UCLA UCLA Previously Published Works

Title

Using excess deaths and testing statistics to improve estimates of COVID-19 mortalities.

Permalink https://escholarship.org/uc/item/5g3307jt

Journal European Journal of Epidemiology, 36(5)

ISSN 2331-8422

Authors

Böttcher, Lucas D'Orsogna, Maria R Chou, Tom

Publication Date 2021-05-01

DOI

10.1007/s10654-021-00748-2

Peer reviewed

COVID-19



Using excess deaths and testing statistics to determine COVID-19 mortalities

Lucas Böttcher^{1,2} · Maria R. D'Orsogna^{1,3} · Tom Chou^{1,4}

Received: 8 January 2021 / Accepted: 5 April 2021 © The Author(s) 2021

Abstract

Factors such as varied definitions of mortality, uncertainty in disease prevalence, and biased sampling complicate the quantification of fatality during an epidemic. Regardless of the employed fatality measure, the infected population and the number of infection-caused deaths need to be consistently estimated for comparing mortality across regions. We combine historical and current mortality data, a statistical testing model, and an SIR epidemic model, to improve estimation of mortality. We find that the average excess death across the entire US from January 2020 until February 2021 is 9% higher than the number of reported COVID-19 deaths. In some areas, such as New York City, the number of weekly deaths is about eight times higher than in previous years. Other countries such as Peru, Ecuador, Mexico, and Spain exhibit excess deaths significantly higher than their reported COVID-19 deaths. Conversely, we find statistically insignificant or even negative excess deaths for at least most of 2020 in places such as Germany, Denmark, and Norway.

Keywords COVID-19 · Excess deaths · Mortality · Test statistics

Introduction

The novel severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) first identified in Wuhan, China in December 2019 quickly spread across the globe, leading to the declaration of a pandemic on March 11, 2020 [1]. As of April 2021, an estimated 130 million people have been infected with COVID-19 disease, with more than 2.8 million deaths across 219 countries [2]. About 104 million people have recovered globally. Properly ascertaining the severity of any infectious

 ☑ Tom Chou tomchou@ucla.edu
 Lucas Böttcher lucasb@ucla.edu

> Maria R. D'Orsogna dorsogna@csun.edu

- ¹ Dept. of Computational Medicine, UCLA, Los Angeles, CA 90095-1766, USA
- ² Computational Social Science, Frankfurt School of Finance and Management, Frankfurt am Main 60322, Germany
- ³ Dept. of Mathematics, California State University at Northridge, Los Angeles, CA 91330-8313, USA
- ⁴ Dept. of Mathematics, UCLA, Los Angeles, CA 90095-1555, USA

disease is crucial for identifying near-future scenarios, and designing intervention strategies. This is especially true for SARS-CoV-2 given the relative ease with which it spreads, due to long incubation periods, asymptomatic carriers, and stealth transmissions [3]. Most measures of severity are derived from the number of deaths, the number of confirmed and unconfirmed infections, and the number of secondary cases generated by a single primary infection, to name a few. Estimating these quantities, determining how they evolve in a population, and how they are to be compared across groups, and over time, is challenging due to many confounding variables and uncertainties [4].

For example, quantifying COVID-19 deaths across jurisdictions must take into account the existence of different protocols in assigning cause of death, cataloging co-morbidities [5], and lag-time reporting. Inconsistencies also arise in the way deaths are recorded, especially when COVID-19 is not the direct cause of death, rather a co-factor leading to complications such as pneumonia and other respiratory ailments [6]. In Italy, the clinician's best judgment is called upon to classify the cause of death of an untested person who manifests COVID-19 symptoms. In some cases, such persons are given postmortem tests, and if results are positive, added to the statistics. Criteria vary from region to region [7]. In Germany, postmortem testing is not routinely employed, possibly explaining the large difference in mortality between the two countries at the onset of the pandemic. In the US, if typical symptoms are observed, the patient's death can be registered as due to COVID-19 even without a positive test [8]. Certain jurisdictions will list dates on which deaths actually occurred, others list dates on which they were reported, leading to potential lag-times. Other countries tally COVID-19 related deaths only if they occur in hospital settings, while others also include those that occur in private and/or nursing homes.

In addition to the difficulty in obtaining accurate and uniform fatality counts, estimating the prevalence of the disease is also a challenging task. Large-scale testing of a population where a fraction of individuals is infected, relies on unbiased sampling, reliable tests, and accurate recording of results. One of the main sources of systematic bias arises from the tested subpopulation: due to shortages in testing resources, or in response to public health guidelines, COVID-19 tests have more often been conducted on symptomatic persons, and the elderly. Such non-random testing may lead to overestimation the infected fraction of the population.

Different types of tests also probe different infected subpopulations. Tests based on reverse-transcription polymerase chain reaction (RT–PCR), whereby viral genetic material is detected primarily in the upper respiratory tract and amplified, probe individuals who are actively infected. Serological tests (such as enzyme-linked immunosorbent assay, ELISA) detect antiviral antibodies and thus measure individuals who have been infected, including those who have recovered.

Finally, different types of tests exhibit significantly different "Type I" (false positive) and "Type II" (false negative) error rates, FPR and FNR, respectively. The accuracy of RT–PCR tests depends on viral load which may be too low to be detected in individuals at the early stages of the infection, and may also depend on which sampling site in the body is chosen. Within serological testing, the kinetics of antibody response and antibody waning are still largely unknown [9, 10]. Instrumentation errors and sample contamination may also result in a considerable number of false positives and/or false negatives. Specifically, at low prevalence, Type I false positive errors can significantly bias the estimation of fatality measures.

Other quantities that are useful in tracking the dynamics of a pandemic include the number of recovered individuals, tested, or untested. These quantities may not be easily inferred from data and need to be estimated from fitting mathematical models such as SIR-type ODEs [11], agestructured PDEs [12], or network/contact models [13–15].

Administration of tests and estimation of all quantities above can vary widely across jurisdictions, making it difficult to properly compare numbers across them. In this paper, we incorporate excess death data, testing statistics, and mathematical modeling to self-consistently compute and compare mortality across different jurisdictions. In particular, we will use excess mortality statistics [16–18] to infer the number of COVID-19-induced deaths in different regions. We then present a statistical testing model to estimate jurisdictionspecific infected fractions and mortalities, their uncertainty, and their dependence on testing bias and errors. Our statistical analyses and source codes are available at [19].

Methods

Mortality measures

Different fatality rate measures have been used to quantify epidemic outbreaks [20]. One of the most common is the case fatality ratio (CFR) defined as the ratio between the number of confirmed "infection-caused" deaths D_c in a specified time window and the number of infections N_c confirmed within the same time window, CFR = D_c/N_c [21]. Depending on how deaths D_c are counted and how infected individuals N_c are defined, the operational CFR may vary and may exceed one if some deaths are not included in N_c .

Another frequently used measure is the infection fatality ratio (IFR) defined as the true number of "infection-caused" deaths $D = D_c + D_u$ divided by the actual number of cumulative infections to date, $N_c + N_u$. Here, D_u is the number of unreported infection-caused deaths within a specified period, and N_u denotes the untested or unreported infections during the same period. Thus, IFR = $D/(N_c + N_u)$.

One major limitation of both CFR and IFR is that they do not account for the time delay between infection and resolution. Both measures may be quite inaccurate, especially early in an outbreak when the number of cases grows faster than the number of deaths and recoveries [12]. An alternative measure that avoids case-resolution delays is the confirmed resolved mortality $M = D_c/(D_c + R_c)$ [12], where R_c is the cumulative number of confirmed recovered cases evaluated in the same specified time window over which D_c is counted. One may also define the true resolved mortality via $\mathcal{M} = D/(D+R)$, the proportion of the actual number of deaths relative to the total number of deaths and recovered individuals during a specified time period. If we decompose $R = R_c + R_u$, where R_c and $R_{\rm u}$ are the confirmed and unreported recovered cases, $\mathcal{M} = (D_c + D_u)/(D_c + D_u + R_c + R_u)$. The confirmed quantities are related through the total confirmed population $N_c = D_c + R_c + I_c$, where I_c the number of living confirmed infecteds. Applying these definitions to any specified time period (typically from the "start" of an epidemic to the date with the most recent case numbers), we observe that CFR $\leq M$ and IFR $\leq M$. After the epidemic has long past, when the number of currently infected individuals I approaches zero, the two fatality ratios and mortality measures converge if the component quantities are defined and

Table 1 Definitions of mortality measures. Quantities with subscript "c" and "u" denote confirmed (*i.e.*, positively tested) and unconfirmed populations. For instance, D_c , R_c , and N_c denote the total number of confirmed dead, recovered, and infected individuals, respectively. $d^{(j)}(i)$ is the number of individuals who have died in the *i*th time window (*e.g.*, day, week) of the *j*th previous year. The mean number of excess deaths \overline{D}_e within all periods i = 1, ..., k is thus

 $\sum_{i=1}^{k} \left[d^{(0)}(i) - \frac{1}{J} \sum_{j=1}^{J} d^{(j)}(i) \right].$ The total number of infection-caused deaths $D_c + D_u$ can be estimated using the excess deaths D_e as detailed in the main text. We have also included raw death numbers/100,000 and the mean excess deaths *r* relative to the mean number of deaths over the same period of time from past years [see Eq. (1)]

Subpopulation	Measure Z			
	Fatality Ratios	Resolved Mortality	Excess Death Indices	
Confirmed	$CFR = \frac{D_c}{N_c}$	$M = \frac{D_{\rm c}}{D_{\rm c} + R_{\rm c}}$	$D_{\rm e}$ per 100,000 : $\frac{D_{\rm c} + D_{\rm u}}{100,000}$	
Total	$IFR = \frac{D_{\rm c} + D_{\rm u}}{N_{\rm c} + N_{\rm u}}$	$\mathcal{M} = \frac{D_{\rm c} + D_{\rm u}}{D_{\rm c} + D_{\rm u} + R_{\rm c} + R_{\rm u}}$	Relative: $\bar{r} = \frac{\sum_{i} \left[d^{(0)}(i) - \frac{1}{J} \sum_{j}^{J} d^{(j)}(i) \right]}{\frac{1}{J} \sum_{j}^{J} \sum_{i} d^{(j)}(i)}$	

measured consistently, $\lim_{t\to\infty} \text{CFR}(t) = \lim_{t\to\infty} M(t)$ and $\lim_{t\to\infty} \text{IFR}(t) = \lim_{t\to\infty} \mathcal{M}(t)$ [12].

The mathematical definitions of the four basic mortality measures Z = CFR, IFR, M, M defined above are given in Table 1 and fall into two categories, confirmed and total. Confirmed measures (CFR and M) rely only on positive test counts, while total measures (IFR and M) rely on projections to estimate the number of infected persons in the total population N. Of the measures listed in Table 1, the fatality ratio CFR and confirmed resolved mortality M do not require estimates of unreported infections, recoveries, and deaths and can be directly derived from the available confirmed counts D_c , N_c , and R_c [22]. Estimation of IFR and the true resolved mortality M requires the additional knowledge on the unconfirmed quantities D_u , N_u , and R_u . We now describe the possible ways to estimate these quantities, along with the associated sources of bias and uncertainty.

Excess death data

An unbiased way to estimate $D = D_c + D_u$, the cumulative number of deaths, is to compare total deaths within a time window in the current year to those in the same time window of previous years, before the pandemic.

Since COVID-19 exhibits appreciable fatality, one may reasonably expect that most "excess" deaths can be attributed to the pandemic [23–27]. Within each affected region, these excess deaths D_e relative to "historical" deaths, are independent of testing limitations and do not suffer from highly variable definitions of COVID-induced death. Thus, D_e is a more inclusive measure of virus-induced deaths than D_c and can be used to estimate the total number of deaths, $D_e \approx D_c + D_u$. Moreover, using data from multiple past years, one can also estimate the uncertainty in D_e . In practice, deaths are tallied daily, weekly [23, 30], or in monthly aggregates [29, 31] with historical records dating back J years so that for every period *i* there are a total of J + 1 values. We denote by $d^{(j)}(i)$ the total number of deaths recorded in period *i* from the *j*th previous year where $0 \le j \le J$ and where j = 0 indicates the current year. To quantify the total cumulative excess deaths we define $d_e^{(j)}(i) = d^{(0)}(i) - d^{(j)}(i)$ as the excess deaths in period *i* relative to that of the *j*th previous year. Since $d^{(0)}(i)$ is the total number of deaths in week *i* of the current year, by definition $d_e^{(0)}(i) \equiv 0$. The excess deaths during week *i* averaged over *J* past years, $\bar{d}_e(i)$, and the associated, unbiased variance $\sigma_e(i)$ [32] are given by

$$\begin{split} \bar{d}_{\rm e}(i) &= \frac{1}{J} \sum_{j=1}^{J} d_{\rm e}^{(j)}(i), \\ \sigma_{\rm e}^2(i) &= \frac{1}{J-1} \sum_{j=1}^{J} \left[d_{\rm e}^{(j)}(i) - \bar{d}_{\rm e}(i) \right]^2. \end{split}$$
(1)

The corresponding quantities accumulated over *k* weeks define the mean and variance of the cumulative excess deaths $\bar{D}_{e}(k)$ and $\Sigma_{e}(k)$:

$$\begin{split} \bar{D}_{e}(k) &= \frac{1}{J} \sum_{j=1}^{J} \sum_{i=1}^{k} d_{e}^{(j)}(i) = \sum_{i=1}^{k} \bar{d}_{e}(i), \\ \Sigma_{e}^{2}(k) &= \frac{1}{J-1} \sum_{j=1}^{J} \sum_{i=1}^{k} \left[d_{e}^{(j)}(i) - \bar{d}_{e}(i) \right]^{2} = \sum_{i=1}^{k} \sigma_{e}^{2}(i), \end{split}$$

$$(2)$$

where deaths and their variances are accumulated from the first to the k^{th} week of the pandemic. The variance in Eqs. (1) and (2) arise from the variability in the number of deaths from the same time period across *J* previous years.

We gathered excess death statistics from over 75 countries and all US states. Some of the data derive from open-source online repositories as listed by official statistical bureaus and health ministries [23–27, 31]; other data are elaborated and tabulated in Ref. [29]. In some countries excess death statistics are available only for a limited number of states or jurisdictions (*e.g.*, Turkey and Indonesia). The US death statistics that



Fig. 1 Examples of seasonal mortality and excess deaths. The evolution of weekly deaths in (a) NYC (over seven years) and (b) Germany (over six years) derived from data in Refs. [28, 29]. Grey solid lines and shaded regions represent the historical numbers of deaths and corresponding confidence intervals defined in Eq. (1). Blue solid lines indicate weekly deaths, and weekly deaths that lie outside the confidence intervals are indicated by solid red lines. The red shaded regions represent statistically significant mean cumulative excess deaths D_e . The reported weekly confirmed deaths $d_c^{(0)}(i)$ (dashed black curves), reported cumulative confirmed deaths $D_c(k)$ (dashed dark red curves), weekly excess deaths $\bar{d}_e(i)$

we use in this study is based on weekly death data between 2015–2019 [31]. For all other countries, the data collection periods are summarized in Ref. [29]. Fig. 1(a-b) shows historical death data for New York City (NYC) and Germany, while Fig. 1(c-d) plots the associated confirmed and excess deaths and their confidence levels computed from Eqs. (1) and (2). We assumed that the cumulative summation is performed from the start of 2020 to the second week of February 2021 (week number k = 7). Significant numbers of excess deaths are clearly evident for NYC, while Germany had not experienced significant excess COVID-19 deaths in the first half of 2020.

To evaluate CFR and M, only data on D_c , N_c , and R_c are required, which are are tabulated by many jurisdictions. We can approximate the numerators of the IFR and \mathcal{M} by using, for example, the mean excess deaths $\bar{D}_e \approx D_c + D_u$ defined in Eq. (2). For the denominators, estimates of the unconfirmed infected N_u and unconfirmed recovered populations R_u are required. In the next two sections we propose methods to estimate N_u using a statistical testing model and R_u using a compartmental population model.

Statistical testing model with bias and testing errors

Testing bias and different sources of uncertainty in disease testing confound the estimation of the true prevalence of a disease. If not corrected, a sampling bias due to preferential testing of symptomatic individuals, healthcare workers, and

(solid grey curves), and cumulative excess deaths $\bar{D}_{\rm e}(k)$ (solid red curves) are plotted in units of per 100,000 in (c) and (d) for NYC and Germany, respectively. The excess deaths and the associated 95% confidence intervals given by the error bars are constructed from historical death data in (a-b) and defined in Eqs. (1) and (2). In NYC there is clearly a significant number of excess deaths that can be safely attributed to COVID-19, while in the first half of 2020 there had been no significant excess deaths in Germany. Excess death greater than reported confirmed deaths [with Germany an exception as shown in (d)]

certain high-risk groups [33] may lead to an overestimation of the fraction or total number of infected individuals. Since the total number of confirmed and unconfirmed cases, $N_c + N_u$, appears in the denominator of the IFR, we develop a statistical model for its estimation in the presence of testing errors and bias in administration of tests.

Although $N_c + N_u$ used to estimate the IFR includes those who have died, it may or may not include those who have recovered. If *S*, *I*, *R*, *D* are the numbers of susceptible, currently infected, recovered, and deceased individuals, the total population is N = S + I + R + D and the infected fraction can be defined as $f = (N_c + N_u)/N = (I + R + D)/N$ for tests that include recovered and deceased individuals (*e.g.*, antibody tests), or $f = (N_c + N_u)/N = (I + D)/N$ for tests that only count currently infected individuals (*e.g.*, RT–PCR tests). If we assume that the total population *N* can be inferred from census estimates, the problem of identifying the number of unconfirmed infected persons N_u is mapped onto the problem of identifying the true fraction *f* of the population that has been infected. Thus, we need to estimate *f* from testing results.

Figure 2 shows a schematic of a hypothetical initial total population of N = 54 individuals in a specified jurisdiction. Without loss of generality we assume there are no unconfirmed deaths, $D_u = 0$, and that all confirmed deaths are equivalent to excess deaths, so that $\bar{D}_e = D_c = 5$. Apart from the number of deceased, we also show the number



Fig. 2 Biased and unbiased testing of a population. A hypothetical scenario of testing a total population of N = 54 individuals within a jurisdiction (solid black boundary). Filled red circles represent the true number of infected individuals who tested positive and the black-filled red circles indicate individuals who have died from the infection. Open red circles denote uninfected individuals who were tested positive (false positives) while filled red circles with dark gray borders are infected individuals who were tested negative (false negatives). In the jurisdiction of interest, five have died of the infection while 16 are truly infected. The true fraction *f* of infected in the entire population is thus f = 16/54 and the true IFR = 5/16. However, under testing (green and blue) samples, a false positive is shown to

of true infected and uninfected subpopulations and label them as true positives, false positives, and false negatives. The true number of infected individuals is $N_c + N_u = 16$ which yields the true fraction f = 16/54 = 0.296 and an IFR = 5/16 = 0.312.

Also shown in Fig. 2 are two examples of sampling. Biased sampling and testing are depicted by the blue contour in which 6 of the 15 individuals are alive and infected, 2 are deceased, and the remaining 7 are healthy. For simplicity, we start by assuming no testing errors. The measured infected fraction of this sample, $\tilde{f}_b = 8/15 = 0.533 > f = 0.296$, is biased since it includes a higher proportion of infected persons than that of the entire jurisdiction. Using this biased measured fraction yields the apparent IFR = $5/(0.533 \times 54) \approx 0.174$, which significantly underestimates the true IFR ≈ 0.312 . A less-biased sample, shown by the green contour, yields an infected fraction of $4/14 \approx 0.286$ and an apparent IFR = $5/(0.286 \times 54) \approx 0.324$ which are much closer to the true fraction f = 0.296 and the true, "correct" IFR ≈ 0.312 .

In both samples discussed above we neglected testing errors such as false positives, as indicated in Fig. 2. Tests that are unable to distinguish false positives from true negatives would yield a larger N_c , resulting in an apparent infected fraction $\tilde{f}_b = 9/15$ and an even smaller apparent IFR ≈ 0.154 , as in the blue sample. By contrast, the false positive testing errors in the green sample would arise. If the measured positive fraction \tilde{f}_b is derived from a biased sample (blue), the estimated *apparent* IFR can be quite different from the true one. For a less biased (more random) testing sample (green), a more accurate estimate of the total number of infected individuals is $N_c + N_u = \tilde{f}_b N = (5/14) \times 54 \approx 19$ when the single false positive in this sample is included, and $\tilde{f}_b N = (4/14) \times 54 \approx 15$ when the single false positive case is excluded, and allows us to more accurately infer the IFR. Note that CFR is defined according to the tested quantities D_c/N_c which are precisely 2/9 and 2/5 for the blue and green sample, respectively, if false positives are considered. When false negatives are known and factored out CFR = 2/8 and 2/4, for the blue and green samples, respectively

yield an apparent infected fraction $\tilde{f}_b = 5/15 \approx 0.333$ and IFR ≈ 0.259 . Given a true infected fraction *f*, we now derive the probability of measuring the value \tilde{f}_b under biased testing and testing errors rates FPR and FNR.

We begin by proposing a parametric form for the apparent or measured (under biased testing) infected fraction

$$f_{\rm b} \equiv f_{\rm b}(f,b) \equiv \frac{fe^b}{f(e^b - 1) + 1}.$$
 (3)

Equation (3) allows us to connect f_b to the true infection fraction f. The bias parameter $-\infty < b < \infty$ in Eq. (3) describes how an infected or uninfected individual might be preferentially selected for testing, with b < 0 (and $f_b < f$) indicating under-testing of infected individuals, and b > 0(and $f_b > f$) representing over-testing of infecteds. A truly random, unbiased sampling arises only when b = 0 and $f_b = f$. Given Q administered tests to date, the number of recorded positive tests \tilde{Q}^+ , error rates FPR, FNR, and true infected fraction f, we derive in the Supplementary Information (SI) the likelihood of observing a positive fraction $\tilde{f}_b := \tilde{Q}^+/Q$

$$P(\tilde{f}_{\rm b}|f,\theta_{\rm T}) \approx \frac{1}{\sqrt{2\pi}\sigma_{\rm T}} \exp\left[-\frac{(\tilde{f}_{\rm b}-\mu)^2}{2\sigma_{\rm T}^2}\right],\tag{4}$$

in which

Table 2 Variables used in statistical testing model. A summary of the main variables that we use in the statistical testing model defined in Eqs. (3)–(6). Above, [0, N] and [0, Q] denotes a set of integer values, while \tilde{f}_b : [0, 1] denotes all rational numbers between 0 and 1. For f_b , FNR, FPR, [0, 1] represents all real numbers between 0 and 1. It is assumed that Q, \tilde{Q}^+ and $\tilde{f}_b := \tilde{Q}^+/Q$ are determined by testing a population of known size *N*

Symbol	Definition
$N \in \mathbb{Z}^+$	Population in jurisdiction
Q:[0, N]	Tests administered
\tilde{Q}^+ : $[0,Q]$	Recorded positive tests
$f = \frac{N_{\rm c} + N_{\rm u}}{N} : [0, 1]$	True fraction of infected individuals
$f_{\rm b} \equiv f_{\rm b}(f, b) : [0, 1]$	Infected fraction under biased testing
$\tilde{f}_{\rm b} := \frac{\tilde{Q}^+}{Q} : [0,1]$	Measured infected fraction under biased testing
$b \in \mathbb{R}$	Testing bias
FPR : [0, 1]	False positive rate
FNR: [0,1]	False negative rate

$$\mu \equiv f_{\rm b}(f,b)(1-{\rm FNR}) + (1-f_{\rm b}(f,b)){\rm FPR},$$

$$\sigma_{\rm T}^2 \equiv \mu(1-\mu)/Q.$$
(5)

Here, μ is the expected value of the measured and biased fraction \tilde{f}_b , and σ_T^2 is its variance. The parameters associated with testing are denoted $\theta_T = \{Q, b, FPR, FNR\}$, and may be time-dependent and may change from sample to sample.

Our formulae Eqs. (3), (4), and (5) provide relationships among the variables and testing parameters. In the testing context, Q, \tilde{Q}^+ , and thus $\tilde{f}_b = \tilde{Q}^+/Q$ will be measured and used to infer f, which we have explicitly separated out in Eq. (4) above. The goal is to use \tilde{f}_b as an estimate of f_b and ultimately estimate the true infected fraction f through Eq. (3). This requires *a-priori* knowledge of the parameters θ_T . For example, FPR and FNR are typically given by test specifications. To estimate b instead, one must conduct tests on a small truly random sample and compare results to those from a more widely used biased one. A summary of the main variables that we use in our statistical testing model is given in Table 2.

For a given measured value \tilde{f}_b , s simple maximum likelihood estimate of *f* can be found by maximizing Eq. (4) with respect to *f*, with all other parameters in θ_T specified,

$$\hat{f} \approx \frac{\tilde{f}_{\rm b} - \rm FPR}{e^b (1 - \rm FNR - \tilde{f}_{\rm b}) + \tilde{f}_{\rm b} - \rm FPR}.$$
(6)

Additional correction terms of $O(1/\sqrt{Q})$ are neglected. Note that although FNRs are typically larger than FPRs, small values of f and \tilde{f}_b imply that \hat{f} and μ are more sensitive to the FPR, as indicated by Eqs. (5) and (6).

If time series data for f_b are available, one can evaluate the estimated testing fractions in Eq. (6) for each time interval. Assuming that serological tests can identify infected individuals long after symptom onset, the latest value of \hat{f} would suffice to estimate corresponding mortality metrics such as the IFR. For RT–PCR testing, one generally needs to track how \tilde{f}_b evolves in time. A rough estimate would be to use the mean of \tilde{f}_b over the whole pandemic period to provide a lower bound of the estimated prevalence \hat{f} .

The fraction \tilde{f}_b measured under biased testing yields only the apparent IFR = $D_e/(\tilde{f}_bN)$ or expected apparent IFR = $\bar{D}_e/(\tilde{f}_bN)$, but Eq. (6) can then be used to estimate the true, corrected IFR $\approx \bar{D}_e/(\hat{f}N)$. For example, under moderate bias $|b| \leq 1$ and assuming FNR, FPR, $\tilde{f}_b \leq 1$, Eq. (6) can be used to relate the expected true IFR to the expected apparent IFR through $\bar{D}_e/(\hat{f}N) \approx (e^b + \text{FPR}/\hat{f})\bar{D}_e/(\tilde{f}_bN)$.

Another commonly used representation of the IFR is IFR = $p(D_c + D_u)/N_c = pD_e/N_c$. This expression is equivalent to IFR = $D_e/(f N)$ if $p = N_c/(N_c + N_u) \approx \tilde{Q}^+/(f N)$ is defined as the fraction of infected individuals that are confirmed [34, 35]. In this alternative representation, the *p* factor implicitly contains the effects of biased testing. Our approach allows the true infected fraction *f* to be directly estimated from \tilde{Q}^+ and *N*.

While the estimate \hat{f} needed to evaluate IFR depends strongly on *b* and FPR, and weakly on FNR, the *uncertainty* in \hat{f} will depend on the uncertainty in the values of *b*, FPR, and FNR. Although statistical models for inferring θ_T from data can be similarly constructed, here, we use a simple linear approximation to propagate uncertainty in the testing parameters to the squared coefficient of variation σ_f^2/\hat{f}^2 of the estimated infected fraction \hat{f} . The uncertainties in the mortality indices *Z* decomposed into the uncertainties in their respective individual components are listed in Table 3.

Using compartmental models to estimate resolved mortalities

Since the number of unreported or unconfirmed recovered individuals R_u required to calculate the total resolved mortality \mathcal{M} is not directly related to excess deaths nor to positive-tested populations, we use an SIR-type compartmental model to relate R_u to other inferable quantities [11]. Both unconfirmed recovered individuals and unconfirmed deaths D_u are related to unconfirmed infected individuals I_u who recover at rate γ_u and die at rate μ_u . The equations for the cumulative numbers of unconfirmed recovered individuals and unconfirmed deaths,

$$\frac{\mathrm{d}R_{\mathrm{u}}(t)}{\mathrm{d}t} = \gamma_{\mathrm{u}}(t)I_{\mathrm{u}}(t), \qquad \frac{\mathrm{d}D_{\mathrm{u}}(t)}{\mathrm{d}t} = \mu_{\mathrm{u}}(t)I_{\mathrm{u}}(t), \tag{7}$$

can be directly integrated to find $R_u(t) = \int_0^t \gamma_u(t')I_u(t')dt'$ and $D_u(t) = \int_0^t \mu_u(t')I_u(t')dt'$. The rates γ_u and μ_u may differ from those averaged over the entire population since testing may be biased towards subpopulations with different values of γ_u and μ_u . If one assumes γ_u and μ_u are approximately constant over the period of interest, we find $R_u/D_u \approx \gamma_u/\mu_u \equiv \gamma$. We now use $D_u \approx D_e - D_c$ to estimate $R_u \approx \gamma(D_e - D_c)$ and write \mathcal{M} as

$$\mathcal{M} = \frac{D_{\rm e}}{D_{\rm e} + R_{\rm c} + \gamma (D_{\rm e} - D_{\rm c})}.$$
(8)

An estimate for the expected value $\overline{\mathcal{M}}$ can be obtained by substituting the excess deaths $D_{\rm e}$ in Eq. (8) with $\overline{D}_{\rm e}$ from historical data.

Thus, a simple SIR model transforms the problem of determining the number of unreported death and recovered cases in $\overline{\mathcal{M}}$ to the problem of identifying the recovery and death rates in the untested population. Alternatively, we can make use of the fact that both the IFR and resolved mortality $\overline{\mathcal{M}}$ should have comparable values and match $\overline{\mathcal{M}}$ to IFR $\approx 0.1 - 1.5\%$ [35–37] by setting $\gamma \equiv \gamma_u/\mu_u \approx 100 - 1000$ (see SI for further information). Note that inaccuracies in confirming deaths may give rise to $D_c > \overline{D}_e$. Since by definition, infection-caused excess deaths must be greater than the confirmed deaths, we set $\overline{D}_e - D_c = 0$ whenever data happens to indicate \overline{D}_e to be less than D_c .

Results

Here, we present much of the available worldwide fatality data, construct the excess death statistics, and compute mortalities and compare them across jurisdictions. We show that standard mortality measures significantly underestimate the death toll of COVID-19 for most regions (see Figs. 1, A1, and A2). We also use the data to estimate uncertainties in the mortality measures and relate them to uncertainties of the underlying components and model parameters.

Excess and confirmed deaths

In NYC, the number of confirmed COVID-19 deaths between March 10, 2020 and December 10, 2020 is 19,694 [38] and thus significantly lower than the 27,938 (95% confidence interval [CI]: 26,516–29,360) reported excess mortality cases [23]. From March 25, 2020 until December 10, 2020, Spain counts 65,673 (99% CI: 91,816–37,061) excess deaths [24], a number that is substantially larger than the officially reported 47,019 COVID-19 deaths [39]. The large difference between excess deaths and reported COVID-19 deaths in Spain and NYC is also observed in Lombardia,



Fig. 3 Excess deaths versus confirmed deaths across different countries/states. The number of excess deaths \bar{D}_e as of March 30, 2021 [29, 31] (counted from January 2020 onwards) versus confirmed deaths D_c across different countries (a) and US states (b). The black solid lines in both panels have slope 1. In (a) the blue solid line is a guide-line with slope 3; in (b) the blue solid line is a least-squares fit of the data with slope 1.087 (95% CI: 1.052–1.121); blue shaded region). All data were updated on March 30, 2021 [22, 29, 31, 40]

one of the most affected regions in Italy. From February 23, 2020 until April 4, 2020, Lombardia reported 8,656 reported COVID-19 deaths [39] but 13,003 (95% CI: 12,335–13,673) excess deaths [27]. Starting April 5 2020, mortality data in Lombardia stopped being reported in a weekly format. In England/Wales, the number of excess deaths from the onset of the COVID-19 outbreak on March 1, 2020 until November 27, 2020 is 70,563 (95% CI: 52,250–88,877) whereas the number of reported COVID-19 deaths in the same time interval is 66,197 [28]. In Switzerland, the number of excess deaths from March 1, 2020 until November 29, 2020 is 5,664 (95% CI: 4,281–7,047) [26], slightly larger than the corresponding 4932 reported COVID-19 deaths [39].

To illustrate the significant differences between excess deaths and reported COVID-19 deaths in various jurisdictions, we plot the excess deaths against confirmed deaths for various countries and US states as of March 30, 2021 in Fig. 3. We observe in Fig. 3a that excess deaths in countries like Egypt, Mexico, Peru, Russia, and South Africa are significantly higher than the corresponding number of confirmed COVID-19 deaths. In particular, they were about three times higher than the number of reported COVID-19 in Egypt, Peru, Russia, and South Africa. For the majority of US states the number of excess deaths is also larger than the number of reported COVID-19 deaths, as shown in Fig. 3b. We performed a least-square fit to calculate the proportionality factor *m* arising in $\bar{D}_e = mD_c$ and found $m \approx 1.087$ (95% CI: 1.052–1.121). That is, across all US states, the number of excess deaths is about 9% larger than the number of confirmed COVID-19 deaths in the period from January 2020 until February 2021.

Estimation of mortality measures and their uncertainties

We now use excess death data and the statistical and modeling procedures to estimate mortality measures Z = IFR, CFR, M, M across different jurisdictions, including all US states and more than 75 countries.¹ Accurate estimates of the confirmed infecteds N_c and the confirmed dead D_c are needed to evaluate the CFR. Values for the parameters Q, FPR, FNR, and b are needed to estimate $N_c + N_u = f N$ in the denominator of the IFR, while \bar{D}_e is needed to estimate the number of infection-caused deaths $D_c + D_u$ that appear in the numerator of the IFR and M. Finally, since we evaluate the resolved mortality M, through Eq. (8), estimates of D_e, D_c, R_c, γ , and FPR, FNR (to correct for testing inaccuracies in D_c and R_c) are necessary. Whenever uncertainties are available or inferable from data, we also include them in our analyses.

Estimates of excess deaths and infected populations themselves suffer from uncertainty encoded in the variances Σ_e^2 and σ_f^2 . The uncertainty in f depends on the uncertainties arising from finite sampling sizes, uncertainty in bias b and uncertainty in test sensitivity and specificity, which are denoted σ_h^2 , σ_I^2 , and σ_{II}^2 , respectively. We use Σ^2 to denote variances in populations and σ^2 to denote variances in intrinsic parameters; covariances with respect to any two variables X, Y are denoted as $\Sigma_{X,Y}$. Variances in the confirmed populations are denoted $\Sigma_{N_c}^2, \Sigma_{R_c}^2$, and $\Sigma_{D_c}^2$ and also depend on uncertainties in testing parameters σ_{I}^{2} and σ_{II}^{2} . The most general approach would be to define a probability distribution or likelihood for observing a value of the mortality index $Z \in [z, z + dz]$. As outlined in the SI, these probabilities depend on the probability densities of the components of the mortalities, which in turn may depend on hyperparameters that define these probability densities. Here, we simply assume uncertainties that are propagated to the mortality indices through variances in the model parameters and hyperparameters [41]. The squared coefficients of variation

of the mortalities are found by linearizing them about the mean values of the underlying components and are listed in Table 3.

To illustrate the influence of different biases *b* on the IFR we use \hat{f} from Eq. (6) in the corrected IFR $\approx D_{\rm e}/(\hat{f}N)$. We model RT–PCR-certified COVID-19 deaths [42] by setting the FPR = 0.05 [43] and the FNR = 0.2 [44, 45]. The observed, possibly biased, fraction of positive tests $\tilde{f}_{\rm b}$ can be directly obtained from corresponding empirical data.

As of February 6, 2021, the average of $\tilde{f}_{\rm h}$ over all tests and across all US states is about 9.3% [46]. The corresponding number of excess deaths is $\bar{D}_e = 536,617$ [29] and the US population is about $N \approx 330$ million [47]. To study the influence of variations in $\tilde{f}_{\rm b}$, in addition to $\tilde{f}_{\rm b} = 0.089$, we also use a slightly larger $\tilde{f}_{\rm b} = 0.15$ in our analysis. In Fig. 4 we use the \bar{D}_{e} from the US and show the expected apparent and corrected IFRs for two values of $\tilde{f}_{\rm b}$ [Fig. 4a] and the coefficient of variation CV_{IFR} [Fig. 4b] as a function of the bias b. For unbiased testing [b = 0 in Fig. 4a], the corrected IFR in the US is 3.6% assuming $\tilde{f}_{\rm b} = 0.089$ and 1.4% assuming $\tilde{f}_{\rm b} = 0.15$. If b > 0, a higher proportion of the infected population is tested, hence, the expected apparent IFR = $\bar{D}_e/(\tilde{f}_b N)$ is smaller than the true IFR, as can be seen by comparing the solid (corrected IFR) and the dashed (apparent IFR) lines in Fig. 4a. For testing biased towards the uninfected population (b < 0), the corrected IFR may be smaller than the apparent IFR. To illustrate how uncertainty in FPR, FNR, and b affect uncertainty in \overline{IFR} , we evaluate CV_{IFR} as given in Table 3.

The first term in uncertainty σ_f^2/\hat{f}^2 given in Eq. (A6) in the SI is proportional to 1/Q and can be assumed to be negligibly small, given the large number Q of tests administered. The other terms in Eq. (A6) are evaluated by assuming $\sigma_b = 0.2, \sigma_I = 0.02$, and $\sigma_{II} = 0.05$ and by keeping FPR = 0.05 and FNR = 0.2. Finally, we infer Σ_e from empirical data using Eq. (2), neglect correlations between D_e and N, and assume that the variation in N is negligible so that $\Sigma_{e,N} = \Sigma_N \approx 0$. Fig. 4b plots $CV_{\overline{IFR}}$ and CV_{D_e} in the US as a function of the underlying bias b. The coefficient of variation CV_{D_e} is about 1%, much smaller than $CV_{\overline{IFR}}$, and independent of b. For the values of b shown in Fig. 4b, $CV_{\overline{IFR}}$ is between 51–68% for $\tilde{f}_b = 0.089$ and between 20–27% for $\tilde{f}_b = 0.15$.

Next, we compared the mortality measures $\overline{\text{IFR}}$, CFR, M, \overline{M} , and \overline{r} listed in Table 1 across numerous jurisdictions. To determine the CFR, we use the COVID-19 data of Refs. [22, 40]. For the expected apparent $\overline{\text{IFR}}$, we use the representation $\overline{\text{IFR}} = p\overline{D}_e/N_c$ discussed above. Although p may depend on the stage of the pandemic, typical estimates range from 4% [48] to 10% [35]. We set p = 0.1 over the lifetime of the pandemic. We can also directly use $\overline{\text{IFR}} = \overline{D}_e/(f N)$, however estimating the corrected $\overline{\text{IFR}}$ requires evaluating

¹ We provide an online dashboard that shows the real-time evolution of CFR and *M* at https://submit.epidemicdatathon.com/#/dashboard

Table 3 Uncertainty propagation for different mortality measures. Table of squared coefficients of variation $CV^2 = \Sigma_Z^2/Z^2$ for the different mortality indices Z derived using standard error propagation expansions [41]. Here, we have used the expected values IFR and $\overline{\mathcal{M}}$ and decomposed the CVs through variances about the mean values of all parameters such as \overline{D}_e . We use $\Sigma_N^2, \Sigma_{N_c}^2, \Sigma_R^2$, and Σ_D^2 to denote the uncertainties in the total population, confirmed cases, recoveries, and deaths, respectively. The variance of the number of excess deaths is Σ_e^2 , which feature in the IFR and \mathcal{M} . The uncertainty in the infected fraction σ_f^2 that contributes to the uncertainty in IFR depends on uncertainties in testing bias and testing errors as shown in Eq. (A6). The term Σ_{D_e,N_e} represents the covariance between D_e, N_e , and similarly for all other covariances $\Sigma_{e,N}$, Σ_{D_e,R_e} , Σ_{R_e,R_u} , $\Sigma_{R_e,Y}$. Since variations in D_e arise from fluctuations in past-year baselines and not from current intrinsic uncertainty, we can neglect correlations between variations in D_e and uncertainty in R_c, R_u . The last two rows represent \mathcal{M} expressed in two different ways, $\Gamma \equiv \bar{D}_e + R_c + R_u$ and $\bar{D}_e + R_c + \gamma(\bar{D}_e - D_c)$, respectively. Moreover, when using the SIR model to replace D_u and R_u with $\bar{D}_e - D_c \ge 0$, there is no uncertainty associated with D_u and R_u in a deterministic model. Thus, covariances cannot be defined except through the uncertainty in the parameter $\gamma = \gamma_u/\mu_u$

Mortality Z	Uncertainties	$CV^2 = \frac{\Sigma_Z^2}{Z^2}$
$CFR = \frac{D_c}{N_c}$	$\Sigma_{D_c}^2, \Sigma_{N_c}^2, \Sigma_{D_c,N_c}$	$\frac{\Sigma_{D_{\rm c}}^2}{D_{\rm c}^2} + \frac{\Sigma_{N_{\rm c}}^2}{N_{\rm c}^2} - \frac{2\Sigma_{D_{\rm c},N_{\rm c}}}{D_{\rm c}N_{\rm c}}$
$\overline{\rm IFR} \approx \frac{\bar{D}_{\rm e}}{f N}$	$\Sigma_{\mathrm{e}}^2, \Sigma_N^2, \Sigma_{\mathrm{e},N}, \sigma_f^2$	$\frac{\sigma_f^2}{\hat{f}^2} + \frac{\Sigma_e^2}{\bar{D}_e^2} + \frac{\Sigma_N^2}{N^2} - \frac{2\Sigma_{e,N}}{\bar{D}_e N}$
$M = \frac{D_{\rm c}}{D_{\rm c} + R_{\rm c}}$	$\Sigma_{D_c}^2, \Sigma_{R_c}^2, \Sigma_{D_c,R_c}$	$M^2 \left(\frac{R_{\rm c}}{D_{\rm c}}\right)^2 \left[\frac{\Sigma_{D_{\rm c}}^2}{D_{\rm c}^2} + \frac{\Sigma_{R_{\rm c}}^2}{R_{\rm c}^2} - \frac{2\Sigma_{R_{\rm c},D_{\rm c}}}{R_{\rm c}D_{\rm c}}\right]$
$\overline{\mathcal{M}} \approx \frac{\bar{D}_{\rm e}}{\bar{D}_{\rm e} + R_{\rm c} + R_{\rm u}}$	$\Sigma_{e}^{2}, \Sigma_{R_{c}}^{2}, \Sigma_{R_{u}}^{2}, \Sigma_{R_{u}}, \Sigma_{R_{c},R_{u}}$	$(1 - \mathcal{M})^2 \frac{\Sigma_{\rm e}^2}{\bar{D}_{\rm e}^2} + \frac{\Sigma_{R_{\rm c}}^2}{\Gamma^2} + \frac{\Sigma_{R_{\rm u}}^2}{\Gamma^2} - \frac{2\Sigma_{R_{\rm c},R_{\rm u}}}{\Gamma^2}$
$\overline{\mathcal{M}} \approx \frac{\bar{D}_{\rm e}}{\bar{D}_{\rm e} + R_{\rm c} + \gamma (\bar{D}_{\rm e} - D_{\rm c})}$	$\Sigma_{R_{\rm c}}^2, \Sigma_{\rm e}^2, \Sigma_{R_{\rm c},\gamma}, \sigma_{\gamma}^2$	$(1 - \mathcal{M})^2 \frac{\Sigma_{\rm e}^2}{\bar{D}_{\rm e}^2} + \frac{\Sigma_{R_{\rm c}}^2}{\Gamma^2} + \frac{(\bar{D}_{\rm e} - D_{\rm c})^2 \sigma_{\gamma}^2}{\Gamma^2} - \frac{2(\bar{D}_{\rm e} - D_{\rm c}) \Sigma_{R_{\rm c},\gamma}}{\Gamma^2}$

the bias *b*. In Fig. 5a, we show the running values (up until March 30, 2021) of the relative excess deaths \bar{r} , the CFR, the apparent IFR, the confirmed resolved mortality *M*, and the true resolved mortality \overline{M} across all regions. In all cases we set p = 0.1, $\gamma = 100$. As illustrated in Fig. 5b, some mortality measures suggest that COVID-induced fatalities are lower in certain countries compared to others, whereas other measures indicate the opposite. For example, the expected total resolved mortality \overline{M} for Brazil is larger than for Russia and Mexico, most likely due to the relatively low number of reported excess deaths as can be seen from Fig. 3a. On the other hand, Brazil's values of CFR, IFR, and *M* are substantially smaller than those of Mexico [see Fig. 5b].

The distributions of all measures Z and relative excess deaths r across jurisdictions are shown in Fig. 5c–g and encode the global uncertainty of these indices. We also calculate the corresponding mean values across jurisdictions, and use the empirical cumulative distribution functions to determine confidence intervals. The average of the mortality indices values across all jurisdictions are $\langle \bar{r} \rangle = 0.121 (95\%$ CI: 0.000–0.594), $\langle CFR \rangle = 0.020 (95\%$ CI: 0.000–0.050), $\langle \overline{IFR} \rangle = 0.004 (95\%$ CI: 0.00–0.02), $\langle M \rangle = 0.039 (95\%$ CI: 0.000–0.242), and $\langle \overline{M} \rangle = 0.021 (95\%$ CI: 0.000–0.094). Here, the CI measures the variation across jurisdictions. To calculate $\langle M \rangle$ and $\langle \overline{M} \rangle$, we excluded countries with incomplete recovery data. The distributions plotted in Fig. 5c–g can be used to inform our analyses of uncertainty or heterogeneity as summarized in Table 3. For example, the overall variance Σ_Z^2 can be determined by fitting the corresponding empirical Z distribution shown in Fig. 5c–g. Table 3 displays how the related CV_Z^2 can be decomposed into separate terms, each arising from the variances associated to the components in the definition of Z. For concreteness, from Fig. 5e we obtain $CV_{\overline{IFR}}^2 = \Sigma_{\overline{IFR}}^2 / \langle \overline{IFR} \rangle^2 \approx 8.05$ which allows us to place an upper bound on σ_b^2 using Eq. (A6), the results of Table 3, and

$$\sigma_b^2 < \frac{(\tilde{f}_b - \text{FPR})^2}{\hat{f}^2 (1 - \hat{f})^2} \text{CV}_{\overline{1FR}}^2 \approx \frac{(\tilde{f}_b - \text{FPR})^2}{\hat{f}^2 (1 - \hat{f})^2} 8.05$$
(9)

or on σ_{I}^2 using $(1 - \hat{f})^2 \sigma_{\text{I}}^2 < (\tilde{f}_{\text{b}} - \text{FPR})^2 \text{CV}_{\overline{\text{IFR}}}^2$.

Finally, to provide more insight into the correlations between different mortality measures, we plot M against CFR and \mathcal{M} against IFR in Fig. 6. For most regions, we observe similar values of M and CFR in Fig. 6a. Although we expect $M \to CFR$ and $\mathcal{M} \to IFR$ towards the end of an epidemic, in some jurisdictions such as the UK, the Netherlands, and Sweden, $M \gg CFR$ due to unreported or incomplete reporting of recovered cases. About 30% of the regions that we show in Fig. 6b have an IFR that is approximately equal to $\overline{\mathcal{M}}$. Again, for regions such as Sweden and the



Fig. 4 Different mortality measures across different regions. (a) The expected apparent (dashed lines) and true (solid lines) $\overline{\text{IFRs}}$ in the US from December 29, 2019 up until February 6, 2021, estimated using excess mortality data. We set $\tilde{f}_b = 0.089, 0.15$ (black, red), $\overline{\text{FPR}} = 0.05$, $\overline{\text{FNR}} = 0.2$, and N = 330 million. For the expected true $\overline{\text{IFR}}$, we use \hat{f} as defined in Eq. (6). Unbiased testing corresponds to setting b = 0. For b > 0 (positive testing bias), infected individuals are overrepresented in the sample population. Hence, the corrected $\overline{\text{IFR}}$ is larger than the apparent $\overline{\text{IFR}}$. If b is sufficiently small (negative testing bias), the expected true $\overline{\text{IFR}}$ may be smaller than the expected apparent $\overline{\text{IFR}}$. (b) The coefficient of variation of \bar{D}_e (dashed line) and $\overline{\text{IFR}}$ (solid lines) with $\sigma_{\text{I}} = 0.02$, $\sigma_{\text{II}} = 0.05$, and $\sigma_b = 0.2$ (see Table 3)

Netherlands, \mathcal{M} is substantially larger than \overline{IFR} because of incomplete reporting of recovered cases.

Discussion

In this paper, we review running COVID mortality metrics starting from the onset of the pandemic through December 2020, or later dates where data were available. In the first few weeks of the initial COVID-19 outbreak in March and April 2020 in the US, the reported death numbers captured only about two thirds of the total excess deaths [17]. This mismatch may have arisen from reporting delays, attribution of COVID-19 related deaths to other respiratory illnesses, and secondary pandemic mortality resulting from delays in necessary treatment and reduced access to health care [17]. We also observe that the number of excess deaths in the Fall months of 2020 have been significantly higher than the corresponding reported COVID-19 deaths in many US states and countries. The weekly numbers of deaths in regions with a high COVID-19 prevalence were up to 8 times higher than in previous years. Among the countries that were analyzed in this study, the ten countries with the largest numbers of excess deaths since the beginning of the COVID-19 outbreak (all numbers per 100,000) are Peru (358), Russia (291), Lithuania (270), North Macedonia (270), Bulgaria (266), Mexico (241), Ecuador (237), Serbia (233), South Africa (217), and Poland (207). The ten countries with the lowest numbers of excess deaths since the beginning of the COVID-19 outbreak are Mongolia (-20), Malaysia (-16), Costa Rica (-15), Australia (-14), Japan (-11), Philippines (-11), New Zealand (-8), Singapore (-6), Mauritius (-6), and Georgia (-4) [29]. From this point of view, as of March, 2021, these countries have not experienced statistically significant higher mortality due to COVID-19.

The proposed use of excess deaths in standard mortality measures may provide a more meaningful estimate of infection-caused deaths, while errors in the estimates of the fraction of infected individuals in a population from testing can be corrected by estimating the testing bias, specificity, and sensitivity.

Underlying our use of excess deaths \bar{D}_{e} for evaluating the disease IFR and mortality \mathcal{M} is the assumption that behavioral changes during the pandemic (social distancing, maskwearing, lockdowns) had no appreciable effect on death. For example, the mean traffic deaths per month in Spain between 2011-2016 is about 174 persons [49], so any pandemic-related changes to traffic volumes would have little impact considering the much larger number of COVID-19 deaths. However, other deaths due to, e.g., increased suicides and deferred medical treatment may contribute significantly to \bar{D}_{e} . One could sharpen estimates of the true COVID-19 deaths by systematically analyzing the statistics of deaths from all reported causes using a standard protocol such as ICD-10 [50]. More research is necessary to disentangle the excess deaths that are directly caused by SARS-CoV-2 infections from those that result from postponed medical treatment [17], increased suicide rates [51], and other indirect factors contributing to an increase in excess mortality. Even if the numbers of excess deaths were accurately reported and known to be caused by a given disease, inferring the corresponding number of unreported cases (e.g., asymptomatic infections), which appears in the definition of the IFR and \mathcal{M} (see Table 1), is challenging and only possible if additional models and assumptions are introduced.

Different mortality measures are sensitive to different sources of uncertainty. Under the assumption that all excess deaths are caused by a given infectious disease (*e.g.*, COVID-19), the underlying error in the determined number of excess deaths can be estimated using historical death statistics from the same jurisdiction. Uncertainties



Fig. 5 Mortality characteristics in different countries and states. (a) Running vales of relative excess deaths \bar{r} , the CFR, the $\overline{\text{IFR}} = p\bar{D}_e/N_c$ with $p = N_c/(N_c + N_u) = 0.1$ [35], the confirmed resolved mortality M, and the expected true resolved mortality \overline{M} (using $\gamma = 100$) are plotted for various jurisdictions. (b) Different mortality measures may be higher or lower in different jurisdictions, providing ambiguous characterization of disease severeness. (c–g) The probability density functions (PDFs) of the mortality measures

shown in (a) and (b). Note that there are only very incomplete recovery data available for certain countries (*e.g.*, US and UK). For countries without recovery data, we could not determine M and \overline{M} . The number of jurisdictions that we used in (a) and (c–g) are 136, 247, 127, 191, and 75 for the respective mortality measures (from left to right). All data from January 2020 onwards were included and last updated on March 30, 2021 [22, 29, 31, 40]

in mortality measures can also be decomposed into the uncertainties of their component quantities, including that of the positive fraction f that in turn depend on uncertainties in the testing parameters. While we have considered only the average or last values of \tilde{f}_{b} , our framework can be straightforwardly extended and dynamically applied across successive time windows, using *e.g.*, Bayesian or Kalman filtering approaches.

As for all epidemic forecasting and surveillance, our methodology depends on the quality of excess death and COVID-19 case data and knowledge of testing parameters. For many countries, the lack of binding international reporting guidelines, testing limitations, and possible data tampering [52] complicates the application of our framework. A striking example of variability is the large discrepancy between mean excess deaths \bar{D}_e and confirmed deaths D_c across many jurisdictions which render mortalities that rely on D_c suspect.

Finally, we have not partitioned the excess deaths or mortalities into subpopulations in age or other attributes such as sex, co-morbidities, occupation, etc. By expanding our testing and modeling approaches on stratified data, one can also straightforwardly infer stratified mortality measures Z, providing additional informative indices for comparison.

Conclusions

Based on the data presented in Figs. 5 and 6, we conclude that the mortality measures \bar{r} , CFR, IFR, M, and \mathcal{M} defined in Table 1 may provide different characterizations of disease severity across jurisdictions due to differences in testing bias and reporting protocols. The propagation of uncertainty and coefficients of variation that we summarize in Table 3 can help quantify and compare errors arising in different mortality measures, thus informing our understanding of the actual death toll of COVID-19. Depending on the stage of an outbreak and the currently available disease monitoring data, certain



Fig. 6 Different mortality measures across different regions. We show the values of *M* and CFR (a) and $\overline{\mathcal{M}}$ (using $\gamma = 100$) and $\overline{\text{IFR}} = p\bar{D}_e/N_c$ with $p = N_c/(N_c + N_u) = 0.1$ [35] (b) for different jurisdictions. The black solid lines have slope 1. If jurisdictions do not report the number of recovered individuals, $R_c = 0$ and M = 1 [light red circles in (a)]. In jurisdictions for which the data indicate $\bar{D}_e < D_c$, we set $\gamma(\bar{D}_e - D_c) = 0$ in the denominator of $\overline{\mathcal{M}}$ which prevents it from becoming negative as long as $\bar{D}_e \ge 0$. All data were counted from January 2020 onwards and last updated on March 30, 2021 [22, 29, 31, 40]

mortality measures are preferable to others. If the number of recovered individuals is being monitored, the resolved mortalities M and \mathcal{M} should be preferred over CFR and IFR, since the latter suffer from errors associated with the time-lag between infection and resolution [12]. For estimating IFR and \mathcal{M} , we propose using excess death data and an epidemic model. In situations in which case numbers cannot be estimated accurately, the relative excess deaths r provides a complementary measure to monitor disease severity. Our analyses of different mortality measures reveal that

• The CFR and *M* are defined directly from confirmed deaths D_c and suffers from variability in its reporting. Moreover, the CFR does not consider resolved cases and is expected to evolve during an epidemic. Although *M* includes resolved cases, it also requires confirmed recovered cases R_c , adding to its variability across jurisdictions. Testing errors affect both D_c and R_c , but if the FNR and FPR are known, they can be controlled using Eq. (A3) given in the SI.

- The IFR requires knowledge of the true cumulative number of disease-caused deaths as well as the true number of infected individuals (recovered or not) in a population. We show how these can be estimated from excess deaths and testing, respectively. Thus, the IFR will be sensitive to the inferred excess deaths and from the testing (particularly from the bias in the testing). Across all countries analyzed in this study, we found a mean IFR of about 0.44% (95% CI: 0.0–2.0%), which is similar to the previously reported values between 0.1 and 1.5% [35–37].
- In order to estimate the true resolved mortality \mathcal{M} , an additional relationship is required to estimate the unconfirmed recovered population R_u . In this paper, we propose a simple SIR-type model in order to relate R_u to measured excess and confirmed deaths through the ratio of the recovery rate to the death rate. The variability in reporting D_c across different jurisdictions generates uncertainty in \mathcal{M} and reduces its reliability when compared across jurisdictions.
- The mortality measures that can most reliably be compared across jurisdictions should not depend on reported data which are subject to different protocols, errors, and manipulation/intentional omission. Thus, the per capital excess deaths and relative excess deaths *r* (see last column of Table 1) should provide the most consistent comparisons of disease mortality across jurisdictions, provided total deaths are accurately tabulated. However, they are the least informative in terms of disease severity and individual risk, for which *M* and *M* are better.
- Uncertainty in all mortalities Z can be decomposed into the uncertainties in component quantities such as the excess death or testing bias. We can use global data to estimate the means and variances in Z, allowing us to put bounds on the variances of the component quantities and/or parameters.

Parts of our framework can be readily integrated into or combined with mortality surveillance platforms such as the European Mortality Monitor (EURO MOMO) project [30] and the Mortality Surveillance System of the National Center for Health Statistics [23] to assess disease burden in terms of different mortality measures and their associated uncertainty.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s10654-021-00748-2.

Acknowledgements LB acknowledges financial support from the Swiss National Fund (P2EZP2_191888). The authors also acknowledge financial support from the Army Research Office (W911NF-18-1-0345), the NIH (R01HL146552), and the National Science Foundation (DMS-1814364, DMS-1814090).

Data availability All datasets used in this study are available from Refs. [23–27, 29, 31]. The CDC excess death numbers and CIs are based on an overdispersed Poisson generalized linear model [31]. For the excess death data of Ref. [29], we determined CIs using the linear regression model class in R. The source codes used in our analyses are publicly available at [19].

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- World Health Organization. WHO Director-General's opening remarks at the media briefing on COVID-19 - 11 March 2020. https://www.who.int/dg/speeches/detail/who-director-general-sopening-remarks-at-the-media-briefing-on-covid-19---11-march-2020, 2020a. Accessed: 2020-04-18.
- COVID-19 statistics. https://www.worldometers.info/coronavirus/, 2020. Accessed: 2021-04-01.
- Xi He, Lau Eric HY, Peng Wu, Xilong Deng, Jian Wang, Xinxin Hao, Chung Lau Yiu, Wong Jessica Y, Yujuan Guan, Xinghua Tan, et al. Temporal dynamics in viral shedding and transmissibility of COVID-19. Nat Med. 2020;26(5):672–5.
- Morris Chris, Reuben Anthony. Coronavirus: Why are international comparisons difficult?, BBC Reality Check. https://www. bbc.com/news/52311014, 2020. Accessed: 2020-09-13.
- Conditions contributing to deaths involving coronavirus disease 2019 (COVID-19), by age group and state, United States. https:// data.cdc.gov/NCHS/Conditions-contributing-to-deaths-involvingcorona/hk9y-quqm, 2020a. Accessed: 2020-09-26.
- Thomas Beaney, Clarke Jonathan M, Vageesh Jain, Kataria Golestaneh Amelia, Gemma Lyons, David Salman, Azeem Majeed. Excess mortality: the gold standard in measuring the impact of COVID-19 worldwide? J Royal SocMed. 2020;113(9):329–34.
- Graziano Onder, Giovanni Rezza, Silvio Brusaferro. Case-fatality rate and characteristics of patients dying in relation to COVID-19 in Italy. JAMA. 2020;323(18):1775–6.
- CDC. Understanding the Numbers: Provisional Death Counts and COVID-19. https://www.cdc.gov/nchs/data/nvss/coronavirus/ Understanding-COVID-19-Provisional-Death-Counts.pdf, 2020b. Accessed: 2020-10-06.
- Dan Jennifer M, Jose Mateus, Kato Yu, Hastie Kathryn M, Dawen Yu Esther, Faliti Caterina E, Alba Grifoni, Ramirez Sydney I, Sonya Haupt, April Frazier, et al. Immunological memory to SARS-CoV-2 assessed for up to 8 months after infection. Science. 2021;371(6529):eabf4063.
- Javier Ibarrondo F, Fulcher Jennifer A, David Goodman-Meza, Julie Elliott, Christian Hofmann, Hausner Mary A, Ferbas Kathie G, Tobin Nicole H, Aldrovandi Grace M, Yang Otto O. Rapid decay of anti-SARS-CoV-2 antibodies in persons with mild Covid-19. New Engl J Med. 2020;383(11):1085–7.
- 11. Keeling Matt J, Rohani Pejman. Modeling infectious diseases in humans and animals. Princeton, NJ. 2011.

- 12. Lucas Böttcher, Mingtao Xia, Tom Chou. Why case fatality ratios can be misleading: individual-and population-based mortality estimates and factors influencing them. Phys Biol. 2020;17:065003.
- Böttcher Lucas, Nagler Jan, Herrmann Hans J. Critical behaviors in contagion dynamics. Phys Rev Lett. 2017;118(8):088301.
- 14. Lucas Böttcher, Nino Antulov-Fantulin. Unifying continuous, discrete, and hybrid susceptible-infected-recovered processes on networks. Phys Rev Res. 2020;2(3):033121.
- Romualdo Pastor-Satorras, Claudio Castellano, Piet Van Mieghem, Alessandro Vespignani. Epidemic processes in complex networks. Rev Mod Phys. 2015;87(3):925.
- Samuel Faust Jeremy, Zhenqiu Lin, Del Rio Carlos. Comparison of estimated excess deaths in new york city during the COVID-19 and 1918 influenza pandemics. JAMA Netw Open. 2020;3(8):e2017527–e2017527.
- Woolf Steven H, Chapman Derek A, Sabo Roy T, Weinberger Daniel M, Latoya Hill. Excess deaths from COVID-19 and other causes, March-April 2020. JAMA. 2020;324(5):510–3.
- Kontis Vasilis, Bennett James E, Rashid Theo, Parks Robbie M, Pearson-Stuttard Jonathan, Guillot Michel, Asaria Perviz, Zhou Bin, Battaglini Marco, Corsetti Gianni, et al. Magnitude, demographics and dynamics of the effect of the first wave of the COVID-19 pandemic on all-cause mortality in 21 industrialized countries. Nat Med. 2020;1–10.
- GitHub repository. https://github.com/lubo93/disease-testing, 2020.
- World Health Organization. Estimating mortality from COVID-19. Global infectious hazard preparedness, WHO Global: Department of Communications; 2020b.
- Zhe Xu, Lei Shi, Yijin Wang, Jiyuan Zhang, Lei Huang, Chao Zhang, Shuhong Liu, Peng Zhao, Hongxia Liu, Li Zhu, et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. Lancet Respir Med. 2020;8(4):420–2.
- Ensheng Dong, Hongru Du, Lauren Gardner. An interactive webbased dashboard to track COVID-19 in real time. Lancet Infect Dis. 2020.
- CDC. Pneumonia and Influenza Mortality Surveillance from the National Center for health statistics mortality surveillance system. https://gis.cdc.gov/grasp/fluview/mortality.html, 2020. Accessed: 2021-03-30.
- MoMo Spain. MoMo Spain. https://momo.isciii.es/public/momo/ dashboard/momo_dashboard.html#datos, 2020. Accessed: 2021-03-30.
- Office for National Statistics. Deaths registered weekly in England and Wales, provisional. https://www.ons.gov.uk/peoplepopulatio nandcommunity/birthsdeathsandmarriages/deaths/datasets/weekl yprovisionalfiguresondeathsregisteredinenglandandwales, 2020. Accessed: 2021-03-30.
- Bundesamt f
 ür Statistik. Sterblichkeit, Todesursachen. https:// www.bfs.admin.ch/bfs/de/home/statistiken/gesundheit/gesun dheitszustand/sterblichkeit-todesursachen.html, 2020. Accessed: 2020-07-16.
- 27. Istituto Nazionale di Statistica. Dati di mortalità: cosa produce l'Istat. https://www.istat.it/it/archivio/240401, 2020. Accessed: 2020-07-08.
- Epidemic Datathon. Summary of historical and current mortality data. https://www.epidemicdatathon.com/data, 2020. Accessed: 2021-03-30.
- 29. The Economist's tracker for COVID-19 excess deaths. https:// github.com/TheEconomist/covid-19-excess-deaths-tracker, 2020a. Accessed: 2021-03-30.
- 30. EURO MOMO. Mortality monitoring in Europe. https://www. euromomo.eu/index.html, 2020. Accessed: 2021-03-30.
- Excess Deaths Associated with COVID-19. https://www.cdc.gov/ nchs/nvss/vsrr/covid19/excess_deaths.htm, 2020b. Accessed: 2021-03-30.

- Holtzman Wayne H. The unbiased estimate of the population variance and standard deviation. Am J Psychol. 1950;63(4):615–7.
- Fenton Norman E, Martin Neil, Magda Osman, Scott McLachlan. COVID-19 infection and death rates: the need to incorporate causal explanations for the data and avoid bias in testing. J Risk Res. 2020;23(7–8):862–5.
- Ruiyun Li, Sen Pei, Bin Chen, Yimeng Song, Tao Zhang, Wan Yang, Jeffrey Shaman. Substantial undocumented infection facilitates the rapid dissemination of novel coronavirus (SARS-CoV-2). Science. 2020;368(6490):489–93.
- Chow Carson C, Chang Joshua C, Gerkin Richard C, Vattikuti Shashaank. Global prediction of unreported SARS-CoV2 infection from observed COVID-19 cases. *medRxiv*, 2020.
- 36. Henrik Salje, Tran Kiem Cécile, Noémie Lefrancq, Noémie Courtejoie, Paolo Bosetti, Juliette Paireau, Alessio Andronico, Nathanaël Hozé, Jehanne Richet, Claire-Lise Dubost, et al. Estimating the burden of SARS-CoV-2 in France. Science. 2020;369(6500):208–11.
- 37. Ioannidis John. The infection fatality rate of COVID-19 inferred from seroprevalence data. *medRxiv*; 2020.
- NYC Health. COVID-19: Data. https://www1.nyc.gov/site/doh/ covid/covid-19-data.page, 2020. Accessed: 2021-03-30.
- Liu Yi. COVID-19 Coronavirus Map repository. https://github. com/stevenliuyi/covid19, 2020. Accessed: 2020-04-20.
- 40. New York Times. Coronavirus (Covid-19) Data in the United States. https://github.com/nytimes/covid-19-data, 2020. Accessed: 2021-03-30.
- 41. Lee Eun Sul. Forthofer Ronald N. SAGE, Thousand Oaks, CA: Analyzing complex survey data; 2006.
- CDC. Guidance for certifying deaths due to coronavirus disease 2019 (COVID–19). https://www.cdc.gov/nchs/data/nvss/vsrg/ vsrg03-508.pdf, 2020a. Accessed: 2020-11-20.
- Watson Jessica, Whiting Penny F, Brush John E. Interpreting a COVID-19 test result. BMJ. 2020;369.
- Yicheng Fang, Huangqi Zhang, Jicheng Xie, Minjie Lin, Lingjun Ying, Peipei Pang, Wenbin Ji. Sensitivity of chest CT for COVID-19: comparison to RT-PCR. Radiology. 2020;296(2):E115-7.
- Wenling Wang, Yanli Xu, Ruqin Gao, Roujian Lu, Kai Han, Guizhen Wu, Wenjie Tan. Detection of SARS-CoV-2 in different types of clinical specimens. JAMA. 2020;323(18):1843–4.
- The COVID Tracking Project. https://covidtracking.com/data/ download/national-history.csv, 2021. Accessed: 2020-03-31.
- Census Bureau Estimates U.S. Population Reached 330 Million Today. https://data.cdc.gov/NCHS/Conditions-contributi ng-to-deaths-involving-corona/hk9y-quqm, 2020. Accessed: 2020-09-26.
- 48. Ali Hortaçsu, Jiarui Liu, Timothy Schwieg. Estimating the fraction of unreported infections in epidemics with a known epicenter:

an application to COVID-19. National Bureau of Economic Research: technical report; 2020.

- EUROSTAT. Death due to transport accidents, by sex. https:// ec.europa.eu/eurostat/databrowser/view/tps00165/default/table? lang=en, 2020. Accessed: 2020-04-20.
- CDC. International Classification of Diseases, Tenth Revision (ICD-10). https://www.cdc.gov/nchs/icd/icd10.htm, 2020b. Accessed: 2020-04-19.
- Leo Sher. The impact of the COVID-19 pandemic on suicide rates. QJM Int J Med. 2020;113(10):707–12.
- Laura Winter. Data fog: Why some countries' coronavirus numbers do not add up. https://www.aljazeera.com/indepth/features/ data-fog-countries-coronavirus-numbers-add-200607065953544. html, 2020. Accessed: 2020-09-14.
- U.S. Food and Drug Administration. EUA authorized serology test performance. https://www.fda.gov/medical-devices/emergencysituations-medical-devices/eua-authorized-serology-test-perfo rmance, 2020.
- 54. Cohen Andrew N, Kessel Bruce, Milgroom Michael G. Diagnosing COVID-19 infection: the danger of over-reliance on positive test results. *medRxiv*; 2020.
- Lassaunière Ria, Frische Anders, Harboe Zitta B, Nielsen Alex CY, Fomsgaard Anders, Krogfelt Karen A, Jørgensen Charlotte S. Evaluation of nine commercial SARS-CoV-2 immunoassays. *medRxiv*; 2020.
- 56. Whitman Jeffrey D, Hiatt Joseph, Mowery Cody T, Shy Brian R, Yu Ruby, Yamamoto Tori N, Rathore Ujjwal, Goldgof Gregory M, Whitty Caroline, Woo Jonathan M, et al. Test performance evaluation of SARS-CoV-2 serological assays. *medRxiv*; 2020.
- 57. Bastos Mayara Lisboa, Tavaziva Gamuchirai, Abidi Syed Kunal, Campbell Jonathon R, Haraoui Louis-Patrick, Johnston James C, Lan Zhiyi, Law Stephanie, MacLean Emily, Trajman Anete, et al. Diagnostic accuracy of serological tests for COVID-19: systematic review and meta-analysis. BMJ. 2020;370.
- 58. Ingrid Arevalo-Rodriguez, Diana Buitrago-Garcia, Daniel Simancas-Racines, Paula Zambrano-Achig, Del Campo Rosa, Agustin Ciapponi, Omar Sued, Laura Martinez-Garcia, Rutjes Anne W, Nicola Low, et al. False-negative results of initial RT-PCR assays for COVID-19: a systematic review. PLoS One. 2020;15(12):e0242958.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.