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Uncovering the Key Role of Distortion in Bioorthogonal Tetrazine Tools That Defy the Reactivity/Stability Trade-Off

Dennis Svatunek,* Martin Wilkovitsch, Lea Hartmann, K. N. Houk, and Hannes Mikula*



ABSTRACT: The tetrazine/trans-cyclooctene ligation stands out from the bioorthogonal toolbox due to its exceptional reaction kinetics, enabling multiple molecular technologies in vitro and in living systems. Highly reactive 2-pyridyl-substituted tetrazines have become state of the art for time-critical processes and selective reactions at very low concentrations. It is widely accepted that the enhanced reactivity of these chemical tools is attributed to the electron-withdrawing effect of the heteroaryl substituent. In contrast, we show that the observed reaction rates are way too high to be explained on this basis. Computational investigation of this phenomenon revealed that distortion of the tetrazine caused by intramolecular repulsive N–N interaction plays a key role in accelerating the cycloaddition step. We show that the limited stability of tetrazines in biological media strongly correlates with the electron-withdrawing effect of the substituent, while



intramolecular repulsion increases the reactivity without reducing the stability. These fundamental insights reveal thus far overlooked mechanistic aspects that govern the reactivity/stability trade-off for tetrazines in physiologically relevant environments, thereby providing a new strategy that may facilitate the rational design of these bioorthogonal tools.

INTRODUCTION

The inverse electron demand Diels-Alder (IEDDA)-initiated ligation of 1,2,4,5-tetrazines (Tz) with strained dienophiles represents a group of exceptionally fast bioorthogonal reactions.¹⁻³ In particular, trans-cyclooctene (TCO) derivatives⁴⁻⁶ provide several orders of magnitude higher reactivity than other dienophiles such as cyclopropenes' or norbornenes.⁸ In the rate-determining step, the tetrazine first reacts with the *trans*-cyclooctene in a [4 + 2]-cycloaddition reaction to form a bicyclic intermediate that rapidly undergoes cycloreversion to give dihydropyridazines as ligation products (Figure 1a). Due to high reaction rates, its biocompatibility, and versatility, the tetrazine/trans-cyclooctene ligation has found broad applications in many fields, in particular enabling selective chemical reactions in living organisms.^{2,3} Very recently, bioorthogonal chemistry has entered phase 1 clinical trials, with tetrazine/trans-cyclooctene reactions currently being tested in humans, aiming for locally restricted prodrug activation to improve the selectivity of chemotherapeutics.^{9,10}

In recent years, a variety of differently substituted Tz have been used for bioorthogonal reactions and in vivo chemistry, including bis-alkyl-substituted Tz,^{11–15} alkyl-aryl-Tz,^{16–19} mono-alkyl-Tz²⁰ (alkyl-H-Tz) as well as highly reactive bisheteroaryl^{21,22} and mono-aryl^{23–26} derivatives (aryl-H-Tz). These applications have motivated and fueled the development of advanced procedures for the synthesis of Tz scaffolds.^{27–31} In particular, 2-pyridyl-Tz are frequently used despite limited



Figure 1. (a) IEDDA-initiated Tz ligation with **TCO**; (b) 2-pyridyl-,^{2,3,32} 4-pyridyl-,³⁶ and recently introduced²⁹ vinyl ether-Tz.

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stability because of their exceptionally high reactivity. This is commonly attributed to the electron-withdrawing effect of the 2-pyridyl substituent, resulting in a lowered orbital energy of the Tz, thereby accelerating the IEDDA cycloaddition (Figure 1b).^{2,3,32} Assuming that the reactivity is indeed controlled by frontier molecular orbital (FMO) interactions, we hypothesized that 4-pyridyl-substituted Tz are even more reactive than their 2-pyridyl analogues.^{33–35} So far, 4-pyridyl-Tz (Figure 1b) have only rarely been used,³⁶ and there is no comparative data on the IEDDA reaction kinetics of 2-pyridyl- and 4-pyridyl-Tz. In addition, vinyl ether-Tz, recently introduced by Fox et al.,²⁹ have been shown to exhibit increased IEDDA reactivity despite the non-electron-withdrawing nature of the substituent (Figure 1b), while also maintaining high stability. Intrigued by these findings, we speculated about the existence of yet overlooked mechanistic aspects other than FMO interactions that have a crucial effect on the reactivity of Tz.

Here, we present the results of a combined experimental and computational approach, finally revealing the key role of Tz distortion in the bioorthogonal reaction with *trans*-cyclo-octenes. We show that this effect accelerates both 2-pyridyl-and vinyl ether-substituted Tz, thereby providing fundamental insight into the underlying mechanism and a molecular basis to describe the reactivity and stability of these bioorthogonal tools.

RESULTS AND DISCUSSION

First, we aimed to study the influence of aryl substituents on the click reactivity with TCO and therefore selected a series of phenyl (Ph)-, 2-pyridyl (2Pyr)-, 3-pyridyl (3Pyr)-, and 4pyridyl (4Pyr)-substituted Tz (Figure 2a) for initial investigations. Theoretical calculations were performed using density functional theory (DFT) at the ω B97X-D/6-311G-(d,p)-SMD(1,4-dioxane) level of theory, and orbital energies



Figure 2. (a) LUMO+1 orbitals and orbital energies of monosubstituted Tz showing a significantly lower LUMO+1 energy for **4Pyr**; (b) rate constants for the reaction of **Ph**, **2Pyr**, **3Pyr**, and **4Pyr** with **TCO** (1,4-dioxane, 25 °C, n = 6, SD < 1%); (c) measured rate constants vs calculated LUMO+1 energy.

were calculated at the HF/6-311+G(d,p)-SMD(1,4-dioxane)// ω B97X-D/6-311G(d,p)-SMD(1,4-dioxane) level of theory. A detailed description of the computational methods used within this study can be found in the Supporting Information. Energies for the reacting LUMO+1³² of Tz range from 1.29 to 0.89 eV. As expected, **Ph** shows the highest orbital energy, followed by **2Pyr** and **3Pyr**, which were calculated to have an almost equal LUMO+1 level, while **4Pyr** shows the lowest orbital energy (Figure 2a). According to FMO theory, **4Pyr** should thus indeed show an increased IEDDA reactivity compared to **2Pyr**.

To compare these results with the measured reaction rates, we have prepared all selected mono-substituted aryl-Tz using a method recently published by Audebert and co-workers.²⁷ The rate constants for the ligation with **TCO**³⁷ were measured by monitoring the IEDDA reactions in 1,4-dioxane at 25 °C by stopped-flow spectrophotometry. The measured rate constants range from 100 M⁻¹ s⁻¹ for **Ph** to 620 M⁻¹ s⁻¹ for **2Pyr** (Figure 2b). While the reactivity trend for **Ph**, **3Pyr**, and **4Pyr** seems to be governed by FMO interactions, **2Pyr** is significantly more reactive (>3-fold) than expected based on the respective orbital energy (Figure 2c). Qualitatively equivalent results were obtained using the calculated Kohn–Sham orbital energies (see Supporting Information, Figure S1). Hence, the high IEDDA reactivity of **2Pyr** cannot be attributed to the electron-withdrawing effect of the 2-pyridyl substituent.

To investigate the origin of the observed reactivity trend for different pyridyl substituents, a DFT study of the respective reactions was performed, showing good correlation between the calculated free energies of activation (ΔG^{\ddagger}) and the experimental values (Figure S2). The calculated transitionstate geometries revealed a slight asynchronicity with very similar forming bond lengths across all four Tz. However, **2Pyr** shows a slightly more synchronous bond formation (Figure 3a).

For a detailed investigation of the barrier heights, we performed a distortion/interaction analysis (also referred to as the activation/strain model) 38 on all four transition states (Figure 3b) using the auto DIAS software package.³⁹ This energy decomposition method was introduced independently by Houk and Bickelhaupt and has successfully been applied to investigate bioorthogonal cycloadditions.^{32,40-42} In this analysis, the energy of activation (ΔE^{\ddagger}) is dissected into two parts, the distortion energy (ΔE_{dist}), which is needed to distort the isolated reactants into transition-state geometry, and the interaction energy (ΔE_{int}) resulting from bringing the two distorted isolated fragments together. The analysis was performed at the transition state as a defined point for each reaction. Performing the distortion/interaction analysis at the transition state geometry can lead to skewed results if transition states at highly different forming bond lengths are compared.⁴³ However, here the forming bond lengths are very similar in all cases, warranting the comparison at the transition state. To validate these results, an analysis at a consistent geometry with fixed bond lengths for all Tz was conducted, which qualitatively provided the same results (see Supporting Information, Table S2). Interaction energies were calculated to be similar for all studied reactions (within 1.1 kcal mol⁻¹) and do not explain the observed differences in IEDDA reactivity. However, ΔE_{dist} for **2Pyr** is about 1.5 kcal mol⁻¹ lower than that for Ph, 3Pyr, and 4Pyr, demonstrating that the increased reactivity of 2Pyr with TCO is mainly caused by a reduced distortion energy (Figure 3b).



Figure 3. (a) Calculated transition-state geometries and free energies of activation (ΔG^{\ddagger} , kcal mol⁻¹) for the reaction of mono-substituted Tz (Ph, 2Pyr, 3Pyr, and 4Pyr) and TCO; (b) distortion/interaction analysis shows that the high reactivity of 2-pyridyl-substituted Tz results from a reduced distortion energy (ΔE_{dist}).

To explain the lowered ΔE_{dist} for aryl substituents with nitrogen atoms in 2-position, we have focused on the geometries encountered at the transition state. In all cases, the aryl moiety is tilted away from the allylic CH₂ of **TCO**. For **Ph**, **3Pyr**, and **4Pyr**, the dihedral angle is approximately 80°, which we reasoned is due to the steric demand of the allylic CH₂. This hypothesis is in agreement with the investigations of analogous reactions with ethylene (no allylic CH₂), showing dihedral angles of approx. 90° (Figures 4a and S3). However, for **2Pyr**, we observed a much stronger tilt in the transition state for the reaction with **TCO**, with a dihedral angle of 63°, which did not change in the reaction with ethylene. These



Figure 4. (a) Dihedral angle in the transition states for the reaction of **Ph** and **2Pyr** with **TCO** and ethylene (for **3Pyr** and **4Pyr**, see Figure S3); (b) calculated energy profiles for the rotation of the aryl-Tz bond showing that N-N repulsion reduces the stabilization energy for **2Pyr**.

observations demonstrate that it is an intrinsic property of 2pyridyl-substituted Tz rather than forced by steric interactions. In fact, the calculated geometry of 2Pyr revealed that a nitrogen-nitrogen interaction destabilizes the reactant. This interaction becomes apparent when looking at the stabilization energies when going from an orthogonal (i.e., dihedral angle between the two aromatic rings of 90°) conformer in the reactant to the stable, coplanar conformer (Figure 4b). For Ph, 3Pyr, and 4Pyr, this stabilization energy based on the conjugation between the ring systems is approximately 6 kcal mol⁻¹. However, for **2Pyr**, the most stable conformation is not planar, and the stabilization is only 3.5 kcal mol^{-1} due to a repulsive interaction between the pyridyl nitrogen and the vicinal Tz nitrogen. This repulsion counteracts the stabilization due to the conjugation of the aromatic systems, leading to an almost flat energy surface between -30 and $+30^\circ$, with a minimum at a dihedral angle of 12° (Figure 4b). At the transition state, this N-N repulsion can be avoided by the rotation of the substituent, thereby increasing the distance between the interacting nitrogen atoms. The nitrogen lone pairs then point in different directions, further reducing the repulsive interaction. In addition, natural bond orbital analysis revealed (i) a weak intermolecular hydrogen bond between the pyridyl nitrogen of 2Pyr and the vinylic CH of TCO, similar to the interactions previously described for bioorthogonal 1,3dipolar cycloadditions⁴⁴ and (ii) an increased $n_N \rightarrow \sigma_{C-N}^*$ donation of the pyridyl lone pair into the σ^* of the Tz C–N bond.⁴⁵ However, both these weak interactions were found to play a minor or negligible role regarding the observed rotation in the transition state (see Supporting Information, Figure S4).

In summary, our results revealed that the high reactivity of 2pyridyl-Tz cannot be explained by the electron-withdrawing nature of the heteroaryl substituent only but moreover by intramolecular N–N repulsion, finally uncovering the mechanistic key role of distortion in Tz ligations.

To confirm that these findings can be translated to physiological conditions, additional IEDDA reactions were carried out in Dulbecco's phosphate buffered saline (DPBS) at 37 °C. Due to the limited stability of the mono-substituted H-Tz in aqueous solution, we have prepared the respective methyl-Tz MePh, Me2Pyr, Me3Pyr, and Me4Pyr and determined the second-order rate constants for the reaction with water-soluble TCO-PEG₄ (Figure 5a). This TCO



Figure 5. (a) Reaction kinetics of MePh, Me2Pyr, Me3Pyr, and Me4Pyr in buffered aqueous solution (DPBS) at 37 °C using watersoluble TCO-PEG₄ (n = 6, SD < 1%); (b) stability of Tz in cell growth medium at 37 °C (n = 3) revealing accelerated degradation with decreasing LUMO+1 energy as a measure for the increased electron-withdrawing character of the (hetero)aryl substituent, promoting the attack of nucleophiles (Nu).

derivative has been prepared starting from axially configured TCO–OH, a frequently used tag for the design of in vivo chemical tools.^{22,46,47} The observed relative reactivity profile almost exactly matches the data as previously obtained for the reactions of the corresponding aryl-H-Tz with **TCO** in 1,4-dioxane (cf. Figure 2b), with **Me2Pyr** showing the highest rate constant in the reaction with **TCO-PEG**₄ (Figure 5a).

It is generally accepted that increasing the reactivity of Tz by using electron-withdrawing substituents leads to reduced stability in aqueous/biological media due to accelerated attack of nucleophiles causing Tz degradation (Figure 5b).^{1-3,48,49} However, when working with buffered aqueous solutions of the aryl-methyl-Tz, we noticed a significantly faster degradation of Me4Pyr compared to Me2Pyr (as indicated by accelerated fading of the characteristic pink color of Tz), despite the higher reactivity of the 2-pyridyl-Tz. To investigate this observation, MePh, Me2Pyr, Me3Pyr, and Me4Pyr were incubated in full cell growth medium (Dulbecco's modified Eagle's medium, DMEM, incl. 10% fetal bovine serum) at 37 °C, and Tz stability was monitored by spectrophotometry (Figure 5b). Indeed, Me4Pyr degraded much faster (30% intact after 12 h) compared to Me3Pyr and Me2Pyr (approx. 85%) and MePh (>95%). These results correlate well with the decreasing LUMO+1 energy³² of the Tz (Figure 5b), which we used as a measure for the increased electron-withdrawing effect of the (hetero)aryl substituent. This finding is in agreement with the hypothesis of accelerated nucleophilic attack leading to Tz degradation. Me2Pyr is thus not only more reactive than

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predicted by FMO theory but also shows a significantly higher stability than expected based on its IEDDA reactivity.

Based on these key mechanistic insights and considering the mentioned vinyl ether-substituted Tz developed by Fox and co-workers (Figure 1b),²⁹ we hypothesized that the observed unexpectedly high reactivity of these Tz is due to the lowered distortion energies caused by intramolecular O–N repulsion (Figure 6a), analogous to N–N repulsion in 2-pyridyl-



Figure 6. (a) Repulsive O–N interaction increases the reactivity of vinyl ether-Tz; (b) computational analysis revealed a reduced rotational barrier for MVE in comparison to MV and a relatively high LUMO+1 energy of 1.46 eV; (c) optimized transition-state geometries (Tz + TCO) and distortion/interaction analysis confirmed O–N repulsion to be the main reason for the increased reactivity of MVE, as indicated by the calculated values for ΔG^{\ddagger} , ΔE^{\ddagger} , and ΔE_{dist} (kcal mol⁻¹).

substituted Tz. Considering the non-electron-withdrawing character of the vinyl ether moiety, this would moreover explain the high stability of these Tz despite the increased IEDDA reactivity. To confirm our assumption, we performed computational investigations using the Tz structures MVE (methylvinyl ether-Tz) and MV (methylvinyl-Tz). The optimized geometries revealed that an oxygen-nitrogen interaction leads to a reduced rotational barrier of 3.9 kcal mol⁻¹ for MVE (similar to 2Pyr) in comparison to 5.1 kcal mol⁻¹ for MV (Figure 6b). The calculated LUMO+1 energies moreover indicate a non-electron-withdrawing character of both substituents (~1.5 eV in contrast to 0.89 eV, as calculated for 4Pyr). The transition-state geometries for the reaction with TCO (Figure 6c) showed a significantly stronger tilt of the vinyl-Tz bond in the case of the vinyl ether-Tz MVE (dihedral angle of 66°). The calculated distortion energies (ΔE_{dist}) finally confirmed O–N repulsion to play a crucial role regarding the potentially increased reactivity of MVE, as indicated by the calculated free energy of activation (ΔG^{\ddagger}) of $16.0 \text{ kcal mol}^{-1}$.

Encouraged by the computational results, we have prepared the vinyl ether-Tz **MeEVE** and the 3,4-dihydro-2*H*-pyran (DHP)-substituted Tz **MeDHP** and **DHP**₂ (Figure 7a; for details on synthetic procedures, see the Supporting Information), but did not obtain sufficient amounts of pure material of the respective bis(vinyl ether)Tz EVE₂ [3,6-bis(1-



Figure 7. (a) Vinyl ether-Tz **MeEVE**, **MeDHP**, and **DHP**₂ and second-order rate constants for the reaction with **TCO-PEG**₄ in buffered aqueous solution (DPBS) at 37 °C (n = 6, SD < 1%); (b) stability of pyridyl- and vinyl ether-Tz under physiological conditions (full cell growth medium, 37 °C, n = 3, SD < 5%) revealing accelerated degradation of pyridyl-Tz in contrast to the exceptional stability of DHP-substituted Tz; (c) 2-pyridyl substituents increase the IEDDA reactivity of Tz, also leading to limited stability due to the electron-withdrawing effect. In contrast, non-electron-withdrawing vinyl ether substituents such as DHP increase the reactivity via intramolecular repulsion, without sacrificing stability.

ethoxyvinyl)Tz; structure not shown]. Second-order rate constants for the reactions of these Tz with TCO-PEG₄ in DPBS at 37 °C were determined by stopped-flow spectrophotometry. The IEDDA reactivity of **MeEVE** (2750 $M^{-1} s^{-1}$) was shown to match the value measured for **Me4Pyr** (2740 $M^{-1} s^{-1}$). In comparison, the cyclic vinyl ether-Tz **MeDHP** was observed to be less reactive (1820 $M^{-1} s^{-1}$), though still significantly faster than the aryl-Tz **MePh** (990 $M^{-1} s^{-1}$). As expected, we observed a high reactivity of the bisvinyl ether-Tz **DHP**₂ (6450 $M^{-1} s^{-1}$), exceeding the rate constant of **Me2Pyr** (5120 $M^{-1} s^{-1}$) by approx. 25% (Figure 7a). These results finally confirm the distortion-induced IEDDA acceleration due to intramolecular O–N repulsion and that increased reactivities similar to pyridyl-Tz can be achieved by using non-electron-withdrawing vinyl ether substituents.

Subsequent investigation of Tz stability in full cell growth medium at 37 °C moreover confirmed the high stability of vinyl ether-Tz **MeEVE** (in accordance with previous findings²⁹) and, in particular, **MeDHP** and **DHP**₂, in contrast to the limited stability of pyridyl-substituted Tz (Figure 7b).

For instance, despite being equal in reactivity, MeDHP is significantly more stable than 4Pyr (>90% vs <15%). Notably, DHP substituents do not lead to decreased stability, as shown by the data obtained for symmetrical DHP₂ and bis(2pyridyl)Tz (2Pyr₂). While installation of a second DHP had no detrimental effect on stability (>90% for both MeDHP and DHP₂), an additional 2-pyridyl substituent resulted in almost complete Tz degradation within 24 h. Despite showing very fast IEDDA reaction with TCO-PEG₄ (69,400 $M^{-1} s^{-1}$), only <1% of intact **2Pyr**₂ was detected at the end of the experiment, in comparison to >90% of DHP_2 (Figure 7b). In applications that require an extended stability of Tz (>10 h), DHP, thus outperforms even highly reactive 2Pyr₂ (Figure S5). Overall, these results confirm that non-electron-withdrawing DHP substituents can be used to significantly increase the IEDDA reactivity of Tz while maintaining a high compound stability (Figure 7c).

CONCLUSIONS

Our detailed investigation of the reactions of pyridyl-Tz with TCOs uncovered the key role of reduced Tz distortion energies caused by repulsive intramolecular interactions. Based on these insights, we have been able to confirm an analogous effect in the case of vinyl ether-Tz and showed that 3,4-dihydro-2*H*-pyran (DHP) substitution increases IEDDA reactivity without accelerating Tz degradation under physiological conditions. Overall, we provide a new mechanistic understanding that may be instrumental in the rational design of next-generation bioorthogonal tools with enhanced reactivity and stability, particularly for strategies that require or benefit from long-term Tz stability.^{9,50-52}

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/jacs.2c01056.

Computational methods, organic synthesis, reaction kinetics, compound stability and characterization, and copies of NMR spectra (PDF)

Coordinates of all calculated geometries (ZIP)

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Notes

The authors declare no competing financial interest.

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REFERENCES

 Nguyen, S. S.; Prescher, J. A. Developing bioorthogonal probes to span a spectrum of reactivities. *Nat. Rev. Chem.* 2020, *4*, 476–489.
 Mayer, S.; Lang, K. Tetrazines in Inverse-Electron-Demand Diels–Alder Cycloadditions and Their Use in Biology. *Synthesis* 2017, *49*, 830–848.

(3) Oliveira, B. L.; Guo, Z.; Bernardes, G. J. L. Inverse electron demand Diels-Alder reactions in chemical biology. *Chem. Soc. Rev.* 2017, *46*, 4895–4950.

(4) Selvaraj, R.; Fox, J. M. trans-Cyclooctene—a stable, voracious dienophile for bioorthogonal labeling. *Curr. Opin. Chem. Biol.* 2013, 17, 753–760.

(5) Darko, A.; Wallace, S.; Dmitrenko, O.; Machovina, M. M.; Mehl, R. A.; Chin, J. W.; Fox, J. M. Conformationally Strained trans-Cyclooctene with Improved Stability and Excellent Reactivity in Tetrazine Ligation. *Chem. Sci.* **2014**, *5*, 3770–3776.

(6) Pigga, J. E.; Rosenberger, J. E.; Jemas, A.; Boyd, S. J.; Dmitrenko,
O.; Xie, Y.; Fox, J. M. Angew. Chem., Int. Ed. 2021, 60, 14975–14980.
(7) Yang, J.; Šečkutė, J.; Cole, C. M.; Devaraj, N. K. Live-Cell Imaging of Cyclopropene Tags with Fluorogenic Tetrazine Cycloadditions. Angew. Chem., Int. Ed. 2012, *S1*, 7476–7479.

(8) Knall, A.-C.; Hollauf, M.; Slugovc, C. Kinetic studies of inverse electron demand Diels–Alder reactions (iEDDA) of norbornenes and 3,6-dipyridin-2-yl-1,2,4,5-tetrazine. *Tetrahedron Lett.* **2014**, *55*, 4763–4766.

(9) Wu, K.; Yee, N. A.; Srinivasan, S.; Mahmoodi, A.; Zakharian, M.; Mejia Oneto, J. M.; Royzen, M. Click activated protodrugs against cancer increase the therapeutic potential of chemotherapy through local capture and activation. *Chem. Sci.* **2021**, *12*, 1259–1271.

(10) Srinivasan, S.; Yee, N. A.; Wu, K.; Zakharian, M.; Mahmoodi, A.; Royzen, M.; Mejía Oneto, J. M. SQ3370 Activates Cytotoxic Drug via Click Chemistry at Tumor and Elicits Sustained Responses in Injected and Non-Injected Lesions. *Adv. Thermoelectr.* **2021**, *4*, 2000243.

(11) Denk, C.; Svatunek, D.; Filip, T.; Wanek, T.; Lumpi, D.; Fröhlich, J.; Kuntner, C.; Mikula, H. Development of a18F-Labeled Tetrazine with Favorable Pharmacokinetics for Bioorthogonal PET Imaging. *Angew. Chem., Int. Ed.* **2014**, *53*, 9655–9659.

(12) Denk, C.; Svatunek, D.; Mairinger, S.; Stanek, J.; Filip, T.; Matscheko, D.; Kuntner, C.; Wanek, T.; Mikula, H. Design, Synthesis, and Evaluation of a Low-Molecular-Weight 11C-Labeled Tetrazine for Pretargeted PET Imaging Applying Bioorthogonal in Vivo Click Chemistry. *Bioconjugate Chem.* **2016**, *27*, 1707–1712.

(13) Versteegen, R. M.; Rossin, R.; ten Hoeve, W.; Janssen, H. M.; Robillard, M. S. Click to Release: Instantaneous Doxorubicin Elimination upon Tetrazine Ligation. *Angew. Chem., Int. Ed.* **2013**, *52*, 14112–14116. (14) Carlson, J. C. T.; Mikula, H.; Weissleder, R. Unraveling Tetrazine-Triggered Bioorthogonal Elimination Enables Chemical Tools for Ultrafast Release and Universal Cleavage. *J. Am. Chem. Soc.* **2018**, *140*, 3603–3612.

(15) Rossin, R.; Versteegen, R. M.; Wu, J.; Khasanov, A.; Wessels, H. J.; Steenbergen, E. J.; ten Hoeve, W.; Janssen, H. M.; van Onzen, A. H. A. M.; Hudson, P. J.; Robillard, M. S. Chemically triggered drug release from an antibody-drug conjugate leads to potent antitumour activity in mice. *Nat. Commun.* **2018**, *9*, 1484.

(16) Fan, X.; Ge, Y.; Lin, F.; Yang, Y.; Zhang, G.; Ngai, W. S. C.; Lin, Z.; Zheng, S.; Wang, J.; Zhao, J.; Li, J.; Chen, P. R. Optimized Tetrazine Derivatives for Rapid Bioorthogonal Decaging in Living Cells. *Angew. Chem., Int. Ed.* **2016**, *55*, 14046–14050.

(17) Werther, P.; Yserentant, K.; Braun, F.; Kaltwasser, N.; Popp, C.; Baalmann, M.; Herten, D. P.; Wombacher, R. Live-Cell Localization Microscopy with a Fluorogenic and Self-Blinking Tetrazine Probe. *Angew. Chem., Int. Ed.* **2020**, *59*, 804–810.

(18) Tu, J.; Svatunek, D.; Parvez, S.; Liu, A. C.; Levandowski, B. J.; Eckvahl, H. J.; Peterson, R. T.; Houk, K. N.; Franzini, R. M. Stable, Reactive, and Orthogonal Tetrazines: Dispersion Forces Promote the Cycloaddition with Isonitriles. *Angew. Chem., Int. Ed.* **2019**, *58*, 9043– 9048.

(19) Sarris, A. J. C.; Hansen, T.; de Geus, M. A. R.; Maurits, E.; Doelman, W.; Overkleeft, H. S.; Codée, J. D. C.; Filippov, D. V.; van Kasteren, S. I. Fast and pH-Independent Elimination of trans-Cyclooctene by Using Aminoethyl-Functionalized Tetrazines. *Chem.—Eur. J.* **2018**, *24*, 18075–18081.

(20) Wilkovitsch, M.; Haider, M.; Sohr, B.; Herrmann, B.; Klubnick, J.; Weissleder, R.; Carlson, J. C. T.; Mikula, H. A Cleavable C₂-Symmetric trans-Cyclooctene Enables Fast and Complete Bioorthogonal Disassembly of Molecular Probes. *J. Am. Chem. Soc.* **2020**, *142*, 19132–19141.

(21) Rossin, R.; Renart Verkerk, P.; van den Bosch, S. M.; Vulders, R. C. M.; Verel, I.; Lub, J.; Robillard, M. S. In Vivo Chemistry for Pretargeted Tumor Imaging in Live Mice. *Angew. Chem., Int. Ed.* **2010**, *49*, 3375–3378.

(22) Stéen, E. J. L.; Jørgensen, J. T.; Denk, C.; Battisti, U. M.; Nørregaard, K.; Edem, P. E.; Bratteby, K.; Shalgunov, V.; Wilkovitsch, M.; Svatunek, D.; Poulie, C. B. M.; Hvass, L.; Simón, M.; Wanek, T.; Rossin, R.; Robillard, M.; Kristensen, J. L.; Mikula, H.; Kjaer, A.; Herth, M. M. Lipophilicity and Click Reactivity Determine the Performance of Bioorthogonal Tetrazine Tools in Pretargeted In Vivo Chemistry. ACS Pharmacol. Transl. Sci. 2021, 4, 824–833.

(23) Meimetis, L. G.; Carlson, J. C. T.; Giedt, R. J.; Kohler, R. H.; Weissleder, R. Ultrafluorogenic Coumarin–Tetrazine Probes for Real-Time Biological Imaging. *Angew. Chem., Int. Ed.* **2014**, *53*, 7531– 7534.

(24) Zeglis, B. M.; Sevak, K. K.; Reiner, T.; Mohindra, P.; Carlin, S. D.; Zanzonico, P.; Weissleder, R.; Lewis, J. S. A Pretargeted PET Imaging Strategy Based on Bioorthogonal Diels-Alder Click Chemistry. J. Nucl. Med. 2013, 54, 1389–1396.

(25) Denk, C.; Wilkovitsch, M.; Aneheim, E.; Herth, M. M.; Jensen, H.; Lindegren, S.; Mikula, H. Multifunctional Clickable Reagents for Rapid Bioorthogonal Astatination and Radio-Crosslinking. *ChemPluschem* **2019**, *84*, 775–778.

(26) García-Vázquez, R.; Battisti, U. M.; Jørgensen, J. T.; Shalgunov, V.; Hvass, L.; Stares, D. L.; Petersen, I. N.; Crestey, F.; Löffler, A.; Svatunek, D.; Kristensen, J. L.; Mikula, H.; Kjaer, A.; Herth, M. M. Direct Cu-mediated aromatic 18F-labeling of highly reactive tetrazines for pretargeted bioorthogonal PET imaging. *Chem. Sci.* **2021**, *12*, 11668–11675.

(27) Qu, Y.; Sauvage, F.-X.; Clavier, G.; Miomandre, F.; Audebert, P. Metal-Free Synthetic Approach to 3-Monosubstituted Unsymmetrical 1,2,4,5-Tetrazines Useful for Bioorthogonal Reactions. *Angew. Chem., Int. Ed.* **2018**, *57*, 12057–12061.

(28) Yang, J.; Karver, M. R.; Li, W.; Sahu, S.; Devaraj, N. K. Metal-Catalyzed One-Pot Synthesis of Tetrazines Directly from Aliphatic Nitriles and Hydrazine. *Angew. Chem., Int. Ed.* **2012**, *51*, 5222–5225. (29) Xie, Y.; Fang, Y.; Huang, Z.; Tallon, A. M.; am Ende, C. W.; Fox, J. M. Divergent Synthesis of Monosubstituted and Unsymmetrical 3,6-Disubstituted Tetrazines from Carboxylic Ester Precursors. *Angew. Chem., Int. Ed.* **2020**, *59*, 16967–16973.

(30) Mao, W.; Shi, W.; Li, J.; Su, D.; Wang, X.; Zhang, L.; Pan, L.; Wu, X.; Wu, H. Organocatalytic and Scalable Syntheses of Unsymmetrical 1,2,4,5-Tetrazines by Thiol-Containing Promotors. *Angew. Chem., Int. Ed.* **2019**, *58*, 1106–1109.

(31) Lambert, W. D.; Fang, Y.; Mahapatra, S.; Huang, Z.; am Ende, C. W.; Fox, J. M. Installation of Minimal Tetrazines through Silver-Mediated Liebeskind–Srogl Coupling with Arylboronic Acids. J. Am. Chem. Soc. 2019, 141, 17068–17074.

(32) Liu, F.; Liang, Y.; Houk, K. N. Theoretical Elucidation of the Origins of Substituent and Strain Effects on the Rates of Diels–Alder Reactions of 1,2,4,5-Tetrazines. *J. Am. Chem. Soc.* **2014**, *136*, 11483–11493.

(33) Shin, Y.-H.; Koh, H.-J.; Um, I.-H. Kinetic Study on Alkaline Hydrolysis of 2-Pyridyl and 4-Pyridyl X-substituted-Benzoates: Effects of Benzoyl Substituent X and Leaving-Group Basicity on Reactivity and Reaction Mechanism. *Bull. Korean Chem. Soc.* **2017**, *38*, 1138–1142.

(34) Hansch, C.; Leo, A.; Taft, R. W. A survey of Hammett substituent constants and resonance and field parameters. *Chem. Rev.* **1991**, *91*, 165–195.

(35) Eicher, T.; Hauptmann, S.; Speicher, A. Six-Membered Heterocycles: Sections 6.10–6.14. In *The Chemistry of Heterocycles*, 2nd ed.; Eicher, T., Hauptmann, S., Speicher, A., Eds.; Wiley-VCH: Weinheim, Germany, 2003; pp 257–310.

(36) Vázquez, A.; Dzijak, R.; Dračínský, M.; Rampmaier, R.; Siegl, S. J.; Vrabel, M. Mechanism-Based Fluorogenic trans-Cyclooctene-Tetrazine Cycloaddition. *Angew. Chem., Int. Ed.* **2017**, *56*, 1334–1337.

(37) Svatunek, D.; Denk, C.; Rosecker, V.; Sohr, B.; Hametner, C.; Allmaier, G.; Fröhlich, J.; Mikula, H. Efficient low-cost preparation of trans-cyclooctenes using a simplified flow setup for photoisomerization. *Monatsh. Chem.* **2016**, 147, 579–585.

(38) Bickelhaupt, F. M.; Houk, K. N. Analyzing Reaction Rates with the Distortion/Interaction-Activation Strain Model. *Angew. Chem., Int. Ed.* **2017**, *56*, 10070–10086.

(39) Svatunek, D.; Houk, K. N. autoDIAS: a python tool for an automated distortion/interaction activation strain analysis. *J. Comput. Chem.* **2019**, *40*, 2509–2515.

(40) Svatunek, D.; Houszka, N.; Hamlin, T. A.; Bickelhaupt, F. M.; Mikula, H. Chemoselectivity of Tertiary Azides in Strain-Promoted Alkyne-Azide Cycloadditions. *Chem. - Eur. J.* **2019**, *25*, 754–758.

(41) Gold, B.; Aronoff, M. R.; Raines, R. T. Decreasing Distortion Energies without Strain: Diazo-Selective 1,3-Dipolar Cycloadditions. *J. Org. Chem.* **2016**, *81*, 5998–6006.

(42) Liang, Y.; Mackey, J. L.; Lopez, S. A.; Liu, F.; Houk, K. N. Control and Design of Mutual Orthogonality in Bioorthogonal Cycloadditions. *J. Am. Chem. Soc.* **2012**, *134*, 17904–17907.

(43) Hamlin, T. A.; Svatunek, D.; Yu, S.; Ridder, L.; Infante, I.; Visscher, L.; Bickelhaupt, F. M. Elucidating the Trends in Reactivity of Aza-1,3-Dipolar Cycloadditions. *Eur. J. Org. Chem.* **2019**, 2019, 378–386.

(44) Dones, J. M.; Abularrage, N. S.; Khanal, N.; Gold, B.; Raines, R. T. Acceleration of 1,3-Dipolar Cycloadditions by Integration of Strain and Electronic Tuning. *J. Am. Chem. Soc.* **2021**, *143*, 9489–9497.

(45) Gold, B.; Dudley, G. B.; Alabugin, I. V. Moderating Strain without Sacrificing Reactivity: Design of Fast and Tunable Noncatalyzed Alkyne–Azide Cycloadditions via Stereoelectronically Controlled Transition State StabilizationModerating Strain without Sacrificing Reactivity: Design of Fast and Tunable Noncatalyzed Alkyne-Azide Cycloadditions via Stereoelectronically Controlled Transition State Stabilization. *J. Am. Chem. Soc.* **2013**, *135*, 1558– 1569.

(46) Rossin, R.; van Duijnhoven, S. M. J.; Läppchen, T.; van den Bosch, S. M.; Robillard, M. S. Trans-Cyclooctene Tag with Improved Properties for Tumor Pretargeting with the Diels–Alder ReactionTrans-Cyclooctene Tag with Improved Properties for Tumor Pretargeting with the Diels-Alder Reaction. *Mol. Pharm.* **2014**, *11*, 3090–3096.

(47) Rossin, R.; van den Bosch, S. M.; ten Hoeve, W.; Carvelli, M.; Versteegen, R. M.; Lub, J.; Robillard, M. S. Highly Reactive trans-Cyclooctene Tags with Improved Stability for Diels–Alder Chemistry in Living SystemsHighly Reactive trans-Cyclooctene Tags with Improved Stability for Diels-Alder Chemistry in Living Systems. *Bioconjugate Chem.* **2013**, *24*, 1210–1217.

(48) Tolshchina, S. G.; Rusinov, G. L.; Charushin, V. N. 1,2,4,5-Tetrazines and Azolo[1,2,4,5]tetrazines: Synthesis and Reactions with Nucleophiles. *Chem. Heterocycl. Compd.* **2013**, *49*, 66–91.

(49) Karver, M. R.; Weissleder, R.; Hilderbrand, S. A. Synthesis and Evaluation of a Series of 1,2,4,5-Tetrazines for Bioorthogonal Conjugation. *Bioconjugate Chem.* **2011**, *22*, 2263–2270.

(50) van Onzen, A. H. A. M.; Versteegen, R. M.; Hoeben, F. J. M.; Filot, I. A. W.; Rossin, R.; Zhu, T.; Wu, J.; Hudson, P. J.; Janssen, H. M.; ten Hoeve, W.; Robillard, M. S. Bioorthogonal Tetrazine Carbamate Cleavage by Highly Reactive trans-Cyclooctene. J. Am. Chem. Soc. 2020, 142, 10955–10963.

(51) Blizzard, R. J.; Backus, D. R.; Brown, W.; Bazewicz, C. G.; Li, Y.; Mehl, R. A. Ideal Bioorthogonal Reactions Using A Site-Specifically Encoded Tetrazine Amino Acid. *J. Am. Chem. Soc.* **2015**, *137*, 10044–10047.

(52) Jang, H. S.; Jana, S.; Blizzard, R. J.; Meeuwsen, J. C.; Mehl, R. A. Access to Faster Eukaryotic Cell Labeling with Encoded Tetrazine Amino Acids. J. Am. Chem. Soc. **2020**, *142*, 7245–7249.