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### Permalink

<https://escholarship.org/uc/item/4xp831st>

### Journal

Journal of Experimental Biology, 226(15)

### ISSN

0022-0949

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### Publication Date

2023-08-01

### DOI

10.1242/jeb.245784

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Peer reviewed

# 1 **Integration of feedforward and feedback control in the neuromechanics of** 2 **vertebrate locomotion: a review of experimental, simulation and robotic studies**

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9  
10 **Keywords:** Neuromechanics, locomotion, sensorimotor control, central pattern generation, reflexes,  
11 feedforward and feedback control.

## 12 13 **Abstract**

14 Animal locomotion is the result of complex and multi-layered interactions between the nervous system, the  
15 musculo-skeletal system and the environment. Decoding the underlying mechanisms requires an  
16 integrative approach. Comparative experimental biology has allowed researchers to study the underlying  
17 components and some of their interactions across diverse animals. These studies have shown that  
18 locomotor neural circuits are distributed in the spinal cord, the midbrain and higher brain regions in  
19 vertebrates. The spinal cord plays a key role in locomotor control because it contains central pattern  
20 generators (CPGs) – systems of coupled neuronal oscillators that provide coordinated rhythmic control of  
21 muscle activation that can be viewed as feedforward controllers – and multiple reflex loops that provide  
22 feedback mechanisms. These circuits are activated and modulated by descending pathways from the  
23 brain. The relative contributions of CPGs, feedback loops and descending modulation, and how these vary  
24 between species and locomotor conditions remain poorly understood. Robots and neuromechanical  
25 simulations can complement experimental approaches by testing specific hypotheses and performing  
26 what-if scenarios. This Review will give an overview of key knowledge gained from comparative vertebrate  
27 experiments, and insights obtained from neuromechanical simulations and robotic approaches. We  
28 suggest that the roles of CPGs, feedback loops and descending modulation vary among animals depending  
29 on body size, intrinsic mechanical stability, time required to reach locomotor maturity and speed effects.  
30 We also hypothesize that distal joints rely more on feedback control compared to proximal joints. Finally,  
31 we highlight important opportunities to address fundamental biological questions through continued  
32 collaboration between experimentalists and engineers.

33 Summary statement: Comparative animal studies and neuromechanical modeling have revealed diversity  
34 in the integration of feedback and feedforward control, related to body size, mechanical stability, time to  
35 locomotor maturity and movement speed.

36  
37 **Short title: Integration of feedforward and feedback control in locomotion**

## Introduction

39 Animal locomotion arises from complex and rich interactions between the nervous system, the  
40 musculoskeletal system and the environment (Dickinson et al., 2000; Nishikawa et al., 2007). Animal  
41 sensorimotor control involves multi-layered and distributed systems, with central networks, reflexes and  
42 mechanics all contributing to sensorimotor responses on varied timescales (Büschges, 2005; Buschmann  
43 et al. 2015; Bidaye et al. 2018; Grillner, 1985; Grillner & El Manira, 2019; Loeb et al., 1999; Nirody, 2023;  
44 Pearson, 1995; Pearson & Gramlich, 2010; Rossignol et al., 2006). Because of this inherent complexity, it  
45 has been a longstanding challenge in biology to rigorously understand the structure, function and  
46 integration of animal sensorimotor systems. Integrative approaches are needed to address this challenge.  
47 Exchange between biology, physics and engineering disciplines can be particularly useful for generating  
48 and rigorously testing hypotheses about how mechanics and control interact to produce agile locomotion.  
49 In this Review, we discuss how the exchange between comparative experimental biology, physics and  
50 engineering can provide fundamental insights into the neuromechanics (see Glossary) of vertebrate  
51 locomotion.

52  
53 Historically, comparative approaches have been essential for providing mechanistic understanding of the  
54 control of movement, and *Journal of Experimental Biology* (JEB) has been an important venue for  
55 comparative studies across diverse taxa. In the 1930s–1940s, Gray and co-authors published numerous  
56 papers on animal movement in JEB, characterizing the locomotor movements, reflex responses and  
57 locomotor pattern generation in the eel, dogfish, toad, leech and earthworm (Gray, 1933, 1936; Gray et al.,  
58 1938; Gray & Lissmann, 1938, 1940; Gray & Sand, 1936). Subsequent studies continued to expand on this  
59 diversity and contribute to mechanistic understanding of sensorimotor control (e.g. Hughes, 1952;  
60 Hughes, 1957; Wilson 1965; Wilson 1967). Many of the early studies were foundational for understanding  
61 both the basic biomechanics and neural control circuitry for movement. At that time, it was often necessary  
62 to study movement mechanics and neural control together, because so little was known about both.

63  
64 In recent decades, the fields of comparative biomechanics and sensorimotor physiology have been  
65 relatively isolated, as each field focused on more detailed mechanistic understanding of specific systems.  
66 Simultaneously, studies of sensorimotor control have increasingly focused on a few genetic animal models,  
67 such as the mouse, zebrafish, fruit fly, and *Caenorhabditis elegans*, which enable the use of the most richly  
68 developed genetic and molecular tools (Bidaye et al. 2018; Fetcho & McLean, 2010; Fouad et al 2018; Kiehn,  
69 2011; Lewis & Eisen, 2003; Zhong et al., 2012; Mantziaris et al. 2020). Nonetheless, this narrow focus has  
70 its drawbacks, and important questions remain about the diversity and specialization of sensorimotor  
71 control across species. We suggest that it is important to continue to expand studies beyond the classic  
72 genetic animal models, because comparative approaches provide opportunities to address fundamental  
73 questions about the evolution and diversity of neuromechanical integration. Critical gaps in understanding  
74 remain in how the different elements of the sensorimotor system are integrated with each other, and how  
75 this integration varies with body size and among animals adapted for different locomotor environments.

76  
77 The interdisciplinary field of comparative neuromechanics has emerged over the last 15 years or so  
78 (Nishikawa et al., 2007; Ting & McKay, 2007), focusing on the study of mechanics and control of movement  
79 in a comparative framework. In this Review, we highlight principles and hypotheses that have emerged from  
80 this field, with a specific focus on sensorimotor and neuromechanical integration in vertebrates. We  
81 discuss current conceptualization of the role of central pattern generators (CPGs; see Glossary) and their  
82 integration with reflexes and limb mechanics, and the interactions between feedforward and feedback  
83 control mechanisms (see Glossary). In the sections below, we first summarize current understanding in

84 biology based on both historical and recent experimental evidence, then discuss contributions from bio-  
85 inspired robotics and simulations, and then highlight current open questions and future directions that are  
86 enabled through direct integration of engineering and experimental biology. Our Review focuses mainly on  
87 vertebrate locomotion; however, we note that many principles of locomotor control have parallels between  
88 vertebrates and invertebrates, and readers are directed to recent reviews of invertebrate systems (Nirody  
89 2023; Mantziaris et al. 2020; Bidaye et al. 2018).

## 90 **Biology: historical experimental evidence, current** 91 **understanding and new hypotheses**

92

### 93 **Integration of CPGs and sensory feedback in the spinal cord**

94 The spinal cord plays a key role in integrating predictive and reactive elements of vertebrate locomotor  
95 control (Grillner & El Manira, 2019; Rossignol et al., 2006). Animal control involves inherently long  
96 sensorimotor delays (see Glossary; More et al., 2010; More & Donelan, 2018). Because of these delays,  
97 animals cannot rely entirely on reactive, feedback-driven control, but must instead use a combination of  
98 predictive and reactive control mechanisms for stable movement (Fig. 1). The spinal cord contains neural  
99 circuits involved in integrating CPGs (see Box 1), reflex responses, and descending and ascending  
100 pathways to the brain (Grillner & El Manira, 2019; Pearson, 1995; Rossignol et al., 2006). CPGs in the spinal  
101 cord provide a type of predictive, feedforward controller for locomotion that helps to overcome delays.  
102 CPGs are activated by a descending 'drive' signal and produce predetermined motor outputs for  
103 locomotion. These outputs provide complex muscle activation for the anticipated mechanical demands,  
104 resulting in, for instance, traveling waves for undulatory locomotion and alternation of stance and swing  
105 phases for legged locomotion. CPGs therefore resemble feedforward controllers in control theory (see  
106 Glossary), because they produce complex and detailed motor commands given a simpler high-level  
107 descending signal for the desired behavior, such as movement at a specific speed. However, CPGs do not  
108 act in isolation. The CPG receives continuous modulation through descending drive commands and reflex  
109 feedback (Grillner et al., 2008; Rossignol et al., 2006). Consequently, the CPG can be thought of as a type  
110 of feedforward controller nested within a feedback control system (Holmes et al., 2006).

111

112 Current evidence suggests that animal locomotion makes use of a nested, multi-layered control  
113 architecture that organizes sensing and action (Grillner & El Manira, 2019; McLean & Dougherty, 2015;  
114 Pearson, 1995; Rossignol et al., 2006). A nested architecture enables separation of task selection (starting,  
115 stopping, gait and speed), task coordination (e.g. generation of coordinated rhythmic leg movements) and  
116 stabilization in response to disturbances. As a first approximation of the hierarchical organization, CPGs  
117 generate rhythmic commands that act as feedforward signals for task coordination, whereas descending  
118 commands activate and modulate CPGs for task selection, and reflexes provide stabilizing responses to  
119 perturbations (Fig. 1). We will later see that neuromechanical models suggest more complexity, with  
120 sensory feedback also contributing to task coordination (Owaki et al., 2013; Thandiackal et al., 2021).  
121 Sensorimotor delays give rise to temporal scaling of control responses based on the spatial distribution of  
122 system elements (Fig. 1). Inner loops deal locally with perturbations, relying on fast peripheral mechanisms,  
123 including intrinsic mechanics (Brown & Loeb, 2000; Daley et al., 2007; Dickinson et al., 2000; Full &  
124 Koditschek, 1999; Jindrich & Full, 2002) and short-latency reflexes (Af Klint et al., 2010; Daley et al., 2009;  
125 Hiebert & Pearson, 1999; Moritz & Farley, 2004). Intermediate loops modify task-level variables, such as  
126 joint and leg stiffness, and outer loops integrate sensorimotor information in the brain to select and switch  
127 among different tasks and update internal models (see Glossary; Pearson & Gramlich, 2010). However,

128 many uncertainties remain in how different control layers are integrated and modulated based on  
129 experience, learning and different locomotor contexts.

130  
131 There was a longstanding historical debate about the relative importance of feedforward and feedback in  
132 control of animal locomotion. Sherrington's work on reduced animal preparations revealed that reflex  
133 actions could generate the component motions necessary for rhythmic walking (Sherrington, 1910). Thus,  
134 Sherrington argued that locomotion could be viewed as the result of a chain of reflexes (Sherrington, 1900,  
135 1906, 1910). Subsequent work by Graham Brown demonstrated that motor patterns for locomotion could  
136 be generated in deafferented preparations and did not require sensory input. Graham Brown argued that  
137 'spinal centers' (later named central pattern generators) acted as the primary unit of motor activity, not the  
138 reflex (Graham Brown, 1911, 1914). He suggested that the reflexes regulate rather than generate motor  
139 activity (Graham Brown, 1911). To this day, the relative roles of central and reflex-generated contributions  
140 to locomotion remain somewhat controversial; however, it has become clear that both mechanisms co-  
141 exist and provide redundancy and flexibility in the system.

142  
143 Subsequent work has confirmed the existence and shared features of spinal CPGs across vertebrates (see  
144 Box 1). Vertebrates also share a common mechanism for descending drive to CPGs to control gait speed  
145 and initiation. Stimulation of the mesencephalic locomotor region, invokes gait initiation and gait  
146 transitions across species, inducing swimming in fish and walking in terrestrial animals (Cabelguen et al.,  
147 2003; Grillner & Wallen, 1985; Shik et al., 1966; Steeves et al., 1987). As the strength of the stimulation  
148 increases, swimming speed increases in the fish, and a quadruped increases speed and transitions  
149 between gaits from a walk to trot to gallop (Shik et al., 1966; Wallén, 1982).

150  
151 In vertebrates, sensory feedback plays an essential role in entraining CPG rhythms to the mechanics of the  
152 body and the interaction with the environment. In the lamprey, periodic bending of the spinal cord results  
153 in entrainment (see Glossary) of the patterns of fictive locomotion (see Glossary), through sensory  
154 feedback from mechanosensitive edge cells in the spinal cord (Grillner et al., 1981; Grillner & Wallén, 1984).  
155 Similarly, proprioceptive feedback entrains gait rhythm and the timing of stance–swing transitions in  
156 quadrupedal gait (Pearson, 2008; Whelan, 1996). In decerebrate and spinal preparations (see Glossary),  
157 increasing belt speed leads to gait transitions from walk to trot and gallop, suggesting a role for sensory  
158 feedback entrainment in gait transitions (Barbeau & Rossignol, 1987; Forssberg et al., 1980; Kriellaars et  
159 al., 1994; Pearson, 2008; Whelan, 1996; Yanagihara et al., 1993). With increasing knowledge of the  
160 interneuronal networks and cellular mechanisms underlying sensorimotor control, it has become clear that  
161 sensory feedback is integrated at multiple levels: as distinct reflexes, as modulators of central pattern  
162 generation and integrating with longer-latency feedback loops to regulate navigation and task selection  
163 (Grillner & El Manira, 2019; Rossignol et al., 2006; Fig. 1).

164  
165 **Diversity in the integration of feedforward, feedback and model-based neural control**  
166 Recent evidence and modeling studies (see below) suggest that the relative contributions of feedforward  
167 and feedback can vary depending on size of the animal, speed, terrain and risks of falling. Below, we discuss  
168 emerging hypotheses about specialization and diversification of control components among vertebrates.  
169 Loosely speaking, we suggest two types of feedforward control, one in the spinal cord, based on CPGs, and  
170 one in higher brain centers, based on internal models, which we will here call (cephalized) model-based  
171 neural control (see Glossary). We call them both feedforward because they represent an anticipatory  
172 component as opposed to the reactive nature of feedback.

173  
174 **Body size and scaling of neuromechanical delays**

175 Body size is one important source of diversity in control mechanisms for locomotion (Fig. 2). Sensorimotor  
176 loop delay increases with body size (M), in proportion to  $M^{0.21}$  (More et al., 2010, 2013; More & Donelan,  
177 2018). Large animals experience longer sensorimotor delays relative to movement durations compared  
178 with the same ratio in small animals. For example, at speeds near the trot–gallop transition, a shrew has a  
179 total delay of 10ms, 25% of its stance duration, but an elephant has a total delay of about 180 ms, 60% of  
180 its stance duration (More & Donelan, 2018). Delay relative to stance duration is important because this  
181 represents the time available to apply force for a corrective response to a disturbance. If an animal cannot  
182 respond to a disturbance within the stance phase, the response requires coordination of multi-step and  
183 multi-legged strategies, which involve longer-latency feedback loops. Delays therefore challenge the ability  
184 of large animals to respond effectively to perturbations, even though they have slower dynamics and step  
185 cycles compared to smaller animals (Mohamed Thangal & Donelan, 2020; More & Donelan, 2018).

186  
187 To compensate for the relatively longer delays, it is expected that larger animals rely more on model-based  
188 control, using sensory feedback and internal models in the brain to generate state estimates for stable  
189 movement. Internal models can allow animals to compensate for delayed and noisy sensory feedback to  
190 predict future states, enabling generation of appropriate motor outputs for stable movement (Todorov,  
191 2004; Todorov & Jordan, 2002; Wolpert & Ghahramani, 2000). The synaptic delays associated with internal  
192 model computations are a higher fraction of the total sensorimotor delay in very small animals (Thangal &  
193 Donelan, 2020; More & Donelan, 2018). Consequently, the benefits of model-based control may not  
194 outweigh the costs of increased computation times in small animals (Thangal & Donelan, 2020; More &  
195 Donelan, 2018). Thus, overall, it is expected that larger animals are likely to have more cephalized model-  
196 based control, with higher brain involvement compared to small animals (Fig. 2), whereas small animals  
197 can achieve agile and robustly stable movement through more spinalized control mechanisms coupled to  
198 intrinsic mechanical reflexes (see Glossary).

199  
200 **Spinalization versus cephalization of feedforward control may relate to mechanical stability and the time**  
201 **to locomotor maturity**

202 Although all vertebrates share similar component systems for locomotor control, a source of diversity  
203 among vertebrates is the degree of involvement of the brain and the complexity of descending drive  
204 modulation (Fig. 3). This distinction may be related to different developmental demands between precocial  
205 and altricial species (see Glossary). Precocial species tend to have relatively small brains as adults;  
206 whereas altricial species tend to have larger adult brain size (Bennett & Harvey, 1985; Garwicz et al., 2009).  
207 We hypothesize that these developmental differences may also be related to the relative degree of  
208 ‘spinalization’ versus ‘cephalization’ of locomotor control. That is, precocial species have more spinalized  
209 locomotor control, relying mainly on spinal CPG networks as a feedforward controller, coupled to intrinsic  
210 mechanics of the body, with short-latency reflexes. In contrast, altricial species have more cephalized  
211 control, relying more on model-based control in the brain, with longer-latency reflexes updating internal  
212 models. Through experience and optimization, animals can optimize the use of efference copy (see  
213 Glossary) and sensory feedback to estimate current states and predict future states to determine desired  
214 motor outputs (Todorov, 2004; Todorov & Jordan, 2002; Wolpert & Ghahramani, 2000). Multiple brain  
215 regions may be involved in the generation, maintenance and updating of internal models, including the  
216 cerebellum and the posterior parietal cortex (Ito, 2008; McVea et al., 2009; Pearson & Gramlich, 2010).  
217 Multi-layered model-based control can overcome delays by predicting mechanical conditions over a wide  
218 range of contexts and adjusting feedforward commands through descending pathways (Nakahira et al.,  
219 2021).

220  
221 The contrast between precocial and altricial species can be appreciated when comparing large ground  
222 birds (such as ostriches and rheas) to humans. Although birds and humans have independently evolved

223 bipedalism, they have converged upon walking and running gaits with similar mechanical and energetic  
224 demands (Gatesy & Biewener, 1991; Roberts et al., 1998; Rubenson et al., 2004; Watson et al., 2011). Yet,  
225 they exhibit important differences in development and sensorimotor control features. Ground birds are  
226 precocial, able to walk and run very shortly after hatching (Muir et al., 1996; Muir & Chu, 2002; Y. U. Ryu &  
227 Bradley, 2009; Smith et al., 2010), whereas humans are exceptionally altricial, requiring many months of  
228 practice to walk without falling. We suggest that these developmental differences also reflect differences  
229 in spinalization versus cephalization of locomotor control. Birds have relatively more spinalized locomotor  
230 control compared to more cephalized control in mammals, including humans. Neurophysiological studies  
231 have demonstrated that spinal circuits are sufficient to generate complete locomotor patterns for self-  
232 supported walking in birds (Ho & O'Donovan, 1993; Sholomenko et al., 1991; Sholomenko & Steeves, 1987;  
233 Ten Cate, 1960). Birds do possess a common descending pathway with other vertebrates, from the  
234 mesencephalic locomotor region to the spinal CPGs, but lack a direct telencephalic–spinal projection  
235 analogous to the mammalian corticospinal tract (Sholomenko & O'Donovan, 1995; Sholomenko & Steeves,  
236 1987; Steeves et al., 1987; Webster & Steeves, 1988). A recent study genetically silenced interneurons in  
237 the dorsal spinal tract in chicks, which increased kinematic variability in walking (Haimson et al., 2021),  
238 suggesting that sensory feedback and descending modulation contribute to stability of walking in birds.  
239 Nonetheless, locomotor control appears to be relatively more spinalized in birds compared to the more  
240 cephalized sensorimotor control observed in mammals.

241  
242 The relatively spinalized versus cephalized locomotor control in ground birds versus mammals,  
243 respectively, might represent different solutions to the problem of neural delays. Cephalization allows more  
244 sophisticated internal models and model-based control; this allows delays to be overcome by predicting  
245 mechanical conditions over a wide range of learned contexts and adjusting descending commands  
246 accordingly. In contrast, spinalization relies on simpler CPG-based feedforward control, with the feedback-  
247 entrained rhythm providing estimates of current state and generation of motor output. We hypothesize that  
248 animals with more spinalized control also tend to have more intrinsically stable biomechanics (Fig. 3).  
249 Among bipeds, birds have a relatively flexed limb posture compared to humans, a forward horizontal  
250 position of the body and high elasticity in the distal leg muscles, features that increase mechanical stability  
251 (Badri-Spröwitz et al., 2022; Daley, 2018; Daley et al., 2009; Daley & Biewener, 2011; Daley & Birn-Jeffery,  
252 2018). Humans, in contrast, have a straight leg posture with the body vertically balanced over the hips,  
253 which is highly unstable without active control.

254  
255 Among quadrupeds there is also diversity in intrinsic mechanical stability that may relate to degree of  
256 cephalization versus spinalization of control. Large ungulates have parasagittal limb posture with relatively  
257 straight legs, a high body center of mass and a relatively narrow base of support. These features benefit  
258 locomotor economy by minimizing the active muscle force required to support body weight against gravity  
259 (Biewener, 1989). However, upright postures are relatively unstable compared to the sprawled posture and  
260 wide base of support typical of many amphibians and lizards (Alexander, 2002; Ijspeert, 2020). Note that  
261 here we mainly refer to static stability (see Glossary) rather than dynamic stability. At least at slow speeds,  
262 static stability in quadrupeds is proportional to the size of the support polygon, and inversely proportional  
263 to the ratio of body height to stance width (Alexander, 2002). We hypothesize that there is a control gradient  
264 in which mechanical stability enables spine-localized feedforward control, whereas mechanical instability  
265 is associated with brain-dominated feedforward control (Fig. 2). We also hypothesize that unstable animals  
266 require better proprioception for monitoring their posture and loads on limbs, and hence have a larger role  
267 for spinal sensing than mechanically stable animals. A similar hypothesis has been proposed in Ryczko et  
268 al. (2020).

269  
270 **Proximo-distal differentiation in control**

271 Across diverse tetrapod animals, limbs have a proximo-distal distribution in muscle–tendon morphology,  
272 which may also result in differentiation in control mechanisms (Daley et al., 2007). The largest, most  
273 powerful muscles are proximal, concentrating mass near the body. Distal muscles have lower mass and  
274 high mass-specific force due to their short, pennate muscle fiber arrangement, and high in-series  
275 compliance (see Glossary). The pennate architecture of distal muscles leads to a load-sensitive  
276 architectural gear ratio (Azizi et al., 2008; Eng et al., 2018; Roberts et al., 2019), which is likely to make them  
277 sensitive to external perturbations. In contrast, the high inertia and lower compliance of the proximal  
278 muscles is likely to make them less sensitive to external perturbations. Based on these mechanical  
279 differences between proximal and distal muscles, we hypothesize that proximal muscles use higher-gain  
280 feedforward control, with length and position feedback entraining CPG rhythm, whereas distal muscles rely  
281 more on intrinsic mechanics (i.e. reflexes) and short-latency load and stretch reflexes (Fig. 4). Indeed,  
282 evidence from cats suggests that proximal muscles at the hip contribute to regulation of stance–swing  
283 transitions, whereas the distal ankle extensors use force feedback to regulate stance load bearing (Donelan  
284 & Pearson, 2004; Gorassini et al., 1994; Hiebert et al., 1994; Pearson et al., 1998). Modeling studies also  
285 support this hypothesis (Dzeladini et al., 2014; see below).

286

## 287 **Using neuromechanical simulations and biorobots to** 288 **investigate animal motor control**

289 One theme that has emerged from neuromechanical studies is the importance of effective tuning of control  
290 to the biomechanical properties of the system and to physical interactions with the environment. However,  
291 the multi-layered, distributed and redundant nature of animal sensorimotor systems makes it challenging  
292 to rigorously understand the relationships and connections among the component systems through  
293 experiments alone. Neuromechanical simulations (i.e. numerical simulations of both neural circuits and  
294 bodies interacting with a virtual environment) can be useful tools to tackle these challenges and test  
295 hypotheses about animal motor control. Additionally, simulated neural circuits can also be tested in the  
296 real world with biomimetic robots used as physical models of animal bodies.

297 Since the modeler is in control of all components, models are ideally suited to study motor control in  
298 animals with an integrative perspective, following Richard Feynman’s famous quote: “What I cannot create,  
299 I do not understand”. Neuromechanical simulations and biorobots present multiple interesting properties:  
300 (i) they allow the modeler to explicitly determine, implement and modify different components (e.g.  
301 feedback loops, CPGs, muscle models); (ii) they provide access to many internal states, including quantities  
302 which are impossible to measure in moving animals; (iii) they offer the option to systematically change  
303 some properties (e.g. sizes and masses); (iv) they allow for repeatable experiments; (v) they allow ‘what-if’  
304 scenarios and testing of motor behaviors not observed in nature; and (vi) they allow for multiple types of  
305 perturbations and lesion studies. Modeling experiments benefit from an iterative approach, with iterations  
306 between animal studies, hypothesis design, model design, (numerical) experiments and predictions (Webb,  
307 2001). They are particularly useful for experiments that cannot be performed on real animals for practical,  
308 financial and/or ethical reasons.

309 The use of neuromechanical simulations and robots to investigate animal behavior has a fairly long history  
310 and finds its roots in early work in cybernetics (Ashby, 1957; Wiener, 2019) and in robotics, for instance,  
311 with Grey Walter’s tortoise robots (Walter, 1950, 1951). Other reviews have addressed the use of  
312 simulations and robots to investigate animal behavior (Dickinson et al., 2000; Floreano et al., 2014; Holmes  
313 et al., 2006; Ijspeert, 2014; Pearson et al., 2006; Webb, 2001, 2020; Ramdya and Ijspeert 2023). Here, we



314 focus in particular on locomotion and on how simulations and robots can help in the investigation of the  
315 interactions between CPGs, sensory feedback loops and descending modulation, and more specifically on  
316 interactions between feedforward and feedback control.

### 317 **Models of lamprey swimming highlight the importance of the CPG in mechanically** 318 **stable locomotion**

319 The lamprey represents a good example of how numerical modeling has contributed to decoding how  
320 neural circuits interact with the body to generate swimming. A first contribution of numerical modeling has  
321 been to decipher the neuronal and network properties of rhythm generation in the local segmental circuits.  
322 The lamprey spinal cord is composed of approximately 100 segments and each segment contains neural  
323 oscillators (see Glossary) that are part of the locomotor CPG. Biophysical models of these segmental  
324 circuits have shown that several mechanisms play a role in rhythm generation, including contralateral  
325 inhibition, frequency adaptation and stretch-sensitive cells (Ekeberg et al., 1991; Hellgren et al., 1992;  
326 Traven et al., 1993; Wallén et al., 1992). The relative importance of these mechanisms likely depends on  
327 the cycle frequency, which varies extensively in the lamprey (Traven et al., 1993; Wallén et al., 1992).

328 Numerical and mathematical models have also investigated the complete CPG circuits, and how traveling  
329 waves of neural activity are generated along the 100 segments of the spinal cord to produce forward  
330 swimming (Buchanan, 1992; Ekeberg, 1993; Ekeberg et al., 1995; Ijspeert, 1996; Williams, 1992a, 1992b;  
331 Cohen et al., 1982; Kopell et al., 1991; Kopell & Ermentrout, 1988; Williams et al., 1990). It is known that  
332 neurons in the local segmental oscillators project up and down the spinal cord, therefore creating couplings  
333 between oscillators. The models have shown that the most likely mechanisms to explain the phase lags  
334 between oscillators are asymmetries of inter-oscillator couplings, and that other potential explanations  
335 such as gradients of intrinsic frequencies and conduction delays are less likely.

336 Finally, models of the lamprey spinal cord have also been connected to simulated bodies in the water, i.e.  
337 to form complete neuromechanical simulations (Fig. 5A), in order to relate CPG activity to actual swimming  
338 behavior (Ekeberg, 1993; Ekeberg et al., 1995; Ekeberg & Grillner, 1999; Ijspeert, 1996; Thandiackal et al.,  
339 2021; Williams & McMillen, 2015). These neuromechanical simulations have demonstrated that the  
340 traveling waves of neural activity indeed generate forward swimming, and that modulating the descending  
341 drive signals applied to the CPG models can change the speed of swimming as well as induce turning, when  
342 symmetric and asymmetric left–right descending pathways are activated (Ekeberg, 1993). These  
343 simulations and robotic experiments (Ijspeert & Crespi, 2007) have shown that lamprey-like swimming can  
344 be obtained using CPG models without sensory feedback, supporting our hypothesis that CPGs play an  
345 important role in mechanically stable locomotion (Fig. 3). In this case the surrounding water provides a  
346 kind of mechanical stability (i.e. by preventing large accelerations).

347 Note that sensory feedback still plays a useful role in lamprey swimming. Sensory feedback is necessary  
348 for handling perturbations in the water, and neuromechanical simulations have shown the role of stretch  
349 receptors (that provide feedback about curvature to the CPG) in coping with changes of flow velocity in the  
350 water (Ekeberg et al. 1995; Ijspeert et al., 1999). Thus, there are specific environments that can only be  
351 crossed when sensory feedback is included, and cannot be crossed with the CPG in open-loop.

352 Remarkably, a lamprey-like robot (Fig. 5B; Thandiackal et al., 2021) has furthermore shown that local  
353 sensory feedback alone (in this case from pressure-sensitive cells in the lamprey skin) could synchronize  
354 the oscillators that comprise the CPG when direct coupling is removed. See Hamlet et al. (2023) for a  
355 similar finding in a neuromechanical simulation with stretch feedback. In other words, sensory feedback

356 presents an alternative and redundant mechanism for traveling wave generation, separate from direct CPG  
357 coupling. This offers a high robustness against lesions, similar to that seen in eels, which can continue  
358 swimming despite full transection of the spinal cord (Wallén, 1982). It also suggests that inter-oscillator  
359 couplings might be less strong than previously thought (see discussion below).

## 360 **Models show that CPGs can induce transitions between swimming and walking in** 361 **amphibians**

362 The transition between swimming and walking has been studied in the salamander using neuromechanical  
363 simulations and robots (Fig. 5C), (Bicanski et al., 2013; Ijspeert, 2001; Ijspeert et al., 2007; Karakasiliotis et  
364 al., 2013, 2016; Knüsel et al., 2020). Because of its amphibious locomotion, the salamander represents an  
365 interesting animal to study the transition from water to ground locomotion and to create a bridge between  
366 aquatic and terrestrial vertebrate studies (Ryczko et al., 2020).

367 Modeling studies in salamanders have demonstrated that the transition from swimming to ground  
368 locomotion can, in principle, be obtained by extending an undulatory swimming circuit with neural  
369 oscillators for the limbs, rather than by creating a completely new locomotor circuit for ground locomotion  
370 (Ijspeert, 2001; Ijspeert et al., 2007). Modeling reveals that the modulation of two descending drives applied  
371 to the left and right sides of the simulated circuits can vary speed, heading and even the type of gait in the  
372 simulated and robotic bodies. Thus, the patterns generated by CPGs are not rigidly fixed but can be  
373 modulated extensively for adaptive locomotion. Modeling also shows that for simple environments,  
374 locomotion can be generated purely in open-loop, i.e. using CPG models without sensory feedback (Ijspeert  
375 et al., 2007). This is due to the high mechanical stability of the sprawling posture of salamanders, with a  
376 large support polygon and a low center of mass. This therefore supports our hypothesis that mechanically  
377 stable locomotion can largely be CPG-driven (Fig. 3). But sensory feedback loops also play an important  
378 role. Local proprioceptive (stretch) feedback can reduce the variability of intersegmental phase lags toward  
379 values appropriate for locomotion and can simplify the generation of different motor behaviors (Knüsel et  
380 al., 2020). Furthermore, sensory feedback could, in principle, also replace direct coupling of CPG oscillators  
381 (Suzuki et al., 2021), similarly to what has been found for lamprey-like swimming (Thandiackal et al., 2021)

## 382 **Models of mammalian legged locomotion inform our understanding of sensory** 383 **feedback and task coordination**

384 The importance of sensory feedback in mammalian legged locomotion has been investigated by Ekeberg  
385 and Pearson who developed a neuromechanical simulation of cat (Fig. 5E) with a focus on hindlimbs  
386 (Ekeberg & Pearson, 2005). They studied two types of feedback involved in determining the duration of the  
387 stance in a leg: one proportional to limb loading and the other proportional to hip extension. Their model  
388 did not include a CPG. The study showed that these two types of sensory feedback can generate interlimb  
389 coordination corresponding to an alternating gait and can handle irregular terrain. The findings also  
390 suggested that limb loading is more important than hip extension in providing sensory feedback. A related  
391 study investigated the role of hip extension feedback and CPGs in rat locomotor adaptation to split-belt  
392 conditions, in which the right and left legs are on belts with different speeds (Fujiki et al., 2018). Under these  
393 conditions, hip extension could be used as a phase-resetting signal to keep the CPG entrained with the  
394 mechanical movements of the legs, such as to adapt to the split-belt conditions. These findings are  
395 consistent with the hypothesized proximo-distal differentiation in control mechanisms summarized in Fig.  
396 4.

397 Controllers inspired by animal CPGs and reflex loops have become popular for the control of legged robots  
398 and have also provided new scientific insights (Aoi et al., 2017; Bellegarda & Ijspeert, 2022; Fukuoka et al.,  
399 2003; Manoonpong et al., 2007). For instance, load feedback similar to that of Ekeberg and Pearson was  
400 used to generate different gaits on a quadruped robot (Fig. 5D) (Owaki et al., 2013; Owaki & Ishiguro, 2017).  
401 Each limb was driven by a local oscillator without direct coupling to other oscillators, representing CPGs  
402 without direct couplings. Local loading feedback was sufficient to generate stable gaits, showing the  
403 importance of the body and physical interactions as mechanisms for synchronizing CPGs. Interestingly,  
404 transitions between walking, trotting and galloping gaits could be produced by simply increasing the  
405 frequency of the oscillators, as previously shown in decerebrated cats (Shik et al., 1966). This shows the  
406 potential role of sensory feedback as a task coordination mechanism (in addition to a role in perturbation  
407 responses). Similar principles have been shown in insect models, for instance, for the stick insect (Cruse,  
408 1990; Cruse et al., 1995; Daun-Gruhn, 2011; Schilling et al., 2013).

409 More detailed models of mammalian CPGs have been developed by Rybak and colleagues (Danner et al.,  
410 2016, 2017; Markin et al., 2016; Rybak et al., 2015). These models reveal how CPG circuits with two layers,  
411 one for rhythm generation and the other for pattern formation, can produce several features of quadruped  
412 locomotion in open loop (without a body); even gait transitions can be produced by the activation of specific  
413 descending pathways (Danner et al., 2017). Thus, it appears that multiple mechanisms could explain gait  
414 transition (consistent with our view that animal neuromechanical systems show high redundancy); some  
415 transitions are mainly induced through descending modulation, whereas others are mainly driven by  
416 sensory feedback. The development of new full-body models of mammalian musculoskeletal systems (see,  
417 for instance, Tata Ramalingasetty et al., 2021), will allow the community to investigate gait generation and  
418 modulation in more detail. Overall, evidence suggests that CPGs and sensory feedback are equally  
419 important in gait generation in quadruped mammals. It is likely that gaits in complex environments require  
420 more complex modulation of descending pathways than in simple environments, and therefore involve  
421 more cephalized, rather than spinalized, control. Similarly, slow gaits might require more descending  
422 modulation than fast gaits, to allow the animal to maintain posture and balance (Fig. 3).

## 423 **Numerical models suggest that biped locomotion relies more on sensory feedback than** 424 **on CPGs**

425 Some of the first neuromechanical simulations of biped locomotion were developed by Taga and  
426 colleagues (Taga, 1995, 1998; Taga et al., 1991). These simulations of human walking demonstrated how  
427 robust locomotion could emerge from the interaction of CPGs and reflexes. Multiple studies have since  
428 explored the interplay of CPGs and reflexes in biped locomotion with simulations and robots (Aoi et al.,  
429 2019; Fujiki et al., 2015; Geyer & Herr, 2010; H. X. Ryu & Kuo, 2021; Van der Noot et al., 2018, 2019). A  
430 particularly influential neuromechanical model developed by Geyer and Herr (2010) demonstrated that a  
431 limited number of reflexes can generate stable locomotion without the need for CPG circuits (Fig. 5G). The  
432 simulated gaits closely match human gait recordings in terms of joint kinematics, ground reaction forces  
433 and muscle activities. Although the model did not include a CPG, it did depend on a finite-state machine  
434 that selectively activates and deactivates reflexes, so that they are only active at particular phases of the  
435 locomotor cycle. One could argue such a gating mechanism represents a similar function as a CPG and  
436 could in fact be implemented in a CPG circuit.

437 The Geyer and Herr model has inspired several follow-up models to investigate gait modulation (Dzeladini  
438 et al., 2014; Ramadan et al., 2022; Russo et al., 2021; Song & Geyer, 2015), 3D walking and running (Wang  
439 et al., 2012), and locomotor pathologies (Bruehl et al., 2022; Ong et al., 2019; Song & Geyer, 2018). Control  
440 of speed is difficult to achieve with a purely sensory-driven circuit, as it requires the modulation of multiple

441 reflex gains with non-linear functions. Dzeladini et al. (2014) demonstrated that adding CPG circuits to the  
442 sensory-driven model of Geyer and Herr could simplify the control of speed (Fig. 5H). They achieved speed  
443 regulation over a large range using simple drive signals that modulated the frequencies and amplitudes of  
444 the CPG oscillations. Interestingly, they obtained the best speed control by adding oscillators only to the  
445 muscles controlling the hip joints. These simulations support the hypothesis proposed by Daley et al.  
446 (2007) – based on perturbations to running gaits of birds – that proximal joints are controlled with higher-  
447 gain feedforward (CPG) signals and distal joints are controlled by higher-gain feedback signals (Fig. 4).

448 Overall, it is clear from these simulation studies that sensory-feedback loops are essential for human  
449 locomotion, supporting our hypothesis that mechanically unstable locomotion is more sensory driven than  
450 CPG driven (Fig. 3). CPGs are likely to contribute mainly to speed and gait modulation. Interestingly, the  
451 simulations of Geyer & Herr (2010) and most follow-up simulations are not capable of producing slow and  
452 very slow walking, because the simulated models simply fall. This suggests more sophisticated balance  
453 control mechanisms are missing in the models, and hence these models do not yet properly implement the  
454 more important role of descending modulation that we hypothesize for unstable locomotion (Fig. 3).

455 The neuromechanics of bipedal locomotion of birds has been less commonly modeled than human  
456 locomotion. One neuromechanical simulation study demonstrated that the sensory-driven control  
457 architectures of Geyer & Herr (2010) and Wang et al. (2012) could be adapted to control an ostrich-like  
458 body (Geijtenbeek et al., 2013). A number of bird-like robots have also been developed (Apgar et al., 2018;  
459 Badri-Spröwitz et al., 2022; Pratt et al., 2001). Among these, BirdBot is particularly interesting, because it  
460 implements a bird-inspired tendon system that uses mechanical coupling to control leg stiffness in the  
461 stance and swing phases (Fig. 5F). The mechanically coupled design provides self-stable and energy-  
462 efficient walking and running with simple CPG-based control and no sensory feedback, highlighting  
463 intrinsically stable features of the avian distal limb. This represents therefore another source of  
464 redundancy, in which the intrinsic musculoskeletal mechanics contributes to intralimb coordination and  
465 balance, aspects that are often considered to be pure control problems.

## 466 **Open questions**

467 Here, we have reviewed historical and current perspectives on the organization of locomotor circuits based  
468 on experimental and modeling evidence. Although there are shared features of the component elements  
469 across vertebrates, there is also diversity in the relative contributions of these components. We have  
470 proposed several hypotheses about how the respective roles of CPGs, reflexes and descending modulation  
471 vary across vertebrates: depending on body size, the mechanical instability of gaits, the speed of  
472 locomotion and the developmental time to locomotor maturity. We have also hypothesized that these roles  
473 can vary within the body, between proximal to distal joints, and depending on the speed and gait. However,  
474 to rigorously confirm or reject these hypotheses, we need further integration of animal experiments and  
475 neuromechanical simulations.

476 Numerical models can be particularly useful to test some of these hypotheses. For instance,  
477 neuromechanical simulations of a pendulum (which can be viewed as a very simple model of a leg) have  
478 shown that periodic behaviors can be obtained by either purely feedback or purely feedforward (CPG-  
479 based) mechanisms; however, circuits that combine both feedback and feedforward contributions are  
480 more robust against unexpected disturbances and sensorimotor noise (Kuo, 2002). Similar results are  
481 obtained with a simulation of biped locomotion (Ryu & Kuo, 2021). An interesting proposition from Kuo and  
482 colleagues is that CPGs can be viewed as ‘state estimators’ that predict the state of limbs (and therefore

483 sensory signals). Based on this, a CPG could be viewed as a “filter for processing sensory information  
484 rather than as a generator of commands” (Kuo, 2002). In our view, this perspective underestimates the role  
485 of CPGs in coordinating and modulating locomotion (e.g. for regulating speed, gait and heading), but it has  
486 the merit of analyzing the tradeoff between feedforward and feedback control using rigorous optimal  
487 estimation principles. Importantly, the potential roles of the CPG as a pattern generator and a state  
488 estimator are not necessarily mutually exclusive – CPGs may act as a type of internal model that filters  
489 sensory inputs, estimates current state and generates rhythmic outputs based on current state estimates.

490 Another important open question is related to the degree of centralization of locomotion control and to the  
491 strength of inter-oscillator couplings within and across animals (Aoi et al., 2017; Holmes et al., 2006; Neveln  
492 et al., 2019; Revzen et al., 2009). Historically, fictive locomotion experiments gave the impression that  
493 locomotor patterns were mainly generated by CPGs, and that inter-oscillator couplings serve as the  
494 mechanism for inter-joint coordination. However, as presented above, modeling has shown that sensory  
495 feedback is another mechanism for synchronization that can replace inter-oscillator couplings (Cruse et  
496 al., 1995; Owaki et al., 2013; Thandiackal et al., 2021, Suzuki et al 2021). Biological evidence also shows  
497 that local sensory feedback is directly integrated into segmentally distributed CPGs (Grillner et al., 1981;  
498 Grillner & Wallén, 1984; Pearson, 2008; Rossignol et al., 2006; Whelan, 1996). Inter-oscillator couplings  
499 might therefore be weaker than previously thought. This relatively decentralized control organization would  
500 allow for flexible motor patterns to be adapted to environmental constraints, through sensing and  
501 modulation of descending pathways.

502 Neveln et al. (2019) have proposed an interesting framework based on mutual information to quantify  
503 centralization in animal locomotion, which could help systematically investigate the degree of  
504 centralization across species and conditions. They suggest that locomotor coordination “could either be  
505 achieved through strong, global coupling with dense connections between components” – representing  
506 high centralization – or “through weak, local coupling with sparse connections”, representing low  
507 centralization. Furthermore, centralization can also be affected by the strength of mechanical coupling and  
508 the organization of sensory feedback, whether it is processed centrally or locally (Holmes et al., 2006). They  
509 tested their approach in simulation, with robots, and in cockroach experiments. This model-free, empirical  
510 method of quantifying centralization will be useful for analyzing future neuromechanical models and  
511 animal experiments. It is likely that the level of centralization depends on the morphology and stability of  
512 locomotion, as well as on environmental conditions (speed of locomotion and complexity of the  
513 environment). Consistent with this, the strength of coupling between legs appears to be speed-dependent  
514 in invertebrates (*Drosophila*), with no coupling at low speeds of walking and high coupling at high speeds  
515 (Berendes et al 2016). Further research is needed to clarify the mechanisms that enable variation in  
516 coupling strength with speed, and to understand the diversity of oscillator coupling strengths across  
517 species with varying locomotor demands.

518 Concerning descending modulation, we do not yet know exactly how many independent descending  
519 pathways exist, how many local spinal locomotor circuits they project to (global versus local joint-specific  
520 projections), and their effect (activating oscillators, changing a joint offset or modulating reflexes, for  
521 instance). From several studies (Arber & Costa, 2022; Ferreira-Pinto et al., 2018; Rossignol et al., 2006), we  
522 know that descending projections present a mix of these properties. Modeling studies have started  
523 exploring how different aspects of legged locomotion (e.g. frequency, step size, ground clearance and  
524 others) can be modulated by descending pathways (Song & Geyer 2015, Bellegarda & Ijspeert 2022,  
525 Ramadan et al. 2022). But more studies are needed to investigate the diversity of descending pathways  
526 across species and how they relate to motor behaviors and mechanical features of the body. Decoding  
527 descending pathways will be particularly important in allowing us to understand what types of voluntary

528 movements an animal can perform (for instance, for gait transitions and for limb placement in visually  
529 guided locomotion). Animals can smoothly switch between steady-state locomotion and highly modulated  
530 locomotion as needed when crossing a complex terrain. It is likely that this is done by switching from  
531 activating a small number of descending pathways to more complex time-varying activations of multiple  
532 descending pathways. This is related to the concept of relatively spinalized versus cephalized control,  
533 discussed above. Also, some animals appear to be better than others at performing fine-tuned movements,  
534 and this may reflect a higher number of descending pathways and a larger role of descending modulation  
535 in mammals than in amphibians, for example (Fig. 3). Integration of experimental and modeling work is  
536 needed to test these ideas.

## 537 **Conclusions and outlook**

538 We envision a bright future for the next 100 years of research in this area, with exciting opportunities to  
539 integrate experiments and modeling to address open questions about the neuromechanical control of  
540 locomotion. Thanks to new imaging techniques and genetic identification methods, future full atlases and  
541 connectomes of spinal circuits and descending and ascending pathways will be tremendously useful to  
542 improve our understanding of the underlying circuits involved in vertebrate locomotion. For instance, by  
543 quantifying the proportion of sensory neurons within the spinal cord and the number of independent  
544 descending pathways (Arber & Costa, 2022; Ferreira-Pinto et al., 2018), it will be possible to more  
545 quantitatively estimate the respective roles of sensory feedback, CPG and descending modulation across  
546 different vertebrate animals. We hope that these techniques will not be limited to classic genetic model  
547 animals (e.g. zebrafish and mice), but also used extensively across diverse species to allow comparison  
548 between different morphologies and locomotor modes. In particular, compared to the rich literature on  
549 terrestrial locomotion, there has been relatively little research on diversity of sensorimotor control  
550 mechanisms among flying vertebrates, which is an important area for further study.

551 Additionally, advances in opto- and chemogenetic techniques represent a tremendous opportunity to  
552 selectively activate or deactivate specific cell types, performing experiments that were previously possible  
553 only in simulation. There are exciting opportunities for new 'virtual twin' experiments that combine  
554 experimental technologies and computing power for simulations; for example, by creating  
555 neuromechanical simulations that replicate animal behavior in real-time, it could become feasible to  
556 conduct state-dependent animal experiments, in which a perturbation is applied when a modeled internal  
557 state from the simulation (e.g. tension in a tendon or phase of an oscillator) reaches a specific threshold.  
558 It might also become possible to create hybrid experiments in which a spinal cord preparation is connected  
559 in closed loop with a musculoskeletal simulation moving in a virtual physics-based environment. In such  
560 an experiment, recorded activity from ventral roots would be used to activate simulated muscles, and virtual  
561 sensory signals from the simulated moving body would be used to stimulate sensory neurons. Such  
562 preparations would allow one to record and investigate spinal cord circuits with all the technologies  
563 available for controlled bench experiments while still approximating *in vivo* conditions of unconstrained  
564 locomotion. These kinds of integrative studies will be essential for testing hypotheses about of the  
565 fundamental principles of locomotion in vertebrates, understanding how control varies among species, and  
566 for guiding functional restoration and therapeutic approaches such as electric epidural stimulation (van  
567 den Brand et al., 2012; Wagner et al., 2018).

## Figure legends:

**Figure 1. Schematic of the neuromechanical system of vertebrates, including the brain, descending drive, spinal networks and intrinsic musculoskeletal mechanics.** Temporal scaling of control arises from the spatial distribution of the system components and delays inherent to animal sensorimotor systems. Central pattern generators (CPGs) in the spinal column receive relatively simple descending signals and generate complex rhythmic motor outputs. The CPG rhythm is entrained by sensory feedback in intact animals but generates fictive locomotor patterns in the absence of feedback. Sensory feedback acts in multiple layers, through 1) short-latency monosynaptic reflexes, 2) entraining CPGs, 3) longer latency multi-synaptic sensory feedback, and 4) ascending pathways that contribute to internal models, task planning and modulation of descending commands. Efferent copy from the spinal networks also contributes input into internal models, enabling prediction of sensory signals that are compared to sensory feedback. The plus symbol indicates summation of multiple signal paths to the motor neurons (MN).

**Figure 2 Hypothesized differences in the integration of mechanics and control between small and large animals.** **A)** Estimated differences in delays between a shrew and an elephant (More and Donelan 2018; Thangal & Donelan, 2020). Small animals have faster reflex responses relative to movement durations, but synaptic delays are a larger fraction of reflex delays (More and Donelan 2018). Large animals have relatively longer inertial delays (Thangal & Donelan, 2020). **B)** These differences may lead small animals to rely more on reflex feedback, with higher-gain short-latency reflexes (indicated by thicker arrows in B) and intrinsic mechanical preflexes for corrective responses. **C)** In contrast, inertial delays exceed reflex delays in the largest animals, suggesting that reflexes and intrinsic mechanics may not be sufficient to allow stable corrective responses. Consequently, it is expected that large animals must rely more on higher-gain sensory input to internal models for predictive control (indicated by thicker arrows in C). Predictive control is enabled by computations in the brain involving many synapses. The ratio of reflex delay to synaptic delay is much greater in large animals compared to small animals, suggesting a lower penalty for increased computational complexity.

**Figure 3: Hypothesized control gradients in the diversity of animal locomotion.** **A)** We hypothesize that the relative roles of spinal sensing and reflexes, CPGs and descending modulation vary among species and between gaits depending on several factors: size, static mechanical stability/instability (estimated based on the ratio between the height of the center of mass and the size of the support polygon), cycle period, which decreases with speed, and time to locomotor maturity, which varies substantially between precocial and altricial species. Animals on the left of these axes rely more on CPGs whereas animals on the right rely more on spinal sensing and reflex, and on descending modulation. We hypothesize that the functional gradients shown exist across taxa; nonetheless, phylogenetic differences are not represented here, and the contributions of descending control likely vary substantially among taxa. The gradients should be interpreted conceptually rather than as an absolute scaling. **B)** Static mechanical instability is related to the ratio of the height of the center of mass compared to the size of the support polygon.

**Figure 4. Hypothesized proximo-distal differentiation in the balance of feedforward and feedback control of limb muscles.** Due to differences in muscle–tendon architecture and inertia of the proximal versus distal limb, it is expected that proximal muscles exhibit higher-gain feedforward control, with length and position feedback entraining the rhythm of the CPG oscillators, influencing stance and swing frequencies. In contrast, distal muscles, with higher compliance and lower inertia, are expected to have higher-gain short-latency reflexes, and higher contributions from intrinsic mechanics ('preflexes'). The width of the arrows showing the reflexes (purple) and CPG (green) are proportional to the hypothesized gains. For clarity, the peripheral circuits are drawn only for the flexors, but similar connections exist for the extensors. The plus symbol indicates summation of multiple signal paths to the motor neurons (MN).

**Figure 5. Robots and neuromechanical simulations.** Multiple robots and neuromechanical simulations have been used to investigate the roles of CPGs, sensory feedback and mechanical properties in the generation of animal locomotion. (A) Ekeberg (1993), image used with permission from Springer Nature.

621 (B) Thandiackal et al. (2021). (C) Ijspeert et al. (2007) and Crespi et al. (2013), image used with  
622 permission from IEEE. (D) Owaki et al. (2013), image used with permission from The Royal Society  
623 Publishing. (E) Ekeberg and Pearson (2005), image used with permission from the American  
624 Physiological Society. (F) Badri-Sprowitz et al. (2022), image reprinted with permission from AAAS. (G)  
625 Geyer and Herr (2010), image used with permission from IEEE. (H) Dzeladini et al. (2014).  
626

## Glossary

**Altricial:** Animals that take extended time after birth to reach locomotor maturity.

**Central pattern generator (CPG):** Neural circuits that can generate the basic rhythmic motor patterns for movement and breathing without any sensory inputs. These circuits produce periodic signals that are often mathematically modeled as oscillators (see below). A defining feature of a CPG is that it can generate a periodic motor output without a periodic input.

**Compliance:** Elastic deformation of a mechanical system. Compliance can be viewed as the opposite of stiffness.

**Control theory:** Field of applied mathematics that deals with the control of dynamical and engineered systems.

**Decerebrate preparations:** An experimental manipulation in which cerebral brain function is eliminated by transection or removal of the cerebrum. The extent to which longer-latency feedback pathways remain intact or eliminated depends on the specific location of the transection.

**Dynamic stability:** Stability of a gait that requires movement to prevent falling (as opposed to static stability, see below). Typically, a dynamically stable gait exhibits convergence to a limit cycle behavior, namely periodic behavior that is robust against (small) perturbations.

**Efferent copy:** A copy of the motor signals that are used as inputs to internal models to predict dynamics and sensory feedback.

**Entrainment:** Synchronization of oscillatory dynamical systems such that they converge to the same frequency and therefore to constant phase differences. Two dynamical systems can mutually entrain each other (and converge to a frequency that is typically an average of their intrinsic frequencies), or one dynamical system can entrain another (and impose its intrinsic frequency on the other).

**Feedback control:** A control pathway in which sensory signals are returned back to generate an error signal that regulates the input commands towards desired output dynamics.

**Feedforward control:** A control pathway that generates a predefined command signal based on the anticipated load and dynamics of the system. In this article, we discuss two types of feedforward control mechanisms: spinal CPGs and supraspinal internal models.

**Fictive locomotion:** The generation of the basic rhythmic muscle activation patterns required for locomotion in isolated spinal cords, such as the alternating activation of flexors and extensors in the



leg of walking animals or the transmission of an undulatory wave down the body in swimming animals.

**Internal models:** Internal neural representations that can predict the interactions between the nervous system, the musculoskeletal system and the environment. Forward internal models can predict causal relationships between actions and their consequences. Inverse internal models can predict which actions are needed to reach particular consequences.

**Model-based control:** Control architecture that uses internal models for performing anticipatory (as opposed to reactive) movements. An example of model-based control in robotics is model-predictive control, which uses a model of the robot and an optimization criterion to define motor commands over a finite-time horizon.

**Neuromechanics:** The scientific field focused on the interactions between biomechanics of the musculoskeletal system and sensorimotor control and their integration for robust and agile movement.

**Oscillators:** Dynamical systems or neural networks that exhibit stable limit cycle behavior, i.e. they produce periodic signals.

**Precocial:** Animals that locomote effectively shortly after birth.

**Preflexes:** The intrinsic dynamic properties of the musculoskeletal system in its activated state that automatically stabilize movements through visco-elastic properties. These form a kind of zero-delay feedback stabilizing movement.

**Sensorimotor delay:** The time lag between the onset of a mechanical perturbation and its reception by a sensory organ to the development of peak muscle force in response to the perturbation. It includes sensing delay, nerve conduction delay, synaptic delay, neuromuscular junction delay, electromechanical delay and muscle force development delay.

**Spinal preparations:** An experimental manipulation in which the brain and brainstem function are completely removed by transection of the spinal cord, typically in the thoracic region.

**Static stability:** Stability of a posture or a gait in which the center of mass is always maintained above the (possibly time-varying) support polygons formed by the contacts between the limbs (or any body parts) and the ground. An animal that is statically stable will not fall when it freezes its posture.

**Box 1: Central Pattern Generators (CPGs)**

CPGs play important roles in the generation of coordinated motor patterns for both vertebrate and invertebrate locomotion. CPGs are neural circuits that can generate the basic rhythmic motor patterns for movement and breathing without any sensory inputs. Locomotor CPG circuits are located in the spinal cord of vertebrates (Grillner and El Manira, 2019), and in the ventral nerve cord of invertebrates (Mantziaris et al., 2020). The existence of CPGs has been demonstrated across diverse vertebrate species through the observation of fictive locomotion in spinal preparations, with all inputs from the brain and periphery transected (Gray, 1936; Gray and Lissmann, 1940; Grillner and Wallén, 1982; Ho and O'Donovan, 1993; Sholomenko et al., 1991; Sholomenko and Steeves, 1987; Ten Cate, 1964, 1965). The CPG circuits in the isolated spinal cord generate fictive swimming in aquatic species (e.g. lamprey, zebrafish), fictive walking in terrestrial species (birds, mammals), and both fictive swimming and fictive walking in amphibious species (e.g. salamanders) (Chevallier et al., 2008; Fetcho and McLean, 2010; Grillner and El Manira, 2019; Grillner and Wallen, 1985; Ryczko et al., 2010; Whelan, 1996). Genetic studies have identified specific subpopulations of interneurons involved in rhythm generation that are conserved across vertebrates (Grillner and El Manira, 2019; Kiehn, 2016; Rybak et al., 2015). Locomotor CPGs are distributed segmentally along the spinal cord as interconnected rhythmic units (Grillner et al., 1995; Kiehn, 2016; McLean and Dougherty, 2015; Rybak et al., 2015), typically one per pair of antagonist muscles or even one per muscle (Cheng et al., 1998). Specific ventrolateral regions of the spinal cord generate flexor/extensor alternation, and specific ventromedial interneurons generate left/right coordination (Kiehn, 2016; McLean and Dougherty, 2015). Note that although there is no direct evidence of CPG circuits in humans, there is ample indirect evidence that humans possess CPG circuits similar to those of other vertebrate species (Minassian et al., 2017).

630

631 **Acknowledgements**

632 We would like to thank Ansgar Büschges, Pavan Ramdya, and two anonymous reviewers for useful  
633 comments on a previous version of the manuscript. Thanks to Brooke Christensen for contributing animal  
634 illustrations for the figures. A.J.I. is financially supported by the EPFL and by the ERC Synergy grant  
635 Salamandra (951477). M.A.D. is supported by UCI and the NSF (2016049, 2021832). Some ideas  
636 represented in this review arose in the workshop on Integrative Organismal Modeling of Movement  
637 supported by the NSF (2040544-to M.A.D.).

638

## References:

- 640 Af Klint, R., Mazzaro, N., Nielsen, J. B., Sinkjaer, T., & Grey, M. J. (2010). Load rather than length sensitive  
641 feedback contributes to soleus muscle activity during human treadmill walking. *Journal of*  
642 *Neurophysiology*, 103(5), 2747–2756.
- 643 Alexander, R. M. (2002). Stability and Manoeuvrability of Terrestrial Vertebrates. *Integrative and*  
644 *Comparative Biology*, 42(1), 158–164. <https://doi.org/10.1093/icb/42.1.158>
- 645 Aoi, S., Manoonpong, P., Ambe, Y., Matsuno, F., & Wörgötter, F. (2017). Adaptive Control Strategies for  
646 Interlimb Coordination in Legged Robots: A Review. *Frontiers in Neurorobotics*, 11.  
647 <https://doi.org/10.3389/fnbot.2017.00039>
- 648 Aoi, S., Ohashi, T., Bamba, R., Fujiki, S., Tamura, D., Funato, T., Senda, K., Ivanenko, Y., & Tsuchiya, K. (2019).  
649 Neuromusculoskeletal model that walks and runs across a speed range with a few motor control  
650 parameter changes based on the muscle synergy hypothesis. *Scientific Reports*, 9(1), 369.  
651 <https://doi.org/10.1038/s41598-018-37460-3>
- 652 Apgar, T., Clary, P., Green, K., Fern, A., & Hurst, J. W. (2018). Fast Online Trajectory Optimization for the  
653 Bipedal Robot Cassie. *Robotics: Science and Systems*.
- 654 Arber, S., & Costa, R. M. (2022). Networking brainstem and basal ganglia circuits for movement. *Nature*  
655 *Reviews Neuroscience*, 23(6), Article 6. <https://doi.org/10.1038/s41583-022-00581-w>
- 656 Ashby, W. R. (1957). *An introduction to cybernetics*. Chapman & Hall Ltd.  
657 <http://dspace.utalca.cl/handle/1950/6344>
- 658 Azizi, E., Brainerd, E. L., & Roberts, T. J. (2008). Variable gearing in pennate muscles. *Proceedings of the*  
659 *National Academy of Sciences*, 105(5), 1745–1750. <https://doi.org/10.1073/pnas.0709212105>
- 660 Badri-Spröwitz, A., Aghamaleki Sarvestani, A., Sitti, M., & Daley, M. A. (2022). BirdBot achieves energy-  
661 efficient gait with minimal control using avian-inspired leg clutching. *Science Robotics*, 7(64),  
662 eabg4055. <https://doi.org/10.1126/scirobotics.abg4055>
- 663 Barbeau, H., & Rossignol, S. (1987). Recovery of locomotion after chronic spinalization in the adult cat.  
664 *Brain Research*, 412(1), 84–95. [https://doi.org/10.1016/0006-8993\(87\)91442-9](https://doi.org/10.1016/0006-8993(87)91442-9)
- 665 Bellegarda, G., & Ijspeert, A. (2022). CPG-RL: Learning Central Pattern Generators for Quadruped  
666 Locomotion. *IEEE Robotics and Automation Letters*, 7(4), 12547–12554.  
667 <https://doi.org/10.1109/LRA.2022.3218167>
- 668 Bennett, P. M., & Harvey, P. H. (1985). Brain size, development and metabolism in birds and mammals.  
669 *Journal of Zoology*, 207(4), 491–509. <https://doi.org/10.1111/j.1469-7998.1985.tb04946.x>
- 670 Berendes, V., Zill, S. N., Büschges, A., & Bockemühl, T. (2016). Speed-dependent interplay between local  
671 pattern-generating activity and sensory signals during walking in *Drosophila*. *Journal of*  
672 *Experimental Biology*, 219(23), 3781–3793. <https://doi.org/10.1242/jeb.146720>
- 673 Bicanski, A., Ryczko, D., Knuesel, J., Harischandra, N., Charrier, V., Ekeberg, Ö., Cabelguen, J.-M., & Ijspeert,  
674 A. J. (2013). Decoding the mechanisms of gait generation in salamanders by combining  
675 neurobiology, modeling and robotics. *Biological Cybernetics*, 107(5), 545–564.  
676 <https://doi.org/10.1007/s00422-012-0543-1>
- 677 Bidaye, S.S., Bockemühl, T. and Büschges, A., 2018. Six-legged walking in insects: how CPGs, peripheral  
678 feedback, and descending signals generate coordinated and adaptive motor rhythms. *Journal of*  
679 *neurophysiology*, 119(2), pp.459-475.
- 680 Biewener, A. A. (1989). Scaling Body Support in Mammals: Limb Posture and Muscle Mechanics. *Science*,  
681 245(4913), 45–48. <https://doi.org/10.1126/science.2740914>
- 682 Brown, I. E., & Loeb, G. E. (2000). A Reductionist Approach to Creating and Using Neuromusculoskeletal  
683 Models. In J. M. Winters & P. E. Crago (Eds.), *Biomechanics and Neural Control of Posture and*

- 684            *Movement* (pp. 148–163). Springer. [https://doi.org/10.1007/978-1-4612-2104-3\\_10](https://doi.org/10.1007/978-1-4612-2104-3_10)
- 685 Bruel, A., Ghorbel, S. B., Di Russo, A., Stanev, D., Armand, S., Courtine, G., & Ijspeert, A. (2022). Investigation  
686 of neural and biomechanical impairments leading to pathological toe and heel gaits using  
687 neuromusculoskeletal modelling. *The Journal of Physiology*, 600(11), 2691–2712.  
688 <https://doi.org/10.1113/JP282609>
- 689 Buchanan, J. T. (1992). Neural network simulations of coupled locomotor oscillators in the lamprey spinal  
690 cord. *Biological Cybernetics*, 66, 367–374.
- 691 Buschmann, T., Ewald, A., von Twickel, A. and Bueschges, A., 2015. Controlling legs for locomotion—  
692 Insights from robotics and neurobiology. *Bioinspiration & biomimetics*, 10(4), p.041001.
- 693 Buschges, A., 2005. Sensory control and organization of neural networks mediating coordination of  
694 multisegmental organs for locomotion. *Journal of neurophysiology*, 93(3), pp.1127-1135.
- 695 Cabelguen, J. M., Bourcier-Lucas, C., & Dubuc, R. (2003). Bimodal locomotion elicited by electrical  
696 stimulation of the midbrain in the salamander *Notophthalmus viridescens*. *The Journal of*  
697 *Neuroscience*, 23(6), 2434–2439.
- 698 Cheng, J., Stein, R. B., Jovanovic, K., Yoshida, K., Bennett, D. J., & Han, Y. (1998). Identification, Localization,  
699 and Modulation of Neural Networks for Walking in the Mudpuppy (*Necturus Maculatus*) Spinal  
700 Cord. *The Journal of Neuroscience*, 18(11), 4295–4304.
- 701 Chevallier, S., Ijspeert, A. J., Ryczko, D., Nagy, F., & Cabelguen, J.-M. (2008). Organisation of the spinal  
702 central pattern generators for locomotion in the salamander: Biology and modelling. *Brain Research*  
703 *Reviews*, 57(1), 147–161. <https://doi.org/10.1016/j.brainresrev.2007.07.006>
- 704 Cohen, A. H., Holmes, P. J., & Rand, R. (1982). The nature of coupling between segmented oscillations and  
705 the lamprey spinal generator for locomotion: A mathematical model. *J. Math. Biol.*, 13, 345–369.
- 706 Crespi, A., Karakasiliotis, K., Guignard, A., & Ijspeert, A. J. (2013). Salamandra Robotica II: An amphibious  
707 robot to study salamander-like swimming and walking gaits. *IEEE Transactions on Robotics*, 29(2),  
708 308–320. <https://doi.org/10.1109/TRO.2012.2234311>
- 709 Cruse, H. (1990). What mechanisms coordinate leg movement in walking arthropods? *Trends in*  
710 *Neurosciences*, 13(1), 15–21.
- 711 Cruse, H., Brunn, D. E., Bartling, Ch., Dean, J., Dreifert, M., Kindermann, T., & Schmitz, J. (1995). Walking: A  
712 complex behavior controlled by simple networks. *Adaptive Behavior*, 3(4), 385–418.
- 713 Daley, M. A. (2018). Understanding the agility of running birds: Sensorimotor and mechanical factors in  
714 avian bipedal locomotion. *Integrative and Comparative Biology*, 58(5), 884–893.
- 715 Daley, M. A., & Biewener, A. A. (2011). Leg muscles that mediate stability: Mechanics and control of two  
716 distal extensor muscles during obstacle negotiation in the guinea fowl. *Philosophical Transactions*  
717 *of the Royal Society B: Biological Sciences*, 366(1570), 1580–1591.
- 718 Daley, M. A., & Birn-Jeffery, A. (2018). Scaling of avian bipedal locomotion reveals independent effects of  
719 body mass and leg posture on gait. *Journal of Experimental Biology*, 221(10), jeb152538.
- 720 Daley, M. A., Felix, G., & Biewener, A. A. (2007). Running stability is enhanced by a proximo-distal gradient  
721 in joint neuromechanical control. *Journal of Experimental Biology*, 210(3), 383–394.  
722 <https://doi.org/10.1242/jeb.02668>
- 723 Daley, M. A., Voloshina, A., & Biewener, A. A. (2009). The role of intrinsic muscle mechanics in the  
724 neuromuscular control of stable running in the guinea fowl. *The Journal of Physiology*, 587(11),  
725 2693–2707.
- 726 Danner, S. M., Shevtsova, N. A., Frigon, A., & Rybak, I. A. (2017). Computational modeling of spinal circuits  
727 controlling limb coordination and gaits in quadrupeds. *ELife*, 6, e31050.  
728 <https://doi.org/10.7554/eLife.31050>
- 729 Danner, S. M., Wilshin, S. D., Shevtsova, N. A., & Rybak, I. A. (2016). Central control of interlimb coordination

730 and speed-dependent gait expression in quadrupeds. *The Journal of Physiology*, 594(23), 6947–  
731 6967. <https://doi.org/10.1113/JP272787>

732 Daun-Gruhn, S. (2011). A mathematical modeling study of inter-segmental coordination during stick insect  
733 walking. *Journal of Computational Neuroscience*, 30(2), 255–278. [https://doi.org/10.1007/s10827-](https://doi.org/10.1007/s10827-010-0254-3)  
734 010-0254-3

735 Dickinson, M. H., Farley, C. T., Full, R. J., Koehl, M. a. R., Kram, R., & Lehman, S. (2000). How Animals Move:  
736 An Integrative View. *Science*, 288(5463), 100–106. <https://doi.org/10.1126/science.288.5463.100>

737 Donelan, J. M., & Pearson, K. G. (2004). Contribution of Force Feedback to Ankle Extensor Activity in  
738 Decerebrate Walking Cats. *Journal of Neurophysiology*, 92(4), 2093–2104.  
739 <https://doi.org/10.1152/jn.00325.2004>

740 Dzeladini, F., van den Kieboom, J., & Ijspeert, A. (2014). The contribution of a central pattern generator in a  
741 reflex-based neuromuscular model. *Frontiers in Human Neuroscience*, 8.  
742 <https://doi.org/10.3389/fnhum.2014.00371>

743 Ekeberg, Ö. (1993). A combined neuronal and mechanical model of fish swimming. *Biological Cybernetics*,  
744 69, 363–374.

745 Ekeberg, Ö., & Grillner, S. (1999). Simulations of neuromuscular control in lamprey swimming. *Philosophical*  
746 *Transactions of the Royal Society B: Biological Sciences*, 354(1385), 895–902.  
747 <https://doi.org/10.1098/rstb.1999.0441>

748 Ekeberg, Ö., Lansner, A., & Grillner, S. (1995). The neural control of fish swimming studied through numerical  
749 simulations. *Adaptive Behavior*, 3(4), 363–384.

750 Ekeberg, Ö., & Pearson, K. (2005). Computer simulation of stepping in the hind legs of the cat: An  
751 examination of mechanisms regulating the stance-to-swing transition. *Journal of Neurophysiology*,  
752 94, 4256–4268.

753 Ekeberg, Ö., Wallén, P., Lansner, A., Traven, H., Brodin, L., & Grillner, S. (1991). A computer-based model for  
754 realistic simulations of neural networks I: The single neuron and synaptic interaction. *Biological*  
755 *Cybernetics*, 65, 81–90.

756 Eng, C. M., Azizi, E., & Roberts, T. J. (2018). Structural Determinants of Muscle Gearing During Dynamic  
757 Contractions. *Integrative and Comparative Biology*, 58(2), 207–218.  
758 <https://doi.org/10.1093/icb/icy054>

759 Ferreira-Pinto, M. J., Ruder, L., Capelli, P., & Arber, S. (2018). Connecting Circuits for Supraspinal Control of  
760 Locomotion. *Neuron*, 100(2), 361–374. <https://doi.org/10.1016/j.neuron.2018.09.015>

761 Fetcho, J. R., & McLean, D. L. (2010). Some principles of organization of spinal neurons underlying  
762 locomotion in zebrafish and their implications. *Annals of the New York Academy of Sciences*,  
763 1198(1), 94–104.

764 Floreano, D., Ijspeert, A. J., & Schaal, S. (2014). Robotics and neuroscience. *Current Biology*, 24(18), R910–  
765 R920. <https://doi.org/10.1016/j.cub.2014.07.058>

766 Fouad, A. D., Teng, S., Mark, J. R., Liu, A., Alvarez-Illera, P., Ji, H., Du, A., Bhirgoo, P. D., Cornblath, E., Guan,  
767 S. A., & Fang-Yen, C. (2018). Distributed rhythm generators underlie *Caenorhabditis elegans*  
768 forward locomotion. *ELife*, 7, e29913. <https://doi.org/10.7554/eLife.29913>

769 Forssberg, H., Grillner, S., Halbertsma, J., & Rossignol, S. (1980). The locomotion of the low spinal cat. II.  
770 Interlimb coordination. *Acta Physiologica Scandinavica*, 108(3), 283–295.  
771 <https://doi.org/10.1111/j.1748-1716.1980.tb06534.x>

772 Fujiki, S., Aoi, S., Funato, T., Sato, Y., Tsuchiya, K., & Yanagihara, D. (2018). Adaptive hindlimb split-belt  
773 treadmill walking in rats by controlling basic muscle activation patterns via phase resetting.  
774 *Scientific Reports*, 8(1), 17341. <https://doi.org/10.1038/s41598-018-35714-8>

775 Fujiki, S., Aoi, S., Funato, T., Tomita, N., Senda, K., & Tsuchiya, K. (2015). Adaptation mechanism of interlimb

776 coordination in human split-belt treadmill walking through learning of foot contact timing: A  
777 robotics study. *Journal of The Royal Society Interface*, 12(110), 20150542.  
778 <https://doi.org/10.1098/rsif.2015.0542>

779 Fukuoka, Y., Kimura, H., & Cohen, A. H. (2003). Adaptive dynamic walking of a quadruped robot on irregular  
780 terrain based on biological concepts. *The International Journal of Robotics Research*, 22(3–4), 187–  
781 202.

782 Full, R. J., & Koditschek, D. E. (1999). Templates and anchors: Neuromechanical hypotheses of legged  
783 locomotion on land. *Journal of Experimental Biology*, 202(23), 3325–3332.  
784 <https://doi.org/10.1242/jeb.202.23.3325>

785 Garwicz, M., Christensson, M., & Psouni, E. (2009). A unifying model for timing of walking onset in humans  
786 and other mammals. *Proceedings of the National Academy of Sciences*, 106(51), 21889–21893.  
787 <https://doi.org/10.1073/pnas.0905777106>

788 Gatesy, S. M., & Biewener, A. A. (1991). Bipedal locomotion: Effects of speed, size and limb posture in birds  
789 and humans. *Journal of Zoology*, 224(1), 127–147.

790 Gatesy, S. M., & Dial, K. P. (1996). Locomotor modules and the evolution of avian flight. *Evolution*, 50(1),  
791 331–340.

792 Geijtenbeek, T., van de Panne, M., & van der Stappen, A. F. (2013). Flexible muscle-based locomotion for  
793 bipedal creatures. *ACM Transactions on Graphics*, 32(6), 206:1-206:11.  
794 <https://doi.org/10.1145/2508363.2508399>

795 Geyer, H., & Herr, H. (2010). A Muscle-Reflex Model That Encodes Principles of Legged Mechanics  
796 Produces Human Walking Dynamics and Muscle Activities. *IEEE Transactions on Neural Systems  
797 and Rehabilitation Engineering*, 18(3), 263–273. <https://doi.org/10.1109/TNSRE.2010.2047592>

798 Gorassini, M. A., Prochazka, A., Hiebert, G. W., & Gauthier, M. J. (1994). Corrective responses to loss of  
799 ground support during walking. I. Intact cats. *Journal of Neurophysiology*, 71(2), 603–610.  
800 <https://doi.org/10.1152/jn.1994.71.2.603>

801 Graham Brown, T. (1911). The Intrinsic Factors in the Act of Progression in the Mammal. *Proceedings of  
802 the Royal Society of London. Series B*, 84(572), 308–319.

803 Graham Brown, T. (1914). On the nature of the fundamental activity of the nervous centres; together with  
804 an analysis of the conditioning of rhythmic activity in progression, and a theory of the evolution of  
805 function in the nervous system. *The Journal of Physiology*, 48(1), 18–46.  
806 <https://doi.org/10.1113/jphysiol.1914.sp001646>

807 Gray, J. (1933). Studies in Animal Locomotion: I. The Movement of Fish with Special Reference to the Eel.  
808 *Journal of Experimental Biology*, 10(1), 88–104. <https://doi.org/10.1242/jeb.10.1.88>

809 Gray, J. (1936). Studies in Animal Locomotion: V. Resistance Reflexes in the Eel. *Journal of Experimental  
810 Biology*, 13(2), 181–191. <https://doi.org/10.1242/jeb.13.2.181>

811 Gray, J., & Lissmann, H. W. (1938). Studies in Animal Locomotion: VII. Locomotory Reflexes in the  
812 Earthworm. *Journal of Experimental Biology*, 15(4), 506–517. <https://doi.org/10.1242/jeb.15.4.506>

813 Gray, J., & Lissmann, H. W. (1940). Ambulatory Reflexes in Spinal Amphibians. *Journal of Experimental  
814 Biology*, 17(2), 237–251. <https://doi.org/10.1242/jeb.17.2.237>

815 Gray, J., Lissmann, H. W., & Pumphrey, R. J. (1938). The Mechanism of Locomotion in the Leech (*Hirudo  
816 Medicinalis* Ray). *Journal of Experimental Biology*, 15(3), 408–430.  
817 <https://doi.org/10.1242/jeb.15.3.408>

818 Gray, J., & Sand, A. (1936). The Locomotory Rhythm of the Dogfish (*Scyllium Canicula*). *Journal of  
819 Experimental Biology*, 13(2), 200–209. <https://doi.org/10.1242/jeb.13.2.200>

820 Grillner, S. (1985). Neural control of vertebrate locomotion – central mechanisms and reflex interaction  
821 with special reference to the cat. In W. J. P. Barnes & M. H. Gladden (Eds.), *Feedback and motor*

- 822 control in invertebrates and vertebrates (pp. 35–56). Croom Helm.
- 823 Grillner, S., Degliana, T., Ekeberg, Ö., El Marina, A., Lansner, A., Orlovsky, G. N., & Wallén, P. (1995). Neural  
824 networks that co-ordinate locomotion and body orientation in lamprey. *Trends in Neuroscience*,  
825 18(6), 270–279.
- 826 Grillner, S., & El Manira, A. (2019). Current Principles of Motor Control, with Special Reference to Vertebrate  
827 Locomotion. *Physiological Reviews*, 100(1), 271–320.  
828 <https://doi.org/10.1152/physrev.00015.2019>
- 829 Grillner, S., McClellan, A., & Perret, C. (1981). Entrainment of the spinal pattern generators for swimming by  
830 mechano-sensitive elements in the lamprey spinal cord in vitro. *Brain Research*, 217(2), 380–386.  
831 [https://doi.org/10.1016/0006-8993\(81\)90015-9](https://doi.org/10.1016/0006-8993(81)90015-9)
- 832 Grillner, S., & Wallén, P. (1982). On Peripheral Control Mechanisms Acting on the Central Pattern Generators  
833 for Swimming in the Dogfish. *Journal of Experimental Biology*, 98(1), 1–22.  
834 <https://doi.org/10.1242/jeb.98.1.1>
- 835 Grillner, S., & Wallén, P. (1984). How Does The Lamprey Central Nervous System Make The Lamprey Swim?  
836 *Journal of Experimental Biology*, 112(1), 337–357. <https://doi.org/10.1242/jeb.112.1.337>
- 837 Grillner, S., & Wallen, P. (1985). Central pattern generators for locomotion, with special reference to  
838 vertebrates. *Annual Review of Neuroscience*, 8(1), 233–261.
- 839 Grillner, S., Wallén, P., Saitoh, K., Kozlov, A., & Robertson, B. (2008). Neural bases of goal-directed  
840 locomotion in vertebrates—An overview. *Brain Res. Rev.*, 57(1), 2–12.  
841 <https://doi.org/10.1016/j.brainresrev.2007.06.027>
- 842 Hamlet, C., Fauci, L., Morgan, J. R., & Tytell, E. D. (2023). Proprioceptive feedback amplification restores  
843 effective locomotion in a neuromechanical model of lampreys with spinal injuries. Proceedings of  
844 the National Academy of Sciences of the United States of America, 120(11), e2213302120.  
845 <https://doi.org/10.1073/pnas.2213302120>
- 846 Haimson, B., Hadas, Y., Bernat, N., Kania, A., Daley, M. A., Cinnamon, Y., Lev-Tov, A., & Klar, A. (2021). Spinal  
847 lumbar dl2 interneurons contribute to stability of bipedal stepping. *ELife*, 10, e62001.  
848 <https://doi.org/10.7554/eLife.62001>
- 849 Hellgren, J., Grillner, S., & Lansner, A. (1992). Computer simulation of the segmental neural network  
850 generating locomotion in lamprey by using populations of network interneurons. *Biological*  
851 *Cybernetics*, 68, 1–13.
- 852 Hiebert, G. W., Gorassini, M. A., Jiang, W., Prochazka, A., & Pearson, K. G. (1994). Corrective responses to  
853 loss of ground support during walking. II. Comparison of intact and chronic spinal cats. *Journal of*  
854 *Neurophysiology*, 71(2), 611–622. <https://doi.org/10.1152/jn.1994.71.2.611>
- 855 Hiebert, G. W., & Pearson, K. G. (1999). Contribution of sensory feedback to the generation of extensor  
856 activity during walking in the decerebrate cat. *Journal of Neurophysiology*, 81(2), 758–770.
- 857 Ho, S., & O'Donovan, M. J. (1993). Regionalization and intersegmental coordination of rhythm-generating  
858 networks in the spinal cord of the chick embryo. *Journal of Neuroscience*, 13(4), 1354–1371.  
859 <https://doi.org/10.1523/JNEUROSCI.13-04-01354.1993>
- 860 Holmes, P., Full, R. J., Koditschek, D., & Guckenheimer, J. (2006). The Dynamics of Legged Locomotion:  
861 Models, Analyses, and Challenges. *SIAM Review*, 48(2), 207–304.
- 862 Hughes, G.M., 1952. The Co-Ordination of insect movements: I The walking movements of insects. *Journal*  
863 *of Experimental Biology*, 29(2), pp.267-285.
- 864 Hughes, G.M., 1957. The co-ordination of insect movements: 11. the effect of limb amputation and the  
865 cutting of commissures in the cockroach (*blatta oiuentalis*). *Journal of Experimental Biology*, 34(3),  
866 pp.306-333.
- 867 Ijspeert, A. J. (1996). *Modelling the Neural Controller of a Swimming Lamprey: A Comparison of Naturally and*

868            *Artificially Evolved Networks* [No 42]. Dept. of Artificial Intelligence, U. of Edinburgh.

869 Ijspeert, A. J. (2001). A connectionist central pattern generator for the aquatic and terrestrial gaits of a  
870 simulated salamander. *Biological Cybernetics*, 84(5), 331–348.  
871 <https://doi.org/10.1007/s004220000211>

872 Ijspeert, A. J. (2014). Biorobotics: Using robots to emulate and investigate agile locomotion. *Science*,  
873 346(6206), 196–203. <https://doi.org/10.1126/science.1254486>

874 Ijspeert, A. J. (2020). Amphibious and Sprawling Locomotion: From Biology to Robotics and Back. *Annual*  
875 *Review of Control, Robotics, and Autonomous Systems*, 3(1), 173–193.  
876 <https://doi.org/10.1146/annurev-control-091919-095731>

877 Ijspeert, A. J., & Crespi, A. (2007). Online trajectory generation in an amphibious snake robot using a  
878 lamprey-like central pattern generator model. *Proceedings - IEEE International Conference on*  
879 *Robotics and Automation*, 262–268. <https://doi.org/10.1109/ROBOT.2007.363797>

880 Ijspeert, A. J., Crespi, A., Ryczko, D., & Cabelguen, J.-M. (2007). From swimming to walking with a  
881 salamander robot driven by a spinal cord model. *Science*, 315(5817), 1416–1420.  
882 <https://doi.org/10.1126/science.1138353>

883 Ijspeert, A. J., Hallam, J., & Willshaw, D. (1999). Evolving swimming controllers for a simulated lamprey with  
884 inspiration from neurobiology. *Adaptive Behavior*, 7(2), 151–172.  
885 <https://doi.org/10.1177/105971239900700202>

886 Ito, M. (2008). Control of mental activities by internal models in the cerebellum. *Nature Reviews*  
887 *Neuroscience*, 9(4), Article 4. <https://doi.org/10.1038/nrn2332>

888 Jacobson, R. D., & Hollyday, M. (1982). Electrically evoked walking and fictive locomotion in the chick.  
889 *Journal of Neurophysiology*, 48(1), 257–270.

890 Jindrich, D. L., & Full, R. J. (2002). Dynamic stabilization of rapid hexapedal locomotion. *Journal of*  
891 *Experimental Biology*, 205(18), 2803–2823. <https://doi.org/10.1242/jeb.205.18.2803>

892 Karakasiliotis, K., Schilling, N., Cabelguen, J.-M., & Ijspeert, A. J. (2013). Where are we in understanding  
893 salamander locomotion: Biological and robotic perspectives on kinematics. *Biological Cybernetics*,  
894 107(5), 529–544. <https://doi.org/10.1007/s00422-012-0540-4>

895 Karakasiliotis, K., Thandiackal, R., Melo, K., Horvat, T., Mahabadi, N. K., Tsitkov, S., Cabelguen, J. M., &  
896 Ijspeert, A. J. (2016). From cineradiography to biorobots: An approach for designing robots to  
897 emulate and study animal locomotion. *Journal of the Royal Society Interface*, 13(119).  
898 <https://doi.org/10.1098/rsif.2015.1089>

899 Kiehn, O. (2011). Development and functional organization of spinal locomotor circuits. *Current Opinion in*  
900 *Neurobiology*, 21(1), 100–109. <https://doi.org/10.1016/j.conb.2010.09.004>

901 Kiehn, O. (2016). Decoding the organization of spinal circuits that control locomotion. *Nature Reviews*  
902 *Neuroscience*, 17(4), 224–238. <https://doi.org/10.1038/nrn.2016.9>

903 Knüsel, J., Crespi, A., Cabelguen, J.-M., Ijspeert, A. J., & Ryczko, D. (2020). Reproducing Five Motor Behaviors  
904 in a Salamander Robot With Virtual Muscles and a Distributed CPG Controller Regulated by Drive  
905 Signals and Proprioceptive Feedback. *Frontiers in Neurorobotics*, 14. Scopus.  
906 <https://doi.org/10.3389/fnbot.2020.604426>

907 Kopell, N., & Ermentrout, G. B. (1988). Coupled oscillators and the design of central pattern generators.  
908 *Mathematical Biosciences*, 90(1), 87–109. [https://doi.org/10.1016/0025-5564\(88\)90059-4](https://doi.org/10.1016/0025-5564(88)90059-4)

909 Kopell, N., Ermentrout, G. B., & Williams, T. L. (1991). On chains of oscillators forced at one end. *SIAM,*  
910 *Journal of Applied Mathematics*, 51(5), 1397–1417.

911 Kriellaars, D. J., Brownstone, R. M., Noga, B. R., & Jordan, L. M. (1994). Mechanical entrainment of fictive  
912 locomotion in the decerebrate cat. *Journal of Neurophysiology*, 71(6), 2074–2086.  
913 <https://doi.org/10.1152/jn.1994.71.6.2074>



- 914 Kuo, A. D. (2002). The Relative Roles of Feedforward and Feedback in the Control of Rhythmic Movements.  
915 *Motor Control*, 6(2), 129–145. <https://doi.org/10.1123/mcj.6.2.129>
- 916 Lewis, K. E., & Eisen, J. S. (2003). From cells to circuits: Development of the zebrafish spinal cord. *Progress*  
917 *in Neurobiology*, 69(6), 419–449. [https://doi.org/10.1016/S0301-0082\(03\)00052-2](https://doi.org/10.1016/S0301-0082(03)00052-2)
- 918 Loeb, G. E. (2001). Learning from the spinal cord. *The Journal of Physiology*, 533(1), 111–117.
- 919 Loeb, G. E., Brown, I. E., & Cheng, E. J. (1999). A hierarchical foundation for models of sensorimotor control.  
920 *Experimental Brain Research*, 126(1), 1–18. <https://doi.org/10.1007/s002210050712>
- 921 Manoonpong, P., Pasemann, F., & Roth, H. (2007). Modular reactive neurocontrol for biologically inspired  
922 walking machines. *International Journal of Robotics Research*, 26(3), 301–331.
- 923 Mantziaris, Charalampos, Till Bockemühl, and Ansgar Büschges. "Central pattern generating networks in  
924 insect locomotion." *Developmental neurobiology* 80, no. 1-2 (2020): 16-30.
- 925 Markin, S. N., Klishko, A. N., Shevtsova, N. A., Lemay, M. A., Prilutsky, B. I., & Rybak, I. A. (2016). A  
926 Neuromechanical Model of Spinal Control of Locomotion. In B. I. Prilutsky & D. H. Edwards (Eds.),  
927 *Neuromechanical Modeling of Posture and Locomotion* (pp. 21–65). Springer New York.  
928 [https://doi.org/10.1007/978-1-4939-3267-2\\_2](https://doi.org/10.1007/978-1-4939-3267-2_2)
- 929 McLean, D. L., & Dougherty, K. J. (2015). Peeling back the layers of locomotor control in the spinal cord.  
930 *Current Opinion in Neurobiology*, 33, 63–70. <https://doi.org/10.1016/j.conb.2015.03.001>
- 931 McVea, D. A., Taylor, A. J., & Pearson, K. G. (2009). Long-Lasting Working Memories of Obstacles  
932 Established by Foreleg Stepping in Walking Cats Require Area 5 of the Posterior Parietal Cortex.  
933 *Journal of Neuroscience*, 29(29), 9396–9404. <https://doi.org/10.1523/JNEUROSCI.0746-09.2009>
- 934 Minassian, K., Hofstoetter, U. S., Dzeladini, F., Guertin, P. A., & Ijspeert, A. (2017). The Human Central Pattern  
935 Generator for Locomotion: Does It Exist and Contribute to Walking? *Neuroscientist*, 23(6), 649–663.  
936 <https://doi.org/10.1177/1073858417699790>
- 937 Mohamed Thangal, S. N., & Donelan, J. M. (2020). Scaling of inertial delays in terrestrial mammals. *PLoS*  
938 *One*, 15(2), e0217188.
- 939 More, H. L., & Donelan, J. M. (2018). Scaling of sensorimotor delays in terrestrial mammals. *Proceedings of*  
940 *the Royal Society B: Biological Sciences*, 285(1885), 20180613.  
941 <https://doi.org/10.1098/rspb.2018.0613>
- 942 More, H. L., Hutchinson, J. R., Collins, D. F., Weber, D. J., Aung, S. K. H., & Donelan, J. M. (2010). Scaling of  
943 sensorimotor control in terrestrial mammals. *Proceedings of the Royal Society B: Biological*  
944 *Sciences*, 277(1700), 3563–3568. <https://doi.org/10.1098/rspb.2010.0898>
- 945 More, H. L., O'Connor, S. M., Brøndum, E., Wang, T., Bertelsen, M. F., Grøndahl, C., Kastberg, K., Hørlyck, A.,  
946 Funder, J., & Donelan, J. M. (2013). Sensorimotor responsiveness and resolution in the giraffe.  
947 *Journal of Experimental Biology*, 216(6), 1003–1011.
- 948 Moritz, C. T., & Farley, C. T. (2004). Passive dynamics change leg mechanics for an unexpected surface  
949 during human hopping. *Journal of Applied Physiology*, 97(4), 1313–1322.
- 950 Muir, G. D., & Chu, T. K. (2002). Posthatching locomotor experience alters locomotor development in chicks.  
951 *Journal of Neurophysiology*, 88(1), 117–123.
- 952 Muir, G. D., Gosline, J. M., & Steeves, J. D. (1996). Ontogeny of bipedal locomotion: Walking and running in  
953 the chick. *The Journal of Physiology*, 493(2), 589–601.
- 954 Nakahira, Y., Liu, Q., Sejnowski, T. J., & Doyle, J. C. (2021). Diversity-enabled sweet spots in layered  
955 architectures and speed–accuracy trade-offs in sensorimotor control. *Proceedings of the National*  
956 *Academy of Sciences*, 118(22), e1916367118. <https://doi.org/10.1073/pnas.1916367118>
- 957 Neveln, I. D., Tirumalai, A., & Sponberg, S. (2019). Information-based centralization of locomotion in animals  
958 and robots. *Nature Communications*, 10(1), 1–11. <https://doi.org/10.1038/s41467-019-11613-y>

- 959 Nishikawa, K., Biewener, A. A., Aerts, P., Ahn, A. N., Chiel, H. J., Daley, M. A., Daniel, T. L., Full, R. J., Hale, M.  
960 E., Hedrick, T. L., Lappin, A. K., Nichols, T. R., Quinn, R. D., Satterlie, R. A., & Szymik, B. (2007).  
961 Neuromechanics: An integrative approach for understanding motor control. *Integrative and*  
962 *Comparative Biology*, 47(1), 16–54. <https://doi.org/10.1093/icb/icm024>
- 963 Nirody, J.A., 2023. Flexible locomotion in complex environments: the influence of species, speed and  
964 sensory feedback on panarthropod inter-leg coordination. *Journal of Experimental Biology*,  
965 226(Suppl\_1), p.jeb245111.
- 966 Ong, C. F., Geijtenbeek, T., Hicks, J. L., & Delp, S. L. (2019). Predicting gait adaptations due to ankle  
967 plantarflexor muscle weakness and contracture using physics-based musculoskeletal simulations.  
968 *PLoS Computational Biology*, 15(10), e1006993. <https://doi.org/10.1371/journal.pcbi.1006993>
- 969 Owaki, D., & Ishiguro, A. (2017). A Quadruped Robot Exhibiting Spontaneous Gait Transitions from Walking  
970 to Trotting to Galloping. *Scientific Reports*, 7(1), Article 1. [https://doi.org/10.1038/s41598-017-](https://doi.org/10.1038/s41598-017-00348-9)  
971 00348-9
- 972 Owaki, D., Kano, T., Nagasawa, K., Tero, A., & Ishiguro, A. (2013). Simple robot suggests physical interlimb  
973 communication is essential for quadruped walking. *Journal of The Royal Society Interface*, 10(78),  
974 20120669. <https://doi.org/10.1098/rsif.2012.0669>
- 975 Pearson, K. G. (1995). Proprioceptive regulation of locomotion. *Current Opinion in Neurobiology*, 5, 786–  
976 791.
- 977 Pearson, K. G. (2008). Role of sensory feedback in the control of stance duration in walking cats. *Brain*  
978 *Research Reviews*, 57(1), 222–227. <https://doi.org/10.1016/j.brainresrev.2007.06.014>
- 979 Pearson, K. G., Ekeberg, Ö., & Büschges, A. (2006). Assessing sensory function in locomotor systems using  
980 neuro-mechanical simulations. *Trends in Neurosciences*, 29(11), 625–631.  
981 <https://doi.org/10.1016/j.tins.2006.08.007>
- 982 Pearson, K. G., & Gramlich, R. (2010). Updating neural representations of objects during walking. *Annals of*  
983 *the New York Academy of Sciences*, 1198(1), 1–9. [https://doi.org/10.1111/j.1749-](https://doi.org/10.1111/j.1749-6632.2009.05422.x)  
984 6632.2009.05422.x
- 985 Pearson, K. G., Misiaszek, J. E., & Fouad, K. (1998). Enhancement and Resetting of Locomotor Activity by  
986 Muscle Afferents. *Annals of the New York Academy of Sciences*, 860(1), 203–215.  
987 <https://doi.org/10.1111/j.1749-6632.1998.tb09050.x>
- 988 Pratt, J., Chew, C. M., Torres, A., Dilworth, P., & Pratt, G. (2001). Virtual Model Control: An intuitive approach  
989 for bipedal locomotion. *The International Journal of Robotics Research*, 20(2), 129–143.
- 990 Ramadan, R., Geyer, H., Jeka, J., Schöner, G., & Reimann, H. (2022). A neuromuscular model of human  
991 locomotion combines spinal reflex circuits with voluntary movements. *Scientific Reports*, 12(1),  
992 Article 1. <https://doi.org/10.1038/s41598-022-11102-1>
- 993 Ramdya, P., & Ijspeert, A. J. (2023). The neuromechanics of animal locomotion: From biology to robotics  
994 and back. *Science Robotics*, 8(78), eadg0279. <https://doi.org/10.1126/scirobotics.adg0279>
- 995 Revzen, S., Koditschek, D. E., & Full, R. J. (2009). Towards Testable Neuromechanical Control Architectures  
996 for Running. In D. Sternad (Ed.), *Progress in Motor Control: A Multidisciplinary Perspective* (pp. 25–55).  
997 Springer US. [https://doi.org/10.1007/978-0-387-77064-2\\_3](https://doi.org/10.1007/978-0-387-77064-2_3)
- 998 Roberts, T. J., Eng, C. M., Sleboda, D. A., Holt, N. C., Brainerd, E. L., Stover, K. K., Marsh, R. L., & Azizi, E.  
999 (2019). The Multi-Scale, Three-Dimensional Nature of Skeletal Muscle Contraction. *Physiology*,  
1000 34(6), 402–408. <https://doi.org/10.1152/physiol.00023.2019>
- 1001 Roberts, T. J., Kram, R., Weyand, P. G., & Taylor, C. R. (1998). Energetics of bipedal running. I. Metabolic  
1002 cost of generating force. *The Journal of Experimental Biology*, 201(19), 2745–2751.
- 1003 Rossignol, S., Dubuc, R., & Gossard, J.-P. (2006). Dynamic Sensorimotor Interactions in Locomotion.  
1004 *Physiological Reviews*, 86(1), 89–154. <https://doi.org/10.1152/physrev.00028.2005>

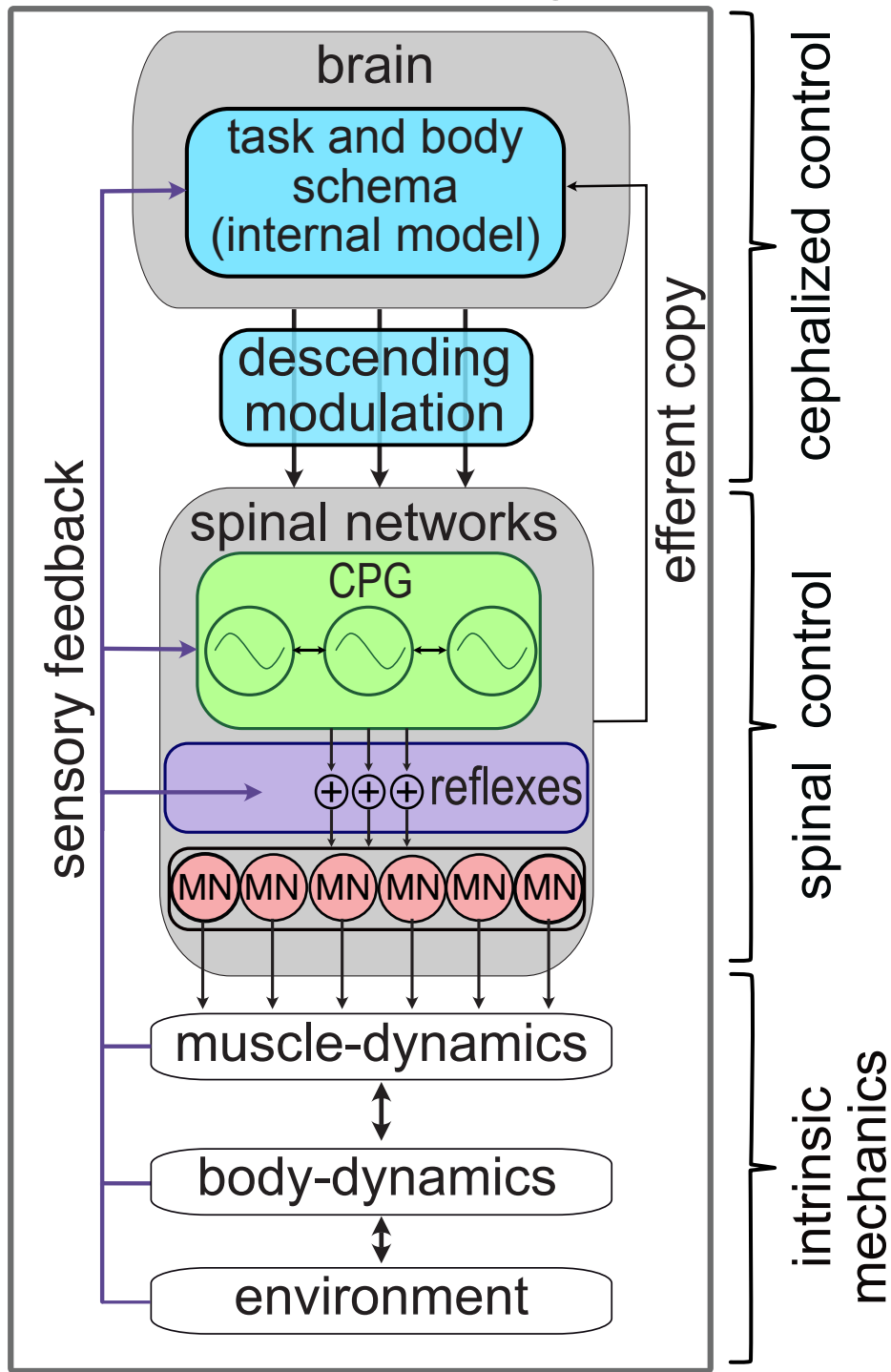
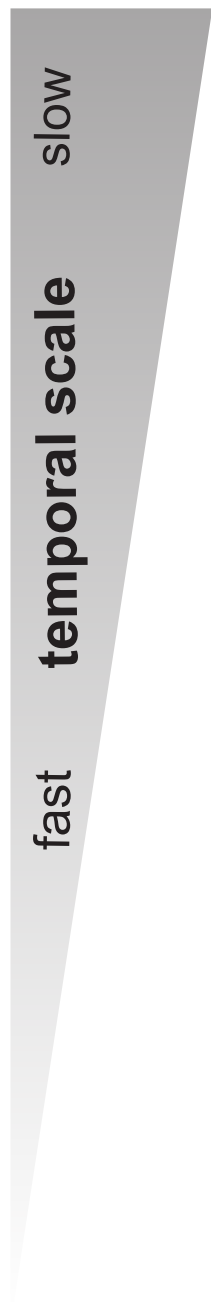
- 1005 Rubenson, J., Heliam, D. B., Lloyd, D. G., & Fournier, P. A. (2004). Gait selection in the ostrich: Mechanical  
1006 and metabolic characteristics of walking and running with and without an aerial phase. *Proceedings*  
1007 *of the Royal Society of London. Series B: Biological Sciences*, 271(1543), 1091–1099.  
1008 <https://doi.org/10.1098/rspb.2004.2702>
- 1009 Ruffolo, J. A., & McClellan, A. D. (2020). Modeling of lamprey reticulospinal neurons: Multiple distinct  
1010 parameter sets yield realistic simulations. *Journal of Neurophysiology*, 124(3), 895–913.  
1011 <https://doi.org/10.1152/jn.00070.2020>
- 1012 Russo, A. D., Stanev, D., Armand, S., & Ijspeert, A. (2021). Sensory modulation of gait characteristics in  
1013 human locomotion: A neuromusculoskeletal modeling study. *PLoS Computational Biology*, 17(5).  
1014 Scopus. <https://doi.org/10.1371/journal.pcbi.1008594>
- 1015 Rybak, I. A., Dougherty, K. J., & Shevtsova, N. A. (2015). Organization of the Mammalian Locomotor CPG:  
1016 Review of Computational Model and Circuit Architectures Based on Genetically Identified Spinal  
1017 Interneurons. *ENeuro*, 2(5). <https://doi.org/10.1523/ENEURO.0069-15.2015>
- 1018 Ryczko, D., Charrier, V., Ijspeert, A., & Cabelguen, J.-M. (2010). Segmental oscillators in axial motor circuits  
1019 of the salamander: Distribution and bursting mechanisms. *Journal of Neurophysiology*, 104(5),  
1020 2677–2692. <https://doi.org/10.1152/jn.00479.2010>
- 1021 Ryczko, D., Simon, A., & Ijspeert, A. J. (2020). Walking with Salamanders: From Molecules to Biorobotics.  
1022 *Trends in Neurosciences*, 43(11), 916–930. <https://doi.org/10.1016/j.tins.2020.08.006>
- 1023 Ryu, H. X., & Kuo, A. D. (2021). An optimality principle for locomotor central pattern generators. *Scientific*  
1024 *Reports*, 11(1), 13140. <https://doi.org/10.1038/s41598-021-91714-1>
- 1025 Ryu, Y. U., & Bradley, N. S. (2009). Precocious locomotor behavior begins in the egg: Development of leg  
1026 muscle patterns for stepping in the chick. *PLoS One*, 4(7), e6111.
- 1027 Schilling, M., Hoinville, T., Schmitz, J., & Cruse, H. (2013). Walknet, a bio-inspired controller for hexapod  
1028 walking. *Biological Cybernetics*, 107(4), 397–419. <https://doi.org/10.1007/s00422-013-0563-5>
- 1029 Sherrington, C. S. (1900). On the innervation of antagonistic muscles. Sixth note. *Proceedings of the Royal*  
1030 *Society of London*, 66(424–433), 66–67. <https://doi.org/10.1098/rspl.1899.0075>
- 1031 Sherrington, C. S. (1906). Observations on the scratch-reflex in the spinal dog. *The Journal of Physiology*,  
1032 34(1–2), 1–50. <https://doi.org/10.1113/jphysiol.1906.sp001139>
- 1033 Sherrington, C. S. (1910). REMARKS ON THE REFLEX MECHANISM OF THE STEP1. *Brain*, 33(1), 1–25.  
1034 <https://doi.org/10.1093/brain/33.1.1>
- 1035 Shik, M. L., Severin, F. V., & Orlovsky, G. N. (1966). Control of walking by means of electrical stimulation of  
1036 the mid-brain. *Biophysics*, 11, 756–765.
- 1037 Sholomenko, G. N., Funk, G. D., & Steeves, J. D. (1991). Locomotor activities in the decerebrate bird without  
1038 phasic afferent input. *Neuroscience*, 40(1), 257–266. [https://doi.org/10.1016/0306-4522\(91\)90188-T](https://doi.org/10.1016/0306-4522(91)90188-T)
- 1040 Sholomenko, G. N., & O'Donovan, M. J. (1995). Development and characterization of pathways descending  
1041 to the spinal cord in the embryonic chick. *Journal of Neurophysiology*, 73(3), 1223–1233.
- 1042 Sholomenko, G. N., & Steeves, J. D. (1987). Effects of selective spinal cord lesions on hind limb locomotion  
1043 in birds. *Experimental Neurology*, 95(2), 403–418. [https://doi.org/10.1016/0014-4886\(87\)90148-8](https://doi.org/10.1016/0014-4886(87)90148-8)
- 1044 Smith, N. C., Jespers, K. J., & Wilson, A. M. (2010). Ontogenetic scaling of locomotor kinetics and  
1045 kinematics of the ostrich (*Struthio camelus*). *Journal of Experimental Biology*, 213(8), 1347–1355.
- 1046 Song, S., & Geyer, H. (2015). A neural circuitry that emphasizes spinal feedback generates diverse  
1047 behaviours of human locomotion. *The Journal of Physiology*, 593(16), 3493–3511.  
1048 <https://doi.org/10.1113/JP270228>
- 1049 Song, S., & Geyer, H. (2018). Predictive neuromechanical simulations indicate why walking performance  
1050 declines with ageing. *The Journal of Physiology*, 596(7), 1199–1210.

- 1051 <https://doi.org/10.1113/JP275166>
- 1052 Steeves, J. D., Sholomenko, G. N., & Webster, D. M. S. (1987). Stimulation of the pontomedullary reticular  
1053 formation initiates locomotion in decerebrate birds. *Brain Research*, 401(2), 205–212.  
1054 [https://doi.org/10.1016/0006-8993\(87\)91406-5](https://doi.org/10.1016/0006-8993(87)91406-5)
- 1055 Suzuki, S., Kano, T., Ijspeert, A. J., & Ishiguro, A. (2021). Spontaneous Gait Transitions of Sprawling  
1056 Quadruped Locomotion by Sensory-Driven Body–Limb Coordination Mechanisms. *Frontiers in*  
1057 *Neurobotics*, 15, 92. <https://doi.org/10.3389/fnbot.2021.645731>
- 1058 Taga, G. (1995). A model of the neuro-musculo-skeletal system for human locomotion. *Biological*  
1059 *Cybernetics*, 73(2), 97–111. <https://doi.org/10.1007/BF00204048>
- 1060 Taga, G. (1998). A model of the neuro-musculo-skeletal system for anticipatory adjustment of human  
1061 locomotion during obstacle avoidance. *Biological Cybernetics*, 78(1), 9–17.
- 1062 Taga, G., Yamaguchi, Y., & Shimizu, H. (1991). Self-organized control of bipedal locomotion by neural  
1063 oscillators in unpredictable environment. *Biological Cybernetics*, 65(3), 147–159.
- 1064 Tata Ramalingasetty, S., Danner, S. M., Arreguit, J., Markin, S. N., Rodarie, D., Kathe, C., Courtine, G., Rybak,  
1065 I. A., & Ijspeert, A. J. (2021). A Whole-Body Musculoskeletal Model of the Mouse. *IEEE Access*, 9,  
1066 163861–163881. <https://doi.org/10.1109/ACCESS.2021.3133078>
- 1067 Ten Cate, J. (1960). Locomotor Movements in the Spinal Pigeon. *Journal of Experimental Biology*, 37(3),  
1068 609–613. <https://doi.org/10.1242/jeb.37.3.609>
- 1069 Ten Cate, J. (1964). Locomotory Movements of the Hind Limbs in Rabbits After Isolation of the  
1070 Lumbosacral Cord. *Journal of Experimental Biology*, 41(2), 359–362.  
1071 <https://doi.org/10.1242/jeb.41.2.359>
- 1072 Ten Cate, J. (1965). Automatic Activity of the Locomotor Centres of the Lumbar Cord in Lizards. *Journal of*  
1073 *Experimental Biology*, 43(1), 181–184. <https://doi.org/10.1242/jeb.43.1.181>
- 1074 Thandiackal, R., Melo, K., Paez, L., Herault, J., Kano, T., Akiyama, K., Boyer, F., Ryczko, D., Ishiguro, A., &  
1075 Ijspeert, A. J. (2021). Emergence of robust self-organized undulatory swimming based on local  
1076 hydrodynamic force sensing. *Science Robotics*, 6(57), eabf6354.  
1077 <https://doi.org/10.1126/scirobotics.abf6354>
- 1078 Ting, L. H., & McKay, J. L. (2007). Neuromechanics of muscle synergies for posture and movement. *Current*  
1079 *Opinion in Neurobiology*, 17(6), 622–628. <https://doi.org/10.1016/j.conb.2008.01.002>
- 1080 Todorov, E. (2004). Optimality principles in sensorimotor control. *Nature Neuroscience*, 7, 907–915.
- 1081 Todorov, E., & Jordan, M. I. (2002). Optimal feedback control as a theory of motor coordination. *Nature*  
1082 *Neuroscience*, 5(11), Article 11. <https://doi.org/10.1038/nn963>
- 1083 Traven, H., Brodin, L., Lansner, A., Ekeberg, Ö., Wallén, P., & Grillner, S. (1993). Computer simulations of  
1084 NMDA and non-NMDA receptors mediated synaptic drive: Sensory and supraspinal modulation of  
1085 neurons and small networks. *J. of Neurophysiology*, 70(2), 695–709.
- 1086 van den Brand, R., Heutschi, J., Barraud, Q., DiGiovanna, J., Bartholdi, K., Huerlimann, M., Friedli, L.,  
1087 Vollenweider, I., Moraud, E. M., Duis, S., Dominici, N., Micera, S., Musienko, P., & Courtine, G. (2012).  
1088 Restoring Voluntary Control of Locomotion after Paralyzing Spinal Cord Injury. *Science*, 336(6085),  
1089 1182–1185. <https://doi.org/10.1126/science.1217416>
- 1090 Van der Noot, N., Ijspeert, A. J., & Ronsse, R. (2018). Bio-inspired controller achieving forward speed  
1091 modulation with a 3D bipedal walker. *International Journal of Robotics Research*, 37(1), 168–196.  
1092 Scopus. <https://doi.org/10.1177/0278364917743320>
- 1093 Van der Noot, N., Ijspeert, A. J., & Ronsse, R. (2019). Neuromuscular model achieving speed control and  
1094 steering with a 3D bipedal walker. *Autonomous Robots*, 43(6), 1537–1554.  
1095 <https://doi.org/10.1007/s10514-018-9814-6>
- 1096 Wagner, F. B., Mignardot, J.-B., Le Goff-Mignardot, C. G., Demesmaeker, R., Komi, S., Capogrosso, M.,

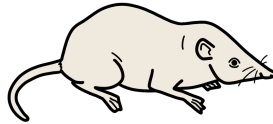
- 1097 Rowald, A., Seáñez, I., Caban, M., Pirondini, E., Vat, M., McCracken, L. A., Heimgartner, R., Fodor, I.,  
1098 Watrin, A., Seguin, P., Paoles, E., Van Den Keybus, K., Eberle, G., ... Courtine, G. (2018). Targeted  
1099 neurotechnology restores walking in humans with spinal cord injury. *Nature*, 563(7729), Article  
1100 7729. <https://doi.org/10.1038/s41586-018-0649-2>
- 1101 Wallén, P. (1982). Spinal mechanisms controlling locomotion in dogfish and lamprey. *Acta Physiologica*  
1102 *Scandinavica. Supplementum*, 503, 1–45.
- 1103 Wallén, P., Ekeberg, Ö., Lansner, A., Brodin, L., Traven, H., & Grillner, S. (1992). A computer-based model for  
1104 realistic simulations of neural networks II: The segmental network generating locomotor  
1105 rhythmicity in the lamprey. *J. of Neurophysiology*, 68, 1939–1950.
- 1106 Walter, W. G. (1950). An Electro-Mechanical « Animal ». *Dialectica*, 4(3), 206–213.
- 1107 Walter, W. G. (1951). A Machine That Learns. *Scientific American*, 185(2), 60–64.
- 1108 Wang, J. M., Hamner, S. R., Delp, S. L., & Koltun, V. (2012). Optimizing locomotion controllers using  
1109 biologically-based actuators and objectives. *ACM Transactions on Graphics*, 31(4), 25:1-25:11.  
1110 <https://doi.org/10.1145/2185520.2185521>
- 1111 Watson, R. R., Rubenson, J., Coder, L., Hoyt, D. F., Propert, M. W., & Marsh, R. L. (2011). Gait-specific  
1112 energetics contributes to economical walking and running in emus and ostriches. *Proceedings of*  
1113 *the Royal Society B: Biological Sciences*, 278(1714), 2040–2046.
- 1114 Webb, B. (2001). Can robots make good models of biological behaviour? *Behavioral and Brain Sciences*,  
1115 24(6).
- 1116 Webb, B. (2020). Robots with insect brains. *Science*, 368(6488), 244–245.  
1117 <https://doi.org/10.1126/science.aaz6869>
- 1118 Webster, D. M., & Steeves, J. D. (1988). Origins of brainstem-spinal projections in the duck and goose.  
1119 *Journal of Comparative Neurology*, 273(4), 573–583.
- 1120 Whelan, P. J. (1996). Control of Locomotion in the deacebrate cat. *Progress in Neurobiology*, 49, 481–515.
- 1121 Wiener, N. (2019). *Cybernetics or Control and Communication in the Animal and the Machine, Reissue of the*  
1122 *1961 second edition*. MIT Press.
- 1123 Williams, T. L. (1992a). Phase coupling by synaptic spread in chains of coupled neuronal oscillators.  
1124 *Science*, 258, 662–665.
- 1125 Williams, T. L. (1992b). Phase coupling in simulated chains of coupled neuronal oscillators representing  
1126 the lamprey spinal cord. *Neural Computation*, 4, 546–558.
- 1127 Williams, T. L., & McMillen, T. (2015). Strategies for swimming: Explorations of the behaviour of a neuro-  
1128 musculo-mechanical model of the lamprey. *Biology Open*, 4(3), 253–258.  
1129 <https://doi.org/10.1242/bio.20149621>
- 1130 Williams, T. L., Sigvardt, K. A., Kopell, N., Ermentrout, G. B., & Rempier, M. P. (1990). Forcing of coupled  
1131 nonlinear oscillators: Studies of intersegmental coordination in the lamprey locomotor central  
1132 pattern generator. *J. of Neurophysiology*, 64, 862–871.
- 1133 Wilson, D.M., 1965. Proprioceptive leg reflexes in cockroaches. *Journal of Experimental Biology*, 43(3),  
1134 pp.397-409.
- 1135 Wilson, D.M., 1967. Stepping patterns in tarantula spiders. *Journal of Experimental Biology*, 47(1), pp.133-  
1136 151.
- 1137 Wolpert, D. M., & Ghahramani, Z. (2000). Computational principles of movement neuroscience. *Nature*  
1138 *Neuroscience*, 3(11), 1212. <https://doi.org/10.1038/81497>
- 1139 Yanagihara, D., Udo, M., Kondo, I., & Yoshida, T. (1993). A new learning paradigm: Adaptive changes in  
1140 interlimb coordination during perturbed locomotion in decerebrate cats. *Neuroscience Research*,  
1141 18, 241–244. [https://doi.org/10.1016/0168-0102\(93\)90060-4](https://doi.org/10.1016/0168-0102(93)90060-4)

1142 Zhong, G., Shevtsova, N. A., Rybak, I. A., & Harris-Warrick, R. M. (2012). Neuronal activity in the isolated  
1143 mouse spinal cord during spontaneous deletions in fictive locomotion: Insights into locomotor  
1144 central pattern generator organization. *The Journal of Physiology*, 590(19), 4735–4759.

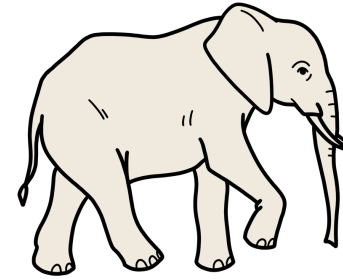
# Neuromechanical system



A



shrew

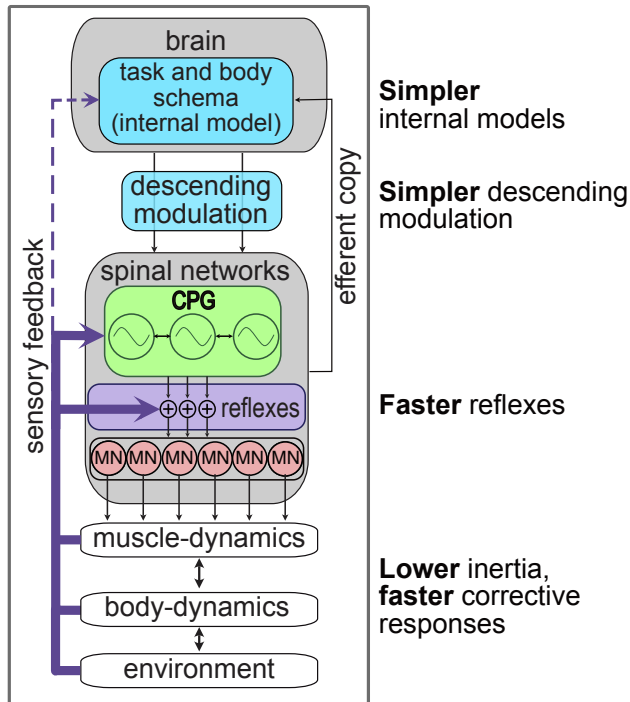


elephant

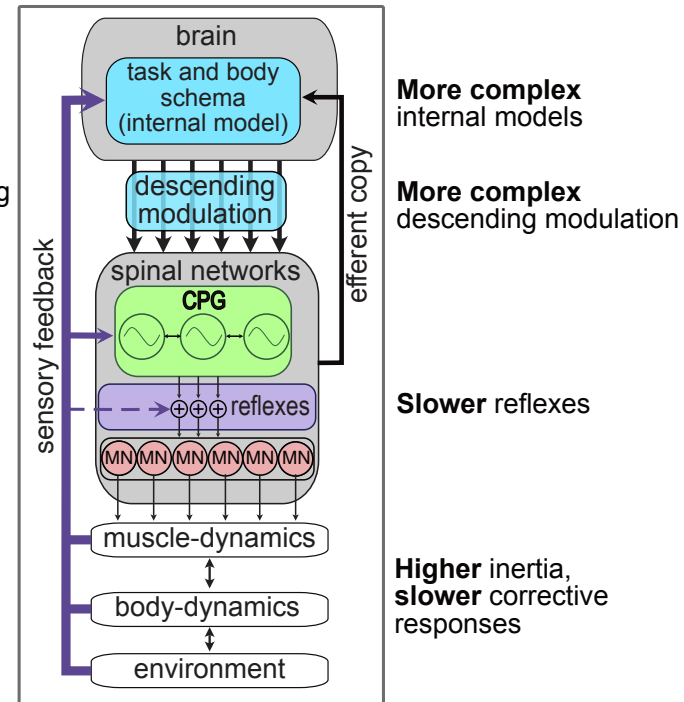
Delay estimates

synaptic	0.7 ms	0.7 ms
monosynaptic reflex	10 ms	180 ms
inertial delays	3–5 ms	200–400 ms
reflex:inertial	2–3.3	0.9–0.5
reflex:synaptic	14	260
reflex:stance duration	0.25	0.6

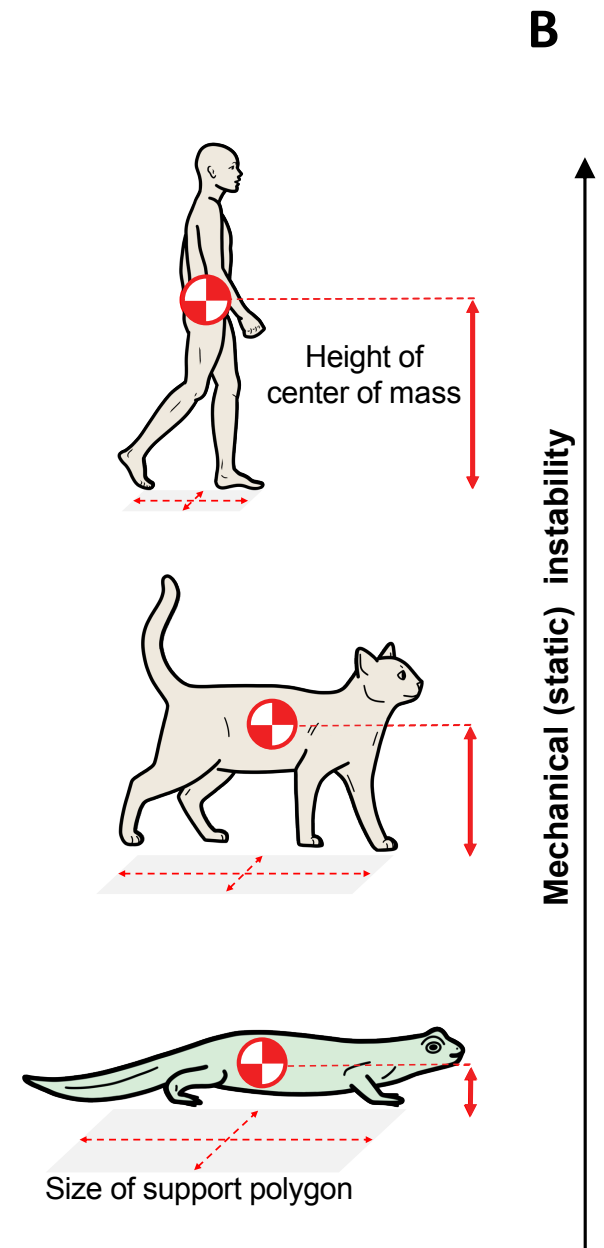
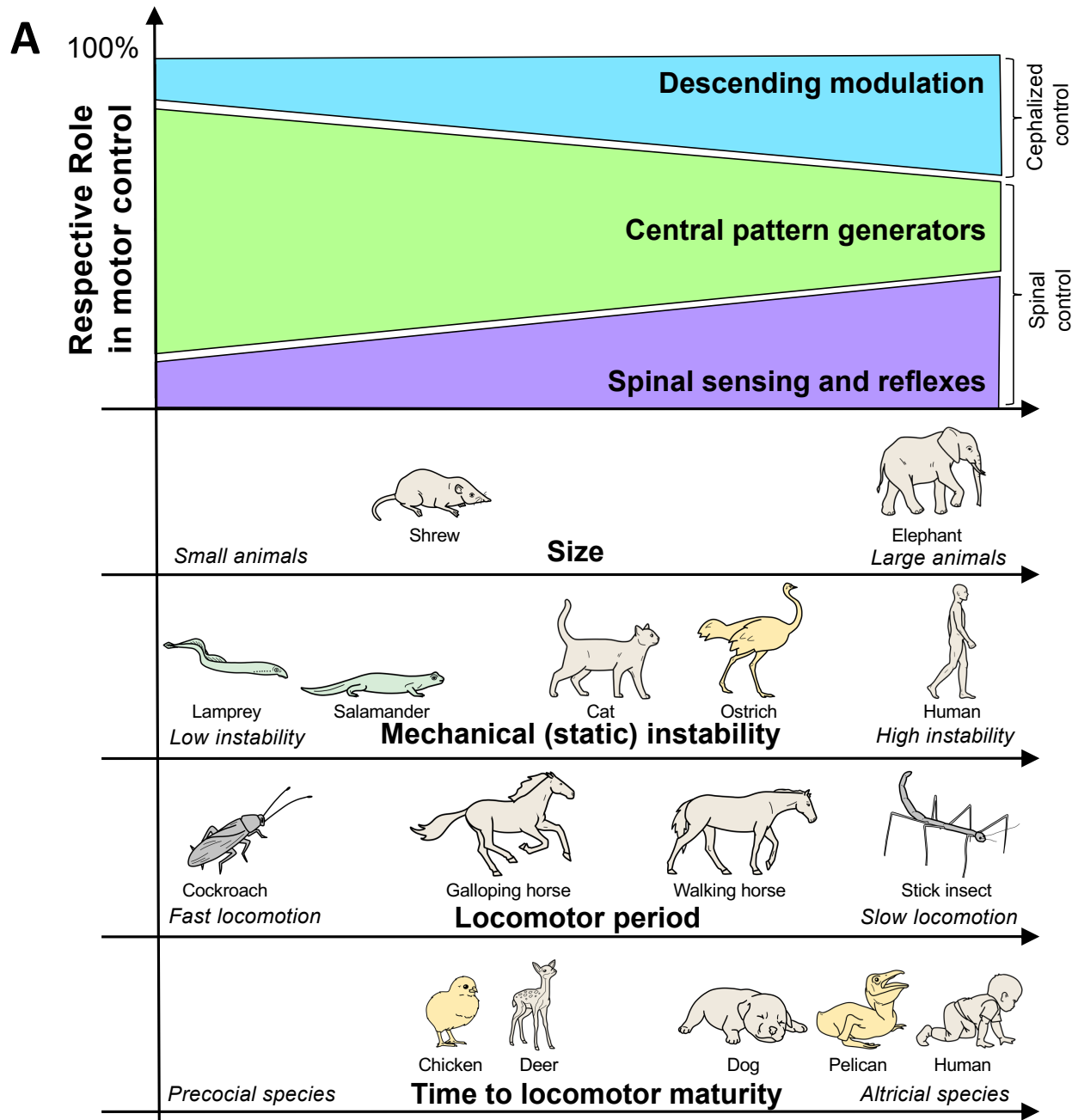
B Small animals



C Large animals







Proximal

Mainly CPG-driven  
Length and position feedback  
High inertia  
Low compliance  
Low 'preflexes'

Distal

Mainly sensory-driven  
Force and velocity feedback  
Low inertia  
High compliance  
High 'preflexes'

