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## RESEARCH ARTICLE

*Trichomonas vaginalis* adherence phenotypes and extracellular vesicles impact parasite survival in a novel *in vivo* model of pathogenesis

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## Abstract

*Trichomonas vaginalis* is a human infective parasite responsible for trichomoniasis—the most common, non-viral, sexually transmitted infection worldwide. *T. vaginalis* resides exclusively in the urogenital tract of both men and women. In women, *T. vaginalis* has been found colonizing the cervix and vaginal tract while in men it has been identified in the upper and lower urogenital tract and in secreted fluids such as semen, urethral discharge, urine, and prostatic fluid. Despite the over 270 million cases of trichomoniasis annually worldwide, *T. vaginalis* continues to be a highly neglected organism and thus poorly studied. Here we have developed a male mouse model for studying *T. vaginalis* pathogenesis *in vivo* by delivering parasites into the murine urogenital tract (MUT) via transurethral catheterization. Parasite burden was assessed *ex-vivo* using a nanoluciferase-based gene expression assay which allowed quantification of parasites pre- and post-inoculation. Using this model and read-out approach, we show that *T. vaginalis* can be found within MUT tissue up to 72 hrs post-inoculation. Furthermore, we also demonstrate that parasites that exhibit increased parasite adherence *in vitro* also have higher parasite burden in mice *in vivo*. These data provide evidence that parasite adherence to host cells aids in parasite persistence *in vivo* and molecular determinants found to correlate with host cell adherence *in vitro* are applicable to infection *in vivo*. Finally, we show that co-inoculation of *T. vaginalis* extracellular vesicles (TvEVs) and parasites results in higher parasite burden *in vivo*. These findings confirm our previous *in vitro*-based predictions that TvEVs assist the parasite in colonizing the host. The establishment of this pathogenesis model for *T. vaginalis* sets the stage for identifying and examining parasite factors that contribute to and influence infection outcomes.

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## Author summary

*Trichomonas vaginalis* is a human infective parasite that is responsible for most common, non-viral, sexually transmitted infection worldwide. Despite the over 270 million cases of *T. vaginalis* infection reported annually worldwide, the parasite continues to be a highly neglected pathogen and thus poorly studied. Despite several attempts to establish an animal model to study *T. vaginalis* infection, reliable infection models are lacking. As a result, our understanding of parasitic features that contribute to establishing and maintaining an infection have been determined by growing cultures of parasites and human host cells in the laboratory and testing parasite: host cell interactions in test tubes. Here we describe the development of a mouse model for studying *T. vaginalis* infection in an animal. We demonstrate that two features of the parasite that affect parasite: host cell interaction in test tubes: (1) its adherence properties and (2) the extracellular vesicles it releases that are taken up by host cells also impact parasite survival in this novel, mice model. The establishment of this animal model for *T. vaginalis* sets the stage for identifying and examining parasite factors that contribute to and influence infection outcomes.

## Introduction

*Trichomonas vaginalis* (*Tv*) is an extracellular, eukaryotic protist and the causative agent of trichomoniasis—the most common, non-viral, sexually transmitted infection worldwide [1]. Worldwide prevalence of trichomoniasis surpasses that of gonorrhea, syphilis, and chlamydia combined with reports of roughly 156 million new cases worldwide and over 276 million cases annually [2–4]. Clinical manifestations of trichomoniasis range from inflammation of the male and female urogenital tracts to more severe complications such as infertility, an increased risk of human immunodeficiency virus (HIV) transmission and adverse pregnancy outcomes in women such as preterm delivery and low infant birth weight [5–12]. While it is assumed that the parasite can infect both sexes with equal frequency [13,14], the asymptomatic nature of infection in men coupled with the unreliable diagnostic testing for men has resulted in an underestimation in infection numbers [15–18]. As men are primarily asymptomatic, they can act as vectors for parasite transmission by passing *Tv* on to their partners [12–15,19]. Additionally, *Tv* has been associated with an increased risk of aggressive prostate cancer, the second leading cause of cancer death among men in the United States [20–24].

To establish infection, *Tv* adheres to the epithelial lining of the urogenital tract by transitioning from a free-swimming ovoid cell into its adherent amoeboid form [25,26]. It is thought that the level of parasite binding to host cells seen *in vitro* mimics the outcome of infection *in vivo* [27], but this has correlation has yet to be investigated. *Tv* adherence to the host occurs through the use of multiple adherence factors. While some of these factors have been identified, such as lipoglycans (LG) [28,29], BspA-like proteins [30], a rhomboid serine protease [31], a cadherin-like protein [32], and a host glycosaminoglycan-interacting protein [33], no single player can fully recapitulate maximal adherence. Therefore, this signifies that parasite adherence to the host epithelium is multifactorial, with additional players still unknown [34,35].

Despite numerous attempts to establish an *in vivo* vaginal mouse model for the study of trichomoniasis [36–40], no reliable model has been established. Variability in the murine vaginal microbiota [41], hormonal variations during the menstrual cycle of women and female mice, as well as differences in vaginal pH, are thought to contribute to the difficulty of establishing a robust *Tv* infection in female mice [39,41]. A male murine model for the study of

prostatitis caused by *Tv* has been reported using Wistar rats [42], suggesting that male mice might also be successfully infected by *Tv*. However, this study primarily focused on pathological changes and failed to provide a method for quantification of parasite burden. Furthermore, there is a smaller repertoire of genetically modified or knock-out rats compared to mice, which would hinder studying host:*Tv* interactions, and the use of rats as an animal model can prove to be more expensive long term.

Here we have established an *in vivo* *T. vaginalis* pathogenesis model using the male mouse urogenital tract (MUT). Using nanoluciferase as a readout for parasite burden, we determined that *Tv* can be found up to 72 hrs post-inoculation in MUT tissues. To our knowledge this is the first study which employs nanoluciferase as a readout for parasite burden in tissues *ex-vivo*. Using this model, we demonstrate that *Tv* strains which are more adherent to host cells *in vitro* result in higher parasite burden *in vivo*, providing the first direct evidence that parasite adherence to host cells affects the ability of the parasite to establish infection *in vivo*. Furthermore, we also demonstrate that the presence of *Tv* extracellular vesicles (EVs) results in higher parasite burden *in vivo*, and confirm previous *in vitro*-based predictions that *Tv*EVs assist the parasite in colonizing the host [43,44].

## Methods

### Ethics statement

Research was conducted under an Institutional Animal Care and Use Committee (IACUC) approved protocol in compliance with the Animal Welfare Act, Public Health Service (PHS) Policy on Humane Care and Use of Laboratory Animals, and other Federal statutes and regulations relating to animals and experiments involving animals. The facility where this research was conducted is accredited by the Association for Assessment and Accreditation of Laboratory Animal Care (AAALAC), International and adheres to principles stated in the Guide for the Care and Use of Laboratory Animals, National Research Council, 2011. Animals meeting pre-established criteria were humanely euthanized in accordance with American Veterinary Medical Association guidelines.

### Mice

BALB/cJ male mice were obtained from The Jackson Laboratory (Bar Harbor, ME) at 8 weeks of age. All procedures were reviewed and approved by the Institutional Animal Care and Use Committee of the University of California, Los Angeles Research Safety and Animal Welfare Administration (protocol ARC 2009–063).

### Parasites and media

*T. vaginalis* strains MSA 1103 and LSU 160 MA and P, derived from LSU 160 [33] were used. MSA 1103 and LSU 160 are clinical isolates from St. Paul Minnesota, USA and New Orleans, Louisiana USA, respectively, and have been described in Lustig et al. 2013 [27]. LSU 160 MA (more adherent) and LSU 160 P (parental) strains were derived from LSU 160 using a novel selection method to create isogenic parasites that differ in their ability to adhere to host cells, as we previously described [33]. Briefly, a clonal population of LSU 160 parasites was passaged for eight weeks by separating parasites that bound to the culture tubes (MA) or passaging without selection to tube-bound parasites (P) daily. The MA parasites were found to be approximately 6-fold more adherent to BPH-1 host cell monolayers, relative to the P parasites [33]. Parasite strain B7RC2, described in Lustig et al. 2013 [27], was used for the isolation of extracellular vesicles (see *Extracellular vesicle (EV) isolation* below). Parasites were cultured in

Diamond's modified Trypticase-yeast extract-maltose (TYM) medium supplemented with 10% horse serum (Sigma-Aldrich), 10 U/mL penicillin/10 µg/mL streptomycin (Gibco), 180 µM ferrous ammonium sulfate, and 28 µM sulfosalicylic acid [45,46] and were grown at 37°C and passaged daily. Overnight cultures of MSA 1103 and LSU 160 P were placed on ice for 10 min and vortexed for 30 sec prior to passaging to a concentration of  $5 \times 10^4$  cells/mL in 15 or 50 mL fresh completed media. LSU 160 MA strain was passaged as previously described [33]. Briefly, an overnight culture of LSU 160 MA was decanted to remove free-swimming parasites from the culture and replaced with fresh completed medium. The culture is then placed on ice for 10 min and passaged as described above.

### Nucleofections with the nanoluciferase expression construct

*T. vaginalis* parasites were nucleofected as previously described [33,47]. Briefly, 100 µg of the *T. vaginalis* specific plasmid, pMasterNeo [48], harboring the nanoluciferase (Nluc) gene [49] was nucleofected into MSA 1103 and LSU 160 parasites using T-cell buffer (Lonza) + D-023 program or V-kit buffer (Lonza) + X-001 program, respectively. Recovered parasites were selected with 100 µg/mL G418 (Invitrogen) 24 hrs post-nucleofection. Presence of the Nluc plasmid was confirmed via nanoluciferase activity assay described below.

### Mouse inoculations using transurethral catheterization

All experiments were performed on 10-14-week-old BALB/c male mice. Transurethral inoculation of the mouse MUT tissue was carried out as previously described [50] with modifications. Briefly, fluid-deprived (4 hrs) mice were anesthetized with isoflurane and catheterized using a sterile polyethylene Intramedic PE-10 catheter (BD Biosciences) lubricated with Surgilube (HR Pharmaceuticals Inc.).  $1 \times 10^8$  Nluc-expressing parasites resuspended in 100 µL RPMI-1640 (+ L-glutamine, + HEPES; Invitrogen) supplemented with 10 U/mL penicillin/10 µg/mL streptomycin, and 10% fetal bovine serum without G418 to avoid any toxicity G418 may have on the recipient mice and injected into the MUT. Mice were kept anesthetized for an additional 30 min post-inoculation to increase the chance of colonization and minimize parasite loss through urination. Infected animals were sacrificed at the indicated times post-inoculation. The MUT tissue (prostate glands, bladder, urethra, and seminal vesicles) were excised and processed for parasite quantification via luciferase activity, protein concentration, or tissue histology as described below.

### Tissue histology

Two mice were euthanized immediately after inoculation with  $5 \times 10^7$  *T. vaginalis* MSA 1103 parasites and the harvested MUT tissue was fixed in 10% formalin overnight at room temperature. The tissues were then rinsed for 15 min under running water and stored in 70% ethanol. Fixed tissues were embedded in paraffin, serially sectioned at a width of 5 µm, and stained with Hematoxylin & Eosin (H&E) staining. Samples were imaged using a BX43 light microscope (Olympus) and Aperio ScanScope AT Turbo Whole Slide Scanning System (Leica Biosystems) and analyzed using the Aperio ImageScope software (Leica Biosystems).

### Tissue processing

Infected mice were euthanized at respective time points and the MUT tissue was excised and placed in pre-weighed tubes containing 5 mL 1x PBS (-CaCl<sub>2</sub>, -MgCl<sub>2</sub>) and placed on ice until ready for further processing. Tubes + MUT were weighed to determine the weight of the tissue and the tissue was then minced and resuspended in 1x MUT lysis buffer (50 mM Tris-HCl, 2

mM EDTA, 0.1% Triton X-100, pH 8 + 1x HALT protease inhibitor (Thermo Fisher)) at a volume of 1 mL for every 0.5 g tissue. Lysates were then sonicated using a Sonic Dismembrator F60 (Fisher Scientific) probe sonicator on ice at 60% power for 5 cycles, 0.5 min on/off, set at setting 12 which was followed by centrifugation at 10,000 rpm at 4°C for 10 min to pellet insoluble material. Supernatant was collected and used for Nluc or Bradford assays further described below.

### Nanoluciferase activity assay

The linear dynamic range of the NanoGlo Luciferase Assay System (Promega) in *T. vaginalis* parasites was determined per the manufacturer's instructions. Briefly, the NanoGlo reagent was prepared by mixing the NanoGlo substrate in NanoGlo buffer at a ratio of 1:50. Serial dilutions of *T. vaginalis*-Nluc parasites were prepared (ranging from 0 parasites/100  $\mu$ L to  $10^6$  parasites/100  $\mu$ L) by diluting parasites in 1x MUT lysis buffer. 100  $\mu$ L of protein dilutions were added to the wells of a 96-well, round bottom white plate (Corning) in triplicate wells and mixed with 100  $\mu$ L NanoGlo reagent per well. Luminescence was read using a Synergy H1 Hybrid Multi Mode (BioTek) plate reader at 460 nm. Data was analyzed using Gen5 Microplate Reader and Imager software (BioTek) and Microsoft Excel.

Measuring luminescence of mouse MUT tissue inoculated with Nluc-expressing *T. vaginalis* parasites was carried out similarly with one minor difference. 100  $\mu$ L of the processed MUT tissue supernatant was added to the wells of a 96-well, round bottom white plate in triplicate and mixed with 100  $\mu$ L NanoGlo reagent. The samples were read and analyzed as described above.

### Bradford assay

To quantify protein concentration of the infected MUT tissue, processed supernatants were diluted 1:10 in 1x MUT lysis buffer and 5  $\mu$ L of the diluents were transferred to a 96-well, flat bottom clear plate (Corning) in triplicate wells. Samples were then mixed with 200  $\mu$ L 1x Bio-Rad Protein Assay (Bio-Rad) reagent. Absorbance was read using a Synergy H1 Hybrid Multi Mode plate reader at 595 nm. Absorbance values of serially diluted bovine serum albumin (BSA) done in triplicate was used to generate a standard curve. Data was analyzed using Gen5™ Microplate Reader and Imager software as well as Microsoft Excel.

### Extracellular vesicle (EV) isolation

Extracellular vesicles (EVs) were isolated from *T. vaginalis* strain B7RC2 as previously described [43,44] and resuspended in 100  $\mu$ L PBS+1x HALT protease inhibitor. After determining the concentration of EV protein using a Pierce BCA Kit (Thermo Scientific), EVs were stored at -80°C until ready for use.

### Co-infection with EVs and parasites

Mouse inoculations using transurethral catheterization were conducted as described above. Prior to inoculating the mice,  $1 \times 10^8$  MSA 1103 Nluc-expressing parasites were pre-incubated with 50  $\mu$ g/ $\mu$ L EVs for 30 min at RT. Mice were sacrificed at 0, 48 and 72 hrs post-infection and MUT tissue was processed and nanoluciferase activity assays were performed as described above. Parasite only controls were treated identically.

### Statistical analyses

Graphs were generated and statistical analyses performed using Prism7 (GraphPad) software. Two-tailed t test was used to determine significance for *in vivo* luminescence/ $\mu$ g protein.



Repeated measures one-way ANOVA was used to determine significance in Nluc signal levels of parasites grown in the absence of G418. One-way ANOVA with Tukey's multiple comparisons test was used to determine significance for *in vivo* luminescence/ $\mu\text{g}$  protein data and total parasite counts following recovery from mouse MUT tissue. Unpaired t-test with Welch's correction was used to determine significance in *in vivo* luminescence/ $\mu\text{g}$  protein data normalized to 0 hr control mice. Data are expressed as  $\pm$  standard deviation ( $\pm$  SD).

## Results

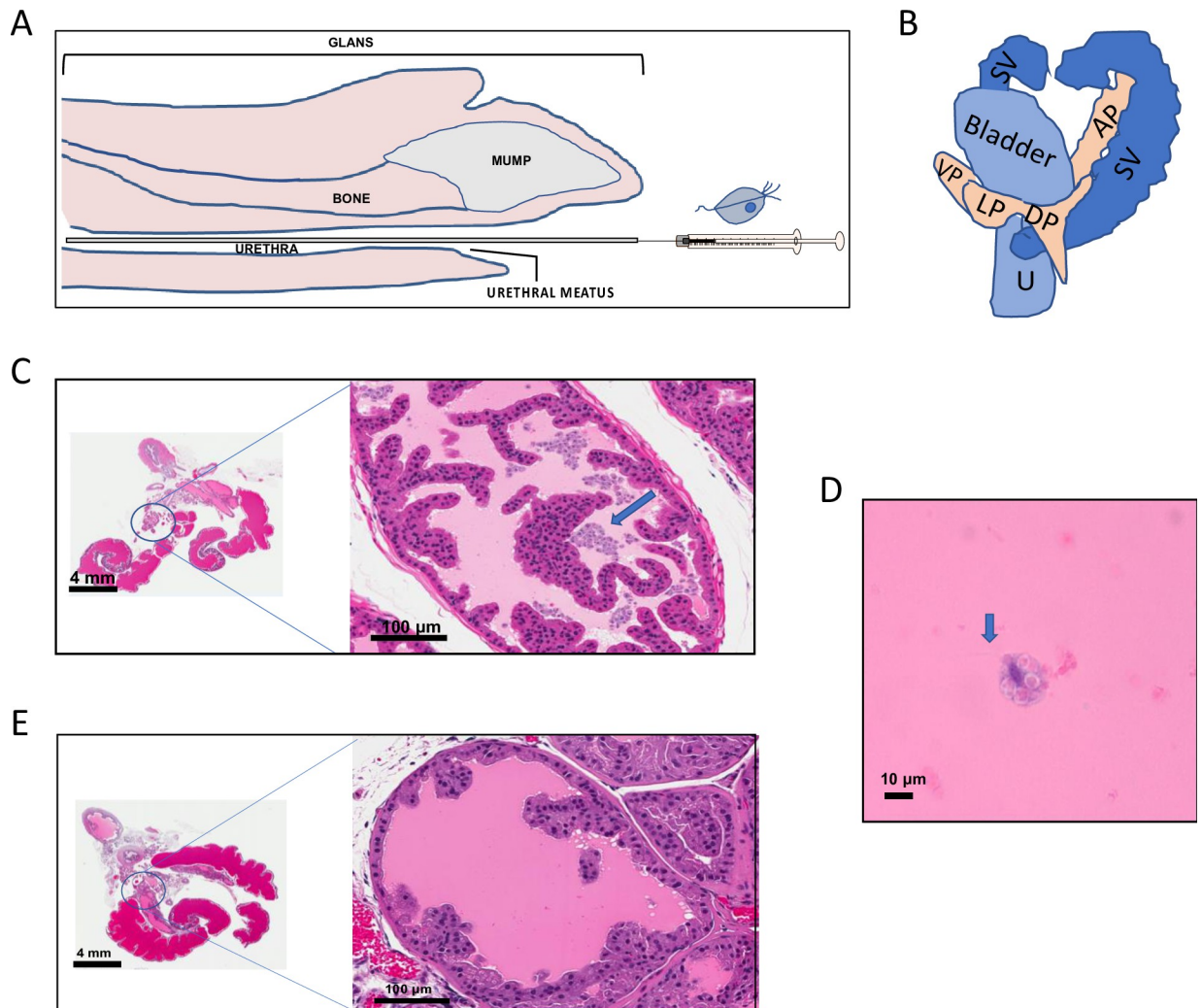
### Delivering *T. vaginalis* into the mouse male urogenital tract

*T. vaginalis* (*Tv*) has been found in the semen, urethral discharge, urine, and prostatic fluids of infected men [15,51]. In this study, we investigated whether the mouse male urogenital tract [MUT] can serve as a tractable *in vivo* model to study *Tv* pathogenesis. Transurethral catheterization was employed to deliver *Tv* to the mouse prostate as this approach has been used to successfully introduce *Escherichia coli* [50,52–54], *Propionibacterium acnes* [55], and *Chlamydia muridarum* [56] into the mouse MUT, as well as delivery of *Tv* to the rat prostate [42]. *Tv* is introduced into the mouse MUT via a syringe and catheter which is inserted past the urethral meatus and into the urethra (Fig 1A) which flows to the bladder, seminal vesicles, and 4 lobes of the mouse prostate (Fig 1B).

To confirm *Tv* delivery into the mouse MUT via transurethral catheterization, two mice were injected with *Tv* strain MSA 1103 parasites, immediately euthanized, and the MUT tissue excised. The tissues were then paraffin embedded and examined using Hematoxylin & Eosin (H&E) staining. Stained tissues were shown to contain parasites (Fig 1C), based on the presence of cells with a pear-shaped morphology and central/anterior elongated nuclei and anterior flagella (Fig 1D). Cells with this morphology were not seen in the mock vehicle control tissue (Fig 1E). A greater number of parasites were observed within the anterior lobes likely due to their larger size and anatomy, as opposed to a preference for this location.

### Nanoluciferase as Quantifiable measure of *Tv* burden in vivo

We initially examined whether available *Tv*-specific antibodies could be used to accurately quantify parasites using immunofluorescent detection with immunohistochemistry but found this to be an unreliable method due to high background levels. As a result, we developed a method which uses parasites expressing the nanoluciferase (Nluc) gene as a quantitative measure of parasite load in inoculated mouse MUT tissue. Nanoluciferase has been used to analyze infection and lifecycles *in vivo* for other eukaryotic pathogens [57,58] and has also been used in *Tv* for an *in vitro* proof-of-function approach [47]. To confirm Nluc expression and detectability as well as determine the linear range of the assay, a serial dilution of the Nluc-harboring *Tv* used in this study were assayed (Fig 2). Three variants of *Tv* strains were examined: the LSU 160 parental (P) and more adherent (MA) isogenic strains, which differ in their abilities to adhere to host cells [33], and the MSA 1132 strain. The LSU 160-P parasites displayed a notable difference in luminescence compared to the LSU 160-MA parasites (Fig 2A). The MSA 1103 parasites displayed a similar linear range to that of the LSU 160- P parasites, however the MSA 1103 maximum luminescence within the linear range was emitted by  $10^5$  parasites rather than  $10^6$  as seen with LSU 160-P (Fig 2B). To determine whether Nluc signals would diminish over time post-inoculation of parasites into the mouse MUT, in the absence of G418 selection of the Nluc plasmid, we grew parasites in the absence of G418 *in vitro* and measured Nluc luminescence over 72 hrs. No differences in luminescence signal were detected (S1 Data).

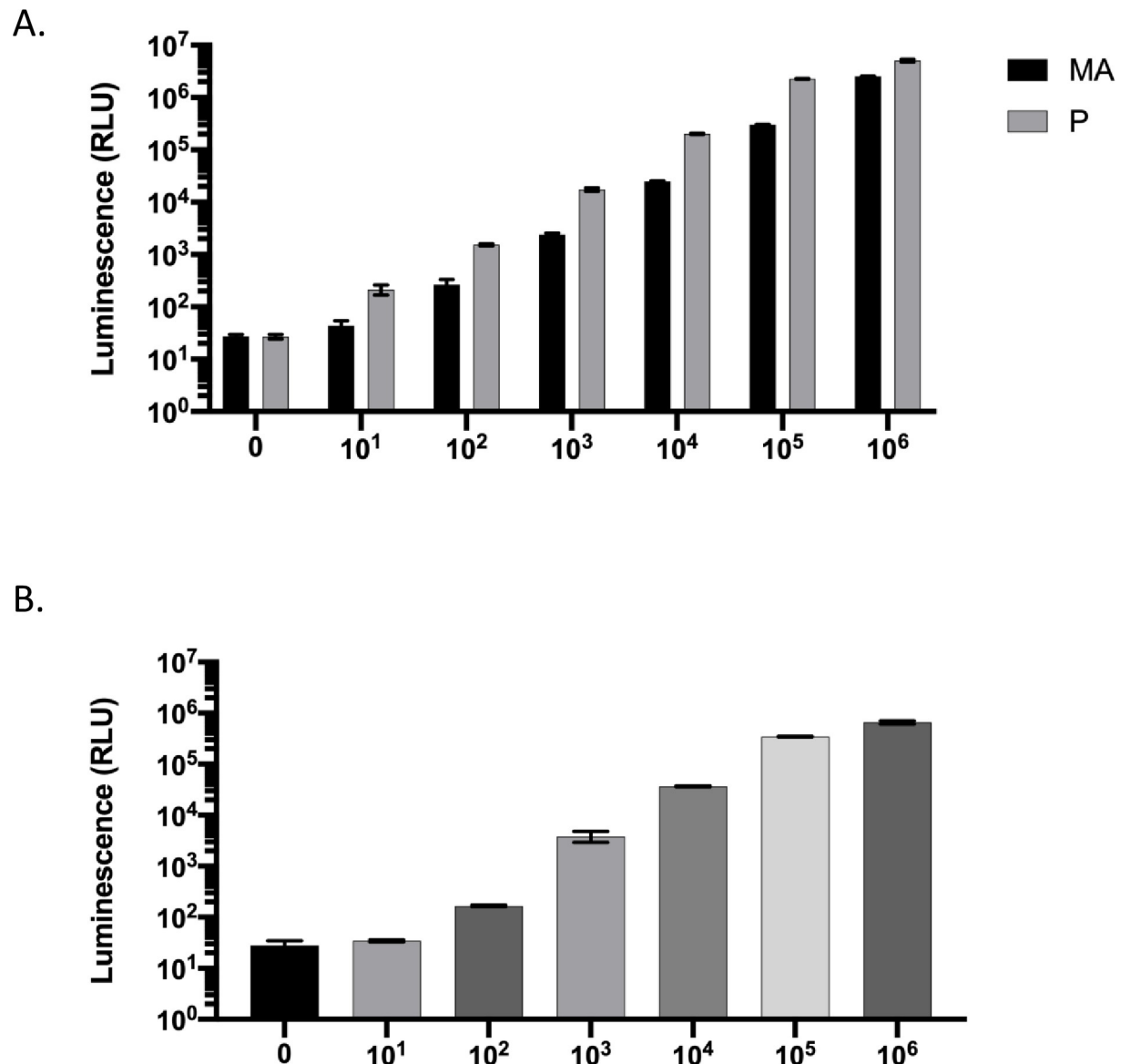


**Fig 1. *Tv* is delivered into the mouse male urogenital tract (MUT) using urethral catheterization.** (A) Diagram of catheterization and *Tv* delivery into the mouse MUT.  $10^8$  Nluc expressing parasites suspended in 100  $\mu$ l RPMI-1640 medium were introduced through the urethral meatus into the urethra via a syringe and polyethylene catheter. Additional mouse penile features such as the glans, penile bone, and the male urogenital mating protuberance (MUMP) are depicted. Mouse penis sketch adapted from a mid-sagittal sectioning of the adult mouse penis [74]. (B) A simplified diagram of a lateral view of the male urogenital system in adult mice modified from Cunha *et al.* [75]. Parasites introduced into the mouse MUT via urethral catheterization would be delivered through the urethra (U) to the anterior (AP), dorsal (DP), lateral (LP), and ventral (VP) prostatic lobes. The bladder and seminal vesicles (SV) are also depicted. Histological validation of *Tv* delivery into the mouse MUT (C) versus the vehicle control. (E) The right panels are a 20X magnification of the left anterior prostate lobe (marked by a circle in the left panels). *T. vaginalis* parasites are indicated by a blue arrow. (D) 100X magnification of *Tv* parasite found in infected mouse MUT tissue. Blue arrow indicates the presence of flagella.

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To confirm that *Tv*-Nluc parasites would yield a detectable signal post-inoculation into the mouse MUT, we introduced  $10^8$  *Tv*-Nluc parasites or RPMI medium into three mice per condition. We then harvested the MUT tissues and homogenized the tissue by mincing and probe sonication to prepare the samples for Nluc and Bradford assay analysis (Fig 3A). *T. vaginalis*-Nluc infected mice display a significantly higher Nluc signal ( $p = 0.0088$ ) at  $96,600 \pm 20,200$  RLU/ $\mu$ g protein compared to the vehicle control at  $4.41 \pm 1.37$  RLU/ $\mu$ g protein (Fig 3B). Together, these data illustrate that *Tv* can be reproducibly delivered into the mouse MUT and that Nluc-expressing parasites can be harvested, and parasite burden measured via luciferase activity and total protein quantification.



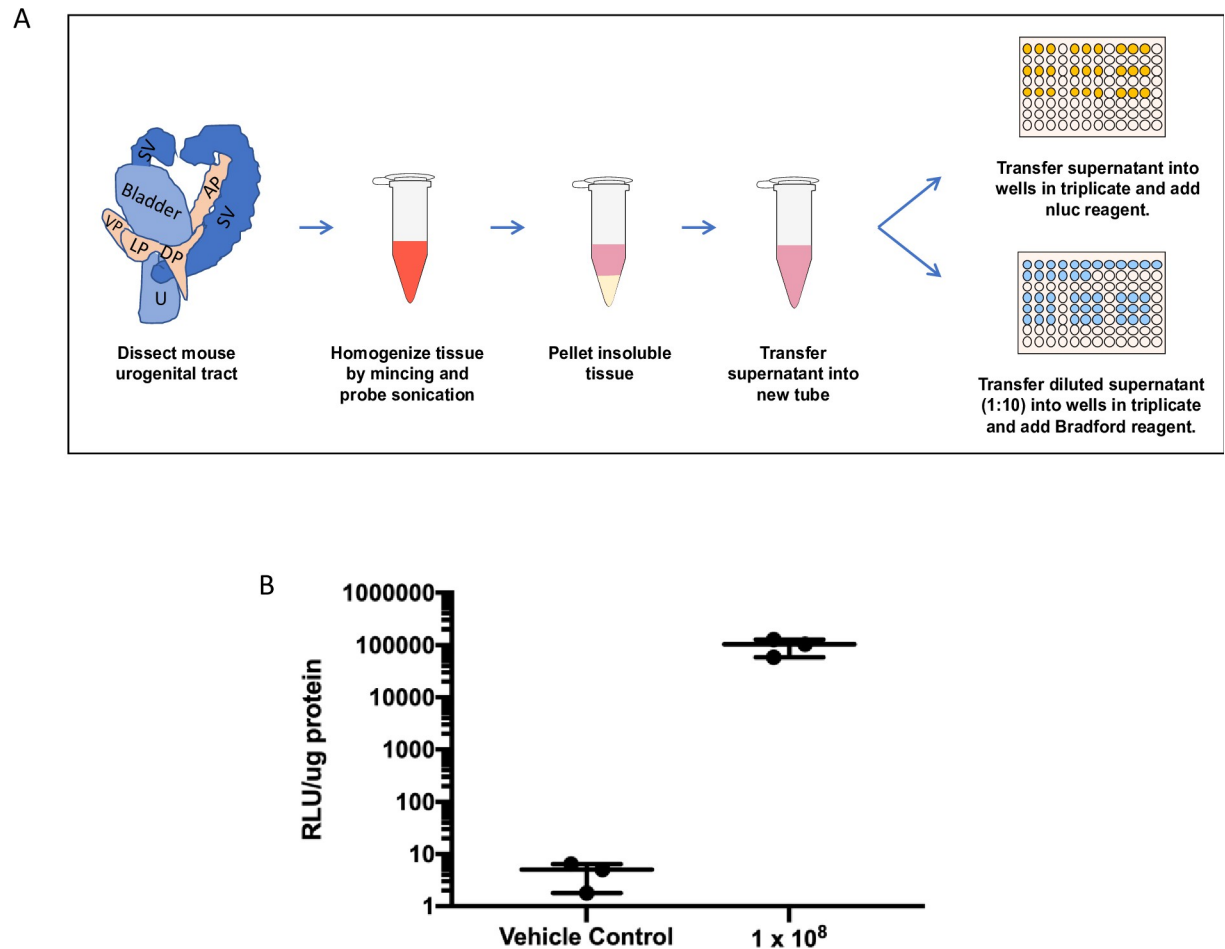


**Fig 2. Linearity of luminescence in different Nluc-expressing *Tv* strains.** *T. vaginalis* strains LSU 160 MA and P (A) and MSA 1103 (B) nucleofected with pMasterNeo-Nluc plasmid were measured for luminescence at varying amounts of parasites from 0 to 10<sup>6</sup> parasites (as indicated on the X axis) to determine the linear range of the assay for each strain. Data shown are averages of luminescence signal from 3 technical replicates with standard deviation for each strain.

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### Quantification of *Tv* in the mouse urogenital tract up to 72 hours post-inoculation

Having established the basis for our male mouse model, we asked whether parasites persist in the mouse MUT over time. Using *Tv* strain MSA 1103, as it has a statistically significant preference for prostate cells *in vitro* [27], we quantified parasite burden at 48 and 72 hrs post-inoculation of the Nluc-expressing *Tv*. Parasite burden decreased from 58,500 ± 12,500 RLU/μg protein at 0 hrs to 12,500 ± 2,720 RLU/μg protein ( $p < 0.0001$ ) at 48 hrs and 1,400 ± 404 RLU/μg protein ( $p < 0.0001$ ) at 72 hrs (Fig 4). Parasite burden could not be measured at time points past 72 hrs as the signal fell on or below the Nluc assay limit of detection (S2 Data).



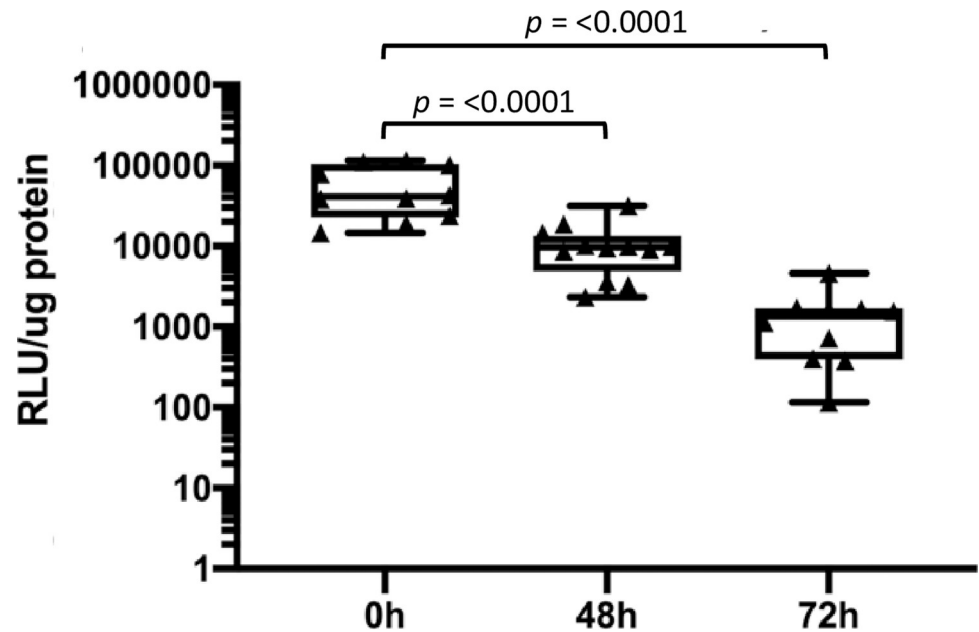
**Fig 3. Quantifying parasite burden using nanoluciferase-expressing *Tv*.** (A) Schematic of mouse MUT tissue sample preparation and quantification of *Tv* parasite burden. Excised mouse MUT tissue is finely minced and suspended in prostate lysis buffer (2 mL/mg tissue). MUT tissue then undergoes additional homogenization via probe sonication to ensure complete tissue dissociation. Insoluble material is pelleted via centrifugation and the supernatant is transferred to a new tube to further quantify luciferase activity and protein concentration using the Nano-Glo luciferase and Bradford assays, respectively. Detailed description found in materials and methods. (B) Mice were inoculated with  $10^8$  parasites ( $n = 3$ ) or RPMI vehicle control ( $n = 3$ ) and promptly euthanized. Samples were processed and analyzed by Nluc and Bradford assays to quantify parasite load. Data is shown as average luminescence signal/ $\mu$ g protein of the sample.

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Furthermore, to confirm that viable parasites were being recovered from the MUT tissue at these time points, *Tv*-inoculated tissue was processed and resuspended in Diamond's modified TYM media to count parasites via hemocytometer. Similar to the Nluc data, parasite numbers also decrease over time (Fig 5). This data demonstrates that Nluc signal reflects parasite numbers, as required to directly compare parasite survival in mice over a 72 hr period.

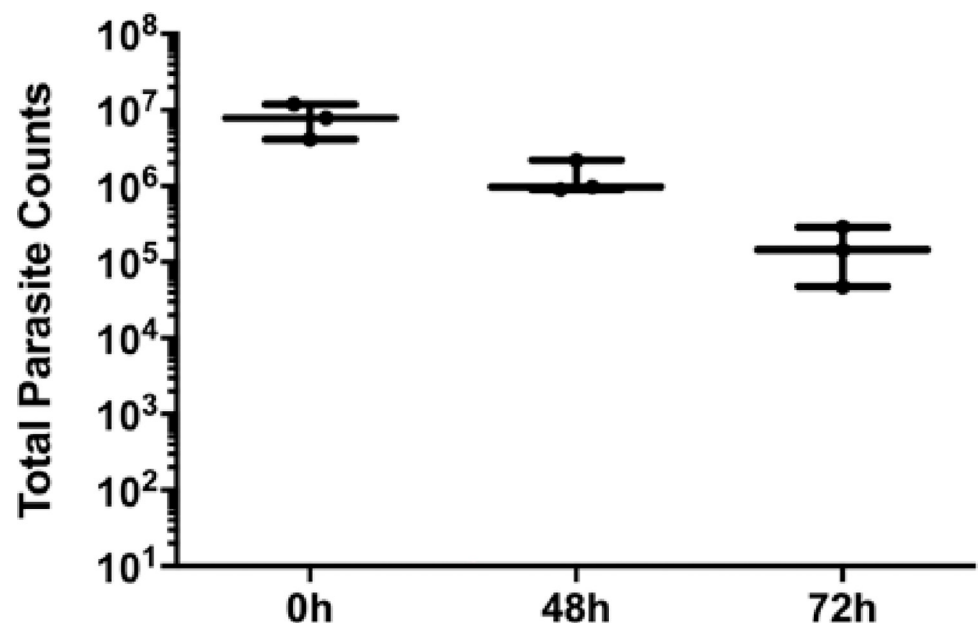
### Increased Host-Cell adherence *in vitro* correlates with increased parasite survival *in vivo*

It is well-established that *T. vaginalis* adherence to host cells *in vitro* varies between strains [27,59,60]. In addition, biochemical and genetic analyses indicate that these strains may differ in other properties in addition to their adherence phenotypes [47,61–64]. To measure parasite infectivity based solely on adherence ability, isogenic strains of *T. vaginalis* with different adherence abilities [33] (more adherent–MA; parental–P) were used to determine if *in vitro*



**Fig 4. *Tv* found within the murine MUT tissue 72 hours post-inoculation.** Quantification of Nluc-expressing MSA 1103 parasites in infected mouse MUT tissues 48 hrs or 72 hrs post-inoculation (as indicated on the X axis). Data are based on 12 infected mice per time point per strain and are averages of luminescence/ $\mu$ g protein of the sample with standard deviation. Data are representative of 3 independent experiments.

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**Fig 5. Viable parasites are recovered from mouse MUT tissue 72 hours post-inoculation.**  $10^8$  *Tv* parasites were introduced into the mouse urogenital tract and the infected tissue was then excised at 0 hr, 48 hrs, and 72 hrs post-inoculation (as indicated on the X axis). Tissue was minced, dissociated using Accutase cell detachment solution, resuspended in completed modified TYM media and counted via hemocytometer. Data shown are the total number of parasites per sample with standard deviation and were carried out using 3 mice per timepoint.

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host cell adherence is correlated with *in vivo* survival. Equivalent numbers of MA and P parasites ( $10^8/100 \mu\text{L}$ ) were delivered into the mouse MUTs using transurethral catheterization and parasite burden was determined 48 hrs and 72 hrs post-inoculation. Eleven to fourteen mice were analyzed for the MA strain at each time point while nine to twelve mice were used for the P strain. MUT tissues were processed, and parasites were quantified as described above.

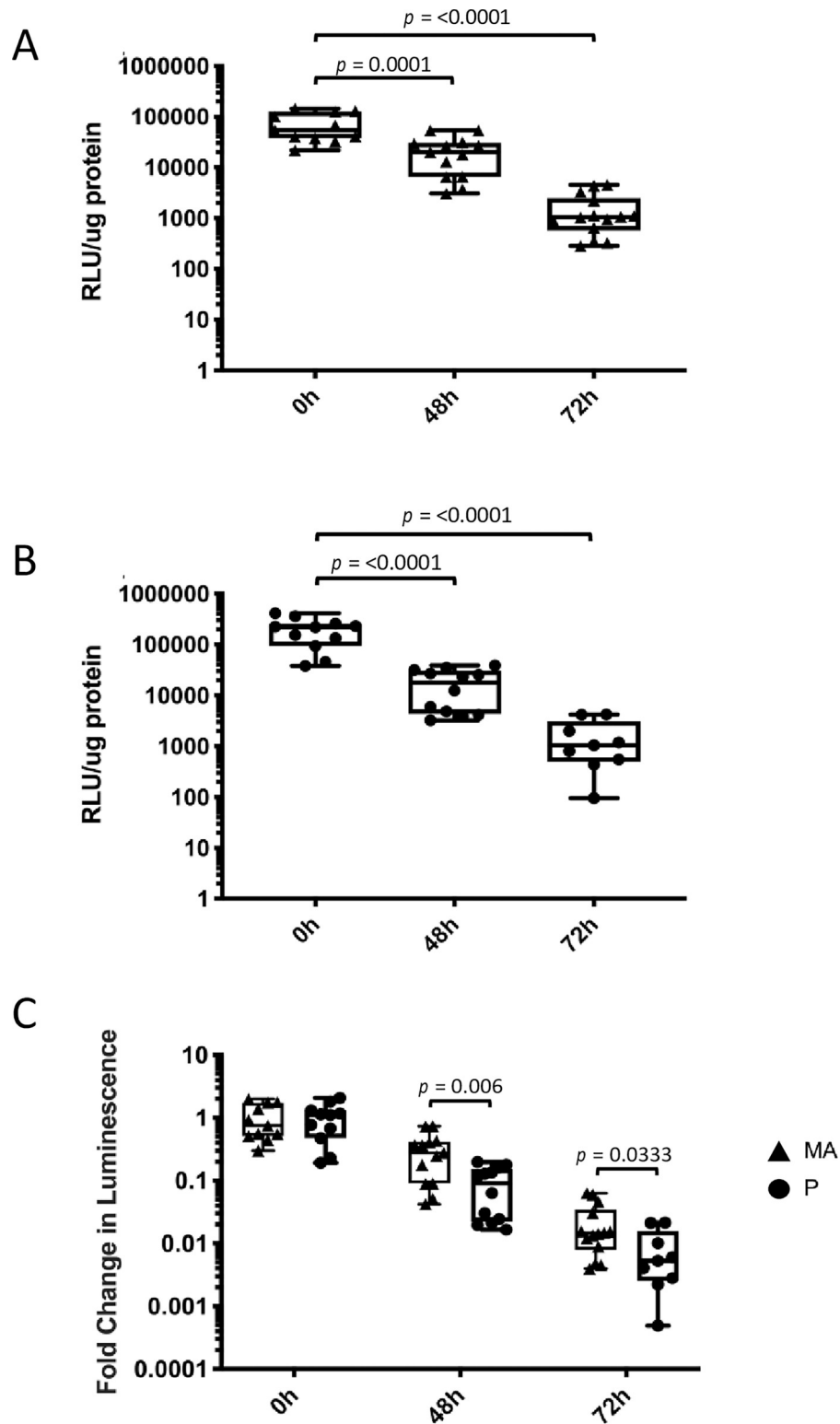
At 48 hrs post-inoculation, MA Nluc signal was  $22,400 \pm 4670 \text{ RLU}/\mu\text{g}$  protein and  $1,590 \pm 389 \text{ RLU}/\mu\text{g}$  at 72 hrs compared to  $72,300 \pm 13,500 \text{ RLU}/\mu\text{g}$  protein seen at 0 hrs (Fig 6A). This is a maximum reduction of  $\sim 1.75$  orders of magnitude in signal per  $\mu\text{g}$  protein between 0 hrs and 72 hrs. A similar trend was observed at 48 and 72 hrs post-inoculation for P parasites, with the signal averaging at  $19,100 \pm 3920 \text{ RLU}/\mu\text{g}$  and  $1620 \pm 523 \text{ RLU}/\mu\text{g}$ , respectively compared to  $198,000 \pm 36,200 \text{ RLU}/\mu\text{g}$  protein seen at 0 hrs (Fig 6B). This is a maximum reduction in signal per  $\mu\text{g}$  protein of  $\sim 2$  orders of magnitude between 0 hrs and 72 hrs. However, taking into account that P parasites express Nluc at higher levels than MA, as seen in Fig 2A, MA and P data was normalized to their respective 0 hrs and compared to see if the enhanced *in vitro* adherence of MA [33] results in increased survival *in vivo*. At both 48 hrs and 72 hrs, MA yields significantly higher parasite burden at  $\sim 3.4$  ( $p = 0.006$ ) and  $\sim 2.7$  ( $p = 0.03$ ) times higher than P, respectively (Fig 6C). These data demonstrate that the MA parasites have higher infection burden *in vivo* than P parasites, consistent with the observed increased adherence of MA parasites, relative to P parasites, to host cells *in vitro* [33]. The higher parasite burden observed for MA parasites indicates that greater adherence to host cells facilitates parasite infectivity and maintenance *in vivo*.

### Parasite extracellular vesicles (EV) increases the survival of parasites *in vivo*

We have demonstrated that extracellular vesicles (EVs) secreted from *Tv* are internalized by host cells [44,65] and increase the adherence of this extracellular parasite to host cells [43] *in vitro*, indicating EVs likely assist in parasite colonization of the host *in vivo*. To determine whether EVs affect the colonization and survival of parasites *in vivo*, we inoculated the mouse MUTs with equivalent amounts ( $1 \times 10^8$ ) of either MSA 1103 parasites preincubated for 30 minutes with  $50 \mu\text{g}/\text{mL}$  EVs or preincubated with the vehicle control, using transurethral catheterization. Parasite burden was then measured at 0, 48 and 72 hrs post-inoculation. At 48 and 72 hrs post-inoculation, MSA 1103 Nluc signal was  $4887 \pm 2167 \text{ RLU}/\mu\text{g}$  protein and  $530 \pm 152 \text{ RLU}/\mu\text{g}$ , respectively, from mice inoculated with parasites only, compared to  $29,965 \pm 18,294 \text{ RLU}/\mu\text{g}$  and  $2150 \pm 1181 \text{ RLU}/\mu\text{g}$ , respectively, from mice co-inoculated with parasites and  $50 \mu\text{g}/\text{mL}$  EVs. This is a maximum increase of  $\sim 1.8$  and  $\sim 1.3$  orders of magnitude in signal per  $\mu\text{g}$  protein, respectively, in mice co-inoculated with parasites and EVs, relative to those inoculated with parasites only (Fig 7). At both 48 and 72 hrs post-inoculation, mice co-inoculated with parasites and EVs had parasite burdens at  $\sim 6.1$  ( $p = 0.0001$ ) and  $\sim 4.1$  ( $p = 0.0001$ ) times higher than mice inoculated with parasites only (Fig 7). These data demonstrate that the presence of EVs significantly increases parasite survival *in vivo*. This increased parasite survival *in vivo* in the presence of EVs is consistent with our observation that EVs increase parasite adherence to host cells *in vitro* [43,66] and likely support the colonization and survival of the parasites in the MUT.

### Discussion

We have developed a male mouse model for the study of *T. vaginalis* (*Tv*) pathogenesis *in vivo*. Using transurethral catheterization, the male urogenital tract (MUT) of BALB/cJ mice was inoculated with *Tv* parasites that express a nanoluciferase gene. After confirming *Tv* delivery



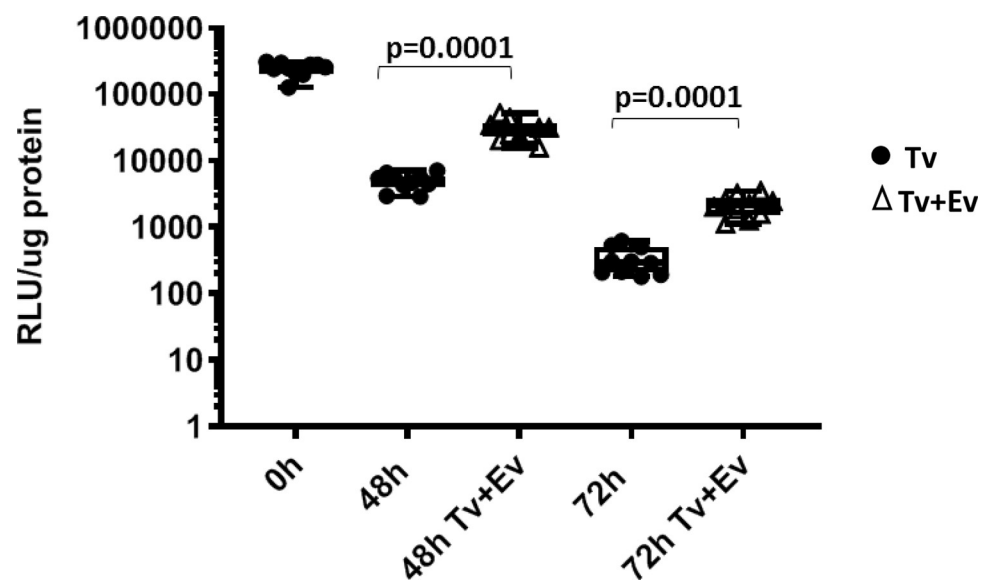
**Fig 6. *Tv* adherence to host cells *in vitro* correlates with an increased parasite burden in the MUT.** Quantification of Nluc-expressing LSU 160 more adherent (MA) (A) or parental (P) (B) parasites in mouse MUT tissues at 48 hrs and 72 hrs post-inoculation (as indicated on the X-axis). 11–14 mice were used for the MA strain at each time point. 9–12 mice were used for the P strain at each time point. Data shown are averages of luminescence/μg protein of the sample with standard deviation and are representative of 3 independent experiments. (C) Comparison of LSU160 MA (▲)

and LSU160 P (●) parasite load quantifications. Data is expressed as fold change in luminescence in the mouse urogenital tract with standard deviation normalized to their respective 0h post-inoculation luminescent signals.

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into the mouse MUT and identifying the linear range of our Nluc assay to allow for signal comparison across different time points post-inoculation (Figs 1–3), live parasites recovered from the MUT tissue 24 hrs and 72 hrs post-inoculation were assessed (Fig 4). Using this method to quantify parasite burden in tissues *ex-vivo*, we demonstrated that parasites could be detected up to 3 days post-inoculation (Fig 5). Having established this mouse model, we tested whether *Tv* isolates with an increased adherence to host cells *in vitro* survived better in the mouse MUT *in vivo* (Fig 6) and whether the addition of parasite EVs increased parasite burden (Fig 7).

To determine whether there is a correlation between the ability of a *Tv* strain to adhere to host cells *in vitro* and survival in the mouse MUT *in vivo*, isogenic strains of *Tv* which differ in their adherence phenotypes to host cells *in vitro* [33] were examined (Fig 6). We found that the more adherent (MA) parasites isolated from inoculated MUT tissues exhibited ~3.4-fold higher signal/ $\mu$ g protein at 48 hrs and ~2.7-fold higher signal/ $\mu$ g protein at 72 hrs compared to their less adherent parental (P) counterparts (Fig 6C). These data indicate that parasite adherence to host cells *in vitro* can be correlated with survival *in vivo*. This is consistent with strains displaying higher levels of *in vitro* adherence [27] which would likely cause a higher titer and more sustained survival. *Tv* is auxotrophic for several essential nutrients and must acquire these from the host or culture media when grown in culture. Nutrient acquisition is thought to require host cell lysis [35] and it has been shown that parasites must adhere to host cells to lyse them [27]. It is notable that in a *Listeria monocytogenes* infection model, highly adherent strains were found to be more infective to the murine liver compared to lowly adherent strains [67]. These data provide the first demonstration that *Tv* adherence to the host epithelium may



**Fig 7. Co-inoculation of parasite secreted EVs with *Tv* increases the parasite burden in the mouse urogenital tract.** Quantification of Nluc-expressing MSA 1103 parasites co-inoculated with either 50  $\mu$ g/ $\mu$ L EVs ( $\square$ ; *Tv*+EV) or parasites only ( $\bullet$ ; *Tv*) in the mouse MUT tissues at 48 and 72 hrs post-inoculation (as indicated on the X-axis). Ten mice were inoculated at each time point per condition. Data shown are averages of luminescence/ $\mu$ g protein of the sample with standard deviation normalized to 0h post-inoculation luminescent signals. Data are representative of 3 independent experiments.

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aid in establishing an infection *in vivo* and underscores the usefulness of using *in vitro* biochemical approaches to identify additional adherence factors which may ultimately inform design of additional therapeutics to treat infections.

We also established a correlation between the ability of parasite secreted EVs to increase the adherence of *Tv* to host cell monolayers *in vitro* [43] and increased survival of the parasite in our murine model (Fig 7). We found that the addition of EVs to parasite inoculations enhanced survival in the mice ~6.1 and ~4.1-fold, 48 and 72 hrs post-inoculation, relative to mice inoculated with parasites alone. These data illustrate for the first time that *Tv*EVs play a role in host:parasite interactions *in vivo* and confirm previous *in vitro*-based predictions that *Tv*EVs assist the parasite in colonizing the host *in vivo* [43,44].

Our male mouse model allows the introduction of *Tv* in the prostate without the need of pretreatments with hormones, antibiotics, corticosteroids or *Lactobacillus*, the predominant bacteria of the human vaginal microbiome [36,68], as is necessary in mouse vaginal infection models [36,37,41,69]. Using the prostate as an infection model has allowed us to avoid issues in reproducibility and parasite titer sustainability as seen in the *Tv* vaginal models monitoring early infection which is thought to arise from the required pretreatments. The vaginal microbiome is also known to affect the outcome of vaginal infections [70,71]. The sterility of the upper male urogenital tract [72] likely contributes to our ability to infect male mice without the need to manipulate the site of infection. Furthermore, this model may be useful for probing the possible role of bacterial infection(s) in the non-sterile urethra for the establishment of *Tv* infection in the male urogenital tract. This is the first male murine model for *Tv* infection to quantify parasite survival and examine the role of parasite adherence properties and the effect of EVs on colonization and survival. Our model differs from the *Tv* rat prostatitis model [42], which was established to examine tissue pathology, host immune cell infiltration and cytokine production. The mouse model described here will allow examination of parasite pathogenic factors, as well as granting access to the increased availability of genetically modified mice and cell lines.

Many *Tv* strains have established symbioses with either the bacterium *Mycoplasma hominis* and/or double-stranded RNA viruses (TVV) [35]. The strains we used for our analyses harbored the *M. hominis* symbiont, however, it is not known whether they also contained TVV. Future experiments, using this mice model and isogenic strains that differ only in the presence or absence of these symbionts, will help define the roles these symbionts play in establishing and maintaining *Tv* infection.

Persistent asymptomatic carrier state infection with *Tv* has been documented in men, whereas parasite clearance within 2 weeks has also been reported [73]. This male model of pathogenesis may allow specific factors of male trichomoniasis and infection outcome to be explored. Future experiments will be necessary to determine whether this model can be modified to establish and examine chronic infections in the mouse prostate and whether such infections are primarily asymptomatic or lead to inflammation and pathology. The ability to study chronic infections may allow better characterization of factors that contribute to adverse outcomes of infection. Information learned using the MUT model may also better inform design of vaginal models which would allow the contribution of the vaginal microbiota in *Tv* pathogenesis to be addressed.

## Supporting information

**S1 Data. Nluc signal is not altered in the absence of G418 selection.** LSU 160 MA-Nluc (A), LSU 160 P-Nluc (B), 1103-Nluc (C) were passaged daily for up to 72 hrs in the absence of G418 selective pressure. Luminescence signal was assayed from  $10^4$  parasites at 48 hrs and 72

hrs (X-axis) post-removal of G418 for each strain and found to have no significant difference in luminescence compared to their respective 0 hr control. Data shown are averages of luminescence signal with standard deviation and were carried out using 3 biological and 3 technical replicates for each strain.

(TIF)

**S2 Data. Nluc signal 4 d post-inoculation lies at the lower limit of Nluc detection.**  $10^8$  *Tv* parasites were introduced into the MUT, followed by excision of the MUT at 0 hr and 4 d post-inoculation (X-axis). The non-axial horizontal line denotes the lower limit of detection for the nanoluciferase assay. Data shown are averages of luminescence/ $\mu$ g protein of the sample with standard deviation using 3 mice per timepoint.

(TIF)

**S3 Data. Numerical values of luminescence in different Nluc-expressing *Tv* strains displayed in Fig 2.**

(XLSX)

**S4 Data. Numerical values for parasite burden displayed in Fig 3.**

(XLSX)

**S5 Data. Numerical values for *Tv* found within the murine MUT tissue displayed in Fig 4.**

(XLSX)

**S6 Data. Numerical values for viable parasites recovered from mouse MUT tissue displayed in Fig 5.**

(XLSX)

**S7 Data. Numerical values for parasites adhering to MUT tissue displayed in Fig 6.**

(XLSX)

**S8 Data. Numerical values for parasites adhering to MUT tissue displayed in Fig 7.**

(XLSX)

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## References

1. Schwebke JR, Burgess D. Trichomoniasis. *Clin Microbiol Rev.* 2004; 17(4):794–803. <https://doi.org/10.1128/CMR.17.4.794-803.2004> PMID: 15489349
2. World Health Organization. Global incidence and prevalence of selected curable sexually transmitted infections: 2008. *Reprod Health Matters.* 2012; 20(40):207–8.
3. World Health Organization. Report on global sexually transmitted infection surveillance, 2018. 2018. p. 1–74. Available from: <https://apps.who.int/iris/handle/10665/277258>.
4. Rowley J, Hoorn S Vander, Korenromp E, Low N, Unemo M, Abu-Raddad LJ, et al. Chlamydia, gonorrhoea, trichomoniasis and syphilis: Global prevalence and incidence estimates, 2016. *Bull World Health Organ.* 2019; 97(8):548–62. <https://doi.org/10.2471/BLT.18.228486> PMID: 31384073
5. Centers for Disease Control and Prevention. STD Surveillance Report 2018. 2019. Available from: <https://www.cdc.gov/nchstp/newsroom/2019/2018-STD-surveillance-report.html>.
6. Petrin D, Delgaty K, Bhatt R, Garber G. Clinical and microbiological aspects of *Trichomonas vaginalis*. *Clin Microbiol Rev.* 1998; 11(2):300–17.
7. Cotch MF, Pastorek JG, Nugent RP, Hillier SL, Gibbs RS, Martin DH, et al. *Trichomonas vaginalis* associated with low birth weight and preterm delivery. *Sex Transm Dis.* 1997; 24(6):353–60.
8. Minkoff HL, Eisenberger-Matityahu D, Feldman J, Burk R, Clarke L. Prevalence and incidence of gynecologic disorders among women infected with human immunodeficiency virus. *Am J Obstet Gynecol.* 1999; 180(4):824–36. [https://doi.org/10.1016/s0002-9378\(99\)70653-8](https://doi.org/10.1016/s0002-9378(99)70653-8) PMID: 10203650
9. Laga M, Manoka A, Kivuvu M, Malele B, Tuliza M, Nzila N, et al. Non-ulcerative sexually transmitted diseases as risk factors for HIV-1 transmission in women: Results from a cohort study. *AIDS.* 1993; 7(1):95–102. <https://doi.org/10.1097/00002030-199301000-00015> PMID: 8442924
10. Cu-Uvin S, Ko H, Jamieson DJ, Hogan JW, Schuman P, Anderson J, et al. Prevalence, Incidence, and Persistence or Recurrence of Trichomoniasis among Human Immunodeficiency Virus (HIV)-Positive Women and among HIV-Negative Women at High Risk for HIV Infection. *Clin Infect Dis.* 2002; 34(10):1406–11. <https://doi.org/10.1086/340264> PMID: 11981738
11. Kissinger P, Adamski A. Trichomoniasis and HIV interactions: A review. *Sex Transm Infect.* 2013; 89(6):426–33. <https://doi.org/10.1136/sextrans-2012-051005> PMID: 23605851
12. Swygard H, Seña AC, Hobbs MM, Cohen MS. Trichomoniasis: Clinical manifestations, diagnosis and management. *Sex Transm Infect.* 2004; 80(2):91–5. <https://doi.org/10.1136/sti.2003.005124> PMID: 15054166
13. Leitsch D. Recent Advances in the *Trichomonas vaginalis* Field [version 1; peer review: 2 approved]. *F1000Research.* 2016; 5(162).
14. Kissinger P. *Trichomonas vaginalis*: A review of epidemiologic, clinical and treatment issues. *BMC Infect Dis.* 2015; 15(1). <https://doi.org/10.1186/s12879-015-1055-0> PMID: 26242185
15. Hobbs MM, Lapple DM, Lawing LF, Schwebke JR, Cohen MS, Swygard H, et al. Methods for Detection of *Trichomonas vaginalis* in the Male Partners of Infected Women: Implications for Control of Trichomoniasis. *J Clin Microbiol.* 2006; 44(11):3994–9.

16. Škerk V, Schönwald S, Granić J, Krhen I, Baršić B, Mareković I, et al. Chronic Prostatitis Caused by *Trichomonas vaginalis*—Diagnosis and Treatment. *J Chemother.* 2002; 14(5):537–8.
17. Gopalkrishnan K, Hinduja IN, Anand Kumar TC. Semen characteristics of asymptomatic males affected by *Trichomonas vaginalis*. *J Vitro Fertil Embryo Transf.* 1990; 7(3):165–7. <https://doi.org/10.1007/BF01135682> PMID: 2380623
18. Kuberski T. *Trichomonas vaginalis* associated with nongonococcal urethritis and prostatitis. *Sex Transm Dis.* 1980; 7(3):135–6.
19. Bowden FJ, Garnett GP. *Trichomonas vaginalis* epidemiology: Parameterising and analysing a model of treatment interventions. *Sex Transm Infect.* 2000; 76(4):248–56. <https://doi.org/10.1136/sti.76.4.248> PMID: 11026878
20. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. *CA Cancer J Clin.* 2020; 70(1):7–30. <https://doi.org/10.3322/caac.21590> PMID: 31912902
21. Mitteregger D, Aberle SW, Makrithathis A, Walochnik J, Brozek W, Marberger M, et al. High detection rate of *Trichomonas vaginalis* in benign hyperplastic prostatic tissue. *Med Microbiol Immunol.* 2012; 201(1):113–6. <https://doi.org/10.1007/s00430-011-0205-2> PMID: 21660495
22. Sutcliffe S, Giovannucci E, Alderete JF, Chang TH, Gaydos CA, Zenilman JM, et al. Plasma antibodies against *Trichomonas vaginalis* and subsequent risk of prostate cancer. *Cancer Epidemiol Biomarkers Prev.* 2006; 15(5):939–45. <https://doi.org/10.1158/1055-9965.EPI-05-0781> PMID: 16702374
23. Stark JR, Judson G, Alderete JF, Mundodi V, Kucknoor AS, Giovannucci EL, et al. Prospective study of *Trichomonas vaginalis* infection and prostate cancer incidence and mortality: Physicians' health study. *J Natl Cancer Inst.* 2009; 101(20):1406–11.
24. Iqbal J, Al-Rashed J, Kehinde EO. Detection of *Trichomonas vaginalis* in prostate tissue and serostatus in patients with asymptomatic benign prostatic hyperplasia. *BMC Infect Dis.* 2016; 16(1):506. <https://doi.org/10.1186/s12879-016-1843-1> PMID: 27660027
25. Crouch ML, Alderete JF. *Trichomonas vaginalis* interactions with fibronectin and laminin. *Microbiology.* 1999; 145(10):2835–43.
26. Pereira-Neves A, Benchimol M. Phagocytosis by *Trichomonas vaginalis*: new insights. *Biol Cell.* 2007; 99(2):87–101. <https://doi.org/10.1042/BC20060084> PMID: 17029588
27. Lustig G, Ryan CM, Secor WE, Johnson PJ. *Trichomonas vaginalis* contact-dependent cytolysis of epithelial cells. *Infect Immun.* 2013; 81(5):1411–9. <https://doi.org/10.1128/IAI.01244-12> PMID: 23429535
28. Bastida-Corcuera FD, Okumura CY, Colocoussi A, Johnson PJ. *Trichomonas vaginalis* lipophosphoglycan mutants have reduced adherence and cytotoxicity to human ectocervical cells. *Eukaryot Cell.* 2005; 4(11):1951–8. <https://doi.org/10.1128/EC.4.11.1951-1958.2005> PMID: 16278462
29. Okumura CYM, Baum LG, Johnson PJ. Galectin-1 on cervical epithelial cells is a receptor for the sexually transmitted human parasite *Trichomonas vaginalis*. *Cell Microbiol.* 2008; 10(10):2078–90.
30. De Miguel N, Lustig G, Twu O, Chattopadhyay A, Wohlschlegel JA, Johnson PJ. Proteome analysis of the surface of *Trichomonas vaginalis* reveals novel proteins and strain-dependent differential expression. *Mol Cell Proteomics.* 2010; 9(7):1554–66. <https://doi.org/10.1074/mcp.M000022-MCP201> PMID: 20467041
31. Riestra AM, Gandhi S, Sweredoski MJ, Moradian A, Hess S, Urban S, et al. A *Trichomonas vaginalis* Rhomboid Protease and Its Substrate Modulate Parasite Attachment and Cytolysis of Host Cells. *PLoS Pathog.* 2015; 11(12):1–25.
32. Chen YP, Riestra AM, Rai AK, Johnson PJ. A novel cadherin-like protein mediates adherence to and killing of host cells by the parasite *Trichomonas vaginalis*. *MBio.* 2019; 10(3):e00720–19. <https://doi.org/10.1128/mBio.00720-19> PMID: 31088924
33. Molgora BM, Rai AK, Sweredoski MJ, Moradian A, Hess S, Johnson PJ. A Novel *Trichomonas vaginalis* Surface Protein Modulates Parasite Attachment via Protein:Host Cell Proteoglycan Interaction. *MBio.* 2021; 12(1):1–17.
34. Ryan CM, De Miguel N, Johnson PJ. *Trichomonas vaginalis*: Current understanding of host-parasite interactions. *Essays Biochem.* 2011; 51(1):161–75.
35. Mercer F, Johnson PJ. *Trichomonas vaginalis*: Pathogenesis, Symbiont Interactions, and Host Cell Immune Responses. *Trends Parasitol.* 2018; 34(8):683–93.
36. Cobo ER, Eckmann L, Corbeil LB. Murine models of vaginal trichomonad infections. *Am J Trop Med Hyg.* 2011; 85(4):667–73. <https://doi.org/10.4269/ajtmh.2011.11-0123> PMID: 21976570
37. McGrory T, Garber GE. Mouse intravaginal infection with *Trichomonas vaginalis* and role of *Lactobacillus acidophilus* in sustaining infection. *Infect Immun.* 1992; 60(6):2375–9.

38. Lushbaugh WB, Blossom AC, Shah PH, Banga AK, Jaynes JM, Cleary JD, et al. Use of intravaginal microbicides to prevent acquisition of *Trichomonas vaginalis* infection in *Lactobacillus*-pretreated, estrogenized young mice. *Am J Trop Med Hyg.* 2000; 63(5–6):284–9. PMID: [11421379](#)
39. Smith JD, Garber GE. *Trichomonas vaginalis* infection induces vaginal CD4+ T-cell infiltration in a mouse model: A vaccine strategy to reduce vaginal infection and HIV transmission. *J Infect Dis.* 2015 Jul 15; 212(2):285–93. PMID: [25616405](#)
40. Riestra AM, Valderrama JA, Patras KA, Booth SD, Quek XY, Tsai C-M, et al. *Trichomonas vaginalis* Induces NLRP3 Inflammasome Activation and Pyroptotic Cell Death in Human Macrophages. *J Innate Immun.* 2019; 11(1):86–98.
41. Meysick KC, Garber GE. Interactions between *Trichomonas vaginalis* and Vaginal Flora in a Mouse Model. *J Parasitol.* 1992; 78(1):157–60.
42. Jang K-S, Han I-H, Lee S-J, Yoo J, Kim Y-S, Sim S, et al. Experimental rat prostatitis caused by *Trichomonas vaginalis* infection. *Prostate.* 2019; 79(4):379–89. <https://doi.org/10.1002/pros.23744> PMID: [30488471](#)
43. Twu O, de Miguel N, Lustig G, Stevens GC, Vashisht AA, Wohlschlegel JA, et al. *Trichomonas vaginalis* Exosomes Deliver Cargo to Host Cells and Mediate Host:Parasite Interactions. *PLoS Pathog.* 2013; 9(7):e1003482.
44. Rai AK, Johnson PJ. *Trichomonas vaginalis* extracellular vesicles are internalized by host cells using proteoglycans and caveolin-dependent endocytosis. *Proc Natl Acad Sci U S A.* 2019; 116(43):21354–60. <https://doi.org/10.1073/pnas.1912356116> PMID: [31601738](#)
45. Diamond LS. The Establishment of Various Trichomonads of Animals and Man in Axenic Cultures. *J Parasitol.* 1957; 43(4):488. PMID: [13463700](#)
46. Fouts AC, Kraus SJ. *Trichomonas vaginalis*: Reevaluation of its clinical presentation and laboratory diagnosis. *J Infect Dis.* 1980; 141(2):137–43.
47. Janssen BD, Chen YP, Molgora BM, Wang SE, Simoes-Barbosa A, Johnson PJ. CRISPR/Cas9-mediated gene modification and gene knock out in the human-infective parasite *Trichomonas vaginalis*. *Sci Rep.* 2018; 8(270).
48. Dyll SD, Koehler CM, Delgadillo-Correa MG, Bradley PJ, Plümper E, Leuenberger D, et al. Presence of a Member of the Mitochondrial Carrier Family in Hydrogenosomes: Conservation of Membrane-Targeting Pathways between Hydrogenosomes and Mitochondria. *Mol Cell Biol.* 2000; 20(7):2488–97. <https://doi.org/10.1128/MCB.20.7.2488-2497.2000> PMID: [10713172](#)
49. Hall MP, Unch J, Binkowski BF, Valley MP, Butler BL, Wood MG, et al. Engineered Luciferase Reporter from a Deep Sea Shrimp Utilizing a Novel Imidazopyrazinone Substrate. *ACS Chem Biol.* 2012; 7:49. <https://doi.org/10.1021/cb3002478> PMID: [22894855](#)
50. Boehm BJ, Colopy SA, Jerde TJ, Loftus CJ, Bushman W. Acute bacterial inflammation of the mouse prostate. *Prostate.* 2012; 72(3):307–17. <https://doi.org/10.1002/pros.21433> PMID: [21681776](#)
51. Gardner WAJ, Culbertson DE, Bennett BD. *Trichomonas vaginalis* in the prostate gland. *Arch Pathol Lab Med.* 1986; 110(5):430–2.
52. Simons BW, Durham NM, Bruno TC, Grosso JF, Schaeffer AJ, Ross AE, et al. A human prostatic bacterial isolate alters the prostatic microenvironment and accelerates prostate cancer progression HHS Public Access. *J Pathol.* 2015; 235(3):478–89. <https://doi.org/10.1002/path.4472> PMID: [25348195](#)
53. Khalili M, Mutton LN, Gurel B, Hicks JL, De Marzo AM, Bieberich CJ. Loss of Nkx3.1 Expression in Bacterial Prostatitis. *Am J Pathol.* 2010; 176(5):2259–68.
54. Wang L, Zoetemelk M, Chitteti BR, Ratliff TL, Myers JD, Srour EF, et al. Expansion of prostate epithelial progenitor cells after inflammation of the mouse prostate. *Am J Physiol—Ren Physiol.* 2015; 308(12):F1421–30. <https://doi.org/10.1152/ajprenal.00488.2014> PMID: [25925259](#)
55. Shinohara DB, Vaghasia AM, Yu S-H, Mak TN, Brüggemann H, Nelson WG, et al. A mouse model of chronic prostatic inflammation using a human prostate cancer-derived isolate of *Propionibacterium acnes*. *Prostate.* 2013; 73(9):1007–15.
56. MacKern-Oberti JP, Motrich RD, Bresler ML, Cejas H, Cuffini C, MacCioni M, et al. Male rodent genital tract infection with *Chlamydia muridarum*: Persistence in the prostate gland that triggers self-immune reactions in genetically susceptible hosts. *J Urol.* 2011; 186(3):1100–6.
57. Silberstein E, Serna C, Fragoso SP, Nagarkatti R, Debrabant A. A novel nanoluciferase-based system to monitor *Trypanosoma cruzi* infection in mice by bioluminescence imaging. *PLoS One.* 2018; 13(4).
58. De Niz M, Stanway RR, Wacker R, Keller D, Heussler VT. An ultrasensitive NanoLuc-based luminescence system for monitoring *Plasmodium berghei* throughout its life cycle. *Malar J.* 2016; 15(1):232.
59. Pachano T, Nieves YR, Lizarraga A, Johnson PJ, Strobl-Mazzulla PH, de Miguel N. Epigenetics regulates transcription and pathogenesis in the parasite *Trichomonas vaginalis*. *Cell Microbiol.* 2017; 19(6):e12716.

60. Nieves YR, Vashisht AA, Corvi MM, Metz S, Johnson PJ, Wohlschlegel JA, et al. Protein palmitoylation plays an important role in *Trichomonas vaginalis* adherence. *Mol Cell Proteomics*. 2018; 17(11):2229–41.
61. Meade JC, Carlton JM. Genetic diversity in *Trichomonas vaginalis*. *Sex Transm Infect*. 2013; 89(6):444–8. <https://doi.org/10.1136/sextrans-2013-051098> PMID: 23702460
62. Bradic M, Warring SD, Tooley GE, Scheid P, Secor WE, Land KM, et al. Genetic indicators of drug resistance in the highly repetitive genome of *Trichomonas vaginalis*. *Genome Biol Evol*. 2017; 9(6):1658–72.
63. Conrad MD, Gorman AW, Schillinger JA, Fiori PL, Arroyo R, Malla N, et al. Extensive genetic diversity, unique population structure and evidence of genetic exchange in the sexually transmitted parasite *Trichomonas vaginalis*. *PLoS Negl Trop Dis*. 2012; 6(3).
64. Kirkcaldy RD, Augustini P, Asbel LE, Bernstein KT, Kerani RP, Mettenbrink CJ, et al. *Trichomonas vaginalis* antimicrobial drug resistance in 6 US cities, STD surveillance network, 2009–2010. *Emerg Infect Dis*. 2012; 18(6):939–43.
65. Artuyants A, Campos TL, Rai AK, Johnson PJ, Dauros-Singorenko P, Phillips A, et al. Extracellular vesicles produced by the protozoan parasite *Trichomonas vaginalis* contain a preferential cargo of tRNA-derived small RNAs. *Int J Parasitol*. 2020; 50(14):1145–55.
66. Twu O, Johnson PJ. Parasite Extracellular Vesicles: Mediators of Intercellular Communication. *PLoS Pathog*. 2014; 10(8):e1004289. <https://doi.org/10.1371/journal.ppat.1004289> PMID: 25166021
67. Kushwaha K, Muriana PM. Analysis of tissue invasiveness of adherent strains of *Listeria monocytogenes* by *in vivo* mouse assay. *Int J Food Microbiol*. 2010; 141(1–2):104–9. <https://doi.org/10.1016/j.ijfoodmicro.2010.03.004> PMID: 20452072
68. Ravel J, Brotman RM, Gajer P, Ma B, Nandy M, Fadrosch DW, et al. Daily temporal dynamics of vaginal microbiota before, during and after episodes of bacterial vaginosis. *Microbiome*. 2013; 1(1):29. <https://doi.org/10.1186/2049-2618-1-29> PMID: 24451163
69. Cappuccinelli P, Lattes C, Cagliani I, Negro Ponzi A. Features of intravaginal *Trichomonas vaginalis* infection in the mouse and the effect of oestrogen treatment and immunodepression. *G Bacteriol Virol Immunol*. 1974; 67(1–6):31–40.
70. Brotman RM. Vaginal microbiome and sexually transmitted infections: An epidemiologic perspective. *J Clin Invest*. 2011; 121(12):4610–7. <https://doi.org/10.1172/JCI57172> PMID: 22133886
71. Petrova MI, van den Broek M, Balzarini J, Vanderleyden J, Lebeer S. Vaginal microbiota and its role in HIV transmission and infection. *FEMS Microbiol Rev*. 2013; 37(5):762–92. <https://doi.org/10.1111/1574-6976.12029> PMID: 23789590
72. Mändar R. Microbiota of male genital tract: Impact on the health of man and his partner. *Pharmacol Res*. 2013; 69(1):32–41. <https://doi.org/10.1016/j.phrs.2012.10.019> PMID: 23142212
73. Krieger JN, Verdon M, Siegel N, Holmes KK. Natural history of urogenital trichomoniasis in men. *J Urol*. 1993; 149(6):1455–8. [https://doi.org/10.1016/s0022-5347\(17\)36414-5](https://doi.org/10.1016/s0022-5347(17)36414-5) PMID: 8501787
74. Rodriguez E, Weiss DA, Yang JH, Menshenina J, Ferretti M, Cunha TJ, et al. New Insights on the Morphology of Adult Mouse Penis. *Biol Reprod*. 2011; 85:1216–21. <https://doi.org/10.1095/biolreprod.111.091504> PMID: 21918128
75. Cunha GR, Donjacour AA, Cooke PS, Mee S, Bigsby RM, Higgins SJ, et al. The endocrinology and developmental biology of the prostate. *Endocr Rev*. 1987; 8(3):338–62. <https://doi.org/10.1210/edrv-8-3-338> PMID: 3308446