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# Cancer Tills the Premetastatic Field: Mechanistic **Basis and Clinical Implications**

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#### **Abstract**

A growing body of work has shown that cancer metastasis is not a random spontaneous event; rather, it is the culmination of a cascade of priming steps through which a subpopulation of the tumor cells acquires invasive traits while readying a permissive environment, termed the "premetastatic niche," in which distant metastases can occur. Signals from the primary tumor mobilize and adapt immune cells as well as directly communicating with distant niche cells to induce a broad spectrum of adaptations in target organs, including the induction of angiogenesis, inflammation, extracellular matrix remodeling, and metabolic reprogramming. Together, these interactions facilitate the formation of a premetastatic niche composed of a variable mix of resident and recruited immune cells, endothelial cells, and stromal cells connected through a complex signaling network that we are only beginning to understand. Here, we summarize the latest findings on how cancer induces and guides the formation of this premetastatic niche as well as potential prognostic markers and therapeutic targets that may lead to a better understanding and effective treatment of metastatic disease. Clin Cancer Res; 22(15); 3725-33. ©2016 AACR.

#### Introduction

Metastasis, the spread of cancer cells from a primary tumor to other organs, is the leading cause of mortality in cancer patients. This is partially due to the limited therapeutic options and the short time window that would allow a successful treatment of clinically detectable metastases. Therefore, there is a great and urgent need to elucidate metastasis-driving molecular and cellular events before and during early stages of metastatic colonization, which may guide development of therapies to prevent or eradicate metastases before they reach an incurable stage. Recent evidence highlights the important role of a premetastatic niche, initially proposed and proven by Kaplan and colleagues (1, 2), in cancer's preparation for metastasis. A premetastatic niche is free of cancer cells but has captured cancer-associated properties that are permissive and sometimes even supportive for cancer cells originating from a foreign tissue to grow. These earliest, noncancerous pathologic changes in a tumor-free organ have the unique potential to serve as prognostic biomarkers and therapeutic targets in the prevention and treatment of metastasis. An overview of the metastatic niche as a whole has been covered in several important reviews on related topics (3-8), including the excellent articles from Psaila and colleagues and Peinado and colleagues on premetastatic niche (9, 10). Here, we focus on the most recent findings that improve our understanding of the premetastatic niche and provide rationales for the development of therapies against cancer metastasis.

## The Premetastatic Niche Model

The long known "seed and soil" hypothesis for metastasis introduced by Paget (11) has been complemented and refreshed by modern cancer research. In the basic framework, migratory tumor cells (the "seeds") leave the primary tumor through intravasation, disseminate throughout the body via the circulation, and eventually engraft in a distant organ that provides an appropriate microenvironment (the "soil"). Recent studies indicate that dissemination of cancer cells from the primary site occurs during early cancer stages but is not sufficient for metastasis (9, 12, 13), emphasizing the essential role of a conducive niche in the target distant organ. The concept of a premetastatic niche was first proposed by Kaplan and colleagues in 2005 after the discovery that bone marrow-derived hematopoietic progenitor cells that are VEGFR1-positive are recruited to future metastatic sites before the tumor cells arrive, where the bone marrow-derived cells (BMDC) promote the chemoattraction and attachment of disseminated cancer cells through mechanisms including the SDF-1/CXCR4 axis (1). Subsequent studies have revealed that in a premetastatic niche, various types of cells together determine the fate of disseminated cancer cells in multiple aspects, including their extravasation, survival, colonization, and aggressive growth. Adaptation of a premetastatic niche prior to the arrival of tumor cells has been recognized as an important means for cancer to facilitate metastasis (6, 10, 14–20). It is worth noting that the premetastatic niche model may cooperate with other models depicting different steps and modes of metastasis (3), such as the tumor self-seeding model proposed by Kim and colleagues (Fig. 1; ref. 21).

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#### **Traits of a Premetastatic Niche**

Recruitment of BMDCs

BMDCs can be mobilized into the circulation and thereby participate in the establishment of a primary or (pre-)metastatic

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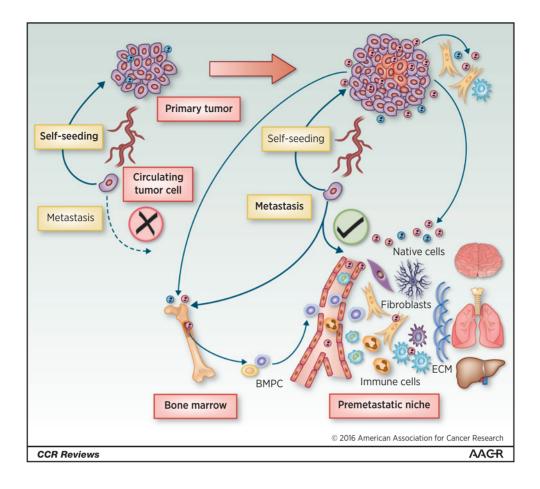


Figure 1.

Tumor-directed formation of a premetastatic niche. Recent studies, including those discussed in this review, suggest that extracellular vesicles (EV) and other factors secreted by the primary tumor initiate and direct the formation of a premetastatic niche. At an early tumor stage (left), circulating tumor cells (CTC) can be detected but are not capable of metastatic colonization due to insufficient invasive traits and/or lack of a permissive metastatic niche. Most CTCs will die, but some may travel back to the primary tumor to evolve a more aggressive phenotype according to the tumor self-seeding model. As the tumor grows and progresses (right), tumor cells experience additional genetic, epigenetic, or environmental alterations, including metabolic stresses (e.g., hypoxia), and secrete a variety of EV-associated and other factors. These tumor-secreted factors, upon release into the circulation, may cause a broad spectrum of systemic effects, including the induction of angiogenesis, inflammation, ECM remodeling, and metabolic reprogramming at a premetastatic site. All types of resident cells in a premetastatic organ can be affected directly or indirectly in this process. New types of noncancerous cells, such as the bone marrow progenitor cells (BMPC), are recruited and often reprogrammed to form the premetastatic niche. Metastatic colonization will succeed once CTCs have acquired sufficient intrinsic potential and a conducive premetastatic niche has been established. Combinatorial therapies targeting both cancer cells and factors driving the formation of premetastatic niche may hold promise for the prevention and treatment of metastasis.

tumor microenvironment as a nonresident cellular component. Factors secreted by primary tumor cells can activate resident fibroblast-like stromal cells at a premetastatic site, resulting in an increased production of the extracellular matrix (ECM) component fibronectin, which enables the adhesion and clustering of migratory BMDCs that express the fibronectin receptor VLA-4 (integrin  $\alpha_4\beta_1$ ), as well as genes related to their mobilization, including MMP9 and Id3, in the premetastatic niche (1). This leads to the expression of SDF-1 in the premetastatic niche, resulting in the recruitment of CXCR4<sup>+</sup> cancer cells. The SDF-1/CXCR4 chemokine axis also participates in the homing of BMDCs. A recent article shows that ECM metalloproteinase inducer (EMMPRIN) in cancer cells can induce the expression and secretion of several factors, such as SDF-1 and VEGF, that mediate BMDC recruitment to the lungs and liver (22). For primary tumors with STAT3 activation, BMDC recruitment can be partially mediated by tumor-secreted factors that are induced by STAT3 signaling, such as IL6 and IL10 (23). These secreted factors lead to a widespread STAT3 activation in premetastatic lungs, activate fibroblasts to produce fibronectin, and induce the formation of clusters of CD11b<sup>+</sup> myeloid-derived suppressor cells (MDSC) in the lungs, resulting in enhanced metastatic growth. MDSCs may also be recruited to the premetastatic lung through hypoxiainduced secreted factors, such as MCP-1 from the primary tumor (24), and the induction of the inflammatory proteins \$100A8 and S100A9 in endothelial and myeloid cells (10). CCL9 is induced through TGFB signaling in myeloid cells in the premetastatic lungs, where it enhances tumor cell survival and promotes metastasis (25). Another recent study has revealed that lysyl oxidaselike 2 (LOXL2) and the bHLH transcription factor E47, which function together to induce epithelial-to-mesenchymal transition (EMT), also contribute to the recruitment of BMDCs to

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premetastatic lungs through transcriptional regulation of fibronectin and cytokines, including GM-CSF (26). TNF $\alpha$ , TGF $\beta$ , and VEGF-A secreted by the primary tumor can induce the expression of S100A8 and S100A9 in premetastatic lung endothelial cells, which act as potent chemoattractants for Mac-1<sup>+</sup> myeloid cells and cancer cells through SAA3-induced Toll-like receptor 4 (TLR4) signaling (17, 18).

#### The heterogeneity of immune cells

It is becoming increasingly apparent that the immune constituents of the premetastatic niche are profoundly different between model systems, even within the same type of cancer, with some model systems showing the recruitment of one major cell type whereas others indicating a larger cross-talking network of cells. In the MMTV-polyoma middle T antigen mouse mammary tumor model, neutrophils were found to be the primary immune cells recruited to the premetastatic lungs, although these cells have a low frequency in the primary tumor microenvironment (27). However, this may be due to the timing of the experiments, as a recent study has shown that immune cells arrive at the premetastatic lung in three separate waves, and some of these cells are only transiently present in the tumors (28). The first wave of immune cells consists of neutrophils and peaks 15 to 30 minutes after tumor cell injection. The second wave is primarily composed of conventional monocytes and peaks 4 hours after tumor cell injection. Finally, nonalveolar macrophages, patrolling monocytes, and dendritic cells (DC) arrive at the premetastatic niche, which peaks at 6 to 24 hours. In mice bearing the primary mammary tumors, infiltration of lung tissues by CD11b<sup>+</sup>Ly6G<sup>+</sup> neutrophils starts before cancer cells can be detected in the lungs and further increases during the metastatic stage. Compared with neutrophils from healthy lungs, tumor-mobilized lung neutrophils are also mature but exhibit minor differences in gene expression. Premetastatic lung neutrophils secrete leukotrienes, which enhance the tumorigenic and metastatic potentials of primary tumor cells (27). Another study shows that mammary tumors induce a systemic expansion and polarization of neutrophils through IL1 $\beta$ -activated, IL17-producing  $\gamma\delta$  T cells in a G-CSF-dependent manner. These neutrophils may help to establish a premetastatic niche, where they suppress CD8<sup>+</sup> T-cell activation to facilitate metastasis (29).

Monocytes/macrophages also contribute to the establishment of a premetastatic niche. Palmitovlated surface antigens on breast cancer secreted exosomes induce NF-kB signaling in macrophages at a premetastatic site through activating TLR2, stimulating macrophages to secrete proinflammatory factors, such as IL6, to promote cancer cell growth (30). Other macrophage-secreted factors, such as granulin, can indirectly support cancer cell growth through the activation of fibroblasts to generate a more permissive niche (31). Macrophages may also comigrate with cancer cells, inducing the expression of Mena in the cancer cells, and the direct interaction between perivascular macrophages, endothelial cells, and Mena-overexpressing tumor cells is significantly correlated with metastatic disease in ER<sup>+</sup>/HER2<sup>-</sup> breast cancer (32). In the premetastatic lymph nodes of a lung tumor model, COX-2 and SDF-1 are induced in DCs, which further increase lymphangiogenesis and the recruitment of regulatory T cells (Treg), suggesting a role of DCs and prostaglandin E2 (PGE2) in the establishment of a premetastatic niche (33). However, DCs can also inhibit metastasis by engulfing tumor-secreted vesicles termed cytoplasts and

traveling to the mediastinal lymph node to activate ovalbuminspecific CD8+ T cells. Depletion of DCs has been shown to increase metastasis (28), highlighting the complexity of the immune components of the premetastatic niche. Similarly, as a part of cancer immunosurveillance, nonclassical "patrolling" CX3CR1<sup>high</sup>CD14<sup>dim</sup>CD16<sup>+</sup> monocytes are enriched in the microvasculature of the lungs, where they inhibit tumor cell adhesion to the vasculature and promote natural killer (NK) cell recruitment and activation to reduce lung metastases. In response to tumor challenge, lung endothelial cells increase the expression of CX3CL1, which attracts the patrolling monocytes expressing CX3CR1 (34). Other myeloid cells contributing to a premetastatic niche include platelets and granulocytes. Platelets form aggregates with circulating tumor cells, which reprogram them to secrete CXCL5 and CXCL7 to recruit granulocytes, forming an early metastatic niche for subsequent metastatic progression (35).

CD8<sup>+</sup> T cells are capable of constraining myeloid cell accumulation in premetastatic lymph nodes by inducing myeloid cell apoptosis. Patients with metastatic melanoma had decreased CD8<sup>+</sup> T-cell infiltration and increased STAT3 in lymph node myeloid cells, suggesting that metastatic tumors may inhibit CD8<sup>+</sup> T-cell expansion or homing to the premetastatic sites (36). Another study shows that complement C5a receptor (C5aR) facilitates metastasis by suppressing CD4<sup>+</sup> and CD8<sup>+</sup>T cells in the lungs and livers. This immunosuppression is mediated by recruitment of MDSCs, regulation of their TGFB and IL10 production, and generation of Treg cells (37). Immunosuppression in the premetastatic niche may also be mediated by factors, such as MCP-1, that are secreted from the primary tumor in response to hypoxia, resulting in the recruitment of MDSCs and immature NK cells, which have reduced cytotoxic activity, to the premetastatic lung (24). The tissue-resident alveolar macrophages, which are accumulated in the premetastatic lungs through C5aR-mediated proliferation instead of recruitment from the circulation, promote cancer metastasis to the lungs by shifting the T-cell population from Th1 towards Th2 to suppress their antitumor activity (38). S100A4 also increases primary tumor growth as well as metastasis by reducing the Th1/Th2 ratio in the lungs in a mammary tumor model (39).

## Reprogramming of stromal cells

The formation of a premetastatic niche not only involves the recruitment of foreign cells, such as immune cells, but also the reprogramming of the resident stromal cells to facilitate metastatic growth. Normal lung fibroblasts express miR-30 family members to restrain MMPs, such as MMP9, to stabilize the lung vasculature (40). Cancer cells reprogram fibroblasts to decrease their expression of miR-30 family members, resulting in enhanced MMP activity, vascular permeability, and metastasis. Factors secreted from the primary tumor induce the expression of αSMA in premetastatic fibroblasts, activating them to induce ECM remodeling through secretion of fibronectin, LOX, and LOXL2, thereby generating a more permissive microenvironment for metastasis (1, 41). The induction of senescence in osteoblasts in the bone increases their secretion of factors, such as IL6, to promote osteoclastogenesis, resulting in increased metastases (42). Premetastatic immune cells may also facilitate the reprogramming of stromal cells. Granulin secreted by CD11b+F4/ 80<sup>+</sup>Ly6G<sup>neg</sup>CCR2<sup>+</sup> metastasis-associated macrophages induces the expression of αSMA in premetastatic hepatic stellate cells

and induces their secretion of ECM remodeling proteins, such as periostin, to enhance metastatic growth (31). This relationship is reciprocal as fibrocytes can secrete CCL2, CCL5, and MMP9 to induce the recruitment of Ly-6C<sup>+</sup>, Ly-6G<sup>low</sup> monocytes into the premetastatic lung to promote metastasis (43). In some instances, cancer may also comigrate with stromal cells, such as fibroblasts, which enhance the viability of cancer cells at the premetastatic site (44).

#### Alterations in the ECM

Alterations in the premetastatic ECM are among the first steps in the formation of the premetastatic niche. Factors secreted by the primary tumor, including exosomes, can induce the accumulation of fibronectin in the premetastatic niche through several mechanisms, including secretion from the primary tumor and reprogramming of fibroblasts (1, 45). Premetastatic niche fibronectin can activate dormant metastatic cancer cells and mediate the recruitment of immune cells and metastatic cancer cells. Binding of VEGFR1<sup>+</sup> BMDCs to fibronectin induces α4β1 integrin signaling, resulting in increased MMP9 expression, enhancing the recruitment of BMDCs and cancer metastasis (1). MMP9 expression in lung endothelial cells and macrophages increases metastasis, enhances lung vascular permeability, and recruits BMDCs and monocytes (1, 10, 40, 43). Hypoxia in the primary tumor induces the secretion of fibronectin and lysyl oxidase (LOX), leading to their accumulation in the premetastatic niche (14). LOX colocalizes with fibronectin-rich regions to recruit CD11b<sup>+</sup> myeloid cells and c-Kit<sup>+</sup> myeloid progenitor cells to the lungs. LOX-mediated collagen cross-linking increases the MMP2 activity in the recruited myeloid cells. MMP2 enhances myeloid cell invasion and mediates collagen IV degradation, releasing collagen IV peptides into the blood, where they act as chemoattractants to generate a positive feedback loop for the recruitment of myeloid cells to the premetastatic niche. Activated fibroblasts in the premetastatic niche, often generated as a result of fibrosis, have increased expression and excretion of LOX and to a lesser extent LOXL2, resulting in increased collagen deposition and ECM stiffening, promoting metastatic cancer cell survival and cancer and immune cell engraftment (41, 46). Hypoxia also induces the secretion of exosomes containing LOXL2 on their outer surface, promoting collagen cross-linking (45). Cancer cells may also secrete such factors as osteopontin to facilitate the recruitment of granulin-expressing immune cells to the premetastatic niche, resulting in increased expression of ECM components and ECM remodeling factors (46).

#### Alterations in the vasculature

Blood vessels in a premetastatic niche directly control the arrest and extravasation of circulating cancer cells and are critical targets for tumor-derived adaptations in preparation for metastasis. Tumor-secreted extracellular vesicles (EV), including exosomes, systemically transfer tumor-derived regulators of the vascular endothelial barriers. Metastatic breast cancer cells, by secreting EV-encapsulated miR-105, downregulate tight junctions in endothelial cells and induce systemic vascular leakiness to promote metastasis (20). EVs secreted by brain metastatic breast cancer cells contain miR-181c, which promotes the destruction of the blood-brain barrier (BBB) through modulation of actin dynamics to facilitate brain metastases (47). In addition to EV-mediated mechanisms, premetastatic lungs express higher levels of angiopoietin-2, MMP3, and

MMP10, which possibly result from cancer-secreted TGFB1 and TNFα, and synergistically induce vascular permeability and the extravasation of circulating cancer cells (48). Another group also found that VEGF secreted by breast cancer cells induces angiopoietin-2 expression in brain microvascular endothelial cells, leading to impaired tight junction structures and increased BBB permeability (49). EMMPRIN expression induces the expression and secretion of SDF-1 and VEGF to induce BMDC-mediated angiogenesis (22). Cancer-secreted VEGF also recruits BMDCs to premetastatic lungs to increase inflammation, angiogenesis, and metastasis through inducing PGE2 production in endothelial cells (50). The peripheral blood plasma and bone marrow plasma from breast cancer patients increase transendothelial migration of breast cancer cells, which may involve systemic factors as well as factors in a premetastatic bone niche. Peripheral blood was only able to increase the migration of nonmetastatic cancer cells, suggesting that it acts through a mechanism that has already been acquired by metastatic cells (51). VEGFR1 expression in benign lymph nodes predicts recurrence of prostate cancer; however, the VEGFR1targeting drug axitinib fails to reduce lymph node VEGFR1, highlighting the need for better targets (52). In addition, CCL2 secreted by the primary tumor enhances CCL2 and CCR2 expression in lung endothelial cells and leukocytes, resulting in enhanced vascular permeability in the lungs through a S100A8-TLR4-mediated pathway (15). Healthy lung fibroblasts express miR-30 family members to inhibit the expression of MMPs, including MMP9, through targeting Skp2, resulting in stabilization of lung vasculature and inhibition of metastasis. Distant tumors are able to decrease the expression of miR-30 family members in premetastatic lung fibroblasts, resulting in vascular destabilization and increased metastasis (40).

The acquisition of EMT is an important step in the development of invasive and metastatic traits in the primary tumor and also results in the secretion of factors that facilitate angiogenesis. EVs secreted by cancer cells that have undergone partial or full EMT have greater enrichment of factors such as Rac1, tissue factor, and ECM remodeling proteins, which can promote endothelial cell proliferation and tube formation (53-55). Furthermore, EVs secreted by mesenchymal-like breast and ovarian cancer cells carry angiogenic molecules to activate endothelial cells through Akt phosphorvlation. Activated endothelial cells, in turn, increase their secretion of vesicles to induce EMT in epithelial cancer cells and promote metastasis (56). Given these findings, the acquisition of EMT in the primary tumor may lead to the release of exosomes that can enhance vascular permeability in the premetastatic niche to facilitate cancer and immune cell engraftment. However, further work must be done to demonstrate that these EMT-induced exosomes exert an effect outside of the primary tumor.

Lymphatic endothelial cells within premetastatic lungs and lymph nodes express CCL5 in response to IL6 secreted by breast cancer, directing cancer cell dissemination into these tissues. Mice treated with breast tumor–conditioned medium show enhanced angiogenesis and lymphangiogenesis in the lymph nodes, as well as enhanced lymphangiogenesis with unchanged angiogenesis in the primary tumors and lungs (57, 58). These results highlight a role of the tissue-residing lymphatic vessels, in addition to blood vessels, in the establishment of a premetastatic vascular niche.

#### Metabolic reprogramming of native cells

In a niche, which in ecology refers to the interactive position of a species in an ecosystem, the competition between different species for limited resources is one of the driving factors for dynamic population changes. When metastatic cancer cells arrive at a distant site, they must compete with the resident niche cells for the nutrients to establish a metastatic colony. Breast cancer cells can secrete EV-encapsulated miR-122, which can be taken up by niche cells, such as lung fibroblasts and astrocytes, to decrease the glucose consumption in these cells by targeting pyruvate kinase (19). This increases the availability of glucose for cancer cells, thus increasing their proliferation and survival to enhance metastasis. Another study shows that colorectal cancer cells, by secreting creatine kinase, convert liver-produced creatine into phosphocreatine that is subsequently taken up to fuel cancer cells during liver metastasis (59). These recent findings demonstrate an active role of noncancerous cells at a premetastatic site in rebalancing the metabolic needs between cancer and niche cells in response to cancer's exploitation of nutrients.

Metabolic stresses, such as hypoxia, are important drivers of tumor progression and also contribute to premetastatic niche formation. HIF1α stabilization under hypoxia induces cancer cells to secrete factors such as MCP-1, G-CSF, TNFα, VEGF, TIMP-1, and MMP9, which promote the recruitment of  $CD11b^{+}/Ly6C^{med}/Ly6G^{+}$  MDSCs as well as  $CD3^{-}/NK1.1^{+}/$ CD11b<sup>low</sup>/CD27<sup>low</sup> immature NK cells with reduced cytotoxicity to the premetastatic lungs and enhance metastasis (24). Hypoxic breast cancer cells secrete LOX, which leads to premetastatic osteolytic lesions and promotes bone metastases through NFATc1-driven osteoclastogenesis independent of RANK ligand (60). For hepatocellular carcinomas, hypoxia and TGFβ induce LOXL2 in the primary tumor and in patient sera, thereby increasing tissue stiffness and promoting cancer cell adhesion and metastasis (61). As an indirect mechanism, hypoxia-induced expression of carbonic anhydrase IX in breast cancer cells leads to the secretion of G-CSF, which mobilizes granulocytic MDSCs to premetastatic lungs and promotes metastasis (62). The effects of other types of stresses in the primary tumors, such as nutrient deprivation, on the establishment of a premetastatic niche are yet to be identified.

## Tumor-Derived Formation of a Premetastatic Niche

## Tumor-secreted EVs (including exosomes)

EVs are released into the extracellular environment by many cell types, including cancer cells. These membrane-encapsulated structures can transfer a variety of cellular materials, including RNA, DNA, and proteins, between adjacent or distant cells (upon systemic delivery via the circulation; refs. 63-66). Many recent studies on EVs focus on exosomes, a subset of EVs that are 30 to 100 nm with an endocytic origin. Cancer cells have been noted for their enhanced secretion of exosomes with altered contents in comparison to their noncancerous counterparts, and as a result cancer-specific serum, exosome miRNAs, and proteins have been proposed as biomarkers for cancer (67-71). Recent studies indicate that exosomes contain fibronectin on their external surface, which facilitated interaction with target cells through heparin sulfate (72). As the accumulation of fibronectin in the premetastatic niche is one of the earliest stages of premetastatic niche formation, exosomes may be the earliest drivers of premetastatic niche formation. Metastatic cancer cells secrete exosomes from their leading edge to promote adhesion and enhance directional trafficking (73). Whether this occurs in vivo remains to be seen. Cancer-secreted EVs can be internalized by other cell types in the primary tumor microenvironment and pre-/metastatic niches. Cargos loaded in these EVs, which to a certain extent reflect the molecular alterations in cancer cells, can be transferred to recipient niche cells to exert profound effects (74-76). Recent EV-tracing studies have indicated that melanoma-derived EVs primarily travel to the tumor-draining lymph nodes, where they are taken up by a protective barrier of subcapsular sinus CD169<sup>+</sup>CD11b<sup>+</sup>SSC<sup>low</sup> macrophages (77). This protective barrier can be compromised by tumor progression or by anticancer treatments, allowing tumor EVs to interact with B cells in the tumor-draining lymph nodes, promoting tumor progression. On the other hand, noncancerous cells in a cancer-hosting niche also secreted EVs to influence cancer behaviors (78, 79). A recent study indicates that cancer-secreted exosomes arriving to a premetastatic niche follow the tropism of their parent cells and that this organotropism in exosome homing is partially determined by the exosomal integrin profile. Mice pretreated with lung-tropic exosomes can shift the metastatic preference of bone-tropic cells to the lungs (80). Some recently reported EVmediated mechanisms that can contribute to the complex intercellular communications at a premetastatic niche are summarized in Table 1.

#### Nonvesicular tumor-secreted factors

Tumor-secreted factors, such as G-CSF, OPN, SDF-1, TNFα, TGFβ, VEGF-A, and PIGF, have long been known to influence a metastatic niche through inducing inflammation, remodeling ECM, altering niche cells, and recruiting immune cells (8, 10, 29, 50). A thorough summary of these factors and their effects on the premetastatic niche has been published (8). More recent work has shown that factors secreted by hypoxic tumor cells, including LOX, LOXL2, and G-CSF, direct a prometastatic niche reprogramming (60-62). LOXL2 has also been shown to collaborate with E47 to induce EMT and increase the secretion of GM-CSF to recruit BMDCs to premetastatic lungs (26). VEGF, another hypoxia-induced factor, has been known to play an important role in cancer growth and metastasis through the induction of angiogenesis in the primary tumor. Tumor-secreted VEGF also increases angiogenesis and recruits MDSCs to the premetastatic lungs through the induction of COX-2 and its downstream target PGE2 in pulmonary endothelial cells (50). In addition, VEGF secreted by metastatic cancer cells can disrupt the BBB by inducing angiopoietin-2 in brain microvascular endothelial cells (49). Together, these studies suggest that the formation of a premetastatic niche may begin as soon as the primary tumor grows large enough for the formation of hypoxic regions and systemic dissemination of hypoxia-induced factors.

## **Clinical Implications**

Many cancer-associated circulating exosomal markers, including those listed in Table 1, have shown promise as a noninvasive means of assessing the metastatic potential of the primary tumor. Serum levels of miR-105 and miR-122 have been shown to be prognostic indicators for metastasis in early-stage breast cancer patients (20, 81). Exosomal miR-181c has been shown to be increased in patients with brain metastases; however, it is

Table 1. Recently reported EV-mediated adaptations in a premetastatic niche

Effector cells	Target cells	EV cargos	Effects	References
Glioblastoma cells	Brain endothelial cells; glioma cells	mRNA (including EGFRvIII), miRNA, angiogenic proteins	Stimulate angiogenesis and glioma cell proliferation	Skog et al. (64)
Melanoma cells	Bone marrow progenitors	MET, TYRP2, VLA-4, HSP70, an HSP90 isoform	Induce vascular leakiness; educate BMDCs to be proangiogenic	Peinado et al. (82)
Multiple types of cancer cells	Endothelial cells	miR-9	Promote endothelial cell migration and tumor angiogenesis	Zhuang et al. (89)
Breast cancer cells	Endothelial cells	miR-105	Induce vascular leakiness	Zhou et al. (20)
Brain metastatic breast cancer cells	Brain endothelial cells	miR-181c	Destroy BBB to promote brain metastases	Tominaga et al. (47)
Breast cancer cells; ovarian cancer cells	Macrophages; monocytes; DCs	Palmitoylated proteins;?a	Induce NFκB- and STAT3-target cytokines	Chow et al. (30); Bretz et al. (90)
Lung cancer cells	Immune cells	miR-21 and miR-29a	Trigger a TLR-mediated prometastatic inflammatory response	Fabbri et al. (91)
Melanoma cells	Sentinel lymph nodes	?	Induce angiogenic pathways, ECM modification, and cancer cell recruitment	Hood et al. (92)
Multiple types of cancer cells	MDSCs	Hsp72	Induce STAT3-dependent immunosuppression	Chalmin et al. (93)
Pancreatic cancer cells	Kupffer cells	MIF	Induce TGFβ secretion and fibronectin production by hepatic stellate cells	Costa-Silva et al. (45)
Lung and pancreatic cancer cells	Myoblasts	miR-21	Promote muscle cell death and cachexia	He et al. (94)
Multiple types of cancer cells	Stromal fibroblasts	TGFβ	Promote differentiation into myofibroblasts	Webber et al. (95)
Breast cancer cells Prostate cancer cells	Lung fibroblasts, astrocytes Prostate fibroblasts	miR-122 miR-100, miR-21, etc.	Suppress glucose metabolism Increase MMP and RANKL expression and fibroblast migration	Fong et al. (19) Sanchez et al. (96)
Pancreatic cancer cells	Lung fibroblasts, lymph node cells, bone marrow cells, endothelial cells	? (require other soluble factors)	Reprogram gene expression to promote metastasis	Jung et al. (97)
Metastatic breast cancer cells	Nonmetastatic breast cancer cells	miR-200	Promote EMT and metastasis	Le et al. (98)
Stromal fibroblast	Breast cancer cells	Cd81	Mobilize autocrine Wnt-planar cell polarity signaling to drive metastasis	Luga et al. (78)
Astrocytes	Breast cancer cells	PTEN-targeting miRNAs	Downregulate PTEN to promote brain metastasis	Zhang et al. (79)

a "?" indicates undetermined/unknown in the cited reference.

unknown whether it is increased at a premetastatic phase (47). Exosomal MET, pMET, TYRP2, VLA-4, and HSP70 have shown a remarkable prognostic value in patients with melanoma (82), whereas exosomal levels of ITG $\beta_4$  and ITG $\alpha_v$  in breast and pancreatic cancer patients at a premetastatic stage are respectively associated with organotropic metastases to the lungs and liver (80).

One of the challenges of studying exosomes *in vivo* is that the exosomes circulating in the blood originate from multiple cell types, including both normal and cancer cells. Glypican-1 (GPC1) has been proposed as a marker of cancer-derived exosomes and has shown promise in the early detection of pancreatic cancer (67). GPC1 outclasses the current clinical standard carbohydrate antigen 19-9 ELISA in discriminating benign pancreatic disease and healthy individuals from patients with pancreatic cancer precursor lesions. Although exosome collection and screening do require more time and procedures than standard serum screens, the exosomal markers offer greater specificity and sensitivity in comparison with unfractionated serum (67).

The enhanced permeability and retention effect (EPR) describes the retention of large (>40–50 kDa) macromolecules within the tumor due to its abnormal vasculature. It is unclear

whether the EPR effect applies to exosomes. Studies have shown that exosome-sized nanoparticles exhibit the EPR effect (83), but the notable increase of cancer-derived exosomes in patient blood indicates that the primary tumor is able to release a substantial number of exosomes, which are not being substantially retained by the tumor. Although established metastases show the EPR effect (84), it is not yet known whether the vasculature in the premetastatic niche may become transformed enough to display this effect before the arrival of cancer cells. This possibility needs to be further elucidated as a potential mechanism that may influence subsequent tissue uptake of exosomes as well as the delivery and retention of therapeutic agents in a premetastatic niche.

Factors and pathways driving the tumor-directed reprogramming of normal niche cells during the establishment of a premetastatic niche are potential therapeutic targets for the prevention and early treatment of metastases. Trebananib targeting Ang-1/2 has been incapable of extending the life of cancer patients receiving chemotherapy, except for ovarian cancer (85–87). However, recent studies suggest that the drug may be used to target brain metastasis. As discussed earlier, cancer cells can induce Ang-2 in brain endothelial cells through the

secretion of VEGF; inhibition of Ang-2 with trebananib reduces tumor-induced BBB disruption in mice (49). Further work needs to be done to determine whether trebananib may increase the survival of patients with brain metastases.

Promising results have been seen in preclinical models with LOX inhibition and the bisphosphonate zoledronic acid in decreasing osteolytic lesions and bone metastasis (60), and with the Alox5 inhibitor zileuton in decreasing leukotrienepromoted lung metastasis (27). Several studies have proposed therapies that inhibit the recruitment of immune cells to the premetastatic niche, including targeting CXCR2 to decrease platelet-mediated granulocyte recruitment (35), C5aR to decrease Treg recruitment (37), and COX-2 to decrease DC recruitment (33). Recent evidence, however, suggests that caution should be used regarding therapies that inhibit the release of immune cells into the circulation (88). CCL2 inhibition is found to reduce metastasis by inhibiting the mobilization of monocytes from bone marrow; however, termination of CCL2 inhibition led to a larger release of monocytes into the blood as well as increased angiogenesis and cancer cell proliferation in the lungs, resulting in reduced survival in treated mice compared with untreated mice (88). Therapies targeting the mobilization of immune cells may need to be given for a prolonged period and combined with other therapies that would overcome the adverse effects. This also suggests that the tumor microenvironments (including premetastatic niches) and other organs harboring tumor-promoting cells (such as the bone marrow) undergo dynamic remodeling in response to targeted therapies, which may result in unpredictable clinical outcome, and these agents need to be carefully evaluated in preclinical models. Nevertheless, further characterization of the causes and phenotypes of premetastatic niches would reveal additional markers with diagnostic and prognostic values and guide the development of new therapies to simultaneously target cancer cells and the premetastatic niche to control cancer metastasis.

#### **Disclosure of Potential Conflicts of Interest**

No potential conflicts of interest were disclosed.

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#### References

- Kaplan RN, Riba RD, Zacharoulis S, Bramley AH, Vincent L, Costa C, et al. VEGFR1-positive haematopoietic bone marrow progenitors initiate the pre-metastatic niche. Nature 2005;438:820-7.
- Kaplan RN, Rafii S, Lyden D. Preparing the "soil": the premetastatic niche. Cancer Res 2006;66:11089–93.
- Valastyan S, Weinberg RA. Tumor metastasis: molecular insights and evolving paradigms. Cell 2011;147:275–92.
- 4. Gupta GP, Massague J. Cancer metastasis: building a framework. Cell 2006;127:679–95.
- Steeg PS. Tumor metastasis: mechanistic insights and clinical challenges. Nat Med 2006;12:895–904.
- Sethi N, Kang Y. Unravelling the complexity of metastasis molecular understanding and targeted therapies. Nat Rev Cancer 2011;11:735–48.
- Chambers AF, Groom AC, MacDonald IC. Dissemination and growth of cancer cells in metastatic sites. Nat Rev Cancer 2002;2:563–72.
- McAllister SS, Weinberg RA. The tumour-induced systemic environment as a critical regulator of cancer progression and metastasis. Nat Cell Biol 2014;16:717–27.
- Psaila B, Lyden D. The metastatic niche: adapting the foreign soil. Nat Rev Cancer 2009;9:285–93.
- Peinado H, Lavotshkin S, Lyden D. The secreted factors responsible for premetastatic niche formation: old sayings and new thoughts. Semin Cancer Biol 2011;21:139–46.
- $11. \ \ Paget S. The \ distribution of secondary growths in cancer of the breast. 1889.$  Cancer Metastasis Rev 1989;8:98–101.
- 12. Alix-Panabieres C, Riethdorf S, Pantel K. Circulating tumor cells and bone marrow micrometastasis. Clin Cancer Res 2008;14:5013–21.
- Podsypanina K, Du YC, Jechlinger M, Beverly LJ, Hambardzumyan D, Varmus H. Seeding and propagation of untransformed mouse mammary cells in the lung. Science 2008;321:1841–4.
- Erler JT, Bennewith KL, Cox TR, Lang G, Bird D, Koong A, et al. Hypoxiainduced lysyl oxidase is a critical mediator of bone marrow cell recruitment to form the premetastatic niche. Cancer Cell 2009;15:35–44.
- Hiratsuka S, Ishibashi S, Tomita T, Watanabe A, Akashi-Takamura S, Murakami M, et al. Primary tumours modulate innate immune signalling to create pre-metastatic vascular hyperpermeability foci. Nat Commun 2013;4:1853.

- Hiratsuka S, Goel S, Kamoun WS, Maru Y, Fukumura D, Duda DG, et al. Endothelial focal adhesion kinase mediates cancer cell homing to discrete regions of the lungs via E-selectin up-regulation. Proc Natl Acad Sci U S A 2011;108:3725–30.
- Hiratsuka S, Watanabe A, Aburatani H, Maru Y. Tumour-mediated upregulation of chemoattractants and recruitment of myeloid cells predetermines lung metastasis. Nat Cell Biol 2006;8:1369–75.
- Hiratsuka S, Watanabe A, Sakurai Y, Akashi-Takamura S, Ishibashi S, Miyake K, et al. The S100A8-serum amyloid A3-TLR4 paracrine cascade establishes a pre-metastatic phase. Nat Cell Biol 2008;10: 1349–55.
- Fong MY, Zhou W, Liu L, Alontaga AY, Chandra M, Ashby J, et al. Breastcancer-secreted miR-122 reprograms glucose metabolism in premetastatic niche to promote metastasis. Nat Cell Biol 2015;17:183–94.
- Zhou W, Fong MY, Min Y, Somlo G, Liu L, Palomares MR, et al. Cancersecreted miR-105 destroys vascular endothelial barriers to promote metastasis. Cancer Cell 2014;25:501–15.
- Kim MY, Oskarsson T, Acharyya S, Nguyen DX, Zhang XH, Norton L, et al. Tumor self-seeding by circulating cancer cells. Cell 2009;139:1315–26.
- Chen Y, Gou X, Kong DK, Wang X, Wang J, Chen Z, et al. EMMPRIN regulates tumor growth and metastasis by recruiting bone marrow-derived cells through paracrine signaling of SDF-1 and VEGF. Oncotarget 2015;6: 32575–85.
- Deng J, Liu Y, Lee H, Herrmann A, Zhang W, Zhang C, et al. S1PR1-STAT3 signaling is crucial for myeloid cell colonization at future metastatic sites. Cancer Cell 2012:21:642–54.
- Sceneay J, Chow MT, Chen A, Halse HM, Wong CS, Andrews DM, et al. Primary tumor hypoxia recruits CD11b+/Ly6Cmed/Ly6G+ immune suppressor cells and compromises NK cell cytotoxicity in the premetastatic niche. Cancer Res 2012;72:3906–11.
- Yan HH, Jiang J, Pang Y, Achyut BR, Lizardo M, Liang X, et al. CCL9 induced by TGFbeta signaling in myeloid cells enhances tumor cell survival in the premetastatic organ. Cancer Res 2015;75:5283–98.
- Canesin G, Cuevas EP, Santos V, Lopez-Menendez C, Moreno-Bueno G, Huang Y, et al. Lysyl oxidase-like 2 (LOXL2) and E47 EMT factor: novel partners in E-cadherin repression and early metastasis colonization. Oncogene 2015;34:951–64.

Clin Cancer Res; 22(15) August 1, 2016

- 27. Wculek SK, Malanchi I. Neutrophils support lung colonization of metastasis-initiating breast cancer cells. Nature 2015;528:413–7.
- Headley MB, Bins A, Nip A, Roberts EW, Looney MR, Gerard A, et al. Visualization of immediate immune responses to pioneer metastatic cells in the lung. Nature 2016;531:513–7.
- 29. Coffelt SB, Kersten K, Doornebal CW, Weiden J, Vrijland K, Hau CS, et al. IL-17-producing gammadelta T cells and neutrophils conspire to promote breast cancer metastasis. Nature 2015;522:345–8.
- Chow A, Zhou W, Liu L, Fong MY, Champer J, Van Haute D, et al. Macrophage immunomodulation by breast cancer-derived exosomes requires Toll-like receptor 2-mediated activation of NF-kappaB. Sci Rep 2014;4:5750.
- Nielsen SR, Quaranta V, Linford A, Emeagi P, Rainer C, Santos A, et al. Macrophage-secreted granulin supports pancreatic cancer metastasis by inducing liver fibrosis. Nat Cell Biol 2016;18:549–60.
- 32. Rohan TE, Xue X, Lin HM, D'Alfonso TM, Ginter PS, Oktay MH, et al. Tumor microenvironment of metastasis and risk of distant metastasis of breast cancer. I Natl Cancer Inst 2014:106:pii:diu136.
- Ogawa F, Amano H, Eshima K, Ito Y, Matsui Y, Hosono K, et al. Prostanoid induces premetastatic niche in regional lymph nodes. J Clin Invest 2014;124:4882–94.
- Hanna RN, Cekic C, Sag D, Tacke R, Thomas GD, Nowyhed H, et al. Patrolling monocytes control tumor metastasis to the lung. Science 2015; 350:985–90.
- Labelle M, Begum S, Hynes RO. Platelets guide the formation of early metastatic niches. Proc Natl Acad Sci U S A 2014;111:E3053–61.
- Zhang W, Zhang C, Li W, Deng J, Herrmann A, Priceman SJ, et al. CD8+ T-cell immunosurveillance constrains lymphoid premetastatic myeloid cell accumulation. Eur J Immunol 2015;45:71–81.
- Vadrevu SK, Chintala NK, Sharma SK, Sharma P, Cleveland C, Riediger L, et al. Complement c5a receptor facilitates cancer metastasis by altering Tcell responses in the metastatic niche. Cancer Res 2014;74:3454–65.
- Sharma SK, Chintala NK, Vadrevu SK, Patel J, Karbowniczek M, Markiewski MM. Pulmonary alveolar macrophages contribute to the premetastatic niche by suppressing antitumor T cell responses in the lungs. J Immunol 2015;194:5529–38.
- Grum-Schwensen B, Klingelhofer J, Beck M, Bonefeld CM, Hamerlik P, Guldberg P, et al. S100A4-neutralizing antibody suppresses spontaneous tumor progression, pre-metastatic niche formation and alters T-cell polarization balance. BMC Cancer 2015;15:44.
- Qi F, He T, Jia L, Song N, Guo L, Ma X, et al. The miR-30 family inhibits pulmonary vascular hyperpermeability in the premetastatic phase by direct targeting of Skp2. Clin Cancer Res 2015;21:3071–80.
- 41. Cox TR, Bird D, Baker AM, Barker HE, Ho MW, Lang G, et al. LOX-mediated collagen crosslinking is responsible for fibrosis-enhanced metastasis. Cancer Res 2013;73:1721–32.
- 42. Luo X, Fu Y, Loza AJ, Murali B, Leahy KM, Ruhland MK, et al. Stromal-initiated changes in the bone promote metastatic niche development. Cell Rep 2016;14:82–92.
- van Deventer HW, Palmieri DA, Wu QP, McCook EC, Serody JS. Circulating fibrocytes prepare the lung for cancer metastasis by recruiting Ly-6C+ monocytes via CCL2. J Immunol 2013;190:4861–7.
- Duda DG, Duyverman AM, Kohno M, Snuderl M, Steller EJ, Fukumura D, et al. Malignant cells facilitate lung metastasis by bringing their own soil. Proc Natl Acad Sci U S A 2010;107:21677–82.
- Costa-Silva B, Aiello NM, Ocean AJ, Singh S, Zhang H, Thakur BK, et al. Pancreatic cancer exosomes initiate pre-metastatic niche formation in the liver. Nat Cell Biol 2015;17:816–26.
- 46. Lu P, Weaver VM, Werb Z. The extracellular matrix: a dynamic niche in cancer progression. J Cell Biol 2012;196:395–406.
- Tominaga N, Kosaka N, Ono M, Katsuda T, Yoshioka Y, Tamura K, et al. Brain metastatic cancer cells release microRNA-181c-containing extracellular vesicles capable of destructing blood-brain barrier. Nat Commun 2015;6:6716.
- Huang Y, Song N, Ding Y, Yuan S, Li X, Cai H, et al. Pulmonary vascular destabilization in the premetastatic phase facilitates lung metastasis. Cancer Res 2009:69:7529–37.
- Avraham HK, Jiang S, Fu Y, Nakshatri H, Ovadia H, Avraham S. Angiopoietin-2 mediates blood-brain barrier impairment and colonization of triple-negative breast cancer cells in brain. J Pathol 2014; 232:369–81.

- Liu S, Jiang M, Zhao Q, Li S, Peng Y, Zhang P, et al. Vascular endothelial growth factor plays a critical role in the formation of the pre-metastatic niche via prostaglandin E2. Oncol Rep 2014;32:2477–84.
- 51. Martinez LM, Vallone VB, Labovsky V, Choi H, Hofer EL, Feldman L, et al. Changes in the peripheral blood and bone marrow from untreated advanced breast cancer patients that are associated with the establishment of bone metastases. Clin Exp Metastasis 2014;31:213–32.
- Pal SK, Vuong W, Zhang W, Deng J, Liu X, Carmichael C, et al. Clinical and translational assessment of VEGFR1 as a mediator of the premetastatic niche in high-risk localized prostate cancer. Mol Cancer Ther 2015;14: 2896–900.
- Garnier D, Magnus N, Lee TH, Bentley V, Meehan B, Milsom C, et al. Cancer cells induced to express mesenchymal phenotype release exosome-like extracellular vesicles carrying tissue factor. J Biol Chem 2012; 287:43565–72.
- 54. Gopal SK, Greening DW, Hanssen EG, Zhu HJ, Simpson RJ, Mathias RA. Oncogenic epithelial cell-derived exosomes containing Rac1 and PAK2 induce angiogenesis in recipient endothelial cells. Oncotarget. 2016 Feb 22. [Epub ahead of print].
- Tauro BJ, Mathias RA, Greening DW, Gopal SK, Ji H, Kapp EA, et al. Oncogenic H-ras reprograms Madin-Darby canine kidney (MDCK) cell-derived exosomal proteins following epithelial-mesenchymal transition. Mol Cell Proteomics 2013;12:2148–59.
- 56. Pasquier J, Thawadi HA, Ghiabi P, Abu-Kaoud N, Maleki M, Guerrouahen BS, et al. Microparticles mediated cross-talk between tumoral and endothelial cells promote the constitution of a pro-metastatic vascular niche through Arf6 up regulation. Cancer Microenviron 2014;7:41–59.
- 57. Lee E, Fertig EJ, Jin K, Sukumar S, Pandey NB, Popel AS. Breast cancer cells condition lymphatic endothelial cells within pre-metastatic niches to promote metastasis. Nat Commun 2014;5:4715.
- Lee E, Pandey NB, Popel AS. Pre-treatment of mice with tumor-conditioned media accelerates metastasis to lymph nodes and lungs: a new spontaneous breast cancer metastasis model. Clin Exp Metastasis 2014;31:67–79.
- Loo JM, Scherl A, Nguyen A, Man FY, Weinberg E, Zeng Z, et al. Extracellular metabolic energetics can promote cancer progression. Cell 2015;160:393–406.
- Cox TR, Rumney RM, Schoof EM, Perryman L, Hoye AM, Agrawal A, et al. The hypoxic cancer secretome induces pre-metastatic bone lesions through lysyl oxidase. Nature 2015;522:106–10.
- Wong CC, Tse AP, Huang YP, Zhu YT, Chiu DK, Lai RK, et al. Lysyl oxidaselike 2 is critical to tumor microenvironment and metastatic niche formation in hepatocellular carcinoma. Hepatology 2014;60:1645–58.
- Chafe SC, Lou Y, Sceneay J, Vallejo M, Hamilton MJ, McDonald PC, et al. Carbonic anhydrase IX promotes myeloid-derived suppressor cell mobilization and establishment of a metastatic niche by stimulating G-CSF production. Cancer Res 2015;75:996–1008.
- Valadi H, Ekstrom K, Bossios A, Sjostrand M, Lee JJ, Lotvall JO. Exosomemediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. Nat Cell Biol 2007;9:654–9.
- Skog J, Wurdinger T, van Rijn S, Meijer DH, Gainche L, Sena-Esteves M, et al. Glioblastoma microvesicles transport RNA and proteins that promote tumour growth and provide diagnostic biomarkers. Nat Cell Biol 2008:10:1470-6.
- Thery C, Zitvogel L, Amigorena S. Exosomes: composition, biogenesis and function. Nat Rev Immunol 2002;2:569–79.
- Redzic JS, Balaj L, van der Vos KE, Breakefield XO. Extracellular RNA mediates and marks cancer progression. Semin Cancer Biol 2014;28: 14–23.
- 67. Melo SA, Luecke LB, Kahlert C, Fernandez AF, Gammon ST, Kaye J, et al. Glypican-1 identifies cancer exosomes and detects early pancreatic cancer. Nature 2015;523:177–82.
- Taylor DD, Gercel-Taylor C. MicroRNA signatures of tumor-derived exosomes as diagnostic biomarkers of ovarian cancer. Gynecol Oncol 2008; 110:13–21.
- Duijvesz D, Luider T, Bangma CH, Jenster G. Exosomes as biomarker treasure chests for prostate cancer. Eur Urol 2011;59:823–31.
- Ogata-Kawata H, Izumiya M, Kurioka D, Honma Y, Yamada Y, Furuta K, et al. Circulating exosomal microRNAs as biomarkers of colon cancer. PLoS One 2014;9:e92921.
- 71. Cheng L, Sharples RA, Scicluna BJ, Hill AF. Exosomes provide a protective and enriched source of miRNA for biomarker profiling

3732 Clin Cancer Res; 22(15) August 1, 2016 Clinical Cancer Research

- compared to intracellular and cell-free blood. J Extracell Vesicles 2014;3. doi: 10.3402/jev.v3.23743.
- 72. Purushothaman A, Bandari SK, Liu J, Mobley JA, Brown EE, Sanderson RD. Fibronectin on the surface of myeloma cell-derived exosomes mediates exosome-cell interactions. J Biol Chem 2016;291:1652–63.
- 73. Sung BH, Ketova T, Hoshino D, Zijlstra A, Weaver AM. Directional cell movement through tissues is controlled by exosome secretion. Nat Commun 2015;6:7164.
- Thuma F, Zoller M. Outsmart tumor exosomes to steal the cancer initiating cell its niche. Semin Cancer Biol 2014;28:39–50.
- Roma-Rodrigues C, Fernandes AR, Baptista PV. Exosome in tumour microenvironment: overview of the crosstalk between normal and cancer cells. Biomed Res Int 2014;2014:179486.
- Aleckovic M, Kang Y. Regulation of cancer metastasis by cell-free miRNAs. Biochim Biophys Acta 2015;1855;24–42.
- Pucci F, Garris C, Lai CP, Newton A, Pfirschke C, Engblom C, et al. SCS macrophages suppress melanoma by restricting tumor-derived vesicle-B cell interactions. Science 2016;352:242–6.
- Luga V, Zhang L, Viloria-Petit AM, Ogunjimi AA, Inanlou MR, Chiu E, et al. Exosomes mediate stromal mobilization of autocrine Wnt-PCP signaling in breast cancer cell migration. Cell 2012;151:1542–56.
- Zhang L, Zhang S, Yao J, Lowery FJ, Zhang Q, Huang WC, et al. Microenvironment-induced PTEN loss by exosomal microRNA primes brain metastasis outgrowth. Nature 2015;527:100–4.
- 80. Hoshino A, Costa-Silva B, Shen TL, Rodrigues G, Hashimoto A, Tesic Mark M, et al. Tumour exosome integrins determine organotropic metastasis. Nature 2015;527:329–35.
- 81. Wu X, Somlo G, Yu Y, Palomares MR, Li AX, Zhou W, et al. De novo sequencing of circulating miRNAs identifies novel markers predicting clinical outcome of locally advanced breast cancer. J Transl Med 2012;10:42.
- 82. Peinado H, Aleckovic M, Lavotshkin S, Matei I, Costa-Silva B, Moreno-Bueno G, et al. Melanoma exosomes educate bone marrow progenitor cells toward a pro-metastatic phenotype through MET. Nat Med 2012;18:883–91.
- 83. Sun D, Zhuang X, Zhang S, Deng ZB, Grizzle W, Miller D, et al. Exosomes are endogenous nanoparticles that can deliver biological information between cells. Adv Drug Deliv Rev 2013;65:342–7.
- Maeda H. Toward a full understanding of the EPR effect in primary and metastatic tumors as well as issues related to its heterogeneity. Adv Drug Deliv Rev 2015;91:3–6.
- Monk BJ, Poveda A, Vergote I, Raspagliesi F, Fujiwara K, Bae DS, et al. Anti-angiopoietin therapy with trebananib for recurrent ovarian cancer (TRINOVA-1): a randomised, multicentre, double-blind, placebo-controlled phase 3 trial. Lancet Oncol 2014;15:799–808.
- 86. Dieras V, Wildiers H, Jassem J, Dirix LY, Guastalla JP, Bono P, et al. Trebananib (AMG 386) plus weekly paclitaxel with or without beva-

- cizumab as first-line therapy for HER2-negative locally recurrent or metastatic breast cancer: a phase 2 randomized study. Breast 2015;24: 182–90.
- Peeters M, Strickland AH, Lichinitser M, Suresh AV, Manikhas G, Shapiro J, et al. A randomised, double-blind, placebo-controlled phase 2 study of trebananib (AMG 386) in combination with FOLFIRI in patients with previously treated metastatic colorectal carcinoma. Br J Cancer 2013;108: 503–11.
- 88. Bonapace L, Coissieux MM, Wyckoff J, Mertz KD, Varga Z, Junt T, et al. Cessation of CCL2 inhibition accelerates breast cancer metastasis by promoting angiogenesis. Nature 2014;515:130–3.
- Zhuang G, Wu X, Jiang Z, Kasman I, Yao J, Guan Y, et al. Tumour-secreted miR-9 promotes endothelial cell migration and angiogenesis by activating the JAK-STAT pathway. EMBO J 2012;31:3513–23.
- Bretz NP, Ridinger J, Rupp AK, Rimbach K, Keller S, Rupp C, et al. Body fluid exosomes promote secretion of inflammatory cytokines in monocytic cells via Toll-like receptor signaling. J Biol Chem 2013; 288:36691–702.
- 91. Fabbri M, Paone A, Calore F, Galli R, Gaudio E, Santhanam R, et al. MicroRNAs bind to Toll-like receptors to induce prometastatic inflammatory response. Proc Natl Acad Sci U S A 2012;109:E2110–6.
- Hood JL, San RS, Wickline SA. Exosomes released by melanoma cells prepare sentinel lymph nodes for tumor metastasis. Cancer Res 2011;71: 3792–801.
- Chalmin F, Ladoire S, Mignot G, Vincent J, Bruchard M, Remy-Martin JP, et al. Membrane-associated Hsp72 from tumor-derived exosomes mediates STAT3-dependent immunosuppressive function of mouse and human myeloid-derived suppressor cells. J Clin Invest 2010;120: 457–71.
- 94. He WA, Calore F, Londhe P, Canella A, Guttridge DC, Croce CM. Microvesicles containing miRNAs promote muscle cell death in cancer cachexia via TLR7. Proc Natl Acad Sci U S A 2014;111:4525–9.
- Webber J, Steadman R, Mason MD, Tabi Z, Clayton A. Cancer exosomes trigger fibroblast to myofibroblast differentiation. Cancer Res 2010;70: 9621–30.
- 96. Sanchez CA, Andahur EI, Valenzuela R, Castellon EA, Fulla JA, Ramos CG, et al. Exosomes from bulk and stem cells from human prostate cancer have a differential microRNA content that contributes cooperatively over local and pre-metastatic niche. Oncotarget 2016;7: 3993–4008.
- 97. Jung T, Castellana D, Klingbeil P, Cuesta Hernandez I, Vitacolonna M, Orlicky DJ, et al. CD44v6 dependence of premetastatic niche preparation by exosomes. Neoplasia 2009;11:1093–105.
- Le MT, Hamar P, Guo C, Basar E, Perdigao-Henriques R, Balaj L, et al. miR-200-containing extracellular vesicles promote breast cancer cell metastasis.
   J Clin Invest 2014;124:5109–28.



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# Cancer Tills the Premetastatic Field: Mechanistic Basis and Clinical **Implications**

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