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### **Permalink** https://escholarship.org/uc/item/5151p50g

**Journal** Nucleic Acids Research, 49(W1)

## ISSN

0305-1048

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## **Publication Date**

2021-07-02

## DOI

10.1093/nar/gkab405

Peer reviewed

# Mergeomics 2.0: a web server for multi-omics data integration to elucidate disease networks and predict therapeutics

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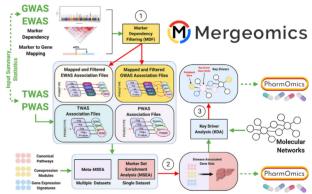
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Received February 28, 2021; Revised April 28, 2021; Editorial Decision April 30, 2021; Accepted May 02, 2021

#### ABSTRACT

The Mergeomics web server is a flexible online tool for multi-omics data integration to derive biological pathways, networks, and key drivers important to disease pathogenesis and is based on the open source Mergeomics R package. The web server takes summary statistics of multi-omics disease association studies (GWAS, EWAS, TWAS, PWAS, etc.) as input and features four functions: Marker Dependency Filtering (MDF) to correct for known dependency between omics markers, Marker Set Enrichment Analysis (MSEA) to detect disease relevant biological processes, Meta-MSEA to examine the consistency of biological processes informed by various omics datasets, and Key Driver Analysis (KDA) to identify essential regulators of disease-associated pathways and networks. The web server has been extensively updated and streamlined in version 2.0 including an overhauled user interface, improved tutorials and results interpretation for each analytical step, inclusion of numerous disease GWAS, functional genomics datasets, and molecular networks to allow for comprehensive omics integrations, increased functionality to decrease user workload, and increased flexibility to cater to user-specific needs. Finally, we have incorporated our newly developed drug repositioning pipeline PharmOmics for prediction of potential drugs targeting disease processes that were identified by Mergeomics. Mergeomics is freely accessible at <a href="http://mergeomics.research.idre.ucla.edu">http://mergeomics.research.idre.ucla.edu</a> and does not require login.

#### **GRAPHICAL ABSTRACT**



#### INTRODUCTION

The advent of omics technologies has made significant strides in unveiling various disease-associated genetic and epigenetic variants, genes, proteins and metabolites. The ever-growing source of multi-omics datasets available including genomics, epigenomics, transcriptomics, proteomics and metabolomics now presents a new challenge of integrating these different data types for more meaningful and holistic interpretation of complex diseases. To conduct a comprehensive investigation of disease pathogenesis, we

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must consider multiple omics layers that contribute to biological complexity (1). The computational pipeline Mergeomics was developed to meet the need for multi-omics integration and functional interpretation to obtain mechanistic understanding. Mergeomics provides flexibility to incorporate the full spectrum of summary statistics (not just top hits) of individual layers of omics or multi-omics data simultaneously along with diverse functional genomics data across data types, studies and species. As such, genomewide association studies (GWAS) as well as epigenome-(EWAS), transcriptome- (TWAS), proteome- (PWAS) and metabolome-wide association studies (MWAS) can all be accommodated.

The development of our Mergeomics tool follows the philosophy of utilizing a systems biology approach to unravel the complex interactions across molecular domains as well as cell types, tissues and organ systems that occur in disease. In particular, we are guided by the omnigenic disease model (2), which states that a large proportion of the genome likely contributes to disease pathogenesis through molecular interactions both within and between tissues. Utilizing this data-driven analysis considering the interactions among different omics layers and tissue contexts will uncover global maps to identify critical targets in disease pathogenesis, which can be followed by experimental approaches to investigate the detailed events that occur through the predicted molecules or pathways.

With the abundance of omics data available, it is unsurprising that various tools or methods have been developed to better integrate and interpret these datasets (3-5). These tools can be broadly categorized into two application categories: multi-omics biomarker predictions of diseases or subtypes (i.e. uncovering correlative or predictive but not necessarily disease-causing features) or mechanistic understanding of disease pathogenesis (i.e. regulators, molecular interactions and processes involved in disease development). Mergeomics focuses on mechanistic modeling but not predictive modeling. In terms of approaches, fusion (such as PFA (6), SNF (7), PSDF (8)), Bayesian (e.g. iCluster (9), PSDF (8), BCC (10)), correlation, multivariate (e.g. MFA (11), IntegrOmics (12), MixOmics (13)), pathway and network methods (PARADIGM (14), SNF (7), iOmicsPASS (15), MiBiOmics (16), Lemon-Tree (17), PaintOmics (18), NetICS (19), Metascape (20)) have been implemented (3-5). Mergeomics falls within the network method category that mainly focuses on understanding disease pathogenesis through uncovering multiple molecular targets within biological processes important to disease. The benefit of a network approach over other integrative options is its ability to provide biological interpretability, which is reliant not on the identification of latent structures through mathematical deconvolution but on the utilization of prior information based on molecular interactions, which can help provide clear targetable options (e.g. genes) in disease. Compared to other tools, Mergeomics not only accommodates diverse data types (GWAS, EWAS, TWAS, PWAS, MWAS) from different sources, studies, or species for a given disease, but also considers relationships between omics layers through functional genomics such as expression quantitative trait loci (eQTLs), molecular pathways, and tissue-specific gene regulatory networks to derive disease networks and predict therapeutics. Mergeomics also uses full summary statistics, not raw data or lists of top associations, as input, thereby reducing the need for raw data processing and harmonization and for pre-determining a specific cutoff to call for significant markers. Mergeomics has the ability to conduct pathway analysis and model gene regulatory networks, protein-protein interaction networks, and transcription factor networks in order to predict and visualize network regulators of disease. These unique features help maximize the utility of existing datasets and overcome limitations of other tools which utilize a narrower range of multi-omics data sources, do not provide mechanistic interpretations, or require programming skills with no intuitive web server for ease of use.

Since the release of the open source Mergeomics R package (https://bioconductor.org/packages/release/bioc/ html/Mergeomics.html) (21) and web server in 2016 (22), this tool has been used to model a diverse set of diseases including cardiometabolic disorders such as non-alcoholic fatty liver disease (23), cardiovascular disease (24-26) and type 2 diabetes (27), autoimmunity including psoriasis (28) and rheumatoid arthritis (29), alcohol dependence (30), brain injury (31), Sjogren's syndrome (32) and environmental contributions to disease (33-35). Importantly, multiple validations of molecular predictions from Mergeomics with in silico, in vitro and in vivo studies highlight the validity and causal nature of the disease network predictions (23,27–28,31,35–40). Due to increasing demand for multi-omics integration and interpretation from scientists with different areas of expertise, we have implemented major revisions and improvements on the Mergeomics web server. Specifically, we have redesigned the user interface, simplified workflows, offered detailed tutorials and case studies, and provided more datasets and network models for utilization. The Mergeomics 2.0 web server offers the scientific community much-improved accessibility to our pipeline, caters to each user's specific goals in multi-omics studies, and addresses a broad range of biological questions, particularly emphasizing a mechanistic understanding of disease pathogenesis and prediction of potential therapeutics based on mechanistic understanding.

#### OVERVIEW AND UPDATES ON THE CORE FUNC-TIONS OF MERGEOMICS

#### **Overview of core functions**

Mergeomics 2.0 features four core functions as previously implemented in version 1.0 with an addition of a new function. First, we provide a preprocessing tool, Marker Dependency Filtering (MDF) to remove omics marker redundancies such as linkage disequilibrium (LD) between single nucleotide polymorphisms (SNPs). Second, Marker Set Enrichment Analysis (MSEA) is used to identify omicsinformed disease processes through the integrations of omics markers such as SNPs with functional genomics, canonical pathways, or co-expression networks. Third, Meta-MSEA runs MSEA on multiple datasets and conducts pathway/network level meta-analysis to retrieve consistent disease processes informed across datasets. Fourth, Key Driver Analysis (KDA) pinpoints network regulators of disease processes based on the topology of biological networks. In Mergeomics 2.0, we added a new functional module called PharmOmics, which takes as input multi-omicsinformed disease pathways or networks from Mergeomics to match with drug signatures to predict potential therapeutic drugs.

#### **Introduction of PharmOmics into Mergeomics 2.0**

We have recently developed a novel species- and tissuespecific network-based drug repositioning tool, PharmOmics, which is based on in vivo molecular studies of drugs (41). PharmOmics is a complementary drug repositioning tool to other existing tools, such as CMap (42)and LINCS L1000 (43), which are mostly based on in vitro cell line data. We provide two drug repositioning methods: network-based drug repositioning and gene overlapbased drug repositioning. Network-based drug repositioning ranks drugs based on the degree of connectivity of genes influenced by drug treatments to disease gene signatures in a given gene network model (44). Gene overlap-based drug repositioning is based on the degree of direct overlap between drug genes and disease genes. Users can directly input their disease pathway results from MSEA (genes from disease pathways are used as input) or KDA (genes from the disease network or significant key drivers (KDs) are used as input). For both MSEA and KDA, specific gene sets can be input into drug repositioning for a more refined analysis. As PharmOmics is based on gene expression studies, inputs are limited to genes or proteins. Users can also input their genes of interest into PharmOmics for drug repositioning analysis without running any other functions in Mergeomics.

#### Flexible workflows using the core functions

Each of the main functions of Mergeomics described above can be utilized as a standalone analysis tool or can be combined into a multi-step workflow with several different cases as portrayed in Figure 1. There are four cases or starting points that a user has the option to select. In case one, the user has one GWAS dataset and is prompted first to run MDF where they provide their association dataset, mapping data (e.g. SNP to gene), and marker dependency data (LD in the case of GWAS) to retrieve corrected SNP associations and mapping files. The MDF step is optional if the user does not wish to correct for LD, although we highly recommend this correction to avoid statistical artefacts due to LD. These results along with a gene set are fed into MSEA to uncover disease-associated pathways, which can be further analyzed in KDA to identify key regulators or PharmOmics for drug repositioning. In case two, the user has EWAS, TWAS, PWAS or MWAS data, and they are led to MSEA, where MDF and marker mapping are optional. As in the GWAS path, results from MSEA can be carried to KDA or PharmOmics. In case three, the user has multiple omics datasets and utilizes Meta-MSEA, which will run MSEA on each dataset and then conduct a meta-analysis across datasets to retrieve consistent biological processes, which can be input into PharmOmics or KDA. Finally, in case four, the user has a gene set and network of interest and can directly run KDA, which will provide KD genes and a

subnetwork visualization of the top KDs, and the KDs or subnetwork can be input into PharmOmics to predict drugs.

#### Update on Marker Dependency Filtering (MDF)

MDF prepares input files for MSEA by correcting for dependency between omics markers and is an optional function. This preprocessing step is most commonly used for GWAS data to correct for LD between SNPs and filter out redundant SNPs, which is critical for removing redundant association signals that can result in statistical and biological artefacts in downstream analysis. Another purpose of MDF is to link the SNPs to potential downstream genes based on functional evidence, such as tissue-specific eOTLs. Correcting for dependency between other omics markers is currently seldom used. However, this feature can be utilized to correct for dependency between other types of markers (methylation sites, transcripts, etc.), if desired. MDF uses as input an association file which details markers (e.g. SNPs) and their disease association strengths (e.g.  $-\log_{10} P$ -values or effect size, note that *P*-values are prohibited as MDF ranks larger values as stronger association strength, which is opposite of *P*-values), a mapping file used for marker to gene mapping (e.g. SNPs are mapped to genes to be enriched for gene sets), and a marker dependency file indicating the dependency between markers (e.g. LD between SNPs, to remove redundant markers) (Figure 2). The resulting corrected association and mapping files are then used as input to MSEA. MDF also allows for the selection of a top percentage of markers (50% or 25% recommended) to be considered in the analysis which reduces noise from low signal markers.

Updates to MDF include an increased number of marker to gene mapping options such as the addition of all available tissue-specific Genotype-Tissue Expression project (GTEx) (45) cis-eQTLs and splicing QTLs (cis-sQTLs) (Table 1), the ability to combine up to five mapping options, and the inclusion of LD files for all 26 populations from 1000 Genomes (1000G) (46) and methylation disequilibrium from EWAS software 2.0 (47). For analysis starting from GWAS data, MDF is a default preprocessing step, but we have included the option to skip MDF. For analytical paths starting from other omics data, users have the option to add MDF if needed.

#### Update on Marker Set Enrichment Analysis (MSEA)

In MSEA, full summary statistics of omics markers such as SNPs from GWAS, epigenetic sites from EWAS, genes from TWAS, proteins from PWAS, or metabolites from MWAS and their disease association values are taken as input and are integrated with functional genomics, canonical pathways, or co-expression networks to retrieve diseaseassociated pathways and networks. MSEA calculates and summarizes enrichment of disease/trait omics markers in sets of functionally related genes, such as canonical pathways and co-expression networks, across a range of statistical cutoffs in the full summary statistics using a chi-squarelike statistic and then uses permutation to determine statistical *P*-values for the enrichment. We emphasize the importance to provide the association strength of the given

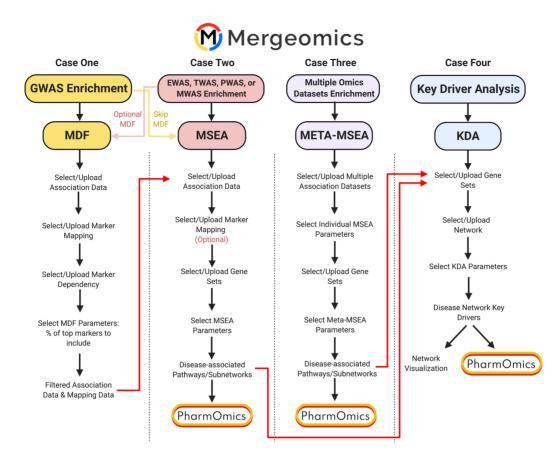


Figure 1. Workflow of Mergeomics. We provide four options on the web server to tailor to the user's data type. Case One: Individual GWAS analysis. For GWAS datasets we advise utilizing the MDF function; however, we also provide the ability to skip MDF and directly run MSEA and follow the workflow to PharmOmics or KDA. Case Two: Individual EWAS, TWAS, PWAS or MWAS analysis. In this case, we directly start at MSEA without MDF; however, we also provide the ability to utilize the MDF function if needed. From here the user can feed the MSEA results into PharmOmics or KDA. Case Three: Multi-omics analysis. If the user has multiple omics of the same type (e.g. two GWAS) or different types (e.g. TWAS and EWAS), they can utilize the MDF function if needed. From here the intervent types (e.g. TWAS and EWAS), they can utilize the MDF function of the same type (e.g. two GWAS) or different types (e.g. TWAS and EWAS), they can utilize the MEA-MSEA function to derive disease-associated pathways and can input their results into PharmOmics or KDA. Case Four: A gene list(s) to run KDA. The user in this case can upload their gene sets of interest and upload or select a network to derive KD genes and visualize top KD subnetworks. The disease subnetwork or significant KDs can be fed into PharmOmics for drug repositioning.

marker wherein a larger number reflects greater association such as  $-\log_{10} P$ -values or effect size to avoid incorrect downstream analysis and interpretability.

MSEA is able to analyze diverse data types, and each has different considerations of inputs which was partly described in the above MDF section (Figure 2). The output from MSEA can be interpreted as omics-informed disease pathways or networks. If GWAS is used, MSEA results can imply causal disease processes since GWAS carries causal inference. For other omics data, the MSEA results can only be interpreted as disease-associated processes but may or may not be causal. Considering GWAS along with other omics data, in our opinion, is a useful way to identify causal genes and processes. We also advise the user to take care in their interpretation of the names or annotations of pathways deemed to be significant (FDR < 0.05) as some can be misleading. Attention to the genes enriched in a given pathway derived from the input dataset should be checked in the gene details output file to confirm whether the pathway name is indeed appropriate as the genes may be more suitable or representative of another biological process. A user can conclude the analysis with results from MSEA or use the MSEA results as input to KDA with a user-defined statistical cutoff to identify network KDs of the disease processes based on molecular network topology.

In Mergeomics 2.0, we added the ability to use diseaseassociated gene sets derived from MSEA as input to PharmOmics for drug repositioning analysis, selecting either specific gene sets or by false discovery rate (FDR) or *P*value threshold, to pinpoint drugs whose gene signatures align with those of the disease-associated gene sets identified by MSEA.

#### Update on Meta-MSEA

Meta-MSEA allows for integration of multiple datasets of the same omics type (e.g. two or more GWAS datasets) or multiple omics types (e.g. GWAS, EWAS, TWAS) and runs MSEA for each omics dataset followed by a meta-analysis. This integration reveals consistencies and differences in biological perturbation across different omics types or different studies of the same omics type.

In Mergeomics 2.0, we improved the guidance of running Meta-MSEA in regard to the differences in preprocessing of the different types of omics data. In addition, we have increased the flexibility of this analysis to allow for spe-

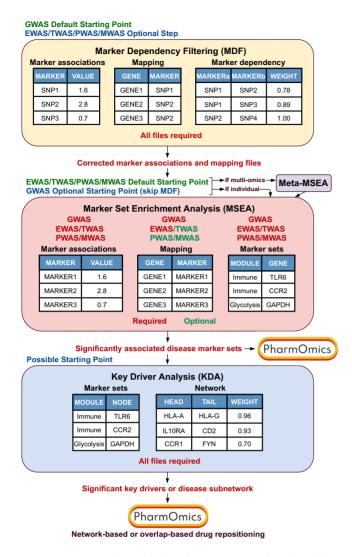


Figure 2. Mergeomics pipeline inputs. MDF is the default starting point for GWAS analysis and is an optional step for EWAS/TWAS/PWAS/MWAS. MDF requires marker-disease associations, a marker-gene mapping file, and a marker dependency file. Users with GWAS data can also skip MDF and run MSEA directly. MDF produces corrected marker-disease associations and marker-gene mapping files containing independent markers that are used for MSEA. For MSEA, required files for all datasets are the marker-disease associations and marker sets (pathway/modules). The marker to gene mapping file is required for GWAS and EWAS and optional for MWAS, TWAS and PWAS. Disease-associated marker sets from MSEA can be fed into KDA, which requires gene sets and a network. KDA can also be a starting point of analysis. Disease-associated gene sets from MSEA or KDs and disease subnetwork from KDA can be fed into PharmOmics for drug repositioning.

cific inputs and parameters for each association data. After each individual omics dataset is added, the user will be able to review which datasets have been successfully uploaded and their individual MSEA parameters with the option to add additional datasets or delete certain datasets, providing an easy way to track all the different inputs. As in the results generated from the individual MSEA, significantly associated gene sets from Meta-MSEA can be used as input to KDA or PharmOmics drug repositioning. We have also implemented user-defined individual MSEA FDR cutoffs to KDA in that the disease-associated pathways must pass all individual MSEA FDR cutoffs as well as the meta-FDR to be used in KDA, allowing the user to focus on the most consistent and robust disease processes across different datasets. In addition, we now provide heterogeneity statistics from Cochran's Q test to indicate the variability between datasets.

#### Update on Key Driver Analysis (KDA)

KDA identifies essential regulators of disease-associated pathways and networks, which are then visualized in the web browser using Cytoscape.js (Figure 3). KDA results can also be downloaded as network files ready to be used on Cytoscape Desktop for further customization of the network visualization. A Chi-square-like statistic,  $\chi = \frac{O-E}{\sqrt{E}-\kappa}$ , is used to identify genes (KDs) that are connected to a significantly larger number of disease-associated genes than what is expected by random chance. O and E represent the observed and expected numbers of disease-associated genes in a hub subnetwork, and E is estimated by  $\frac{N_k N_p}{N_l}$ where  $N_p$  is the disease gene set size,  $N_k$  is the hub degree, and N is the full network order. KDs represent prioritized disease regulatory genes based on network topology. In numerous recent applications of Mergeomics, top KDs have been shown to be causal for diseases based on experimental evidence (23, 27, 36), thereby supporting their importance. KDA can be utilized as a follow up analysis to MSEA or Meta-MSEA, and it can also be used as an independent analysis using a gene list of interest and a given network as inputs. For instance, the user can upload a list of curated disease genes and choose or upload a relevant network to run KDA to identify how the disease genes interact in the network and whether there are key hub nodes in the network that regulate the disease genes.

In Mergeomics 2.0, we added the ability to visualize input gene overlap with a given network, if any, in the case that no KDs were found. The user can therefore be better informed on the reason for the lack of KD hits based on the distribution and connectivity of the input genes in the network. If few input genes are in the network or the input genes are widely dispersed in the network, KDs may not be identified. We have additionally increased the number of sample tissuespecific networks (Table 1). As we have done similarly with MSEA and Meta-MSEA, disease subnetworks or significant KDs from KDA can be used directly for PharmOmics drug repositioning, and users can further customize which processes in the subnetwork are used in drug repositioning for a more focused analysis.

#### DATA AND SAMPLE INPUT UPDATES

We have significantly augmented the amount of Mergeomics-ready sample files with commonly used datasets and will continue to actively update sample files to enrich data resources on a monthly basis.

In Mergeomics 2.0, we include over 20 GWAS datasets from a broader range of diseases from metabolic syndrome

Table 1.	Sample resources	on Mergeomics web serve	er. Complete list in Supplementa	ry Tables S1–S4

General data category	Data type	Specifics	Citation
Association data	GWAS	Alzheimer's disease	(71)
		Attention deficit hyperactivity disorder	(72)
		Alcohol dependence	(73)
		Body mass index	(74)
		Breast cancer	(75)
		Coronary artery disease	(76)
		Fasting glucose	(77)
		Heart failure	(77)
		High density lipoproteins (HDL)	(78)
		Low density lipoproteins (LDL)	(79)
		Major depressive disorder	(80)
		Parental lifespan	(81)
		Parkinson's disease	(82)
		Psoriasis	(83)
		Severe illness in COVID-19	(84)
		Schizophrenia	(85)
		Stroke	(86)
		Systemic lupus erythematosus	(87)
		Type 2 diabetes	(88)
		Total cholesterol	(79)
		Triglycerides	(79)
	EWAS	Birth weight	(89)
		Maternal anxiety	(90)
		Social communication	(91)
		Psoriasis	(62,63)
Marker mapping	Chromosomal distance	10kb, 20kb, 50kb	(46)
Warker mapping	Regulome	RegulomeDB (ENCODE)	(92)
	eQTL	49 tissue types	(45)
	sQTL	49 tissue types	(45)
Marker dependency	Linkage disequilibrium	26 populations at $r^2 > 0.5$ and $>0.7$	(46)
	Methylation disequilibrium	$r^2 > 0.5$	(47)
Marker sets	Canonical (knowledge based)	KEGG	(50)
		Reactome	(51)
		BioCarta	(52)
		MSigDB	(49)
		GO	(53)
		BioPlanet	(55)
		WikiPathways	(54)
	Data-driven (co-expression)	24 tissue specific modules	(45,56–57)
		(WGCNA/MEGENA)	(50.02.00)
Networks	Gene regulatory human and mouse composite (Bayesian)	Adipose, blood, brain, kidney, liver, muscle	(58,93–98)
	Gene regulatory (GIANT)	Adipose, blood, brain, kidney, liver, muscle	(61)
	Protein-protein interaction	STRING	(59)
	Transcription factor-target (FANTOM5)	Adipose, blood, brain, kidney, liver, muscle	(60)

to psychiatric disorders (Table 1; detailed data sources and links in Supplementary Table S1). For omics dependency filtering options, we have added the full array of LD data from 26 human populations studied in 1000G (46) with LD above 0.5 and 0.7 for SNP filtering to remove redundant SNPs in high LD and have also provided an example methylation disequilibrium data file for correction of EWAS data. For SNP to gene mapping options, we have added all tissue-specific cis-eQTL and cis-sQTL mapping files from the GTEx version 8 (q-value < 0.05) (45), which inform on the SNPs associated with gene expression level changes (eQTL) or differential splicing (sQTL). In addition, we offer ENCODE regulatory gene mapping (48) and various chromosomal location-based mapping options (Table 1; Supplementary Table S2). Moreover, we have increased the number of curated pathways from version 1 to include all gene sets from Molecular Signatures Database (MSigDB) (49) such as KEGG (50), Reactome (51), Biocarta (52) canonical pathways, chemical and genetic perturbation, microRNA and transcription factor targets, and cell type marker signatures, Gene Ontology (53), Wikipathways (54) and Bioplanet (55), among others (Table 1; Supplementary Table S3). To complement knowledge-based pathways, we include our data-driven tissue-specific co-expression network modules utilizing GTEx transcriptome datasets and coexpression network construction tools MEGENA (56) and WGCNA (57) (Table 1; details of data sources, methods, and parameters used to construct networks in Supplementary Table S3). Finally, we have constructed tissue-specific Bayesian gene regulatory networks (58) and include them as sample networks on the web server. We also provide human protein-protein interaction networks (59), transcription factor networks (60) and GIANT networks (61) (Table 1; Supplementary Table S4). Sample files are available to download from our sample resources page (http://mergeomics. research.idre.ucla.edu/samplefiles.php), and further clarification on correct formatting of input data is detailed on the web server and in Figure 2.

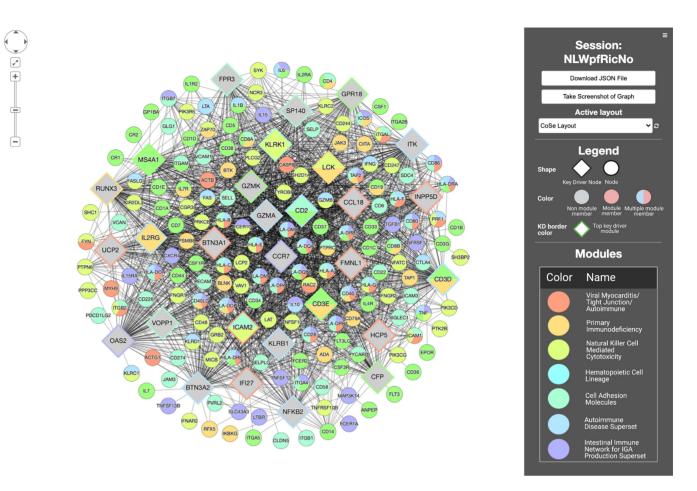


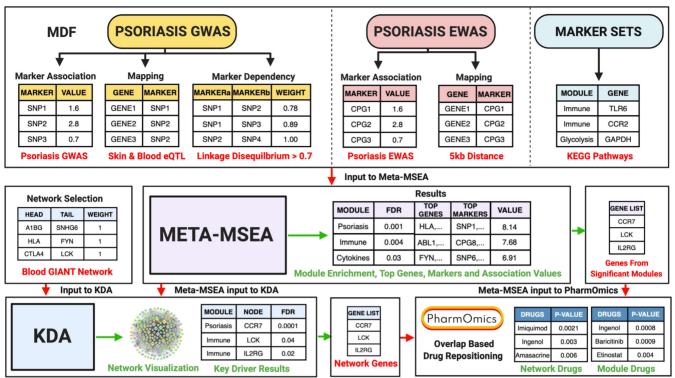
Figure 3. Top KDs network visualization. Screenshot of the in-browser interactive network visualization (using Cytoscape.js) directed from the KDA results page. The colors of the nodes represent member genes of a disease-associated pathway. The diamond shaped nodes represent KD genes, where the border color represents the top pathway that is regulated by the KD. If a node has multiple colors, it is part of two or more disease-associated pathways, and if a node is grey, it does not belong to the disease pathways (non-member genes) but is present in the input network.

#### **GENERAL UPDATES**

We have completely redesigned the user interface for a much more intuitive guidance of the use of the pipeline for different omics data types. To start the pipeline, users are presented with four workflow options in regard to their data: (i) GWAS, (ii) EWAS, TWAS, PWAS or MWAS, (iii) multiple of the same or different types of omics data and (iv) a gene set list (user can run KDA or PharmOmics). The separation of GWAS from other omics datasets is for the additional need to correct for LD and link SNPs to candidate genes through MDF, which is not required or is optional for other omics datasets. For EWAS, a marker to gene mapping file is required if the user uploads epigenetic markers such as CpG probes. For MWAS, a metabolite to gene mapping file is optional but not required if the user uses metabolite sets as the marker sets to be tested. Marker mapping is not needed for TWAS and PWAS as the markers (genes and proteins) match the gene sets. This workflow design clearly delineates what is needed for each specific data type, which is more intuitive for the user. We have also improved the fluidity and presentation of the pipeline workflow as each collapsible step appears below the previous in a vertical format so that the user can revisit input files, parameters, and results of previous steps in the pipeline and choose to rerun a step at any point in the pipeline. A workflow map with navigation links is also generated on the left sidebar to help visualize the steps taken and downstream paths.

We have improved the system that allows users to return to their session where results of analyses can be revisited or continued onto the next step using a unique tracking ID number that is valid for up to 48 hours after the start of their session. The user can also choose to have their results emailed upon completion of the analysis, which is not mandatory but is recommended because the tracking ID allows the user to reload their session and retrieve completed jobs in case a crash occurs. Because later steps of the pipeline, KDA and PharmOmics, can be run independently, downloadable result files from MSEA and KDA can be uploaded directly to the desired next step in the analysis (e.g. MSEA to KDA/PharmOmics or KDA to PharmOmics).

In addition, we have improved case-specific responsiveness of the web server to better inform the user such as errorchecking of user uploaded files to ensure the file is formatted correctly and providing feedback on user results such as whether the results are substantial enough to be used in the next step of the analysis. Across all applications of Merge-



#### **MULTIPLE OMICS DATA ENRICHMENT**

**Figure 4.** Meta-MSEA use case study overview. To showcase the function and output of the web server, we utilized multiple human psoriasis GWAS and EWAS data and ran the multiple omics data workflow (Case 3 in Figure 1, Meta-MSEA). Firstly, we uploaded the psoriasis GWAS data, mapped the SNPs to genes using a combined skin and blood eQTL file, and filtered for LD > 0.7 to remove redundant SNPs in LD. Next, we uploaded our psoriasis EWAS association datasets and mapped the CpG sites to genes based on a 5 kb distance. Finally, we uploaded KEGG pathways with a psoriasis control set. Pathway enrichment results are produced, and each pathway's top genes, markers, and corresponding association values are displayed. Psoriasis-associated pathways are used as input into KDA as well as PharmOmics drug repositioning (using genes from significant pathways/modules). In the KDA, along with the Meta-MSEA input, we chose the blood GIANT network option and ran the KDA providing KD results and visualization (Figure 3) and additionally utilized the network genes as an input into PharmOmics. Finally, two sets of drug repositioning results were produced using gene overlap-based drug repositioning in PharmOmics: one based on the genes of significant pathways from the Meta-MSEA results and the other based on the KDA subnetwork genes.

omics 2.0 we have provided an improved review of analysis inputs and parameters and new interactive tables with pagination, sorting, and search features (Figure 5). We also implemented real-time runtime analysis output and progress updates, and this job log including any errors that occurred is available for download at the conclusion of the analysis. Finally, we have improved multi-device usage including on tablets and phones such that it can be appropriately viewed on different screen sizes. We further improved the tutorial to explain input file preparation, parameter setting, and the underlying methods of each computational function and provide video tutorials to demonstrate the different pipeline options.

#### USE CASE: IDENTIFYING PATHOGENIC PATHWAYS AND NETWORKS FOR PSORIASIS BASED ON MULTI-OMICS DATA

The use case described here utilizes publicly available GWAS and EWAS data to perform Meta-MSEA and subsequently KDA to find pathogenic pathways and regulators of psoriasis (Figure 4). All data used in this example are provided as sample data on the web server which can

be downloaded (http://mergeomics.research.idre.ucla.edu/ samplefiles.php). GWAS of psoriasis was obtained from dbGAP database (www.ncbi.nlm.nih.gov/gap) with accession phs000019.v1.p1, and two EWAS of psoriasis were obtained from GEO (GSE31835 and GSE63315) (62,63). For preprocessing of the GWAS data, we use the top 50% of SNPs ranked by -log<sub>10</sub> P-value and correct for LD between SNPs using MDF with the psoriasis GWAS summary statistics as the marker associations, combined skin and blood eQTLs as the SNP to gene mapping, and the 1000G CEU LD structure containing SNPs with  $r^2 > 0.7$  as the marker dependency file. For the EWAS data, CpG sites are mapped to adjacent genes within 5 kb. Next, we chose canonical pathways from the KEGG database and a positive control gene set from the NHGRI-EBI GWAS catalog (64) for psoriasis as the pathways or marker sets to be examined. We ran Meta-MSEA across the GWAS and two EWAS datasets. At the conclusion of Meta-MSEA, a set of results files and a summary table display are generated on the webpage detailing the pathways ranked by meta P-value and their top markers and corresponding mapped genes (Figure 5A; Supplementary Table S5).

SHOW 5 C ENTRIES									S	EARCH:		
		MSEA: P-	MSEA:	4554						_N	lodule T	on
Module ID			FDR <sup>†↓</sup>	Modu	lle Top Gene		Mod	ule Top M	larker <sup>†↓</sup>		ciation	
Psoriasis GWAS Catalog Control Set		2.87e-11	0.00%	BIPSORS	CDSN,HLA- 51C1 MICA MICB N		rs12191877 rs12191877 rs		191877 rs	s  50.69 50.69 50.69 50		
Cytokine Cytokine Receptor Interaction		1.06e-10	0.00%	CSF3 C)	KCR1 TGFBR2 EGFI		cg21432842 cg21004129		1004129 c	c  13.39 9.94 8.24 8.2		8.2
Graft Versus Host Disease		2.21e-10	0.00%	CDSN,HLA-BIHCG27,HLA- C,POUSF3]HLA- DRB5]HLA- DMAJAGER,HLA- DQA1,HLA-DQB1,HLA- DQB1-AS1,HLA- DQB2,HLA-DRB1,HLA- DQB2,HLA-DRB1,HLA- DRB6J		rs12191877 rs2524163 rs8			50.69 19.27 16.96 14			
Natural Killer Cell Mediated Cytotoxicity		2.27e-10	0.00%	CDSN,HLA- B MICB MICA HCG27,HLA- C,POU5F1 CD48		rs12191877 rs12191877 rs			50.69 50.69 50.69 1			
Hematopoietic Cell Lineage		7.31e-10	0.00%	HLA- DR55 C5F3 AGER,HLA- DQA1,HLA-DQB1,HLA- DQB1-A51,HLA- DQB2,HLA-DR51,HLA- DR86 HLA-DRA1CD55		rs8365 cg21432842 rs9266		[16.96]13.39]12.05]1				
Showing 1 to 5 of 187 entries								Previous	1 2	3 4	5	38
SHOW 10 CENTRIES										SEARCH	4:	
Merge Module ID		Key Driver Node		alue <sup>†↓</sup>			Module Genes	KE Subnet Gen		Module an Subnetwor Overlap		Fo rich
Autoimmune Disease Sup	erset	CD2	2.3	2e-59	6.98e-56		52	461		27		12
Viral Myocarditis/Tight Junction/Autoimmune		ICAM2	3.5	8e-59	6.98e-56		65	35	393			10
Autoimmune Disease Sup	erset	NFKB2	2.1	7e-51	2.82e-48		52	392		21		11
Viral Myocarditis/Tight Junction/	Autoimmune	BTN3A1	4.5	9e-50	4.47e-47		65	65 316		23		12
Autoimmune Disease Superset		GZMA	1.9	3e-49	1.50e-46	1.50e-46		52 35		56 25		14
Viral Myocarditis/Tight Junction/		INPP5D		2e-48	1.51e-45		65 3		29		22	
Autoimmune Disease Sup		IL2RG		3e-47	1.26e-44		52	40		24		10
Intestinal Immune Network for IGA Production Superset		OA52	5.8	0e-45	1.26e-42		48	48	39	24		11
Intestinal Immune Network for IGA Production Superset		IL2RG	1.9	2e-44	3.56e-42		48	44	54	24		11
Hematopoletic Cell Line	300	GZMK	35	9e-36	1.70e-34		84	28	14	27		12
Showing 1 to 10 of 3,894 entries	-0-								Previous 1	2 3 4	15_	33
_												
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Database	Method <sup>†↓</sup>	Drug TJ	Species <sup>⊤↓</sup>	Cell Line	Study TI PharmOmics	Dos		uration <sup>⊤↓</sup>	Score T	Ratio	value <sup>T↓</sup>	
GSE61551, GSE61552, GSE61554, GSE61555, GSE45514, GSE45551	Characteristic direction	inhibitor	Mus musculus	integument	meta	me					2.1400E- 20	
L1000_CPC006_HA1E_6H	Characteristic direction	Ingenol	Homo sapiens	HA1E	In Vitro	10 ι	IM	6hr	0.042	10.901	1.0300E- 20	
GSE61551, GSE61552, GSE61554, GSE61555	Characteristic direction	Baricitinib	Mus musculus	integument	PharmOmics meta	Pharmo		harmOmics meta	0.041	4.122	1.1500E- 20	
GSE80028	Characteristic direction	Imiquimod	Homo sapiens	integument	PharmOmics meta	Pharma		harmOmics meta	0.041	14.631	1.6700E- 20	
GSE80028	Characteristic	Toll-like receptor	Homo	integument	PharmOmics	Pharm	Omics F	harmOmics	0.041	14.631	1.6700E-	
GSE61551, GSE61552,	direction Characteristic	agonist Immunosuppressant	sapiens Mus	integument	meta PharmOmics	Pharm		meta harmOmics	0.040	4.055	20 2.2600E-	
GSE61554, GSE61555, GSE51194	direction		musculus	-	meta	me	ta	meta			18	
L1000_CPC011_PC3_24H	Characteristic direction	Pizotifen	Homo sapiens	PC3	In Vitro	10 .	iM	24hr	0.039	11.017	2.1800E- 18	
GSE49872, E-GEOD-6196, GSE61551, GSE61552, GSE61554, GSE61555, GSE49872	Characteristic direction	Anti-inflammatory	Mus musculus	integument	PharmOmics meta	Pharm		harmOmics meta	0.038	3.894	1.2000E- 16	
drugMatrix_Affy	limma	Indomethacin	Rattus norvegicus	kidney	In Vivo	9.6 m	g/kg	5d	0.038	3.216	1.2000E- 08	

**Figure 5.** Output files from Meta-MSEA, KDA, and PharmOmics based on the case study of psoriasis outlined in Figure 4. Tables are interactive with pagination, search, and sort functions. Result files are downloadable from links on the webpage above the output tables (not shown). (A) Example Meta-MSEA output from the psoriasis use case. The table shown details the significance of association of each pathway/module and the top markers and corresponding association strengths that contributed to the module association. There are two additional tables which can be displayed by clicking on the tabs to the right of 'Module Results' at the top. The second table shows the significance and details of merged modules after merging redundant pathways (termed 'Supersets'), and these non-overlapping gene sets are used as input to KDA. The third table shows the individual significance values for each omics dataset included in this Meta-MSEA of one GWAS and two EWAS of psoriasis. (B) Example KDA output from the psoriasis use case. The table shown records the significance of KDs, the pathways/modules that they regulate based on network topology, and details of the local subnetwork such as the number of KD subnetwork genes and number of pathway/module gene overlap with the KD subnetwork. Merged pathways/modules are represented by the term 'Superset', which means they are comprised of multiple redundant (significant gene overlap) pathways. (C) Example PharmOmics drug repositioning output using a gene overlap-based analysis between disease pathways and drug signatures. Gene overlap-based drug repositioning queries, tissue or cell line, whether the study was done *in vitro* or *in vivo*, the dose and time regimen, the Jaccard score, and statistical significance of the drug study including species, tissue or cell line, whether the study was done *in vitro* or *in vivo*.

As shown in Figure 5A, 'Cytokine cytokine receptor interaction', 'Graft versus host disease' and 'Natural killer cell mediated cytotoxicity' were three of the top pathways identified among others. Following Meta-MSEA, KDA was run with default parameters using non-redundant supersets (pathways that were merged due to significant overlap in gene members) significantly associated with psoriasis from Meta-MSEA and a blood GIANT Bayesian gene regulatory network (61) (chosen due to the relevance of the immune system to psoriasis) to identify KDs of the disease related gene sets. At the conclusion of the KDA, a table is produced on the webpage listing the KDs and significance of enrichment of psoriasis-associated gene sets in their network neighborhood (Figure 5B; Supplementary Table S6). For example, *ICAM2* is identified as the KD for the viral myocarditis/tight junction/autoimmune pathway, and CD2 is identified as a KD for the Autoimmune Disease Superset. By default, the top five KDs and their local subnetworks from each gene set is included in the interactive subnetwork visualization in the browser (Figure 3).

With addition of the PharmOmics pipeline to the Mergeomics web server, we ran two drug repositioning analyses: one directly from the MSEA results and the other considering the whole subnetwork derived from the KDA (Figure 5C; Supplementary Table S7). In this case study, we do not consider gene expression direction changes (upregulation or downregulation) in psoriasis and therefore will simply be utilizing genes involved in disease without considering if they are protective or pathogenic; thus, our predicted drug list will contain drugs that can induce as well as drugs that can potentially treat psoriasis. In addition, PharmOmics interrogates all drug signatures regardless of the tissue or species, and the user can choose to focus on the relevant drug studies for their given dataset. For example, we mainly focused on drugs that were studied in integument tissue, due to its relevance to psoriasis. In the top 10 repositioned drugs derived from psoriasis associated gene sets from Meta-MSEA, we find 8/10 to have prior association with a role in psoriasis pathogenesis (Imiquimod (65)) or treatment including broad options suggesting classes of drugs such as anti-inflammatory, immunosuppressant, JAK inhibitors, and anti-rheumatic drugs and more specific options such as Baricitinib (66), Ingenol (67), and Etinostat (68) (Figure 5C; Supplementary Table S7). Similarly, using the psoriasis subnetwork from the KDA highlights Imiquimod and Ingenol within the top 10 drugs, and the remainder of the results are broad categories such as JAK inhibitors, anti-inflammatory drugs, and anti-rheumatic drugs (Supplementary Table S8), each of which are actively being investigated in the treatment of psoriasis (69,70). The predicted drugs can form new hypotheses for experimental testing.

#### **FUTURE DIRECTIONS**

The web server will continue to actively incorporate the most up-to-date public resources including multi-omics association data, functional genomics data such as eQTLs or protein QTLs (pQTLs), knowledge-based pathways, gene co-expression networks, and gene regulatory networks on a monthly basis. We will also include single cell networks when available to understand the gene regulatory connections within a given cell type or between cell types rather than across a whole tissue, which will offer higher resolution molecular mechanisms of disease pathogenesis. Cell type level association data derived from single cell omics studies can be used in the current platform. We will also continue incorporating additional analytical functions into the web server such as different forms of meta-analysis that can be conducted within the Meta-MSEA tool as well as adding new features to better accommodate analysis of data types that are currently not considered or well tested, such as gut microbiome and spatial transcriptomics data.

#### CONCLUSION

Thanks to advancements in technologies, the number of multi-omics data (GWAS, EWAS, TWAS, PWAS and MWAS) increases exponentially. The systems biology approach to interrogate multi-tissue multi-omics data has become a promising method to understand biology in a datadriven way and sheds light on the hidden mechanisms. However, the computational knowledge and skills required to perform such integrative analysis are often considered as a hurdle to many biologists. Therefore, the Mergeomics web server was developed to lower this barrier to enable fellow researchers to dive into multi-omics systems biology. The current update, Mergeomics 2.0, is a versatile webbased tool that provides multi-omics data integration using a pathway- and network-based approach. The improvements we made support a wide range of pre-calculated networks and data for all steps of the pipeline to fulfill a variety of needs and research purposes. In addition, the new user interface presents a more intuitive and flexible environment that greatly improves its ease of use. In addition to a detailed tutorial, each step of the pipeline contains embedded guidance to facilitate the user experience. We believe that the Mergeomics 2.0 and systematics approach applied here will accelerate our understanding of complex diseases and guide therapeutics.

#### DATA AVAILABILITY

Sample resources are available on our sample resources page on the Mergeomics web server (http://mergeomics. research.idre.ucla.edu/samplefiles.php), and the R package for Mergeomics can be found on (https://bioconductor.org/ packages/release/bioc/html/Mergeomics.html).

#### SUPPLEMENTARY DATA

Supplementary Data are available at NAR Online.

#### ACKNOWLEDGEMENTS

We would like to thank Dr Yuqi Zhao for guiding the use case example. We thank Yanning Zuo, Russell Littman, Neil Hsu, Jenny Cheng, I-Hsin Tseng, Caden McQuillen, Yutian Zhao, Zara Saleem, Hengjian Li and Carissa Zhu for their suggestions and testing of the web server.

#### FUNDING

X.Y. is supported by NIH R01 [NS117148, NS111378, DK117850, HL145708, HL147883, HD100298]; Montgomery Blencowe is supported by the American Heart Association Predoctoral Fellowship; Sung-min Ha is supported by the UCLA QCBio Collaboratory Postdoc Fellowship. Funding for open access charge: National Institutes of Health [DK117850, HD100298, HL145708, HL147883, NS111378, NS117148].

Conflict of interest statement. None declared.

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