

UCSF

UC San Francisco Previously Published Works

Title

Genome-wide association study identifies TH1 pathway genes associated with lung function in asthmatic patients

Permalink

<https://escholarship.org/uc/item/56f0945p>

Journal

Journal of Allergy and Clinical Immunology, 132(2)

ISSN

0091-6749

Authors

Li, Xingnan
Hawkins, Gregory A
Ampleford, Elizabeth J
[et al.](#)

Publication Date

2013-08-01

DOI

10.1016/j.jaci.2013.01.051

Peer reviewed



Published in final edited form as:

J Allergy Clin Immunol. 2013 August ; 132(2): 313–320.e15. doi:10.1016/j.jaci.2013.01.051.

Genome-wide association study identifies T_H1 pathway genes associated with lung function in asthmatic patients

Xingnan Li, PhD, MS^a, Gregory A. Hawkins, PhD^a, Elizabeth J. Ampleford, PhD^a, Wendy C. Moore, MD^a, Huashi Li, MS^a, Annette T. Hastie, PhD^a, Timothy D. Howard, PhD^a, Homer A. Boushey, MD^b, William W. Busse, MD^c, William J. Calhoun, MD^d, Mario Castro, MD^e, Serpil C. Erzurum, MD^f, Elliot Israel, MD^g, Robert F. Lemanske Jr, MD^h, Stanley J. Szeffler, MDⁱ, Stephen I. Wasserman, MD^j, Sally E. Wenzel, MD^k, Stephen P. Peters, MD, PhD^a, Deborah A. Meyers, PhD^a, and Eugene R. Bleeker, MD^a

© 2013 American Academy of Allergy, Asthma & Immunology

Corresponding author: Xingnan Li, PhD, MS, Center for Genomics and Personalized Medicine Research, Wake Forest School of Medicine, Medical Center Blvd, Winston-Salem, NC 27157. xinli@wakehealth.edu..

Disclosure of potential conflict of interest: X. Li, G. A. Hawkins, H. Li, and T. D. Howard have received grants from the National Institutes of Health (NIH). W. C. Moore has received grants from the National Heart, Lung, and Blood Institute (NHLBI) and was a subinvestigator in clinical trials performed at Wake Forest University for Aerovance, Amgen, AstraZeneca, Boehringer Ingelheim, Centocor, Ception, Forest, Genentech, GlaxoSmithKline, MedImmune, Novartis, Pfizer, Sanofi-Aventis, and Wyeth. A. T. Hastie has received a grant from the NHLBI. H. A. Boushey has received research support from the NIH/NHLBI; has consultant arrangements with Merck, GlaxoSmithKline, Genentech, Kalbios, Pharmaxis, and Johnson & Johnson; has received research support from GlaxoSmithKline and Genentech; has received payment for lectures from the Allergy, Asthma, and Immunology Foundation of Northern California and Breathe California; and has received royalties from the McGraw Hill Companies. W. W. Busse has a board membership with Merck; has consultant arrangements with Amgen, Novartis, GlaxoSmithKline, MedImmune, and Genentech; has received grants from the NIH/National Institute of Allergy and Infectious Diseases and the NIH/NHLBI; and has received royalties from Elsevier. W. J. Calhoun has consultant arrangements with Genentech and receives grants from the Federal Emergency Management Agency. M. Castro has received grants from the NIH, the American Lung Association, Asthmatx/Boston Scientific, Amgen, Ception/Cephalon/Teva, Genentech, MedImmune, Merck, Novartis, GlaxoSmithKline, Sanofi-Aventis, and Vectura; has received travel support from the NIH; has consultant arrangements with Asthmatx/Boston Scientific, Genentech, IPS, Pulmagen, and Sanofi-Aventis; has received payment for lectures from Pfizer, Merck, GlaxoSmithKline, Genentech, and Asthmatx/Boston Scientific; and receives royalties from Elsevier. E. Israel has consultant arrangements with Abbott, Amgen, Cowen & Co, Infinity Pharmaceuticals, MedImmune (now AstraZeneca), Merck, newMentor, NKT Therapeutics, Ono Pharmaceuticals, Regeneron Pharmaceuticals, Schering-Plough, TEVA Specialty Pharmaceuticals, Gilead Sciences, Johnson & Johnson, and Agenzia Italiana del Farmico; has provided expert testimony for Campbell, Campbell, Edwards & Conroy, Diedrich & Donohue, Ficksman & Conley, Ryan Ryan Deluca LLP, and Sullway & Hollis; has received grants from Aerovance, Amgen, i3 Research (Biota), Genentech, MedImmune, and Novartis; has received payment for lectures from Merck, the Spanish Society of Allergy & Immunology, the Western Society of Allergy, Asthma & Immunology, and the World Allergy Congress; and has received royalties from UpToDate. R. F. Lemanske has received grants, travel support, and fees for participation in review activities from the NIH; has consultant arrangements with Merck, Sepracor, SA Voney and Associates LTD, GlaxoSmithKline, American Institute of Research, Genentech, Double Helix Development, and Boehringer Ingelheim; is employed by the University of Wisconsin School of Medicine and Public Health; has received grants from the NHLBI and Pharmaxis; has received payment for lectures from the Michigan Public Health Institute, Allegheny General Hospital, the American Academy of Pediatrics, West Allegheny Health Systems, California Chapter 4 of the American Academy of Pediatrics, the Colorado Allergy Society, the Pennsylvania Allergy and Asthma Association, Harvard Pilgrim Health, the California Society of Allergy, the New York City Allergy Society, the World Allergy Organization, the American College of Chest Physicians, and the APAPARI; has received payment for manuscript preparation from the American Academy of Allergy, Asthma & Immunology; and has received royalties from Elsevier and UpToDate. S. J. Szeffler has received grants, travel support, payment for participation in review activities, and payment for writing or reviewing the manuscript from the NHLBI; has consultant arrangements with Merck, Genentech, Boehringer Ingelheim, and GlaxoSmithKline; has received grants from GlaxoSmithKline; has received payment for lectures from Merck; has received payment for manuscript preparation from Genentech; and has a submitted patent for a β -adrenergic receptor polymorphism for the CARE Network. S. I. Wasserman has received grants from the NIH and the American Lung Association, is President of the American Board of Allergy and Immunology, and has provided expert testimony for Scharf, Banks, Marmor in Chicago, Illinois. S. P. Peters has received grants from the NIH and the NHLBI; has consultant arrangements with AstraZeneca, Aerocrine, Airsonett, AB, GlaxoSmithKline, Merck, Targacept, and TEVA; has received payment for lectures from Integrity CE and Merck; and has received royalties from UpToDate.

The rest of the authors declare that they have no relevant conflicts of interest.

Winston-Salem, NC, San Francisco and La Jolla, Calif, Madison, Wis, Galveston, Tex, St Louis, Mo, Cleveland, Ohio, Boston, Mass, Denver, Colo, and Pittsburgh, Pa

^athe Center for Genomics and Personalized Medicine Research, Wake Forest School of Medicine, Winston-Salem; ^bthe Department of Medicine, University of California at San Francisco; ^cthe Department of Medicine, University of Wisconsin, Madison; ^dthe Department of Internal Medicine, University of Texas Medical Branch at Galveston; ^ethe Department of Medicine, Washington University School of Medicine, St Louis; ^fthe Lerner Research Institute, Cleveland; ^gthe Pulmonary and Critical Care Division, Brigham and Women's Hospital and Harvard Medical School, Boston; ^hthe Clinical Science Center, University of Wisconsin School of Medicine and Public Health, Madison; ⁱthe Department of Pediatrics, National Jewish Health, Denver; ^jthe Department of Medicine, University of California at San Diego, La Jolla; ^kthe Department of Medicine, University of Pittsburgh.

Abstract

Background—Recent meta-analyses of genome-wide association studies in general populations of European descent have identified 28 loci for lung function.

Objective—We sought to identify novel lung function loci specifically for asthma and to confirm lung function loci identified in general populations.

Methods—Genome-wide association studies of lung function (percent predicted FEV₁ [ppFEV₁], percent predicted forced vital capacity, and FEV₁/forced vital capacity ratio) were performed in 4 white populations of European descent (n = 1544), followed by meta-analyses.

Results—Seven of 28 previously identified lung function loci (*HHIP*, *FAM13A*, *THSD4*, *GSTCD*, *NOTCH4-AGER*, *RARB*, and *ZNF323*) identified in general populations were confirmed at single nucleotide polymorphism (SNP) levels ($P < .05$). Four of 32 loci (*IL12A*, *IL12RB1*, *STAT4*, and *IRF2*) associated with ppFEV₁ ($P < 10^{-4}$) belong to the T_H1 or IL-12 cytokine family pathway. By using a linear additive model, these 4 T_H1 pathway SNPs cumulatively explained 2.9% to 7.8% of the variance in ppFEV₁ values in 4 populations ($P = 3 \times 10^{-11}$). Genetic scores of these 4 SNPs were associated with ppFEV₁ values ($P = 2 \times 10^{-7}$) and the American Thoracic Society severe asthma classification ($P = .005$) in the Severe Asthma Research Program population. T_H2 pathway genes (*IL13*, *TSLP*, *IL33*, and *IL1RL1*) conferring asthma susceptibility were not associated with lung function.

Conclusion—Genes involved in airway structure/remodeling are associated with lung function in both general populations and asthmatic subjects. T_H1 pathway genes involved in anti-virus/bacterial infection and inflammation modify lung function in asthmatic subjects. Genes associated with lung function that might affect asthma severity are distinct from those genes associated with asthma susceptibility.

Keywords

Lung function; FEV₁; asthma; T_H1; IL12A; IL12RB1; STAT4; IRF2

Asthma is a chronic inflammatory respiratory disease with a high level of genetic and phenotypic heterogeneity. Phenotypically, asthma can be classified as mild, moderate, or severe according to National Asthma Education and Prevention Program, Global Initiative for Asthma, or American Thoracic Society (ATS) guidelines.¹⁻³ Recent work has described 5 distinct asthma severity phenotypes through cluster analysis.⁴ Measures of lung function,

particularly percent predicted FEV₁ (ppFEV₁) values, are essential for categorizing asthma severity by using either current guidelines or cluster methodologies.¹⁻⁴

Three recent large meta-analyses of genome-wide association studies (GWASs) in general populations of European descent have identified 28 loci for lung function⁵⁻⁷: *HHIP*, *HTR4*, *INTS12-GSTCD-NPNT*, *TNS1*, *C10orf11*, *CDC123*, *MECOM*, and *ZKSCAN3-ZNF323* for FEV₁ and *ADAM19*, *DAAM2*, *FAM13A*, *GPR126*, *HHIP*, *HTR4*, *NOTCH4-AGER-PPT2*, *PID1*, *PTCH1*, *THSD4*, *ARMC2*, *CCDC38*, *CDC123*, *CFDP1*, *HDAC4*, *KCNE2*, *LRP1*, *MFAP2*, *MMP15*, *NCR3*, *RARB*, *SPATA9*, and *TGFB2* for FEV₁/forced vital capacity (FVC) ratio. In a previous study of candidate genes, we have shown that *HHIP* is associated with ppFEV₁ and percent predicted forced vital capacity (ppFVC) values in asthmatic subjects.⁸ A recent GWAS of lung function decrease has suggested that genetic profiles for longitudinal and cross-sectional lung function differ between subjects with and without asthma.⁹

In this study we performed meta-analyses of GWASs of lung function (ppFEV₁, ppFVC, and FEV₁/FVC ratio) in 4 European American asthmatic populations to determine whether the 28 loci identified in the previous GWASs for normal variation of lung function are important components affecting abnormal lung function in asthmatic subjects. More importantly, we explored whether some novel genes have large effects on lung function unique to asthmatic subjects.

METHODS

Study subjects

Subjects with mild-to-severe asthma recruited at the National Heart, Lung, and Blood Institute (NHLBI)-funded Severe Asthma Research Program (SARP) centers were carefully characterized, including baseline spirometry with a medication withhold before testing.^{3,4,8} Similar baseline spirometry was performed in subjects with severe or difficult-to-treat asthma from The Epidemiology and Natural History of Asthma: Outcomes and Treatment Regimens (TENOR) multicenter study.^{8,10,11} Asthmatic subjects who were studied in the NHLBI's Collaborative Studies on the Genetics of Asthma (CSGA) by the Wake Forest investigators using a similar protocol were included in the analyses.^{8,12} Asthmatic patients from 2 NHLBI-funded Asthma Clinical Research Network (ACRN) clinical trials (the Tiotropium Bromide as an Alternative to Increased Inhaled Corticosteroid in Patients Inadequately Controlled on a Lower Dose of Inhaled Corticosteroids [TALC] and Best Adjustment Strategy for Asthma in Long Term [BASALT] trials) were also included.¹³ The patients from the BASALT trials (ClinicalTrials.gov no. NCT00495157) had mild-to-moderate asthma. The patients from the TALC trials (ClinicalTrials.gov no. NCT00565266) had poorly controlled disease on a lower dose of inhaled corticosteroid.¹³ All studies were approved by the appropriate institutional review boards at the participating sites, including obtaining appropriate informed consent.

DNA was isolated by using standard protocols, and single nucleotide polymorphism (SNP) genotyping was performed with the Illumina HumanCNV370 BeadChip (Illumina, San Diego, Calif) for TENOR samples. SARP and CSGA samples were genotyped with the Illumina HumanHap1M BeadChip. ACRN samples were genotyped with the Illumina HumanOmniExpress700k BeadChip. Genotyping for all studies was performed with BeadStudio or GenomeStudio (Illumina).

Statistical analysis

Quality control processes of GWASs for the SARP, CSGA, and TENOR populations have been described previously.⁸ Quality control of ACRN cases was performed in a similar manner.

ppFEV₁ and ppFVC values were calculated based on Hankinson formula.¹⁴ Outliers (outside of 3× SDs of ppFEV₁, ppFVC, and FEV₁/FVC ratio values) were removed. ppFEV₁, ppFVC, and FEV₁/FVC ratio values were standardized with a mean of 0 and an SD of 1. A linear additive model was used for analysis of standardized ppFEV₁, ppFVC, and FEV₁/FVC ratio values in the SARP, CSGA, TENOR, and ACRN populations separately by using PLINK (version 1.06, <http://pngu.mgh.harvard.edu/purcell/plink/>)¹⁵ adjusted for significant principal components (EIGENSTRAT, version 3.0, <http://genepath.med.harvard.edu/~reich/Software.htm>).¹⁶ Association plots were generated with SNAP (version 2.0; <http://www.broad.mit.edu/mpg/snap/>).¹⁷

Meta-analyses of linear regression slope β with 369,771 SNPs that were shared in all studies from 4 European American populations (SARP, CSGA, TENOR, and ACRN) were performed, with weights proportional to the inverse variance of β using METAL software (<http://www.sph.umich.edu/csg/abecasis/metal/>). *P* values with genomic control adjustment from each population were used for meta-analysis to reduce genomic inflation.

Joint analysis of the 4 T_H1 pathway SNPs (rs7636840 in *IL12A*, rs388159 in *IL12RB1*, rs925847 in *STAT4*, and rs3756089 in *IRF2*) was performed in the SARP population. Genetic scores were defined by the number of risk alleles present in these 4 SNPs. A linear or logistic model was used for analysis of ppFEV₁ values and percentages of subjects with severe asthma by using the ATS asthma severity classification,³ with genetic scores as defined.

RESULTS

Subjects' demographics

Although the 4 studies were all composed of European American subjects with asthma and use similar clinical approaches to phenotyping cases, they differed in the proportion of subjects with severe asthma. SARP is a cohort enriched for subjects with severe asthma but also well balanced with subjects with mild-to-moderate asthma. TENOR focused on subjects with difficult-to-treat or severe asthma. CSGA included all levels of severity but primarily milder levels. The ACRN study is composed of the TALC and BASALT trials, in which BASALT had subjects with mild-to-moderate asthma but TALC focused on subjects with poorly controlled asthma receiving a low dose of inhaled corticosteroids. These phenotypic differences are reflected in baseline lung function (see Table E1 in this article's Online Repository at www.jacionline.org). Only subjects with an age of enrollment of 12 years or greater were included in the analysis to reduce the age effect on lung development. The SARP cohort is well phenotyped with a relatively large sample size (n=618) and has a broad range of lung function, and thus SARP was used as the primary population for this analysis.

After a quality control process, 1,544 subjects from the 4 studies were analyzed with the shared 369,771 SNPs for association with predrug spirometric measures of lung function: ppFEV₁, ppFVC, and FEV₁/FVC ratio. ppFEV₁ was the primary phenotype in this study because it is the lung function measure most used for clinical studies in asthmatic subjects.

Meta-analysis of GWASs of ppFEV₁

Forty-four SNPs from 32 independent loci were suggestively associated with ppFEV₁ values ($P < 10^{-4}$; Fig 1 and see Table E2 and Fig E1 in this article's Online Repository at www.jacionline.org). rs7670758, which is located approximately 60 kb upstream of *HHIP*, was associated with ppFEV₁ values ($P = 9.5 \times 10^{-5}$). The minor allele A of rs7670758 was associated with decreased ppFEV₁ values. This effect direction for rs7670758 is consistent with previous findings in general populations,⁵⁻⁷ supporting our study design and statistical analysis.

Four of 32 loci associated with ppFEV₁ values were within or near important T_H1 or IL-12 cytokine family pathway genes: *IL12A* (rs7636840 and rs11918254 in strong linkage disequilibrium [LD]: $r^2 = 0.94$), *IL12RB1* (rs388159), *STAT4* (rs925847), and *IRF2* (rs3756089; Table I and see Table E2 and Figs E2-E5 in this article's Online Repository at www.jacionline.org). Four T_H1 pathway SNPs were consistently associated with ppFEV₁ values in the SARP, CSGA, TENOR, and ACRN populations (Tables I and II and see Table E3 in this article's Online Repository at www.jacionline.org).

These 4 T_H1 pathway SNPs cumulatively explained 2.9% to 7.8% of the variance in ppFEV₁ values in 4 populations, with a meta-analysis P value of 3×10^{-11} (Table II). The possession of an increased number of risk alleles (genetic scores) for these 4 SNPs inversely correlated with ppFEV₁ values ($P = 2 \times 10^{-7}$) and positively correlated with the ATS asthma severity classification ($P = .005$) in the SARP population (Fig 2). In the SARP cohort ppFEV₁ values displayed a stepwise decrease from 81.9 (SD, 20.1) to 76.6 (SD, 21.1) and then to 65.1 (SD, 22.3), and the percentage of subjects with severe asthma (based on ATS classification) showed an increase from 40.2% to 49.4% and then to 64.3% with the increasing number of risk alleles (Fig 2).

Meta-analyses of GWASs of ppFVC and FEV₁/FVC ratio values

Thirty-nine SNPs were suggestively associated with ppFVC values ($P < 10^{-4}$, see Table E4 in this article's Online Repository at www.jacionline.org). Two SNPs downstream of *IL12RB1* (rs12984174 and rs388159 in moderate LD: $r^2 = 0.75$) ranked as number 1 and number 9. rs388159 was also the top candidate SNP associated with ppFEV₁ values (Table I), making it the most robust finding in this study.

Thirty-one SNPs were suggestively associated with FEV₁/FVC ratio values ($P < 10^{-4}$, see Table E5 in this article's Online Repository at www.jacionline.org). rs11032873, which is located between *APIP* and *EHF*, was associated with FEV₁/FVC ratio values ($P = 3.1 \times 10^{-5}$). The *APIP-EHF* region has been identified as a modifier locus of lung disease severity in patients with cystic fibrosis.¹⁸ Two adenylate cyclase family genes (*ADCY2* and *ADCY9*) were associated with FEV₁/FVC ratio values ($P = 5.6 \times 10^{-5}$ or 6.3×10^{-5} for rs12659620 or rs2230739, respectively). rs3130696, which is located upstream of *HLA-C*, was associated with FEV₁/FVC ratio values ($P = 7.3 \times 10^{-5}$).

Replication of 28 normal lung function loci

Two meta-analyses of GWASs in approximately 20,000 healthy subjects of European descent have identified 12 loci for lung function^{6,7}: *HHIP*, *HTR4*, *INTS12-GSTCD-NPNT*, and *TNSI* for FEV₁ and *ADAM19*, *DAAM2*, *FAM13A*, *GPR126*, *HHIP*, *HTR4*, *NOTCH4-AGER-PPT2*, *PID1*, *PTCH1*, and *THSD4* for FEV₁/FVC ratio (see Table E6 in this article's Online Repository at www.jacionline.org). One more recent meta-analysis of GWASs in approximately 90,000 healthy subjects of European descent has identified 16 novel loci for lung function⁵: *C10orf11*, *CDC123*, *MECOM*, and *ZKSCAN3-ZNF323* for FEV₁ and *ARMC2*, *CCDC38*, *CDC123*, *CFDP1*, *HDAC4*, *KCNE2*, *LRP1*, *MFAP2*, *MMP15*, *NCR3*,

RARB, SPATA9, and *TGFB2* for FEV₁/FVC ratio (see Table E7 in this article's Online Repository at www.jacionline.org).

In this study we replicated 7 of 28 lung function loci at SNP levels ($P < .05$) for the same phenotypes and in the same effect direction in asthma: *HHIP* (rs1980057 and rs7670758), *FAM13A* (rs2869967 and rs6825998), *THSD4* (rs12899618), *GSTCD* (rs17035917 and rs6820671), *NOTCH4-AGER* (rs206015), *RARB* (rs1529672 and rs7616278), and *ZNF323* (rs6922111 and rs1416920, see Tables E6 and E7).

Comparison of asthma genes and lung function genes

Although lung function is an essential intermediate phenotype distinct between asthmatic patients and healthy subjects, the genes associated with lung function were largely not associated with asthma susceptibility in the GABRIEL study (extracted from the European Genome-Phenome Archive; <http://www.cng.fr/gabriel>; accession no. EGAS00000000077).¹⁹

In this study the 4 T_H1 pathway SNPs associated with lung function were not associated with asthma susceptibility in the GABRIEL,¹⁹ TENOR,¹¹ or SARP/CSGA²⁰ studies (see Table E8 in this article's Online Repository at www.jacionline.org). Six SNPs (in *ORMDL3-GSDMB*, *IL33*, *IL1RL1-IL18R1*, *TSLP*, *IL13*, and *HLA-DRA*) associated with asthma susceptibility¹⁹⁻²¹ were not associated with ppFEV₁ values (see Table E9 in this article's Online Repository at www.jacionline.org).

Two-step asthma progression genetic model

On the basis of this study and previous findings, we suggest a 2-step asthma progression genetic model (Fig 3). In the first step genetic variants in T_H2 pathway genes (*IL13*, *TSLP*, *IL33*, and *IL1RL1*), interacting with environmental factors, induce T_H2-dominant response and atopy, which distinguish subjects with asthma from healthy subjects. In the second step genetic variants in the T_H1 or IL-12 cytokine family pathway (*IL12A*, *IL12RB1*, *STAT4*, and *IRF2*), airway structure/remodeling (*HHIP*, *FAM13A*, *THSD4*, *GSTCD*, *NOTCH4-AGER*, *RARB*, and *ZNF323*), and/or other mechanisms interacting with environmental factors lead to lower lung function and increased asthma severity.

DISCUSSION

In this study we investigated genetic variants related to lung function in 4 European American asthmatic populations ($n = 1544$) using GWASs and meta-analyses. Although the sample size is relatively small compared with that of the studies in general populations, the current study is the first and largest GWAS of lung function in comprehensively phenotyped subjects with asthma. Further replication in well-phenotyped independent populations is essential to confirm our findings. Three predrug spirometric measures of lung function (ppFEV₁, ppFVC, and FEV₁/FVC ratio) were studied. FEV₁ is the volume of air that can be expired forcibly in 1 second after full inspiration. FVC is the volume of air that can be expired forcibly after full inspiration. The FEV₁/FVC ratio is the ratio of FEV₁ to FVC. Both FEV₁ and FVC values can be decreased in patients with respiratory diseases but might not be in the same proportion. These 3 measures are correlated but might reflect fine differences in lung function. The FEV₁/FVC ratio and FEV₁ reflect airway obstruction and the grade of airway obstruction and thus are used together to define chronic obstructive pulmonary disease. ppFEV₁ was the primary phenotype in this study because it is highly correlated with asthma severity and mostly used for clinical studies in asthmatic subjects. Although no SNP reached genome-wide significance ($P = 5 \times 10^{-8}$), multiple SNPs were suggestively associated with ppFEV₁, ppFVC, or FEV₁/FVC ratio values ($P < 10^{-4}$, see

Tables E2, E4, and E5). In general, the results for these 3 measurements agreed well, although the top SNPs were not always the same in the 3 measurements (see Table E2). More importantly, multiple top SNPs belong to the same pathway and are functionally related to lung function. The same effect direction of these SNPs in all 4 studied populations further supports our results. Although a single SNP with a modest P value does not meet the strict requirements for association in a GWAS analysis, multiple independent loci mapped to the same biological pathway are very unlikely to be a result of chance, and thus pathway analysis is an important complementary approach to a GWAS.²²

For example, 4 of 32 loci associated with ppFEV₁ values are T_H1 or IL-12 cytokine family pathway genes: *IL12A*, *IL12RB1*, *STAT4*, and *IRF2* (Table I). Although these 4 SNPs were not significant individually at the genome-wide level, cumulatively they explained 2.9% to 7.8% of the variance in ppFEV₁ values ($P = 3 \times 10^{-11}$, Table II). In the well-phenotyped SARP population genetic scores of these 4 SNPs were inversely associated with ppFEV₁ values ($P = 2 \times 10^{-7}$) and positively associated with the ATS asthma severity classification ($P = .005$). The risk allele of *IL12RB1*, *STAT4*, and *IRF2* is its major allele, and the risk allele of *IL12A* is its minor allele. In general, the combinations of risk alleles involving *IRF2* were correlated with higher ppFEV₁ values, and those including *IL12A* were associated with lower ppFEV₁ values (see Table E10 in this article's Online Repository at www.jacionline.org). IL-12 is a key cytokine that modulates innate and adaptive immune responses. IL-12 is a heterodimer composed of the p35 subunit (encoded by *IL12A*) and the p40 subunit (encoded by *IL12B*). The coexpression and dimerization of the IL-12RB1 and IL-12RB2 proteins led to the formation of the high-affinity IL-12 receptor. The main role of IL-12 is to activate IFN- γ (IFNG) production. The response of lymphocytes to IL-12 is mediated by *STAT4*, which induces the expression of *IL12RB2* and *IRF1*. *IRF1* is a transcription activator of the genes induced by IFN- α (*IFNA1*), IFN- β (*IFNB1*), and IFNG, whereas *IRF2* competitively inhibits the expression of genes activated by *IRF1*. The IL-12–*STAT4*–IFNG signaling pathway is essential for the differentiation of naive T_H cells into T_H1 cells. GWASs in patients with autoimmune diseases have consistently identified IL-12–*STAT4*–IFNG signaling pathway genes. For example, SNPs in the *IL12A* region have been associated with celiac disease,^{23,24} primary biliary cirrhosis,²⁵⁻²⁷ and multiple sclerosis.²⁸ SNPs in the *STAT4* region have been associated with systemic lupus erythematosus,²⁹⁻³¹ systemic sclerosis,³² rheumatoid arthritis,^{33,34} and primary biliary cirrhosis.²⁷ Candidate gene studies have indicated that *IL12RB1*, *STAT4*, and *IRF2* were associated with asthma, but the association results are not consistent.³⁵

Several genes associated with lung development or height were identified (see Tables E2, E4, and E5), including *HHIP*,⁵⁻⁷ *SULF1*,³⁶ and *SYN3*,^{36,37} although ppFEV₁ and ppFVC values were already adjusted for height. Similarly, GWASs of lung function in general populations also identified multiple genes associated with height,⁵⁻⁷ indicating the importance of height in determining lung volume or pulmonary function.

Two adenylate cyclase family genes (*ADCY2* and *ADCY9*) were suggestively associated with FEV₁/FVC ratio values (see Table E5). The same nonsynonymous coding SNP (rs2230739 or Ile772Met) in *ADCY9* has been shown to be significantly associated with the difference in improvement of FEV₁ values with the response to β -agonist with or without corticosteroid treatment.³⁸ In a candidate gene study *ADCY2* was associated with both chronic obstructive pulmonary disease and FEV₁/FVC ratio values independently of smoking effect.³⁹

Previously, we have shown that a subset of genes that regulate lung function in general populations might be associated with abnormal lung function in subjects with asthma.⁸ In this study this list has been extended to 7 of 28 lung function loci at the SNP level: *HHIP*

(rs1980057 and rs7670758), *FAM13A* (rs2869967 and rs6825998), *THSD4* (rs12899618), *GSTCD* (rs17035917 and rs6820671), *NOTCH4-AGER* (rs206015), *RARB* (rs1529672 and rs7616278), and *ZNF323* (rs6922111 and rs1416920, see Tables E6 and E7). Among these 7 genes, several might be involved in airway structure and airway remodeling: (1) *HHIP* might influence embryonic lung-branching morphogenesis⁴⁰; (2) *RARB* is an inhibitor of the perinatal formation of pulmonary alveoli⁴¹; and (3) *NOTCH4* and *THSD4* might be involved in angiogenesis and airway remodeling.^{42,43} Furthermore, both *TIMP3* and *CITED2*, which are associated with ppFEV₁ values (see Table E2), might be involved in hypoxia-inducible factor 1 α /vascular endothelial growth factor-induced hypoxia and angiogenesis.^{44,45}

In the GABRIEL study¹⁹ the 12 lung function loci^{6,7} were not associated with asthma susceptibility. In this study the 4 T_H1 pathway SNPs associated with lung function were not associated with asthma susceptibility (see Table E8); conversely, 6 SNPs (in *ORMDL3-GSDMB*, *IL33*, *IL1RL1-IL18R1*, *TSLP*, *IL13*, and *HLA-DRA*) conferring asthma susceptibility¹⁹⁻²¹ were not associated with ppFEV₁ values (see Table E9). These results indicate that genes associated with lung function that can modify asthma severity might be distinct from those genes associated with asthma susceptibility. Thus we hypothesize that asthma would develop in a subject with susceptibility loci but that more severe asthma with reduced lung function requires the presence of additional severity or pulmonary function loci.

GWASs in subjects with autoimmune diseases have consistently identified *IL12A*²³⁻²⁸ and *STAT4*^{27,29-34} regions. The top SNPs of *STAT4* identified in autoimmune diseases are not in strong LD with the top SNPs identified in this study, indicating that distinct transcription regulation mechanisms might exist in different tissues for autoimmune diseases and asthma. However, evidence from genetic association studies indicates the opposite direction of effect size for *STAT4* between autoimmune diseases and ppFEV₁ values. For example, the minor allele T of rs10168266 in *STAT4* is the risk allele for primary biliary cirrhosis ($P = 4.2 \times 10^{-5}$)²⁴ but the protective allele for ppFEV₁ ($P = .028$). The minor allele A of rs3821236 is the risk allele for systemic lupus erythematosus ($P = 8.5 \times 10^{-11}$)³⁰ but the protective allele for ppFEV₁ ($P = .036$). The minor allele T of rs7574865 is the risk allele for systemic lupus erythematosus in subjects of Chinese ($P = 5.2 \times 10^{-42}$)³¹ or European ($P = 9.0 \times 10^{-14}$)²⁹ descent but possibly the protective allele for ppFEV₁ ($P = .093$). The minor allele T of rs925847 is the protective allele for ppFEV₁ ($P = 8.2 \times 10^{-5}$) and eczema ($P = .014$)⁴⁶ but the major risk allele for ulcerative colitis in the Korean population ($P = .025$).⁴⁷ In general, the expression of T_H1 pathway genes is believed to be increased in subjects with autoimmune diseases, and thus the expression of T_H1 pathway genes might be positively correlated with ppFEV₁ values or negatively correlated with asthma severity (Fig 3).

The T_H1 pathway primarily acts against viruses and bacterial infections. Respiratory tract virus infection might initiate and exacerbate asthma. The decrease in production of IFN- γ in response to viral infection of monocytes at birth predicts the susceptibility to respiratory tract illness during the first year of life.⁴⁸ The weaker T_H1 responses to viral infection are associated with asthma exacerbations and suggest that viral infection can exacerbate a pre-existing T_H2-dominated lung disease.⁴⁹ In a study of acute asthma exacerbations in children,⁵⁰ the expression of T_H1 pathway genes is decreased during exacerbation, with deficits in baseline lung function. Rhinovirus-induced clinical phenotypes are more severe in asthmatic subjects than healthy control subjects and indicate impaired T_H1 or augmented T_H2 immune responses.⁵¹ The weaker T_H1 response might strengthen the T_H2-dominated response in asthmatic subjects, whereas the impaired T_H1 response against viral infection might extend the inflammation process and lead to airway remodeling (Fig 3).

We must emphasize the complexity of immunologic pathways. Although we speculated that the T_H1 pathway might be important for lung function in asthmatic subjects, the genes identified (*IL12A*, *IL12RB1*, *STAT4*, and *IRF2*) could be involved in other pathways. For example, IL-12p35, encoded by *IL12A*, is a ligand subunit shared by IL-12 and IL-35, where IL-35 might be involved in the regulatory T-cell pathway.⁵² IL-12Rβ1, which is encoded by *IL12RB1*, is a receptor subunit shared by IL-12 and IL-23, where IL-23 is a proinflammatory cytokine involved in the T_H17 pathway.⁵² More comprehensively, we can label *IL12A*, *IL12RB1*, *STAT4*, and *IRF2* as IL-12 cytokine family pathway genes.

On the basis of this study and previous findings, we propose a simplified 2-step genetic model of asthma progression (Fig 3). Genetic variants in T_H2 pathway genes (*IL13*, *TSLP*, *IL33*, and *ILIRL1*) induce T_H2-dominant response, atopy, and asthma susceptibility. Genetic variants in T_H1 or IL-12 cytokine family pathway genes (*IL12A*, *IL12RB1*, *STAT4*, and *IRF2*), airway structure/remodeling (*HHIP*, *FAM13A*, *THSD4*, *GSTCD*, *NOTCH4-AGER*, *RARB*, and *ZNF323*), and/or other mechanisms affect lung function and increase asthma severity and asthma exacerbation.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

We acknowledge contributions from all investigators, staff, and participants in the SARP, TENOR, CSGA, and ACRN studies.

Severe Asthma Research Program (SARP) centers were supported by National Institutes of Health (NIH) grants HL69116, HL69130, HL69149, HL69155, HL69167, HL69170, HL69174, HL69349, UL1RR024992, M01RR018390, M01RR07122, M01RR03186, HL87665, and HL091762. Genetic studies for SARP and Collaborative Studies on the Genetics of Asthma (CSGA) were funded by NIH grant HL87665 and Go Grant RC2HL101487. The clinical The Epidemiology and Natural History of Asthma: Outcomes and Treatment Regimens (TENOR) study was supported by Genentech and Novartis Pharmaceuticals Corporation, and the genetic studies were funded by NIH HL76285, HL87665, and Go Grant RC2HL101487.

Abbreviations used

ACRN	Asthma Clinical Research Network
ATS	American Thoracic Society
BASALT	Best Adjustment Strategy for Asthma in Long Term
CSGA	Collaborative Studies on the Genetics of Asthma
FVC	Forced vital capacity
GWAS	Genome-wide association study
LD	Linkage disequilibrium
NHLBI	National Heart, Lung, and Blood Institute
ppFEV₁	Percent predicted FEV ₁
ppFVC	Percent predicted forced vital capacity
SARP	Severe Asthma Research Program
SNP	Single nucleotide polymorphism

TALC	Tiotropium Bromide as an Alternative to Increased Inhaled Corticosteroid in Patients Inadequately Controlled on a Lower Dose of Inhaled Corticosteroids
TENOR	The Epidemiology and Natural History of Asthma: Outcomes and Treatment Regimens

REFERENCES

1. National Asthma Education and Prevention Program. Expert panel report 3: guidelines for the diagnosis and management of asthma. National Heart, Lung, and Blood Institute; Bethesda: 2007. Publication no. 07–4051. Available at: <http://www.nhlbi.nih.gov/guidelines/asthma> [Accessed July 26, 2012]
2. Global Initiative for Asthma: global strategy for asthma management and prevention (GINA). [Accessed July 26, 2012] National Institutes of Health/National Heart, Lung, and Blood Institute, updated 2008. Available at: <http://www.ginasthma.org>
3. Moore WC, Bleecker ER, Curran-Everett D, Erzurum SC, Ameredes BT, Bacharier L, et al. Characterization of the severe asthma phenotype by the National Heart, Lung, and Blood Institute's Severe Asthma Research Program. *J Allergy Clin Immunol.* 2007; 119:405–13. [PubMed: 17291857]
4. Moore WC, Meyers DA, Wenzel SE, Teague WG, Li H, Li X, et al. Identification of asthma phenotypes using cluster analysis in the Severe Asthma Research Program. *Am J Respir Crit Care Med.* 2010; 181:315–23. [PubMed: 19892860]
5. Soler Artigas M, Loth DW, Wain LV, Gharib SA, Obeidat M, Tang W, et al. Genome-wide association and large-scale follow up identifies 16 new loci influencing lung function. *Nat Genet.* 2011; 43:1082–90. [PubMed: 21946350]
6. Hancock DB, Eijgelsheim M, Wilk JB, Gharib SA, Loehr LR, Marcianti KD, et al. Meta-analyses of genome-wide association studies identify multiple loci associated with pulmonary function. *Nat Genet.* 2010; 42:45–52. [PubMed: 20010835]
7. Repapi E, Sayers I, Wain LV, Burton PR, Johnson T, Obeidat M, et al. Genome-wide association study identifies five loci associated with lung function. *Nat Genet.* 2010; 42:36–44. [PubMed: 20010834]
8. Li X, Howard TD, Moore WC, Ampleford EJ, Li H, Busse WW, et al. Importance of hedgehog interacting protein and other lung function genes in asthma. *J Allergy Clin Immunol.* 2011; 127:1457–65. [PubMed: 21397937]
9. Imboden M, Bouzigon E, Curjuric I, Ramasamy A, Kumar A, Hancock DB, et al. Genome-wide association study of lung function decline in adults with and without asthma. *J Allergy Clin Immunol.* 2012; 129:1218–28. [PubMed: 22424883]
10. Dolan CM, Fraher KE, Bleecker ER, Borish L, Chipps B, Hayden ML, et al. Design and baseline characteristics of the epidemiology and natural history of asthma: Outcomes and Treatment Regimens (TENOR) study: a large cohort of patients with severe or difficult-to-treat asthma. *Ann Allergy Asthma Immunol.* 2004; 92:32–9. [PubMed: 14756462]
11. Li X, Howard TD, Zheng SL, Haselkorn T, Peters SP, Meyers DA, et al. Genome-wide association study of asthma identifies RAD50-IL13 and HLA-DR/DQ regions. *J Allergy Clin Immunol.* 2010; 125:328–35. [PubMed: 20159242]
12. Xu J, Meyers DA, Ober C, Blumenthal MN, Mellen B, Barnes KC, et al. Genome-wide screen and identification of gene-gene interactions for asthma-susceptibility loci in three U.S. populations: collaborative study on the genetics of asthma. *Am J Hum Genet.* 2001; 68:1437–46. [PubMed: 11349227]
13. Peters SP, Kunselman SJ, Icitovic N, Moore WC, Pascual R, Ameredes BT, et al. Tiotropium bromide step-up therapy for adults with uncontrolled asthma. *N Engl J Med.* 2010; 363:1715–26. [PubMed: 20979471]
14. Hankinson JL, Odencrantz JR, Fedan KB. Spirometric reference values from a sample of the general U.S. population. *Am J Respir Crit Care Med.* 1999; 159:179–87. [PubMed: 9872837]

15. Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet.* 2007; 81:559–75. [PubMed: 17701901]
16. Price AL, Patterson NJ, Plenge RM, Weinblatt ME, Shadick NA, Reich D. Principal components analysis corrects for stratification in genome-wide association studies. *Nat Genet.* 2006; 38:904–9. [PubMed: 16862161]
17. Johnson AD, Handsaker RE, Pulit SL, Nizzari MM, O'Donnell CJ, de Bakker PI. SNAP: a web-based tool for identification and annotation of proxy SNPs using HapMap. *Bioinformatics.* 2008; 24:2938–9. [PubMed: 18974171]
18. Wright FA, Strug LJ, Doshi VK, Commander CW, Blackman SM, Sun L, et al. Genome-wide association and linkage identify modifier loci of lung disease severity in cystic fibrosis at 11p13 and 20q13.2. *Nat Genet.* 2011; 43:539–46. [PubMed: 21602797]
19. Moffatt MF, Gut IG, Demenais F, Strachan DP, Bouzigon E, Heath S, et al. A large-scale, consortium-based genomewide association study of asthma. *N Engl J Med.* 2010; 363:1211–21. [PubMed: 20860503]
20. Li X, Ampleford EJ, Howard TD, Moore WC, Torgerson DG, Li H, et al. Genome-wide association studies of asthma indicate opposite immunopathogenesis direction from autoimmune diseases. *J Allergy Clin Immunol.* 2012; 130:861–8.e7. [PubMed: 22694930]
21. Torgerson DG, Ampleford EJ, Chiu GY, Gauderman WJ, Gignoux CR, Graves PE, et al. Meta-analysis of genome-wide association studies of asthma in ethnically diverse North American populations. *Nat Genet.* 2011; 43:887–92. [PubMed: 21804549]
22. Lee PH, O'Dushlaine C, Thomas B, Purcell SM. INRICH: interval-based enrichment analysis for genome wide association studies. *Bioinformatics.* 2012; 28:1797–9. [PubMed: 22513993]
23. Hunt KA, Zernakova A, Turner G, Heap GA, Franke L, Bruinenberg M, et al. Newly identified genetic risk variants for celiac disease related to the immune response. *Nat Genet.* 2008; 40:395–402. [PubMed: 18311140]
24. Dubois PC, Trynka G, Franke L, Hunt KA, Romanos J, Curtotti A, et al. Multiple common variants for celiac disease influencing immune gene expression. *Nat Genet.* 2010; 42:295–302. [PubMed: 20190752]
25. Hirschfield GM, Liu X, Xu C, Lu Y, Xie G, Gu X, et al. Primary biliary cirrhosis associated with HLA, IL12A, and IL12RB2 variants. *N Engl J Med.* 2009; 360:2544–55. [PubMed: 19458352]
26. Liu X, Invernizzi P, Lu Y, Kosoy R, Bianchi I, Podda M, et al. Genome-wide meta-analyses identify three loci associated with primary biliary cirrhosis. *Nat Genet.* 2010; 42:658–60. [PubMed: 20639880]
27. Mells GF, Floyd JA, Morley KI, Cordell HJ, Franklin CS, Shin SY, et al. Genome-wide association study identifies 12 new susceptibility loci for primary biliary cirrhosis. *Nat Genet.* 2011; 43:329–32. [PubMed: 21399635]
28. De Jager PL, Jia X, Wang J, de Bakker PI, Ottoboni L, Aggarwal NT, et al. Meta-analysis of genome scans and replication identify CD6, IRF8 and TNFRSF1A as new multiple sclerosis susceptibility loci. *Nat Genet.* 2009; 41:776–82. [PubMed: 19525953]
29. Hom G, Graham RR, Modrek B, Taylor KE, Ortmann W, Garnier S, et al. Association of systemic lupus erythematosus with C8orf13-BLK and ITGAM-ITGAX. *N Engl J Med.* 2008; 358:900–9. [PubMed: 18204098]
30. Graham RR, Cotsapas C, Davies L, Hackett R, Lessard CJ, Leon JM, et al. Genetic variants near TNFAIP3 on 6q23 are associated with systemic lupus erythematosus. *Nat Genet.* 2008; 40:1059–61. [PubMed: 19165918]
31. Han JW, Zheng HF, Cui Y, Sun LD, Ye DQ, Hu Z, et al. Genome-wide association study in a Chinese Han population identifies nine new susceptibility loci for systemic lupus erythematosus. *Nat Genet.* 2009; 41:1234–7. [PubMed: 19838193]
32. Radstake TR, Gorlova O, Rueda B, Martin JE, Alizadeh BZ, Palomino-Morales R, et al. Genome-wide association study of systemic sclerosis identifies CD247 as a new susceptibility locus. *Nat Genet.* 2010; 42:426–9. [PubMed: 20383147]

33. Stahl EA, Raychaudhuri S, Remmers EF, Xie G, Eyre S, Thomson BP, et al. Genome-wide association study meta-analysis identifies seven new rheumatoid arthritis risk loci. *Nat Genet.* 2010; 42:508–14. [PubMed: 20453842]
34. Kochi Y, Okada Y, Suzuki A, Ikari K, Terao C, Takahashi A, et al. A regulatory variant in CCR6 is associated with rheumatoid arthritis susceptibility. *Nat Genet.* 2010; 42:515–9. [PubMed: 20453841]
35. Bosse Y, Hudson TJ. Toward a comprehensive set of asthma susceptibility genes. *Annu Rev Med.* 2007; 58:171–84. [PubMed: 16907639]
36. N'Diaye A, Chen GK, Palmer CD, Ge B, Tayo B, Mathias RA, et al. Identification, replication, and fine-mapping of Loci associated with adult height in individuals of African ancestry. *PLoS Genet.* 2011; 7:e1002298. [PubMed: 21998595]
37. Lango Allen H, Estrada K, Lettre G, Berndt SI, Weedon MN, Rivadeneira F, et al. Hundreds of variants clustered in genomic loci and biological pathways affect human height. *Nature.* 2010; 467:832–8. [PubMed: 20881960]
38. Tantisira KG, Small KM, Litonjua AA, Weiss ST, Liggett SB. Molecular properties and pharmacogenetics of a polymorphism of adenylyl cyclase type 9 in asthma: interaction between beta-agonist and corticosteroid pathways. *Hum Mol Genet.* 2005; 14:1671–7. [PubMed: 15879435]
39. Hardin M, Zielinski J, Wan ES, Hersh CP, Castaldi PJ, Schwinder E, et al. CHRNA3/5, IREB2, and ADCY2 are associated with severe COPD in Poland. *Am J Respir Cell Mol Biol.* 2012; 47:203–8. [PubMed: 22461431]
40. Warburton D, Bellusci S, De Langhe S, Del Moral PM, Fleury V, Mailleux A, et al. Molecular mechanisms of early lung specification and branching morphogenesis. *Pediatr Res.* 2005; 57:26R–37R.
41. Massaro GD, Massaro D, Chan WY, Clerch LB, Ghyselinck N, Chambon P, et al. Retinoic acid receptor-beta: an endogenous inhibitor of the perinatal formation of pulmonary alveoli. *Physiol Genom.* 2000; 4:51–7.
42. Favre CJ, Mancuso M, Maas K, McLean JW, Baluk P, McDonald DM. Expression of genes involved in vascular development and angiogenesis in endothelial cells of adult lung. *Am J Physiol Heart Circ Physiol.* 2003; 285:H1917–38. [PubMed: 12842817]
43. Chen H, Herndon ME, Lawler J. The cell biology of thrombospondin-1. *Matrix Biol.* 2000; 19:597–614. [PubMed: 11102749]
44. Qi JH, Ebrahem Q, Moore N, Murphy G, Claesson-Welsh L, Bond M, et al. A novel function for tissue inhibitor of metalloproteinases-3 (TIMP3): inhibition of angiogenesis by blockage of VEGF binding to VEGF receptor-2. *Nat Med.* 2003; 9:407–15. [PubMed: 12652295]
45. Tien ES, Davis JW, Vanden Heuvel JP. Identification of the CREB-binding protein/ p300-interacting protein CITED2 as a peroxisome proliferator-activated receptor alpha coregulator. *J Biol Chem.* 2004; 279:24053–63. [PubMed: 15051727]
46. Sharma S, Poon A, Himes BE, Lasky-Su J, Sordillo JE, Belanger K, et al. Association of variants in innate immune genes with asthma and eczema. *Pediatr Allergy Immunol.* 2012; 23:315–23. [PubMed: 22192168]
47. Moon CM, Cheon JH, Kim SW, Shin DJ, Kim ES, Shin ES, et al. Association of signal transducer and activator of transcription 4 genetic variants with extra-intestinal manifestations in inflammatory bowel disease. *Life Sci.* 2010; 86:661–7. [PubMed: 20176035]
48. Sumino K, Tucker J, Shahab M, Jaffee KF, Visness CM, Gern JE, et al. Antiviral IFN-gamma responses of monocytes at birth predict respiratory tract illness in the first year of life. *J Allergy Clin Immunol.* 2012; 129:1267–73. [PubMed: 22460071]
49. Mallia P, Johnston SL. How viral infections cause exacerbation of airway diseases. *Chest.* 2006; 130:1203–10. [PubMed: 17035457]
50. Bosco A, Ehteshami S, Stern DA, Martinez FD. Decreased activation of inflammatory networks during acute asthma exacerbations is associated with chronic airflow obstruction. *Mucosal Immunol.* 2010; 3:399–409. [PubMed: 20336062]

51. Message SD, Laza-Stanca V, Mallia P, Parker HL, Zhu J, Keadze T, et al. Rhinovirus-induced lower respiratory illness is increased in asthma and related to virus load and Th1/2 cytokine and IL-10 production. *Proc Natl Acad Sci U S A*. 2008; 105:13562–7. [PubMed: 18768794]
52. Vignali DA, Kuchroo VK. IL-12 family cytokines: immunological playmakers. *Nat Immunol*. 2012; 13:722–8. [PubMed: 22814351]

Key messages

- Genes involved in airway structure/remodeling are associated with lung function in both general populations and subjects with asthma.
- T_H1 pathway genes involved in anti-virus/bacterial infection and inflammation modify lung function in asthmatic subjects.

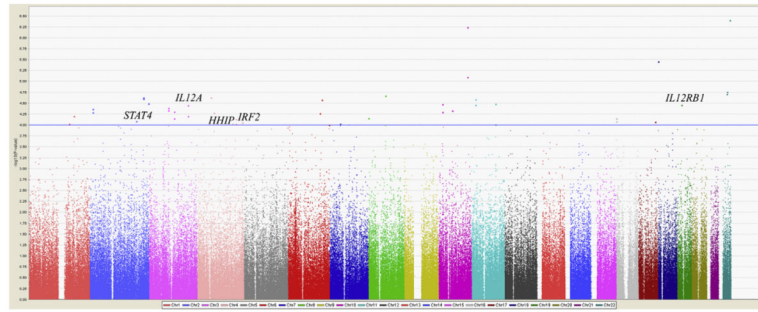


FIG 1. Genome-wide association of ppFEV₁ values with 369,771 SNPs in 4 European American populations. The *color scale* of the *x-axis* represents chromosomes. Negative log-transformed meta-analysis *P* values are shown on the *y-axis*. The *blue horizontal line* is drawn at a *P* value of 10^{-4} .

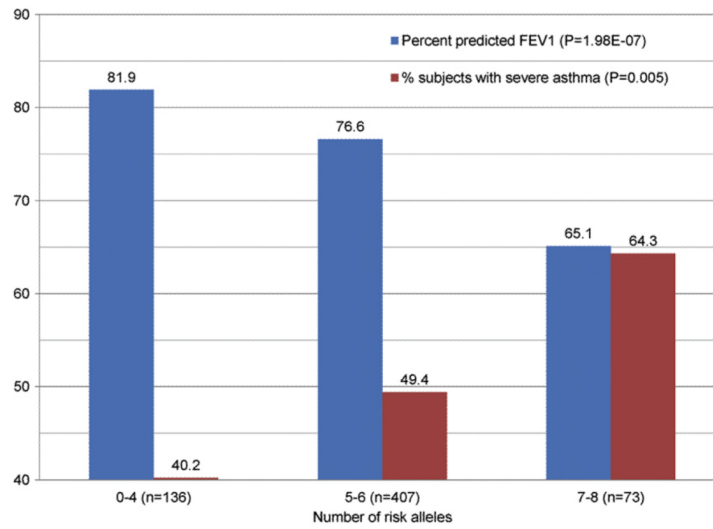
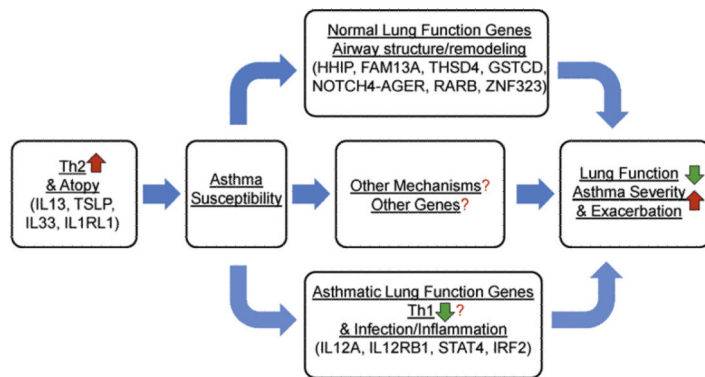


FIG 2. Joint analysis of 4 T_H1 pathway SNPs in *IL12A*, *IL12RB1*, *STAT4*, and *IRF2* for ppFEV₁ values in the SARP population. *Blue bars* represent ppFEV₁, and *purple bars* represent percentages of subjects with severe asthma based on ATS classification.

**FIG 3.**

Two-step asthma progression genetic model. The *red arrow* indicates higher gene expression levels or protein activities. The *green arrow* indicates lower gene expression levels or protein activities. The *question mark* indicates the lack of available experimental evidence.

TABLE I

Meta-analysis results of 4 T_H1 pathway SNPs associated with ppFEV₁

SNP	Gene	Chromosome	BP	Location	Ref/alt allele	Alt freq	Alt effect	Direction [*]	P value
rs388159	<i>IL12RB1</i>	19	18002996	3' [†]	C/T	0.1767	0.1943	++++	3.47E-05
rs7636840	<i>IL12A</i>	3	161161335	5' [‡]	G/T	0.1725	-0.1945	----	3.58E-05
rs925847	<i>STAT4</i>	2	191605785	Intron	C/T	0.2774	0.1579	++++	8.17E-05
rs3756089	<i>IRF2</i>	4	185552786	Intron	C/T	0.0818	0.2595	++++	8.79E-05

* Direction: +/- indicates that the alternative allele is correlated with ppFEV₁ values positively/negatively in the 4 cohorts, and the order of the populations is SARP, CSGA, ACRN, and TENOR.

[†] rs388159 is located between *IL12RB1* and *ARDDC2*.

[‡] rs7636840 is located between *IL12A* and *SCHIP1*.

TABLE IIFour TH1 pathway SNPs in a prediction model for ppFEV₁ values

SNP	Gene	SARP	CSGA	ACRN	TENOR
rs7636840	<i>IL12A</i>	0.0031	0.93	0.0031	0.15
rs388159	<i>IL12RB1</i>	0.0028	0.28	0.15	0.011
rs925847	<i>STAT4</i>	0.032	0.00087	0.49	0.055
rs3756089	<i>IRF2</i>	0.009	0.079	0.0035	0.45
Linear regression (<i>P</i> value/ <i>R</i> ²)*		9.30E-06/.046	0.0043/.078	0.00063/.065	0.013/.029
Meta-analysis <i>P</i> value [†]			3.31E-11		

* Linear regression (*P*value/*R*²): linear regression of ppFEV₁ values versus 4 TH1 pathway SNPs.

[†] Meta-analysis *P*value: Meta-analysis of *P*values with weights proportional to sample size.