

UCLA

UCLA Previously Published Works

Title

Pulmonary Fibrosis Stakeholder Summit: A Joint NHLBI, Three Lakes Foundation, and Pulmonary Fibrosis Foundation Workshop Report.

Permalink

<https://escholarship.org/uc/item/5092667m>

Journal

American Journal of Respiratory and Critical Care Medicine, 209(4)

Authors

Hariri, Lida

Hogaboam, Cory

Jenkins, R

et al.

Publication Date

2024-02-15

DOI

10.1164/rccm.202307-1154WS

Copyright Information

This work is made available under the terms of a Creative Commons Attribution-NonCommercial-NoDerivatives License, available at

<https://creativecommons.org/licenses/by-nc-nd/4.0/>

Peer reviewed

WORKSHOP REPORT

Pulmonary Fibrosis Stakeholder Summit

A Joint NHLBI, Three Lakes Foundation, and Pulmonary Fibrosis Foundation Workshop Report

6 Sydney B. Montesi¹, Christian R. Gomez³, Michael Beers⁴, Robert Brown⁵, Ishanu Chattopadhyay⁶, Kevin R. Flaherty⁷, Christine Kim Garcia¹⁰, Brigitte Gomperts¹¹, Lida P. Hariri^{1,2}, Cory M. Hogaboam¹⁴, R. Gisli Jenkins¹⁵, Naftali Kaminski¹⁶, Grace Hyun J. Kim^{12,13}, Melanie Königshoff¹⁷, Martin Kolb¹⁸, Darrell N. Kotton¹⁹, Jonathan A. Kropski²⁰, Joseph Lasky^{21,22}, Chelsea M. Magin^{23,24,25}, Toby M. Maher²⁷, Mark McCormick²¹, Bethany B. Moore⁸, Cheryl Nickerson-Nutter²⁸, Justin Oldham⁷, Anna J. Podolanczuk²⁹, Ganesh Raghu³⁰, Ivan Rosas³¹, Steven M. Rowe^{32,33}, William T. Schmidt²¹, David Schwartz²⁶, Jessica E. Shore²¹, Cathie Spino⁹, J. Matthew Craig³, and Fernando J. Martinez²⁹

Abstract

Despite progress in elucidation of disease mechanisms, identification of risk factors, biomarker discovery, and the approval of two medications to slow lung function decline in idiopathic pulmonary fibrosis and one medication to slow lung function decline in progressive pulmonary fibrosis, pulmonary fibrosis remains a disease with a high morbidity and mortality. In recognition of the need to catalyze ongoing advances and collaboration in the field of pulmonary fibrosis, the NHLBI, the Three Lakes Foundation, and the Pulmonary Fibrosis Foundation hosted the Pulmonary Fibrosis Stakeholder Summit on

November 8–9, 2022. This workshop was held virtually and was organized into three topic areas: 1) novel models and research tools to better study pulmonary fibrosis and uncover new therapies, 2) early disease risk factors and methods to improve diagnosis, and 3) innovative approaches toward clinical trial design for pulmonary fibrosis. In this workshop report, we summarize the content of the presentations and discussions, enumerating research opportunities for advancing our understanding of the pathogenesis, treatment, and outcomes of pulmonary fibrosis.

Keywords: interstitial lung disease, pulmonary fibrosis

In 2012, the NHLBI conducted a workshop to define areas of future research direction in idiopathic pulmonary fibrosis (IPF) (1). The subsequent decade saw major advances in the understanding of disease pathogenesis, identification of molecular mediators promoting fibrosis, elucidation of genetic risk factors, development of imaging-based biomarkers, defining the risk of combined immunosuppression in patients with IPF, and demonstration of the efficacy of nintedanib and pirfenidone for slowing lung function decline in patients with IPF and those with other forms of progressive interstitial lung disease (ILD). Despite this,

pulmonary fibrosis remains a disease with a high morbidity and mortality; currently approved therapies only slow the rate of disease progression (2–4). Significant work is required to ensure timely diagnosis of individuals with pulmonary fibrosis, identify treatments that will halt or reverse fibrosis, and improve the lives of individuals living with this disease.

On November 8–9, 2022, the NHLBI, the Three Lakes Foundation, and the Pulmonary Fibrosis Foundation convened the Pulmonary Fibrosis Stakeholder Summit. The goal of this summit was to identify scientific gaps and future basic and clinical

research directions related to pulmonary fibrosis. This virtual summit was divided into three sessions addressing topics related to 1) novel models and research tools to better study pulmonary fibrosis and uncover new therapeutic targets, 2) early disease risk factors and methods to improve diagnosis, and 3) innovative approaches to clinical trial design for pulmonary fibrosis. The topics for presentation were selected by the summit organizers, and efforts were made to have a broad representation of content and speakers. Here we summarize the content of the summit and present future research opportunities identified by collective discussion.

(Received in original form July 6, 2023; accepted in final form December 19, 2023)

3 This article is open access and distributed under the terms of the Creative Commons Attribution Non-Commercial No Derivatives License 4.0. For commercial usage and reprints, please e-mail Diane Gern (dgern@thoracic.org).

This summit was cosponsored by the NHLBI, Three Lakes Foundation, and Pulmonary Fibrosis Foundation, with funding provided by the NHLBI.

Correspondence and requests for reprints should be addressed to Sydney B. Montesi, M.D., Massachusetts General Hospital, 55 Fruit Street, BUL-148, Boston, MA 02114. E-mail: sbmontesi@partners.org.

Am J Respir Crit Care Med Vol 209, Iss 4, pp 362–373, Feb 15, 2024

Copyright © 2024 by the American Thoracic Society

Originally Published in Press as DOI: 10.1164/rccm.202307-1154WS on December 19, 2023

Internet address: www.atsjournals.org

¹Division of Pulmonary and Critical Care Medicine and ²Department of Pathology, Massachusetts General Hospital, Boston, Massachusetts; ³Division of Lung Diseases, National Heart, Lung, and Blood Institute, National Institutes of Health, Bethesda, Maryland; ⁴Pulmonary and Critical Care Division, University of Pennsylvania, Philadelphia, Pennsylvania; ⁵Program in Neurotherapeutics, University of Massachusetts Chan Medical School, Worcester, Massachusetts; ⁶Department of Medicine, University of Chicago, Chicago, Illinois; ⁷Division of Pulmonary and Critical Care Medicine, ⁸Department of Microbiology and Immunology, and ⁹Department of Biostatistics, University of Michigan, Ann Arbor, Michigan; ¹⁰Division of Pulmonary, Allergy, and Critical Care Medicine, Columbia University Irving Medical Center, New York, New York; ¹¹Department of Pediatrics, David Geffen School of Medicine, ¹²Center for Computer Vision and Imaging Biomarkers, Department of Radiological Sciences, David Geffen School of Medicine, and ¹³Department of Biostatistics, Fielding School of Public Health, University of California, Los Angeles, Los Angeles, California; ¹⁴Women's Guild Lung Institute, Department of Medicine, Cedars-Sinai Medical Center, Los Angeles, California; ¹⁵National Heart and Lung Institute, Imperial College London, London, United Kingdom; ¹⁶Pulmonary, Critical Care and Sleep Medicine, Department of Medicine, Yale School of Medicine, New Haven, Connecticut; ¹⁷Pulmonary, Allergy, Critical Care and Sleep Medicine, University of Pittsburgh, Pittsburgh, Pennsylvania; ¹⁸Division of Respiratory, McMaster University, Hamilton, Ontario, Canada; ¹⁹Center for Regenerative Medicine, Boston University and Boston Medical Center, Boston, Massachusetts; ²⁰Division of Allergy, Pulmonary and Critical Care Medicine, Vanderbilt University Medical Center, Nashville, Tennessee; ²¹Pulmonary Fibrosis Foundation, Chicago, Illinois; ²²Department of Medicine, Tulane University, New Orleans, Louisiana; ²³Department of Bioengineering, ²⁴Department of Pediatrics, ²⁵Division of Pulmonary Sciences and Critical Care Medicine, Department of Medicine, and ²⁶Department of Medicine, University of Colorado Anschutz Medical Campus, Aurora, Colorado; ²⁷Keck School of Medicine, University of Southern California, Los Angeles, California; ²⁸Three Lakes Foundation, Northbrook, Illinois; ²⁹Division of Pulmonary and Critical Care, Weill Cornell Medical College, New York, New York; ³⁰Division of Pulmonary, Sleep and Critical Care Medicine, University of Washington, Seattle, Washington; ³¹Pulmonary, Critical Care and Sleep Medicine, Baylor College of Medicine, Houston, Texas; and ³²Department of Medicine and ³³Gregory Fleming James Cystic Fibrosis Research Center, University of Alabama at Birmingham, Birmingham, Alabama

ORCID IDs: 0000-0001-5323-9507 (C.R.G.); 0000-0002-2149-3209 (M.B.); 0000-0001-7686-0291 (K.R.F.); 0000-0002-0771-1249 (C.K.G.); 0000-0002-1809-723X (B.G.); 0000-0002-7929-2119 (R.G.J.); 0000-0001-5917-4601 (N.K.); 0000-0003-1225-3489 (G.H.J.K.); 0000-0001-9414-5128 (M. Königshoff); 0000-0003-3837-1467 (M. Kolb); 0000-0002-8923-1344 (J.A.K.); 0000-0002-6988-8584 (C.M.M.); 0000-0001-7192-9149 (T.M.M.); 0000-0003-3051-745X (B.B.M.); 0000-0003-4957-8869 (J.O.); 0000-0002-9559-1485 (A.J.P.); 0000-0001-6743-8443 (D.S.); 0000-0002-2412-3182 (F.J.M.).

Novel Models and Research Tools to Better Study Pulmonary Fibrosis and Uncover New Therapies

Despite numerous experimental studies describing the efficacy of novel compounds for reducing fibrosis, therapies that halt or reverse pulmonary fibrosis in patients remain elusive. The limitations of traditional models of pulmonary fibrosis to recapitulate human lung architecture and cellular composition and reflect the phenotypic and mechanistic features of human disease have been a major hurdle hampering drug development. The ability to model the highly complex biological phenomena of pulmonary fibrosis is crucial to better understand the biology of the disease, identify new drug targets, and test the efficacy of novel or repurposed compounds (5, 6).

Novel Animal Models

There is ongoing work to develop models that more accurately mirror the evolution from early injury through fibrogenesis to resolution. Moreover, animal models to study nonresolving, progressive fibrosis and early disease time points are also needed. Ferrets may serve as a model for studying a persistent fibrotic phenotype and elucidating the connection between MUC5B expression and abnormal lung remodeling. A gain-of-function *MUC5B* promoter variant is

recognized as the dominant genetic risk factor in IPF, and ferrets have a distribution of MUC5B-producing submucosal glands similar to humans and express high amounts of *MUC5B* with native presence of the risk-conferring rs35705950 TT promoter variant (7). Human cell types, absent in mice, that participate in aberrant repair have also been identified in ferrets (8). Ferrets exposed to bleomycin exhibit persistent fibrosis with histopathology like human IPF with prominent airway remodeling, proximalization of the distal airway spaces, and MUC5B-rich honeycomb cystic structures that resemble honeycomb cysts in humans (7, 9). Limitations to the ferret model include increased cost and model complexity because of longer ferret lifespan, fewer ferret-specific reagents, and need for more drug compound in therapeutic experiments than in murine models.

Genetic models with intrinsic defects may enable early events in the fibrosis cascade culminating from epithelial cell dysfunction to be studied; these may better mimic pulmonary fibrosis initiation in humans. Mutations in the alveolar epithelial type 2 (AT2) cell restricted surfactant protein C (SP-C) gene (*SFTPC*) have been identified in a subset of patients with IPF (10). *Sftpc* mouse models, such as knock-in murine models expressing either a trafficking (SP-C^{I73T}) or a BRICHOS misfolding (SP-C^{C121G}) mutation, offer an additional and disease-relevant preclinical platform to explore IPF pathogenesis and therapeutic

discovery (11, 12). Expression of either mutation in adult mice results in spontaneous fibrosis and recapitulates many disease-defining elements, including activation of lung tissue repair-associated pathways, heterogeneous fibrotic histology with features of usual interstitial pneumonia, restrictive lung physiology, and elevation of relevant IPF biomarkers (i.e., MMP7) (11, 12).

Despite the recognition of the need for novel models, traditional murine models continue to offer vast opportunities to study mechanisms driving pulmonary fibrosis and could importantly assist in the interrogation of the findings elucidated by single-cell profiling (13). Murine models enable genetic manipulation or deletion of cell lines and can be used to delineate the roles of specific fibroblast states, macrophage subtypes, and endothelial or epithelial cells within tissue injury and repair. These genetic models could then be shared with the research community more easily and at a lower cost than many other novel models to catalyze discovery.

Three-Dimensional Models of Pulmonary Fibrosis

Lung tissue-based precision-cut lung slices (PCLSs) have emerged as a promising model system for chronic lung diseases (14). PCLSs are 300–500-μm-thick sections of lung tissue that can be generated from explanted or resected lung tissue (15). PCLSs model lung

structure and function in its native three-dimensional (3D) environment, thus reflecting natural interactions between cells, molecules, and the extracellular matrix (ECM) *ex vivo*. Subjecting healthy donor lungs to a fibrotic cocktail treatment (transforming growth factor- β , platelet-derived growth factor-AB, lysophosphatidic acid, and tumor necrosis factor- α) over a time course of 5–7 days results in an *ex vivo* model of pulmonary fibrosis initiation (16) that can be used to screen candidate interventions (17–20). To further develop PCLSs as a robust tool, standardization of PCLS generation and culture is needed. Studies altering the structure and microenvironment of the PCLS, using stretch or novel biomaterials to alter stiffness and matrix–cell communication, would be helpful to better mimic fibrotic disease. Techniques that allow genetic manipulation in human lung tissue would further maximize the use of PCLSs for mechanistic and therapeutic studies.

Hydrogel biomaterials can be engineered to control and study cell–matrix interactions in real time. Advances in lung decellularization techniques have fueled a growing interest in biomaterials from decellularized ECM. To study the influence of cell–matrix interactions on fibrotic cellular activation *in vitro*, a new class of photoaddressable hybrid hydrogels containing a dynamically tunable polyethylene glycol backbone and clickable decellularized ECM has been developed (21, 22). These materials support on-demand spatiotemporal control over local mechanical properties in 3D cultures facilitating epithelium–fibroblast interactions. Precisely designed microenvironments can facilitate controlled biological studies aimed at understanding the dynamic cell–matrix interactions that occur during fibrosis-related ECM remodeling (23). Current models may not fully recapitulate the *in vivo* ECM or the *in vivo* inflammatory milieu and lack an air–liquid interface.

Organoids are 3D structures that mimic the organ cellular and structural microenvironment and enable cell–cell interactions to be studied in a tissue-like environment. Lung organoids have been developed using a bead-based microsccaffold lung cell coculture approach to generate a tissue-like structure for disease modeling and studying epithelial–mesenchymal interactions (24–27). Using combinations of primary and induced pluripotent stem cell

(iPSC)-derived cell types and ECM, a 3D model was created with a phenotype resembling IPF. Progressive fibrosis was seen over time in culture, which correlated with increased senescent cells and a senescence-associated secretory phenotype. Such models could be scalable and amenable to high-throughput drug screening with newer 3D imaging techniques and machine learning algorithms. Current models lack vasculature and air flow. To study cell–cell interactions across multiple cell types, advances in model complexity are needed to enable the addition of cell types such as endothelial and immune cells.

Pluripotent Stem Cells

Methods for the generation of iPSCs from peripheral blood and the directed differentiation of iPSCs into a variety of lung epithelial lineages have been developed for disease modeling or cell-based therapies (28–31). Reprogramming patient-specific somatic cells can provide an inexhaustible source of disease-specific iPSCs for disease modeling, drug screens, or cell reconstitution. For example, by using patient-derived cells that carry a disease-causing mutation, *SFTPC*^{L73T}, known to be expressed solely in AT2 cells, mechanisms associated with the inception of type 2 alveolar epithelial cell dysfunction can be elucidated, including potential druggable target pathways for therapeutic intervention (32). To move from epithelium-only models of disease inception to more complex models of epithelial–mesenchymal crosstalk, models have been developed where iPSC-derived epithelium can be cultured together with mesenchymal lineages in 3D cultures (33). These models offer an opportunity for more complex studies of disease pathogenesis, including measures of fibrogenesis, the potential to employ CRISPR gene editing to examine the impact of specific polymorphisms or mutations, and the ability to screen for drugs that ameliorate either epithelial dysfunction or the fibrotic mesenchymal response. Important gaps to be addressed include the development of standardized differentiation procedures for all lung cell types and cocultures of multiple cell types to better mimic the *in vivo* cellular environment of interest.

Single-Cell Profiling

Single-cell profiling technologies offer unprecedented opportunities to profile DNA, mRNA, and proteins at a single-cell resolution (34). The application of single-cell RNA sequencing led to the recognition that

reduced numbers of AT1 and AT2 cells and increased numbers of airway epithelial and systemic venous endothelial cells were hallmarks of human pulmonary fibrosis (35–38). Cell populations that were not described previously, such as the profibrotic macrophages (38, 39) or the aberrant basaloid cells, were identified (37, 40, 41). An atlas of IPF cell data has been publicly shared in a user-friendly data-sharing and dissemination portal (www.IPFCellAtlas.com) (42). However, limitations to single-cell data acquired to date include an overdependence on end-stage lung tissues and the lack of samples from a diverse patient population with various etiologies of pulmonary fibrosis.

Specific compartments such as pulmonary lymph nodes have not yet been studied and may be important to fully understand disease mechanisms and to better characterize the role of the immune system in pulmonary fibrosis. In addition to single-cell or nuclear or spatial transcriptomics, single-cell resolution metabolomics, genomics, and epigenomics should also be applied. Development of standards for data preservation and sharing are critically required to make the data widely accessible. With the emergence of these complex datasets, computational models of the human fibrotic lung could be used to simulate the effects of disease-modifying perturbations on fibrosis and potentially help in prioritizing drug targets and compounds.

Standardized collection of well-characterized samples stored in media or conditions that allow single-cell profiling across the spectrum of pulmonary fibrosis is vitally needed. Such well-characterized collection of cells and tissue would add great value for both single-cell profiling and use in other model systems discussed above. This endeavor should extend beyond lung explants to identify resources for lung tissue from patients with preclinical pulmonary fibrosis, familial pulmonary fibrosis, and non-IPF ILD to maximize impact. Clinically performed bronchoscopies and lung biopsies represent opportunities for sample collection, and the development of a live-cell bank was collectively identified as a resource to facilitate future mechanistic inquiry.

Translating Preclinical Fibrosis Models to Human Disease

The translation of preclinical models to human disease in pulmonary fibrosis

remains a “valley of death” in drug development (43). Modeling of lung fibrosis is a vastly complex task, and even the most robust translational system can only ever characterize a portion of the true disease biology (44). There is no perfect model (Table 1). Deriving translational value from preclinical modeling requires using models correctly and timing interventions appropriately (45). A small fraction of studies evaluating candidate IPF treatments in the bleomycin rodent model have tested the drugs as therapeutic and not as preventive agents (46). There is a lack of published preclinical data from disease models for most of the compounds tested in human studies during the last decade that eventually failed to demonstrate favorable results. The lack of access to the experimental details makes it impossible to understand the predictive value of the models for future clinical trial success. Publishing these results would have immense value to the scientific community.

There is an opportunity to have greater integration between diverse stakeholders to translate discoveries related to the pathobiology of pulmonary fibrosis into new drugs. There are several novel investigational therapies in the clinical development pipeline. It may be beneficial to foster discussion to promote incorporating the preclinical models discussed here for early testing of experimental compounds. Such a collaborative approach may help select which agents are more likely to succeed in clinical trials while helping us understand which, if any, of these newer models have high predictive value of demonstration of efficacy in humans.

Early Disease Risk Factors and Methods to Improve Diagnosis

IPF and other fibrotic ILDs develop insidiously over years. In most cases, a diagnosis of pulmonary fibrosis is made only after considerable disease progression has transpired. Early detection and consequently therapeutic intervention may provide opportunities to preserve lung function and improve overall survival. However, for this to occur, greater recognition of risk factors for pulmonary fibrosis, development of methods to enable early diagnosis, and validation of biomarkers for near-term disease course prediction are essential.

Interstitial Lung Abnormalities

Research over the past decade supports the presence of interstitial lung abnormalities (ILAs), which are incidentally detected interstitial changes on chest computed tomography (CT), as a risk factor for ILD (47–54). Individuals with ILAs share certain genetic risk factors with adults diagnosed with IPF and other fibrotic ILDs. For example, the *MUC5B* variant (rs35705950), short telomere length, and IPF-related rare genetic variants are linked with ILAs (50, 55, 56). The presence of ILAs is independently associated with a 1.3 - 2.7 hazard ratio for death (49). In a cohort of first-degree relatives of patients with familial pulmonary fibrosis, ILA progression occurred in 20% of participants, with half (10%) developing incident ILD over 5 years (57). In another familial cohort, worsening respiratory symptoms occurred in 40%, and worsening fibrosis measured by quantitative CT occurred in 33%, over 4 years (58). Despite a greater awareness of the potential clinical impact of and risk for ILA progression (53, 59), the drivers of progression from ILAs to clinically significant pulmonary fibrosis remain poorly understood. Important knowledge gaps remain, such as how to integrate demographic characteristics, radiologic patterns, and genetic information to identify individuals with the highest risk of near-term disease progression. Such knowledge is essential to enabling future therapeutic efforts focused on prevention of pulmonary fibrosis (60) and represents an important unaddressed opportunity with significant implications for individual risk prediction.

Common and Rare Genetic Variants

The past decade has seen considerable advances in our understanding of rare and common genetic variants associated with IPF (61–63). A *MUC5B* promoter variant is observed in over 30% of subjects with familial or sporadic IPF and can identify individuals at risk for preclinical pulmonary fibrosis (9, 50, 61). Up to two dozen common genetic variants have been shown to contribute to the risk of IPF (62, 64). Rare variants in genes related to surfactant processing and telomere/chromosomal biology have been implicated in familial pulmonary fibrosis and have also been observed in patients with “sporadic” IPF, although typically at somewhat lower prevalence than is seen in familial cohorts (65–69). These findings raise the possibility that a subset of sporadic IPF cases represent

the “index” case in a family, but no studies to date have yet evaluated the impact of such variants on disease risk in the children of patients with IPF.

Major gaps remain in our understanding of genetic risk factors. How do common genetic variants and environmental risk factors contribute to the development, progression, and phenotypic heterogeneity of pulmonary fibrosis? How do rare and common genetic variants interact to mediate and modulate disease risk? What is the role of testing for genetic variants when assessing risk for pulmonary fibrosis in individuals with a family history of sporadic or familial pulmonary fibrosis, and how should results of genetic testing inform recommendations for screening for pulmonary fibrosis? There remains a missing heritability because common and rare variants do not completely explain the heritable risk for this disease. To date, genetic studies have focused on non-Hispanic White people. Greater representation of patients across ethnicities is crucial for a comprehensive elucidation of genetic drivers of pulmonary fibrosis.

Blood-based Biomarkers and Progressive Pulmonary Fibrosis

Multiple candidate blood-based biomarkers have been studied for disease course prediction in IPF. However, there remains no clinically used blood-based biomarker for individual disease course prediction in IPF. Although nearly all patients with IPF progress after diagnosis, variable proportions of those with non-IPF ILD develop a progressive fibrotic phenotype, termed “progressive pulmonary fibrosis” (PPF), with a survival that closely resembles IPF (70, 71). Currently, there is no ability to predict which individuals will develop PPF, and this represents a critical gap in knowledge with significant therapeutic implications. Criteria proposed to identify PPF consist of clinical features that often precede death or lung transplant, including categorical decline in FVC and DL_{CO}; increasing extent of fibrosis on high-resolution CT; worsening respiratory symptoms, including cough and dyspnea; and combinations of these features IPF (70, 71). These conventional measures of ILD progression are easily applied and have been used to successfully test the impact of therapeutic interventions on PPF (4). A PPF clinical practice guideline was recently published (70) that was met with considerable skepticism by the international

community. Criticism of these opinion-based criteria stemmed from their reliance on retrospective cohort data, heavy extrapolation from IPF, and arbitrarily assigned measurement periods (72–74). Recent studies have corroborated these concerns, finding that near-term FVC decline and long-term transplant-free survival were highly heterogeneous, depending on the PPF criterion satisfied (71, 75). The uncertainty regarding the optimal criteria or their timing jeopardizes impactful research and efficient clinical care. As such, ongoing research is needed to better understand this observed phenotypic heterogeneity and define PPF.

Composite models are limited in their ability to predict near-term disease progression (76), suggesting a better ability to identify advanced disease than biologically active disease. Because current criteria for PPF are based on functional and imaging criteria reflective of short-term disease progression (70), biomarkers with a greater predictive ability for near-term disease progression over longer-term survival are essential. Using an unbiased proteomic approach, 17 biomarkers were associated with near-term disease progression, defined as $\geq 10\%$ FVC decline, death, or transplant over 1 year, in both discovery and validation cohorts of participants with non-IPF ILD (77), suggesting that a multibiomarker panel may be needed for near-term disease prediction. Additional research to identify clinically actionable blood-based biomarkers or a combination of multimodal biomarkers remains an important unmet need to enable earlier therapeutic intervention for both IPF and non-IPF ILD. To facilitate data sharing, samples collected under sponsored research could be made available and centrally analyzed to identify and validate the performance of biomarkers across studies.

Radiomics and Emerging Imaging Technology

The past decade has seen increasing research into CT-based radiomics of pulmonary fibrosis. Image texture features from 3D CT datasets are converted to numerical values for mathematical analyses using advanced computational techniques. Single-time point radiomic scores, such as the CALIPER (Computer-Aided Lung Informatics for Pathology Evaluation and Ratings)-derived vessel-related structure scores, have been shown to predict survival in IPF (78). Short-term changes in radiomic scores have also

been associated with survival; for example, an increase in quantitative lung fibrosis score $\geq 4\%$ over 6 months was associated with a reduced progression-free survival in IPF (79). Several radiomic scores, such as quantitative lung fibrosis and quantitative interstitial lung disease, have been used to detect treatment effect in recent IPF or systemic sclerosis-associated ILD clinical trials (80, 81).

Standardization of high-resolution CT acquisitions across study protocols and clinical centers is needed to enable various radiomic measurements to be compared across large numbers of individuals and ILD subtypes. Variations in CT acquisition (sequence parameters, radiation dose, inspiratory air volume) affect imaging outputs (82). The development of a large repository of CT datasets in combination with disease outcomes would facilitate validation of radiomic algorithms for disease course prediction for both established disease and at risk for disease populations. This information could be combined with other omics-based biomarkers for elucidation of pathobiology and development of individual risk prediction.

An imaging-based approach with potential utility for early microscopic diagnosis and disease monitoring over time is endobronchial optical coherence tomography (EB-OCT). EB-OCT is a bronchoscope-compatible, rapid imaging technology that provides microscopic resolution 200 times greater than CT in lung tissue volumes 100 times larger than surgical lung biopsy (83, 84) and exhibited 100% sensitivity and specificity for both histopathologic usual interstitial pneumonia

and clinical IPF diagnoses in patients undergoing diagnostic surgical lung biopsy; there was high agreement between EB-OCT and histopathologic ILD pattern (85). Polarization-sensitive EB-OCT also detects birefringence and fiber orientation in organized tissues, such as collagen, and distinguished destructive fibrosis, nondestructive interstitial fibrosis, and normal parenchyma in participants with ILD (86).

The optimal imaging modalities to best detect early, clinically significant disease and how this information can be incorporated to better predict individual disease course remain to be determined. There was collective consensus on the need for ongoing development of tools to enhance early diagnosis and disease course prediction. It is likely that a multimodal strategy would be needed to best inform individual disease risk for clinical decision making (Figure 1). Such tools would be of great relevance not only to individuals with ILAs or with a family history of pulmonary fibrosis but also to those at risk of developing pulmonary fibrosis in the setting of autoimmune-related diseases such as systemic sclerosis and rheumatoid arthritis.

Machine Learning for IPF Risk Prediction

Emerging research highlights the potential utility of machine learning technology for secondary disease prevention. Delays in diagnosis and initial misdiagnosis are common in patients with IPF because of nonspecific symptoms that overlap with those of more common pulmonary diseases (87).

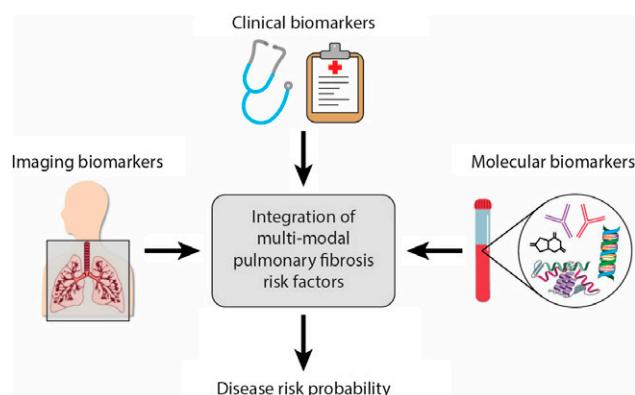


Figure 1. Integration of multimodal risk factors for disease risk probability. The schematic illustrates the integration of imaging, clinical, and molecular information to determine disease risk for an individual patient. Disease risk could pertain to the development of pulmonary fibrosis for those with interstitial lung abnormalities or a family history of pulmonary fibrosis or to the development of disease progression for those with established pulmonary fibrosis.

Using comorbidity signatures from the electronic health records of individuals with IPF, the zero-burden comorbidity risk score for IPF was developed as a screening tool for diagnosis of IPF (88). This machine learning algorithm leverages available data in the electronic health records. The zero-burden comorbidity risk score for IPF predicted IPF diagnosis up to 4 years in the future with good accuracy and, when applied 1 year before diagnosis, achieved positive likelihood ratios exceeding 30 at 99% specificity.

Innovative Approaches to Clinical Trial Design for Pulmonary Fibrosis

Traditional drug development pathways can take up to 10 years (double the median survival of IPF) to evaluate a single treatment. Favorable early-phase trial results are no guarantee of late-phase efficacy, with several late-phase trials recently discontinued because of lack of efficacy for IPF (e.g., the ISABELA [A Clinical Study to Test How Effective and Safe GLPG1690 Is for Subjects with IPF When Used Together with Standard of Care], STARScape [A Study to Evaluate the Efficacy and Safety of Recombinant Human Pentraxin-2 (rhPTX-2; PRM-151) in Participants with Idiopathic Pulmonary Fibrosis], and ZEPHYRUS [Evaluation of Efficacy and Safety of Pamrevlumab in Participants with IPF] trials) (89). Use of background treatment with nintedanib or pirfenidone affects the potential magnitude of the treatment effect of new agents, and the feasibility of recruitment can be more difficult because of changing practice patterns and intense competition for participation of a limited number of patients.

Improving the Efficiency of Early-Phase Trial Design

There remains a need to improve the efficiency of not only late-phase but also early-phase trials. This would enable multiple candidate drugs to be tested in early-phase studies in a shorter time frame to identify the most promising ones to move to late-stage trials. One way to facilitate this is to maximize biomarker assessments to obtain pharmacodynamic information. Several proof-of-concept studies have been conducted that have incorporated assessment of target engagement of

investigational therapies in participants with IPF. Reduction in alveolar macrophage expression of galectin-3 was detected using an inhaled galectin-3 inhibitor in a phase I/IIA study (90). Use of ^{18}F -fluorodeoxyglucose positron emission technology (PET) demonstrated a reduction in ^{18}F -fluorodeoxyglucose uptake in fibrotic lung regions, confirming metabolic effects of a phosphatidylinositol-3-kinase/mammalian target of rapamycin inhibitor in a phase I study (91). An $\alpha_v\beta_6$ -specific PET probe confirmed target engagement of an inhaled inhibitor of $\alpha_v\beta_6$ integrin in a small study of eight participants (92). Other developments that may increase efficiency include Bayesian analysis, which incorporates prior control data into the current trial of interest (93).

Molecular imaging may provide important insight within clinical trials of pulmonary fibrosis, especially within early-phase trial design. A number of molecular probes have been developed for noninvasive assessment of fibrosis (94). PET probes targeting type I collagen and $\alpha_v\beta_6$ integrin and a fibroblast-activating protein inhibitor probe have been translated into use in humans with pulmonary fibrosis (95–99). In addition to enabling assessment of target expression and engagement, molecular imaging also could be used to assist with dose selection or early assessment of treatment response and provide a noninvasive means for a molecular-based cohort enrichment strategy (100). Blood-based biomarkers may also provide an early window into *in vivo* drug effects. Routine incorporation of biomarkers into early-phase trials can also serve to validate candidate biomarkers for identification of individual treatment response.

Innovations in Trial Design

Innovative concepts in trial design have been introduced for therapeutic development for pulmonary fibrosis. Pragmatic trials that increase the number of potential study participants and provide more generalizable results have been demonstrated to be feasible in IPF. CleanUp-IPF (Study of Clinical Efficacy of Antimicrobial Therapy Strategy Using Pragmatic Design in Idiopathic Pulmonary Fibrosis) used limited inclusion and exclusion criteria, completed enrollment ahead of schedule, and produced results similar to those of the EME-TIPAC (Treating Pulmonary Fibrosis with Co-trimoxazole) trial, a placebo-controlled traditional explanatory trial that also

employed cotrimoxazole in participants with IPF (101, 102). The currently enrolling PRECISIONS (Prospective Treatment Efficacy in IPF Using Genotype for Nac Selection) study includes several pragmatic elements and represents the first pharmacogenetic trial in IPF while leveraging a partnership with the Pulmonary Fibrosis Foundation to facilitate recruitment and use of biospecimens through the Pulmonary Fibrosis Foundation Patient Registry (103).

Other forms of trial design, such as umbrella, basket, or platform trials, have been pioneered in oncology (104) and may hold promise for use for pulmonary fibrosis. Adaptive platform trials have been used for other chronic and rare diseases, such as amyotrophic lateral sclerosis (105). An adaptive platform trial allows the evaluation of multiple treatments within an integrated clinical trial infrastructure using a single main protocol and integrated statistical framework. Interventions may be administered in combination, with the list of available treatment arms changing over time as some are found to be effective, ineffective, or harmful and as new treatments become available (104, 106, 107). The ability of the platform trial design to address combination treatment strategies and heterogeneity of treatment benefit across subgroups makes this approach particularly relevant for the evaluation of interventions for pulmonary fibrosis. In addition, the stream of information acquired during the study period can trigger specific changes or adaptations in the trial structure in real time, such as altering randomization proportions or early termination of an arm or subpopulation for demonstration of efficacy, futility, or harm, according to prespecified decision rules (106). Fundamentally, this means that an adaptive platform trial can enable the most information on the therapies that are most effective and that, theoretically, outcomes for patients involved in the trial should improve over time.

Trial Endpoints and Statistical Considerations

Important progress in understanding the strengths and limitations of potential endpoints in late-phase trials in IPF has been made over the last decade. Approval of nintedanib and pirfenidone by regulatory agencies was based on efficacy for slowing the rate of FVC decline over 52 weeks of treatment (2, 3). Other studies have used composite endpoints, such as progression-

Table 1. Novel Models and Tools to Study Pulmonary Fibrosis

Model/Tool	Advantages	Limitations
Ferret model	Pathology recapitulates characteristic key features of pulmonary fibrosis, including persistent fibrosis, prominent MUC5B expression in distal airways, and aberrant repair	Cost and complexity of ferret model Lifespan 5–10 yr with onset of geriatric diseases between 3 and 4 yr Fewer ferret-specific reagents available to characterize the model Requires more compound for drug testing than mouse models Therapeutic applications not yet demonstrated
<i>Sftpc</i> mouse models	Develop spontaneous fibrosis Model early disease Elaborate cytokines and biomarkers detected in pulmonary fibrosis patients Can be used to benchmark drug efficacy	<i>Sftpc</i> locus unavailable for lineage tracing Does not model bronchiolization
Precision-cut lung slices	Natural composition of cells and ECM Live imaging of cell–cell and cell–ECM interaction <i>ex vivo</i> Early stages of fibrosis can be induced Can be used for drug discovery and validation	Limited culture time No ventilation/perfusion No homing of nonresident cells Limited value for translation in relation to route of administration of therapeutic agents (e.g., inhaled or systemic administration)
Hydrogel biomaterials	Enable manipulation of mechanical properties Biochemical changes can be decoupled from biophysical changes Can probe cell–matrix interactions Can create sex-specific models Many cell types can be included Can be used for drug discovery and validation	No air–liquid interface No cyclic stretch Current models may not capture the complexity of the <i>in vivo</i> ECM or <i>in vivo</i> inflammatory milieu
Lung organoids	Enable epithelial–mesenchymal cellular interactions to be studied High-throughput analysis possible for drug screening and biological readouts Many cell types can be included Mechanotransduction forces can be modeled	No vasculature or airflow Does not model bronchiolization Takes a reductionist approach; not all cellular and matrix components are present No homing of nonresident cells
Pluripotent stem cells	Inexhaustible source of cells for generating lung lineages of interest for disease modeling, drug screening, or cell-based therapies Patient-specific, editable, and scalable Allows production of initially normal patient-derived cells to replay or recapitulate disease onset/emergence <i>in vitro</i>	Lack of standardized differentiation procedures for all lung cell types Differentiation protocols for some relevant cell types (e.g., AT2 cells and lung mesenchymal lineages) are currently a work in progress Cocultures of multiple cell types needed to fully recapitulate <i>in vivo</i> environment
Single cell profiling	Allows discovery of novel cellular phenotypes and states associated with fibrosis Provides a detailed atlas of molecular changes and cellular interactions that occur in lung fibrosis Information obtained can be used to orient therapy development focused on specific cell populations in the fibrotic lung	Dependent on tissue availability Only captures transcriptional regulation, not post-transcriptional effects (e.g., mRNA stability or post-translational modifications) Most results to date have been from end-stage lung tissue and are limited in ethnic, racial, and geographical representation

Definition of abbreviation: ECM = extracellular matrix.

free survival, commonly defined as a composite of decline in FVC or death, as a primary endpoint in IPF studies (108, 109). However, composite endpoints often use different components and thresholds, making cross-study comparisons difficult. Discordant effects of a drug on various components of a composite endpoint can obscure efficacy, and a less important component can drive treatment differences, dominating the analysis and interpretation. Although clinically meaningful, the

development of acute exacerbations of IPF as an endpoint is better suited as a secondary endpoint than as a primary endpoint because of the large sample size needed to demonstrate a significant treatment effect for decreasing the frequency of acute exacerbations of IPF. Mortality as a primary endpoint has the highest clinical relevance and meaning to patients; however, a mortality primary endpoint often requires a larger sample size and prolonged trial duration, limiting overall feasibility (110, 111).

Statistical considerations in endpoint selection include improving the inherent information available in an endpoint (e.g., survival [time to event] instead of mortality [binary], recurrent events instead of time to first event) and the relevance of an endpoint (e.g., hierarchical composite endpoint [win ratio] instead of time to first component of a composite endpoint) (112, 113). Exploring home-based functional measures (e.g., home spirometry) may provide more repeated measures, and the potential loss in precision

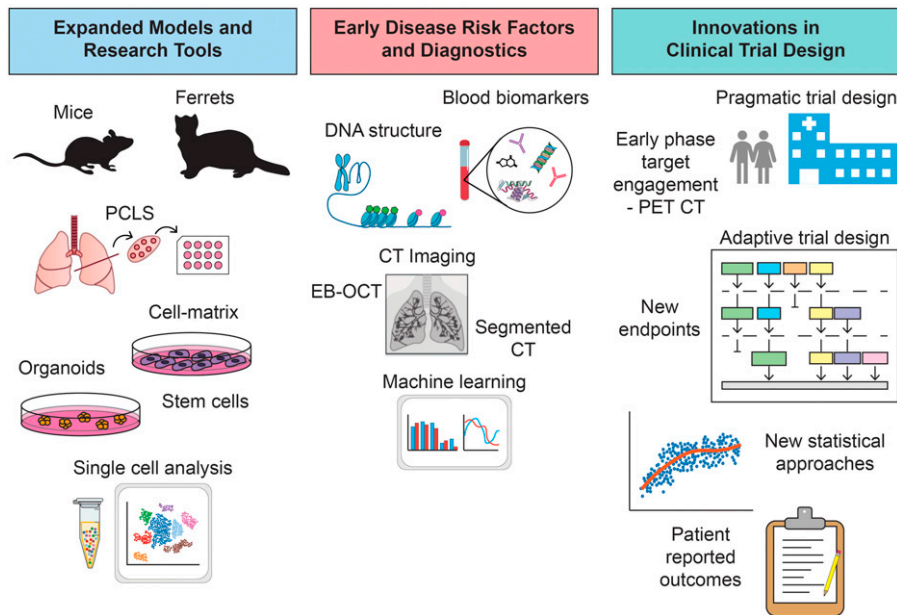


Figure 2. Tools, diagnostics, and innovations to advance discovery, diagnosis, and drug development for pulmonary fibrosis. CT = computed tomography; EB-OCT = endobronchial optical coherence tomography; PCLS = precision-cut lung slices; PET = positron emission tomography.

relative to clinic-based measures may be counterbalanced by increased repeated measures and less dropout (114). However, this approach remains an active research area. The application of more sophisticated statistical methods can also lead to new

insights for endpoint selection. Machine and statistical learning methods can be used to assist with biomarker discovery through data-driven subgroup identification (115). Novel data integration methods allow better prediction capabilities of biomarkers by

borrowing information from internal auxiliary data (116–118) or by incorporating external information from other studies (119–122). These methods work for both cross-sectional and longitudinal analyses, boosting statistical power while accounting for the study population heterogeneity. Increased incorporation of biomarker discovery into clinical trials and sharing of individual-level data from trials, registries, and cohort studies will allow more patient-level meta-analyses and data integration opportunities to inform subsequent clinical trial design.

Patients living with pulmonary fibrosis experience complex and interrelated symptoms of dyspnea, cough, anxiety, and depression that affect quality of life. Patient-reported outcome (PRO) measures enable assessment of patient symptoms and quality of life (123). PROs validated for use in IPF and ILD include the King's Brief Interstitial Lung Disease Questionnaire, the Living with Idiopathic Pulmonary Fibrosis questionnaire, and the Fatigue Severity Scale (124–126). Including PROs in the management of patients with IPF and in clinical trials evaluating the utility of new pharmacotherapies adds a critical, patient-centered dimension to efficacy assessment in clinical trials. Further development, validation, and inclusion of PROs as a measure within clinical trials is needed to strengthen inclusion of patient

Table 2. Research Opportunities for Advancing Pulmonary Fibrosis

Novel models and research tools to better study pulmonary fibrosis and uncover new therapies

- Develop models that recapitulate the evolution from injury through fibrogenesis to resolution
- Test drug candidates in the established fibrotic phase of disease
- Expand collection of live cells at the time of routine clinical procedures (i.e., bronchoscopy)
- Develop a live-cell bank to examine different mutations or gene variants as they relate to disease pathogenesis
- Standardize protocols for tissue and sample collection to yield samples from multiple fibrotic lung diseases at various clinical stages
- Develop a central repository for single-cell analyses of tissue and models

Early disease factors and methods to improve diagnosis

- Determine risk factors and mechanistic drivers of disease progression from preclinical disease to pulmonary fibrosis
- Develop tools to predict individual risk for progressive pulmonary fibrosis in non-idiopathic pulmonary fibrosis interstitial lung disease
- Increase representation across ethnicities for elucidation of genetic risk factors for pulmonary fibrosis
- Determine optimal strategies for screening/surveillance in high-risk populations for pulmonary fibrosis (familial pulmonary fibrosis families, connective tissue disease)
- Define recommendations for incorporating genetic testing into clinical practice for individuals with established pulmonary fibrosis
- Standardize high-resolution computed tomography acquisition parameters to facilitate large-scale radiomic analyses
- Develop tools to enhance early diagnosis and disease course prediction

Innovative approaches to clinical trial design for pulmonary fibrosis

- Incorporate novel imaging and blood-based biomarkers into early-phase clinical trials
- Leverage innovative statistical methods to enable adaptive trial design
- Develop a trial platform to assess multiple candidate therapies for pulmonary fibrosis across a range of subtypes of pulmonary fibrosis
- Synergize trial design to facilitate data sharing and cross-validation of clinical trial results
- Increase diversity and inclusion in recruitment and enrollment of patients with pulmonary fibrosis into clinical trials
- Develop and validate clinical trial endpoints to support the conduct of adequate and well-controlled studies

experience and quality of life as essential components of treatment efficacy.

Summary

Although great progress has been made over the past decade, there remains much work to be done to improve the lives of patients living with pulmonary fibrosis. This summit provided a platform for investigators, sponsors, physicians, and patients to share

innovative ideas with the pulmonary fibrosis community with the goal of ultimately improving outcomes for patients with pulmonary fibrosis (Figure 2). It is our hope that the collective research needs put forth (Table 2) will catalyze collaboration and discoveries that will one day make curing pulmonary fibrosis a reality. ■

Author disclosures are available with the text of this article at www.atsjournals.org.

Acknowledgment: More than 100 individuals from around the world attended this virtual summit. The authors acknowledge the many individuals whose input contributed to the content of this report. The findings, knowledge gaps, and opportunities described herein represent a summary of individual opinions and ideas expressed during the workshop. The summary does not represent a consensus opinion or directive made to or by the NHLBI or the NIH. The authors also acknowledge Susan Sheng for her assistance with figure illustrations.

References

- Blackwell TS, Tager AM, Borok Z, Moore BB, Schwartz DA, Anstrom KJ, et al. Future directions in idiopathic pulmonary fibrosis research. An NHLBI workshop report. *Am J Respir Crit Care Med* 2014;189:214–222.
- King TE Jr, Bradford WZ, Castro-Bernardini S, Fagan EA, Glaspole I, Glassberg MK, et al.; ASCEND Study Group. A phase 3 trial of pirfenidone in patients with idiopathic pulmonary fibrosis. *N Engl J Med* 2014;370:2083–2092.
- Richeldi L, du Bois RM, Raghu G, Azuma A, Brown KK, Costabel U, et al.; INPULSIS Trial Investigators. Efficacy and safety of nintedanib in idiopathic pulmonary fibrosis. *N Engl J Med* 2014;370:2071–2082.
- Flaherty KR, Wells AU, Cottin V, Devaraj A, Walsh SLF, Inoue Y, et al.; INBUILD Trial Investigators. Nintedanib in progressive fibrosing interstitial lung diseases. *N Engl J Med* 2019;381:1718–1727.
- Jenkins RG, Moore BB, Chambers RC, Eickelberg O, Königshoff M, Kolb M, et al.; ATS Assembly on Respiratory Cell and Molecular Biology. An official American Thoracic Society workshop report: use of animal models for the preclinical assessment of potential therapies for pulmonary fibrosis. *Am J Respir Cell Mol Biol* 2017;56:667–679.
- Bonnaud P, Fabre A, Frossard N, Guignabert C, Inman M, Kuebler WM, et al. Optimising experimental research in respiratory diseases: an ERS statement. *Eur Respir J* 2018;51:1702133.
- Lever JEP, Phillips S, Bakshi S, Lever JMP, Crossman DK, Crowley MR, et al. Mucus matters: bleomycin induced pulmonary fibrosis in ferrets is sustained and recapitulates features of human idiopathic pulmonary fibrosis [abstract]. *Am J Respir Crit Care Med* 2020;A5989.
- Basil MC, Cardenas-Diaz FL, Kathirya JJ, Morley MP, Carl J, Brumwell AN, et al. Human distal airways contain a multipotent secretory cell that can regenerate alveoli. *Nature* 2022;604:120–126.
- Yang IV, Fingerlin TE, Evans CM, Schwarz MI, Schwartz DA. MUC5B and idiopathic pulmonary fibrosis. *Ann Am Thorac Soc* 2015;12(Suppl 2):S193–S199.
- Lawson WE, Grant SW, Ambrosini V, Womble KE, Dawson EP, Lane KB, et al. Genetic mutations in surfactant protein C are a rare cause of sporadic cases of IPF. *Thorax* 2004;59:977–980.
- Nureki SI, Tomer Y, Venosa A, Katzen J, Russo SJ, Jamil S, et al. Expression of mutant Sftpc in murine alveolar epithelia drives spontaneous lung fibrosis. *J Clin Invest* 2018;128:4008–4024.
- Katzen J, Wagner BD, Venosa A, Kopp M, Tomer Y, Russo SJ, et al. An SFTPC BRICHOS mutant links epithelial ER stress and spontaneous lung fibrosis. *JCI Insight* 2019;4:e126125.
- Moore BB, Lawson WE, Oury TD, Sisson TH, Raghavendran K, Hogaboam CM. Animal models of fibrotic lung disease. *Am J Respir Cell Mol Biol* 2013;49:167–179.
- Alsafadi HN, Uhl FE, Pineda RH, Bailey KE, Rojas M, Wagner DE, et al. Applications and approaches for 3D precision-cut lung slices: disease modeling and drug discovery. *Am J Respir Cell Mol Biol* 2020;62:681–691.
- Gerckens M, Alsafadi HN, Wagner DE, Lindner M, Burgstaller G, Königshoff M. Generation of human 3D lung tissue cultures (3D-LTCs) for disease modeling. *J Vis Exp* 2019;(144):e58437.
- Alsafadi HN, Staab-Weijnitz CA, Lehmann M, Lindner M, Peschel B, Königshoff M, et al. An ex vivo model to induce early fibrosis-like changes in human precision-cut lung slices. *Am J Physiol Lung Cell Mol Physiol* 2017;312:L896–L902.
- Lehmann M, Buhl L, Alsafadi HN, Klee S, Hermann S, Mutze K, et al. Differential effects of nintedanib and pirfenidone on lung alveolar epithelial cell function in ex vivo murine and human lung tissue cultures of pulmonary fibrosis. *Respir Res* 2018;19:175.
- Ahangari F, Becker C, Foster DG, Chioiccioli M, Nelson M, Beke K, et al. Saracatinib, a selective Src kinase inhibitor, blocks fibrotic responses in preclinical models of pulmonary fibrosis. *Am J Respir Crit Care Med* 2022;206:1463–1479.
- Tsoyi K, Liang X, De Rossi G, Ryter SW, Xiong K, Chu SG, et al. CD148 deficiency in fibroblasts promotes the development of pulmonary fibrosis. *Am J Respir Crit Care Med* 2021;204:312–325.
- Gerckens M, Schorpp K, Pelizza F, Wögrath M, Reichau K, Ma H, et al. Phenotypic drug screening in a human fibrosis model identified a novel class of antifibrotic therapeutics. *Sci Adv* 2021;7:eabb3673.
- Saleh KS, Hewawasam R, Šerbedžija P, Blomberg R, Noreldeen SE, Edelman B, et al. Engineering hybrid-hydrogels comprised of healthy or diseased decellularized extracellular matrix to study pulmonary fibrosis. *Cell Mol Bioeng* 2022;15:505–519.
- Petrou CL, D'Ovidio TJ, Bölükbas DA, Tas S, Brown RD, Allawzi A, et al. Clickable decellularized extracellular matrix as a new tool for building hybrid-hydrogels to model chronic fibrotic diseases in vitro. *J Mater Chem B Mater Biol Med* 2020;8:6814–6826.
- Bailey KE, Pino C, Lennon ML, Lyons A, Jacot JG, Lammers SR, et al. Embedding of precision-cut lung slices in engineered hydrogel biomaterials supports extended ex vivo culture. *Am J Respir Cell Mol Biol* 2020;62:14–22.
- Wilkinson DC, Alva-Ornelas JA, Sucre JMS, Vijayaraj P, Durra A, Richardson W, et al. Development of a three-dimensional bioengineering technology to generate lung tissue for personalized disease modeling. *Stem Cells Transl Med* 2017;6:622–633.
- Wilkinson DC, Melody M, Meneses LK, Hope AC, Dunn B, Gomperts BN. Development of a three-dimensional bioengineering technology to generate lung tissue for personalized disease modeling. *Curr Protoc Stem Cell Biol* 2018;46:e56.
- Sucré JMS, Wilkinson D, Vijayaraj P, Paul M, Dunn B, Alva-Ornelas JA, et al. A three-dimensional human model of the fibroblast activation that accompanies bronchopulmonary dysplasia identifies Notch-mediated pathophysiology. *Am J Physiol Lung Cell Mol Physiol* 2016;310:L889–L898.
- Sucré JMS, Vijayaraj P, Aros CJ, Wilkinson D, Paul M, Dunn B, et al. Posttranslational modification of β -catenin is associated with pathogenic fibroblastic changes in bronchopulmonary dysplasia. *Am J Physiol Lung Cell Mol Physiol* 2017;312:L186–L195.
- Sommer AG, Rozelle SS, Sullivan S, Mills JA, Park SM, Smith BW, et al. Generation of human induced pluripotent stem cells from peripheral blood using the STEMCCA lentiviral vector. *J Vis Exp* 2012;(68):4327.
- Jacob A, Morley M, Hawkins F, McCauley KB, Jean JC, Heins H, et al. Differentiation of human pluripotent stem cells into functional lung alveolar epithelial cells. *Cell Stem Cell* 2017;21:472–488, e10.
- Hawkins FJ, Suzuki S, Beermann ML, Barilla C, Wang R, Villacorta-Martin C, et al. Derivation of airway basal stem cells from human pluripotent stem cells. *Cell Stem Cell* 2021;28:79–95, e8.

31. Alysandratos K-D, Herriges MJ, Kotton DN. Epithelial stem and progenitor cells in lung repair and regeneration. *Annu Rev Physiol* 2021;83:529–550.
32. Alysandratos K-D, Russo SJ, Petcherski A, Taddeo EP, Acín-Pérez R, Villacorta-Martin C, *et al.* Patient-specific iPSCs carrying an SFTPC mutation reveal the intrinsic alveolar epithelial dysfunction at the inception of interstitial lung disease. *Cell Rep* 2021;36:109636.
33. Sharma A, Sances S, Workman MJ, Svendsen CN. Multi-lineage human iPSC-derived platforms for disease modeling and drug discovery. *Cell Stem Cell* 2020;26:309–329.
34. Adams TS, Marlier A, Kaminski N. Lung cell atlases in health and disease. *Annu Rev Physiol* 2023;85:47–69.
35. Morse C, Tabib T, Sembrat J, Buschur KL, Bittar HT, Valenzi E, *et al.* Proliferating SPP1/MERTK-expressing macrophages in idiopathic pulmonary fibrosis. *Eur Respir J* 2019;54:1802441.
36. Reyman PA, Walter JM, Joshi N, Anekalla KR, McQuattie-Pimentel AC, Chiu S, *et al.* Single-cell transcriptomic analysis of human lung provides insights into the pathobiology of pulmonary fibrosis. *Am J Respir Crit Care Med* 2019;199:1517–1536.
37. Adams TS, Schupp JC, Poli S, Ayaub EA, Neumark N, Ahangari F, *et al.* Single-cell RNA-seq reveals ectopic and aberrant lung-resident cell populations in idiopathic pulmonary fibrosis. *Sci Adv* 2020;6:eaba1983.
38. Wendisch D, Dietrich O, Mari T, von Stillfried S, Ibarra IL, Mittermaier M, *et al.*; Deutsche COVID-19 OMICS Initiative (DeCOI). SARS-CoV-2 infection triggers profibrotic macrophage responses and lung fibrosis. *Cell* 2021;184:6243–6261, e27.
39. Justet A, Zhao AY, Kaminski N. From COVID to fibrosis: lessons from single-cell analyses of the human lung. *Hum Genomics* 2022;16:20.
40. Habermann AC, Gutierrez AJ, Bui LT, Yahn SL, Winters NI, Calvi CL, *et al.* Single-cell RNA sequencing reveals profibrotic roles of distinct epithelial and mesenchymal lineages in pulmonary fibrosis. *Sci Adv* 2020;6:eaba1972.
41. Valenzi E, Tabib T, Papazoglou A, Sembrat J, Trejo Bittar HE, Rojas M, *et al.* Disparate interferon signaling and shared aberrant basaloid cells in single-cell profiling of idiopathic pulmonary fibrosis and systemic sclerosis-associated interstitial lung disease. *Front Immunol* 2021;12:595811.
42. Neumark N, Cosme C Jr, Rose K-A, Kaminski N. The idiopathic pulmonary fibrosis cell atlas. *Am J Physiol Lung Cell Mol Physiol* 2020;319:L887–L893.
43. Seyhan AA. Lost in translation: the valley of death across preclinical and clinical divide—identification of problems and overcoming obstacles. *Transl Med Commun* 2019;4:18.
44. Yanagihara T, Chong SG, Vierhout M, Hirota JA, Ask K, Kolb M. Current models of pulmonary fibrosis for future drug discovery efforts. *Expert Opin Drug Discov* 2020;15:931–941.
45. Kolb P, Upagupta C, Vierhout M, Ayaub E, Bellaye PS, Gauldie J, *et al.* The importance of interventional timing in the bleomycin model of pulmonary fibrosis. *Eur Respir J* 2020;55:1901105.
46. Moeller A, Ask K, Warburton D, Gauldie J, Kolb M. The bleomycin animal model: a useful tool to investigate treatment options for idiopathic pulmonary fibrosis? *Int J Biochem Cell Biol* 2008;40:362–382.
47. Podolanczuk AJ, Oelsner EC, Barr RG, Bernstein EJ, Hoffman EA, Easthausen LJ, *et al.* High-attenuation areas on chest computed tomography and clinical respiratory outcomes in community-dwelling adults. *Am J Respir Crit Care Med* 2017;196:1434–1442.
48. Podolanczuk AJ, Oelsner EC, Barr RG, Hoffman EA, Armstrong HF, Austin JHM, *et al.* High attenuation areas on chest computed tomography in community-dwelling adults: the MESA study. *Eur Respir J* 2016;48:1442–1452.
49. Putman RK, Hatabu H, Araki T, Gudmundsson G, Gao W, Nishino M, *et al.*; Evaluation of COPD Longitudinally to Identify Predictive Surrogate Endpoints (ECLIPSE) Investigators; COPDGene Investigators. Association between interstitial lung abnormalities and all-cause mortality. *JAMA* 2016;315:672–681.
50. Hunninghake GM, Hatabu H, Okajima Y, Gao W, Dupuis J, Latourelle JC, *et al.* MUC5B promoter polymorphism and interstitial lung abnormalities. *N Engl J Med* 2013;368:2192–2200.
51. Washko GR, Hunninghake GM, Fernandez IE, Nishino M, Okajima Y, Yamashiro T, *et al.*; COPDGene Investigators. Lung volumes and emphysema in smokers with interstitial lung abnormalities. *N Engl J Med* 2011;364:897–906.
52. Zhang Y, Wan H, Richeldi L, Zhu M, Huang Y, Xiong X, *et al.* Reticulation is a risk factor of progressive subpleural nonfibrotic interstitial lung abnormalities. *Am J Respir Crit Care Med* 2022;206:178–185.
53. Rose JA, Menon AA, Hino T, Hata A, Nishino M, Lynch DA, *et al.* Suspected interstitial lung disease in COPDGene Study. *Am J Respir Crit Care Med* 2023;207:60–68.
54. Hatabu H, Hunninghake GM, Richeldi L, Brown KK, Wells AU, Remy-Jardin M, *et al.* Interstitial lung abnormalities detected incidentally on CT: a position paper from the Fleischner Society. *Lancet Respir Med* 2020;8:726–737.
55. Putman RK, Axelsson GT, Ash SY, Sanders JL, Menon AA, Araki T, *et al.* Interstitial lung abnormalities are associated with decreased mean telomere length. *Eur Respir J* 2022;60:2101814.
56. Hobbs BD, Putman RK, Araki T, Nishino M, Gudmundsson G, Gudnason V, *et al.* Overlap of genetic risk between interstitial lung abnormalities and idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med* 2019;200:1402–1413.
57. Salisbury ML, Hewlett JC, Ding G, Markin CR, Douglas K, Mason W, *et al.* Development and progression of radiologic abnormalities in individuals at risk for familial ILD. *Am J Respir Crit Care Med* 2019;201:1230–1239.
58. Steele MP, Peljto AL, Mathai SK, Humphries S, Bang TJ, Oh A, *et al.* Incidence and progression of fibrotic lung disease in an at-risk cohort. *Am J Respir Crit Care Med* 2023;207:587–593.
59. Park S, Choe J, Hwang HJ, Noh HN, Jung YJ, Lee J-B, *et al.* Long-term follow-up of interstitial lung abnormality: implication in follow-up strategy and risk thresholds. *Am J Respir Crit Care Med* 2023;208:858–867.
60. Kim JS, Montesi SB, Adegunsaye A, Humphries SM, Salisbury ML, Hariri LP, *et al.* Approach to clinical trials for the prevention of pulmonary fibrosis. *Ann Am Thorac Soc* 2023;20:1683–1693.
61. Seibold MA, Wise AL, Speer MC, Steele MP, Brown KK, Loyd JE, *et al.* A common MUC5B promoter polymorphism and pulmonary fibrosis. *N Engl J Med* 2011;364:1503–1512.
62. Fingerlin TE, Murphy E, Zhang W, Peljto AL, Brown KK, Steele MP, *et al.* Genome-wide association study identifies multiple susceptibility loci for pulmonary fibrosis. *Nat Genet* 2013;45:613–620.
63. Allen RJ, Stockwell A, Oldham JM, Guillen-Guio B, Schwartz DA, Maher TM, *et al.*; International IPF Genetics Consortium. Genome-wide association study across five cohorts identifies five novel loci associated with idiopathic pulmonary fibrosis. *Thorax* 2022;77:829–833.
64. Allen RJ, Guillen-Guio B, Croot E, Kraven LM, Moss S, Stewart I, *et al.* Genetic overlap between idiopathic pulmonary fibrosis and COVID-19. *Eur Respir J* 2022;60:2103132.
65. Zhang D, Povysil G, Kobeissy PH, Li Q, Wang B, Amelotte M, *et al.* Rare and common variants in *KIF15* contribute to genetic risk of idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med* 2022;206:56–69.
66. Dressen A, Abbas AR, Cabanski C, Reeder J, Ramalingam TR, Neighbors M, *et al.* Analysis of protein-altering variants in telomerase genes and their association with MUC5B common variant status in patients with idiopathic pulmonary fibrosis: a candidate gene sequencing study. *Lancet Respir Med* 2018;6:603–614.
67. Petrovski S, Todd JL, Durheim MT, Wang Q, Chien JW, Kelly FL, *et al.* An exome sequencing study to assess the role of rare genetic variation in pulmonary fibrosis. *Am J Respir Crit Care Med* 2017;196:82–93.
68. Sutton RM, Bittar HT, Sullivan DI, Silva AG, Bahudhanapati H, Parikh AH, *et al.* Rare surfactant-related variants in familial and sporadic pulmonary fibrosis. *Hum Mutat* 2022;43:2091–2101.
69. Moore C, Blumhagen RZ, Yang IV, Walts A, Powers J, Walker T, *et al.* Resequencing study confirms that host defense and cell senescence gene variants contribute to the risk of idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med* 2019;200:199–208.
70. Raghu G, Remy-Jardin M, Richeldi L, Thomson CC, Inoue Y, Johkoh T, *et al.* Idiopathic pulmonary fibrosis (an update) and progressive pulmonary fibrosis in adults: an official ATS/ERS/JRS/ALAT clinical practice guideline. *Am J Respir Crit Care Med* 2022;205:e18–e47.
71. Pugashetti JV, Adegunsaye A, Wu Z, Lee CT, Srikrishnan A, Ghodrati S, *et al.* Validation of proposed criteria for progressive pulmonary fibrosis. *Am J Respir Crit Care Med* 2023;207:69–76.
72. Cottin V, Brown KK, Flaherty KR, Wells AU. Progressive pulmonary fibrosis: should the timelines be taken out of the definition? *Am J Respir Crit Care Med* 2022;206:1293–1294.

73. Johansson KA, Kolb M, Fisher JH, Walsh SLF. Progressive pulmonary fibrosis: putting the cart before the horse. *Am J Respir Crit Care Med* 2022;206:1294–1295.
74. Khor YH, Farooqi M, Hambly N, Kolb M, Ryerson CJ; Austin ILD Registry and CARE-PF Investigators. Patient characteristics and survival for progressive pulmonary fibrosis using different definitions. *Am J Respir Crit Care Med* 2023;207:102–105.
75. Oldham JM, Lee CT, Wu Z, Bowman WS, Pugashetti JV, Dao N, et al. Lung function trajectory in progressive fibrosing interstitial lung disease. *Eur Respir J* 2022;59:2101396.
76. Ley B, Bradford WZ, Vittinghoff E, Weycker D, du Bois RM, Collard HR. Predictors of mortality poorly predict common measures of disease progression in idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med* 2016;194:711–718.
77. Bowman WS, Newton CA, Linderholm AL, Neely ML, Pugashetti JV, Kaul B, et al. Proteomic biomarkers of progressive fibrosing interstitial lung disease: a multicentre cohort analysis. *Lancet Respir Med* 2022;10:593–602.
78. Jacob J, Bartholmai BJ, Rajagopalan S, van Moersel CHM, van Es HW, van Beek FT, et al. Predicting outcomes in idiopathic pulmonary fibrosis using automated computed tomographic analysis. *Am J Respir Crit Care Med* 2018;198:767–776.
79. Kim GHJ, Weigt SS, Belperio JA, Brown MS, Shi Y, Lai JH, et al. Prediction of idiopathic pulmonary fibrosis progression using early quantitative changes on CT imaging for a short term of clinical 18-24-month follow-ups. *Eur Radiol* 2020;30:726–734.
80. Kim GHJ, Goldin JG, Hayes W, Oh A, Soule B, Du S. The value of imaging and clinical outcomes in a phase II clinical trial of a lysophosphatidic acid receptor antagonist in idiopathic pulmonary fibrosis. *Ther Adv Respir Dis* 2021;15:17534666211004238.
81. Roofeh D, Lin CJF, Goldin J, Kim GH, Furst DE, Denton CP, et al.; focuSSced Investigators. Tocilizumab prevents progression of early systemic sclerosis-associated interstitial lung disease. *Arthritis Rheumatol* 2021;73:1301–1310.
82. Wu X, Kim GH, Salisbury ML, Barber D, Bartholmai BJ, Brown KK, et al. Computed tomographic biomarkers in idiopathic pulmonary fibrosis. The future of quantitative analysis. *Am J Respir Crit Care Med* 2019;199:12–21.
83. Hariri LP, Adams DC, Wain JC, Lanuti M, Muniappan A, Sharma A, et al. Endobronchial optical coherence tomography for low-risk microscopic assessment and diagnosis of idiopathic pulmonary fibrosis in vivo. *Am J Respir Crit Care Med* 2018;197:949–952.
84. Yun SH, Tearney GJ, Vakoc BJ, Shishkov M, Oh WY, Desjardins AE, et al. Comprehensive volumetric optical microscopy in vivo. *Nat Med* 2006;12:1429–1433.
85. Nandy S, Raphaely RA, Muniappan A, Shih A, Roop BW, Sharma A, et al. Diagnostic accuracy of endobronchial optical coherence tomography for the microscopic diagnosis of usual interstitial pneumonia. *Am J Respir Crit Care Med* 2021;204:1164–1179.
86. Nandy S, Bergei SR, Keyes CM, Muniappan A, Auchincloss HG, Lanuti M, et al. Polarization-sensitive endobronchial optical coherence tomography for microscopic imaging of fibrosis in interstitial lung disease. *Am J Respir Crit Care Med* 2022;206:905–910.
87. Collard HR, Tino G, Noble PW, Shreve MA, Michaels M, Carlson B, et al. Patient experiences with pulmonary fibrosis. *Respir Med* 2007;101:1350–1354.
88. Onishchenko D, Marlowe RJ, Ngufor CG, Faust LJ, Limper AH, Hunninghake GM, et al. Screening for idiopathic pulmonary fibrosis using comorbidity signatures in electronic health records. *Nat Med* 2022;28:2107–2116.
89. Maher TM, van der Aar EM, Van de Steen O, Allamassey L, Desrivot J, Dupont S, et al. Safety, tolerability, pharmacokinetics, and pharmacodynamics of GLPG1690, a novel autotaxin inhibitor, to treat idiopathic pulmonary fibrosis (FLORA): a phase 2a randomised placebo-controlled trial. *Lancet Respir Med* 2018;6:627–635.
90. Hirani N, MacKinnon AC, Nicol L, Ford P, Schambye H, Pedersen A, et al. Target inhibition of galectin-3 by inhaled TD139 in patients with idiopathic pulmonary fibrosis. *Eur Respir J* 2021;57:2002559.
91. Lukey PT, Harrison SA, Yang S, Man Y, Holman BF, Rashidnasab A, et al. A randomised, placebo-controlled study of omipalisib (PI3K/mTOR) in idiopathic pulmonary fibrosis. *Eur Respir J* 2019;53:1801992.
92. Maher TM, Simpson JK, Porter JC, Wilson FJ, Chan R, Eames R, et al. A positron emission tomography imaging study to confirm target engagement in the lungs of patients with idiopathic pulmonary fibrosis following a single dose of a novel inhaled $\alpha\text{v}\beta 6$ integrin inhibitor. *Respir Res* 2020;21:75.
93. Richeldi L, Azuma A, Cottin V, Hesselinger C, Stowasser S, Valenzuela C, et al.; 1305-0013 Trial Investigators. Trial of a preferential phosphodiesterase 4B inhibitor for idiopathic pulmonary fibrosis. *N Engl J Med* 2022;386:2178–2187.
94. Désogère P, Montesi SB, Caravan P. Molecular probes for imaging fibrosis and fibrogenesis. *Chemistry* 2019;25:1128–1141.
95. Montesi SB, Izquierdo-Garcia D, Désogère P, Abston E, Liang LL, Digumarthy S, et al. Type I collagen-targeted positron emission tomography imaging in idiopathic pulmonary fibrosis: first-in-human studies. *Am J Respir Crit Care Med* 2019;200:258–261.
96. Kimura RH, Wang L, Shen B, Huo L, Tummers W, Filipp FV, et al. Evaluation of integrin $\alpha\text{v}\beta 6$ cysteine knot PET tracers to detect cancer and idiopathic pulmonary fibrosis. *Nat Commun* 2019;10:4673.
97. Lukey PT, Coello C, Gunn R, Parker C, Wilson FJ, Saleem A, et al. Clinical quantification of the integrin $\alpha\text{v}\beta 6$ by [^{18}F]FB-A20FMDV2 positron emission tomography in healthy and fibrotic human lung (PETAL Study). *Eur J Nucl Med Mol Imaging* 2020;47:967–979.
98. Yang P, Luo Q, Wang X, Fang Q, Fu Z, Li J, et al. Comprehensive analysis of fibroblast activation protein expression in interstitial lung diseases. *Am J Respir Crit Care Med* 2023;207:160–172.
99. Bergmann C, Distler JHW, Treutlein C, Tascilar K, Müller A-T, Atzinger A, et al. ^{68}Ga -FAPi-04 PET-CT for molecular assessment of fibroblast activation and risk evaluation in systemic sclerosis-associated interstitial lung disease: a single-centre, pilot study. *Lancet Rheumatol* 2021;3:e185–e194.
100. Montesi SB, Désogère P, Fuchs BC, Caravan P. Molecular imaging of fibrosis: recent advances and future directions. *J Clin Invest* 2019;129:24–33.
101. Martinez FJ, Yow E, Flaherty KR, Snyder LD, Durheim MT, Wisniewski SR, et al.; CleanUP-IPF Investigators of the Pulmonary Trials Cooperative. Effect of antimicrobial therapy on respiratory hospitalization or death in adults with idiopathic pulmonary fibrosis: the CleanUP-IPF randomized clinical trial. *JAMA* 2021;325:1841–1851.
102. Wilson AM, Clark AB, Cahn T, Chilvers ER, Fraser W, Hammond M, et al.; EME-TIPAC team. Effect of co-trimoxazole (trimethoprim-sulfamethoxazole) vs placebo on death, lung transplant, or hospital admission in patients with moderate and severe idiopathic pulmonary fibrosis: the EME-TIPAC randomized clinical trial. *JAMA* 2020;324:2282–2291.
103. Podolanczuk AJ, Kim JS, Cooper CB, Lasky JA, Murray S, Oldham JM, et al.; PRECISIONS Study Team. Design and rationale for the prospective treatment efficacy in IPF using genotype for NAC selection (PRECISIONS) clinical trial. *BMC Pulm Med* 2022;22:475.
104. Woodcock J, LaVange LM. Master protocols to study multiple therapies, multiple diseases, or both. *N Engl J Med* 2017;377:62–70.
105. Paganoni S, Berry JD, Quintana M, Macklin E, Saville BR, Detry MA, et al.; Healey ALS Platform Trial Study Group. Adaptive platform trials to transform amyotrophic lateral sclerosis therapy development. *Ann Neurol* 2022;91:165–175.
106. Angus DC, Alexander BM, Berry S, Buxton M, Lewis R, Paoloni M, et al.; Adaptive Platform Trials Coalition. Adaptive platform trials: definition, design, conduct and reporting considerations. *Nat Rev Drug Discov* 2019;18:797–807.
107. Berry SM, Connor JT, Lewis RJ. The platform trial: an efficient strategy for evaluating multiple treatments. *JAMA* 2015;313:1619–1620.
108. Raghu G, Brown KK, Bradford WZ, Starko K, Noble PW, Schwartz DA, et al.; Idiopathic Pulmonary Fibrosis Study Group. A placebo-controlled trial of interferon gamma-1b in patients with idiopathic pulmonary fibrosis. *N Engl J Med* 2004;350:125–133.
109. King TE Jr, Brown KK, Raghu G, du Bois RM, Lynch DA, Martinez F, et al. BUILD-3: a randomized, controlled trial of bosentan in idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med* 2011;184:92–99.
110. Raghu G, Collard HR, Anstrom KJ, Flaherty KR, Fleming TR, King TE Jr, et al. Idiopathic pulmonary fibrosis: clinically meaningful primary endpoints in phase 3 clinical trials. *Am J Respir Crit Care Med* 2012;185:1044–1048.

111. King TE Jr, Albera C, Bradford WZ, Costabel U, du Bois RM, Leff JA, *et al.*; Implications for the Design and Execution of Clinical Trials. All-cause mortality rate in patients with idiopathic pulmonary fibrosis. Implications for the design and execution of clinical trials. *Am J Respir Crit Care Med* 2014;189:825–831.
112. Finkelstein DM, Schoenfeld DA. Combining mortality and longitudinal measures in clinical trials. *Stat Med* 1999;18: 1341–1354.
113. Pocock SJ, Ariti CA, Collier TJ, Wang D. The win ratio: a new approach to the analysis of composite endpoints in clinical trials based on clinical priorities. *Eur Heart J* 2012;33:176–182.
114. Russell A-M, Adamali H, Molyneaux PL, Lukey PT, Marshall RP, Renzoni EA, *et al.* Daily home spirometry: an effective tool for detecting progression in idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med* 2016;194:989–997.
115. Lipkovich I, Dmitrienko A, D'Agostino Sr BR. Tutorial in biostatistics: data-driven subgroup identification and analysis in clinical trials. *Stat Med* 2017;36:136–196.
116. Han P, Taylor JMG, Mukherjee B. Integrating information from existing risk prediction models with no model details. *Can J Stat* 2023;51: 355–374.
117. Chen C, Han P, He F. Improving main analysis by borrowing information from auxiliary data. *Stat Med* 2022;41:567–579.
118. Han P, Lawless JF. Empirical likelihood estimation using auxiliary summary information with different covariate distributions. *Stat Sin* 2019;29:1321–1342.
119. Taylor JMG, Choi K, Han P. Data integration: exploiting ratios of parameter estimates from a reduced external model. *Biometrika* 2022; 110:119–134.
120. Zhai Y, Han P. Data integration with oracle use of external information from heterogeneous populations. *J Comput Graph Stat* 2022;31:1001–1012.
121. Kundu P, Tang R, Chatterjee N. Generalized meta-analysis for multiple regression models across studies with disparate covariate information. *Biometrika* 2019;106:567–585.
122. Chatterjee N, Chen Y-H, Maas P, Carroll RJ. Constrained maximum likelihood estimation for model calibration using summary-level information from external big data sources. *J Am Stat Assoc* 2016; 111:107–117.
123. Swigris JJ, Fairclough D. Patient-reported outcomes in idiopathic pulmonary fibrosis research. *Chest* 2012;142:291–297.
124. Sinha A, Patel AS, Siegert RJ, Bajwah S, Maher TM, Renzoni EA, *et al.* The King's Brief Interstitial Lung Disease (KBILD) questionnaire: an updated minimal clinically important difference. *BMJ Open Respir Res* 2019;6:e000363.
125. Swigris JJ, Andrae DA, Churney T, Johnson N, Scholand MB, White ES, *et al.* Development and initial validation analyses of the Living with Idiopathic Pulmonary Fibrosis Questionnaire. *Am J Respir Crit Care Med* 2020;202:1689–1697.
126. Aronson KI, Martin-Schwarze AM, Swigris JJ, Kolenic G, Krishnan JK, Podolanczuk AJ, *et al.*; Pulmonary Fibrosis Foundation. Validity and reliability of the Fatigue Severity Scale in a real-world interstitial lung disease cohort. *Am J Respir Crit Care Med* 2023;208:188–195.