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Possibility and Challenges of Conversion of Current Virus Species Names to Linnaean Binomials

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Abstract.—Botanical, mycological, zoological, and prokaryotic species names follow the Linnaean format, consisting of an italicized Latinized binomen with a capitalized genus name and a lower case species epithet (e.g., Homo sapiens). Virus species names, however, do not follow a uniform format, and, even when binomial, are not Linnaean in style. In this thought exercise, we attempted to convert all currently official names of species included in the virus family Arenaviridae and the virus order Mononegavirales to Linnaean binomials, and to identify and address associated challenges and concerns. Surprisingly, this endeavor was not as complicated or time-consuming as even the authors of this article expected when conceiving the experiment. [Arenaviridae; binomials; ICTV; International Committee on Taxonomy of Viruses; Mononegavirales; virus nomenclature; virus taxonomy.]

Botanical, mycological, zoological, and prokaryotic species names follow a Latinized binomi(n)al format (i.e., binomial nomenclature) first introduced, formally and systematically, by Carl Linnaeus in 1753 (Linnaeus 1753). This format consists of two italicized words (a binomen or binary combination or scientific/Latin name), with the first capitalized word naming the genus to which the species belongs ("genus name") and the second lower case word denoting the species ("specific

name/species epithet") (International Committee on Systematic Bacteriology 1992; International Association for Plant Taxonomy 2011; International Commission on Zoological Nomenclature 2012). Typical examples for binomial species names are *Arabidopsis thaliana*, *Saccharomyces cerevisiae*, *Homo sapiens*, or *Escherichia coli*. Such species names have several advantages over most alternative species-naming conventions, including: (i) they are internationally recognizable as they do not

change in typography even in texts using non-Latin alphabets or other scripts; (ii) they are easier to remember than alphanumerical schemes or "names"; (iii) they may be explicative, that is, each part of the binomial name may relate specific characteristics of species members; (iv) they are in an extinct language that evolves very slowly if at all, ensuring the stability of names and of their etymology; and (v) they frequently differ from the names of species members (e.g., *Homo sapiens* ↔ humans), thereby emphasizing the logical difference between taxon (a concept of the mind) and taxon member (a concrete physical entity) (Drebot et al. 2002; Calisher and Mahy 2003; Van Regenmortel 2003, 2006, 2007, 2016a; Kuhn and Jahrling 2010).

In contrast, viral species names do not follow a uniform format, except for a requirement to be italicized and to have the first word capitalized (King et al. 2012; International Committee on Taxonomy of Viruses 2013). The few existing binomial virus species names are not Latinized and typically begin with the capitalized species epithet followed by the lower case name of the genus to which the species belongs (e.g., Lassa mammarenavirus), thus inverting the order of the Linnaean format. The mismatch between the various formats of virus species names and the species name format of all other official taxonomies reflects the complicated history of virus taxonomy and nomenclature.

Importantly, the lack of a uniform virus speciesnaming scheme impedes the development comprehensive biological databases. Because all species names currently in use for animals, fungi, plants, prokaryotes, and protozoans follow the Linnaean binomial format, software can easily recognize the genus name and species epithet in a properly formatted database entry: the first word in the < species > entry field will be the genus name, the second, following a space, will be the species epithet. A virus species name that is not following the Linnaean format will be parsed erroneously. For instance, a species name such as the current "Measles virus" will lead the software to assume that the species epithet "Measles" is a genus and that the word "virus" is a species epithet. Likewise, a species name such as the current non-Latinized binomial "Lassa mammarenavirus" will lead the software to assume that "Lassa" and "mammarenavirus" are genus and species epithet, respectively, rather than vice versa ("Mammarenavirus" is the genus in current virus taxonomy). Consequently, virus taxonomy is currently excluded from many bioinformatic projects, such as BioCode (Greuter et al. 2011). Alignment of virus taxonomy with other taxonomies was originally considered by the founders of the International Committee on Nomenclature of Viruses (ICNV), the predecessor of the International Committee on Taxonomy of Viruses (ICTV). Numerous Linnaean species names were published in the first and second ICNV/ICTV Reports (Wildy 1971; Fenner 1976), but this practice was abandoned thereafter. Linnaean

virus species names have been commonly used in the past [e.g., "Herpesvirus simiae" (Hummeler et al. 1959; Huemer et al. 2003)] or are currently used [e.g., "Pandoravirus salinus" (Philippe et al. 2013)], but none are officially approved at present. Indeed, Matthews stated in 1985 that "[n]othing releases adrenalin more readily for many virologists than the suggestion that virus species names should be latinized [sic]" (Matthews 1985a), a view that was especially prominent among plant virologists at the time (Matthews 1983, 1985a, 1985b; Milne 1984).

During the most recent, largest-to-date ICTV meeting in February 2016, a straw poll among the attending ICTV Study Group chairs revealed an interest in discussing the advantages and disadvantages of virus species name conversion to the Linnaean format (International Committee on Taxonomy of Viruses 2016). This majority vote to revisit the issue somewhat contrasts with the published view that such a conversion "is unlikely to be a welcome alternative [to current virus species names]" (Van Regenmortel 2016a). Whether such a conversion is even plausible is unclear. Over the years, a number of potential challenges, conceptual and practical, have been postulated. One conceptual debate is whether viruses are alive, that is, considered to be organisms (for contrasting views, see Koonin and Starokadomskyy 2016; Van Regenmortel 2016a, 2016b), and, therefore, whether or not Linnaean virus species names would give the impression that this dispute has been settled in favor of viruses being part of the living world. The most relevant practical concern is the fear that Latinization is so complex that only a few experts, not accessible to typical ICTV Study Groups, could achieve it and that converting the existing virus species names would become a Herculean endeavor (Van Regenmortel 2016a).

Here we set the conceptual questions aside and, as a proof of principle, demonstrate the results of a practical team effort of devising Linnaean binomial species names for all currently accepted species of the family *Arenaviridae* and the order *Mononegavirales*. Without attempting to impose the resulting names as a new standard, we seek to provide a more solid basis for consideration of the principle of applying the Linnaean format to virus nomenclature using a case study of how such conversion could be accomplished if the virology community chooses to do so.

The family *Arenaviridae* currently includes two ICTV-approved genera and 34 species for viruses that infect bats, eulipotyphla, rodents, and snakes (Radoshitzky et al. 2015; Adams et al. 2016). All arenavirus species names are non-Latinized binomials, which led us to believe that these names could be easily converted to the Linnaean format. The order *Mononegavirales* currently includes eight ICTV-approved families, 34 genera, and 141 species for viruses that infect animals, plants, and fungi (Adams et al. 2016; Afonso et al. 2016). Mononegaviral species naming conventions differ among families and, as a whole, reflect the current nonuniformity of species naming across viral taxa; it

was *a priori* unclear whether a consistent conversion is practically achievable in this case.

FORMAL REQUIREMENTS FOR LINNAEAN BINOMIALS

Linnaean binomials are Latinized, that is, both name components follow Latin grammar, although the components themselves can be derived from languages other than Latin. The genus name must be treated as a Latin singular noun in the nominative case, whereas the species epithet may be an adjective or a noun, in both cases either in the nominative or genitive case. Correct formation of a species name, therefore, requires a basic understanding of Latin grammar. However, the grammatical complexities of virus species names are less intricate than those of nonviral taxonomies; by convention, all virus genus names end in the suffix "virus." Consequently, the question of the declension of the cognate species epithet is simplified to only one gender, as the Latin word "virus," meaning slime or poison, is a noun of the neuter gender (unexpectedly for a word ending in "-us").

SPECIES NAME CONVERSION: SPECIFIC CHALLENGES AND POSSIBLE SOLUTIONS

Conversion of True Non-Latinized Binomial Species Names to Linnaean Binomials

Most currently accepted arenaviral and several currently accepted mononegaviral species names follow a true binomial, albeit non-Linnaean, format originally proposed by van Regenmortel et al. (2010). While italicized names are not Latinized, these names consist of only two words, with the first word a capitalized species epithet and the second word a lower case genus name (e.g., Lassa mammarenavirus, Human metapneumovirus). Most viral species epithets in current non-Latinized binomials within and outside of Arenaviridae and Mononegavirales refer to geographic areas (e.g., "Lassa," "Zaire"), to host taxa (e.g., Sclerotinia), or to vernacular host names (e.g., alethinophid, human). Conversion to the Linnaean format is, therefore, straightforward and achieved by switching the word order and converting the species epithet to a Latinized word in the proper case (e.g., zairense: nominative singular neuter of the suffix "-ensis," indicating a place of origin, attached to the root "zaire"; sclerotiniae: genitive singular of "sclerotinia," after the host genus; and hominis: genitive singular of "homo," also after the host genus). One encountered problem is the presence of diacritical marks in several species names (e.g., Junín mammarenavirus, Sabiá mammarenavirus). Conversion to Linnaean binomials can be achieved, however, by either dropping diacritical marks or by changing the name to contain a geographic location not requiring a diacritic (note also that the ICTV recently ratified a Code rule to disallow the use of diacritics and other special characters in new taxon names). Examples for this type of possible conversion are shown in Tables 1 and 2 in dark green (online version only).

Conversion of Non-Latinized Binomial-like Species Names to Linnaean Binomials

Several currently accepted arenaviral mononegaviral species names are similar to binomials in structure, with a species epithet and a genus name as outlined above, but consist of more than two words (e.g., in the names Soybean cyst nematode socyvirus and Lymphocytic choriomeningitis mammarenavirus, "Soybean cyst nematode" and "Lymphocytic choriomeningitis" serve as the respective species epithets). Conversion to true binomials, therefore, requires reduction, contraction, or replacement of words in addition to Latinization. Conversion of names that contain virus host information could be achieved by, for example, referring to the host taxa: soybean cyst nematodes are members of the genus Heterodera (Soybean cyst nematode socyvirus → Socyvirus heteroderae), alfalfa is the member of the species Medicago sativa (Alfalfa dwarf cytorhabdovirus → Cytorhabdovirus medicagonis), and barley is a member of the species Hordeum vulgare (Barley yellow striate Cytorhabdovirus hordei). mosaic cytorhabdovirus → Geographical locations that consist of multiple words may be contracted using Latin words (e.g., Tai Forest ebolavirus, referring to Taï Forest in Côte d'Ivoire, could be converted to Ebolavirus silvataiense: "silva" is forest in Latin).

Other current non-Latinized binomial-like species names include numbers. These numbers are located either between species epithet and genus name (e.g., Alethinophid 1 reptarenavirus, Psittaciform 1 bornavirus) or after the genus name (e.g., Sonchus cytorhabdovirus 1). Conversion of these names would be more challenging, especially for sets of species names that differ in numbers but are otherwise identical. One straightforward way would be to convert the numbers into Latinized alphanumericals (*Psittaciform 1 bornavirus* \rightarrow *Bornavirus* alphapsittaciforme). This conversion approach may be problematic as long alphanumerical lists (e.g., alphapsittaciforme, betapsittaciforme, gammapsittaciforme, etc.) are nondescript and, therefore, difficult to memorize. A more elegant conversion could be achieved if the numbers were omitted entirely and new species names formed were based on the known properties of the associated member viruses. Examples for these types of possible conversion are shown in Tables 1 and 2 in light green (online version only).

In addition, some current non-Latinized binomiallike species names contain genus names that do not refer to existing genera. For instance, the two species names *Atlantic salmon paramyxovirus* and *Avian paramyxovirus* 2 imply that these species belong to a genus "*Paramyxovirus*," which does not exist (there is only a family *Paramyxoviridae*). These names could easily be converted to Linnaean binomials, in analogy to other non-Latinized binomials, for instance by simply replacing the genus names with the correct ones

TABLE 1. Possible Linnaean binomial names for all currently accepted arenaviral species

Current ICTV-approved species name	Possible converted Linnaean species name ^a	(Unchanged) member virus name and abbreviation
Genus Mammarenavirus		
Allpahuayo mammarenavirus	Mammarenavirus allpahuayense	Allpahuayo virus (ALLV)
Amaparí mammarenavirus	Mammarenavirus amapariense	Amaparí virus (AMAV)
Bear Canyon mammarenavirus	Mammarenavirus saltusursinense ^b	Bear Canyon virus (BCNV)
Chapare mammarenavirus	Mammarenavirus chaparense	Chapare virus (CHAPV)
Cupixi mammarenavirus	Mammarenavirus cupixense	Cupixi virus (CUPXV)
Flexal mammarenavirus	Mammarenavirus flexalense	Flexal virus (FLEV)
Gairo mammarenavirus	Mammarenavirus gairense	Gairo virus (GAIV)
Guanarito mammarenavirus	Mammarenavirus guanaritense	Guanarito virus (GTOV)
Ippy mammarenavirus	Mammarenavirus ippyense	Ippy virus (IPPYV)
Junín mammarenavirus	Mammarenavirus iuninense	Junín virus (JUNV)
Lassa mammarenavirus	Mammarenavirus lassaense	Lassa virus (LASV)
Latino mammarenavirus	Mammarenavirus latinum	Latino virus (LATV)
Lujo mammarenavirus	Mammarenavirus luioense	Lujo virus (LUJV)
Luna mammarenavirus	Mammarenavirus lunaense	Luna virus (LUAV)
Lunk mammarenavirus	Mammarenavirus lunkense	Lunk virus (LNKV)
Lymphocytic choriomeningitis	Mammarenavirus	lymphocytic choriomeningitis virus (LCMV)
mammarenavirus	choriomeningitidis	, ,
Machupo mammarenavirus	Mammarenavirus machupense	Machupo virus (MACV)
Mariental mammarenavirus	Mammarenavirus marientalense	Mariental virus (MRLV)
Merino Walk mammarenavirus	Mammarenavirus viamerinense	Merino Walk virus (MRWV)
Mobala mammarenavirus	Mammarenavirus mobalaense	Mobala virus (MOBV)
Mopeia mammarenavirus	Mammarenavirus mopeiense	Mopeia virus (MOPV), Morogoro virus (MORV)
Okahandja mammarenavirus	Mammarenavirus okahandiense	Okahandja virus (OKAV)
Oliveros mammarenavirus	Mammarenavirus oliverosense	Oliveros virus (OLVV)
Paraná mammarenavirus	Mammarenavirus paranaense	Paraná virus (PRAV)
Pichindé mammarenavirus	Mammarenavirus pichindense	Pichindé virus (PICHV)
Pirital mammarenavirus	Mammarenavirus piritalense	Pirital virus (PIRV)
Sabiá mammarenavirus	Mammarenavirus sabiaense	Sabiá virus (SBAV)
Tacaribe mammarenavirus	Mammarenavirus tacaribense	Tacaribe virus (TCRV)
Tamiami mammarenavirus	Mammarenavirus tamiamense	Tamiami virus (TMMV)
Wenzhou mammarenavirus	Mammarenavirus wenzhouense	Wēnzhōu virus (WENV)
Whitewater Arroyo	Mammarenavirus arroyense	Catarina virus (CTNV), Big Brushy Tank virus
mammarenavirus		(BBRTV), Skinner Tank virus (SKTV), Tonto Creek virus (TTCV), Whitewater Arroyo virus (WWAV)
Genus Reptarenavirus		
Alethinophid 1 reptarenavirus	Reptarenavirus portaureae ^c	Golden Gate virus (GOGV)
Alethinophid 2 reptarenavirus	Reptarenavirus helsinkii	ROUT virus (ROUTV), University of Helsinki virus (UHV)
Alethinophid 3 reptarenavirus	Reptarenavirus californiae	CAS virus (CASV)

Note: Dark green (online version only) depicts current true binomial names and their Linnaean counterparts; light green (online version only) depicts current binomial-like names and their Linnaean counterparts.

(e.g., Atlantic salmon paramyxovirus \rightarrow Atlantic salmon aquaparamyxovirus \rightarrow Aquaparamyxovirus salmonis). Examples for these types of possible conversion are shown in Tables 1 and 2 in purple (online version only).

Conversion of Non-binomial Species Names to Linnaean Binomials

Numerous mononegaviral species names are not yet in any binomial or binomial-like format and often differ from the names of their member viruses only in italicization (e.g., measles virus is a member of the species *Measles virus*). Conversion of these names to Linnaean binomials would simply require the replacement of the word "virus" with a genus name and then following the steps outlined for other names above (*Measles virus* \rightarrow *Measles morbillivirus* \rightarrow *Morbillivirus rubeolae*: "rubeola" is measles in Latin). Examples for this type of possible conversion are shown in Table 2 in purple (online version only).

^aThese names are for illustration purposes only and may not be the same as the names that ICTV Study Groups would officially propose if a Linnaean binomial species naming convention were implemented.

bsaltusursinense = "from the Bear Canyon."

^cportaureae = "of the Golden Gate."

TABLE 2. Possible Linnaean binomial names for all currently accepted mononegaviral species

TABLE 2. Possible Linnaean binomial names for all currently accepted mononegaviral species				
Current ICTV-approved species name	Possible converted Linnaean species name ^a	(Unchanged) member virus name and abbreviation		
Family Bornaviridae				
Elapid 1 bornavirus	Bornavirus elapsoideae ^b	Loveridge's garter snake virus 1 (LGSV-1)		
Mammalian 1 bornavirus	Bornavirus crocidurae	Borna disease viruses 1/2 (BoDV-1/2)		
Psittaciform 1 bornavirus	Bornavirus alphapsittaciforme	parrot bornaviruses 1/2/3/4/7 (PaBV-1/2/3/4/7)		
Psittaciform 2 bornavirus	Bornavirus betapsittaciforme	parrot bornavirus 5 (PaBV-5)		
Passeriform 1 bornavirus	Bornavirus alphapasseriforme	canary bornaviruses 1/2/3 (CnBV-1/2/3)		
Passeriform 2 bornavirus	Bornavirus betapasseriforme	estrildid finch bornavirus 1 (EsBV-1)		
Waterbird 1 bornavirus	Bornavirus avisaquaticae	aquatic bird bornaviruses 1/2 (ABBV-1/2)		
Family Filoviridae				
Lloviu cuevavirus	Cuevavirus lloviense	Lloviu virus (LLOV)		
Bundibugyo ebolavirus	Ebolavirus bundibugyoense	Bundibugyo virus (BDBV)		
Reston ebolavirus	Ebolavirus restonense	Reston virus (RESTV)		
Sudan ebolavirus	Ebolavirus sudanense	Sudan virus (SUDV)		
Taï Forest ebolavirus	Ebolavirus silvataiense	Taï Forest virus (TAFV)		
Zaire ebolavirus	Ebolavirus zairense	Ebola virus (EBOV)		
Marburg marburgvirus	Marburgvirus marburgense	Marburg virus (MARV), Ravn virus (RAVV)		
Family Mymonaviridae Sclerotinia sclerotimonavirus	Sclerotimonavirus sclerotiniae	Sclerotinia sclerotiorum negative-stranded RNA virus 1 (SsNSRV-1)		
Family Nyamiviridae				
Midway nyavirus	Nyavirus midwayense	Midway virus (MIDWV)		
Nyamanini nyavirus	Nyavirus nyamaninense	Nyamanini virus (NYMV)		
Sierra Nevada nyavirus	Nyavirus sierranevadense	Sierra Nevada virus (SNVV)		
Soybean cyst nematode socyvirus	Socyvirus heteroderae	soybean cyst nematode virus 1 (SbCNV-1)		
Family Paramyxoviridae				
Atlantic salmon paramyxovirus	Aquaparamyxovirus salmonis	Atlantic salmon paramyxovirus (AsaPV)		
Newcastle disease virus	Avulavirus avisprimum	avian paramyxovirus 1 (APMV-1)		
Avian paramyxovirus 2	Avulavirus avissecundum	avian paramyxovirus 2 (APMV-2)		
Avian paramyxovirus 3	Avulavirus avistertium	avian paramyxovirus 3 (APMV-3)		
Avian paramyxovirus 4	Avulavirus avisquartum	avian paramyxovirus 4 (APMV-4)		
Avian paramyxovirus 5	Avulavirus avisquintum	avian paramyxovirus 5 (APMV-5)		
Avian paramyxovirus 6	Avulavirus avissextum	avian paramyxovirus 6 (APMV-6)		
Avian paramyxovirus 7	Avulavirus avisseptimum	avian paramyxovirus 7 (APMV-7)		
Avian paramyxovirus 8	Avulavirus avisoctavum	avian paramyxovirus 8 (APMV-8)		
Avian paramyxovirus 9	Avulavirus avisnonum	avian paramyxovirus 9 (APMV-9)		
Avian paramyxovirus 10	Avulavirus avisdecimum	avian paramyxovirus 10 (APMV-10)		
Avian paramyxovirus 11	Avulavirus avisundecimum	avian paramyxovirus 11 (APMV-11)		
Avian paramyxovirus 12	Avulavirus avisduodecimum	avian paramyxovirus 12 (APMV-12)		
Fer-de-Lance paramyxovirus	Ferlavirus bothropsi	Fer-de-Lance virus (FDLV)		
Cedar henipavirus	Henipavirus cedarense	Cedar virus (CedV)		
Ghanaian bat henipavirus	Henipavirus ghanense	Kumasi virus (KV)		
Hendra virus	Henipavirus hendrense	Hendra virus (HeV)		
Mojiang henipavirus	Henipavirus moiangense	Mòjiāng virus (MojV)		
Nipah virus	Henipavirus nipahense	Nipah virus (NiV)		
Canine distemper virus	Morbillivirus canis	canine distemper virus (CDV)		
Cetacean morbillivirus	Morbillivirus cetaceae	cetacean morbillivirus (CeMV)		
Feline morbillivirus	Morbillivirus felis	feline morbillivirus (FeMV)		
Measles virus	Morbillivirus hominis	measles virus (MeV)		
Peste-des-petits-ruminants virus	Morbillivirus caprinae	peste-des-petits-ruminants virus (PPRV)		
Phocine distemper virus	Morbillivirus phocinae	phocine distemper virus (PDV)		
Rinderpest virus	Morbillivirus bovinae	rinderpest virus (RPV)		
Bovine parainfluenza virus 3 Human parainfluenza virus 1	Respirovirus bovistertium	bovine parainfluenza virus 3 (BPIV-3) human parainfluenza virus 1 (HPIV-1)		
Human parainfluenza virus 3	Respirovirus parainfluenzaeprimum Respirovirus parainfluenzaetertium	human parainfluenza virus 1 (HPIV-3)		
Porcine parainfluenza virus 1	Respirovirus suisprimum	porcine parainfluenza virus 3 (11114-3)		
Sendai virus	Respirovirus muris	Sendai virus (SeV)		
Human parainfluenza virus 2	Rubulavirus parainfluenzaesecundum	human parainfluenza virus 2 (HPIV-2)		
Human parainfluenza virus 2 Human parainfluenza virus 4	Rubulavirus parainfluenzaequartum	human parainfluenza viruses 4a/b (HPIV-4a/b)		
Mapuera virus	Rubulavirus mapuerense	Mapuera virus (MapV)		
Mumps virus	Rubulavirus parotitidis ^c	mumps virus (MuV), bat mumps virus		
Parainfluenza virus 5	Rubulavirus parainfluenzaequintum	parainfluenza virus 5 (PIV-5)		
Porcine rubulavirus	Rubulavirus suis	La Piedad Michoacán Mexico virus (LPMV)		
Simian virus 41	Rubulavirus macacae	simian virus 41		

Current ICTV-approved species name	Possible converted Linnaean species name ^a	(Unchanged) member virus name and abbreviation
Family Pneumoviridae		
Avian metapneumovirus	Metapneumovirus avis	turkey rhinotracheitis virus
Human metapneumovirus	Metapneumovirus hominis	human metapneumovirus (HMPV)
Bovine respiratory syncytial virus	Orthopneumovirus bovis	bovine respiratory syncytial virus (BRSV)
Human respiratory syncytial virus	Orthopneumovirus hominis	human respiratory syncytial viruses A2/B1/S2 (HRSV-A2/B1/S2)
Murine pneumonia virus	Orthopneumovirus muris	murine pneumonia virus (MPV)
Family Rhabdoviridae		1616 1 6 4 4 7 7 7 7
Alfalfa dwarf cytorhabdovirus	Cytorhabdovirus medicagonis	alfalfa dwarf virus (ADV)
Barley yellow striate mosaic cytorhabdovirus	Cytorhabdovirus hordei	barley yellow striate mosaic virus (BYSMV), maize sterile stunt virus (MSSV), wheat chlorotic streak virus (WCSV)
Broccoli necrotic yellows cytorhabdovirus	Cytorhabdovirus brassicae	broccoli necrotic yellows virus (BNYV)
Festuca leaf streak cytorhabdovirus	Cytorhabdovirus festucae	festuca leaf streak virus (FLSV)
Lettuce necrotic yellows cytorhabdovirus	Cytorhabdovirus lactucanecante ^d	lettuce necrotic yellows virus (LNYV)
Lettuce yellow mottle cytorhabdovirus	Cytorhabdovirus lactucamaculante ^e	lettuce yellow mottle virus (LYMoV)
Northern cereal mosaic cytorhabdovirus	Cytorhabdovirus cerealisborei ^f	northern cereal mosaic virus (NCMV)
Sonchus cytorhabdovirus 1	Cytorhabdovirus sonchi	sonchus virus (SonV)
Strawberry crinkle cytorhabdovirus	Cytorhabdovirus fragariae	strawberry crinkle virus (SCV)
Wheat American striate mosaic cytorhabdovirus	Cytorhabdovirus tritici	wheat American striate mosaic virus (WASMV)
Coffee ringspot dichorhavirus	Dichorhavirus coffeae	coffee ringspot virus (CoRSV)
Orchid fleck dichorhavirus	Dichorhavirus orchidaceae	orchid fleck virus (OFV)
Adelaide River ephemorovirus	Ephemerovirus flumenadelaidense	Adelaide River virus (ARV)
Berrimah ephemerovirus	Ephemerovirus berrimahense	Berrimah virus (BRMV)
Bovine fever ephemerovirus	Ephemerovirus bubulifebris ^g	bovine ephemeral fever virus (BEFV)
Kotonkan ephemerovirus	Ephemerovirus kotonkani	kotonkan virus (KOTV)
Obodhiang ephemerovirus	Ephemerovirus obodhiangense	Obodhiang virus (OBOV)
Aravan lyssavirus	Lyssavirus aravanense	Aravan virus (ARAV)
Australian bat lyssavirus	Lyssavirus ballinense	Australian bat lyssavirus (ABLV)
Bokeloh bat lyssavirus	Lyssavirus bokelohense	Bokeloh bat lyssavirus (BBLV)
Duvenhage lyssavirus	Lyssavirus duvenhagei	Duvenhage virus (DUVV)
European bat 1 lyssavirus European bat 2 lyssavirus	Lyssavirus alphaeuropense Lyssavirus betaeuropense	European bat lyssavirus 1 (EBLV-1) European bat lyssavirus 2 (EBLV-2)
Ikoma lyssavirus	Lyssavirus ikomense	Ikoma lyssavirus (IKOV)
Irkut lyssavirus	Lyssavirus irkutense	Irkut virus (IRKV)
Khujand lyssavirus	Lyssavirus khuiandense	Khujand virus (KHUV)
Lagos bat lyssavirus	Lyssavirus lagosense	Lagos bat virus (LBV)
Mokola lyssavirus	Lyssavirus mokolense	Mokola virus (MOKV)
Rabies lyssavirus	Lyssavirus rabies	rabies virus (RABV)
Shimoni bat lyssavirus	Lyssavirus shimonense	Shimoni bat virus (SHIBV)
West Caucasian bat lyssavirus	Lyssavirus occidenscaucasense	West Caucasian bat virus (WCBV)
Hirame novirhabdovirus	Novirhabdovirus paralichthyos	Hirame rhabdovirus (HIRV)
Oncorhynchus 1 novirhabdovirus	Novirhabdovirus salmonidae	infectious hematopoietic necrosis virus (IHNV)
Snakehead novirhabdovirus	Novirhabdovirus channae	snakehead rhabdovirus (SHRV)
Oncorhynchus 2 novirhabdovirus	Novirhabdovirus piscicidante ^h	viral hemorrhagic septicemia virus (VHSV)
Datura yellow vein nucleorhabdovirus	Nucleorhabdovirus daturae	datura yellow vein virus (DYVV)
Eggplant mottled nucleorhabdovirus Maize fine streak nucleorhabdovirus	Nucleorhabdovirus solanimelongenae Nucleorhabdovirus zealineante	eggplant mottled dwarf virus (EMDV) maize fine streak virus (MSFV)
Maize Iranian mosaic	Nucleorhabdovirus zeairanense	maize Iranian mosaic virus (MIMV)
nucleorhabdovirus	N. 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	
Maize mosaic nucleorhabdovirus	Nucleorhabdovirus zeamosaicante	maize mosaic virus (MMV)
Potato yellow dwarf nucleorhabdovirus	Nucleorhabdovirus solanituberosi	potato yellow dwarf virus (PYDV)
Rice yellow stunt nucleorhabdovirus	Nucleorhabdovirus oryzae	rice yellow stunt virus (RYSV), rice transitory
Sonchus yellow net nucleorhabdovirus	Nucleorhabdovirus retesonchi	yellowing virus (RTYV) sonchus yellow net virus (SYNV)
Sowthistle yellow vein	Nucleorhabdovirus venasonchi	sowthistle yellow vein virus (SYVV)
nucleorhabdovirus		, (-2)
Taro vein chlorosis nucleorhabdovirus	Nucleorhabdovirus colocasiae	taro vein chlorosis virus (TaVCV)
into cetti cittorooto itticicorimonocti no	1 there en in the concentration	
Anguillid perhabdovirus	Perhabdovirus anguillae	eel virus European X (EVEX), eel virus American

TABLE 2. Continued				
Current ICTV-approved species name	Possible converted Linnaean species name ^a	(Unchanged) member virus name and abbreviation		
Perch perhabdovirus	Perhabdovirus percae	perch rhabdovirus (PRV)		
Sea trout perhabdovirus	Perhabdovirus truttae	lake trout rhabdovirus (LTRV), Swedish sea trout rhabdovirus (SSTV)		
Drosophila affinis sigmavirus	Sigmavirus drosophilaeaffinis	Drosophila affinis sigmavirus (DAffSV)		
Drosophila ananassae sigmavirus	Sigmavirus drosophilaeananassae	Drosophila ananassae sigmavirus (DAnaSV)		
Drosophila immigrans sigmavirus	Sigmavirus drosophilaeimmigrantis	Drosophila immigrans sigmavirus (DImmSV)		
Drosophila melanogaster sigmavirus	Sigmavirus drosophilaemelanogastris	Drosophila melanogaster sigmavirus (DMelSV)		
Drosophila obscura sigmavirus	Sigmavirus drosophilaeobscurae	Drosophila obscura sigmavirus (DObsSV)		
Drosophila tristis sigmavirus	Sigmavirus drosophilaetristis	Drosophila tristis sigmavirus (DTriSV)		
Muscina stabulans sigmavirus	Sigmavirus muscinaestabulantis	Muscina stabulans sigmavirus (MStaSV)		
Pike fry sprivivirus	Sprivivirus esocis	pike fry rhabdovirus (PFRV), grass carp rhabdovirus (GrCRV), Tench rhabdovirus (TenRV)		
Carp sprivivirus	Sprivivirus cyprinidae	spring viremia of carp virus (SVCV)		
Coastal Plains tibrovirus	Tibrovirus planiorae ⁱ	Coastal Plains virus (CPV)		
Tibrogargan tibrovirus	Tibrovirus tibrogarganense	Bivens Arm virus (BAV), Tibrogargan virus (TIBV)		
Durham tupavirus	Tupavirus durhamense	Durham virus (DURV)		
Tupaia tupavirus	Tupavirus tupaiae	tupaia virus (TUPV)		
Lettuce big-vein associated varicosavirus	Varicosavirus lactucavenamagna	lettuce big-vein associated virus (LBVaV)		
Alagoas vesiculovirus	Vesiculovirus alagoasense	vesicular stomatitis Alagoas virus (VSAV)		
Carajas vesiculovirus	Vesiculovirus caraiasense	Carajás virus (CJSV)		
Chandipura vesiculovirus	Vesiculovirus chandipurense	Chandipura virus (CHPV)		
Cocal vesiculovirus	Vesiculovirus cocalense	Cocal virus (COCV)		
Indiana vesiculovirus	Vesiculovirus indianense	vesicular stomatitis Indiana virus (VSIV)		
Isfahan vesiculovirus	Vesiculovirus isfahanense	Isfahan virus (ISFV)		
Maraba vesiculovirus	Vesiculovirus marabense	Maraba virus (MARAV)		
New Jersey vesiculovirus	Vesiculovirus newierseyense	vesicular stomatitis New Jersey virus (VSNJV)		
Piry vesiculovirus	Vesiculovirus piryense	Piry virus (PIRYV)		
Flanders virus	Conversion not possible because not assigned to a genus	Flanders virus (FLAV)		
Ngaingan virus	Conversion not possible because not assigned to a genus	Ngaingan virus (NGAV)		
Wongabel virus	Conversion not possible because not assigned to a genus	Wongabel virus (WONV)		
Family Sunviridae	3 0			
Reptile sunshinevirus 1	Sunshinevirus reptilis	Sunshine Coast virus (SunCV)		
Unassigned				
Xincheng anphevirus	Anphevirus xinchengense	Xīnchéng mosquito virus (XcMV)		
Lishi arlivirus	Arlivirus lishiense	Lishì spider virus 2 (LsSV-2)		
Sanxia wastrivirus	Wastrivirus sanxiense	Sānxiá water strider virus 4 (SxWSV-4)		
Tacheng chengtivirus	Chengtivirus tachengense	Tăchéng tick virus 6 (TcTV-6)		
Wenzhou crustavirus	Crustavirus wenzhouense	Wēnzhōu crab virus 1 (WzCV-1)		

Note: Dark green (online version only) depicts current true binomial names and their Linnaean counterparts; light green (only version only) depicts current binomial-like names and their Linnaean counterparts; purple (online version only) depicts current non-binomial names and their Linnaean counterparts

Conversion of Free-Floating Species Names to Linnaean Binomials

Three mononegaviral species names (*Flanders virus*, *Ngaingan virus*, and *Wongabel virus*) are "free-floating" within the family *Rhabdoviridae*, that is, they have not been assigned to any of the current genera (purple

in Table 2). Consequently, these names cannot be converted to Linnaean binomials until genera for them are established (a proposal to include these species in genera is currently in preparation). The implementation of Linnaean binomial virus species names by the ICTV would, therefore, require a change in the International Code of Virus Classification and

^aThese names are for illustration purposes only and may not be the same as the names that ICTV Study Groups would officially propose if a Linnaean binomial species naming convention were implemented.

^bFor species names referring to host names of their virus members, we used the singular possessive of the lowest taxon these hosts belong to.

^cFor species names referring to disease caused by their virus members, we used Latin translations for the disease names.

 $^{^{\}rm d} {\it lactucane cante} = "lettuce-killing".$

elactucamaculante = "lettuce-sullying".

fcerealisborei = "of the northern cereal".

g bubulifebris = "of the bovine fever".

hpiscicidante= "fish-killing".

iplaniorae = "of the plain of the coast".

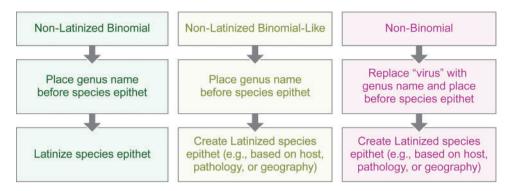


FIGURE 1. Stepwise flow chart for conversion of current virus species names to the Linnaean binomial format. Colors correspond to those used in Tables 1 and 2 (online version only).

Nomenclature (ICVCN) mandating genus assignment or establishment concomitant with species establishment.

DISCUSSION

The results of the exercise described here suggest that conversion of current virus species names to Linnaean binomials is a practical task that could be achieved rapidly by individual ICTV Study Groups. The draft conversion of the 34 arenaviral and 141 mononegaviral species names (Tables 1 and 2) was achieved within three work days by a single individual (T.S.P.) following a simple flow chart (Fig. 1).

The main logistical challenge turned out to be the identification of the etymological origins of some words in current species names (e.g., "Piry" in *Piry vesiculovirus*; "Aravan" in *Aravan lyssavirus*) for proper Latinization. Subsequent discussion between the members of all relevant ICTV Study Groups to refine or replace the draft preliminary names by correcting and devising alternatives took an additional 2–3 weeks.

Clearly, the Linnaean names listed in Tables 1 and 2 are not without controversy and could be easily ameliorated. As often is the case in nomenclature, a number of questions remains, such as whether meaning should be closely attached to names. If future scientific discoveries contradict the species name (e.g., if a virus species is named after the presumed host of the member virus and later research demonstrates a wider host spectrum or the presumed host not to be the host), how concerned should scientists be with the mismatch? Another question is whether certain within-taxa consistencies are preferable (e.g., if most species within a genus are named after the hosts of their members, then perhaps all species names within that genus should be devised that way). Actual ICTV-supported conversion of current virus species names to the Linnaean format would likely result in more extensive ICTV Study Group discussions on which types of names should be chosen. Nevertheless, the overall process described herein can most likely be extrapolated to virus species beyond the family *Arenaviridae* and order Mononegavirales, albeit with some virus taxon-specific complexities and concerns. Thus, the current Master List of ICTV-approved species names (3704 in 2016) could

likely be processed within a few months or even in a much shorter time, if a concerted effort is undertaken by the relevant ICTV Study Groups.

ICTV Study Group members either already possess or can acquire the basic knowledge of Latin needed for most name conversions, and the guidance of experts with a more extensive background in taxonomy-oriented Latin would be advantageous. Some creativity would be required in instances of sets of virus species names that differ from each other by only a number or an alphanumerical and species name epithets that consist of merely a number or a letter—a situation that barely exists in other, nonviral, taxonomies.

Linnaean-style binomials (genus names preceding the species epithet) but without Latinization have been proposed as a practical alternative to the more radical complete binomial nomenclature of viruses by some authors of this article. Following those proposals, most current species names could be converted into new names that would not look as unfamiliar to the virologists as properly declined Latinized names. Importantly, uniform grammar of these names would also allow incorporation of viral taxa into comprehensive taxonomic databases. However, Latinized binomials may have important advantages over non-Latinized binomials. Throughout the history of science, different languages have dominated scientific publications, from Arabic and Greek to Italian, French, and German. While today English is today considered the unofficial language of international science, this preference may quite possibly change in the future. Moreover, many researchers publish findings in their native languages in specialty journals.

In contrast to English or other spoken languages, Latin is static, that is, the meaning of Latin words will not change over time or depend on geographic areas. Therefore, the meaning of Latin words is quite precise no matter when they are used. In zoology and botany, this Latin name stability allows easy translation of an animal or plant name in a manuscript written in a foreign language because of the common Latinized species name. For instance, the German "Meerschweinchen (Cavia porcellus)," Chinese "豚鼠 (Cavia porcellus)," and English "guinea pig (Cavia porcellus)" instantly clarify

that "Meerschweinchen," "豚鼠," and "guinea pig" refer to the same animal belonging to the species Cavia porcellus. This advantage may even apply within a given language. For instance, in American English, "catamount," "cougar," "mountain lion," and "puma" are all synonymous names for the felid belonging to the species Puma concolor. The four names map to a unique Latin name of a species, but they do not necessarily map uniquely to specific words in every other extant language. Such name diversity is also present in virology. For instance, in Chinese, both "伊波拉病毒" and "埃博拉病毒" are in common use for the English "Ebola virus," but both Chinese names could immediately be interrelated via the associated virus species name. Using a universal, "dead" language such as Latin for species names may not only keep biological taxonomy consistent, but also keep it neutral with respect to emergent globally dominant languages. As an additional advantage, Latinized species names will appear foreign to most readers. The distinct appearance of these names could make it easier for nonspecialists to easily distinguish names of viruses (physical entities that are typically labeled in a spoken language, such as English) and the names of species (concepts of the mind).

Different taxonomic codes for different kingdoms utilize additional rules helping to avoid various kinds of complication and confusion. For instance, the International Code of Botanical Nomenclature prohibits the use of tautonymic species names, that is, names in which the genus name is identical in spelling with the species epithet; such names, however, are allowed in zoological taxonomy (e.g., Gorilla gorilla, Rattus rattus). Tautonymic species names currently do exist in mononegavirus taxonomy (e.g., Marburg marburgvirus).

Botanical and zoological, but not prokaryotic, species names are typically followed by the "authority," that is, information on who first published the description and name of the species (in botany, for instance, "Arabidopsis thaliana (L.) Heynh."; in zoology, for instance, "Mus musculus Linnaeus, 1758"). In the case of virus species names, considerations may be put forward to omit any authoritative information, as is done in prokaryotic taxonomy.

Another source of contention in nonviral taxonomies is "priority," that is, the problem of choosing the official name between two distinct species names after it is determined that both names refer to the same member organism. Priority would unlikely be a significant issue for virus taxonomy at this time as the number of ICTVaccepted species is still relatively low. The history of establishing virus species spans only a few decades, rather than hundreds of years, and such names are generally better documented than in older, nonviral, taxonomies. An opportunity exists to update the ICVCN with new rules on resolving potential naming conflicts once they arise. In contrast to other taxonomies, a mechanism is already in place for virus taxonomists to modify the ICVCN annually if necessary. It should also be noted that "authority" and "priority" are concepts that apply in taxonomies in which the first valid

publication of a taxon determines the application of taxon names. Viruses are explicitly excluded from such a process at the moment, because the use of taxon names is determined by the ICTV through its approval process.

We conclude that conversion of current virus species names to the Linnaean format would not be as complicated or time-consuming as even the authors of this article had expected when this thought experiment was initiated. The current ICVCN rules also do not prevent the adoption of Linnaean binomials, and such names can be proposed officially by virologists or ICTV Study Groups. However, the ICVCN would have to be amended if uniform application of Linnaean binomials was desired. Whether such an amendment should be pursued will require extensive consultation across the virology community and will be the decision of the ICTV. The actual Linnaean species names to be chosen for each virus would ultimately be the purview of the various ICTV Study Groups and the ICTV membership.

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REFERENCES

- Adams M.J., Lefkowitz E.J., King A.M.Q., Gorbalenya A.E., Harrach B., Harrison R.L., Knowles N.J., Kropinski A.M., Krupovic M., Kuhn J.H., Mushegian A.R., Nibert M., Sabanadzovic S., Sanfaçon H., Siddell S.G., Simmonds P., Zerbini F.M., Davison A.J. 2016. Ratification vote on taxonomic proposals to the International Committee on Taxonomy of Viruses (2016). Arch. Virol. 161:
- Afonso C.L., Amarasinghe G.K., Bányai K., Bào Y., Basler C.F., Bavari S., Bejerman N., Blasdell K.R., Briand F.-X., Briese T., Bukreyev A., Calisher C.H., Chandran K., Chéng J., Clawson A.N., Collins P.L., Dietzgen R.G., Dolnik O., Domier L.L., Dürrwald R., Dye J.M., Easton A.J., Ebihara H., Farkas S.L., Freitas-Astúa J., Formenty P., Fouchier R.A.M., Fù Y., Ghedin E., Goodin M.M., Hewson R., Horie M., Hyndman T.H., Ji?ng D., Kitajima E.W., Kobinger G.P., Kondo H., Kurath G., Lamb R.A., Lenardon S., Leroy E.M., Li C.-X., Lin X.-D., Liú L., Longdon B., Marton S., Maisner A., Mühlberger E., Netesov S.V., Nowotny N., Patterson J.L., Payne S.L., Paweska J.T., Randall R.E., Rima B.K., Rota P., Rubbenstroth D., Schwemmle M., Shi M., Smither S.J., Stenglein M.D., Stone D.M., Takada A., Terregino C., Tesh R.B., Tian J.-H., Tomonaga K., Tordo N., Towner J.S., Vasilakis N., Verbeek M., Volchkov V.E., Wahl-Jensen V., Walsh J.A., Walker P.J., Wang D., Wang L.-F., Wetzel T., Whitfield A.E., Xiè J., Yuen K.-Y., Zhang Y.-Z., Kuhn J.H. 2016. Taxonomy of the order *Mononegavirales*: update 2016. Arch. Virol. 161:2351–2360.
- Calisher C.H., Mahy B.W. 2003. Taxonomy: get it right or leave it alone.
- Am. J. Trop. Med. Hyg. 68:505–506. Drebot M.A., Henchal E., Hjelle B., LeDuc J.W., Repik P.M., Roehrig J.T., Schmaljohn C.S., Shope R.E., Tesh R.B., Weaver S.C., Calisher C.H. 2002. Improved clarity of meaning from the use of both formal species names and common (vernacular) virus names in virological literature. Arch. Virol. 147:2465-2472.
- Fenner F. 1976. Classification and nomenclature of viruses second report of the International Committee on Taxonomy of Viruses. Intervirology 7:1-115.
- Greuter W., Garrity G., Hawksworth D.L., Jahn R., Kirk P.M., Knapp S., McNeill J., Michel E., Patterson D.J., Pyle R., B.J. Tindall. 2011. Draft BioCode (2011): principles and rules regulating the naming of organisms. Taxon 60:201-212.
- Huemer H.P., Wechselberger C., Bennett A.M., Falke D., Harrington L. 2003. Cloning and expression of the complement receptor glycoprotein C from Herpesvirus simiae (herpes B virus): protection from complement-mediated cell lysis. J. Gen. Virol. 84: 1091-1100.
- Hummeler K., Davidson W.L., Henle W., Laboccetta A.C., Ruch H.G. 1959. Encephalomyelitis due to infection with Herpesvirus simiae (herpes B virus); a report of two fatal, laboratory-acquired cases. N. Engl. J. Med. 261:64-68.
- International Association for Plant Taxonomy. 2011. International Code of Nomenclature for algae, fungi, and plants (Melbourne Code). Oberreifenberg, Germany: Koeltz Scientific Books (Regnum Vegetabile; vol. 154). Available from: URL http://www.iapt-taxon. org/nomen/main.php.
- International Commission on Zoological Nomenclature. 2012. International Code of Zoological Nomenclature. 4th ed. London, UK: The International Trust for Zoological Nomenclature. Available from: URL http://www.iczn.org/iczn/index.jsp#.

- International Committee on Systematic Bacteriology. International Code of Nomenclature of Bacteria: Bacteriological Code, 1990 Revision. Washington, DC: ASM Press.
- International Committee on Taxonomy of Viruses. 2013. The International Code of Virus Classification and Nomenclature, February 2013. Available from: URL http://www.ictvonline.org/ codeOfVirusClassification.asp.
- International Committee on Taxonomy of Viruses. 2016. Brief notes of the meeting of the ICTV Executive Committee (EC) and Study Group (SG) chairs held on 1-2 February 2016 at Hinxton Hall, Wellcome Trust Genome Campus, Cambridgeshire, UK. Available from: URL http://talk.ictvonline.org/meetings/ sg_hinxton/w/sg_meeting
- King A.M.Q., Adams M.J., Carstens E.B., Lefkowitz E.J. 2012. The International Code of Virus Classification and Nomenclature. In: King A.M.Q., Adams M.J., Carstens E.B., Lefkowitz E.J., editors. Virus taxonomy-ninth report of the International Committee on Taxonomy of Viruses. London, UK: Elsevier/Academic Press. p. 1273-1277
- Koonin E.V., Starokadomskyy P. 2016. Are viruses alive? The replicator paradigm sheds decisive light on an old but misguided question. Stud. Hist. Philos. Biol. Biomed. Sci. 59:125-134.
- Kuhn J.H., Jahrling P.B. 2010. Clarification and guidance on the proper usage of virus and virus species names. Arch. Virol. 155:445-453.
- Linnaeus C. 1753. Species plantarum. Stockholm, Sweden [Latin]: Laurentius Salvius.
- Matthews R.E.F. 1983. A critical appraisal of viral taxonomy. Boca Raton (FL): CRC Press.
- Matthews R.E.F. 1985a. Viral taxonomy. Microbiol. Sci. 2:74-76.
- Matthews R.E.F. 1985b. Viral taxonomy for the nonvirologist. Annu. Rev. Microbiol. 39:451-474.
- Milne R.G. 1984. The species problem in plant virology. Microbiol. Sci.
- Philippe N., Legendre M., Doutre G., Coute Y., Poirot O., Lescot M., Arslan D., Seltzer V., Bertaux L., Bruley C., Garin J., Claverie J.M., Abergel C. 2013. Pandoraviruses: amoeba viruses with genomes up to 2.5 Mb reaching that of parasitic eukaryotes. Science 341:281–286.
- Radoshitzky S.R., Bao Y., Buchmeier M.J., Charrel R.N., Clawson A.N., Clegg C.S., DeRisi J.L., Emonet S., Gonzalez J.-P., Kuhn J.H., Lukashevich I.S., Peters C.J., Romanowski V., Salvato M.S., Stenglein M.D., de la Torre J.C. 2015. Past, present, and future of arenavirus taxonomy. Arch. Virol. 160:1851-1874.
- Van Regenmortel M.H. 2003. Viruses are real, virus species are manmade, taxonomic constructions. Arch. Virol. 148:2481–2488.
- Van Regenmortel M.H. 2006. Virologists, taxonomy and the demands of logic. Arch. Virol. 151:1251-1255.
- Van Regenmortel M.H. 2007. Virus species and virus identification: past and current controversies. Infect. Genet. Evol. 7:133-144.
- Van Regenmortel M.H., Burke D.S., Calisher C.H., Dietzgen R.G., Fauquet C.M., Ghabrial S.A., Jahrling P.B., Johnson K.M., Holbrook M.R., Horzinek M.C., Keil G.M., Kuhn J.H., Mahy B.W., Martelli G.P., Pringle C., Rybicki E.P., Skern T., Tesh R.B., Wahl-Jensen V., Walker P.J., Weaver S.C. 2010. A proposal to change existing virus species names to non-Latinized binomials. Arch. Virol. 155:1909-1919.
- Van Regenmortel M.H.V. Forthcoming 2016a. Classes, taxa and categories in hierarchical virus classification: a review of current debates on definitions and names of virus species. Bionomina 10.
- van Regenmortel M.H.V. 2016b. The metaphor that viruses are living is alive and well, but it is no more than a metaphor. Stud. Hist. Philos. Biol. Biomed. Sci. 59:117-124.
- Wildy P. 1971. Classification and nomenclature of viruses first report of the International Committee on Nomenclature of Viruses. Basel, Switzerland: S. Karger.