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Metagenomes from Experimental Hydrologic Manipulation of Restored Coastal Plain Wetland Soils (Tyrell County, North Carolina)

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ABSTRACT Hydrologic changes modify microbial community structure and ecosystem functions, especially in wetland systems. Here, we present 24 metagenomes from a coastal freshwater wetland experiment in which we manipulated hydrologic conditions and plant presence. These wetland soil metagenomes will deepen our understanding of how hydrology and vegetation influence microbial functional diversity.

icrobial community structure, soil physicochemical properties, and the abundance and composition of vegetation interact to influence biogeochemical functions (1). Changes in wetland hydrology due to drought, draining, and rewetting cause shifts in soil redox potential, microbial community composition, and associated ecosystem processes such as greenhouse gas (GHG) production (2-6). Vegetation also affects microbial processes by facilitating the transport of oxygen into the root zone and the transport of methane from the root zone into the atmosphere (7, 8). Here, we present wetland soil metagenomes from a mesocosm experiment in which we manipulated hydrology and plant presence to examine microbial community responses. These data complement targeted amplicon sequencing data, GHG fluxes, and soil physicochemical properties from the wetland mesocosm experiment (9).

We collected wetland soils from a restored coastal freshwater wetland at the Timberlake Observatory for Wetland Restoration on the Albemarle Peninsula in Tyrell County, North Carolina (latitude, 35.8959; longitude, -76.1658). We collected soils from three locations with different water table levels (-20 cm, -10 cm, and 0 cm) (10). We altered redox conditions by manipulating hydrology over 8 weeks. The experimental design and sampling details were published by Bledsoe and Peralta (9). We collected and combined six soil cores (3-cm diameter, 10-cm depth) from plant or no-plant areas. We completed 16S rRNA amplicon sequencing (NCBI BioProject PRJNA636184), GHG flux measurements, and soil environmental analyses (9). We sequenced metagenomes that represented the most distinct microbial communities based on amplicon sequencing, and we chose the following samples: wetland soils sampled from field sites at which the water table measured -20 cm and 0 cm, to capture "mesocosm baseline" (n=8) functional diversity, and a subset of samples at the end of the 8-week hydrologic manipulation (i.e., prolonged drying or wetting only) in the presence or absence of vegetation (n = 16) (9).

We used the Qiagen DNeasy PowerMax soil kit to extract genomic DNA from freeze-dried soils. Purified DNA products were sent to the U.S. Department of Energy (DOE) Joint Genome Institute (JGI) for sequencing and analysis. Metagenomes were sequenced at the DOE JGI (GitHub [see SupplementalTableS1_MetagenomeSummary_ Peralta_et_al.csv at https://doi.org/10.5281/zenodo.4042110]), and project information

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Peralta et al.

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TABLE 1 Annotation and gene-calling summary for the 24 metagenomes

Gene type and database used ^a	No. (mean ± SD)
CDSs	992,951 ± 305,299
rRNAs	775 ± 173
Other genes	$4,878 \pm 2,340$
CDSs with COGs	$633,925 \pm 194,834$
CDSs with SMART	$127,732 \pm 38,536$
CDSs with SUPFAM	641,238 ± 189,914
CDSs with CATH FunFam	$509,413 \pm 154,168$
CDSs with Pfam	603,115 ± 187,921

 $[^]a$ CDSs, coding sequences; COGs, Clusters of Orthologous Groups; SUPFAM, superfamily; FunFam, functional families.

can be accessed under GOLD (11) study project accession number Gs0142547 and NCBI BioProject accession number PRJNA641216. Plate-based DNA library preparation for Illumina sequencing was performed according to published protocols in GitHub (see Supplemental_Methods_Details_MRA_WetlandSoilMetagenomes.txt at https://doi.org/10.5281/zenodo.4042110). The sequencing project resulted in 583.2 Gbp of raw sequence data. The average read length for each metagenome is found in GitHub (see SupplementalTableS1_MetagenomeSummary_Peralta_et_al.csv at https://doi.org/10.5281/zenodo.4042110). These data were processed using the DOE JGI Metagenome Annotation Pipeline using IMG/M v.5.0.9 (12–14). Initial sequence quality control details can be found in GitHub (see Supplemental_Methods_Details_MRA_Wetland SoilMetagenomes.txt at https://doi.org/10.5281/zenodo.4042110). Illumina reads were quality control filtered according to the protocol described in GitHub (see Supplemental_Methods_Details_MRA_WetlandSoilMetagenomes.txt at https://doi.org/10.5281/zenodo.4042110).

Annotation and gene calling resulted in $600,507 \pm 172,049$ annotated contigs per sample (mean \pm standard deviation [SD]), with a mean of 998,604 gene features (SD, 307,431 gene features) identified in each (Table 1). Based on phylogenetic associations (determined on the basis of bidirectional best hits to genes in other genomes), the estimated alpha diversity across all metagenomes is $10,365 \pm 837$ operational taxonomic units (OTUs) (mean \pm SD) (GitHub [see SupplementalTableS1_Metagenome Summary_Peralta_et_al.csv at https://doi.org/10.5281/zenodo.4042110]).

Data availability. Metagenomes were sequenced at the DOE JGI, and the study information can be found under GOLD study project accession number Gs0142547 and NCBI BioProject accession number PRJNA641216. Additional sample-specific metagenomestatistics can be found in GitHub (see Supplemental Table S1_Metagenome Summary_Peralta_et_al.csv at https://doi.org/10.5281/zenodo.4042110). Details on metagenomic library preparation and sequence filtering can be found in GitHub (see Supplemental_Methods_Details_MRA_Wetland Soil Metagenomes.txt at https://doi.org/10.5281/zenodo.4042110).

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REFERENCES

- Jansson JK, Hofmockel KS. 2020. Soil microbiomes and climate change. 1.
 Nat Rev Microbiol 18:35–46. https://doi.org/10.1038/s41579-019-0265-7.
- 2. Peralta AL, Ludmer S, Matthews JW, Kent AD. 2014. Bacterial community

response to changes in soil redox potential along a moisture gradient in restored wetlands. Ecol Eng 73:246–253. https://doi.org/10.1016/j.ecoleng.2014.09.047.

Volume 9 lssue 41 e00882-20 mra.asm.org **2**



- 3. Lazcano C, Deol AS, Brummell ME, Strack M. 2020. Interactive effects of vegetation and water table depth on belowground C and N mobilization and greenhouse gas emissions in a restored peatland. Plant Soil 448: 299–313. https://doi.org/10.1007/s11104-020-04434-2.
- Bansal S, Johnson OF, Meier J, Zhu X. 2020. Vegetation affects timing and location of wetland methane emissions. J Geophys Res Biogeosci 125: e2020JG005777.
- Ou Y, Rousseau AN, Wang L, Yan B, Gumiere T, Zhu H. 2019. Identification
 of the alteration of riparian wetland on soil properties, enzyme activities and
 microbial communities following extreme flooding. Geoderma 337:
 825–833. https://doi.org/10.1016/j.geoderma.2018.10.032.
- Peralta AL, Ludmer S, Kent AD. 2013. Hydrologic history influences microbial community composition and nitrogen cycling under experimental drying/wetting treatments. Soil Biol Biochem 66:29–37. https:// doi.org/10.1016/j.soilbio.2013.06.019.
- Chanton JP. 2005. The effect of gas transport on the isotope signature of methane in wetlands. Org Geochem 36:753–768. https://doi.org/10 .1016/j.orggeochem.2004.10.007.
- Carmichael MJ, Bernhardt ES, Bräuer SL, Smith WK. 2014. The role of vegetation in methane flux to the atmosphere: should vegetation be included as a distinct category in the global methane budget? Biogeochemistry 119:1–24. https://doi.org/10.1007/s10533-014-9974-1.
- Bledsoe RB, Peralta AL. 2020. Dynamic hydrology and plant-mediated effects reduce greenhouse gas emissions and alter wetland microbial communities. bioRxiv https://doi.org/10.1101/2020.06.29.178533.

- Ardón M, Morse JL, Doyle MW, Bernhardt ES. 2010. The water quality consequences of restoring wetland hydrology to a large agricultural watershed in the southeastern coastal plain. Ecosystems 13:1060–1078. https://doi.org/10.1007/s10021-010-9374-x.
- Mukherjee S, Stamatis D, Bertsch J, Ovchinnikova G, Katta HY, Mojica A, Chen I-MA, Kyrpides NC, Reddy TBK. 2019. Genomes Online Database (GOLD) v.7: updates and new features. Nucleic Acids Res 47:D649 –D659. https://doi.org/10.1093/nar/gky977.
- Markowitz VM, Chen I-MA, Chu K, Szeto E, Palaniappan K, Pillay M, Ratner A, Huang J, Pagani I, Tringe S, Huntemann M, Billis K, Varghese N, Tennessen K, Mavromatis K, Pati A, Ivanova NN, Kyrpides NC. 2014. IMG/M 4 version of the integrated metagenome comparative analysis system. Nucleic Acids Res 42:D568–D573. https://doi.org/10.1093/nar/gkt919.
- Huntemann M, Ivanova NN, Mavromatis K, Tripp HJ, Paez-Espino D, Tennessen K, Palaniappan K, Szeto E, Pillay M, Chen I-MA, Pati A, Nielsen T, Markowitz VM, Kyrpides NC. 2016. The standard operating procedure of the DOE-JGI Metagenome Annotation Pipeline (MAP v.4). Stand Genomic Sci 11:17. https://doi.org/10.1186/s40793-016-0138-x.
- Chen I-MA, Markowitz VM, Chu K, Palaniappan K, Szeto E, Pillay M, Ratner A, Huang J, Andersen E, Huntemann M, Varghese N, Hadjithomas M, Tennessen K, Nielsen T, Ivanova NN, Kyrpides NC. 2017. IMG/M: integrated genome and metagenome comparative data analysis system. Nucleic Acids Res 45:D507–D516. https://doi.org/10 .1093/nar/gkw929.

Volume 9 lssue 41 e00882-20 mra.asm.org **3**