# UC Davis UC Davis Previously Published Works

## Title

The biogeography of community assembly: latitude and predation drive variation in community trait distribution in a guild of epifaunal crustaceans

Permalink https://escholarship.org/uc/item/56k7w5bg

**Journal** Proceedings of the Royal Society B, 289(1969)

**ISSN** 0962-8452

## **Authors**

Gross, Collin P Duffy, J Emmett Hovel, Kevin A <u>et al.</u>

**Publication Date** 

2022-02-23

## DOI

10.1098/rspb.2021.1762

Peer reviewed

## 1 The biogeography of community assembly: latitude and predation drive variation in

- 2 community trait distribution in a guild of epifaunal crustaceans
- 3
- 4 Authors
- Collin P. Gross Department of Evolution and Ecology, University of California, Davis, CA,
   USA. <u>https://orcid.org/0000-0002-0896-8476</u>. colgross@ucdavis.edu\*
- J. Emmett Duffy Tennenbaum Marine Observatories Network, MarineGEO, Smithsonian
   Environmental Research Center, Edgewater, MD, USA. <u>https://orcid.org/0000-0001-8595-</u>
   6391. duffye@si.edu
- Kevin A. Hovel Department of Biology, San Diego State University, San Diego, CA, USA.
   https://orcid.org/0000-0002-1643-1847. khovel@sdsu.edu
- Melissa R. Kardish Department of Evolution and Ecology, University of California, Davis,
   CA, USA. <u>https://orcid.org/0000-0002-2729-9167</u>. mrkardish@ucdavis.edu
- Pamela L. Reynolds DataLab: Data Science and Informatics, University of California, Davis, CA, USA. <u>https://orcid.org/0000-0002-0177-3537</u>. plreynolds@ucdavis.edu
- Christoffer Boström Department of Environmental and Marine Biology, Åbo Akademi
   University, Åbo, Finland. <u>https://orcid.org/0000-0003-2845-8331</u>. cbostrom@abo.fi
- Katharyn E. Boyer Estuary & Ocean Science Center and Department of Biology, San
   Francisco State University, San Francisco, CA, USA. <u>https://orcid.org/0000-0003-2680-</u>
   2493. katboyer@sfsu.edu
- 8. Mathieu Cusson Sciences fondamentales and Québec Océan, Université du Québec à Chicoutimi, Chicoutimi, QC, Canada. <u>https://orcid.org/0000-0002-2111-4803</u>.
   mathieu.cusson@uqac.ca
- Johan Eklöf Department of Ecology, Environment and Plant Sciences (DEEP), Stockholm
   University, Stockholm, Sweden. <u>https://orcid.org/0000-0001-6936-0926</u>. johan.eklof@su.se
- 10. Aschwin H. Engelen CCMAR, Universidade do Algarve, Faro, Portugal.
   https://orcid.org/0000-0002-9579-9606. aengelen@ualg.pt
- 11. Britas Klemens Eriksson University of Groningen, Groningen, Netherlands.
   <u>https://orcid.org/0000-0003-4752-922X</u>. b.d.h.k.eriksson@rug.nl
- 30 12. F. Joel Fodrie Institute of Marine Sciences, University of North Carolina at Chapel Hill,
   31 Morehead City, NC, USA. <u>https://orcid.org/0000-0001-8253-9648</u>. jfodrie@unc.edu
- 32 13. John N. Griffin Department of Biosciences, Swansea University, Wales, UK.
   33 <u>https://orcid.org/0000-0003-3295-6480</u>. j.n.griffin@swansea.ac.uk
- 14. Clara M. Hereu Universidad Autónoma de Baja California, Mexicali, Baja California,
   Mexico. <u>https://orcid.org/0000-0002-2088-9295</u>. chereu@uabc.edu.mx
- 36 15. Masakazu Hori Fisheries Research and Education Agency, Hatsukaichi, Hiroshima, Japan.
   37 <u>https://orcid.org/0000-0002-4677-9377. mhori@affrc.go.jp</u>
- 16. A. Randall Hughes Department of Marine and Environmental Sciences, Northeastern
   University, Nahant, MA, USA. rhughes@northeastern.edu
- 40 17. Mikhail V. Ivanov Department of Ichthyology and Hydrobiology, St Petersburg State
  41 University, St Petersburg, Russia. <u>https://orcid.org/0000-0002-8277-7387</u>.
  42 ivmisha@gmail.com
- 43 18. Pablo Jorgensen Instituto de Ciencias Polares, Ambiente y Recursos Naturales,
- 44 Universidad Nacional de Tierra del Fuego, Ushuaia, Tierra del Fuego, Antártida e Islas del 45 Atlántico Sur, Argentina. https://orcid.org/0000-0002-6018-7124. pjorgensen@untdf.edu.ar

- 46 19. Claudia Kruschel University of Zadar, Zadar, Croatia. <u>https://orcid.org/0000-0003-4255-</u>
   47 <u>8400</u>. ckrusche@unizd.hr
- 48 20. Kun-Seop Lee Department of Biological Sciences, Pusan National University, Busan,
   49 South Korea. https://orcid.org/0000-0003-0431-1829. klee@pusan.ac.kr
- 50 21. Jonathan Lefcheck Tennenbaum Marine Observatories Network, MarineGEO, Smithsonian
   51 Environmental Research Center, Edgewater, MD, USA. <u>https://orcid.org/0000-0002-8787-</u>
   52 <u>1786. lefcheckj@si.edu</u>
- 53 22. Karen McGlathery Department of Environmental Sciences, University of Virginia,
   54 Charlottesville, VA, USA. <u>kjm4k@virginia.edu</u>
- 23. Per-Olav Moksnes Department of Marine Sciences, University of Gothenburg, Goteborg,
   Sweden. <u>https://orcid.org/0000-0001-8611-7848</u>. per.moksnes@marine.gu.se
- 57 24. Masahiro Nakaoka Hokkaido University, Akkeshi, Hokkaido, Japan.
   <u>https://orcid.org/0000-0002-5722-3585</u>. nakaoka@fsc.hokudai.ac.jp
- 59 25. Mary I. O'Connor Biodiversity Research Centre and Department of Zoology, University of
- British Columbia, Vancouver, BC, Canada. <u>https://orcid.org/0000-0002-3133-0913</u>.
  oconnor@zoology.ubc.ca
- 62 26. Nessa E. O'Connor School of Natural Sciences, Trinity College Dublin, Dublin, Ireland.
   63 <u>https://orcid.org/0000-0002-3133-0913</u>. n.oconnor@tcd.ie
- 64 27. Jeanine L. Olsen University of Groningen, Groningen, Netherlands. j.l.olsen@rug.nl
- Robert J. Orth Virginia Institute of Marine Science, College of William and Mary,
   Gloucester Point, VA, USA. <u>https://orcid.org/0000-0003-2491-7430</u>. jjorth@vims.edu
- 67 29. Bradley J. Peterson School of Marine and Atmospheric Sciences, Stony Brook University,
  68 Stony Brook, NY, USA. <u>https://orcid.org/0000-0001-5942-8253</u>.
  69 bradley.peterson@stonybrook.edu
- 30. Henning Reiss Nord University, Bodø, Norway. <u>https://orcid.org/0000-0003-1393-0269</u>.
   henning.reiss@nord.no
- 72 31. Francesca Rossi Centre National de la Récherche Scientifique, ECOSEAS Laboratory,
   73 Université de Cote d'Azur, Nice, France. <u>https://orcid.org/0000-0003-1928-9193</u>.
   74 francesca.rossi@cnrs.fr
- Jennifer Ruesink Department of Biology, University of Washington, Seattle, WA, USA.
   <u>https://orcid.org/0000-0001-5691-2234</u>. ruesink@uw.edu
- 33. Erik E. Sotka. Grice Marine Laboratory, College of Charleston, Charleston, SC, USA.
   <u>https://orcid.org/0000-0001-5167-8549</u>. sotkae@cofc.edu
- 34. Jonas Thormar Institute of Marine Research, His, Norway. https://orcid.org/ 0000-0002 7925-3822. jonas.thormar@hi.no
- 35. Fiona Tomas IMEDEAS (CSIC), Esporles, Islas Baleares, Spain. https://orcid.org/0000 0001-8682-2504. <u>fiona@imedea.uib-csic.es</u>
- 83 36. Richard Unsworth Department of Biosciences, Swansea University, Wales, UK.
   84 <u>https://orcid.org/0000-0003-0036-9724</u>. r.k.f.unsworth@swansea.ac.uk
- 85 37. Erin P. Voigt Department of Biology, San Diego State University, San Diego, CA, USA.
   <u>https://orcid.org/0000-0003-3415-7842</u>. epvoigt@ncsu.edu
- 38. Matthew A. Whalen Hakai Institute, Campbell River, BC, Canada, and University of
  British Columbia, Vancouver, BC, Canada. <u>https://orcid.org/0000-0002-5262-6131</u>.
  <u>matt.whalen@hakai.org</u>
- 90 39. Shelby L. Ziegler Moss Landing Marine Laboratories, Moss Landing, CA, USA.
- 91 <u>https://orcid.org/0000-0001-7218-0811</u>. shelbylziegler@gmail.com

92	40. John J. Stachowicz – Department of Evolution and Ecology, University of California, Davis,
93	CA. USA. https://orcid.org/0000-0003-2735-0564. jistachowicz@ucdavis.edu
94	, <u></u>
05	*Company and in a systhem Department of Evolution and Ecology University of Colifornia Devia
95	Corresponding author: Department of Evolution and Ecology, University of California, Davis
96	2320 Storer Hall, One Shields Ave, Davis, CA 95616
97	(206) 619-3913 <u>colgross@ucdavis.edu</u>
98	
99	
100	
101	
102	
102	
103	
104	
105	
106	
107	
108	
109	
110	
111	
112	
113	
114	
115	
116	
117	
110	
110	
119	
120	
121	
122	
123	
124	
125	
126	
127	
128	
129	
130	
131	
122	
132	
133	
134	
135	
136	
137	
138	

#### 139 Abstract

While considerable evidence exists of biogeographic patterns in the intensity of species interactions, the influence of these patterns on variation in community structure is less clear. Studying how the distributions of traits in communities vary along global gradients can inform how variation in interactions and other factors contribute to the process of community assembly. Using a model selection approach on measures of trait dispersion in crustaceans associated with eelgrass (Zostera marina) spanning 30° of latitude in two oceans, we found that dispersion strongly increased with increasing predation and decreasing latitude. Ocean and epiphyte load appeared as secondary predictors; Pacific communities were more overdispersed while Atlantic communities were more clustered, and increasing epiphytes were associated with increased clustering. By examining how species interactions and environmental filters influence community structure across biogeographic regions, we demonstrate how both latitudinal variation in species interactions and historical contingency shape these responses. Community trait distributions have implications for ecosystem stability and functioning, and integrating large-scale observations of environmental filters, species interactions, and traits can help us predict how communities may respond to environmental change. 

#### 170 Introduction

171 Community ecology is fundamentally concerned with the assembly and maintenance of 172 diversity across space and time. Key to this endeavour is the idea that the composition of a local 173 community is the result of multiple ecological filters selecting species from a regional pool (Poff 174 1997; Thompson et al. 2020). Different kinds of filters apply different kinds of selective 175 pressures on the species pool, and because species' traits are what allow them to pass through 176 filters, studying the distribution of traits within the community can help us understand how these 177 filters act on the species pool as a whole. Strong environmental filters (i.e., abiotic filters sensu 178 Kraft et al. 2015) such as climate are thought to act on large spatial scales to constrain trait 179 diversity such that species are more alike (clustered) in traits that respond to these factors than 180 we would expect under a purely random assembly process (Webb et al. 2002; Cavender-Bares et 181 al. 2009; Starko et al. 2020; Thompson et al. 2020). Biotic filters, such as competition, then act 182 at smaller spatial scales to enhance or reduce trait diversity among species with broadly similar 183 abiotic tolerances, depending on which traits are affected (Mayfield & Levine 2010). When traits 184 related to the acquisition of distinct resources are considered, competition for these resources 185 drives the distribution of traits to be wider than expected by chance (overdispersed) as there are 186 multiple resource niche optima that can be occupied (Webb et al. 2002; Cavender-Bares et al. 187 2009; Pavoine & Bonsall 2011). In contrast, competition for a single, dominant limiting resource 188 can also act as a filter, selecting for traits related to acquiring this resource to converge around an 189 optimal value, because species deviating from the optimum are otherwise competitively 190 excluded. All else being equal, as richness increases, an increase in trait dispersion may point to 191 stronger stabilising mechanisms and limiting similarity, while a decrease in trait dispersion can 192 suggest stronger equalising mechanisms promoting unstable coexistence. (Chesson 2000; 193 Mayfield & Levine 2010).

194 Despite well-known geographic patterns in the strength of both biotic interactions and 195 environmental filters (Schemske *et al.* 2009; Reynolds *et al.* 2018; Longo *et al.* 2019; Zvereva & 196 Kozlov 2021), few studies have examined the global-scale consequences of geographic variation 197 in these filters for community trait distributions (Ford & Roberts 2018, Skeels *et al.* 2020). In 198 particular, intense predation, competition, and mutualistic interactions at lower latitudes 199 (Freestone & Osman 2011; Longo *et al.* 2019; Zvereva & Kozlov 2021), may lead to the 200 predominance of biotic interactions over environmental filters in structuring low-latitude 201 communities. This may cause stronger trait clustering near the poles that shifts towards more 202 overdispersed communities at lower latitudes. On the other hand, selection for tolerance of 203 extreme heat conditions could also cause trait clustering at low latitudes. Finally, patterns in 204 community structure along latitudinal gradients could be dominated by idiosyncratic and 205 historically-contingent effects of predators, prey, competitors, and mutualists that vary among 206 biogeographic provinces (Sanford & Bertness 2009; Mittelbach & Schemske 2015; Ford & 207 Roberts 2019; Whalen et al. 2020). Local abiotic factors, habitat complexity, assemblage 208 composition, and adaptation to these local factors could further obscure broader geographic 209 patterns of community assembly (Sanford & Bertness 2009; Lavender et al. 2017), stressing the importance of assessing patterns across multiple independent species pools. For example, the 210 211 effects of regional gradients in predation may be overshadowed by local increases in habitat 212 complexity, which can decrease predation pressure (Reynolds et al. 2018) and increase trait 213 dispersion as species assort into disparate microhabitat niches (Best & Stachowicz 2014). 214 Understanding trait distributions and their drivers should provide insight into the likely responses 215 of communities to environmental fluctuations or perturbations in the same way that 216 understanding the diversity of traits within a population can inform us on its evolutionary 217 potential (Cadotte et al. 2011; Rumm et al. 2018).

218 Here we examine geographic patterns in the trait distribution of epifaunal invertebrates 219 living on eelgrass throughout the northern hemisphere to assess the extent and causes of 220 geographic variation in the drivers of the assembly of these communities. Eelgrass (Zostera 221 marina) is the world's most widespread species of temperate seagrass, a marine angiosperm 222 found throughout the Northern Hemisphere from 30° to 67° N latitude in both the Atlantic and 223 Pacific Oceans (den Hartog 1970; Green & Short 2003). Much of the animal community in 224 eelgrass beds is made up of invertebrate mesograzers that primarily feed on the epiphytic 225 microalgae fouling the seagrass blades (Valentine & Duffy 2006). Competition for food and 226 microhabitat space occurs among mesograzers, and can significantly affect community 227 composition (Edgar 1990; Best et al. 2013; Best & Stachowicz 2014; Amundrud et al. 2015). 228 Peracarid crustaceans (amphipods, isopods, and tanaids) are the most widespread, abundant, and 229 species-rich mesograzer taxon in these eelgrass beds, and they experience elevated predation in 230 low-latitude eelgrass beds (Reynolds et al. 2018) which could either cause clustering of 231 communities around traits that increase resistance or tolerance to predation, or cause dispersion

232 of communities due to competition for enemy-free space. Z. marina's wide range across latitudes 233 provides an opportunity to assess the role of gradients of ecological filters on global scales 234 without the confounding influence of changing habitat type. We predicted: (1) that trait 235 dispersion would increase with decreasing latitude as species interactions become more intense 236 and (2) that abiotic filters would be strongest and result in clustering at higher latitudes and 237 where biotic interactions are weak. While marine systems often show non-linear variation in 238 species diversity and interaction strength with latitude (peaking at mid-latitudes; Chaudhary et 239 al. 2017; Whalen et al. 2020), our predictions are reasonable within the range of latitudes 240 occupied by eelgrass ( $\sim$ 30-70°N). We test these predictions in separate ocean basins with largely 241 unique fauna, allowing us to assess whether the unique histories of these zoogeographic 242 provinces result in different patterns and drivers of trait distribution in each ocean basin (Roy et 243 al. 2009; Dyer & Forister 2019).

#### 244 Methods

245 Study design and sample collection. Between May and September 2014, we sampled 42 246 sites across the range of Z. marina, spanning 30 degrees of latitude along both coasts of Eurasia 247 and North America (30.4°N to 60.1°N; Fig. 1) to characterize the biological and physical 248 structure of eelgrass beds using standardized measurements. We implemented a hierarchical 249 sampling design consisting of two oceans (Atlantic and Pacific), each with two coasts (east and 250 west), each with 6-14 sites, each with 20 plots, for a total of 840 plots in 42 sites sampled as part of the Zostera Experimental Network (ZEN; Fig. S1). Plots were 1 m<sup>2</sup> and spaced 2 m apart at 251 252 each site. Along each coastline, sites were separated by 4.9 km (Virginia, USA) to 485.4 km 253 (Washington State, USA) of water.

254 Assessing eelgrass habitat characteristics. We sampled eelgrass biomass by haphazardly 255 placing and pushing a 20-cm diameter core tube 20 cm into the sediment within each plot. We 256 gathered all shoots rooted within the core bottom area into the core tube to ensure that no shoots 257 were cut off during sampling. We then removed the shoots from the sediment, transferred the 258 core contents into a mesh bag. In the lab, we rinsed the core contents, removed fouling algae and 259 sediment from the eelgrass tissue, and separated above- and belowground biomass by cutting the 260 plant above the rhizome. In addition to eelgrass, we also removed all of the macroalgae from the 261 plot. All eelgrass and macroalgal tissue was dried to a constant weight at 60°C and weighed.

From five haphazardly collected eelgrass shoots per plot, we also collected 3-cm lengths of
tissue from a healthy, unfouled inner leaf and processed these samples for tissue nitrogen using a
CHN analyser (Thermo Fisher Scientific Inc., Waltham, MA, USA).

We quantified eelgrass habitat structure at the plot level by measuring shoot density and canopy height. We estimated shoot density by counting the number of shoots emerging within a 20-cm diameter ring placed haphazardly in the plot. In plots where density was particularly low (less than 50 shoots m<sup>-2</sup>, about 5% of plots), we counted all of the shoots in the plot. We measured canopy height by haphazardly collecting five shoots from each plot and measuring their length from the tip of the longest leaf to the leaf sheath.

We sampled epiphyte load on the eelgrass blades by selecting four shoots from each plot and removing them from the substrate either by gently uprooting or clipping at the meristem and placing them in a plastic bag on ice for transport. In the lab, we scraped both sides of all the leaves with a glass slide to remove fouling material, which was then filtered, transferred to an aluminium pan, dried to a constant weight at 60°C, and weighed.

276 Measuring predation intensity. Predation intensity was quantified by tethering locally-277 collected prey ("gammarid" amphipods) in each plot for 24 hours. These data and methods are 278 reported in detail in Reynolds et al. (2018). Briefly, each individual amphipod was glued to a 10-279 cm piece of monofilament line 0.133 mm in diameter (Berkley Fireline<sup>TM</sup>, Spirit Lake, IA, USA) 280 tied to a transparent acrylic stake anchored in the sediment, so that it could swim freely in the 281 water column and cling to adjacent eelgrass blades. After 24 hours, we removed the stakes and 282 scored prey as present (uneaten) or absent (eaten); partially-consumed prey were considered 283 eaten, and moulted prey were excluded from analyses. Site-level predation was calculated by 284 averaging scores across plots.

285 Abiotic environmental variables. To characterize the abiotic environment experienced by 286 epifauna across the range of eelgrass, we measured in-situ temperature and salinity at each site at 287 the time of sampling. To characterize the overall abiotic environment of each site, we also 288 retrieved estimates of annual mean sea surface temperature (SST), photosynthetically active 289 radiation (PAR), and surface chlorophyll A (Chl a) from the surrounding region, available in the 290 Bio-ORACLE data set (Tyberghein et al. 2012). These data were taken from monthly readings of the Aqua-MODIS and SeaWiFS satellites at a 9.6 km<sup>2</sup> spatial resolution from 2002 to 2009. We 291 292 used the raster package in R v. 3.6.3 (Hijmans & Etten 2020; R Development Core Team 2021)

to extract the annual mean SST, SST range, PAR, and Chl a from all cells within 10 km of each site, and averaged these cell-level estimates to generate site-level predictors. Other water quality parameters, including dissolved nitrate and other nutrients, were spatially interpolated based on surface measurements in the World Ocean Database 2009 (Garcia *et al.* 2010).

297 Epifaunal community composition. To sample the macrofauna associated with the 298 eelgrass blades, we carefully placed an open-mouthed fine-mesh drawstring bag (500 µm mesh, 299 18 cm diameter) over a clump of shoots in the centre of the plot so that the mouth of the bag was 300 flush with the sediment surface. We then cut the shoots where they emerged from the sediment 301 and quickly closed the drawstring to capture the shoots and associated animals. The shoots were 302 transferred to the lab on ice, rinsed and hand-inspected to dislodge the epifauna, which were then 303 passed through a 1-mm sieve and ultimately transferred into 70% ethanol. Epifauna were then 304 identified to the lowest possible taxonomic level (typically species). Epifaunal abundance was 305 standardized by the aboveground biomass of the eelgrass sample from which they were 306 collected.

307 We scored all peracarids (amphipods, isopods, and tanaids) for a series of traits based on 308 information available in the literature, including body size, fecundity, body shape, living habit, 309 motility, bioturbation, and diet components (Table 1, Appendix 1 for literature). Due to a paucity 310 of data on intraspecific trait variation for most species, literature values were assumed to be 311 representative for all individuals in our study. For subsequent analyses, we categorized each of 312 these traits as related to microhabitat or dietary niche; we also performed analyses with all traits 313 ungrouped. While we acknowledge that these broad categories may overlap, we elected to sort 314 traits into these categories because they represent two potential components of trait dispersion 315 exhibited by peracarids in field studies and laboratory experiments (Best et al. 2013; Best & 316 Stachowicz 2014). Correlations among traits were generally weak, save for strong positive 317 relationships between eating live seagrass tissue and macroalgae, detritivory and consuming 318 seagrass detritus, and suspension feeding and bioturbation (Fig. S2).

319 *Characterizing community dispersion.* For all the peracarid species observed in our 320 dataset, we used the trait dataset to generate three matrices of Gower distances between species: 321 one of all traits, one for diet traits, and one for microhabitat traits using the FD package in R 322 (Laliberté *et al.* 2014). Using subsets of these matrices for communities at the site level (summed 323 across 20 plots at each site, n = 42), we measured the trait distance between species as the Mean Pairwise Distance (MPD) and Mean Nearest Taxon Distance (MNTD) for each set of traits
(Webb *et al.* 2002; Sessa *et al.* 2018). MPD is the average of the trait distances between all pairs
of species found within a given sample unit (site), while MNTD is the average minimum
distance between species pairs in a site. Both are independent of species richness, but the two
metrics can behave differently depending on the clustering of species in trait space within a
sample (Sessa *et al.* 2018).

330 To determine whether the observed species traits in each community differed from those 331 expected by chance, we standardized MPD and MNTD against null distributions generated 332 according to two permutation algorithms. The first, independent swap, is a semi-constrained 333 model that randomly re-assembles the sample-by-species community matrix while maintaining 334 the species richness of each sample and the presence/absence of each species across samples. 335 The second, tip shuffle, is a more constrained model that directly shuffles the traits of the species 336 in the community while maintaining richness, occurrence, and trait distances between 337 community members, effectively moving the tip labels on a trait dendrogram. Imposing more 338 constraints on permutation controls for patterns in the data that are not directly relevant to the 339 question at hand, such as species richness, occurrence, or identity, ultimately reducing type I error rates (Swenson 2014). Because of the relatively low overlap in species pools across the 340 341 range of our study, comparing the results relative to both types of models can be informative of 342 the importance of species identity in these types of permutations, and also facilitate comparison 343 with other studies in which the independent swap algorithm has been used together with less 344 constrained permutations (e.g., Best and Stachowicz 2014). These permutations were each 345 completed 999 times for each community, and null distributions of MPD and MNTD were 346 generated based on values calculated from randomized communities.

We examined the effect of the species pool on community dispersion, using varying degrees of constraint on the matrix and trait dendrogram used to generate null distributions. To make comparisons among sites, we permuted within the global species pool (all sites) and oceanlevel Atlantic and Pacific species pools. Using a global pool in our permutations is appropriate because while all species were not present in all regions, there were no traits that were exclusive to any region (Fig. S2).

Each observed value of community trait distance was then compared to the corresponding null distribution by calculating the standard effect size (SES<sub>MPD</sub> or SES<sub>MNTD</sub>). A positive value of 355 SES indicates that the observed community trait distance (as measured by MPD or MNTD) is 356 greater than the null mean, meaning that community members are more dissimilar than expected 357 under a random draw (overdispersion), while a negative SES indicates that trait distance is less 358 than the null mean, meaning that community members are more similar to each other than 359 expected under a random draw (clustering). MPD, MNTD, null distributions and SES values 360 were calculated using the picante package in R (Kembel *et al.* 2010).

361 Data analysis. Two distance metrics (MPD and MNTD), two permutation algorithms 362 (independent swap and tip shuffle), three species pools (global, Pacific, and Atlantic), and three 363 trait sets (all, diet, and microhabitat) totalled 36 sets of SES values. However due to missing diet 364 data for some species, we were unable to calculate diet SES<sub>MNTD</sub> with the tip shuffle algorithm, 365 leaving us with a total of 33 sets. For each distance metric, algorithm, species pool, and trait set, 366 SES values were used as response variables in a set of 16 linear models incorporating latitude, 367 ocean, continental margin (east vs. west), in-situ temperature and salinity, annual mean and 368 range of SST, total crustacean abundance and median crustacean size, epifaunal and peracarid 369 richness, macroalgal biomass, average predation intensity, epiphyte load, Chl a, PAR, water 370 column nitrate, mean leaf nitrogen content, and two axes of eelgrass habitat structure as derived 371 from a principal component analysis incorporating shoot density, leaf sheath width and length, 372 longest leaf length, and aboveground biomass (PC1 and 2, Fig. S4) as predictor variables, as well 373 as select interactions between them (Table 2). Predictors were log-, square-root-, or arcsin-374 transformed where appropriate to conform to a normal distribution based on Shapiro-Wilk 375 normality tests and visual examinations of histograms. Collinearity of predictors was accounted 376 for using variance inflation factors (VIF) for variables in composite models using the car 377 package in R (Fox & Weisberg 2019). Predictors with a VIF greater than five were removed 378 from composite models. We also examined the effects of predictors on the SES of individual 379 traits to understand what traits may drive the patterns we see across environmental gradients 380 (Appendix 2).

We ranked these initial hypothesis-driven models of SES using AICc scores (MuMIn package; Bartoń 2020), and then incorporated predictors from the three lowest-scoring models of each set into a set of composite models to examine the combined effects of multiple predictor types. We then used backwards elimination to select the lowest-scoring model from these  $^{385}$  composite models. Where two models had a  $\Delta$ AICc less than 3 units, we selected the model with the fewest parameters for interpretation.

#### 387 **Results**

388 Peracarid assemblages at Pacific sites had greater trait dispersion than Atlantic sites, and 389 dispersion increased with increasing predation and decreasing latitude, though there were some 390 differences among the two oceans that we outline below. Across our sites, we found a total of 391 105 species, 55 of which were found in the Atlantic, and 60 of which were found in the Pacific, 392 with 10 species found in both oceans. There were 15 species in the Northwest Pacific, 48 species 393 in the Northeast Pacific, 36 species in the Northwest Atlantic, and 24 species in the Northeast 394 Atlantic (Fig. S3). The patterns and predictors of trait dispersion were robust across SES metrics 395 and permutation algorithms (Table S1; Fig. S5); here we present and interpret the results of 396 model selection on SES<sub>MNTD</sub> calculated using the tip shuffle algorithm, with exceptions 397 presented where relevant.

398 Dispersion of traits by ocean basin. Of the set of all traits examined, communities at 399 Atlantic sites were on average clustered (SES < 0) relative to the global null, particularly for 400 body size and living habit (Fig. A2-2) – species clustered around a mean body size of 14.09 mm 401 (47.5% smaller than the mean Pacific body size), and most were free-living. Communities at 402 Pacific sites were overdispersed (SES > 0) on average relative to the global null (Fig. 2, Table 403 S1, Table S2). This pattern held for both metrics and null models but was significant only for 404 SES<sub>MPD</sub> (SES<sub>MPD</sub> independent swap  $t_{38.097} = 2.43$ , p = 0.020; SES<sub>MPD</sub> tip shuffle  $t_{38.242} = 2.31$ , p =405 0.027; two-sample t tests). Within the global pool, the separate calculations of SES using 406 microhabitat and feeding traits showed a similar pattern; for microhabitat traits, Pacific 407 communities were more overdispersed and Atlantic communities more clustered (SES<sub>MNTD</sub> tip 408 shuffle  $t_{35.654} = 3.64$ , p = 0.00086; Fig. 2).

409 *Correlates of among-site variation in trait dispersion.* Predation intensity, latitude, 410 epiphyte load, and ocean basin (within the global species pool) were the strongest and most 411 consistent predictors of SES across all species pools and all trait sets (Table S1, Fig. S5). In-situ 412 temperature, bed characteristics, epifaunal richness, continental margin, nitrate, and salinity also 413 appeared occasionally (less than 30% of models) across the best models of SES. Mean annual 414 sea surface temperature, epifaunal richness, salinity, nitrate, in-situ temperature, and crustacean 415 abundance also varied significantly with latitude (Fig. S8). 416 In all of the best models, peracarid communities at sites with higher predation intensity 417 had more overdispersed traits, whereas those with less intense predation had more clustered traits 418 relative to a random draw from the species pool (Fig. 3a, Table S1, Fig. S5a-c). Predation 419 (removal of amphipod baits) varied from 20% in Quebec to 100% in Sweden, San Francisco 420 Bay, Ireland, Korea, and British Columbia; the average predation rate was significantly greater in 421 the Pacific than in the Atlantic Ocean (Table S3, Fig. S7, S8), but this did not translate to a 422 difference in the effect of predation on dispersion across the two basins when permuting within 423 the global pool (p = 0.48; Fig. 3a). Across the three species pools, the predation effect was 424 stronger on average when permuting within the Pacific than the Atlantic or global pools, (Fig. 425 S5a), and strongest in models of the dispersion of all traits together (Fig. S5b).

426 As predicted, trait dispersion decreased with increasing latitude in the best models (global 427 species pool, microhabitat traits); communities became more clustered at higher latitude, while 428 communities toward the equatorward edge of Z. marina's range were more overdispersed (Fig. 429 3b, Fig. S5d-f). These latitude effects were stronger in the Pacific Ocean than in the Atlantic  $(F_{1,38} = 7.95, p = 0.0076; Fig. 3b)$  although they did not appear in the top models when 430 431 permuting within the Pacific species pool (Fig. S5d); the best model including latitude was 1.3 432 AICc units better than the top model, but it was not selected as the top model because of the 433 small difference in AICc score and greater number of parameters. Like predation, the latitude 434 effect was strongest in models including all traits together (Fig. S5e).

Communities were more clustered (more negative SES) at sites with high epiphyte loads,
but this effect was most obvious in the Atlantic species pool when only microhabitat traits were
considered (Fig. 3c; Fig. S5g-h). There was rarely an effect of epiphyte load on SES when using
other species pools (Fig. S5g, Table S1) and never for diet traits (Fig. S5h).

439 **Discussion** 

Using a global dataset of eelgrass-associated peracarid crustaceans, we found a strong
increase in community trait dispersion with decreasing latitude and increasing predation (Fig. 3a,
b). Latitudinal clines in different ecological filters have been well-characterized in a wide variety
of systems (Schemske *et al.* 2009; Reynolds *et al.* 2018; Zvereva & Kozlov 2021), particularly
temperature and the strength of species interactions (Schemske *et al.* 2009; Longo *et al.* 2019;
Zvereva & Kozlov 2021), both of which decrease at high latitudes. Stronger biotic interactions,
in particular stabilizing interactions (*sensu* Chesson 2000), at lower latitudes may select for an

overdispersed community (Webb *et al.* 2002; Mayfield & Levine 2010; Pavoine & Bonsall
2011), while stronger abiotic filters (or relatively weaker biotic filters) at either end of range (e.g.
cold at the poleward edge or hot at the equatorward edge) could select for a clustered community
(Webb *et al.* 2002; Cavender-Bares *et al.* 2009; Kraft *et al.* 2015). We found similar total
numbers of species in the two oceans (Fig. S3) given similar sampling effort, and all traits were
found in both oceans, so the differences we observe among oceans are not simply the result of
different diversities in the underlying species pool.

454 Several lines of evidence point to the relatively greater effect of biotic interactions over 455 temperature in structuring our communities. First, temperature rarely appeared as a significant 456 factor in our best models (Fig. 3d, Table S1). Second, latitudinal clines in dispersion were more 457 dependent on ocean basin than continental margins, which differ significantly in their 458 temperature gradients (western side of oceans are warmer at an equivalent latitude; Fig. 3b; 459 Reynolds et al. 2018). Third, predation in this system decreases with latitude, as it does in many 460 others (Reynolds et al. 2018; Longo et al. 2019; Zvereva & Kozlov 2021). Fourth, we observed 461 greater dispersion in living habit, motility, and macroalgae consumption at lower latitudes (Fig. 462 A2-1b-d), all of which can be reasonably linked to stabilizing competition for food or enemy-463 free space. Finally, for some traits (body size, fecundity), we would expect clustering at both 464 ends of a thermal gradient, but around different optima: large-bodied and highly fecund 465 peracarids at cool sites, and small-bodied peracarids that produce fewer eggs at warm sites 466 (Sainte-Marie 1991; Jaramillo et al. 2017). However, in ectotherms like peracarids, decreases in 467 temperature at higher latitudes are less likely to be strong drivers of community structure than 468 increases in temperature at lower latitudes as a result of asymmetrical performance curves 469 (Martin & Huey 2008; Vasseur et al. 2014). While we saw that high-latitude sites tended to have 470 species with high fecundity (65 to <135 eggs per brood; part of a general trend for clustered sites 471 to have high or very high fecundity; Fig. A2-1a), we saw no similar trend towards clustering at 472 low latitudes around low fecundity values or any other traits.

The decline in trait dispersion with latitude was significantly greater in the Pacific than the Atlantic. This difference in latitudinal clines and trait dispersion more generally between the two ocean basins (Fig. 2, Fig. 3b) may be in part due to differences in these assemblages' biogeographic and evolutionary histories (Mittelbach & Schemske 2015). First, glaciation in the north Atlantic during the last Ice Age means that many of the areas in which eelgrass now occurs 478 would have been colonized after glaciers retreated (Vermeij 1991; Olsen et al. 2004), leaving 479 less time for in-situ adaptation and specialization that might lead to increased trait dispersion 480 (Cavender-Bares et al. 2009). Similarly, given Z. marina's origin in the Pacific and more recent 481 Pleistocene expansion into the Atlantic (Olsen et al. 2004), we might also generally expect 482 Atlantic species to have colonized eelgrass from other Atlantic-native habitats, perhaps 483 predisposing them to be less overdispersed in their traits as they cluster around a single mean. 484 Consistent with this, we found that species in Atlantic sites were clustered around a smaller mean 485 body size, which may be selected for by the denser eelgrass habitat in the Atlantic (Fig. S4, Fig. 486 A2-2a; (Bartholomew et al. 2000). Finally, gastropod relative abundance increases with latitude, 487 and gastropods are a more abundant and speciose component of the epifaunal community in the 488 north Atlantic than in the Pacific (Gross *et al.* unpublished). Competition with gastropods for 489 epiphytes or other shared resources may push the peracarids there into a more constrained area of 490 trait space, leading to the clustering we observed.

The precise impacts of these and other historical factors are difficult to quantify but may be further investigated with analyses of phylogenetic dispersion or more detailed studies of trait distributions in the regional species pool (Denelle *et al.* 2019; Skeels *et al.* 2020). However, we currently lack a phylogeny of peracarids with sufficient resolution and taxon sampling with which to evaluate underlying differences in phylogenetic diversity between the two ocean basins. We do note that richness of species, genera and families did not vary substantially between the ocean basins (Fig. S3).

498 One of the most striking results of our study was the positive effect of predation intensity 499 on community dispersion among sites that was consistent in both oceans (Fig. 3a); peracarid 500 species were more dissimilar in their traits than expected by chance in sites with high predation 501 intensity. This effect appeared across trait sets, species pools, dispersion metrics and methods 502 (Table S1), although we rarely saw this signal at the level of individual traits (Table A2-1, Fig. 503 A2-3). Changes in predator community structure, predation intensity, or both could lead to an 504 increase in competition for predator-free space, an ecological selective filter that may result in 505 overdispersion, particularly with respect to microhabitat and predator avoidance traits (Best & 506 Stachowicz 2014). Herbivorous arthropods in both marine and terrestrial systems are known to 507 select their microhabitat niches based largely on their effectiveness as shelter from predators 508 rather than the availability or quality of food (Bernays & Graham 1988; Duffy & Hay 1991;

509Lasley-Rasher *et al.* 2011). Consequently, competition for enemy-free space can be an important510factor structuring communities. Alternatively, predation could affect trait dispersion by reducing511competition (Pianka 1966; Amundrud *et al.* 2015), but we would expect this to lead to an512increase in dispersion from strongly clustered (SES < 0) to random communities (SES = 0) as</td>513stabilizing competition lessened, rather than the observed shift from clustered to overdispersed

514 (SES > 0, Fig. 3a, Fig. S5b).

515 Latitudinal patterns of species interactions are now broadly appreciated (Schemske et al. 516 2009; Freestone & Osman 2011; Reynolds et al. 2018; Longo et al. 2019; Whalen et al. 2020; 517 Zvereva & Kozlov 2021), but rarely are these results explicitly connected to variation in the 518 structure of communities. By examining both how species interactions and environmental drivers 519 vary within a single habitat type across a broad geographic gradient, we demonstrate an 520 important role for latitudinal variation in species interactions in driving patterns of community 521 assembly. Diversity in important traits can increase the completeness with which epiphytes are 522 removed, leading to increased seagrass growth (Duffy et al. 2003), an effect that is strongest in 523 the presence of predators (Duffy et al. 2005). More generally, trait clustering and dispersion have 524 implications for redundancy, stability, and ecosystem functioning (Cavender-Bares et al. 2009; 525 Cadotte et al. 2011; Leibold et al. 2017). For instance, communities may be less resilient to 526 environmental change if they are clustered by environmental filters (Cadotte et al. 2011, Rumm 527 et al. 2018). Clustering that occurs as a result of equalizing mechanisms (sensu Chesson 2000) 528 can weaken the relationship between diversity and ecosystem functioning, or certain ecosystem 529 functions may be enhanced in communities with overdispersed effect traits, especially if 530 diversity-function relationships arise through complementarity (Leibold et al. 2017; Thompson 531 et al. 2020). Thus, historical contingency and broad-scale ecological drivers may play an 532 important role in constraining not only the assembly of local communities, but the resulting trait 533 diversity can affect the functioning of the entire ecosystem. This approach, if applied broadly, 534 offers the potential for developing a predictive understanding of how entire communities respond 535 to environmental change.

#### 536 References

Amundrud, S.L., Srivastava, D.S. & O'Connor, M.I. (2015). Indirect effects of predators control
 herbivore richness and abundance in a benthic eelgrass (Zostera marina) mesograzer
 community. *Journal of Animal Ecology*, 84, 1092–1102.

- Bartholomew, A., Diaz, R.J. & Cicchetti, G. (2000). New dimensionless indices of structural
   habitat complexity: predicted and actual effects on a predator<sup>1</sup>s foraging success. *Marine Ecology Progress Series*, 206, 45–58.
- 543 Bartoń, K. (2020). Multi-Model Inference. R. .
- Bernays, E. & Graham, M. (1988). On the Evolution of Host Specificity in Phytophagous
  Arthropods. *Ecology*, 69, 886–892.
- Best, R.J., Caulk, N.C. & Stachowicz, J.J. (2013). Trait vs. phylogenetic diversity as predictors
   of competition and community composition in herbivorous marine amphipods. *Ecology Letters*, 16, 72–80.
- Best, R.J. & Stachowicz, J.J. (2014). Phenotypic and phylogenetic evidence for the role of food
   and habitat in the assembly of communities of marine amphipods. *Ecology*, 95, 775–786.
- Cadotte, M.W., Carscadden, K. & Mirotchnick, N. (2011). Beyond species: functional diversity
   and the maintenance of ecological processes and services. *Journal of Applied Ecology*,
   48, 1079–1087.
- Cavender-Bares, J., Kozak, K.H., Fine, P.V.A. & Kembel, S.W. (2009). The merging of
   community ecology and phylogenetic biology. *Ecology Letters*, 12, 693–715.
- Chaudhary, C., Saeedi, H. & Costello, M.J. (2017). Marine Species Richness Is Bimodal with
   Latitude: A Reply to Fernandez and Marques. *Trends in Ecology & Evolution*, 32, 234–
   237.
- Chesson, P. (2000). Mechanisms of Maintenance of Species Diversity. *Annu. Rev. Ecol. Syst.*,
  31, 343–366.
- Denelle, P., Violle, C. & Munoz, F. (2019). Distinguishing the signatures of local environmental
   filtering and regional trait range limits in the study of trait–environment relationships.
   *Oikos*, 128, 960–971.
- Duffy, J.E. & Hay, M.E. (1991). Food and Shelter as Determinants of Food Choice by an
   Herbivorous Marine Amphipod. *Ecology*, 72, 1286–1298.
- Duffy, J.E., Richardson, J.P. & Canuel, E.A. (2003). Grazer diversity effects on ecosystem
   functioning in seagrass beds. *Ecology Letters*, 6, 637–645.
- Duffy, J.E., Richardson, J.P. & France, K.E. (2005). Ecosystem consequences of diversity
   depend on food chain length in estuarine vegetation. *Ecology Letters*, 8, 301–309.
- 570 Dyer, L.A. & Forister, M.L. (2019). Challenges and advances in the study of latitudinal gradients
   571 in multitrophic interactions, with a focus on consumer specialization. *Current Opinion in* 572 *Insect Science*, Ecology Parasites/Parasitoids/Biological control, 32, 68–76.
- Edgar, G.J. (1990). Population regulation, population dynamics and competition amongst mobile
  epifauna associated with seagrass. *Journal of Experimental Marine Biology and Ecology*,
  144, 205–234.
- Ford, B.M. & Roberts, J.D. (2018). Latitudinal gradients of dispersal and niche processes
  mediating neutral assembly of marine fish communities. *Mar Biol*, 165, 94.
- Ford, B.M. & Roberts, J.D. (2019). Evolutionary histories impart structure into marine fish
   heterospecific co-occurrence networks. *Global Ecology and Biogeography*, 28, 1310–
   1324.
- Fox, J. & Weisberg, S. (2019). An R Companion to Applied Regression. 3rd edn. Sage, Thousand
  Oaks, CA.
- Freestone, A.L. & Osman, R.W. (2011). Latitudinal variation in local interactions and regional
   enrichment shape patterns of marine community diversity. *Ecology*, 92, 208–217.

- Garcia, H.E., Locarnini, R.A., Boyer, T.P., Antonov, J.I., Zweng, M.M., Baranova, O.K., *et al.*(2010). *World Ocean Atlas 2009, Volume 4: Nutrients (phosphate, nitrate, silicate).*World Ocean Atlas 2009. U.S. Government Printing Office, Washington, D.C.
- Green, E.P. & Short, F.T. (2003). World Atlas of Seagrasses. University of California Press,
   Berkeley, CA, USA.
- den Hartog, C. (1970). *The seagrasses of the world*. North Holland Publishing Co., Amsterdam.
- 591 Hijmans, R.J. & Etten, J. van. (2020). raster: Geographic Data Analysis and Modeling. R.
- Jaramillo, E., Dugan, J.E., Hubbard, D.M., Contreras, H., Duarte, C., Acuña, E., *et al.* (2017).
   Macroscale patterns in body size of intertidal crustaceans provide insights on climate
   change effects. *PLOS ONE*, 12, e0177116.
- Kembel, S.W., Cowan, P.D., Helmus, M.R., Cornwell, W.K., Morlon, H., Ackerly, D.D., *et al.*(2010). Picante: R tools for integrating phylogenies and ecology. *Bioinformatics*, 26,
  1463–1464.
- Kraft, N.J.B., Adler, P.B., Godoy, O., James, E.C., Fuller, S. & Levine, J.M. (2015). Community
  assembly, coexistence and the environmental filtering metaphor. *Functional Ecology*, 29,
  592–599.
- Laliberté, É., Legendre, P. & Shipley, B. (2014). Measuring functional diversity (FD) from
   multiple traits, and other tools for functional ecology. R. .
- Lasley-Rasher, R.S., Rasher, D.B., Marion, Z.H., Taylor, R.B. & Hay, M.E. (2011). Predation
   constrains host choice for a marine mesograzer. *Marine Ecology Progress Series*, 434,
   91–99.
- Lavender, J.T., Dafforn, K.A., Bishop, M.J. & Johnston, E.L. (2017). An empirical examination
   of consumer effects across twenty degrees of latitude. *Ecology*, 98, 2391–2400.
- Leibold, M.A., Chase, J.M. & Ernest, S.K.M. (2017). Community assembly and the functioning
   of ecosystems: how metacommunity processes alter ecosystems attributes. *Ecology*, 98,
   909–919.
- Longo, G.O., Hay, M.E., Ferreira, C.E.L. & Floeter, S.R. (2019). Trophic interactions across 61
   degrees of latitude in the Western Atlantic. *Global Ecology and Biogeography*, 28, 107–
   117.
- Martin, T.L. & Huey, R.B. (2008). Why "Suboptimal" Is Optimal: Jensen's Inequality and
   Ectotherm Thermal Preferences. *The American Naturalist*, 171, E102–E118.
- Mayfield, M.M. & Levine, J.M. (2010). Opposing effects of competitive exclusion on the
   phylogenetic structure of communities. *Ecology Letters*, 13, 1085–1093.
- Mittelbach, G.G. & Schemske, D.W. (2015). Ecological and evolutionary perspectives on
   community assembly. *Trends in Ecology & Evolution*, 30, 241–247.
- Olsen, J.L., Stam, W.T., Coyer, J.A., Reusch, T.B.H., Billingham, M., Boström, C., *et al.* (2004).
   North Atlantic phylogeography and large-scale population differentiation of the seagrass
   Zostera marina L. *Molecular Ecology*, 13, 1923–1941.
- Pavoine, S. & Bonsall, M.B. (2011). Measuring biodiversity to explain community assembly: a
  unified approach. *Biological Reviews*, 86, 792–812.
- Pianka, E.R. (1966). Latitudinal Gradients in Species Diversity: A Review of Concepts. *The American Naturalist*, 100, 33–46.
- Poff, N.L. (1997). Landscape Filters and Species Traits: Towards Mechanistic Understanding
   and Prediction in Stream Ecology. *Journal of the North American Benthological Society*,
   16, 391–409.
- 630 R Development Core Team. (2021). R: A Language and Environment for Statistical Computing.

- Reynolds, P.L., Stachowicz, J.J., Hovel, K., Boström, C., Boyer, K., Cusson, M., *et al.* (2018).
   Latitude, temperature, and habitat complexity predict predation pressure in eelgrass beds
   across the Northern Hemisphere. *Ecology*, 99, 29–35.
- Roy, K., Hunt, G., Jablonski, D., Krug, A.Z. & Valentine, J.W. (2009). A macroevolutionary
  perspective on species range limits. *Proceedings of the Royal Society B: Biological Sciences*, 276, 1485–1493.
- Rumm, A., Foeckler, F., Dziock, F., Ilg, C., Scholz, M., Harris, R.M.B., *et al.* (2018). Shifts in
   mollusc traits following floodplain reconnection: Testing the response of functional
   diversity components. *Freshwater Biology*, 63, 505–517.
- Sainte-Marie, B. (1991). A review of the reproductive bionomics of aquatic gammaridean
   amphipods: variation of life history traits with latitude, depth, salinity and superfamily.
   *Hydrobiologia*, 223, 189–227.
- Sanford, E. & Bertness, M. (2009). Latitudinal gradients in species interactions. In: *Marine Macroecology* (eds. Witman, J. & Roy, K.). University of Chicago Press, pp. 357–391.
- Schemske, D.W., Mittelbach, G.G., Cornell, H.V., Sobel, J.M. & Roy, K. (2009). Is There a
  Latitudinal Gradient in the Importance of Biotic Interactions? *Annual Review of Ecology, Evolution, and Systematics*, 40, 245–269.
- Sessa, E.B., Chambers, S.M., Li, D., Trotta, L., Endara, L., Burleigh, J.G., *et al.* (2018).
  Community assembly of the ferns of Florida. *American Journal of Botany*, 105, 549–564.
- 650 Skeels, A., Esquerré, D. & Cardillo, M. (2020). Alternative pathways to diversity across
  651 ecologically distinct lizard radiations. *Global Ecology and Biogeography*, 29, 454–469.
- Starko, S., Demes, K.W., Neufeld, C.J. & Martone, P.T. (2020). Convergent evolution of niche
   structure in Northeast Pacific kelp forests. *Functional Ecology*, 34, 2131–2146.
- Swenson, N.G. (2014). Functional and Phylogenetic Ecology in R. Use R! Springer-Verlag, New
   York.
- Thompson, P., Guzman, M., De Meester, L., Horváth, Z., Ptacnik, R., Vanschoenwinkel, B., *et al.* (2020). A process-based metacommunity framework linking local and regional scale community ecology. *Ecology Letters*, 23.
- Tyberghein, L., Verbruggen, H., Pauly, K., Troupin, C., Mineur, F. & Clerck, O.D. (2012). Bio ORACLE: a global environmental dataset for marine species distribution modelling.
   *Global Ecology and Biogeography*, 21, 272–281.
- Valentine, J.F. & Duffy, J.E. (2006). The Central Role of Grazing in Seagrass Ecology. In:
   *SEAGRASSES: BIOLOGY, ECOLOGYAND CONSERVATION* (eds. LARKUM, A.W.D.,
   ORTH, R.J. & DUARTE, C.M.). Springer Netherlands, Dordrecht, pp. 463–501.
- Vasseur, D.A., DeLong, J.P., Gilbert, B., Greig, H.S., Harley, C.D.G., McCann, K.S., *et al.* (2014). Increased temperature variation poses a greater risk to species than climate
   warming. *Proceedings of the Royal Society B: Biological Sciences*, 281, 20132612.
- Vermeij, G.J. (1991). Anatomy of an Invasion: The Trans-Arctic Interchange. *Paleobiology*, 17,
   281–307.
- Webb, C.O., Ackerly, D.D., McPeek, M.A. & Donoghue, M.J. (2002). Phylogenies and
  Community Ecology. *Annual Review of Ecology and Systematics*, 33, 475–505.
- Whalen, M.A., Whippo, R.D.B., Stachowicz, J.J., York, P.H., Aiello, E., Alcoverro, T., *et al.*(2020). Climate drives the geography of marine consumption by changing predator
  communities. *PNAS*, 117, 28160–28166.

Zvereva, E.L. & Kozlov, M.V. (2021). Latitudinal gradient in the intensity of biotic interactions
 in terrestrial ecosystems: Sources of variation and differences from the diversity gradient
 revealed by meta-analysis. *Ecology Letters*, 24, 2506–2520.

#### 679 Acknowledgements

680 We thank the many lab and field assistants that participated in this research and whose

- 681 contributions of time and effort were invaluable for making this project happen. The manuscript
- 682 was improved with comments from SP Lawler, ED Sanford, SY Strauss, and two anonymous
- referees. This research was funded by National Science Foundation grants to JED, JJS, and KAH
- 684 (NSF-OCE 1336206, OCE 1336905, and OCE 1336741). CB was funded by the Åbo Akademi
- 685 University Foundation. This manuscript was prepared as a chapter for CPG's doctoral
- 686 dissertation.
- 687

678

#### 688 Tables

Table 1. Traits used in analyses of ZEN peracarid communities. Full citations, as well as sources

690 for individual species traits, are listed in Appendix 1.

Trait	Туре	Values	Category	Interpretation	Citations
Maximum fecundity (number of eggs)	Ordered categorical	Very low (0 to <18), Low (18 to <31), Medium (31 to <65), High (65 to <135), Very high (>135)	Neither	Competitive ability, population resilience, population density	Sainte-Marie 1991, Best and Stachowicz 2013, Lefcheck and Duffy 2015, Ashford <i>et al.</i> 2018
Maximum adult length	Continuous	2-50 mm	Microhabitat	Susceptibility to predators, ability to occupy physical space	Sainte-Marie 1991, Best and Stachowicz 2013, Lefcheck and Duffy 2015, Ashford <i>et al.</i> 2018
Body shape	Categorical	Laterally compressed, Dorsoventrally compressed, Vermiform	Microhabitat	Ability to occupy physical space, palatability	Lefcheck and Duffy 2015
Living habit	Categorical	Free, Parasite/direct commensal, Tube/burrow dweller	Microhabitat	Degree of substrate association, substrate type, population density	Best and Stachowicz 2013, Ashford <i>et al.</i> 2018
Motility	Categorical	Swimmer, Crawler	Microhabitat	Susceptibility to predators, dispersal ability, degree of substrate association	Lefcheck and Duffy 2015, Ashford <i>et al.</i> 2018
Bioturbator?	Binary		Microhabitat	Degree of substrate association, substrate type	Ashford et al. 2018

Microalgae feeding	Binary	Diet			
Macroalgae feeding	Binary	Diet		Duffy and Harvilicz 2001, Best and Stachowicz 2012, 2013	
Seagrass feeding	Binary	Diet			
Seagrass detritus feeding	Binary	Diet	Dietary niche		
Suspension feeding	Binary	Diet	partitioning		
Detritivory, deposit feeding	Binary	Diet		2013	
Carnivory, parasitism, scavenging	Binary	Diet			

- 691
- Table 2. A priori models used to analyse site-level SES values. These 16 models were separately
- applied to 33 sets of SES values for different trait distance metrics, permutation algorithms,
- 694 species pools, and trait sets, for a total of 528 models.

Model name			Predictors		
Biogeography 1	Latitude				
Biogeography 2	Latitude	Continental Margin	Ocean		
Biogeography 3	Latitude	Continental Margin	Latitude × Continental Margin		
Biogeography 4	Latitude	Continental Margin	Ocean	Latitude × Continental Margin	
Biogeography 5	Latitude	Continental Margin	Ocean	Latitude × Continental Margin	Latitude × Ocean
Abiotic Environment	in-situ Temperature	in-situ Salinity	Mean Leaf % N		
Temperature Regime 1	Mean SST				
Temperature Regime 2	SST Range				
Temperature Regime 3	Mean SST	SST Range	Mean SST × SST Range		
Community	log(Mean Standard Total Crustacean Abundance)	Median Crustacean Size	-		
Total Biodiversity	log(Site Epifaunal Richness)				
Peracarid Biodiversity	log(Site Peracarid Richness)				
Habitat	PC1	PC2	log(Macroalgal Biomass + 1)		
Predation	arcsin(Mean Amphipod Predation)		Diomass + 1)		
Resource 1	log(Mean Epiphyte load)	log(Mean Chl a)			
Resource 2	$\sqrt{NO_2}$	Mean PAR			

695

#### 696 Figure legends

Figure 1. *Zostera* Experimental Network (ZEN) sites used in our analyses. Sites spanned 30° of
latitude on the Pacific and Atlantic coasts of North America and Eurasia, including the

Baltic and Mediterranean seas, covering most of the range of *Zostera marina* (eelgrass).
Colours indicate trait dispersion (SES<sub>MNTD</sub> calculated using the tip shuffle algorithm);
positive values of SES<sub>MNTD</sub> indicate greater dispersion in traits than expected from a
random draw from the global species pool, whereas negative values of SES<sub>MNTD</sub> indicate
clustering in traits relative to a random draw. See Fig. S1 for more detailed information
about site locations.

705 Figure 2. Trait dispersion (SES<sub>MNTD</sub>) in eelgrass-associated peracarid crustacean communities 706 across trait sets. In general, communities at sites in the Pacific Ocean were more 707 overdispersed, while communities at Atlantic sites were less dispersed than expected. The 708 dashed horizontal line represents an SES<sub>MNTD</sub> value of 0, indicating random assembly. 709 Asterisks indicate means significantly different from zero (two-tailed one-sample t tests; 710 see table S2); error bars represent standard errors. Figure shows SES<sub>MNTD</sub> calculated according to the Tip Shuffle permutation algorithm; results were comparable across 711 712 permutation algorithms and SES values.

713 Figure 3. The effects of predation (a), latitude (b), epiphyte load (c), and in-situ temperature (d) 714 on trait dispersion (SES<sub>MNTD</sub> using the tip shuffle algorithm) in univariate analyses. In all 715 of the best models of dispersion, sites with higher predation intensity had more 716 overdispersed communities, while those with lower predation intensity had more clustered communities (a;  $R^2 = 0.15$ , p = 0.012). In the best models that had a non-zero 717 718 latitude effect, sites at lower latitudes had more overdispersed communities, while those 719 at higher latitudes had more clustered communities. This effect was stronger in the Pacific than the Atlantic species pool (b;  $R^2 = 0.36$ , interaction p = 0.0076). In the best 720 721 models with a non-zero epiphyte effect, sites where eelgrass had lower epiphyte density 722 had more overdispersed communities, while sites with more heavily fouled blades had 723 clustered communities (c; plot shows SES<sub>MNTD</sub> for microhabitat traits in the Atlantic species pool;  $R^2 = 0.15$ , p = 0.046). In-situ temperature appeared only sporadically across 724 725 permutations and dispersion metrics, and was not significant for total trait dispersion ( $\mathbb{R}^2$ ) = 0.0094, p = 0.54). The dashed horizontal line represents an SES value of 0, indicating 726 727 random assembly; sites in **bold** italics are those for which SES is significantly different 728 from 0 at  $\alpha = 0.05$ . See Fig. S1 for an explanation of site codes.

729







#### Supplemental tables and figures

Table S1. Top models of trait dispersion (SES) selected by backwards elimination by AICc scores. Models highlighted in grey are discussed in the main text. (table uploaded in separate Excel document).

Table S2. Results of t-tests comparing average SES values within ocean basins to zero. SES values are calculated relative to the global species pool; p values in bold represent significance at an  $\alpha$  level of 0.05.

Ocean	Metric	Permutation Algorithm	Trait Set	Mean SES	t	df	p
Pacific	MPD	Independent Swap	All	0.393	2.27	19	0.0352
			Microhabitat	0.404	2.41	19	0.0261
			Diet	0.415	1.66	19	0.114
		Tip Shuffle	All	0.363	2.12	19	0.0479
			Microhabitat	0.412	2.86	19	0.0101
			Diet	0.381	1.56	19	0.135
	MNTD	Independent Swap	All	0.155	0.589	19	0.563
			Microhabitat	0.686	3.48	19	0.00254
			Diet	-0.0449	-0.155	17	0.879
		Tip Shuffle	All	0.221	0.855	19	0.403
			Microhabitat	0.737	3.73	19	0.00143
			Diet	0.263	0.684	14	0.505
Atlantic	MPD	Independent Swap	All	-0.156	-1.07	21	0.295
			Microhabitat	-0.0959	-0.518	21	0.61
			Diet	-0.0998	-0.532	20	0.601
		Tip Shuffle	All	-0.699	-4.23	21	0.000375
			Microhabitat	-0.505	-2.55	21	0.0185
			Diet	-0.382	-2.06	20	0.0531
	MNTD	Independent Swap	All	-0.364	-2.4	21	0.026
			Microhabitat	-0.314	-1.74	21	0.0974
			Diet	-0.272	-1.77	19	0.0935
		Tip Shuffle	All	-0.358	-2.35	21	0.0285
			Microhabitat	-0.3327	-1.69	21	0.011
			Diet	0.00933	0.0394	14	0.9691

Table S3. Average predation rate and epiphyte load across ocean basins. Values in the first two rows are mean  $\pm$  standard deviation. Values in the third row represent the results of two-sample t-tests on untransformed (predation) and log-transformed (epiphytes) data across oceans.

Ocean	Prop. Tethered	Prey Removed	g Epipinytes g Eelgrass		
Pacific	0.80	$\pm 0.20$	0.30 :	± 0.31	
Atlantic	0.64	$\pm 0.24$	0.13 :	± 0.12	
Difference	t = 2.18	p = 0.037	t = 1.13	p = 0.27	

Pron Tethered Prev Removed g Eninhytes g Eelgrass<sup>-1</sup> Oc \_\_\_\_



Figure S1. Hierarchical design of the ZEN 2014 seagrass ecosystem survey. Sites are nested in one of 22 areas: KO = South Korea; JS = southern Japan (Seto Inland Sea); JN = northern Japan (Hokkaido); SD = San Diego Bay, US; MX = Mexico (Pacific Baja California); SF = San Francisco Bay, US; BB = Bodega and Tomales Bays, US; OR = Oregon, US, BC = British Columbia, Canada; WA = Washington State, US; NC = North Carolina (Back Sound), US; VA = York River, Virginia, US; ES = Virginia Eastern Shore, US; LI = Long Island, US; MA = Massachusetts, US; QU = Quebec (St. Lawrence Estuary), Canada; PO = Algarve, Portugal; FR = Mediterranean France; UK = Wales, UK; IR = Ireland; FI = Archipelago Sea, Finland; SW = Swedish west coast. Numbers in parentheses indicate the number of sites in a given area.



Figure S2. Principal coordinates analysis (PCoA) biplot of peracarid species in our global species pool, based on Gower distances. Solid symbols represent species in trait space, with symbol shape and colour corresponding to where they were found in our samples; hollow symbols represent centroids for categorical traits. Traits were fairly independent, and few were strongly correlated. Additionally, locale was not a significant predictor of where a given species fell in trait space (PERMANOVA; pseudo  $F_{9,95} = 0.98$ , p = 0.51). In other words, there were no traits that were particularly distinct to regions. Amphi-Pacific and Amphi-Atlantic distributions refer to species that occur in both the western and eastern margins of the Pacific and Atlantic Oceans, respectively.



Figure S3. Peracarid species richnesses across the four coastlines observed in this study. 55 species were collected from Atlantic sites and 60 species were collected from Pacific sites. Of these, 15 species were collected from the Northwest Pacific, 48 species from the Northeast Pacific, 36 species from the Northwest Atlantic, and 24 species from the Northeast Atlantic. There were 37 genera in 24 families in the Pacific and 40 genera in 22 families in the Atlantic.



Figure S4. Principal component biplot for eelgrass habitat structure across sites. Most of the variation in eelgrass was between short canopies of dense shoots and taller canopies of sparser shoots. The first two principal components accounted for 85.64% of the total variation in habitat structure at the site level. Eelgrass beds in the Atlantic Ocean were mostly characterized by small, densely packed shoots, while those in the Northwest Pacific contained larger, sparser shoots. Northeast Pacific sites contained both of these bed types.



Figure S5. Effects of predation (a-c), latitude (d-f), and epiphyte load (g-h) in best models of site-level trait dispersion (SES values) across three species pools (a, d, g), 3 sets of traits (b, e, g), two permutation algorithms, and two dispersion metrics (c, f, h). Columns show mean effect sizes (across best models selected by AICc) averaged across species pools, trait sets, algorithms, and metrics where appropriate; error bars represent standard errors.



Figure S6. Maps of salinity (a), water column nitrate (b), in-situ temperature (c), and crustacean abundance (d) across sites and ocean basins. Of these predictor, only crustacean abundance was significantly greater in the Pacific ( $R^2 = 0.076$ , p = 0.043). See Fig. S1 for more detailed information about site locations.



Figure S7. Maps of predation intensity (a), epiphyte load (b), mean annual sea surface temperature (c), and epifaunal species richness (d) across sites and ocean basins. None of these predictors varied significantly by ocean basin. See Fig. S1 for more detailed information about site locations.



Figure S8. Predictors used in models of dispersion, including predation intensity (a), epiphyte load (b), mean sea surface temperature (c), epifaunal richness (d), salinity (e), water column nitrate (f), in-situ temperature (g), and crustacean abundance (h), plotted against latitude. Without accounting for other variables, latitude was a significant predictor of mean sea surface temperature ( $R^2 = 0.58$ , p < 0.0001), site epifaunal richness (log-transformed;  $R^2 = 0.15$ , p = 0.0062), salinity ( $R^2 = 0.16$ , p = 0.0056), nitrate (square root-transformed;  $R^2 = 0.26$ , p = 0.00034), in-situ temperature ( $R^2 = 0.074$ , p = 0.046), and crustacean abundance (log-transformed;  $R^2 = 0.092$ , p = 0.029). Points represent sites, color-coded by ocean; Atlantic sites are in red, Pacific sites are in blue.