82]. Additionally, increased *VLDLR* expression in macrophages was found to promote adipose tissue inflammation and impaired glucose tolerance in obese mice [83]. In another study, it has been reported that *VLDLR* expression was a factor in adipose tissue inflammation, where this inflammation was only reduced in obese *VLDLR*-deficient mice fed a high-fat diet [84]. *VLDLR* has also been found to modulate fibrin-dependent leukocyte transmigration and thereby promote inflammation [85,86].

However, several studies have identified mechanisms for reducing the risk of *VLDLR*-attributed inflammation and hepatic steatosis. For example, Fuchs et al. discovered that a lack of adipose triglyceride lipase protected mice from endoplasmic reticulum (ER) and hepatic stress in the presence of high TG levels [87]. Zarei et al. observed that FGF21 may protect against hepatic steatosis by attenuating ER stress-induced *VLDLR* upregulation [88]. Recent results with *VLDLR* knockout mice have also indicated that *VLDLR* is not a major factor in fatty liver formation, particularly during protein restriction [89]. Thus, although there is a potential risk of hepatic steatosis resulting from excessive TG uptake in hypertriglyceridemic states [90], it is possible that compensatory mechanisms, such as increased fatty acid  $\beta$ -oxidation or reduced hepatic lipogenesis, may act to mitigate hepatic TG accumulation.

## 5. NOVEL GENETIC MEDICINE APPROACHES TOWARD TARGETING *VLDLR*

Despite the previous hurdles discussed above, gene therapy targeting hepatic *VLDLR* expression remains a promising modality to effectively treat and manage atherogenic lipid disorders. Thus, upcoming genetic medicine approaches for achieving sustained hepatic expression of *VLDLR* may benefit from incorporating the following features:

### 5.1. Integrative technology to improve durability of treatment effect

For atherogenic lipid disorders, the lack of durable treatment effects observed with AAV gene therapy in dividing hepatocytes can often be circumvented by using gene integrating or editing technology. However, the main potential safety risks with these methods are related to the site of gene integration. For example, some gene editing methods with CRISPR/Cas9 involve the introduction of double-stranded breaks (DSBs). These DSBs cause extensive genomic rearrangements (chromothripsis) that can drive the rapid acquisition of multiple cancer-causing mutations simultaneously. Chromothripsis can promote tumorigenesis in many tissue types, including ones relevant for therapeutic editing [91,92]. Although some gene editing systems that utilize base editors or prime editors do not introduce DSBs [93–95], these systems are limited in their degree of DNA editing to small genomic regions. The immune response induced by Cas9 protein itself can also be a problem. It is possible that this is due to the presence of certain peptides in Cas9 that may act as MHC-binding epitopes. It should be remembered that Cas9 is a protein of bacterial origin and can have an immunogenic effect in mammals [96].

New generations of gene integration methodology may be able to avoid these concerns and limitations. For example, a transposon-based system that allows genetic material to be transferred to a specific site in a host organism's chromosome may represent a viable methodology for durable and safe integrating gene therapy [97,98].

Currently available non-mammalian transposon systems include fish-derived Sleeping Beauty [79,99,100] and insect-derived PiggyBac [101–103], with newer ones on the horizon. A potential drawback for such methods is that the DNA recognition sequence may be found throughout the human genome and thus the gene would not be targeted to a specific site. This is particularly problematic for the Sleeping Beauty transposon system because the recognition sequence consists of a short dinucleotide sequence (TA), and more recent studies have uncovered the ability of Sleeping Beauty transposon to integrate into non-TA dinucleotides under certain conditions [104,105]. An ideal transposon system would be one that is mammal-derived without an immunogenic effect with the capability to insert genetic material of unlimited size at a site-specific genomic target.

### 5.2. VLDLR liver-specific targeting via LNP-based delivery

Targeted delivery to the liver is crucial to the success of ectopic VLDLR hepatic expression. While AAV vectors exhibit tropism to specific cell types or tissues by using different capsid proteins, their specificity is not absolute [81,106]. Although lipid nanoparticles (LNPs) lack the tissue tropism of AAV vectors [107–109], their preferential accumulation in the liver, through an apoE-dependent process [110], is ideal for delivering VLDLR to the liver for the treatment of hyperlipidemia. To further enhance the specificity of this targeting, hepatocyte-specific ligands such as N-acetyl-D-galactosamine (GalNAc) can be incorporated into the LNPs to target hepatocyte receptors [111]. In addition, LNP-based delivery systems theoretically offer unlimited packaging capacity [112–114], while viral vectors have a cargo size limitation (4.7 kB for AAVs) [115]. Moreover, non-viral vectors—such as cationic lipids and LNPs—are potentially safer than viral vectors due to the absence of immunogenic viral proteins.

## 6. CONCLUSION

Hepatic *VLDLR* expression as a therapeutic target is highly promising, exhibiting many features that make it amenable to genetic therapies aimed at reducing levels of TG and Lp(a). Typically, gene therapy is used to correct a genetic defect, where a gene is either not functioning or is overly functional. However, hepatic VLDLR targeting instead represents a genetic medicine approach for lipoprotein lowering therapy that would replace or augment traditional pharmacological (small molecule) drugs that primarily treat the effects of a disordered system or disease state. While further studies will be required to assess the clinical efficacy and potential adverse effects of such a therapeutic agent in humans, its successful development and implementation could present an important opportunity to lower pathologic lipoprotein levels and reduce CVD risk.

## FUNDING

This work was supported by SalioGen Therapeutics.

## CREDIT AUTHOR STATEMENT

**RM Krauss:** Writing- Original draft preparation, Writing- Reviewing and Editing. **JT Lu:** Writing- Original draft preparation, Writing- Reviewing and Editing. **CM Clary:** Writing- Reviewing and Editing. **JJ Higgins:** Conceptualization, Writing- Reviewing and Editing. **R Tabibiazar:** Conceptualization, Writing- Reviewing and Editing. All authors approved the final version of the manuscript.

## DATA AVAILABILITY

No data was used for the research described in the article.

## ACKNOWLEDGMENTS

RM Krauss and JT Lu drafted the manuscript. JJ Higgins and CM Clary revised the manuscript. R Tabibiazar provided consultation and expertise. The authors would also like to acknowledge Kathryn Verhoeven for editorial support during manuscript preparation.

## CONFLICT OF INTEREST

RM Krauss is a scientific advisor to SalioGen Therapeutics, Virta Health Corp., Day Two, and Seraphina Therapeutics. JT Lu, JJ Higgins, CM Clary, and R Tabibiazar are full-time employees of SalioGen Therapeutics.

## REFERENCES

- [1] Tóth PP, Potter D, Ming EE. Prevalence of lipid abnormalities in the United States: the National Health and Nutrition examination survey 2003-2006. *Journal of Clinical Lipidology* 2012;6:325–30.
- [2] Soppert J, Lehrke M, Marx N, Jankowski J, Noels H. Lipoproteins and lipids in cardiovascular disease: from mechanistic insights to therapeutic targeting. *Adv Drug Deliv Rev* 2020;159:4–33.
- [3] Pedersen SB, Langsted A, Nordestgaard BG. Nonfasting mild-to-moderate hypertriglyceridemia and risk of acute pancreatitis. *JAMA Intern Med* 2016;176:1834–42.
- [4] Michos ED, McEvoy JW, Blumenthal RS. Lipid management for the prevention of atherosclerotic cardiovascular disease. *N Engl J Med* 2019;381:1557–67.
- [5] Silverman MG, Ference BA, Im K, Wiviott SD, Giugliano RP, Grundy SM, et al. Association between lowering LDL-C and cardiovascular risk reduction among different therapeutic interventions: a systematic review and meta-analysis. *J Am Med Assoc* 2016;316:1289–97.
- [6] Vallejo-Vaz AJ, Robertson M, Catapano AL, Watts GF, Kastelein JJ, Packard CJ, et al. Low-density lipoprotein cholesterol lowering for the primary prevention of cardiovascular disease among men with primary elevations of low-density lipoprotein cholesterol levels of 190 mg/dL or above: analyses from the WOSCOPS (west of scotland coronary prevention study) 5-year randomized trial and 20-year observational follow-up. *Circulation* 2017;136:1878–91.
- [7] Grundy SM, Stone NJ, Bailey AL, Beam C, Birtcher KK, Blumenthal RS, et al. 2018 AHA/ACC/AACVPR/AAPA/ABC/ACPM/ADA/AGS/APhA/ASPC/NLA/PCNA guideline on the management of blood cholesterol. *J Am Coll Cardiol* 2019;73:e285–350.
- [8] Ginsberg HN, Packard CJ, Chapman MJ, Borén J, Aguilar-Salinas CA, Averna M, et al. Triglyceride-rich lipoproteins and their remnants: metabolic insights, role in atherosclerotic cardiovascular disease, and emerging therapeutic strategies—a consensus statement from the European Atherosclerosis Society. *Eur Heart J* 2021;42:4791–806.
- [9] Krauss RM, King SM. Remnant lipoprotein particles and cardiovascular disease risk. *Best Pract Res Clin Endocrinol Metabol* 2022;101682. <https://doi.org/10.1016/j.beem.2022.101682> [Pre-Print].
- [10] Gill PK, Dron JS, Hegele RA. Genetics of hypertriglyceridemia and atherosclerosis. *Curr Opin Cardiol* 2021;36:264–71.
- [11] Hernandez P, Passi N, Modarressi T, Kulkarni V, Soni M, Burke F, et al. Clinical management of hypertriglyceridemia in the prevention of cardiovascular disease and pancreatitis. *Curr Atherosclerosis Rep* 2021;23:72.

## Review

- [12] Ballantyne CM, Olsson AG, Cook TJ, Mercuri MF, Pedersen TR, Kjekshus J. Influence of low high-density lipoprotein cholesterol and elevated triglyceride on coronary heart disease events and response to simvastatin therapy in 4S. *Circulation* 2001;104:3046–51.
- [13] Miller M, Cannon CP, Murphy SA, Qin J, Ray KK, Braunwald E. Impact of triglyceride levels beyond low-density lipoprotein cholesterol after acute coronary syndrome in the PROVE IT-TIMI 22 trial. *J Am Coll Cardiol* 2008;51:724–30.
- [14] Ginsberg HN, Elam MB, Lovato LC, Crouse 3rd JR, Leiter LA, Linz P, et al. Effects of combination lipid therapy in type 2 diabetes mellitus. *N Engl J Med* 2010;362:1563–74.
- [15] Marston NA, Giugliano RP, Im K, Silverman MG, O'Donoghue ML, Wiviott SD, et al. Association between triglyceride lowering and reduction of cardiovascular risk across multiple lipid-lowering therapeutic classes. *Circulation* 2019;140:1308–17.
- [16] Kim NH, Kim SG. Fibrates revisited: potential role in cardiovascular risk reduction. *Diabetes Metab J* 2020;44:213–21.
- [17] Das Pradhan A, Glynn RJ, Fruchart J-C, MacFadyen JG, Zaharris ES, Everett BM, et al. Triglyceride lowering with pefmifibate to reduce cardiovascular risk. *N Engl J Med* 2022;387:1923–34.
- [18] Virani SS. The fibrates story - a tepid end to a PROMINENT drug. *N Engl J Med* 2022;387:1991–2.
- [19] Gao Y, Shen W, Lu B, Zhang Q, Hu Y, Chen Y. Upregulation of hepatic VLDLR via PPAR $\alpha$  is required for the triglyceride-lowering effect of fenofibrate. *JL R (J Lipid Res)* 2014;55:1622–33.
- [20] Sherratt SCR, Libby P, Bhatt DL, Mason RP. A biological rationale for the disparate effects of omega-3 fatty acids on cardiovascular disease outcomes. *Prostagl Leukot Essent Fat Acids* 2022;182:102450.
- [21] Yokoyama M, Origasa H, Matsuzaki M, Matsuzawa Y, Saito Y, Ishikawa Y, et al. Effects of eicosapentaenoic acid on major coronary events in hypercholesterolaemic patients (JELIS): a randomised open-label, blinded endpoint analysis. *Lancet* 2007;369:1090–8.
- [22] Nicholls SJ, Lincoff AM, Garcia M, Bash D, Ballantyne CM, Barter PJ, et al. Effect of high-dose omega-3 fatty acids vs corn oil on major adverse cardiovascular events in patients at high cardiovascular risk: the STRENGTH randomized clinical trial. *J Am Med Assoc* 2020;324:2268–80.
- [23] Gaba P, Bhatt DL, Steg PG, Miller M, Brinton EA, Jacobson TA, et al. Prevention of cardiovascular events and mortality with icosapent ethyl in patients with prior myocardial infarction. *J Am Coll Cardiol* 2022;79:1660–71.
- [24] Bhatt DL, Steg PG, Miller M, Brinton EA, Jacobson TA, Ketchum SB, et al. Cardiovascular risk reduction with icosapent ethyl for hypertriglyceridemia. *N Engl J Med* 2019;380:11–22.
- [25] Doi T, Langsted A, Nordestgaard BG. A possible explanation for the contrasting results of REDUCE-IT vs. STRENGTH: cohort study mimicking trial designs. *Eur Heart J* 2021;42:4807–17.
- [26] Steg PG, Bhatt DL. The reduction in cardiovascular risk in REDUCE-IT is due to eicosapentaenoic acid in icosapent ethyl. *Eur Heart J* 2021;42:4865–6.
- [27] Doi T, Langsted A, Nordestgaard BG. Mineral oil and icosapent ethyl may jointly explain the between arm difference of cardiovascular risk in REDUCE-IT. *Eur Heart J* 2021;42:4867–8.
- [28] Mason RP, Eckel RH. Mechanistic insights from REDUCE-IT STRENGTHen the case against triglyceride lowering as a strategy for cardiovascular disease risk reduction. *Am J Med* 2021;134:1085–90.
- [29] Nurmohamed NS, Dallinga-Thie GM, Stroes ESG. Targeting apoC-III and ANGPTL3 in the treatment of hypertriglyceridemia. *Expert Rev Cardiovasc Ther* 2020;18:355–61.
- [30] Mohamed F, Botha TC, Raal FJ. Inhibition of angiopoietin-like 3 for the management of severe hypercholesterolemia. *Curr Opin Lipidol* 2021;32:213–8.
- [31] Gaudet D, Alexander VJ, Baker BF, Brisson D, Tremblay K, Singleton W, et al. Antisense inhibition of apolipoprotein C-III in patients with hypertriglyceridemia. *N Engl J Med* 2015;373:438–47.
- [32] Gaudet D, Brisson D, Tremblay K, Alexander VJ, Singleton W, Hughes SG, et al. Targeting APOC3 in the familial chylomicronemia syndrome. *N Engl J Med* 2014;371:2200–6.
- [33] Alexander VJ, Xia S, Hurh E, Hughes SG, O'Dea L, Geary RS, et al. N-acetyl galactosamine-conjugated antisense drug to APOC3 mRNA, triglycerides and atherogenic lipoprotein levels. *Eur Heart J* 2019;40:2785–96.
- [34] Butler AA, Price CA, Graham JL, Stanhope KL, King S, Hung YH, et al. Fructose-induced hypertriglyceridemia in rhesus macaques is attenuated with fish oil or ApoC3 RNA interference. *JLR (J Lipid Res)* 2019;60:805–18.
- [35] Raposeiras-Roubin S, Rosselló X, Oliva B, Fernández-Friera L, Mendiguren JM, Andrés V, et al. Triglycerides and residual atherosclerotic risk. *J Am Coll Cardiol* 2021;77:3031–41.
- [36] Khera AV, Everett BM, Caulfield MP, Hantash FM, Wohlgemuth J, Ridker PM, et al. Lipoprotein(a) concentrations, rosuvastatin therapy, and residual vascular risk: an analysis from the JUPITER trial (justification for the use of statins in prevention: an intervention trial evaluating rosuvastatin). *Circulation* 2014;129:635–42.
- [37] Sahebkar A, Reiner Ž, Simental-Mendía LE, Ferretti G, Cicero AF. Effect of extended-release niacin on plasma lipoprotein(a) levels: a systematic review and meta-analysis of randomized placebo-controlled trials. *Metabolism* 2016;65:1664–78.
- [38] O'Donoghue ML, Fazio S, Giugliano RP, Stroes ESG, Kanevsky E, Gouni-Berthold I, et al. Lipoprotein(a), PCSK9 inhibition, and cardiovascular risk. *Circulation* 2019;139:1483–92.
- [39] Korneva VA, Kuznetsova TY, Julius U. Modern approaches to lower lipoprotein(a) concentrations and consequences for cardiovascular diseases. *Biomedicines* 2021;9.
- [40] Iannuzzo G, Tripaldella M, Mallardo V, Morgillo M, Vitelli N, Iannuzzi A, et al. Lipoprotein(a) where do we stand? From the physiopathology to innovative therapy. *Biomedicines* 2021;9.
- [41] Greco MF, Sirtori CR, Corsini A, Ezhev M, Sampietro T, Ruscica M. Lipoprotein(a) lowering-from lipoprotein apheresis to antisense oligonucleotide approach. *J Clin Med* 2020;9.
- [42] Swerdlow DI, Rider DA, Yavari A, Wikström Lindholm M, Campion GV, Nissen SE. Treatment and prevention of lipoprotein(a)-mediated cardiovascular disease: the emerging potential of RNA interference therapeutics. *Cardiovasc Res* 2021;118:1218–31.
- [43] Mazhar F, Hjemdahl P, Clase CM, Johnell K, Jernberg T, Sjölander A, et al. Intensity of and adherence to lipid-lowering therapy as predictors of major adverse cardiovascular outcomes in patients with coronary heart disease. *J Am Heart Assoc* 2022;11:e025813.
- [44] Waßmuth S, Rohe K, Noack F, Noutsias M, Treede H, Schlitt A. Adherence to lipid-lowering therapy in patients with coronary heart disease from the state of saxony-anhalt, Germany. *Vasc Health Risk Manag* 2019;15:477–83.
- [45] Tiebel O, Oka K, Robinson K, Sullivan M, Martinez J, Nakamura M, et al. Mouse very low-density lipoprotein receptor (VLDLR): gene structure, tissue-specific expression and dietary and developmental regulation. *Atherosclerosis* 1999;145:239–51.
- [46] Sakai J, Hoshino A, Takahashi S, Miura Y, Ishii H, Suzuki H, et al. Structure, chromosome location, and expression of the human very low density lipoprotein receptor gene. *J Biol Chem* 1994;269:2173–82.
- [47] Gene [Internet]. Bethesda (MD): National Library of Medicine (US), National Center for Biotechnology Information; 2004. -2022 [cited 2022 09 10], Available from: <https://www.ncbi.nlm.nih.gov/gene/7436>.
- [48] Wyne KL, Pathak K, Seabra MC, Hobbs HH. Expression of the VLDL receptor in endothelial cells. *Arterioscler Thromb Vasc Biol* 1996;16:407–15.

- [49] Iwasaki T, Takahashi S, Takahashi M, Zenimaru Y, Kujiraoka T, Ishihara M, et al. Deficiency of the very low-density lipoprotein (VLDL) receptors in streptozotocin-induced diabetic rats: insulin dependency of the VLDL receptor. *Endocrinology* 2005;146:3286–94.
- [50] Schneider WJ. Lipoprotein receptors. In: Ridgway ND, McLeod RS, editors. *Biochemistry of lipids, lipoproteins and membranes*. ed. Boston: Elsevier; 2016. p. 489–518.
- [51] Dlugosz P, Nimpf J. The Reelin receptors apolipoprotein E receptor 2 (ApoER2) and VLDL receptor. *Int J Mol Sci* 2018;19.
- [52] Feldman M. Liver physiology and energy metabolism. In: Roy-Chowdhury N, Roy-Chowdhury J, editors. *Sleisenger and Fordtran's gastrointestinal and liver disease*. Boston: Elsevier; 2020. p. 1135–53.
- [53] Goudriaan JR, Espiritu Santo SM, Voshol PJ, Teusink B, van Dijk KW, van Vlijmen BJ, et al. The VLDL receptor plays a major role in chylomicron metabolism by enhancing LPL-mediated triglyceride hydrolysis. *JLR (J Lipid Res)* 2004;45:1475–81.
- [54] Ruiz J, Kouiavskaya D, Migliorini M, Robinson S, Saenko EL, Gorlatova N, et al. The apoE isoform binding properties of the VLDL receptor reveal marked differences from LRP and the LDL receptor. *JLR (J Lipid Res)* 2005;46:1721–31.
- [55] Mahley RW. Apolipoprotein E: from cardiovascular disease to neurodegenerative disorders. *J Mol Med* 2016;94:739–46.
- [56] Dergunov AD, Novoselov AV, Visvikis S, Siest G, Yakushkin WV, Tsibulsky V. The composition, structural properties and binding of very-low-density and low-density lipoproteins to the LDL receptor in normo- and hypertriglyceridemia: relation to the apolipoprotein E phenotype. *Biol Chem* 2005;386:441–52.
- [57] Yu Y, Kuang YL, Lei D, Zhai X, Zhang M, Krauss RM, et al. Polyhedral 3D structure of human plasma very low density lipoproteins by individual particle cryo-electron tomography1. *JLR (J Lipid Res)* 2016;57:1879–88.
- [58] Ruby MA, Goldenson B, Orasanu G, Johnston TP, Plutzky J, Krauss RM. VLDL hydrolysis by LPL activates PPAR-alpha through generation of unbound fatty acids. *JLR (J Lipid Res)* 2010;51:2275–81.
- [59] Ramms B, Patel S, Nora C, Pessentheiner AR, Chang MW, Green CR, et al. ApoC-III ASO promotes tissue LPL activity in the absence of apoE-mediated TRL clearance. *JLR (J Lipid Res)* 2019;60:1379–95.
- [60] Heidemann BE, Koopal C, Bots ML, Asselbergs FW, Westerink J, Visseren FLJ. The relation between VLDL-cholesterol and risk of cardiovascular events in patients with manifest cardiovascular disease. *Int J Cardiol* 2021;322:251–7.
- [61] Castillo-Núñez Y, Morales-Villegas E, Aguilar-Salinas CA. Triglyceride-rich lipoproteins: their role in atherosclerosis. *Rev Investig Clin* 2022;74:61–70.
- [62] Packard CJ, Boren J, Taskinen M-R. Causes and consequences of hypertriglyceridemia. *Front Endocrinol* 2020;11:252.
- [63] Chait A, Subramanian S. Hypertriglyceridemia: pathophysiology, role of genetics, consequences, and treatment. In: Feingold KR, Anawalt B, Boyce A, Chrousos G, de Herder WW, Dhatariya K, et al., editors. *Endotext [Internet]*. South Dartmouth (MA): MDText.com, Inc.; 2000–2022.
- [64] de Pretis N, Amadio A, Frulloni L. Hypertriglyceridemic pancreatitis: epidemiology, pathophysiology and clinical management. *United Euro Gastroenterol J* 2018;6:649–55.
- [65] Schmidt K, Noureen A, Kronenberg F, Utermann G. Structure, function, and genetics of lipoprotein (a). *JLR (J Lipid Res)* 2016;57:1339–59.
- [66] Kronenberg F. Lipoprotein(a). In: von Eckardstein A, Binder CJ, editors. *Prevention and treatment of atherosclerosis: improving state-of-the-art management and search for novel targets*. Cham: Springer; 2022. p. 201–32 (CH).
- [67] Dentali F, Gessi V, Marcucci R, Gianni M, Grandi AM, Franchini M. Lipoprotein(a) as a risk factor for venous thromboembolism: a systematic review and meta-analysis of the literature. *Semin Thromb Hemost* 2017;43:614–20.
- [68] Ugovšek S, Šebestjan M. Lipoprotein(a)-The crossroads of atherosclerosis, atherothrombosis and inflammation. *Biomolecules* 2021;12:26–34.
- [69] Orsó E, Schmitz G. Lipoprotein(a) and its role in inflammation, atherosclerosis and malignancies. *Clin Res Cardiol Suppl* 2017;12:31–7.
- [70] Kronenberg F. Human genetics and the causal role of lipoprotein(a) for various diseases. *Cardiovasc Drugs Ther* 2016;30:87–100.
- [71] Vavuranakis MA, Jones SR, Cardoso R, Gerstenblith G, Leucker TM. The role of Lipoprotein(a) in cardiovascular disease: current concepts and future perspectives. *Hellenic J Cardiol* 2020;61:398–403.
- [72] Varvel S, McConnell JP, Tsimikas S. Prevalence of elevated Ip(a) mass levels and patient thresholds in 532 359 patients in the United States. *Arterioscler Thromb Vasc Biol* 2016;36:2239–45.
- [73] McCormick SP. Lipoprotein(a): biology and clinical importance. *Clin Biochem Rev* 2004;25:69–80.
- [74] Takahashi S. Triglyceride rich lipoprotein -LPL-VLDL receptor and Ip(a)-VLDL receptor pathways for macrophage foam cell formation. *J Atherosclerosis Thromb* 2017;24:552–9.
- [75] Kosaka S, Takahashi S, Masamura K, Kanehara H, Sakai J, Tohda G, et al. Evidence of macrophage foam cell formation by very low-density lipoprotein receptor: interferon-gamma inhibition of very low-density lipoprotein receptor expression and foam cell formation in macrophages. *Circulation* 2001;103:1142–7.
- [76] Kobayashi K, Oka K, Forte T, Ishida B, Teng B, Ishimura-Oka K, et al. Reversal of hypercholesterolemia in low density lipoprotein receptor knockout mice by adenovirus-mediated gene transfer of the very low density lipoprotein receptor. *J Biol Chem* 1996;271:6852–60.
- [77] Kozarsky KF, Jooss K, Donahee M, Strauss 3rd JF, Wilson JM. Effective treatment of familial hypercholesterolemia in the mouse model using adenovirus-mediated transfer of the VLDL receptor gene. *Nat Genet* 1996;13:54–62.
- [78] Oka K, Pastore L, Kim IH, Merched A, Nomura S, Lee HJ, et al. Long-term stable correction of low-density lipoprotein receptor-deficient mice with a helper-dependent adenoviral vector expressing the very low-density lipoprotein receptor. *Circulation* 2001;103:1274–81.
- [79] Turunen TA, Kurkipuro J, Heikura T, Vuorio T, Hytonen E, Izsvak Z, et al. Sleeping beauty transposon vectors in liver-directed gene delivery of LDLR and VLDLR for gene therapy of familial hypercholesterolemia. *Mol Ther* 2016;24:620–35.
- [80] Rader DJ. Gene therapy for familial hypercholesterolemia. *Nutr Metabol Cardiovasc Dis* 2001;11(Suppl 5):40–4.
- [81] Colella P, Ronzitti G, Mingozzi F. Emerging issues in AAV-mediated in vivo gene therapy. *Mol Ther Methods Clin Dev* 2018;8:87–104.
- [82] Jo H, Choe SS, Shin KC, Jang H, Lee JH, Seong JK, et al. Endoplasmic reticulum stress induces hepatic steatosis via increased expression of the hepatic very low-density lipoprotein receptor. *Hepatology* 2013;57:1366–77.
- [83] Shin KC, Hwang I, Choe SS, Park J, Ji Y, Kim JL, et al. Macrophage VLDLR mediates obesity-induced insulin resistance with adipose tissue inflammation. *Nat Commun* 2017;8:1087.
- [84] Nguyen A, Tao H, Metrione M, Hajri T. Very low density lipoprotein receptor (VLDLR) expression is a determinant factor in adipose tissue inflammation and adipocyte-macrophage interaction. *J Biol Chem* 2014;289:1688–703.
- [85] Yakovlev S, Mikhailenko I, Cao C, Zhang L, Strickland DK, Medved L. Identification of VLDLR as a novel endothelial cell receptor for fibrin that modulates fibrin-dependent transendothelial migration of leukocytes. *Blood* 2012;119:637–44.
- [86] Yakovlev S, Medved L. Dual functions of the fibrin  $\beta$ N-domains in the VLDL receptor-dependent pathway of transendothelial migration of leukocytes. *Thromb Res* 2022;214:1–7.
- [87] Fuchs CD, Claudel T, Kumari P, Haemmerle G, Pollheimer MJ, Stojakovic T, et al. Absence of adipose triglyceride lipase protects from hepatic endoplasmic reticulum stress in mice. *Hepatology* 2012;56:270–80.

## Review

- [88] Zarei M, Barroso E, Palmer X, Dai J, Rada P, Quesada-Lopez T, et al. Hepatic regulation of VLDL receptor by PPARbeta/delta and FGF21 modulates non-alcoholic fatty liver disease. *Mol Metabol* 2018;8:117–31.
- [89] Oshio Y, Hattori Y, Kamata H, Ozaki-Masuzawa Y, Seki A, Tsuruta Y, et al. Very low-density lipoprotein receptor increases in a liver-specific manner due to protein deficiency but does not affect fatty liver in mice. *Sci Rep* 2021;11:8003.
- [90] Nassir F, Rector RS, Hammoud GM, Ibdah JA. Pathogenesis and prevention of hepatic steatosis. *Gastroenterol Hepatol* 2015;11:167–75.
- [91] Leibowitz ML, Papathanasiou S, Doerfler PA, Blaine LJ, Sun L, Yao Y, et al. Chromothripsis as an on-target consequence of CRISPR-Cas9 genome editing. *Nat Genet* 2021;53:895–905.
- [92] Leibowitz ML, Zhang CZ, Pellman D. Chromothripsis: a New mechanism for rapid karyotype evolution. *Annu Rev Genet* 2015;49:183–211.
- [93] Komor AC, Badran AH, Liu DR. Editing the genome without double-stranded DNA breaks. *ACS Chem Biol* 2018;13:383–8.
- [94] Eid A, Alshareef S, Mahfouz MM. CRISPR base editors: genome editing without double-stranded breaks. *Biochem J* 2018;475:1955–64.
- [95] Anzalone AV, Randolph PB, Davis JR, Sousa AA, Koblan LW, Levy JM, et al. Search-and-replace genome editing without double-strand breaks or donor DNA. *Nature* 2019;576:149–57.
- [96] Cheng X, Fan S, Wen C, Du X. CRISPR/Cas9 for cancer treatment: technology, clinical applications and challenges. *Brief Funct Genom* 2020;19:209–14.
- [97] Ivics Z, Li MA, Mátés L, Boeke JD, Nagy A, Bradley A, et al. Transposon-mediated genome manipulation in vertebrates. *Nat Methods* 2009;6:415–22.
- [98] Kleckner N, Roth J, Botstein D. Genetic engineering in Vivo using translocatable drug-resistance elements: New methods in bacterial genetics. *J Mol Biol* 1977;116:125–59.
- [99] Wilber A, Frandsen JL, Geurts JL, Largaespada DA, Hackett PB, McIvor RS. RNA as a source of transposase for Sleeping Beauty-mediated gene insertion and expression in somatic cells and tissues. *Mol Ther* 2006;13:625–30.
- [100] Ochmann MT, Ivics Z. Jumping ahead with sleeping beauty: mechanistic insights into cut-and-paste transposition. *Viruses* 2021;13.
- [101] Platt 2nd RN, Vandewege MW, Ray DA. Mammalian transposable elements and their impacts on genome evolution. *Chromosome Res* 2018;26:25–43.
- [102] Li X, Ewiss H, Hice RH, Malani N, Parker N, Zhou L, et al. A resurrected mammalian hAT transposable element and a closely related insect element are highly active in human cell culture. *Natl Acad Sci U S A* 2013;110:E478–87.
- [103] Wei M, Mi CL, Jing CQ, Wang TY. Progress of transposon vector system for production of recombinant therapeutic proteins in mammalian cells. *Front Bioeng Biotechnol* 2022;10:879222.
- [104] Zhou Y, Ma G, Yang J, Gao Z, Guo Y. The integration preference of sleeping beauty at non-TA site is related to the transposon end sequences. *Front Genet* 2021;12:639125.
- [105] Guo Y, Zhang Y, Hu K. Sleeping Beauty transposon integrates into non-TA dinucleotides. *Mobile DNA* 2018;9:8.
- [106] Sands MS. AAV-mediated liver-directed gene therapy. *Methods Mol Biol* 2011;807:141–57.
- [107] Meng N, Grimm D. Membrane-destabilizing ionizable phospholipids: novel components for organ-selective mRNA delivery and CRISPR–Cas gene editing. *Signal Transduct Targeted Ther* 2021;6:206.
- [108] Sago CD, Krupczak BR, Lokugamage MP, Gan Z, Dahlman JE. Cell subtypes within the liver microenvironment differentially interact with lipid nanoparticles. *Cell Mol Bioeng* 2019;12:389–97.
- [109] Kularatne RN, Crist RM, Stern ST. The future of tissue-targeted lipid nanoparticle-mediated Nucleic acid delivery. *Pharmaceuticals* 2022;15.
- [110] Sebastiani F, Yanez Arteta M, Lerche M, Porcar L, Lang C, Bragg RA, et al. Apolipoprotein E binding drives structural and compositional rearrangement of mRNA-containing lipid nanoparticles. *ACS Nano* 2021;15:6709–22.
- [111] Debacker AJ, Voutilä J, Catley M, Blakey D, Habib N. Delivery of oligonucleotides to the liver with GaINAC: from research to registered therapeutic drug. *Mol Ther* 2020;28:1759–71.
- [112] Zhao Y, Huang L. Lipid nanoparticles for gene delivery. *Adv Genetics* 2014;88:13–36.
- [113] Kulkarni JA, Cullis PR, van der Meel R. Lipid nanoparticles enabling gene therapies: from concepts to clinical utility. *Nucleic Acid Therapeut* 2018;28:146–57.
- [114] Li L, Hu S, Chen X. Non-viral delivery systems for CRISPR/Cas9-based genome editing: challenges and opportunities. *Biomaterials* 2018;171:207–18.
- [115] Maestro S, Weber ND, Zabaleta N, Aldabe R, Gonzalez-Aseguinolaza G. Novel vectors and approaches for gene therapy in liver diseases. *JHEP Rep* 2021;3:100300.