

UC Berkeley

UC Berkeley Previously Published Works

Title

Spatially resolved GHz magnetization dynamics of a magnetite nano-particle chain inside a magnetotactic bacterium

Permalink

<https://escholarship.org/uc/item/5qc3z1cr>

Journal

Physical Review Research, 3(3)

ISSN

2643-1564

Authors

Feggeler, Thomas
Meckenstock, Ralf
Spoddig, Detlef
[et al.](#)

Publication Date

2021-07-01

DOI

10.1103/physrevresearch.3.033036

Copyright Information

This work is made available under the terms of a Creative Commons Attribution License, available at <https://creativecommons.org/licenses/by/4.0/>

Peer reviewed

Spatially resolved GHz magnetization dynamics of a magnetite nano-particle chain inside a magnetotactic bacterium

Thomas Feggeler,¹ Ralf Meckenstock,¹ Detlef Spoddig,¹ Benjamin W. Zingsem,^{1,2} Hendrik Ohldag,^{3,4} Heiko Wende,¹ Michael Farle,^{1,5} Michael Winklhofer,⁶ and Katharina J. Ollefs¹

¹*Faculty of Physics and Center for Nanointegration Duisburg-Essen (CENIDE),
University of Duisburg-Essen, 47048 Duisburg, Germany**

²*Ernst Ruska Centre for Microscopy and Spectroscopy with Electrons and Peter Grünberg Institute,
Forschungszentrum Jülich GmbH, 52425 Jülich, Germany*

³*SLAC National Accelerator Laboratory, 94025 Menlo Park, CA, United States*

⁴*Department of Physics, University of California Santa Cruz, Santa Cruz CA 95064, United States[†]*

⁵*Kirensky Institute of Physics, Federal Research Center KSC SB RAS, Russia*

⁶*School of Mathematics and Science, University of Oldenburg, 26129 Oldenburg, Germany*

Understanding magnonic properties of non-periodic magnetic nanostructures requires real-space imaging of ferromagnetic resonance modes with spatial resolution well below the optical diffraction limit and sampling rates in the 5-100 GHz range. Here, we demonstrate element-specific Scanning Transmission X-ray Microscopy detected Ferromagnetic Resonance (STXM-FMR) applied to a chain of dipolarly coupled Fe_3O_4 nano-particles (40-50 nm particle size) inside a single cell of a magnetotactic bacterium, *Magnetospirillum magnetotacticum*. The ferromagnetic resonance mode of the nanoparticle chain driven at 6.748 GHz and probed with 50 nm X-ray focus size was found to have a uniform phase response but non-uniform amplitude response along the chain segments due to the superposition of dipolar coupled modes of chain segments and individual particles, in agreement with micromagnetic simulations.

I. INTRODUCTION

Magnonics deals with the controlled excitation and detection of spin waves in magnetic media [1–7]. Due to the low energy nature of spin wave quanta (magnons), magnonic devices have been suggested to overcome key challenges of current charge-based computer information processing, such as power consumption and heat dissipation [8]. Recently, magnetic nanoparticle chains were proposed as magnonic logic gates on the sub-micron scale, which would enable unprecedented transistor counts in a single central processing unit [9]. For visual control of the functionality of submicron-sized magnon-based logic devices, it is essential to have a technique that can probe the magnetization dynamics on the 1-100 GHz scale with few nm resolution to determine the local phase and amplitude of excited magnons. Real-time observations using scanning electron microscopy with polarization analysis [10] have reached a temporal resolution (1 ns) sufficient to track relatively slow (10^2 MHz) dynamic magnetization processes involving vortex cores in soft magnetic materials. Faster dynamical processes can be studied with pump-probe schemes repeated for a sequence of delay times with each repetition starting from the same initial state and accumulating many pump-probe pulse cycles

in a stroboscopic procedure. Pump-probe schemes are used in time-resolved photoemission electron microscopy (PEEM) [11, 12] and ultrafast transmission electron microscopes [13, 14], capable of probing transient demagnetization dynamics of nanodiscs with sub-100 nm resolution and sub-picoseconds delay time resolution [15]. Here we use Scanning Transmission X-ray Microscopy detected Ferromagnetic Resonance (STXM-FMR), where the magnetic specimen is driven by a continuous wave excitation, while temporal sampling is performed stroboscopically at a higher harmonic [16, 17]. The technique offers element specificity, ≤ 50 nm spatial resolution, and sampling rates of tens of GHz, allowing for spatial detection of resonant responses at up to 10 GHz [16]. By exploiting these features of STXM-FMR, we were able to detect a spatially non-uniform ferromagnetic resonance mode across a single chain of magnetite (Fe_3O_4) nanoparticles in a magnetotactic bacterium. Thus, we demonstrate the feasibility to detect ferromagnetic resonance in a nanomagnetic system with ≤ 50 nm spatial resolution.

II. EXPERIMENTAL DETAILS

With the STXM-FMR setup at the Stanford Synchrotron Radiation Lightsource (SSRL), FMR excitations in the linear regime are measured element specifically by employing the X-ray Magnetic Circular Dichroism (XMCD) effect [19, 20] in the transverse XMCD geometry [18]. The XMCD effect, defined as difference in X-ray absorption of left and right hand circularly polarized X-rays, is a measure of the magnetization M of the probed material projected onto the k -vector of the inci-

* Present address: Advanced Light Source, Lawrence Berkeley National Laboratory, Berkeley CA 94720, United States; tfeeggeler@lbl.gov

† Present address: Advanced Light Source, Lawrence Berkeley National Laboratory, Berkeley CA 94720, United States and Department of Material Sciences and Engineering, Stanford University, Stanford CA 94305, United States

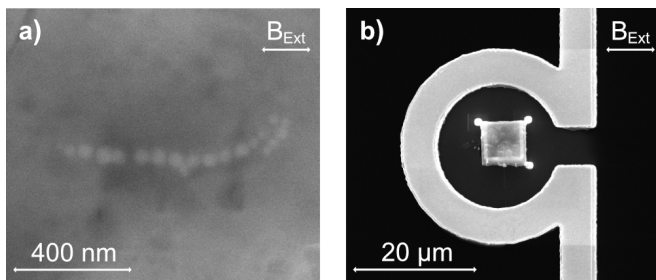


FIG. 1. a) Electron micrograph of the Fe_3O_4 nano-particle chain selected for STXM-FMR. The structure was deposited on a TEM grid and cut out with FIB. b) Cutout area including the nano-particle chain positioned in the center of the micro-resonator (light grey "omega" shape Au conduit). The direction of the magnetic field B_{Ext} is indicated.

dent X-rays. The resonant magnetic excitation is realized by applying linearly polarized continuous microwaves b_{mw} to the sample, polarized parallel to the propagation direction of the X-rays, while the static magnetic field B_{Ext} is orientated perpendicular to the k -vector of the X-rays. For the homogeneous excitation with high conventional detection sensitivity a micro-resonator setup is used [21].

The sample contains a chain of 19 Fe_3O_4 [23–26] nanoparticles with particle sizes of 40 - 50 nm [26] arranged in a segment of 15 particles and a smaller segment of 4 particles offset from this segment by a distance of about 25 nm (Figure 1 a). The nanoparticle chain grew naturally by biomineralization inside a bacterial cell of *Magnetospirillum magnetotacticum* [26–28]. The bacteria were obtained from Leibniz Institute DSMZ-German Collection of Microorganisms and Cell Cultures [22] as actively growing culture and positioned on a Transmission Electron Microscopy (TEM) grid (Figure 1 a). The grid with the nanoparticles was cutout by Focused Ion Beam (FIB) milling and placed at the center of the micro-resonator loop (Figure 1 b). The external magnetic field $B_{\text{Ext}} = 0.167 \pm 0.002$ T (Figure 1) corresponding to the resonance field predicted by our micromagnetic simulations, was applied in the sample plane along the direction of the chain axis as indicated in Figure 1. The sample was probed under dry conditions at room temperature at the Fe L_3 -edge (708 eV [29]) with a focus of ≤ 50 nm, a step size of 25 nm and a dwell time of five seconds per pixel with constant polarization of the X-rays. The microwave excitation at 6.748 GHz with a power of 25 dBm was chosen as the $(14+1/6)^{\text{th}}$ harmonics of the synchrotron frequency.

In the STXM-FMR measurements two sets of 6 consecutive images each are recorded pixel-by-pixel, which correspond to 6 images recorded with microwaves on and 6 images with microwave off, respectively. Between two consecutive images there is a time difference of 24.7 ps and a phase difference of ($\Delta\varphi = 60^\circ$), yielding to the monitoring of 6 time steps of the microwave cycle [16]. To extract the microwave induced X-ray transmission the

natural logarithm of the microwave off data divided by the microwave on data was taken. The data was normalized to the average intensity with consecutive minimum-maximum normalization. The phase and amplitude response at each pixel was obtained by fitting a sine function to the time dependent data [37]. The result is coded in the Hue-Saturation-Brightness (HSB) format to represent the phase relative to the time $t = 0$ ps as hue [30], the amplitude as brightness and the fit accuracy as saturation [37].

From the scanning electron micrographs (Figure 1 a), we designed a three-dimensional computer model of the nano-particle chain (single particle size 50 nm) using the micromagnetic simulation package MuMax3 [31, 32]. Taking into account the not perfectly stoichiometric Fe_3O_4 with the known decrease of the Verwey transition temperature [33], the saturation magnetisation M_{Sat} and the magnetocrystalline anisotropy constant K_1 [34], the dynamic simulation was calculated according to [34] with $M_{\text{sat}} = 465$ kA/m, and a first order cubic anisotropy constant $K_1 = -1.0 \times 10^4$ J/m³. The exchange stiffness was set to $A = 1.32 \times 10^{-11}$ J/m [35] and the simulation grid was defined as $200 \times 80 \times 12$ cells with a cell size of $(5 \text{ nm})^3$.

III. RESULTS AND DISCUSSION

In the FMR absorption spectrum simulated at 6.748 GHz (Figure 2 a), we observe three resonances at 0.169 T, 0.176 T and 0.187 T, which is surprising given that a linear chain usually behaves as a uniaxial magnet with one resonance mode per field orientation [36]. Evidently variations in the orientation and strength of the dipolar coupling between the particles in a chain produces complex magnon dispersion relations with multiple resonance lines and even magnonic band gaps, where resonances involving the same group of particles have incommensurate spatial mode profiles [9]. The variations in dipolar coupling are most pronounced in chain geometries with irregular particle spacings [9]. Indeed, for the chain at hand, the micromagnetic simulation confirms pronounced variations in dipolar coupling strength (Figure 2 b) along the chain axis, ranging from 0.450 T between adjacent particles (orange/red color) down to 0.050 T in the gap between the two chain segments. This essentially dynamically decouples the small four-particle segment on the left from the main chain segment. The decoupling becomes apparent in the snapshots of the dynamic micromagnetic simulation (Figure 2 c), where the oscillation amplitude is large in the main segment and small in the four-particle segment. Likewise, at the far right end of this chain, the oscillation amplitudes are comparatively small (Figure 2 c), which is due to the disturbance of the simple linear chain structure by two pairs of particles in side-by-side position perpendicular to the chain axis. In both side-by-side pairs, it is the lower particle that is more strongly coupled to the curved chain

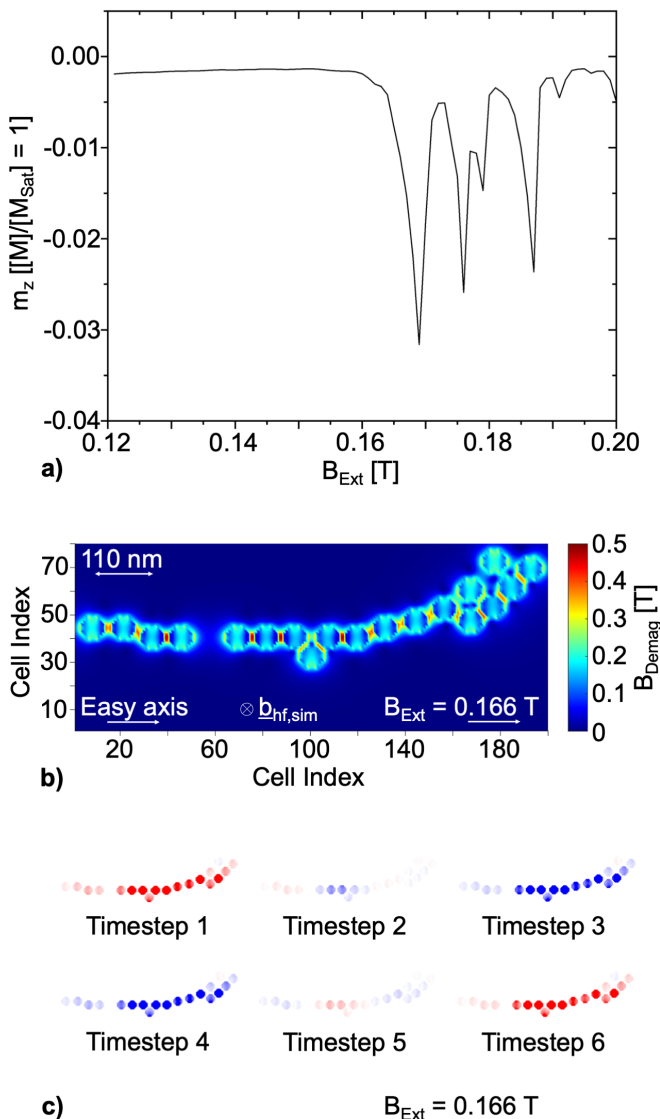


FIG. 2. a) Simulated FMR absorption spectrum in terms of the normalized m_z component of the magnetization over the field range $B_{\text{Ext}} = 0.12 \text{ T} - 0.2 \text{ T}$. b) Spatial distribution of the simulated inner demagnetization and outer stray-field at an applied static magnetic bias field of 0.166 T . c) Six timesteps ($\Delta t \sim 24.7 \text{ ps}$) of the dynamic micromagnetic simulation, displaying a spatially inhomogeneous oscillation response. The normalized m_z -out-of-plane-component of the magnetization is color-coded ($-z$: blue, $+z$: red).

axis (compare with Figure 2 b), so that it can be excited more easily in the main resonance mode of the chain. Thus, on the basis of our micromagnetic simulations, we expect a non-homogeneous microwave response along the particle chain.

Indeed, the experimentally determined resonance amplitudes (Figure 3 a, c) are strongest (high brightness purple, white) in the main segment of the chain, but almost vanish in the left four-particle segment extending

from $x = 14$ to $x = 22 \text{ px}$ at $y = 10 \text{ px}$). Although individual particles cannot be resolved due to the limited focus (50 nm) at the Fe L_3 , the upcurving bright white intensity pattern ($x = 25$ to $x = 48 \text{ px}$ and $y = 6$ to $y = 12 \text{ px}$) tracks the curvature of the main segment of the chain. We note that in Figure 3a) and c) we find a non-zero phase and amplitude not only at the location of the chain but almost everywhere in the field of view. The reason for this observation is that there is a small crosstalk between the applied microwave and the (shielded) avalanche photodiode. Since the setup described in [16] represents a high quality factor lock-in amplifier tuned to the exciting microwave frequency, even the smallest fraction of microwave power at the diode will be detected. In consequence also without a sample a non-zero signal is observed. To remove the visibility of this background we discriminate the detected channel with a value of 250000 counts (see supporting information), revealing a clearly distinguishable resonant response of the particle chain with the highest amplitude along the upcurving chain segment. The phase map (Figure 3 a, b) reveals a largely uniform resonant response of the particle chain (270° to 300°) and is consistent with the time snapshots of the micromagnetic simulation (Figure 2 c). For comparison between experiment and theory, we evaluated the snapshot sequences from the micromagnetic simulations in the way as the experimental image sequences to obtain phase and amplitude maps (Figure 3 e and 3 f). All simulation results are pictured at $B_{\text{Ext}} 166 \text{ mT}$, at which the best match to the experimental data is observed. Deviations between calculated and observed maps can be ascribed to the fact that not all small features of the 3D particles are reflected in the computer model, resulting in equally sized particles in the micromagnetic model, while the particle sizes and spacing in the real chain have some variability, particularly at the right end of the chain. This leads to variations in dipolar coupling strength (comp. Figure 2 b) and in consequence in the observed resonance modes.

The calculated phase response tallies with the observed one, except for the particle at the far upper right, which oscillates antiphase to the rest of the chain (Figure 3 e), albeit with very low amplitude (Figure 3 f). As can be seen in the amplitude map, the response of the left four-particle segment indeed is weak, which explains the lack of signal in this region in the measured amplitude map. As inferred from the stray field map above (Figure 2 b), the particles that are vertically offset from the central chain axis and more weakly coupled to the chain, are weakly excited and thus have low oscillation amplitudes (Figure 3 f).

In summary, we observe a uniform phase but non-uniform amplitude response of the nano-particle chain. With the STXM-FMR technique, we thus demonstrate for the first time a measurement of a ferromagnetic resonance mode of a nano-particle chain in a spatially resolved manner ($\leq 50 \text{ nm}$). This technique allows to element-specifically examine the functionality of both periodic and non-periodic magnetic nanostructures of fu-

Experiment

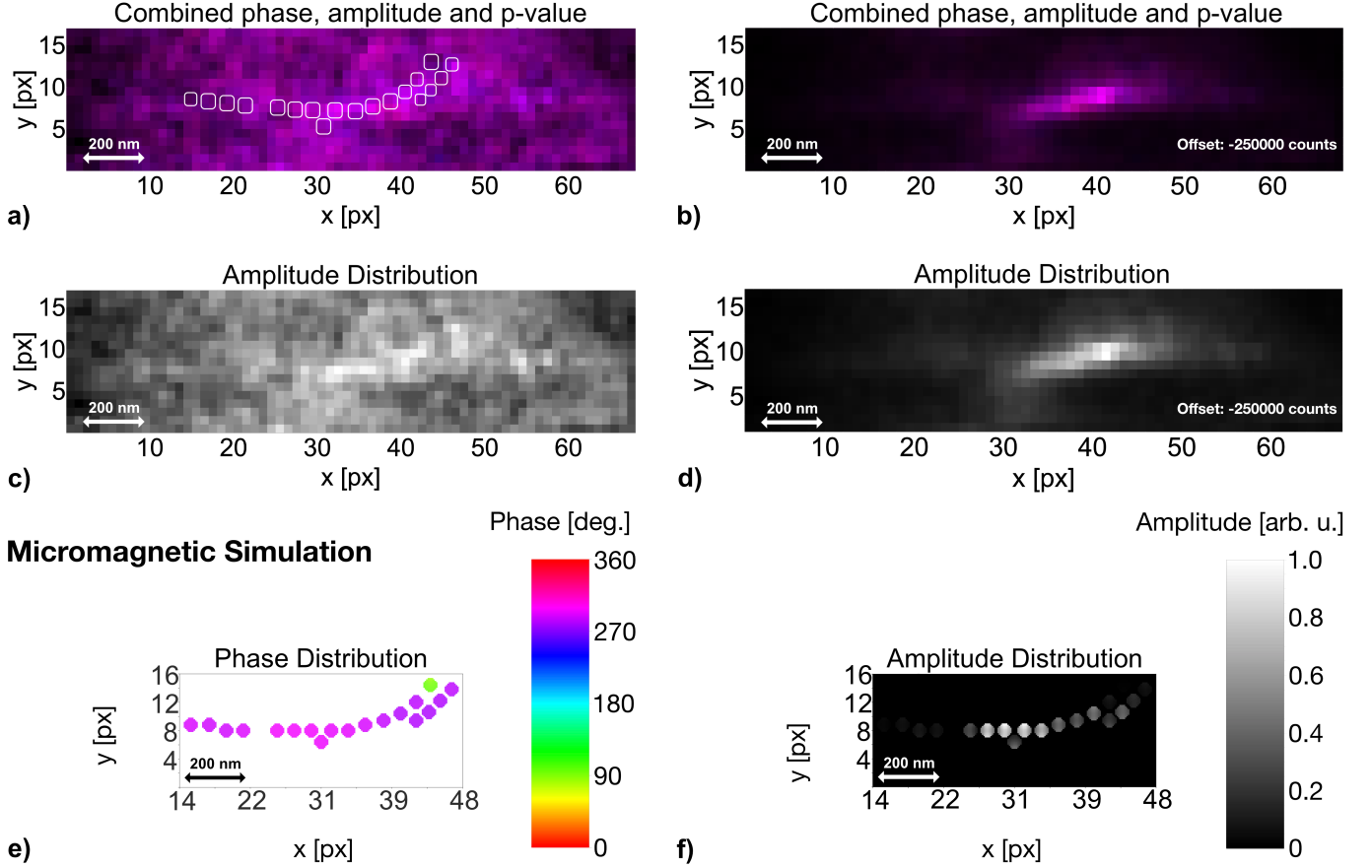


FIG. 3. a) Result of the pixel-by-pixel sinusoidal fit analysis for the STXM-FMR data at B_{Ext} 167 mT at $f = 6.748$ GHz. The phase relative to $t = 0$ ps, the amplitude and the fit accuracy are coded as hue, brightness and saturation. The estimated particle positions are indicated by rounded squares. b) Subfigure a) after subtraction of an offset of 250000 counts. c), d) Normalised amplitude distribution extracted from a), b) respectively. e), f) Phase and amplitude distribution of the micromagnetic simulation at B_{Ext} 166 mT at $f = 6.748$ GHz.

ture magnonic and spintronic devices, like for magnon spin transistors [39] and biomagnonics [9, 40], complementing spatially-resolved, non-element-specific, experimental techniques and micromagnetic simulations. A new STXM-FMR setup taking advantage of the increased spatial resolution of 10 nm [41] at the Advanced Light Source (ALS) will open the pathway to image X-band GHz magnetization dynamics of nanostructures on the ≤ 20 nm scale.

IV. ACKNOWLEDGEMENT

The authors thank the German Research Foundation (DFG project: OL513/1-1, 321560838) and the Austrian Science Fund (FWF project: I 3050-N36) for financial support. We gratefully acknowledge A. Ney for fruitful discussions. The use of the Stanford Synchrotron Radiation Lightsource, SLAC National Accelerator Laboratory, is supported by the U.S. Department of Energy, Office of Science, Office of Basic Energy Sciences under Contract No. DE-AC02-76SF00515.

V. SUPPORTING INFORMATION AVAILABLE:

Experimental information on the time resolved setup and considerations on the oscillating background.

- [1] Chumak, A. V.; Serga, A. A. and Hillebrands, B., Magnonic crystals for data processing. *J. Phys. D: Appl. Phys.* **2017**, 50, 244001.
- [2] Chumak, A. V.; Vasyuchka, V. I. and Hillebrands, B., Magnon spintronics. *Nat. Phys.* **2015**, 11, 453461.
- [3] Urazhdin, S.; Demidov, V. E.; Ulrichs, H.; Kendziorczyk, T.; Kuhn, T.; Leuthold, H.; Wilde, G. and Demokritov, S. O., Nanomagnonic devices based on the spin-transfer torque. *Nat. Nanotech.* **2014**, 9, 509513.
- [4] Vogt, K.; Fradin, F. Y.; Pearson, J. E.; Bader, S. D.; Hillebrands, B.; Hoffmann, A.; and Schultheiss, H., Realization of a spin-wave multiplexer. *Nat. Commun.* **2014**, 5, 3727.
- [5] Sato, N.; Sekiguchi, K. and Nozaki, Y., Electrical demonstration of spin-wave logic operation. *Appl. Phys. Expr.* **2013**, 6, 063001.
- [6] Kruglyak, V.V.; Demokritov, S.O., and Grundler, D., Magnonics. *J. Phys. D: Appl. Phys.* **2010**, 43, 264001.
- [7] Khitun, A.; Bao, M. and Wang, K. L., Magnonic logic circuits. *J. Phys. D: Appl. Phys.* **2010**, 43, 264005.
- [8] Denning, P. J. and Lewis, T. G., Exponential laws of computing growth. *Commun. ACM* **2016**, 60, 54-65.
- [9] Zingsem, B. W.; Feggeler, T.; Terwey, A.; Ghaisari, S.; Spoddig, D.; Faivre, D.; Meckenstock, R.; Farle, M.; Winklhofer, M., Biologically encoded magnonics. *Nat. Commun.* **2019**, 85, 4345.
- [10] Frömter, R.; Kloodt, F.; Röler, S.; Frauen, A.; Staeck, P.; Cavicchia, D. R.; Bocklage, L.; Röbbisch, V.; Quandt, E. and Oepen, H. P., Time-resolved scanning electron microscopy with polarization analysis. *Appl. Phys. Lett.* **2016**, 108, 142401.
- [11] Krasnyuk, A.; Oelsner, A.; Nepijko, S. A.; Kuksov, A.; Schneider, C. M. and Schönhense, G., Time-resolved photoemission electron microscopy of magnetic field and magnetization changes. *Appl. Phys. A-Mater.* **2003**, 76, 863-868.
- [12] Vogel, J.; Kuch, W.; Bonfim, M.; Camarero, J.; Penne, Y.; Offi, F.; Fukumoto, K.; Kirschner, J.; Fontaine, A. and Pizzini, S., Time-resolved magnetic domain imaging by x-ray photoemission electron microscopy. *Appl. Phys. Lett.* **2003**, 82, 2299.
- [13] Park, H. S.; Spencer Baskin, J.; Zewail, A. H., 4D Lorentz Electron Microscopy Imaging: Magnetic Domain Wall Nucleation, Reversal, and Wave Velocity. *Nano Lett.* **2010**, 10, 3796-3803.
- [14] Schliep, K. B.; Quarterman, P.; Wang, J.-P. and Flannigan, D. J., Picosecond Fresnel transmission electron microscopy. *Appl. Phys. Lett.* **2017**, 110, 222404.
- [15] Rubiano da Silva, N.; Möller, M.; Feist, A.; Ulrichs, H.; Ropers, C.; and Schäfer, S., Nanoscale Mapping of Ultrafast Magnetization Dynamics with Femtosecond Lorentz Microscopy. *Phys. Rev. X* **2018**, 8, 031052.
- [16] Bonetti, S.; Kukreja, R.; Chen, Z.; Spoddig, D.; Ollefs, K.; Schöppner, Ch.; Meckenstock, R.; Ney, A.; Pinto, J.; Houanche, Frisch, J.; Stöhr, J.; Dürr, H. A.; Ohldag, H., Microwave soft x-ray microscopy for nanoscale magnetization dynamics in the 510 GHz frequency range. *Rev. Sci. Instr.* **2015**, 86, 093703.
- [17] Schaffers, T.; Meckenstock, R.; Spoddig, D.; Feggeler, T.; Ollefs, K.; Schöppner, Ch.; Bonetti, S.; Ohldag, H.; Farle, M.; and Ney, A., The combination of microresonators with spatially resolved ferromagnetic resonance. *Rev. Sci. Instrum.* **2017**, 88, 093703.
- [18] Goulon, J.; Rogalev, A.; Wilhelm, F.; Jaouen, N.; Goulon-Ginet, C.; Brouder, C., X-ray detected ferromagnetic resonance in thin films. *Eur. Phys. J. B* **2006**, 53, 169-184.
- [19] Dürr, H. A.; Eimüller, T.; Elmers, H.-J.; Eisebitt, S.; Farle, M.; Kuch, W.; Matthes, F.; Mertins, M.; Mertins, H.-C.; Oppeneer, P. M.; Plucinski, L.; Schneider, C. M.; Wende H.; Wurth, W.; Zabel H., A Closer Look Into Magnetism: Opportunities With Synchrotron Radiation. *IEEE Trans. Magn.* **2009**, 45, 15.
- [20] Van der Laan, G.; Figueroa, A. I., X-ray magnetic circular dichroismA versatile tool to study magnetism. *Coordin. Chem. Rev.* **2014**, 95, 277-278.
- [21] Narkowicz, R.; Suter, D.; and Stonies, R., Planar microresonators for EPR experiments. *J. Magn. Reson.* **2005**, 175, 275-284.
- [22] DSMZ, *Magnetospirillum magnetotacticum DSM 3856 2019*.
- [23] Zhu, X.; Hitchcock, A. P.; Bazylinski, D. A.; Denes, P.; Joseph, J.; Lins, U.; Marchesini, S.; Shiu, H. W.; Tylizszczak, T.; and Shapiro, D. A., Measuring spectroscopy and magnetism of extracted and intracellular magnetosomes using soft X-ray ptychography. *Proc. Natl. Acad. Sci. U. S. A.* **2016**, 113, E8219-E8227.
- [24] Posfai, M.; Lefvre, C. T.; Trubitsyn, D.; Bazylinski, D. A.; and Frankel, R. B., Phylogenetic significance of composition and crystal morphology of magnetosome minerals. *Front. Microbiol.* **2013**, 4, 1-15.
- [25] Frankel, R. B.; Blakemore, R. and Wolfe, R. S., Magnetite in Freshwater Magnetotactic Bacteria. *Science* **1979**, 203, 1355-1356.
- [26] Maratea, D. and Blakemore, R., *Aquaspirillum magnetotacticum* sp. nov., a Magnetic Spirillum. *Int. J. Syst. Bacteriol.* **1981**, 31, 452-455.
- [27] Faivre, D. and Schüler, D., Magnetotactic Bacteria and Magnetosomes. *Chem. Rev.* **2008**, 108, 4875-4898.
- [28] Schleifer, K. H.; Schüler, D.; Spring, S.; Weizenegger, M.; Amann, R.; Ludwig, W.; and Köhler, M., The genus *Magnetospirillum* gen. nov., description of *Magnetospirillum gryphiswaldense* sp. nov. and transfer of *Aquaspirillum magnetotacticum* to *Magnetospirillum magnetotacticum* comb. nov. *Syst. Appl. Microbiol.* **1991**, 14, 379-385.
- [29] Thompson, A. C.; Kirz, J.; Attwood, D. T.; Gullikson, E. M.; Howells, M. R.; Kortright, J. B.; Liu, Y.; Robinson, A. L.; Underwood, J. H.; Kim, K.-J.; Lindau, I.; Pianetta, P.; Winick, H.; Williams, G. P.; Scofield, J. H., *X-Ray Data Booklet*. 3rd ed., Lawrence Berkeley National Laboratory University of California: Berkeley CA, **2009**.
- [30] Fairchild, M.D., *Color Appearance Models*. 3rd ed., John Wiley & Sons: Chichester, 2013.
- [31] Vansteenkiste, A.; Leliaert, J.; Dvornik, M.; Helsen, M.; Garcia-Sanchez, F. and Van Waeyenberge, B., The design and verification of MuMax3. *AIP Adv.* **2014**, 4, 107133.
- [32] Exl, L.; Bance, S.; Reichel, F.; Schreffl, T.; Stimming, H. P.; and Mauser, N. J., LaBonte's method revisited: An effective steepest descent method for micromagnetic energy minimization. *J. Appl. Phys.* **2014**, 115, 17D118.

- [33] Aragón, R., Buttrey, D. J., Shepherd, J. P., Honig, J. M., Influence of nonstoichiometry on the Verwey transition, *Phys. Rev. B* **1985**, 31, 430-436.
- [34] Kąkol, Z., Honig, J. M., Influence of deviations from ideal stoichiometry on the anisotropy parameters of magnetite $Fe_{3(1-\delta)}O_4$, *Phys. Rev. B* **1989**, 40, 9090-9097.
- [35] Wu, H.-C., Arora, S. K., Mryasov, O. N., Shvets, I. V., Antiferromagnetic interlayer exchange coupling between Fe_3O_4 layers across a nonmagnetic MgO dielectric layer, *Appl. Phys. Lett.* **2008**, 92, 182502.
- [36] Charilaou, M.; Winklhofer, M. and Gehring, A. U., Simulation of ferromagnetic resonance spectra of linear chains of magnetite nanocrystals. *J. Appl. Phys.* **2011**, 109, 093903.
- [37] Zingsem, B.; Feggeler, T.; Meckenstock, R.; Schaffers, T.; Pile, S.; Ohldag, H.; Farle, M.; Wende, H.; Ney, A.; Ollefs, K., *arXiv:1901.10595 [cond-mat.str-el]*, **2019**.
- [38] Schaffers, T.; Feggeler, T.; Pile, S.; Meckenstock, R.; Buchner, M.; Spoddig, D.; Ney, V.; Farle, M.; Wende, H.; Wintz, S.; Weigand, M.; Ohldag, H.; Ollefs, K.; Ney, A., Evaluation protocol for revealing magnonic contrast in STXM-FMR measurements. *Nanomaterials (Basel)* **2019**, 9, 1-13.
- [39] Chumak, A. V.; Serga, A. a. and Hillebrands, B., Magnon transistor for all-magnon data processing. *Nat. Commun.* **2014**, 5, 4700.
- [40] Barman, A.; Gubbiotti, G.; Ladak, S.; Adeyeye, A. O.; Krawczyk, M.; Gräfe, J.; Adelman, C.; Cofofana, S.; Naeemi, A.; Vasyuchka, V. I.; Hillebrands, B.; Nikitov, S. A.; Yu, H.; Grundler, D.; Sadovnikov, A.; Grachev, A. A.; Sheshukova, S. E.; Duquesne, J.-Y.; Marangolo, M.; Gyorgy, C.; Porod, W.; Demidov, V. E.; Urazhdin, S.; Demokritov, S.; Albisetti, E.; Petti, D.; Bertacco, R.; Schultheiss, H.; Kruglyak, V. V.; Poimanov, V. D.; Sahoo, A. K.; Sinha, J.; Yang, H.; Muenzenberg, M.; Moriyama, T.; Mizukami, S.; Landeros, P.; Gallardo, R. A.; Carlotti, G.; Kim, J.-V.; Stamps, R. L.; Camley, R. E.; Bivas, R.; Otani, Y.; Yu, W.; Yu, T.; Bauer, G. E. W.; Back, C. H.; Goetz, S. U.; Dobrovolskiy, O. V.; van Dijken, S.; Budinska, B.; Qin, H.; Chumak, A.; Khitun, A.; Nikonov, D. E.; Young, I. A.; Zingsem, B. and Winklhofer, M., The 2021 Magnonics Roadmap. *J. Phys.: Condens. Matter* **2021**, in press, DOI: 10.1088/1361-648X/abec1a.
- [41] Chao, W., Fischer, P.; Tyliczszak, T.; Rekawa, S.; Anderson, E.; and Naulleau, P., Real space soft x-ray imaging at 10 nm spatial resolution. *Opt. Express* **2012**, 20, 9777-9783.