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Complete Genome Sequence of the Type Strain *Pectobacterium punjabense* SS95, Isolated from a Potato Plant with Blackleg Symptoms

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ABSTRACT *Pectobacterium punjabense* is a newly described species causing blackleg disease in potato plants. Therefore, by the combination of long (Oxford Nanopore Technologies, MinION) and short (Illumina MiSeq) reads, we sequenced the complete genome of *P. punjabense* SS95^T, which contains a circular chromosome of 4.793 Mb with a GC content of 50.7%.

The family *Pectobacteriaceae* encompasses pectinolytic plant pathogens that represent a threat to economically important vegetable crops and ornamental plants. *Pectobacterium* spp. are responsible for rotting diseases such as carrot or melon soft rot and potato blackleg and soft rot (1). Over the past decade, advances in genomics have allowed the scientific community to clarify the taxonomic position of many *Pectobacterium* species by either reexamining biological resources in the international collections or sampling a wider range of environments, from plants to surface waters (2–4). The type strain *Pectobacterium punjabense* SS95 (CFBP 8604, LMG 30622) was isolated from potato plants showing blackleg symptoms collected from Punjab, Pakistan, in 2017 (5). Serially diluted samples were plated onto crystal violet pectate (CVP) agar medium, and plates were incubated for 48 h at 28°C (6). Bacterial colonies producing pitting on CVP were purified on nutrient agar (beef extract [3 g], peptone [5 g], glucose [2.5 g], and agar [15 g per liter]). Genomic DNA was extracted using a MasterPure complete DNA purification kit (Epicentre, Madison, WI, USA). DNA quantification and quality control were performed using a Qubit 2.0 fluorometer and 1.0% agarose gel electrophoresis. Whole-genome shotgun DNA sequencing of *P. punjabense* SS95 was performed by a combination of Illumina MiSeq and Oxford Nanopore Technologies (ONT) MinION sequencing. The library was prepared using the Nextera DNA Flex kit (Illumina), and sequencing was performed using the MiSeq reagent kit v.2 with paired-end chemistry (2 × 150 bp). The sequence reads were trimmed to remove adapter sequences with Cutadapt v.1.15, and quality control was performed using FastQC v.0.11.5 (<http://www.bioinformatics.babraham.ac.uk/projects/fastqc>).

The ONT library preparation and sequencing were performed at the GeT-PlaGe core facility (INRA, Toulouse), according to the manufacturer's instructions, by following the 1D native genomic DNA barcoding protocol (EXP-NBD103 and SQK-LSK108). At each step, the DNA was quantified using the Qubit double-stranded DNA (dsDNA) high-sensitivity (HS) assay kit (Life Technologies). The DNA purity was tested using the NanoDrop spectrophotometer (Thermo Fisher), and the size distribution and degrada-

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