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**Differential predation alters pigmentation in threespine stickleback
(*Gasterosteus aculeatus*)**

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We thank the two anonymous reviewers and the associate editor for their thoughtful comments. Below you will find each of the comments made by the reviewers (in bold) and the action that we have taken to satisfy any concerns (marked by >).

Reviewer 1:

The manuscript reports a study that investigates the effects of predation on specific colour pattern elements in threespine stickleback. The authors use first and second generation hybrids between two ecotypes (benthic and limnetic) in predator and non-predator treatments. The results show that predation had an effect on the striped colour pattern element, with a higher contrast of stripes on individuals in the predator treatment.

Overall, I found the manuscript to be interesting and largely well written. I have a few concerns which I have detailed below in the order in which they appear in the manuscript.

1) With regards to the use of the term ‘cryptic colouration’ early in the introduction (line 41). Initially there is no justification for the stripe/green colouration to be assigned as ‘cryptic’. A justification comes later in the introduction (line 90) but this two points need to be more strongly reconciled for the introduction to make sense to the reader. I see no mention of how these colours might be perceived by the predators which is the basis of the manuscript. In order to understand how predation is acting on visual signals it is standard to incorporate the visual parameters of the trout (or a close relative). In this case, I am willing to accept that the colours being investigated are at least detectable by the predators, however, this needs to be stated in the manuscript (preferably in the introduction, by stating something like ‘our study does not determine how this colouration is perceived by the predator, but it is reasonable to assume that these colours can be seen by the predator based on.....’).

>The words cryptic and conspicuous have been removed from the early part of the manuscript’s introduction and downplayed throughout the manuscript.

>There is strong evidence to suggest cutthroat trout are able to perceive a wide variety of wavelengths of light (and correspondingly colours). Close relatives of the cutthroat trout (e.g. rainbow trout and most species of anadromous salmon) have been shown to possess (and express) five or six opsin genes which allow the perception of most wavelengths of light (Rennison et al., 2012). Correspondingly, these species are predicted to be at least tetrachromatic. Visual cues are also known to be key for trout foraging (Mazur & Beauchamp, 2003). Stickleback express four opsin genes (Rennison et al., 2015), thus trout would likely have as good or better wavelength discriminatory abilities. A statement indicating this has been incorporated in the introduction – **Line 59 onwards.**

2) There is no mention of the relative proportions of the phenotypes at the start of the experiment. The assumption is that all of the ponds started with the same abundance of each phenotype, but there is no mention of this in the text. It seems entirely possible that an unfortunate experimental bias with regards to the starting populations resulted in some of the effects seen. Were any steps taken to control the frequency of phenotypes in each pond pair? I can see how determining the frequency of the intermediate phenotypes would be difficult, but a line or two addressing this seems like a fundamental addition (as it is, the authors refer to ‘a few’ of each phenotype – line 120).

>Unfortunately, it is impossible to know the precise starting frequencies of the phenotypes because the traits we were interested were not expressed at the start of the experiment prior to selection (i.e. when the fish were fry or very young juveniles) and the nuptial coloration is only expressed during the breeding season; this is why we surveyed adults and made our collection in the breeding season. However, all of the F₁ fish were intermediate in these traits and were heterozygous for loci that are differentiated between their pure benthic and limnetic parents (i.e. at tens of thousands if not hundreds of thousands of loci – genome sequencing suggests that large portions of the benthic and limnetic genomes harbour fixed differences (Schluter unpublished)). Pigmentation traits are also likely to be highly polygenic - thus heavily skewed trait distributions would be unlikely to be generated from breeding events between heterozygous parents.

>Given that full-sibling F₁s were evenly distributed between a pair of ponds it is unlikely that there would arise significantly differential trait distributions between paired ponds. Since we had 4 replicate pairs of ponds it would be particularly unlikely that all treatment ponds would show the same random skew by chance. Many previous benthic-limnetic F₂ crosses (e.g. Arnegard *et al.*, 2014; Conte *et al.*, 2015) have shown that there are individuals produced that are very benthic in their phenotype, individuals that are very limnetic, and individuals that have phenotypes intermediate relative to either pure ecotype. A statement has been added to reflect this information – **Lines 175 onwards**

Additionally, the reader has no idea how big the starting populations, (‘F₁ fish were left to reproduce naturally in the ponds..’ – line 113) or the final populations, were in each pond. Because of this the authors are asking the readers to assume that predation actually occurred (rather than differential mortality or sampling biases for example).

>The population sizes of each pond were estimated through mark recapture at several points during the experiment (Rudman *et al.*, 2016). On average there were 1834 fish per pond at the beginning of the experiment. There was not a significant difference in population size between the trout and control treatments. Just preceding the survey of colouration the control ponds had an average population size of 1262 fish, a ~25%

reduction in population size. In contrast the trout predation ponds had an average population of 710 fish, a ~65% reduction in population size. We believe that the stark difference in population declines between treatments combined with visual observations of successful predation events provide strong evidence for actual predation. Statements indicating the starting and pre-sampling population size, and evidence of predation have now been added – [Line 163](#).

3) I found the first two paragraphs of the results section a little difficult to follow. As it is the reader has to jump between figures in order to follow the current format. Please can this be rectified by referring to each of the figures in the order that they appear.

>The results have now been reordered to make this more intuitive.

4) Eye blueness is discussed in the results and discussion. I would really like to see the author's predictions of the effects of predation on eye blueness included briefly in the introduction (if that is what is meant by 'nuptial colouration' in line 76 please can the authors be more specific).

>This has been clarified [Lines 100-101 & 126-131](#)

5) The discussion fails to put the study into a broader context regarding predation and colouration. This would be a nice addition to the manuscript, at the moment the discussion has too much of a narrow focus in my opinion. Also, how do differences in body shape between the limnetic and benthic phenotypes contribute to the results? This should be discussed as surely some phenotypes could escape predation more effectively.

>The breadth of discussion has been widened. We now discuss the implications of our turbidity findings with regards to more general shifts in fish community and discuss animal crypsis more generally. Since we didn't collect body shape data from these individuals we cannot say whether or how shape and colour co-varied. In general, benthic and limnetic fish have different body shapes and some of these differences are thought to be due to predation – limnetics are streamlined whereas benthics are thought to have tail build for quick burst swimming. A sentence has been added to the discussion addressing this [Line 328](#).

Additional comments:

Line 34: Any colour trait (structural or pigmented) that contributes to crypsis can be favoured by natural selection (not just pigment based colour).

>The word structural has been added here to reflect this.

Line 40: Colour conspicuousness is a function of the visual system viewing it.

Here the authors imply that they are testing conspicuous colouration but immediately below the authors write about cryptic colouration. The introduction needs to talk about components of a colour pattern rather than assigning them to being cryptic or conspicuous without providing the justification.

>The word crypsis and its discussion has been moved towards the end of the introduction and is now more explicitly and thoroughly addressed throughout the manuscript.

Line 41: I don't understand why the authors assume that pigment traits are cryptic. Whether a colour is cryptic or not really depends on who is viewing it. It would be better to talk about the potential function of colours rather than prematurely deciding that they are cryptic (after all, the authors state whether these colours are cryptic this has not been tested – starting line 63).

>As addressed above the word crypsis and its discussion has been moved towards the end of the introduction and is now more explicitly and thoroughly addressed.

Line 49: Please mention that these species are sympatric somewhere in the introduction. It is currently implies rather than stated.

>This has been added to [line 48](#).

Line 73: The authors are not showing 'divergence' here, but rather differences.

>The word divergence has been changed to differences.

Line 74: I am missing the justification for these predictions. What makes stripes and green pigmentation cryptic in this system? Is it because of the microhabitat, the visual system of the predators, prey/predator behaviour? There are a number of reasons that colours can be considered conspicuous or cryptic and this is often dependent on the context under which they are viewed. The justification of the predictions needs to be developed further.

>Clearer and evidence-based predictions are now explicitly made – [Lines: 93 – 102, 126-130](#).

Line 90: If this is the justification that I was missing in the previous comment please can the last two paragraphs of the introduction be rearranged so that the justification of the predictions and the predictions are more closely linked. Also, this statement needs a reference; what is the evidence that 'few patterns or colours would be conspicuous in highly turbidity environments..'?

>This is now more explicitly linked to the predictions as outlined above ([Lines: 93 – 102, 126-130](#))

Line 96: This sentence reads as though the ponds would be greener and more striped.

>This sentence has been revised to reflect our intended meaning – that the *fish* in these ponds would be more striped and green.

Line 106: How were the families ‘split’? Was it randomly? By phenotype? This is actually quite an important detail given that this step can strongly dictate the results.

>The F₁ fish were all intermediate in phenotype, thus the family was split in half randomly. A statement indicating this has been added – [Line 146](#)

Line 107: The ponds didn’t receive the same number of individuals; they received between 23 and 31. Please remove ‘the same number of individuals’ from this sentence.

>Each pond *within* a pair received the same number of fish. The number of fish between pairs varied. The sentence has been revised to reflect this.

Line 113: Were F1 fish remove from the tanks? If so, this isn’t clear. If not, then the authors are not testing the effects on the F2 generation alone as implied in the introduction (line 69).

>F₁ fish were not removed from the ponds. However, they had little or no impact on the experiment. This is because most stickleback live for only one year, correspondingly the vast majority of the F₁ fish died following the breeding season. Perhaps one or two F₁ fish survived the breeding season in each pond – in contrast there were ~1834 F₂ fish in each pond, thus any hypothetically surviving F₁ would comprise about 0.01% of the population. We are also confident that none of the fish phenotyped for colour or pigmentation were F₁ individuals. As any remaining F₁s would be substantially larger than the 10-11 month old F₂ fish. A statement indicating this has been added – [Line 158](#)

Line 120: ‘A few’ isn’t very scientific. If the authors have the number of individuals of the three phenotypes (benthic, limnetic and intermediate) they should put them here. If not then a sentence or two addressing how this was initially controlled should be added.

>The word few has been removed and as described above a statement addressing the initial distribution has been added.

Line 124: It would be useful to know how many individuals were in the tanks at the start of the experiment and at the sampling period. The strength of the predation (and that predation did occur) can be assessed by determining the difference between the population at the start and end of the experiment

(accounting for natural mortality which could be judged from the controls).

>As described above a statement addressing the initial and pre-sampling population size has been added.

Also the readers will want to know what fraction of the final population 100 individuals represents (presumably this is both the F1 and F2 generation?). Additionally, it seems important to know how these individuals were caught (given the differences in feeding behaviour and tendency to sort by depth) – could the sampling have been subject to any bias?

>As described above this was only the F2 generation. A statement on how we conducted pond sampling has been added – [Line 186](#).

Line 125: Please be explicit in stating how many generations 9-10 months represents? It is implied that this represents a single generational time period, but it is not stated. Also, why was this time period chosen?

>Stickleback generally live one year, the fish sampled were about 1 year old. The reason for choosing this time point was that this is when the fish are reproductively mature and breeding, thus expressing the nuptial coloration and pigmentation patterns we were interested in. This information has been integrated at various points in the manuscript.

Line 136: I am curious as to whether the authors considered comparing the number of individuals with and without stripes between the treatments?

>We hadn't previously considered looking at the proportion of individuals with or without stripes, the reason is that you have to define a somewhat arbitrary cut-off differential pigmentation to determine whether an individual is or is not striped. However, we did try this during the revision and it yields the same results of our more quantitative measure. 31-41% of fish in predation ponds were striped compared to 6—20%. Thus, we have kept our original quantitative measure rather than a binary one.

Line 140: Contrast should be calculated with regards to the visual system that is viewing it. At the very least this should be acknowledged and it stated that contrast was determined by absolute differences between two colours.

>To avoid confusion associated with the word contrast in the visual perception literature we have replace our use of "stripe contrast" with "degree of lateral barring", which has been used previously (e.g. by Greenwood *et al.* 2011) to describe the horizontal stripes in threespine stickleback. We now also make it clear that the degree of barring was determined by estimating the absolute differences between the two colours. [Line 209](#)

Line 145: Why was the total iris area not used? Is the outer edge the most

colourful part? Please can a line be added explaining this?

>Eye size varies tremendously among individuals, this was a way to standardize the area surveyed. Additionally, in the photos there was often glare (reflection of the illumination) at the top of the iris, the standardization was also implemented to avoid the effect of this, as it varied from fish to fish. A statement indicating this has been added – [Line 215](#)

Line 157: Please include the sample sizes for the males classed as reproductive.

>There were 163 individuals designated as reproductive males. As statement indicating this has been added – [Line 227](#)

Line 176: I am not clear what data went into the one sample t-test. Was it the paired difference between the control and treatment ponds?

>Yes, the paired differences (which is the same as a paired t-test). A statement indicating this has been added – [Line 254 onwards](#).

Line 203: There was a trend in two of the families, not four as is implied by this sentence.

>This has been revised.

Line 214: The assumption that the stripes are cryptic is again made here. The authors are testing whether stripes can be considered cryptic.

>The use of the word cryptic and the basis for our hypotheses have been revised throughout the manuscript.

With the positive correlation between eye blueness and greenness how are the authors sure that greenness isn't related to nuptial colour or reproductive status. Really it seems that the authors are investigating the functions of different aspects of a colour pattern rather than cryptic vs nuptial.

>Previous work (e.g. Clarke & Schluter, 2011) has clearly shown that both male and female fish exhibit green pigmentation (*i.e* unlike blue eyes it isn't sexually dimorphic) and that fish exhibit this trait outside of the breeding season. One explanation for the correlation could be that fish that are in good enough condition to maintain bright dorsal pigmentation can also maintain bright blue eye pigmentation (*i.e.* both traits are to some degree condition dependent). We agree these are different aspects of colour pattern and we now try not to contrast cryptic vs nuptial as much throughout the manuscript.

Line 246: If red pigmentation indicates reproductively active individuals, I don't understand how nuptial colour was measured in reproductively active males if

there weren't enough males with red pigmentation in the populations. I am clearly missing something! Whatever that is, please can it be added to the methods or discussion.

>Red is indeed one nuptial colour trait; however, it can be a relatively rare phenotype in some populations (Reimchen, 1989 Evolution) (and even expressed in females in other populations). In wild benthic and limnetic populations many male individuals in reproductive condition (i.e. exhibiting the pigmented eye, with mature testis and building nests) do not exhibit red coloration. Whether due to genetic or plastic (e.g. condition or parasite load (Bolnick et al., 2015 PloS One)) effects a relatively small fraction of the males in our experiment exhibited red throat pigmentation. Since so few individuals exhibited the trait in either treatment we choose not to conduct an analysis of this trait due to the small sample size. A section in the methods has been expanded to explain this more thoroughly. [Lines 232 - 238](#)

Reviewer: 2

Comments to the Author General comments:

In this interesting study, the authors explored whether differential predation by trout contributes to differences in pigmentation in sticklebacks. The authors used a within-generation selection experiment on F2 benthic-limnetic hybrids. After 10 months, they compared the pigmentation of fish under trout predation to control fish and found that stickleback were more striped in ponds with trout. Fish in ponds with trout foraged more on benthic invertebrates, which released zooplankton from predation and decreased phytoplankton abundance, which in turn decreased turbidity. The authors found that greater stripe contrast was negatively correlated with the magnitude of turbidity across pond replicates. A more benthic diet, which they used as a proxy for habitat use, was also correlated with greater stripe contrast and green dorsal pigmentation. These patterns suggest that differential exposure to predation, and the cascading effects on turbidity and habitat use, may explain divergence in cryptic body pigmentation between benthic and limnetic ecotypes.

The across-generation experimental approach is excellent, and the system is ideal for testing the hypotheses. Photographing 100 individuals per pond is also a strength of the work that allowed the authors to get precise estimates of their response variables. The manuscript is also well-written and the study appears to have been carried out well.

My primary concerns are: 1) I found the description of the statistical methods to be somewhat vague and confusing, and think that more detail of analyses is needed; 2) I wonder if the number of analyses could be reduced by using ANCOVA (see comment below); and 3) the discussion is almost entirely focused on sticklebacks, and for a broad journal, I would expect more discussion of the

broad implications to the field.

Minor comment: L 160-165: Some additional details of when this occurred and how often turbidity was measured would be helpful.

>The measurement was taken the month before the phenotyping was done – a statement indicating this was added to [Line 240](#)

L 172-180: I find this description to be vague (or possibly just awkwardly worded). For example, you state that ‘Significance testing of pigmentation treatment effects was done using one-sample t-tests’. Would it be possible to state what you are testing for biologically in the stats description? E.g. ‘To determine if trout predation influenced pigmentation, we used t-tests in which we determined if the control and predation ponds differed in striping.’ (or something similar).

>This has been revised as suggested.

L 174-175, ‘Treatment effects were estimated within each of the four F1 families (i.e. within the control and predation ponds that were paired)’: Does this mean that individual fish are being treated as replicates? Or are ponds your level of replication?

>Ponds are the replicate as indicated by the 3 degrees of freedom for our test statistics. This has been clarified in the methods section – [Line 258](#)

Also, does this mean that a separate analysis was run for each family (which seems to be implied in L 194 but not elsewhere)? If so, it seems like a single analysis for each response variable would be more appropriate. For example, why not run a single ANOVA (or ANCOVA) for each response variable in which family and predation treatment are factors and stripe contrast? I would think that turbidity and diet could also be included as covariates in such a model. Perhaps I am missing or misunderstanding something, though.

>No, a single test was run for each trait. This has been clarified in the methods section – [Line 256](#).

L 231-232: Good point.

>Thank you.

Discussion in general: The discussion focuses largely on sticklebacks. Do your findings relate to work in other systems? Are there any bigger-picture implications to our understanding of the expression of variation in general?

>The discussion has now been thoroughly re-written in an attempt to broaden the implications of the findings.

1 **Abstract**

2 Animal pigmentation plays a key role in many biological interactions, including
3 courtship and predator avoidance. Sympatric benthic and limnetic ecotypes of
4 threespine stickleback (*Gasterosteus aculeatus*) exhibit divergent pigment patterns.
5 To test whether differential predation by cutthroat trout contributes to the differences
6 in pigmentation seen between the ecotypes, we used a within-generation selection
7 experiment on F₂ benthic-limnetic hybrids. After 10 months of differential selection,
8 we compared the pigmentation of fish under trout predation to control fish not
9 exposed to trout predation. We found that stickleback exhibited more lateral barring
10 in ponds with trout predation. Ponds with trout were also less turbid, a greater degree
11 of barring was negatively correlated with the magnitude of turbidity across pond
12 replicates. A more benthic diet, a proxy for habitat use, was also correlated with
13 greater lateral barring and green dorsal pigmentation. These patterns suggest that
14 differential exposure to cutthroat trout predation may explain divergence in body
15 pigmentation between benthic and limnetic ecotypes.

16 **Key words:** natural selection, pigmentation, patterning, adaptation, species
17 interactions

18 **Introduction**

19 Colouration and pigmentation patterns have long been considered important traits in
20 animals (Dice & Blossom, 1937; Endler, 1978), as these traits are well known to
21 mediate intra- and inter-specific interactions. In many species, different patches of
22 colour across an animal's body enable an individual to distinguish its own species
23 from another and among the individuals of its own species. For example, male
24 nuptial colouration influences reproductive outcomes in many taxa; often females
25 prefer brightly coloured males over dull ones (Ciccotto & Mendelson, 2016), and
26 colouration can indicate quality or reproductive status (Houde, 1987). Colouration
27 can also be important for mediating the outcome of interspecific interactions such as
28 predation (Godin & McDonough, 2003). Body colouration is often used for
29 camouflage, where species have adapted to their environment in such a way that
30 they are matched to their surroundings and can avoid detection by a predator
31 (Endler, 1978; Slagsvold & Dale, 1995, Sherratt *et al.*, 2004).

32 Pigment and structural traits that function in predator avoidance are predicted
33 to be favoured by natural selection in the presence of visual predators, while
34 conspicuous visual signals, such as bright nuptial colours that attract mates, are
35 thought to be selected against when visual predators are present (Endler, 1983). A
36 cost of conspicuous male ornamentation has been shown in guppies, where fish
37 under higher predation pressures have evolved duller colouration (Godin &
38 McDonough, 2003). However, it remains unclear how often bright colouration is
39 disfavoured and cryptic colouration favoured. Here, we sought to test whether two
40 non-reproductive pigment traits and one nuptial pigment trait were favoured or
41 disfavoured in the presence of predators, and whether pigment traits evolve
42 independently. To determine the effect of predation-based natural selection on

43 pigment traits, we conducted a manipulative experiment using hybrid benthic-limnetic
44 threespine stickleback that varied in pigmentation. By manipulating the presence or
45 absence of a visual predator, we could make progress in identifying the mechanisms
46 driving the evolution of pigmentation. We also used the experimental design to
47 assess how habitat use and turbidity influence pigmentation.

48 Sympatric benthic and limnetic threespine stickleback (*Gasterosteus*
49 *aculeatus*) are an excellent system to examine the interaction between pigmentation
50 and predation-based natural selection. Benthic and limnetic stickleback exhibit
51 divergent pigmentation for two male nuptial traits, a red throat patch and blue iris; for
52 both traits male limnetics are generally brighter and more colourful than male
53 benthics (Boughman *et al.*, 2005; Albert *et al.*, 2007). Year-round there is variation
54 between the species in body colouration and lateral barring (black vertical stripes)
55 (Clarke & Schluter, 2011; Greenwood *et al.*, 2011). Benthic and limnetic stickleback
56 also experience differential predation (Schluter & McPhail, 1992); benthic fish are
57 primarily preyed upon by invertebrate predators, whereas limnetic fish are primarily
58 preyed upon by cutthroat trout (*Onchorhynchus clarkii*) (Schluter & McPhail, 1992).

59 The two different suites of predators that each species is exposed to have
60 distinct prey detection methods. Cutthroat trout use vision as a core sensory system
61 for prey detection; trout are known to rely heavily on visual cues during pursuit of
62 their prey (Vogel & Beauchamp, 1999) and hunting success declines with increasing
63 turbidity (Vogel & Beauchamp, 1999; Mazur & Beauchamp, 2003). Cutthroat trout are
64 predicted to be tetrachromatic (Bowmaker and Kunz, 1987; Rennison *et al.*, 2012),
65 and thus should be able to detect a wide variety of wavelengths and discriminate
66 among a multitude of colours. In contrast, the invertebrate predators of threespine
67 stickleback are largely ambush predators and are less dependent on visual cues for

68 prey detection (Foster *et al.*, 1988). Thus, exposure to these distinct predators could
69 contribute to divergence in the colouration and patterning of benthic and limnetic
70 species.

71 Differences between benthic and limnetic stickleback in non-reproductive
72 colouration and patterning have been hypothesized to be important for camouflage in
73 the presence of vertebrate predators (Clarke & Schluter, 2011; Greenwood *et al.*,
74 2011), but this has not been directly tested. The dorsal colouration of benthic
75 stickleback is more closely matched (*i.e.* has less contrast) to the littoral background,
76 than that of limnetic stickleback (Clarke & Schluter, 2011); this suggests that within
77 the littoral habitat the green dorsal colouration of benthics may be more cryptic than
78 the limnetic colouration. Neither species shows significant pigment matching to the
79 pelagic background (Clarke & Schluter, 2011). The lateral barring exhibited by
80 stickleback may play a role in predation avoidance either through background
81 matching in a spatially complex environment (Josef *et al.*, 2012), as disruptive
82 colouration (Cuthill *et al.*, 2005), or through motion dazzle camouflage (when high-
83 contrast geometric patterns interrupt the motion detection systems of a visual
84 predator) (Thayer, 1909). A variety of factors have been hypothesized to underlie
85 reduced nuptial colouration in some stickleback populations, including differential
86 predation pressure (Semler, 1971), increased turbidity and carotenoid deficiency
87 (Reimchen, 1989), yet direct tests of these hypotheses have been lacking.

88 To determine the effect of differential predation on pigmentation traits we used
89 hybrid F₂ benthic - limnetic stickleback in a selection experiment conducted under
90 semi-natural conditions in artificial ponds. Four experimental ponds were exposed to
91 cutthroat trout predation and four ponds were kept as trout-free controls. After ten
92 months of differential predation, differences in colour and the degree of lateral barring

93 were estimated. Based on the observed matching between benthic colouration
94 (green dorsal pigmentation) and the littoral habitat (Clarke & Schluter, 2011), we
95 predicted that green pigmentation would be favoured in the trout predation treatment
96 where background matching may be more beneficial. The hypothesized role of lateral
97 barring in predation avoidance, led us to predict that barring should be more common
98 in the presence of vertebrate predation. Based on previous work suggesting that
99 bright nuptial colouration is selected against in presence of predation (e.g. Semler,
100 1971; Endler, 1978), we predicted that the bright blue eye displayed by many
101 reproductive males would be disfavoured in the trout predation treatment.

102 The visual environment under which pigment signals are viewed is an
103 important determinate of whether a signal appears to be cryptic or conspicuous
104 (Hemmings, 1965); this is because visibility depends on the contrast between a
105 signal, the background it is viewed upon and any medium between the two objects
106 (Hemmings, 1965). Increased turbidity is one factor that can reduce the visibility of
107 visual displays and signals by diminishing the contrast between an object and the
108 background; this is due to the scattering of light, and through an overall reduction of
109 light penetrance within the water column (Lythgoe, 1979; Utne-Palm, 2002). As a
110 result, under turbid conditions signals that would have high contrast and appear
111 bright in clear water may appear less conspicuous. Previous work has shown that
112 increased turbidity leads to a reduction in bright nuptial colouration (Reimchen, 1989;
113 Seehausen *et al.*, 1997) and reduced reliance on colour based signals during
114 courtship (Luyten & Lily, 1985; Seehausen *et al.*, 1997; Engström-Öst & Candolin,
115 2007).

116 To further explore our hypothesis that lateral barring and dorsal pigmentation
117 could be beneficial for predation avoidance (camouflage), we considered the effect of

118 turbidity and diet (a proxy for habitat usage) on the magnitude of divergence in colour
119 pigmentation and patterning. In this experiment, it was previously shown that the
120 addition of cutthroat trout led to a shift in stickleback habitat use and diet, which also
121 affected the turbidity of the ponds (Rudman *et al.*, 2016). The shift in turbidity was the
122 result of a trophic cascade: in ponds with trout, the stickleback foraged more on
123 benthic invertebrates, which released zooplankton from predation and decreased
124 phytoplankton abundance, thereby decreasing turbidity (Rudman *et al.*, 2016). The
125 opposite was seen in control ponds, where stickleback foraged more heavily on
126 zooplankton. Given the observed differences in turbidity between the treatments
127 (Rudman *et al.*, 2016) and the known effect of increased turbidity on the visibility of
128 pigment patterns and bright colouration (*e.g.* Hemmings, 1965), we predicted that
129 under turbid conditions the utility of bright nuptial colouration in mate displays would
130 be reduced and the necessity for pigmentation that aids in camouflage would be
131 lessened. To determine whether habitat use affected colouration, we examined the
132 relationship between diet (estimated by stomach contents) and pigmentation. We
133 considered the proportion of zooplankton vs benthic invertebrates in the diet, as this
134 would indicate where fish most often foraged. Again, considering the observed
135 matching between green dorsal pigmentation and the littoral habitat (Clarke &
136 Schluter, 2011), we predicted that increased green pigmentation would be favoured
137 by individuals that more frequently exploited the littoral habitat.

138

139 **Methods**

140 **Experimental design**

141 In spring 2011, four benthic females were artificially crossed with four limnetic
142 male threespine stickleback from Paxton Lake (Texada Island, British Columbia) to
143 create four F₁ benthic-limnetic hybrid families. These F₁-hybrid offspring were reared
144 under common laboratory conditions in 100 L tanks for one year. In spring 2012,
145 these F₁ fish were introduced into 8 semi-natural experimental ponds located on the
146 University of British Columbia campus. Each F₁-hybrid family was randomly split in
147 half and introduced into a pair of ponds. See Supplementary Figure 1 for a depiction
148 of the experimental design. Each pond within a pair received the same number of
149 individuals. However, different pond pairs received different numbers of individuals
150 depending on the original F₁ family size (between 23 - 31 individuals were added per
151 pond). The experimental ponds were 15 x 25m in size with a maximal depth of 6m
152 (see Arnegard *et al.*, 2014 for further details on the pond structure). Each pond
153 contained a natural assemblage of food resources and vegetation. Prior to fish
154 introduction the eight ponds were paired based on count surveys of macrophyte
155 coverage, phytoplankton, zooplankton and insect abundance. In spring 2012 the F₁
156 fish reproduced naturally within the ponds to create the focal F₂-hybrid generation. In
157 September 2012 two coastal cutthroat trout (*Onchorhynchus clarkii*) were introduced
158 into one randomly chosen pond within each pond pair. The majority of the F₁ fish
159 died following the 2012 breeding season; however due to size differences between
160 two-year-old F₁ fish and the approximately one-year-old F₂ cohort we are confident
161 that any rare F₁ survivors did not contribute to the sample of fish we phenotyped for
162 the analysis.

163 At the beginning of the experiment, on average, there were 1834 F₂ fish per
164 pond, with no significant difference in the population size of fish in trout treatment
165 pond vs. control ponds (Rudman *et al.*, 2016). After 7 months of the experiment, and

166 immediately prior to the breeding season (and phenotyping time point), the
167 stickleback population size in control ponds had been reduced on average by 25%, in
168 contrast there was an average 65% reduction in population size for trout treatment
169 ponds (Rudman *et al.*, 2016). This differential mortality between treatments,
170 combined with observed predation events, provides strong evidence that the trout
171 were active predators over the course of the experiment.

172 The purpose of using hybrids in the experiment was to increase the genetic
173 variation available for selection to act upon. F_2 hybrids specifically were instrumental
174 in this study as they had experienced two generations of recombination, which
175 allowed unlinked traits to segregate independently. By establishing the ponds using
176 full-sibling F_1 crosses, that were intermediate in the pigmentation phenotypes and
177 heterozygous at loci that are differentiated between their pure benthic and limnetic
178 parents, we were able to generate F_2 individuals that exhibited phenotypic variation.
179 Previous benthic-limnetic F_2 crosses (*e.g.* Arnegard *et al.*, 2014; Conte *et al.*, 2015)
180 have shown that there are individuals produced in an F_2 cross that are very benthic in
181 their phenotype, that are very limnetic, and that have phenotypes intermediate
182 relative to either pure ecotype (with an approximately normal distribution of
183 phenotypes). The F_1 experimental design ensured that starting frequencies of each
184 phenotype would be very similar between treatment and control ponds within a pair.

185 **Pond Sampling**

186 In May and June of 2013 (after nine to ten months of natural selection) adult (~1 year
187 old) reproductively mature F_2 stickleback were caught using a combination of un-
188 baited minnow traps, open water seining, and dip netting. One hundred F_2 individuals
189 were randomly sub-sampled from all of the captured individuals from each pond (800

190 individuals total) and were retained for phenotyping before being returned to the pond
191 of origin.

192 **Phenotyping of pigmentation traits**

193 The F₂ individuals were photographed using a Nikon D300 camera with a 60mm
194 macro lens (Nikon, Melville, NY). The photos were illuminated with ambient light, the
195 camera flash and an external ring flash. The camera settings were ISO 200,
196 automatic white balance, 2.5 second exposure and F22. Prior to analysis a white
197 balance was applied in Photoshop (Adobe Creative Suite 5 and 6) to all pictures.
198 Quantitative analysis was done in ImageJ (<https://imagej.nih.gov/ij/download.html>)
199 with the additional Color_Histogram.jar plugin ([https://imagej.nih.gov/ij/plugins/color-](https://imagej.nih.gov/ij/plugins/color-histogram.html)
200 [histogram.html](https://imagej.nih.gov/ij/plugins/color-histogram.html)). From the pictures, eye colouration, dorsal colouration and the
201 degree of lateral barring were measured.

202 The degree of barring along the lateral flank was determined by estimating the
203 absolute differences between light and dark patches. This was done by selecting two
204 squares 20x20 pixels in size, with one square placed on a dark patch, the second
205 one was placed on the brighter area between two of the vertical bars. When an
206 individual did not have any barring, we selected two squares at the average distance
207 found between vertical bars when present. From these two squares the colour mode
208 (a value of brightness and intensity between 0 – 255 where 0 is black and 255 is
209 white) was recorded. We then calculated the absolute difference in mode between
210 squares; more pronounced barring yielded a higher absolute difference in brightness.
211 To evaluate dorsal colouration, we selected an area of 20x150 pixels in length and
212 placed it directly above the pectoral fin joint for consistency. Within this area, the
213 mean green pixel number (dorsal greenness) was estimated.

214 To estimate the blue colouration of the iris, a segmented line of 15-pixel width
215 was captured around the pupil, and the mean number of blue and red pixels was
216 extracted from the area. The segmented line was used to standardize the area
217 surveyed and minimize effects of light reflection off of the top of the eye. Male eye
218 blueness was estimated by dividing the mean blue pixel number by the mean red
219 pixel number. To consider whether male nuptial colouration had diverged in the
220 experiment it was necessary to classify individuals as reproductive males. From
221 photos, the sex and reproductive state of some individuals could unequivocally be
222 determined, for other individuals this was less certain. To identify all individuals that
223 were reproductive males we plotted red pigmentation against blue pigmentation
224 (both colours are indicative of male reproductive state) then used Gaussian Mixture
225 Modelling for model-based clustering, using the *mclust* package (Fraley *et al.*, 2012).
226 Using this method, we could identify two trait clusters that differentiated
227 the previously sexed individuals. We then used these clusters to classify the
228 individuals of unknown or ambiguous sex/reproductive state; there were 163
229 individuals classified as reproductive males and 639 as females or non-reproductive
230 males. We then proceeded with the nuptial colouration analysis only considering the
231 163 individuals putatively classified as reproductive males.

232 We did not evaluate red throat colour in males, which is an important mate
233 choice cue in some populations of threespine stickleback (*e.g.* Bakker & Mundwiler,
234 1992). Our reasoning for omitting red throat pigmentation was that only a small
235 proportion of males in either treatment group exhibited the trait. We are not sure why
236 red throats were rare among our pond fish, one possible explanation is parasites;
237 parasitic infections have been shown to contribute to reduced red pigmentation in
238 sticklebacks (Bolnick *et al.*, 2015).

239 **Ecological data**

240 Water turbidity was assessed in April 2013 (the month preceding the pigmentation
241 phenotyping) by measuring phytoplankton abundance using spectrofluorometry
242 ~10cm below surface. The data was then converted into $\mu\text{g l}^{-1}$ phytoplankton by
243 applying a lab standard calibration curve (see (Rudman *et al.*, 2016) for full details).
244 To quantify diet, 10 fish were collected in December 2012 from each pond using a
245 combination of dip-netting and seining. Fish were euthanized and preserved in 95%-
246 ethanol. Prey items in the stomach were identified to the lowest feasible taxonomic
247 unit and the length of each item was measured using an ocular micrometer (see
248 (Rudman *et al.*, 2016) for full details). We then used these taxonomic classification
249 data to quantify the proportion of the diet that was comprised of zooplankton. It
250 should be noted that colour measurements and stomach content data were not
251 collected from the same individuals.

252 **Statistical Analyses**

253 All analyses were done in R (R Development Core Team 2017) and R Studio version
254 3.2.3 (R Studio Team 2015). To determine if trout predation influenced pigmentation,
255 we used a paired t-test (two-sided with a null of zero); this allowed us to determine if
256 the control and predation ponds differed significantly in each pigmentation trait (*i.e.*
257 there were three tests run, one for each trait). In the analysis control and treatment
258 ponds were paired by F_1 family. Ponds were used as our level of replication; thus,
259 our test statistics are based on three degrees of freedom. To look for an association
260 between ecological data (diet and water turbidity) and pigment traits we estimated
261 correlation coefficients using Pearson's product-moment correlations.

262

263 Results

264 There was a greater degree of barring along the lateral flank of stickleback from trout
265 predation ponds relative to those from the paired control ponds (Figure 1A; mean =
266 22.55, $t_3 = 4.24$, $P = 0.024$, 95% CI: 5.64 – 39.46). Across the replicate pond pairs
267 there was no significant treatment effect on the greenness of dorsal pigmentation
268 (Figure 1B; mean = 6.85, $t_3 = 1.90$, $P = 0.15$, 95% CI: -4.62 – 18.32), although there
269 was a significant effect in two of the four pairs (Figure 1B). There was a non-
270 significant trend of a reduced blue eye colouration in reproductive males, with an
271 effect seen in two predation treatment ponds (Figure 1C; mean = -0.14, $t_3 = -2.59$, P
272 = 0.08, 95% CI: -0.31 – 0.03).

273 Among the ponds of both treatments there was a significant negative
274 relationship between the degree of barring and the proportion of zooplankton in the
275 diet (Figure 2A; $r = -0.764$, $t_6 = -2.90$, $P = 0.027$, 95% CI: -0.96 – -0.13). The extent of
276 barring was also negatively correlated with water turbidity (Figure 2B; $r = -0.903$, $t_6 = -$
277 5.16, $P = 0.0025$, 95% CI: -0.98 – -0.55). Thus, fish with a greater degree of barring
278 were found in ponds with lower turbidity and were less likely to consume
279 zooplankton; *i.e.* a more benthic habitat usage.

280 There was a significant negative correlation between the proportion of
281 zooplankton in the diet and dorsal greenness (Figure 3A; $r = -0.803$, $t_6 = -3.30$, $P =$
282 0.016, 95% CI: -0.96 – -0.23), suggesting that fish with greener backs were more
283 common in ponds where fish consumed less zooplankton. However, there was no
284 significant correlation between the dorsal greenness and water turbidity. (Figure 3B; r
285 = -0.56, $t_6 = -1.66$, $P = 0.15$, 95% CI: -0.907 – 0.2387). There was also no correlation

286 between dorsal greenness and the degree of barring ($r = -0.045$, $P > 0.05$), which
287 suggests the two pigmentation traits were evolving independently in the F_2 hybrids.

288 The environmental factors of diet and turbidity did not explain patterns of
289 divergence in eye colour; there was no significant correlation between the eye
290 colouration and water turbidity or stomach content (turbidity $r = 0.26$, $t_6 = 0.6$, $P =$
291 0.57 , 95%-CI: $-0.61 - 0.85$; diet $r = -0.19$, $t_6 = -0.42$, $P = 0.69$, 95% CI: $-0.82 - 0.66$).
292 There was also no significant correlation between the degree of barring and eye-
293 blueness ($r = -0.01$, $P = 0.91$). There was a weak positive correlation between eye-
294 blueness and dorsal greenness (Supplementary Figure 2; $r = 0.35$, $t_{161} = 4.68$, $P <$
295 0.001 , 95% CI: $0.2 - 0.47$), individuals with greener backs tended to have bluer eyes.

296 Discussion

297 *Evidence that predation causes selection on colour*

298 The aim of our study was to determine whether pigment traits in threespine
299 sticklebacks shifted in response to the introduction of a vertebrate predator. To
300 accomplish this, we used an experiment that manipulated the presence of cutthroat
301 trout, which are thought to differentially encounter wild benthic and limnetic
302 stickleback. The traits we focused on were, lateral barring and dorsal pigmentation in
303 males and females, and blue nuptial eye colouration in reproductive males. We found
304 repeated differentiation in the two non-reproductive pigmentation traits, but not in the
305 blue eye pigmentation. It seems plausible that these non-reproductive pigmentation
306 traits aided in predation avoidance through crypsis. Yet, the precise mechanisms by
307 which increased lateral barring and perhaps increased green pigmentation provide a
308 selective advantage remains to be determined.

309 In the wild, benthic and limnetic stickleback differ in their pigmentation patterns
310 (Boughman *et al.*, 2005; Albert *et al.*, 2007; Clarke & Schluter, 2011; Greenwood *et*
311 *al.*, 2011) and their exposure to cutthroat trout (Schluter & McPhail, 1992). In the
312 experiment, fish were significantly more barred in the predator treatment ponds
313 relative to paired control ponds. Despite a trend, there was no significant difference in
314 dorsal greenness between the trout and control treatments. These results suggest
315 that the presence of cutthroat trout may directly or indirectly select for increased
316 pigmentation (particularly increased lateral barring). In the wild, differential exposure
317 of benthic and limnetic stickleback to cutthroat trout may be a key mechanism
318 underlying the divergence of pigmentation between these ecotypes. This
319 corresponds with previous work which has shown that predation plays an important
320 role in explaining differences in pigmentation between species or populations (*e.g.*
321 Endler, 1991; Stuart-Fox *et al.*, 2004)

322 Experimental fish were not reared in a common garden after exposure to trout.
323 As a result, we cannot definitively say whether the shifts in pigmentation we observe
324 are due to genetic changes or a result of phenotypic plasticity. Previous work on the
325 lateral bar trait in freshwater threespine stickleback has identified quantitative trait
326 loci explaining over 30% of the variance (Greenwood *et al.*, 2011), suggesting that it
327 is heritable to some degree. The heritability of green dorsal pigmentation remains to
328 be determined. It is also important to keep in mind that there may have been
329 covariance in additional unmeasured traits, such as body shape which has been
330 hypothesized to affect the probability of escape from predation (Walker, 1997), so we
331 cannot rule out a correlated response.

332 ***Crypsis as the mechanism behind the observed colour change***

333 Evidence from a variety of taxa suggests that crypsis plays a substantial role
334 in the evolution of colour variation between populations or species (Endler, 1978).
335 There are three types of pigmentation thought to be useful for avoiding detection by
336 predators. The first is object mimicry, resemblance to a common object in the
337 environment (such as a leaf or twig) (Allen and Cooper, 1985). Object mimicry is not
338 likely to be the mechanism that stickleback would be utilizing when considering the
339 pigmentation traits examined in this study and will not be discussed here further. The
340 second is background matching (Endler 1984), when an animal takes on colouration
341 useful in blending into the local background. The third is disruptive colouration (Cott,
342 1940), which is when dark pigment elements make the detection of body shape more
343 difficult. The latter two mechanisms could plausibly contribute to the observed shifts
344 in pigment phenotype between the treatments during the experiment.

345 Background matching is an important mechanism of predation avoidance in a
346 variety of taxa (Stevens & Merilaita, 2011). In benthic stickleback background
347 matching has been suggested to underlie the advantage of green dorsal
348 pigmentation in the littoral environment (Clarke & Schluter, 2011). Our findings
349 suggest that background matching may indeed provide a selective advantage for
350 stickleback in the presence of predators. In a few cases disruptive colouration has
351 been found to increase survival in the presence of visual predators (*e.g.* Schafer &
352 Stobbe, 2006; Stevens & Cuthill, 2006), in others a lack of support for this
353 mechanism has been found (*e.g.* Silberglied *et al.*, 1980). Disruptive colouration,
354 such as striping or barring, is thought to be particularly useful for generalist taxa, as
355 they may encounter more variable visual backgrounds (Ruxton *et al.*, 2004; Sherratt
356 *et al.*, 2005). Our results suggest that lateral barring in threespine stickleback may be
357 another example of the advantage of such disruptive pigmentation; although direct

358 tests of this will be required to confirm whether this is indeed the mechanism by
359 which lateral barring confers an advantage in this species.

360 ***The role of predators in shaping nuptial colouration***

361 Nuptial colouration is often thought to be costly (Andersson, 1994); bright
362 colours in the presence of predators may bring unwanted attention and thus be
363 disfavoured in high predation environments (Zuk & Kolloru, 1998). In a variety of taxa
364 it has been shown that predators lead to duller nuptial colouration (e.g. Godin &
365 McDonough, 2003; Husak *et al.*, 2005; Giery *et al.*, 2015). We found no significant
366 difference in male eye colouration between treatments. It is possible that nuptial
367 colouration in sticklebacks is unaffected by predation. However, we cannot rule out
368 the possibility that we failed to detect divergence due to misclassification of
369 reproductive status (and perhaps sex) or due to changes in the effect over the course
370 of the breeding season. Additionally, nuptial colouration may also have been more
371 strongly affected if the experiment were conducted over a longer time period as was
372 done in guppies (Godin & McDonough, 2003); given that the experiment was
373 conducted within a generation, there may not have been sufficient time for adaptation
374 of this trait.

375 ***Correlations between colour pigments and the light environment***

376 Colouration is a visual signal which strongly depends on light transmission and
377 visibility in the water (Wilkins *et al.*, 2016) and the background upon which signals
378 are viewed (Abrahams & Kattenfeld, 1997). Ponds containing trout were less turbid
379 (Rudman *et al.*, 2016) than control ponds. In the low turbidity trout predation ponds
380 visibility would be high; as a result, fish that were greener and /or barred would likely
381 exhibit reduced contrast against background light. Under these conditions, reduced

382 contrast against the background would potentially aid in predation avoidance.
383 Consistent with this, we found that fish with lateral barring and green dorsal
384 pigmentation were favoured in the presence of cutthroat trout (although not
385 significantly for green pigmentation) and that turbidity was strongly negatively
386 correlated with both pigmentation traits.

387 In a variety of fish species increased turbidity has been shown to have
388 important implications for the expression of pigmentation traits (Reimchen, 1989;
389 Seehausen *et al.*, 1997), predation risk (Utne-Palm, 2002) and mate choice (Luyten
390 & Liley, 1985; Seehausen *et al.*, 1997; Engström-Öst & Candolin, 2007). Human
391 activities such as logging, and farming have been shown to cause eutrophication
392 (Sharpley *et al.*, 2003), which in turn can lead to increased turbidity. In this
393 experiment we show that turbidity associated with a trophic cascade can also directly
394 or indirectly affect pigmentation traits. This suggests that changes in the composition
395 of a local fish community can have broad reaching phenotypic effects that include
396 pigmentation, such shifts in pigmentation could have important secondary effects on
397 predation risk and mate choice.

398 Green dorsal pigmentation in wild benthic stickleback is well matched to the
399 littoral habitat (Clarke & Schluter, 2011). Fish in trout predation ponds exhibited a
400 more benthic diet (lower proportion of zooplankton in the diet), and presumably fed
401 more often in the littoral habitat (Rudman *et al.*, 2016). If background matching is
402 important for benthic stickleback it would be predicted that fish that spent more time
403 in the littoral habitat would have more benthic-like pigmentation (increased barring
404 and/or green dorsal pigmentation). This is indeed what we find to be the case; there
405 was a significant positive association between benthic diet, a proxy for littoral habitat
406 use (Wund *et al.*, 2012), and both pigmentation traits. Unfortunately, because

407 turbidity and habitat use (diet) covary in our study, we are unable to distinguish
408 whether one or both environmental factors mediated the proposed cryptic effects we
409 found here.

410 ***Correlations among traits***

411 To determine whether the pigmentation traits could change independently of
412 one another, we analysed the correlations between them. We found that lateral
413 barring and dorsal pigmentation were uncorrelated, and thus likely to evolve
414 independently. However, there was a weak correlation between blueness of the eye
415 and dorsal greenness. It is possible that this association constrained the divergence
416 of these traits and could explain the weaker pattern of differentiation between
417 treatments for dorsal pigmentation. Given that this experiment used F₂ hybrids, we
418 do not have the resolution needed to determine whether this association is due to
419 tight genetic linkage (which may have varied among F₁ families) or due to the
420 pleiotropic effects of a locus on both traits. Alternatively, if these traits are costly to
421 produce or maintain covariance could be explained if both traits were to some degree
422 condition dependent (*i.e.* high condition individuals were able to produce and
423 maintain a bright blue eye and green dorsal pigmentation). Further analyses must be
424 conducted to distinguish between these options.

425 ***Conclusion***

426 Using a controlled manipulative experiment, we show that lateral barring (and
427 perhaps green dorsal pigmentation) is favoured in the presence of trout (and/or
428 disfavoured in the absence of trout). We suggest the shift in lateral barring is likely
429 adaptive as it arises across independent replicates. Differential predation did not
430 have the same effect on blue eye pigmentation, a male nuptial trait that varies in the

431 wild. Lateral barring and dorsal pigmentation were associated with littoral habitat use
432 and decreased turbidity, which suggest that crypsis may be the key mechanism
433 mediating the observed shifts. These findings suggest that cutthroat trout predation
434 may be a factor contributing to the divergence of pigmentation between benthic and
435 limnetic stickleback ecotypes.

436

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445

446 **Data sharing statement**

447 All raw data will be archived in dryad doi: to be determined

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605 Figure Captions:

606

607 **Figure 1.** Effect of trout predation on four pigmentation traits. (A) Barring on the lateral
608 flank (mode-difference). (B) Dorsal greenness (mean green pixels). (C) Male eye
609 blueness (ratio of blue to red pixels). Colour is consistent across panels and indicates
610 ponds derived from the same F₁ family (paired ponds). * indicates a significant
611 treatment effect.

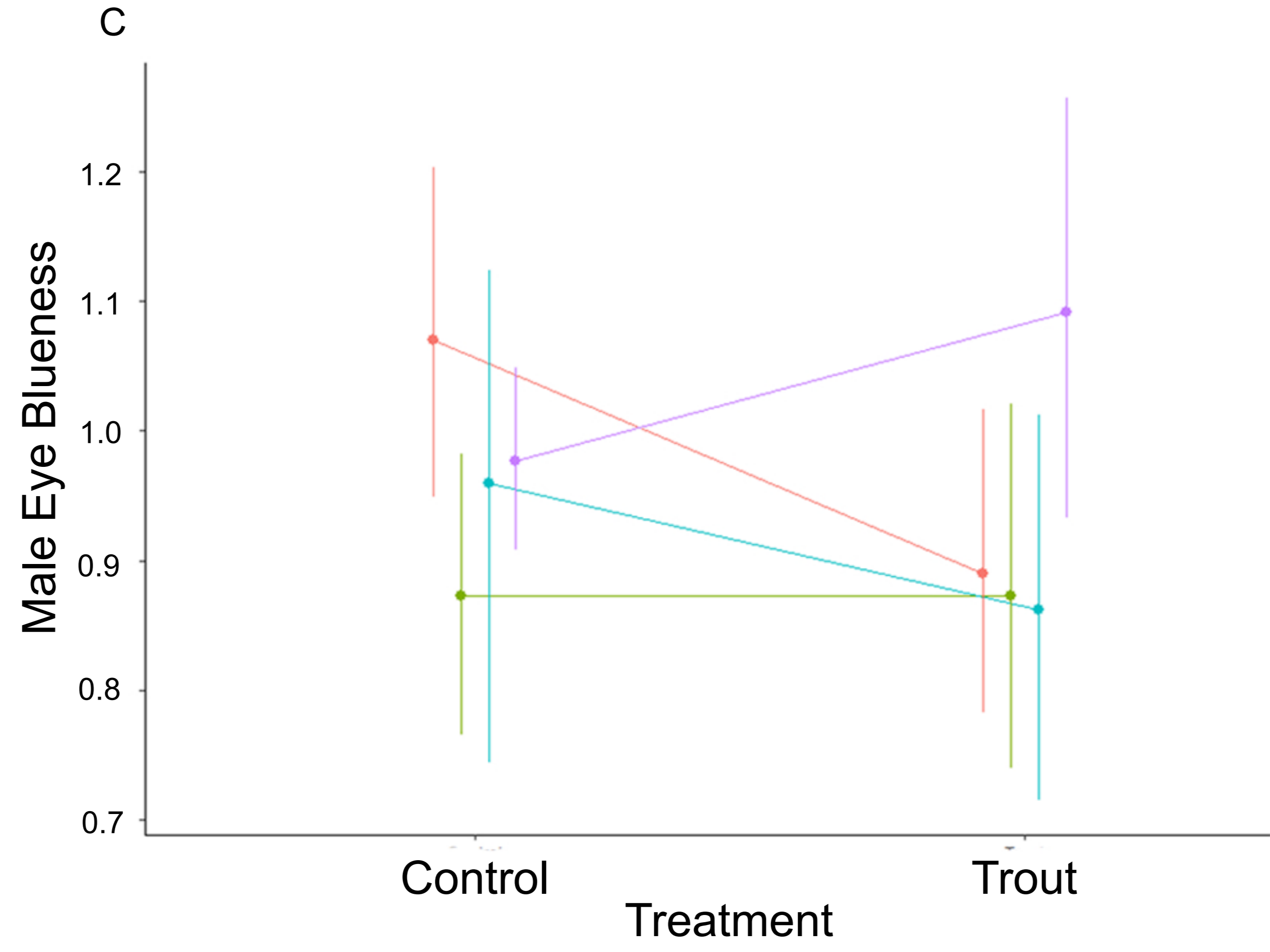
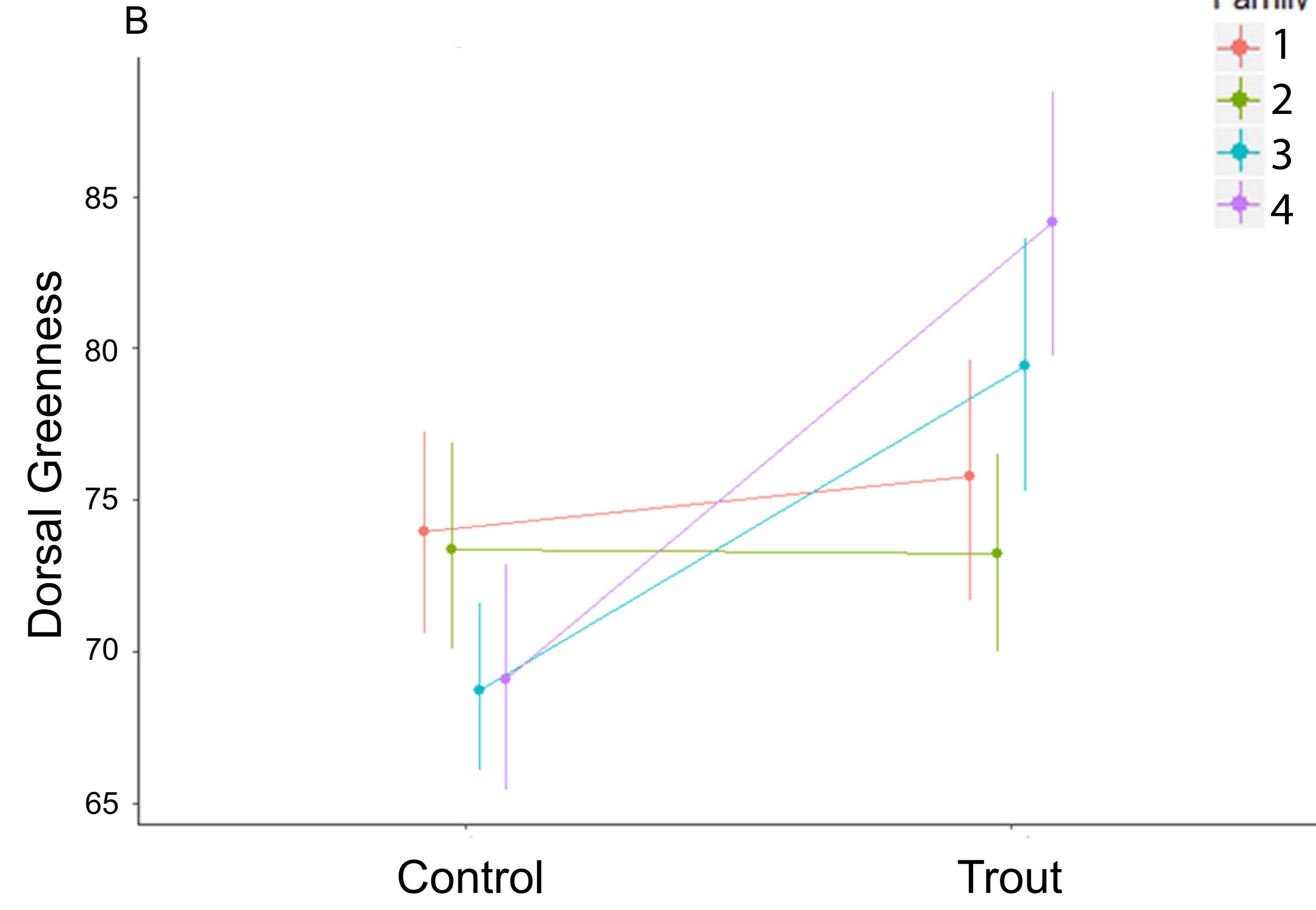
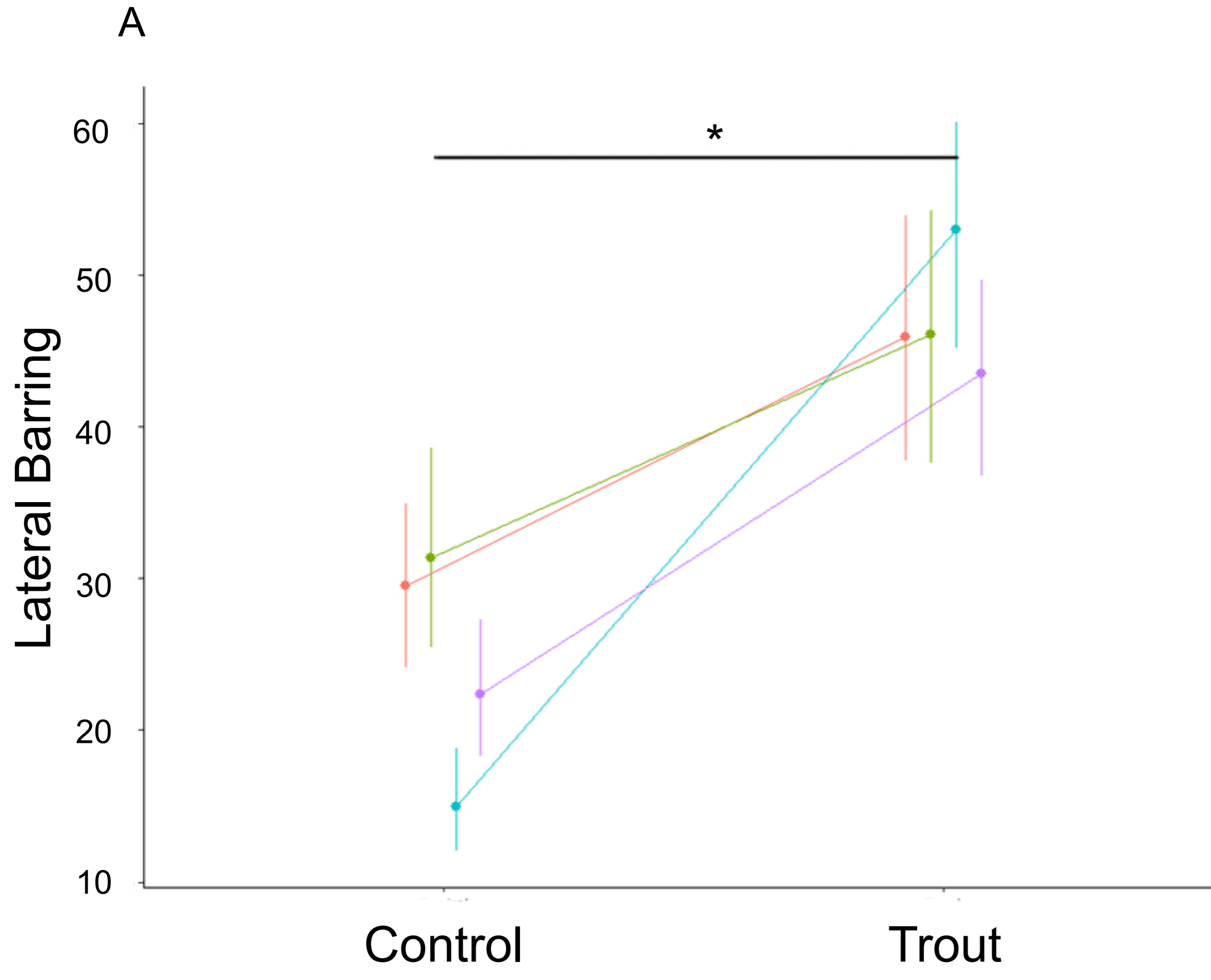
612 **Figure 2.** Relationship between barring on the lateral flank (mode-difference) and (A)
613 proportion of zooplankton in the diet and (B) water turbidity ($\mu\text{g l}^{-1}$ phytoplankton). In
614 both panels triangles indicate trout treatment ponds and circles indicate control ponds.

615 **Figure 3.** Relationship between dorsal greenness (mean green pixels) and (A) the
616 proportion of zooplankton in the diet and (B) water turbidity ($\mu\text{g l}^{-1}$ phytoplankton). In
617 both panels triangles indicate trout treatment ponds and circles indicate control ponds.

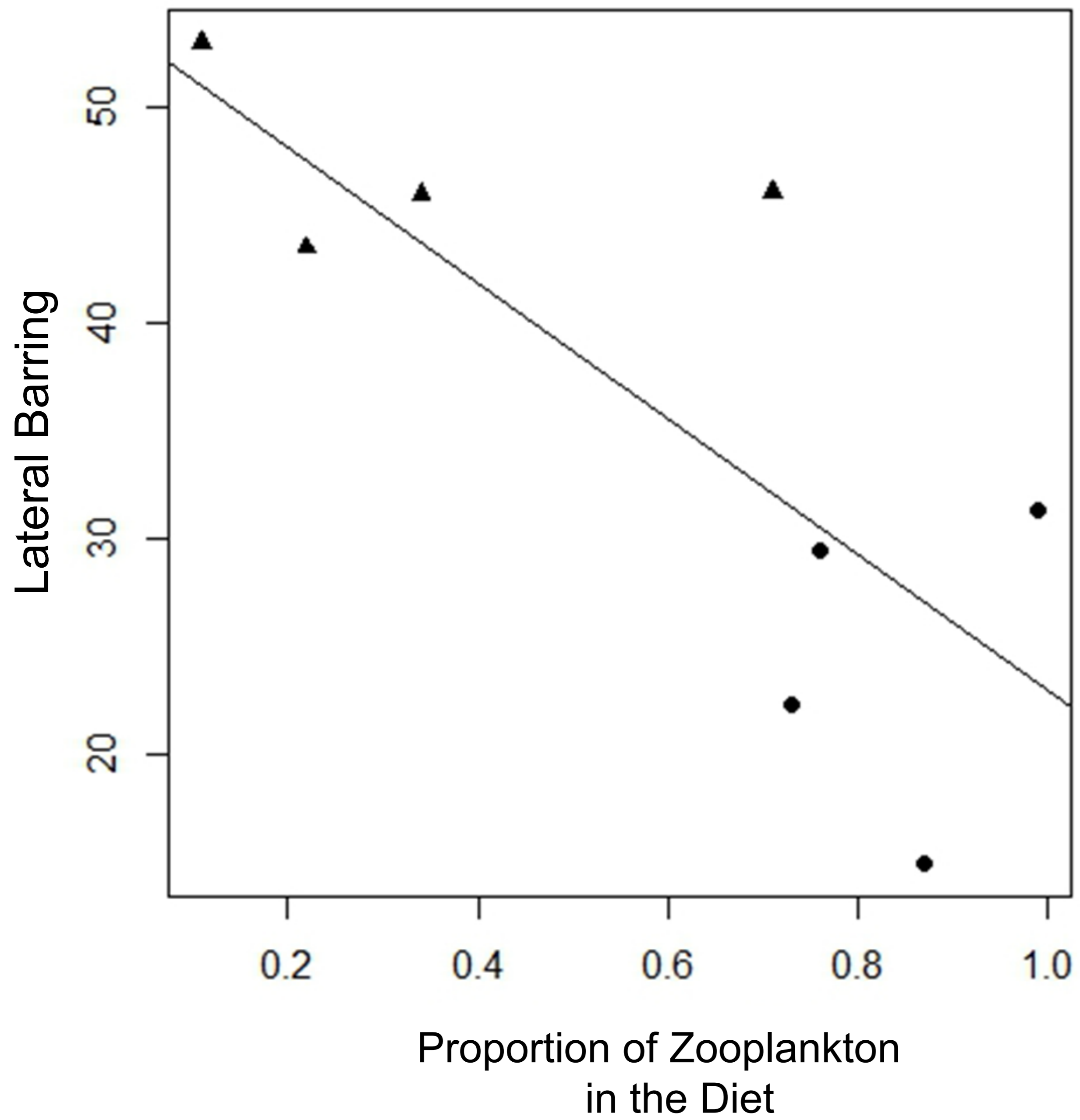
618 **Supplementary Figure 1.** Experimental set-up. Blue circles represent the four initial
619 F₁ families that were split into a trout pond (T) and a control pond (C). 100 F₂
620 individuals were photographed and analysed from each pond.

621 **Supplementary Figure 2.** Relationship between dorsal greenness (mean green
622 pixels) and male eye blueness (ratio of blue to red pixels).

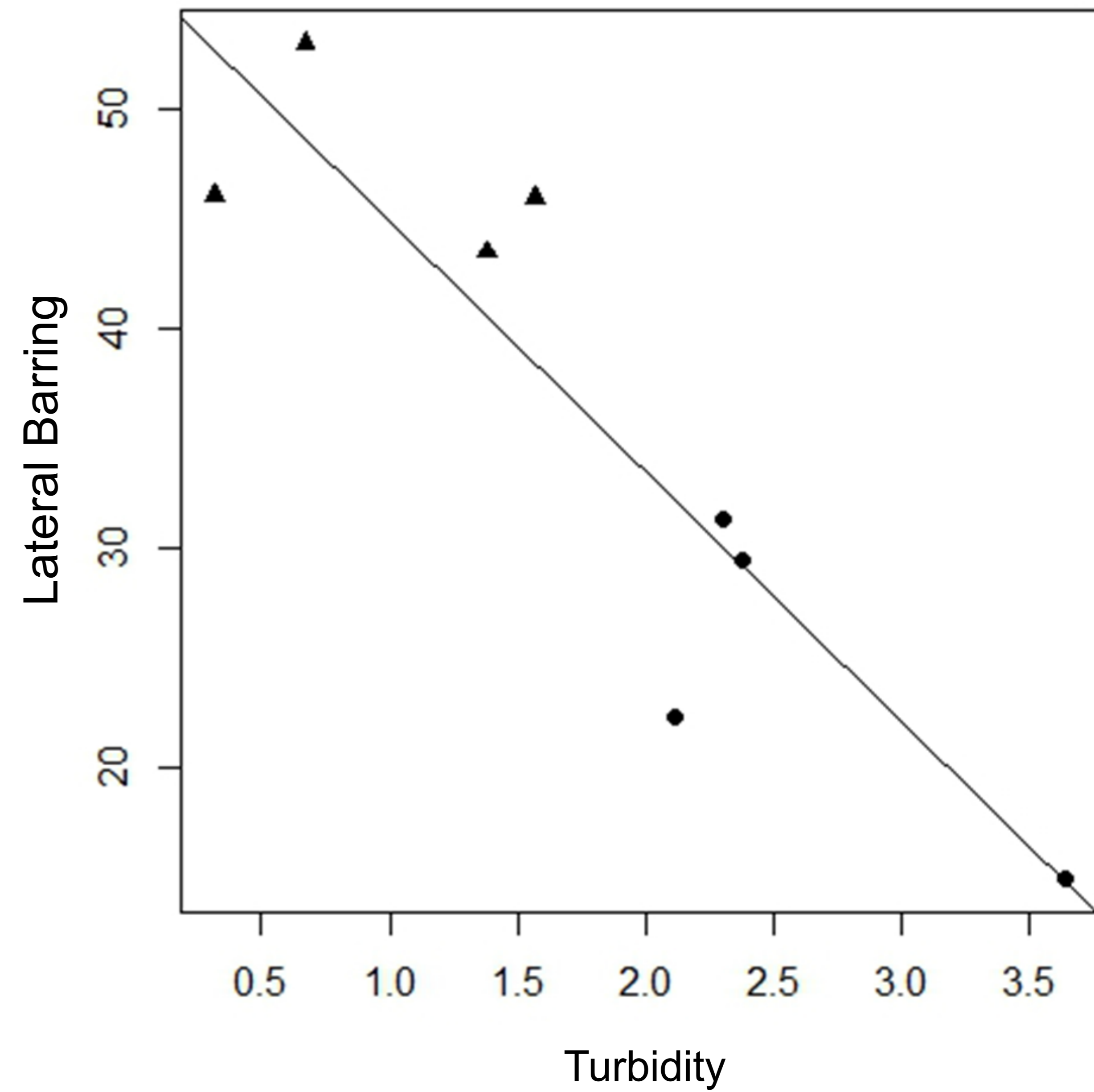
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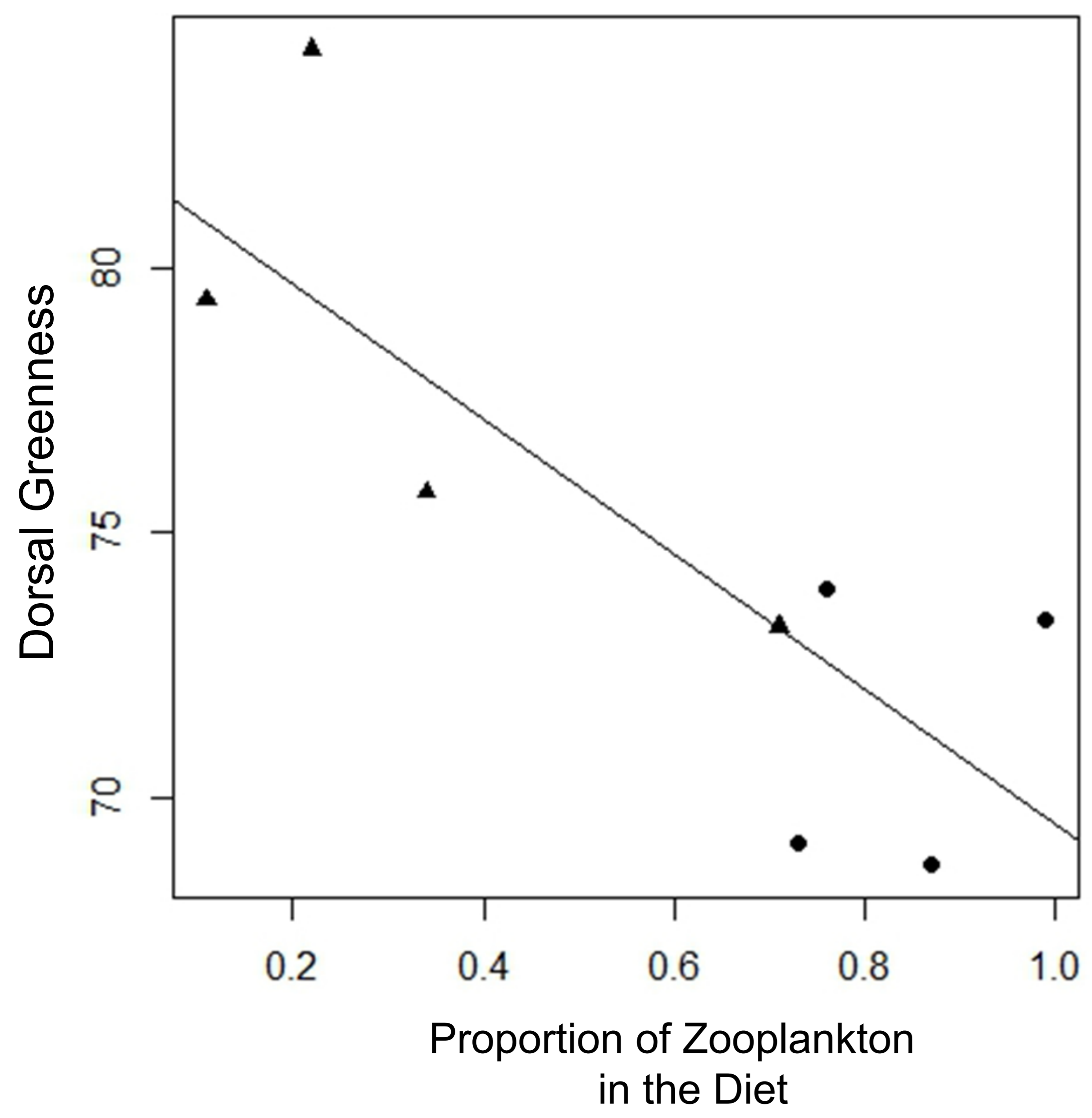
A



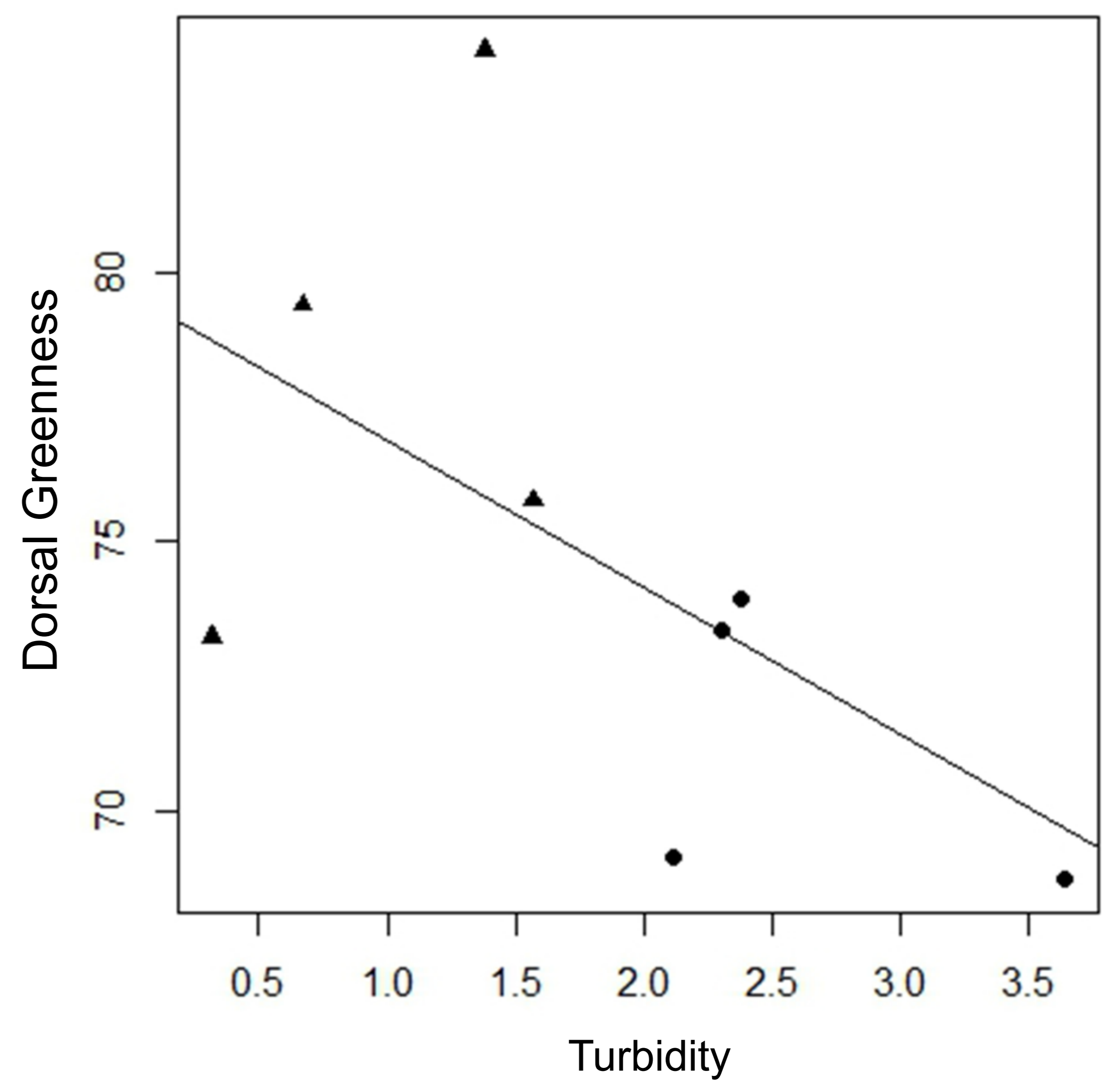
B



A



B



Set-up

