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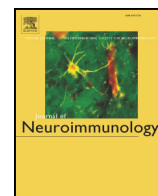
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Blood-CSF barrier and compartmentalization of CNS cellular immune response in HIV infection

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ABSTRACT

HIV infection is persistent in the CNS, to evaluate the compartmentalization of the CNS immune response to HIV, we compared soluble markers of cellular immunity in the blood and CSF among HIV[−] ($n = 19$) and HIV⁺ ($n = 68$), as well as among HIV participants with or without CSF pleocytosis. Dysfunction of the blood cerebrospinal fluid barrier (BCSFB) was common in HIV participants. CSF levels of TNF α , IFN γ , IL-2, IL-6, IL-7, IL-10, IP-10, MIP-1 α , MIP-1 β , and RANTES were significantly higher in participants with CSF pleocytosis ($P < 0.05$); serum levels of these biomarkers were comparable. The CNS immune response is compartmentalized, and remains so despite the BCSFB dysfunction during HIV infection; it is markedly reduced by virology suppression, although BCSFB dysfunction persists on this subgroup.

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1. Introduction

The CNS barriers, i.e. blood-brain barrier (BBB) and the blood-CSF barrier (BCSFB) are highly selective physiological barriers that restrict the passage of macromolecules from circulating blood to the CNS. These barriers protect neurons from constant changes in the concentration of blood constituents without restricting the transport of nutrients and metabolic products to and from the brain, respectively, as well as the bi-directional communication with the immune system (de Vries et al., 1997; Ballabh et al., 2004; Hamilton et al., 2007; Stamatovic et al., 2008; Abbott et al., 2010; Banks, 2015). Reductions in CSF production, release, and flow rate impair the BCSFB function, thereby increasing the ratio of CSF albumin to serum albumin (albumin quotient Q_{Alb}) (Kuehne et al., 2013).

In recent years, there has been a resurgence of interest in the physiology of the brain barriers, which may play critical roles in a wide range of neurological disorders, including those associated with HIV infection (Saunders et al., 2008). Studies conducted prior to the

development of highly active anti-retroviral therapy demonstrated that dysfunction of the BCSFB was common in HIV participants, particularly in those with increased CSF white blood cell (WBC) count (pleocytosis) (Marshall et al., 1988; Petito and Cash, 1992). More recent studies also found evidence of BCSFB dysfunction (Andersson et al., 2001; Calcagno et al., 2014), which can occur early in infection, and persist even with highly active anti-retroviral therapy (Calcagno et al., 2015).

HIV is a systemic infection, of which the nervous system is an important component (Brew et al., 1997). Indeed, HIV is highly neurotropic and neurovirulent, and causes persistent infection in the CNS. Invasion of the CNS during acute HIV infection (Resnick et al., 1988; Spudich et al., 2011; Valcour et al., 2013) initiates an inflammatory cascade that eventually compartmentalizes the CNS immune response even with anti-retroviral therapy. Thus we hypothesize that the inflammatory response in HIV infection is stronger in CNS than peripheral blood, characterizing compartmentalization of CNS cellular immune response; despite the dysfunction of the BCSFB during HIV infection. By compartmentalization of immune responses we mean evidence of immune activity, including production of signaling molecules such as chemokines and interleukins, that is CNS specific and would be missed by studying only plasma or serum.

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To evaluate the compartmentalization of the CNS immune response to HIV, we compared soluble markers of cellular immunity in the blood and CSF among uninfected and HIV-infected individuals, as well as among HIV participants with or without CSF pleocytosis. We selected biomarkers on the basis of their roles in cellular immunity, neuroinflammation, chemotaxis, and CNS HIV pathobiology (Nath, 1999; Genis et al., 1992; King et al., 2006; Campbell et al., 2007a; Abbas and Herbein, 2013). We also investigated the BCSFB function, and identified the origin of CSF biomarkers.

We anticipated that the data would provide important pathophysiological insights into the neurological impact of HIV and the specific immune response in the CNS. This study extends previous studies (Price et al., 2013; Peterson et al., 2014) by examining cellular immunity in the CSF and serum of participants infected with non-B HIV subtypes, noting that HIV-infected samples we tested were collected from the same geographic area. Indeed, previous studies focused almost exclusively on the effects of HIV-1 subtype B on inflammatory and chemotaxis biomarkers (Genis et al., 1992; Cinque et al., 2007; Brew and Letendre, 2008; Hagberg et al., 2010; Yuan et al., 2013).

2. Materials and methods

This study was a cross-sectional survey of stored CSF and serum samples, and was approved by institutional review boards at University of California San Diego, Hospital de Clínicas-UFPR in Brazil, and Brazil National IRB (CONEP). Participants and methods were described previously (de Almeida et al., 2013, 2016).

2.1. Subjects

A total of 87 paired CSF and serum samples were analyzed. Demographic characteristics, HIV status, and co-infections, if any, are summarized in Table 1 for HIV-positive ($n = 68$) and HIV-negative ($n = 19$) volunteers.

Table 1
Demographics, HIV infection characteristics, and co-morbidities of HIV participants and uninfected volunteers.

	HIV + ($n = 68$)	HIV – ($n = 19$)	<i>P</i>
<i>Demographics</i>			
Age, years	43 (35; 48)	41 (38; 50)	0.91
Education, years	8 (5; 11)	12 (11; 15)	0.0001
Gender, n male (%)	33 (49)	14 (74)	0.07
<i>HIV status and treatment</i>			
AIDS, n (%)	55 (81)	–	–
Duration of infection, months	99 (29; 140)	–	–
Current CD4	369 (201; 534)	–	–
Nadir CD4	92 (37; 267)	–	–
Log plasma HIV RNA ^a	1.7 (1.7; 3.5)	–	–
Plasma HIV RNA < 50 copies/mL, n (%)	38 (56)	–	–
Log CSF HIV RNA	1.7 (1.7; 2.8)	–	–
CSF HIV RNA < 50 copies/mL, n (%)	36 (53)	–	–
On CART ^b , n (%)	55 (81)	–	–
CPE ^c	8 (6; 9)	–	–
Adherence ^d , n (%)	51 (93)	–	–
<i>Co-morbidities</i>			
HCV ^e , n (%)	12 (18)	0	–
Log plasma HCV RNA, n	2.9 (1.7; 5.9)	0	–

Participants co-infected with HCV were not on treatment with interferon-gamma. Data are median (IQR) or number of cases (%).

^a Plasma viral load, log₁₀ c/mL.

^b CART, combination anti-retroviral therapy.

^c CPE, anti-retroviral CNS penetration effectiveness (Letendre et al., 2010).

^d Anti-retroviral treatment adherence was evaluated using AIDS clinical trial (ACTG) adherence questionnaire (4-day recall).

^e Hepatitis C virus (HCV) status was assessed by antibody testing (Abbott-Architect).

2.1.1. HIV participants

HIV participants were recruited at Hospital de Clínicas, Universidade Federal do Paraná, Curitiba, Paraná, Brazil. Individuals with opportunistic CNS infections were excluded. All volunteers provided blood and CSF samples, and underwent serological testing to confirm HIV status before enrollment, in accordance with guidelines published by the Brazilian Ministry of Health (2009). For participants with clinically resistant infection, the infecting HIV strain was genotyped using *pol* sequences, while *env* sequences were used for all other participants. Genotyping indicated that 27 individuals were infected with HIV subtype B, and 40 with non-B HIV subtypes (C-26, BF-10, BC-1, CF-1, and F-2). In one participant, the HIV subtype could not be determined.

2.1.2. Uninfected controls

As lumbar punctures could not be performed in uninfected volunteers in Brazil, we recruited a control group of 19 age-matched HIV-negative individuals at HIV Neurobehavioral Research Center, University of California San Diego. These volunteers were free of neurological comorbidities, and tested negative on serological tests for hepatitis C virus and syphilis. The neurochemical criteria for inclusion into the control group were CSF WBC count ≤ 5 cells/mm³, CSF total protein ≤ 45 mg/dL, and CSF glucosid ≥ 55 mg/dL.

2.2. Laboratory methods

Lumbar punctures were performed aseptically using an atraumatic spinal needle. CSF total protein and glucose were quantified by benzethonium chloride and hexokinase/G-6-PDH (Architect-Abbott, IL), respectively. Total WBC/mm³ was determined in fresh, uncentrifuged CSF by manual counting in a Fuchs-Rosenthal chamber. CSF pleocytosis was defined as WBC > 5 cells/mm³. For differential leukocyte counts, CSF samples were concentrated by Shandon Cytospin (Pittsburgh, USA), mounted, and stained by May-Grünwald-Giemsa technique. Aliquots of CSF and serum were refrozen and stored at -80 °C for subsequent batch testing for cytokines and chemokines.

2.2.1. Clinical laboratory parameters

HIV RNA levels in serum and CSF were quantified by branched DNA assay (Siemens) with nominal limit of detection 50 copies/mL. CD4 counts were quantified by flow cytometry (FACSCalibur-Multitest), while nadir CD4 was extracted from medical records.

2.2.1.1. CSF and serum biomarkers. RANTES was quantified by high-sensitivity enzyme-linked immunosorbent assay (R&D Systems, Minneapolis, MN). TNF α , IFN γ , IL-1 β , IL-2, IL-4, IL-6, IL-7, IL-10, MCP-1, MIP-1 α , MIP-1 β , and IP-10 were quantified by multiplex bead suspension array immunoassays (EMD Millipore, Billerica, MA), in which primary antibodies are immobilized to dyed fluorescent microspheres. The analytic sensitivity of these assays ranged between 0.1 and 10.1 pg/mL. All samples were assayed concurrently in duplicate according to manufacturers' instructions.

2.2.1.2. Blood-CSF barrier function. CSF and serum albumin were quantified by the nephelometric method (Dade Behring BNII, Deerfield, IL). The functional integrity of the blood-CSF barrier was then assessed by the CSF albumin/serum albumin quotient, which was calculated as $Q_{Alb} = Alb_{CSF} / Alb_{serum}$. The upper limit of the reference range for Q_{Alb} is age-dependent, was calculated for each participant according to the equation: $4 + \text{age in years} / 15$ (Reiber and Peter, 2001). The albumin and IgG hyperbolic function was calculated using the Reibergram plot (Reiber, 1995). Albumin leakage across the blood-brain barrier over 24 h was also calculated (Tourtellotte et al., 1989).

2.3. Data analyses

Demographic data, HIV disease characteristics, and CSF biochemical, cytological, and virological measures were compared between groups using independent samples *t*-tests for continuous variables and Fisher's exact test for binary and categorical variables (gender, AIDS diagnostic, ART, HCV serostatus, HIV RNA in plasma and CSF). The demographic data and CSF biochemical and cytological measures were compared between the HIV-positive and the HIV-negative control groups using similar methods. The distribution of plasma HIV viral loads was highly skewed so the Wilcoxon rank-sum test was used.

In order to detect the source of the biomarkers in CSF was calculated the coefficient of variation (CV) for each biomarker in CSF and serum in the normal control group; to the evaluation of the inter-individual variation. The comparison of biological variation of molecule concentration can be used to compare functionally connected compartments of the same group; if we could expect variation of propagation for molecules due to the passage from one compartment to the other (Kuehne et al., 2013).

The CSF and serum biomarker values were log₁₀-transformed to normalize their distributions, and presented in terms of mean (SD), since they are approximately normal on the log scale.

The biomarkers were compared between the HIV-positive and HIV-negative control groups as well as between the HIV-positive samples with pleocytosis in the CSF and HIV-positive samples with a normal CSF WBC count. We used a multivariable linear regression (adjusted analysis), controlling for plasma HIV viral load suppression, and nadir CD4 count, which has been shown in previous studies to be associated with increased soluble biomarkers of inflammation and chemotaxis in HIV (Gisslen et al., 1994; Mooney et al., 2015; Noel et al., 2014). In a second moment results were adjusted for Q_{Alb} for the comparison of groups HIV + and HIV –; and Q_{Alb} in addition to suppressed plasma and CSF viral load, and CD4 nadir for the comparison of the groups with and without CSF pleocytosis.

Correlations between variables were calculated using Spearman's rank-order correlation. Results were considered statistically significant at the 5% alpha level. Cohen's *d* effect sizes (and 95% confidence intervals) were reported for differences between groups.

3. Results

As shown in Table 1, ART was prescribed to 55 (81%) of the HIV + participants. The most frequent ART regimen (33 participants; 60%) contained a ritonavir-boosted HIV protease inhibitor (PI) plus two nucleoside/nucleotide reverse transcriptase inhibitors (NRTI; NtRTI); 18 participants (33%) received a non-nucleoside RT inhibitor (NNRTI) with 2 NRTIs, and 4 (7%) participants received other regimen types. Darunavir was not available in Brazil during the time that these samples were collected. Because of this the Brazilian National HIV Therapy protocol recommended that patients with resistance receive a regimen of an NNRTI combined with 2 NRTIs and ritonavir-boosted lopinavir. The number of participants on ARV and plasma HIV RNA <50 copies/mL was 37 (67%).

3.1. CSF profile and HIV RNA levels

The biochemical, cytological, and virological characteristics of the CSF in uninfected and HIV participants are summarized in Table 2. CSF pleocytosis was detected exclusively in 20/68 (29.4%, $P < 0.0001$) among HIV participants. CSF pleocytosis can be attributed solely to HIV infection, since we excluded other opportunistic or co-infections by standard etiological investigation methods. All participants in this study were ambulatory volunteers without signs and symptoms of active opportunistic disease or co-infections. In addition, all CSF samples with pleocytosis underwent standard microbiological tests to exclude coinfections including gram smear and culture for bacterial infection;

Table 2

Biochemical, cytological, and virological characteristics of the CSF in HIV-positive and uninfected volunteers. Significant differences are highlighted in bold.

	HIV + (n = 68)	HIV – (n = 19)	P
WBC, cells/mm ³	2.1 (0.6; 7.2)	2 (1;2.5)	0.4024
WBC count >5 cells/mm ³	20 (29%)	0	<0.0001
Glucose, mg/dL	57 (53; 62)	63 (59; 71)	0.0003
Total protein, mg/dL	40 (32; 46)	30 (26; 38)	0.0026
Total protein >45 mg/dL	20 (29%)	0	0.0048
Albumin, mg/dL	22.4 (16.4; 28.9)	18 (15; 24)	0.019
Albumin quotient, Q _{Alb}	0.0064 (0.0049; 0.0097)	0.005 (0.004; 0.006)	0.0023
Lactic acid, mmol/L	1.6 (1.5; 1.8)	–	–
RBC, cells/mm ³	0.5 (0; 7.5)	2.0 (1.0, 4.0)	0.3993
Log CSF HIV RNA	1.7 (1.7; 2.8)	–	–
Viral load <50	35	–	–
HIV RNA CSF > blood	12 (18%)	–	–

CSF glucose was significantly higher in HIV-negative volunteers than in HIV participants, although levels in both groups were below reference range. Data are median (IQR) or number of cases (%).

direct CSF smear, culture and latex agglutination tests for *C. neoformans* capsular antigen; Ziehl smear, culture and PCR for *M. tuberculosis*; and PCR for HSV, enterovirus, VZV, CMV, HHV6, HHV7, EBV and JC virus.

3.2. Inflammatory biomarkers

3.2.1. Comparison between HIV-positive and HIV-negative groups

Cytokine and chemokine levels are listed in Table 3. In CSF, 9 of 13 cytokines and chemokines were significantly elevated ($P < 0.05$) in HIV participants than in uninfected controls. In contrast, only 4 of 13 markers were significantly elevated in serum (Table 3).

In unadjusted regression models, CSF levels of the cytokines TNF α , IFN γ , IL-1 β , IL-2, IL-7, and IL-10 were significantly elevated in HIV participants than in uninfected individuals. In contrast, only TNF α and IFN γ were significantly elevated in serum. CSF levels of the chemokines IP-10, MCP-1, and MIP-1 α were also significantly elevated in HIV participants than in uninfected controls, while IP-10 and RANTES were significantly elevated in serum. Standardized differences in CSF between HIV-positive and HIV-negative subjects (effect sizes) were larger, on average, than differences in serum between the two groups (Table 3). Adjusted analysis for CSF/serum albumin quotient (Q_{alb}) there was no significant difference between the groups for all cytokines and chemokines measured ($P > 0.05$).

Collectively, the data indicate that the cellular immune response differs qualitatively between CSF and serum, as more types of cytokines and chemokines were stimulated in CSF than in serum. Also levels of some cytokines and chemokines were also higher in CSF than in serum, clearly indicating intrathecal synthesis (Table 3). Hence, we conclude that the immunological response is stronger in CSF than in serum.

In a subgroup analysis of plasma virologically suppressed individuals ($n = 37$), MIP-1 β was significantly elevated compared to HIV negative subjects ($P = 0.027$).

3.2.2. Comparison between HIV participants with or without CSF pleocytosis

CSF and serum cytokines and chemokines in HIV participants with or without CSF pleocytosis are summarized in Table 4. After adjusting for CD4 nadir and viral load suppression in plasma and CSF, most cytokines, including TNF α , IFN γ , IL1 β , IL-2, IL-6, IL-7, and IL-10, were elevated, along with the chemokines IP10, MIP-1 α , MIP-1 β and RANTES, in the CSF of HIV participants with CSF pleocytosis than in participants without pleocytosis. Additionally, CSF levels of MCP-1 tended to be higher in participants with pleocytosis, although differences were not statistically significant. In contrast, serum cytokines and chemokines were comparable. These findings are consistent with the idea that the immune response to HIV differs between blood and CNS.

Table 3 HIV-positive (n = 68) and HIV-negative (n = 19) levels of cellular chemotaxis and inflammatory biomarkers in cerebrospinal fluid and serum. Significant differences are in **bold** typeface.

	CSF				Serum				CSF/serum			
	HIV (+)	HIV (-)	Cohen's d (95% CI) ^b	P ^a	HIV (+)	HIV (-)	Cohen's d (95% CI) ^b	P ^a	HIV (+)	HIV (-)	Cohen's d (95% CI) ^b	P ^a
Cytokines												
TNFα	0.33 (0.48)	0.04 (0.24)	0.66 (0.3, 1.02)	0.0006	0.94 (0.21)	0.79 (0.18)	0.72 (0.2, 1.23)	0.0071	-0.61 (0.47)	-0.75 (0.26)	0.33 (-0.19, 0.84)	0.2100
IFN-γ	0.27 (0.43)	0.07 (0.29)	0.50 (-0.02, 1.02)	0.0230	0.27 (0.45)	0.56 (0.47)	-0.62 (-1.13, -0.1)	0.0200	0.001 (0.63)	-0.48 (0.53)	0.79 (0.28, 1.31)	0.0029
IL1β	-0.11 (0.03)	-0.1 (0.0)	-0.31 (-0.58, -0.04)	0.0270	-0.09 (0.07)	-0.02 (0.18)	-0.60 (-1.44, 0.23)	0.1500	-0.02 (0.08)	-0.07 (0.18)	0.5 (-0.31, 1.32)	0.2100
IL2	0.02 (0.09)	0 (0.0)	0.27 (0, 0.54)	0.0540	0.06 (0.31)	0.08 (0.19)	-0.07 (-0.58, 0.45)	0.7500	-0.04 (0.33)	-0.08 (0.19)	0.13 (-0.38, 0.65)	0.6100
IL4	0.65 (0)	0.65 (0)	0 (0, 0)	1.0000	0.69 (0.16)	0.77 (0.39)	-0.35 (-1.18, 0.48)	0.3900	0.03 (0.16)	-0.11 (0.39)	0.35 (-0.48, 1.18)	0.3900
IL6	0.16 (0.34)	0.09 (0.27)	0.22 (-0.3, 0.73)	0.4100	0.07 (0.29)	0.08 (0.26)	-0.04 (-0.56, 0.47)	0.8700	0.09 (0.45)	0.01 (0.35)	0.19 (-0.32, 0.71)	0.4600
IL7	0.28 (0.20)	0.17 (0.14)	0.60 (0.09, 1.12)	0.0220	0.64 (0.37)	0.65 (0.28)	-0.04 (-0.55, 0.48)	0.8800	-0.36 (0.41)	-0.49 (0.33)	0.32 (-0.2, 0.84)	0.2200
IL10	0.25 (0.41)	0.04 (0.0)	0.56 (0.29, 0.84)	0.0001	0.31 (0.45)	0.30 (0.40)	0.01 (-0.51, 0.53)	0.9700	-0.06 (0.58)	-0.26 (0.40)	0.37 (-0.15, 0.88)	0.1600
Chemokines												
IP10	3.26 (0.75)	2.98 (0.20)	0.42 (0.12, 0.72)	0.0067	2.98 (0.35)	2.57 (0.2)	1.27 (0.75, 1.78)	< 0.0001	0.29 (0.67)	0.41 (0.20)	-0.21 (-0.52, 0.1)	0.1900
MCP1	3.16 (0.19)	3.05 (0.12)	0.62 (0.1, 1.13)	0.0200	2.73 (0.21)	2.63 (0.22)	0.46 (-0.05, 0.98)	0.0770	0.43 (0.27)	0.42 (0.28)	0.04 (-0.47, 0.56)	0.8800
MIP1α	1.10 (0.14)	1.01 (0.06)	0.66 (0.31, 1.01)	0.0004	0.61 (0.29)	0.64 (0.29)	-0.11 (-0.63, 0.4)	0.6600	0.49 (0.34)	0.37 (0.27)	0.37 (-0.15, 0.88)	0.1600
MIP1β	1.09 (0.28)	0.95 (0.30)	0.49 (-0.03, 1)	0.0650	1.71 (0.27)	1.72 (0.19)	-0.05 (-0.56, 0.47)	0.8500	-0.62 (0.33)	-0.77 (0.24)	0.48 (-0.04, 0.99)	0.0690
RANTES	0.58 (0.62)	0.37 (0.57)	0.34 (-0.18, 0.85)	0.2000	4.48 (0.26)	4.75 (0.2)	-1.09 (-1.6, -0.57)	< 0.0001	-3.9 (0.67)	-4.38 (0.57)	0.73 (0.22, 1.25)	0.0060

Values are log transformed and presented in mean (SD).

^a Unadjusted analysis. Adjusted analysis for CSF/serum albumin quotient (Q_{Alb}); there was no significant difference between the groups for all cytokines and chemokines measured (all P > 0.05); for CSF/serum ratio after adjustment for Q_{Alb}, there was significant difference for CSF/serum ratio of IFN-γ, IL-1β and IP-10 (P = 0.035; 0.027 and 0.0009 respectively).

^b Group differences presented as Cohen's d effect sizes and 95% confidence intervals.

We note that in participants with pleocytosis, the cytokines IFNγ, IL-1β, IL-2, IL-6, and IL-10, as well as the chemokines IP-10, MCP-1, and MIP-1α, were higher in CSF than in serum, indicating intrathecal synthesis (Table 4).

Finally, statistically significant differences in TNFα, IL-10, IP-10 (all P < 0.0001), MIP-1α (P = 0.0069), MIP-1β (P = 0.014), and RANTES (P = 0.00015) persisted after adjusting for Q_{Alb}, CD4 nadir, and viral load suppression in plasma and CSF.

3.3. Origin of CSF biomarkers

In uninfected controls, the coefficients of variation for IFNγ, IL-7, and MCP-1 were much smaller in CSF (0.81, 0.32, and 0.24, respectively) than in serum (1.25, 0.57, and 0.49, respectively), suggesting that these molecules are not derived from blood, but from the brain or the leptomeninges (Kuehne et al., 2013). In contrast, we found that the coefficient of variation for albumin was higher in CSF (0.299) than in serum (0.063), in line with published data (Kuehne et al., 2013). Indeed, a molecule that passes from blood to CSF should have a larger inter-individual variation in CSF than in blood due to variability in CSF flow and transport across barriers (Reiber et al., 2012). Thus, albumin can be used as positive control for this analysis, because CSF albumin is exclusively blood-derived. For all other biomarkers (IL-6, TNFα, IP-10, MIP-1α, MIP-1β, and RANTES) the variability was higher in CSF than in serum. These results, however, are inconclusive, and may indicate either transport from blood or limited regulatory control of release from the brain (Reiber et al., 2012). For IL-1β, IL-2, IL-4, and IL-10, the coefficient of variation is smaller in CSF than in blood, although levels were below the assay detection limits in the uninfected volunteers.

In HIV participants, the levels of the cytokines IL-6 and the chemokines IP-10, MCP-1, and MIP-1α were higher in CSF than in serum, also indicating intrathecal synthesis (Table 3).

3.4. Blood-CSF barrier function

Q_{Alb} was significantly higher in HIV participants than in uninfected controls, and was measured to be 6.4 × 10⁻³ (4.9 × 10⁻³; 9.7 × 10⁻³) in the former and 5 × 10⁻³ (4 × 10⁻³; 6 × 10⁻³) in the latter (Table 2; Fig. 1; P = 0.0023). In HIV participants with and without CSF pleocytosis, Q_{Alb} was determined to be 7.9 × 10⁻³ (6.3 × 10⁻³; 11.6 × 10⁻³) and 6.1 × 10⁻³ (4.5 × 10⁻³; 9.2 × 10⁻³), respectively (P = 0.014). Based on age-dependent reference values, there was BCSFB dysfunction in 30 (44%) HIV-positive individuals, but not in uninfected controls (P = 0.0002). BCSFB dysfunction was observed in 13 (65%) HIV participants with CSF pleocytosis, as well as in 17 (35%) participants with normal white blood cell count in the CSF (P = 0.033). Q_{Alb} and albumin Tourtellotte's formula are summarized in Fig. 1A and B, respectively; the Reibergram plot is shown in Fig. 2.

In a subgroup analysis of plasma virologically suppressed individuals (n = 37), Q_{Alb} and albumin Tourtellotte's formula remained significantly elevated compared to HIV negative subjects (P = 0.001 and 0.002).

In HIV participants, the CSF WBC count was weakly but positively correlated with Q_{Alb} (r_s = 0.27; P = 0.0275), indicating that CSF WBC is not a marker of BCSFB function. However, there was strong correlation between CSF total protein and Q_{Alb} (r_s = 0.8; P < 0.0001).

3.5. Correlation of CSF inflammatory biomarkers with CSF flow rate; CSF or plasma HIV RNA

Q_{Alb}, a measure of CSF flow rate, was moderately and positively correlated with CSF for the majority of cytokines and chemokines studied on the HIV+ participants, r_s and P values ranged from r_s = 0.289 (P = 0.017) to r_s = 0.483 (P < 0.0001), for IL-7 and TNFα respectively. The correlation was not significant for IL-1B, IL-4, and IL-10. Similar results were obtained when biomarkers were tested for correlation with albumin Tourtellotte's formula. These correlations may explain why

Table 4 Comparison of cerebrospinal fluid (CSF) and serum chemotaxis and inflammatory biomarkers in HIV-positive samples with an increased CSF WBC count (*n* = 20), and normal CSF WBC count (*n* = 48). Significant differences are in bold typeface.

	CSF				Serum				CSF/serum				
	WBC > 5		WBC < 5		WBC > 5		WBC < 5		WBC > 5		WBC < 5		P ^a
	Cohen's d (95% CI) ^b	P ^a	Cohen's d (95% CI) ^b	P ^a	Cohen's d (95% CI) ^b	P ^a	Cohen's d (95% CI) ^b	P ^a	Diff (95% CI) ^b				
Cytokines													
TNFα	0.81 (0.56)	0.13 (0.26)	1.65 (1.06, 2.23)	<0.0001	1.03 (0.19)	0.90 (0.21)	0.5 (-0.08, 1.09)	0.09	-0.22 (0.54)	-0.77 (0.33)	1.24 (0.65, 1.82)	<0.0001	
IFN γ	0.52 (0.54)	0.17 (0.33)	0.63 (0.04, 1.21)	0.0360	0.34 (0.57)	0.25 (0.40)	0.35 (-0.23, 0.94)	0.23	0.19 (0.84)	-0.08 (0.51)	0.14 (-0.44, 0.72)	0.6300	
IL1β	-0.11 (0.05)	-0.1 (0.02)	-0.37 (-0.96, 0.21)	0.210	-0.07 (0.14)	-0.1 (0)	0.34 (-0.24, 0.93)	0.25	-0.05 (0.14)	-0.01 (0.02)	-0.48 (-1.06, 0.11)	0.1100	
IL2	0.08 (0.16)	0 (0)	0.86 (0.28, 1.45)	0.0044	0.02 (0.12)	0.08 (0.36)	-0.15 (-0.73, 0.44)	0.62	0.06 (0.22)	-0.08 (0.36)	0.36 (-0.22, 0.95)	0.2200	
IL4	0.65 (0)	0.65 (0)	-0.07 (-0.66, 0.51)	0.8100	0.72 (0.22)	0.67 (0.14)	0.3 (-0.29, 0.88)	0.32	-0.06 (0.22)	-0.02 (0.14)	-0.3 (-0.88, 0.29)	0.3200	
IL6	0.35 (0.41)	0.08 (0.28)	0.75 (0.17, 1.34)	0.0120	0.07 (0.26)	0.06 (0.31)	-0.09 (-0.67, 0.49)	0.76	0.28 (0.45)	0.02 (0.44)	0.61 (0.03, 1.2)	0.0400	
IL7	0.41 (0.17)	0.23 (0.19)	0.72 (0.14, 1.31)	0.0160	0.53 (0.36)	0.69 (0.37)	-0.28 (-0.86, 0.31)	0.35	-0.13 (0.41)	-0.46 (0.38)	0.63 (0.05, 1.22)	0.0340	
IL10	0.62 (0.56)	0.09 (0.18)	1.58 (1, 2.17)	<0.0001	0.48 (0.45)	0.23 (0.43)	0.57 (-0.01, 1.16)	0.06	0.14 (0.75)	-0.14 (0.49)	0.49 (-0.1, 1.07)	0.1000	
Chemokines													
IP10	4.0 (0.95)	2.97 (0.37)	1.53 (0.94, 2.11)	<0.0001	3.16 (0.35)	2.90 (0.32)	0.46 (-0.12, 1.05)	0.12	0.82 (0.89)	0.07 (0.38)	1.33 (0.75, 1.92)	<0.0001	
MCP1	3.24 (0.22)	3.12 (0.16)	0.48 (-0.1, 1.07)	0.1000	2.71 (0.19)	2.74 (0.21)	0.23 (-0.36, 0.81)	0.44	0.53 (0.27)	0.39 (0.26)	0.18 (-0.4, 0.76)	0.5400	
MIP1α	1.21 (0.2)	1.05 (0.07)	1.13 (0.55, 1.72)	0.0003	0.63 (0.35)	0.59 (0.27)	0.38 (-0.21, 0.96)	0.20	0.57 (0.43)	0.46 (0.29)	0.09 (-0.49, 0.68)	0.7600	
MIP1β	1.21 (0.25)	1.04 (0.28)	0.97 (0.38, 1.55)	0.0016	1.64 (0.3)	1.74 (0.26)	0.07 (-0.52, 0.65)	0.82	-0.43 (0.33)	-0.70 (0.31)	0.75 (0.17, 1.34)	0.0130	
RANTES	1.14 (0.79)	0.34 (0.32)	1.43 (0.85, 2.02)	<0.0001	4.51 (0.23)	4.47 (0.27)	0.23 (-0.35, 0.82)	0.43	-3.36 (0.84)	-4.13 (0.41)	1.17 (0.59, 1.76)	0.0002	

Values are log transformed, presented in mean (SD).
^a Adjusted for plasma and HIV viral load suppression, and CD4 nadir. Unadjusted and adjusted analysis adding CSF/serum albumin quotient (Q_{alb}) to plasma and HIV viral load suppression, and CD4 nadir in the multivariable model are not shown.
 After adding Q_{alb} in the adjustment multivariable model, persisted significance for TNFα, IL-10, IP-10, MIP-1α, MIP-1β, and RANTES (*P* < 0.0001; <0.0001; 0.0069; 0.014; 0.00015 respectively) and for the CSF/serum TNFα, IL-7, IP-10, MIP-1β, MIP-1α and RANTES (*P* = 0.002; 0.045; 0.0007; 0.029; 0.0016 respectively) for all the other results *P* > 0.05.
^b Group differences presented as Cohen's d effect sizes and 95% confidence intervals.

incorporation of Q_{alb} into regression models diminishes the significance of a majority of biomarkers. Hence, we conclude that BCSFB dysfunction critically affects and accounts for levels of inflammatory biomarkers in CSF.

CSF and plasma viral load are important as HIV antigen load may drive cytokine responses. Indeed, CSF HIV RNA was moderately and positively correlated with levels of CSF IFNγ (*r*_s = 0.472, *P* < 0.0001); IL-10 (*r*_s = 0.380, *P* = 0.0013); IL-6 (*r*_s = 0.415, *P* = 0.0004); IL-7 (*r*_s = 0.518, *P* < 0.0001); TNFα (*r*_s = 0.618, *P* < 0.0001); IP-10 (*r*_s = 0.562, *P* < 0.0001); MCP-1 (*r*_s = 0.333, *P* = 0.0055); MIP-1α (*r*_s = 0.488, *P* < 0.0001); and RANTES (*r*_s = 0.458, *P* < 0.0001). Plasma HIV RNA was moderately and positively correlated with CSF IFNγ (*r*_s = 0.369, *P* = 0.002); IL-6 (*r*_s = 0.263, *P* = 0.031); IL-7 (*r*_s = 0.464, *P* < 0.0001); TNFα (*r*_s = 0.418, *P* = 0.0004); IP-10 (*r*_s = 0.474, *P* < 0.0001); MIP-1α (*r*_s = 0.397, *P* = 0.0008); RANTES (*r*_s = 0.284, *P* = 0.019). The majority of cases (26; 81%) with detectable HIV CSF viral load also had detectable HIV RNA in blood. To account for this our statistical analysis adjusted for blood viral load.

4. Discussion

In this cross-sectional analysis, unadjusted regression models detected differences in biomarkers in CSF, but not in serum, between HIV-positive and HIV-negative volunteers. This result strongly suggested compartmentalization of the CNS inflammatory response to HIV. In addition, we observed dysfunction of the BCSFB in HIV participants, especially among participants with CSF pleocytosis. The functional integrity of this barrier is generally defined by diffusion and CSF flow rate, the latter being a function of CSF production and release, and is typically assessed by measuring CSF albumin. Dysfunction of the BCSFB is generally due to a decrease in CSF flow rate, rather than structural disruption (Reiber et al., 2012).

In HIV participants, CSF levels of a majority of inflammation biomarkers were positively correlated with CSF/serum albumin ratio. Consequently, incorporation of this ratio into regression models diminished the differences between HIV-positive and HIV-negative CSF, and between HIV participants with or without CSF pleocytosis. These observations indicate that CSF flow rate plays a critical role in CNS inflammation, presumably by facilitating influx of inflammatory molecules, white blood cells, HIV particles, and HIV proteins from peripheral blood, and thereby exacerbating and sustaining cellular immunity and inflammation in the CNS. If the BCSFB dysfunction is cause or consequence of the inflammation can't be answered by this study. Presumably probably both process are involved, this observation is shared by others authors (Saunders et al., 2008). CNS barriers actively regulate brain homeostasis, and specifically respond to events in peripheral tissues and in the brain parenchyma. Indeed, these activities should be considered to fully characterize CNS diseases. Consequently, there has been a resurgence of interest in the possibility that barrier dysfunction may be involved in different neurological disorders (Saunders et al., 2008).

Our data indicate that in HIV participants, cytokine levels were altered more drastically in the CSF than in the serum. Although this phenomenon was observed for both pro-inflammatory (TNFα and IFNγ) and anti-inflammatory (IL-10) cytokines, the effect was stronger in the former. Levels of IL-7, an important inducer of autoimmune reactions that elicit a strong response to IL-17, i.e., Th-17 response were also elevated in CSF. In contrast, only pro-inflammatory cytokines (TNFα and IFNγ) were altered in the serum. Taken together, the data support the notion that the CNS is a primary site of HIV infection, and are consistent with the hypothesis that the CNS reacts immunologically when stimulated appropriately, although the system is considered immunologically privileged (Barker and Billingham, 1977; Kreutzberg, 1996).

Although CNS HIV infection originates from viral and immune components transported into the CNS, the infection eventually becomes

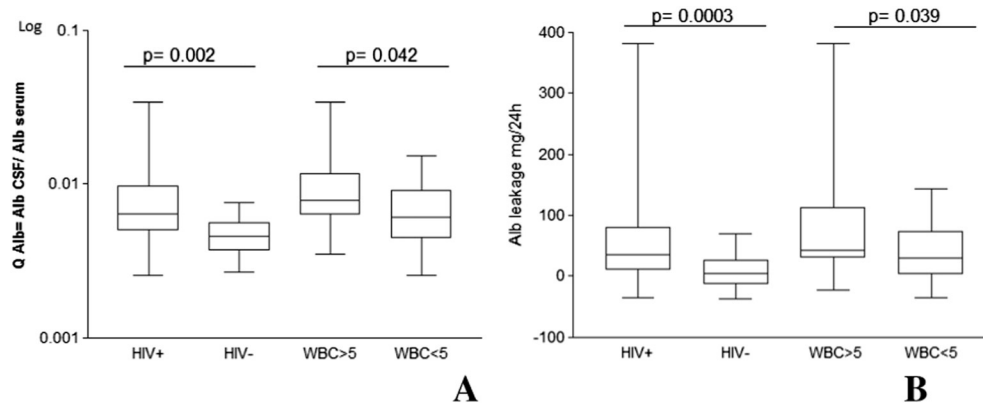


Fig. 1. Albumin quotient, Q_{Alb} (A) and albumin Tourtellotte's formula (B) in HIV positive and uninfected volunteers; and in HIV positive participants with or without CSF pleocytosis (WBC > 5 cell/mm³). Q_{Alb} and albumin Tourtellotte's formula were strongly and positively correlated ($r_s = 0.99$; $P < 0.0001$), in accordance with Sydulko et al. (1993).

“compartmentalized” and acquires characteristics distinct from those observed in the blood. Thus, viral load and levels of immune components may be determined by independent processes in the CSF and blood (Price et al., 2013). Accordingly, a previous study demonstrated that inflammatory reactions in the CSF and serum are comparable during primary HIV infection, but eventually diverge as the infection progresses and as HIV-associated neurocognitive disorders develop. In addition, persistent CSF and blood inflammation despite HIV suppression suggests incomplete control of systemic and CNS infection (Peterson et al., 2014).

We note that all chemokines stimulated by HIV infection are induced via IFN γ , for instance, IP-10 and MCP-1. Similarly, RANTES, was almost

twice as high in the CSF in HIV participants, although this difference was not statistically significant. MIP-1 α is also elevated, although this chemokine is down-regulated by IFN γ in vitro (Sherry et al., 1998). In addition, IP-10 and RANTES levels were elevated in the serum of HIV participants.

The origins of most CSF biomarkers tested in this study are unknown, and tracing such origins may help elucidate the dynamics of these biomarkers. CSF molecules can be derived exclusively from blood, brain parenchyma or from the leptomeninges (Kuehne et al., 2013; Reiber, 2001; Reiber et al., 2012). Our data indicate that IL-6, IL-7, IFN γ , MCP-1, MIP-1 α , and IP-10 are derived from intrathecal synthesis, either because the concentration was higher in CSF than in serum (IL-6, IP-10, MCP-1, and MIP-1 α), or because the coefficient of variation was smaller in CSF than in serum (IFN γ , IL-7, and MCP-1). The latter result suggests that inter-individual variations in CSF biomarkers do not depend on serum concentration. Thus, we believe that CSF/serum quotients for these biomarkers primarily derived from the brain are not informative (Reiber, 2001, 2003). However, for proteins derived primarily from blood, the CSF/serum quotients may indeed be informative. We note that coefficients of variation for molecules derived from blood increase with transport into CSF because of variability in barrier traffic and CSF flow rate (Kuehne et al., 2013).

Further, we note that several proteins we analyzed are of small size, including IFN γ (17 kDa), IL-1 β (17.5 kDa), IL-2 (15 kDa), IL-4 (17 kDa), IL-6 (25 kDa), IL-7 (17 kDa), IL-10 (20 kDa), TNF α (18 kDa), IP-10 (8.7 kDa), MCP-1 (8 kDa), MIP-1 α (20 kDa), MIP-1 β (8–14 kDa), and RANTES (7.8 kDa). Hence, these proteins may also cross the blood-CSF barrier independent of its integrity, as diffusion through an intact barrier may occur along a concentration gradient (Reiber, 2001, 2003; Reiber et al., 2012), although such diffusion may be affected by changes in CSF flow rate. Our data are in line with these observations, as CSF levels of all cytokines and chemokines tested, with the exception of IL-1B, IL-4, and IL-10, were positively correlated with Q_{Alb} .

This paper has several positive points; we believe that the sample size was sufficiently large, and consisted of 87 samples, of which 68 were obtained from HIV participants, and 19 from uninfected controls. Nevertheless, we detected significant medium- and large-effect sizes between infected and uninfected volunteers, and between HIV participants with or without CSF pleocytosis. These effects are expressed in terms of Cohen's d and 95% confidence intervals. Notably, Cohen's effect sizes were mostly large in comparisons between HIV participants with or without pleocytosis. Furthermore, the samples included participants infected with non-B HIV subtypes, in contrast to previously published papers. Although HIV-1 subtypes differences on biomarkers were not the aim of this study, comparisons on the same panel of inflammatory biomarkers between HIV-1 subtype B and C were published on a previous paper, we note that inflammatory biomarkers were found to be

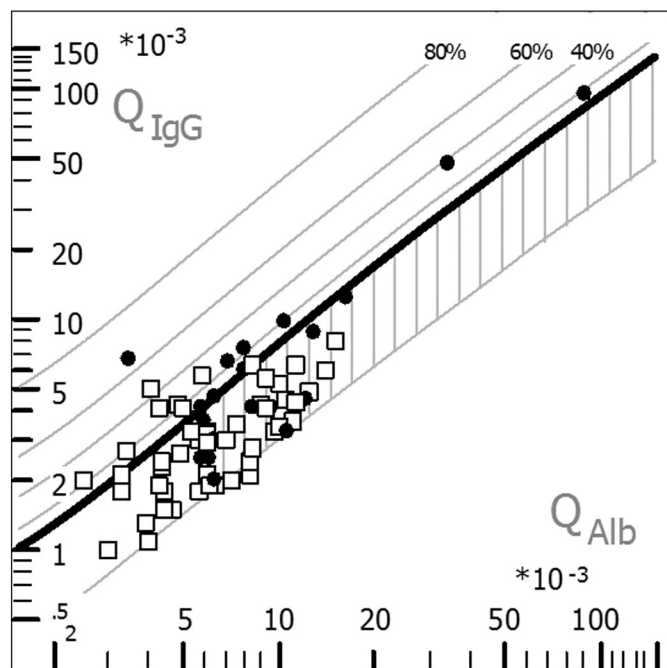


Fig. 2. Hyperbolic function (Reibergram plot) of blood-CSF barrier function and intrathecal IgG synthesis in HIV positive participants. Hyperbolic functions are a consequence of nonlinear interaction between molecular flux and CSF flow rate as derived from the laws of diffusion (Reiber, 1995). Filled circles are participants HIV+ with CSF WBC > 5 cell/mm³, while open squares are participants HIV+ with CSF WBC < 5 cell/mm³. The age-corrected normal range of Q_{Alb} was 7×10^{-3} , as indicated in the graphic. BCSFB dysfunction (area with vertical stripes) was observed in 12 (60%) participants with white blood cell count > 5 cell/mm³, as well as in 20 (41.67%) participants with white blood cell count < 5 cell/mm³ ($\chi^2 P = 0.192$). Intrathecal IgG synthesis was present in nine cases in both groups, at a prevalence of 45% and 18.75% respectively ($\chi^2 P = 0.05$). BCSFB barrier dysfunction was noted in five of these participants, all of whom presented pleocytosis. The plot was generated with CSF Research software.

comparable between participants infected with these subtypes (de Almeida et al., 2016).

Finally, previous papers did not fully integrate multiple results from studies of the BCSFB during HIV infection, and did not reach the conclusion that a dysfunctional barrier helps to maintain and increase the divergence of the immune response in CSF and blood. Nevertheless, this study is not without limitations, and we emphasize that results are cross-sectional, and not longitudinal.

As conclusion, our findings indicate that despite dysfunction of the BCSFB, cellular immune response remains highly compartmentalized. Thus markers of cellular immune responses to HIV in CSF are distinct from those in serum. Furthermore, chemokines and cytokines were frequently elevated in the CSF of HIV participants, in comparison to uninfected controls. CSF chemokines and cytokines correlated with CSF and plasma HIV RNA, suggesting that HIV antigen load may drive cytokine responses. These results support the notion that the CNS behaves as a separate immunologic compartment. This compartmentalized immune response is markedly reduced by virology suppression, although BCSFB dysfunction persists on this subgroup. Finally, we found that dysfunction of the BCSFB critically contributes to the strong CNS inflammatory response to HIV, presumably by allowing transport of WBC, serum molecules, HIV particles, and HIV proteins into the CNS.

Abbreviations

BBB	blood-brain barrier
BCSFB	blood-CSF barrier
CART	combination anti-retroviral therapy
CPE	CNS anti-retroviral penetration effectiveness
IFN	interferon
IL	interleukin
IP-10	interferon gamma-induced protein 10
Q _{Alb}	albumin quotient
MCP	monocyte chemoattractant protein
MIP	macrophage inflammatory protein
RANTES	regulated on activation, normal T cell expressed and secreted
TNF	tumor necrosis factor
WBC	white blood cell

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