Title

Towards multiomic analysis of oral mucosal pathologies

Permalink

https://escholarship.org/uc/item/5s62174w

Journal

Seminars in Immunopathology, 45(1)

ISSN

1863-2297

Authors

Einhaus, Jakob
Han, Xiaoyuan
Feyaerts, Dorien
et al.

Publication Date

2023

DOI

10.1007/s00281-022-00982-0

Peer reviewed

REVIEW

Towards multiomic analysis of oral mucosal pathologies

Jakob Einhaus[1,2] · Xiaoyuan Han[3] · Dorien Feyaerts[1] · John Sunwoo[4] · Brice Gaudilliere[1] ·
Somayeh H. Ahmad[5] · Nima Aghaeepour[1] · Karl Bruckman[5] · David Ojcius[3] · Christian M. Schürch[2] ·
Dyani K. Gaudilliere[5]

Abstract

Oral mucosal pathologies comprise an array of diseases with worldwide prevalence and medical relevance. Affecting a confined space with crucial physiological and social functions, oral pathologies can be mutilating and drastically reduce quality of life. Despite their relevance, treatment for these diseases is often far from curative and remains vastly understudied. While multiple factors are involved in the pathogenesis of oral mucosal pathologies, the host's immune system plays a major role in the development, maintenance, and resolution of these diseases. Consequently, a precise understanding of immunological mechanisms implicated in oral mucosal pathologies is critical (1) to identify accurate, mechanistic biomarkers of clinical outcomes; (2) to develop targeted immunotherapeutic strategies; and (3) to individualize prevention and treatment approaches. Here, we review key elements of the immune system's role in oral mucosal pathologies that hold promise to overcome limitations in current diagnostic and therapeutic approaches. We emphasize recent and ongoing multiomic and single-cell approaches that enable an integrative view of these pathophysiological processes and thereby provide unifying and clinically relevant biological signatures.

Keywords Multiomics · Oral pathology · Immunology · Cytomics · Transcriptomics · Proteomics · Metabolomics · Microbiome · Mass cytometry

This article is a contribution to the special issue on: Single-cell and spatial multi-omics in clinical outcomes studies - Guest Editor: Brice Gaudillière

✉ Dyani K. Gaudilliere
dyani00@stanford.edu

1 Department of Anesthesiology, Perioperative & Pain Medicine, School of Medicine, Stanford University, Stanford, CA, USA

2 Department of Pathology and Neuropathology, University Hospital and Comprehensive Cancer Center Tübingen, Tübingen, Germany

3 Arthur A. Dugoni School of Dentistry, University of the Pacific, San Francisco, CA, USA

4 Division of Head and Neck Surgery, Department of Otolaryngology, School of Medicine, Stanford University, Stanford, CA, USA

5 Division of Plastic and Reconstructive Surgery, Department of Surgery, School of Medicine, Stanford University, 770 Welch Road, Palo Alto, CA 94304, USA

Introduction

Oral mucosal pathology is a large but understudied field that has important implications for the health and quality of life of billions of people worldwide. The most common pathologies affecting the oral mucosa fall under three categories, including (1) benign, precancerous, and malignant neoplasms (e.g., fibromas, leukoplakias, and squamous cell carcinoma), (2) bacterial, fungal, and viral infectious diseases (e.g., periodontitis, candidiasis, and herpes simplex virus), and (3) autoimmune disorders (e.g., oral lichen planus, recurrent aphthous stomatitis, pemphigus vulgaris, and mucous membrane pemphigoid). Oral pathologies affect a carefully designed barrier of the human body between a strongly bacterially colonized environment of the oral cavity and the bordering, highly vascularized and multifunctional mucosa. The mucosal integrity is crucial for central human functions such as food intake, taste, speech, breathing, and esthetics, making disability in this area truly crippling to quality of life. The localization and functionality of the mucosa as a barrier at the interface of contrasting

environments imply a strong presence and involvement of the host's immune system in the pathophysiology of most oral mucosal pathologies. An integrated examination of the immune system's complex role in health and pathology of the oral mucosa is essential to advance the management of oral diseases.

Recently, advances in high-throughput transcriptomic, proteomic, metabolomic, and cytomic technologies have enabled the characterization of the complexity of localized and systemic diseases [1–5]. In this review, we focus on the interplay between oral pathology triggers and the host's immune response mechanisms in the development, maintenance, progression, and resolution of pathological processes in the oral cavity. Importantly, we highlight recent technological advances that allow a multiomic assessment of immunological events involved in oral pathologies and the improvement of clinical care towards targeted immunotherapies and diagnostics.

Immune involvement in oral pathologies

The mucosa of the oral cavity represents an important physiological barrier of the human body and, therefore, demonstrates a high level of immune cell presence along its surface under healthy conditions. In consequence, emerging oral pathologies show a particularly strong immune involvement. The deployed functional defense mechanisms on one hand and failure, dysfunction, or hyperfunction of the host's immune response on the other determine the development

and maintenance of pathological states. In the following paragraphs, we elucidate the elemental role of immune cells in the most common neoplastic, infectious, and autoimmune diseases of the oral cavity (Fig. 1) and demonstrate how the immune system's involvement in oral pathologies can be leveraged to enhance early detection, prevention, treatment, and resolution of these diseases.

Neoplasms

Neoplasms or abnormal growths of tissue can occur anywhere in the body, but the oral mucosa is particularly prone to neoplastic processes, either from genetic, reactive, environmental, or unknown triggers. Oral cavity cancer, most commonly squamous cell carcinoma, accounts for approximately 2% of all cancer diagnoses [6, 7]. In many patients, oral cavity squamous cell carcinoma (OSCC) is associated with alcohol and tobacco use, a small but increasing number of cases is driven by human papilloma virus (HPV)[8], and a third category of patients have no known risk factors. Pathogenetically, the chronic insult by carcinogens leads to the development of precancerous and dysplastic lesions that develop into malignancies. Mutagenesis is also driven by chronic inflammation in the context of bacterial or viral infection [9], which is demonstrated by the strong association between persistent inflammation in chronic periodontitis and OSCC [10]. As these carcinogenic influences act on the oral cavity as a whole (field cancerization theory), cancer recurrence and metastasis, as well as synchronous or asynchronous secondary malignancies represent difficult
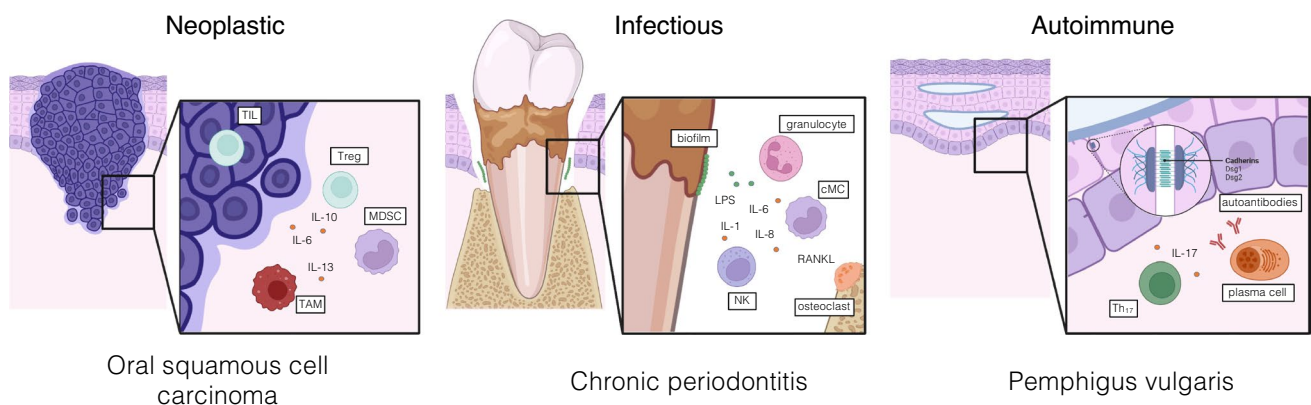

Fig. 1 Immune involvement at the mucosal barrier. Immune cells are heavily involved in diseases of the oral mucosa. In neoplastic malignancies (*left*) such as oral cavity squamous cell carcinoma (OSCC), tumor invasion results in a pronounced immune reaction in the surrounding tissue. Abundance and composition of anti-tumor immune infiltrates in the tumor microenvironment, including tumor-associated macrophages (TAM) and tumor-infiltrating lymphocytes (TIL), as well as levels of immunosuppressive myeloid-derived suppressor cells (MDSCs) and regulatory T cells, represent prognostically relevant markers. In infectious diseases (*middle*), such as chronic peri-

odontitis, the activation of immune cells, such as neutrophils and monocytes, by dysbiotic bacteria leads to a pronounced localized immune response resulting in the destruction of soft tissue and bone. Autoimmune diseases of the oral mucosa (*right*), such as pemphigus vulgaris, are characterized by autoreactive immune cells and autoantibodies targeting adhesive junctions in the oral mucosa. By disrupting the epithelial integrity, the inflammatory process leads to chronic blistering and painful ulcers and increases the mucosal susceptibility to bacterial infection and tissue destruction

clinical challenges and contribute to the poor prognosis (50–60% 5-year survival rate) of OSCC [11–13]. Current treatment regimes, primarily relying on surgical removal of the tumor in combination with adjuvant radiation and chemotherapy for higher tumor stages, leave room for optimization to improve outcomes, personalize treatment, and increase quality of life.

Despite many efforts to identify biological determinants of disease evolution in OSCC, existing predictive tools in patient surveillance and treatment are limited. To bridge this knowledge gap, understanding immune cell-mediated mechanisms is a high-yield approach due to the important role of the immune system in the development, progression, and metastasis of OSCC at the local and systemic levels. Locally within the tumor microenvironment, the mutual influence of innate and adaptive immune cells and cancer cells is a key determinant of tumorigenesis and the response to treatment [14, 15]. On the other hand, systemic immune dysfunction, such as immunodeficiencies and immunosuppressive treatment, increase the risk for many malignancies, including OSCC, particularly with HPV-driven pathogenesis [16, 17]. A better understanding of immune mechanisms and involvement, both detrimental and beneficial, in the pathogenesis of OSCC can be leveraged to improve early detection, prediction of outcomes, and treatment optimization strategies.

For instance, a proteomic analysis of saliva comparing patients with OSCC to healthy individuals identified significant differences in the abundance of molecules related to the acute inflammatory response and regulation of humoral immune responses (e.g., complement factors, serotransferrin, and fibrinogen) [18]. These salivary diagnostics can be useful in improving early detection protocols, as well as for tumor surveillance after initial treatment. For example, towards early detection and risk stratification, antibody positivity against HPV16 oncogenic proteins E6/E7 in the plasma years before diagnosis is highly associated with the development of HPV-positive OSCC [19]. For tumor surveillance, salivary levels of HPV DNA after surgical tumor removal have demonstrated remarkable accuracy for the prediction of OSCC recurrence [20, 21].

An analysis of the immune infiltrates at the tumor invasive front represents a promising path to predict outcomes more accurately than current tumor classification systems: expression of T cell subset markers and their distribution in and around the tumor can be useful in developing an immune scoring system that differentiates patients based on their survival [22]. In addition to the importance of histological examination of tumor tissue to quantify the immune involvement in OSCC, systemic-scale analyses reveal distinct immune changes that occur in patients with OSCC. Peripheral blood immune signatures are strongly tilted towards a state of immunosuppression, and multiple studies have found an increase in peripheral regulatory CD4$^+$ T cells

and myeloid-derived suppressor cells in patients with OSCC [23, 24]. These patterns of immunosuppression also recur in proteomic analyses of saliva samples that show increased concentrations of immunosuppressive IL-10 and IL-13 [25]. Suppression of anti-tumoral immune responses mechanistically should result in a worse prognosis, and indeed evidence suggests that regulatory T cells could play a role in the recurrence of OSCC [26]. However, the potential of these distinct immune signatures to predict clinical outcomes and treatment response has only been partially exploited.

As for many other cancers, immunotherapies are increasingly incorporated into clinical protocols as adjuvant or neoadjuvant treatment options. Immune checkpoint inhibitors, such as PD-1/PD-L1 or CTLA-4 inhibitors, leverage the host's antitumoral immune response by redirecting existing immune defense mechanisms against tumor cells. Similarly, the discovery of regulatory immune receptors on tumor-fighting T cells or monocyte-derived suppressor cells, such as Vista, Tim-3, and Lag [27–29], has extended the repertoire of immunomodulatory protein targets. However, treatment success is highly variable and not all patients benefit from immune checkpoint inhibitor therapy [30]. This inconsistent treatment response may be partially explained by the fact that PD-1/PD-L1 expression is highly variable and modulated by inflammatory and hypoxic conditions in the tumor microenvironment [31, 32]. However, expression of PD-1/PD-L1 alone represents an insufficient predictor of treatment success [33]. In-depth functional and phenotypic analysis of the abundant immune populations (e.g., cytotoxic CD8$^+$ T cells [34] and tumor-associated macrophages [35, 36]) at the tumor invasive front of OSCC can help identify how immune presence influences the response to immunotherapies [37, 38]. With a wider spectrum of available immunotherapies, including checkpoint inhibitors or growth-factor receptor antibodies, patient stratification approaches based on tumor phenotype and immune microenvironment are necessary to tailor the best treatment to each individual patient [39].

Infections

Of the up to 700 species of microbes present in the oral cavity, including bacteria, fungi, viruses, and protozoa, the bacterial colonies are best characterized and have many implications for oral and systemic health. The diversity of bacterial species reflects the presence of multiple biological niches with varying conditions, from hard tissues on which the bacteria are arranged in biofilms to mucosal surfaces, from aerobic to (more pathologically) anaerobic environments (e.g., in deepening periodontal pockets under accumulated calculus) [40, 41]. This strong bacterial presence holds the pathogenetic potential for widespread diseases of the oral cavity. One of the most pertinent oral mucosal infections is periodontitis, which manifests as either a localized

or generalized, acute or chronic process. In chronic periodontitis, inflammation triggered by bacteria, such as the gram-negative, facultative anaerobe *Porphyromonas gingivalis*, ultimately causes breakdown of connective tissue and alveolar bone around the teeth. Despite the initiation by bacteria, it is mainly the host's inflammatory immune response that determines the destructive character of the disease [42]. Bacterial virulence factors, e.g., lipopolysaccharide (LPS), directly activate the host's immune cells via Toll-like receptor (TLR) 2 and TLR4 on the surface of innate immune cells, leading to release of pro-inflammatory mediators resulting in the characteristic tissue destruction. Chronic periodontitis is highly prevalent, affecting 46% of the U.S. population [43], and it is epidemiologically associated with many other systemic conditions. In the case of cardiovascular disease, periodontal bacteria can exacerbate these conditions by translocating into the bloodstream and directly promoting the formation of atherosclerotic plaques through innate and adaptive immune mechanisms [44, 45]. Furthermore, immune cells activated locally by bacteria at the gingival sulcus circulate systemically and contribute to adverse pregnancy outcomes (preterm birth, preeclampsia), diabetes, Alzheimer's disease, and some cancers. [46–52] To date, bacterial diseases of the mouth are often refractory to available treatments, particularly in the case of periodontitis, leading to continued insult to the mouth and body from a prolonged, infected state.

In chronic lesions of periodontitis, disease progression towards tissue destruction and bone loss is driven by complex interactions between periodontal bacteria and the host's proinflammatory immune response. The release of cytokines such as receptor activator of nuclear factor kB ligand (RANK-L), interleukin (IL)-1β, IL-6, tumor necrosis factor (TNF)-α, and prostaglandin E_2a as well as increased proteolytic enzyme expression (e.g., of matrix metallopeptidase-13) by activated immune cells are immunological hallmarks of the disease [53]. Additionally, increased reactive oxygen species released by activated neutrophils and reduced antioxidative compensation, both locally and systemically, crucially contribute to periodontal pathogenesis [54]. In proteomic analyses of blood and gingival crevicular fluid, these factors can aid in the surveillance and prediction of periodontitis progression [55]. Similarly, phenotypic and functional analyses of the immune infiltrates at gingival lesions have increased our knowledge of the underlying pathomechanisms in periodontitis. Plasma cells represent one of the most predominant immune cell subsets in periodontitis and have been demonstrated to exert an important role in the initiation of osteoclastogenesis [56–58]. To determine the effectiveness of different treatment approaches, tracking the dynamic evolution of the disease over time is of crucial importance. A transcriptomic longitudinal study of periodontitis in a primate model identified gene expression patterns in the gingival tissue that demarcated phases of initiation, progression, or remission in chronic periodontitis [59]. Therefore, temporal resolution can unveil biomarkers for disease resolution or treatment success. Of particular interest are systemic immune shifts that can indicate the outcome of current, suboptimal treatment approaches, which mainly consist of scaling, root planing, and potentially local antibiotic treatment, or trials of innovative, novel therapeutics [60]. By capturing single-cell immune activation at a system level, peripheral blood signatures of active chronic periodontitis and disease remission can be recorded (Fig. 2). A recent study using suspension mass cytometry, i.e., cytometry by time-of-flight mass spectrometry (CyTOF), analysis of peripheral blood in patients with periodontitis showed heightened innate immune signaling in response to *P. gingivalis*-LPS and IL-2, 4, and 6, while adaptive immune branches showed marked inhibition of JAK/STAT signaling pathways, changes which were found to be reversible after standard treatment [61]. Such high-dimensional approaches can point towards hallmarks of localized inflammation, mechanistic links to systemic disease, and biomarkers for patient surveillance after treatment.

Autoimmune conditions

Autoimmune diseases of the oral cavity are less prevalent than neoplastic and infectious diseases, but they cause marked reduction in quality of life, and their treatment options are often limited [62]. Oral lichen planus, recurrent aphthous stomatitis, pemphigus vulgaris, and mucous membrane pemphigoid are among the most frequently occurring autoimmune pathologies that affect the oral mucosa, and all suffer from incomplete understanding and/or lack of treatment options. Curative treatments for these diseases are often non-existent, and symptomatic management is typically achieved with blunt immunosuppressive treatments, such as topical steroids, and avoidance of exacerbating lifestyle factors, such as stress or dietary triggers [63].

One of the most common of these disorders, oral lichen planus, is a CD8^{+} T cell-mediated inflammatory condition with no known cause or cure [64, 65]. Activation of cytotoxic CD8^{+} T cell and T helper cells through antigens presented on basal keratinocytes trigger a cascade of cytokine release (e.g., TNF-α for the recruitment of other inflammatory immune cell subsets), cytotoxicity against keratinocytes (e.g., via granzyme B and Fas-ligand), and destruction of vital tissue structure (e.g., by matrix metalloproteinases) [66]. On a transcriptomic level, RNA-sequencing has allowed for identification of the dysregulated genes in oral lichen planus, which are mostly involved in T cell activation and the Wnt signaling pathway in keratinocytes [67]. Another highly prevalent disease, recurrent aphthous
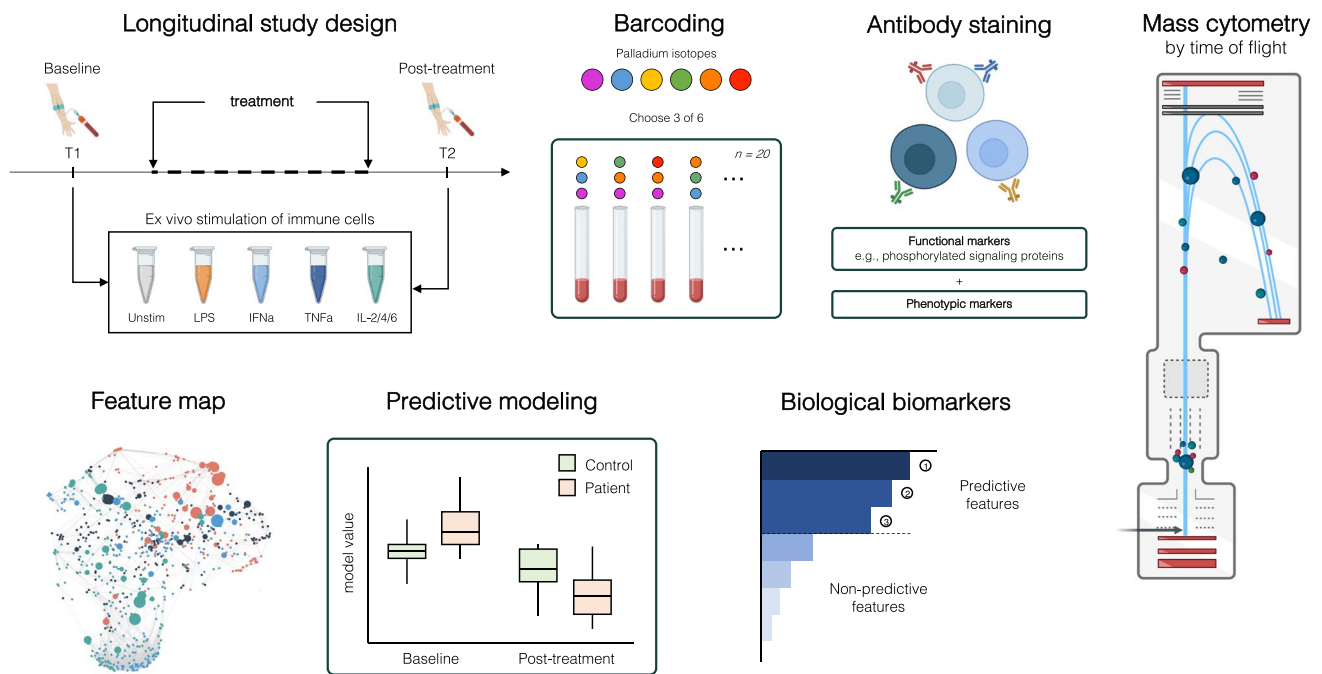
Fig. 2 Systemic immune profiling in longitudinal studies using suspension mass cytometry (CyTOF). Using CyTOF, systemic immune signatures can be profiled in a longitudinal study design, e.g., before and after treatment. In a streamlined workflow, collected blood samples are barcoded for batch processing, stained with antibodies for phenotypic and functional markers and analyzed using CyTOF. The acquired single-cell data can be visualized and interpreted using clustering algorithms, and machine learning approaches can produce and validate reliable predictive models. In the end, the most predictive individual features are derived as biomarkers for disease, treatment success, or outcome

stomatitis, resembles oral lichen planus in its T cell-mediated inflammatory pathophysiology and episodically affects up to 20% of the population [68, 69].

Alternatively, autoreactive B cell subsets and antibody-releasing plasma cells can take center stage in autoimmune diseases: in oral pemphigus vulgaris, IgG autoantibodies are directed against members of the cadherin class of cell–cell adhesion molecules (e.g., desmoglein 1 and 3) and induce the formation of blisters by activating p38, MAPK, and mTOR signaling in keratinocytes, prompting cytoskeleton collapse and disrupting intercellular junctions [70]. Analysis of the transcriptome of pemphigus vulgaris lesions showed an IL-17A–dominated immune signature, and further analyses confirmed an increase in Th17 immune cells which contribute to the induction of desmoglein-specific autoantibody production by B cells [71, 72]. Despite the well-characterized pathophysiology of pemphigus vulgaris, corticosteroids still represent the most commonly used therapy for symptomatic management, while the development of targeted immune therapies is still in the early stages [73, 74], including promising results by targeting Bruton's tyrosine kinase in autoreactive B cells with novel small molecule inhibitors [75]. Finally, mucous membrane pemphigoid, a similarly presenting blistering disease affecting the skin and mucous membranes, is also characterized by autoantibodies attacking epithelial structures, but the antigen targets differ from those in pemphigus vulgaris and are more variable, as they can include intracellular (BPAg1), transmembrane (BPAg2, integrins), or extracellular (collagen VII) proteins [76, 77]. Beyond the involvement of autoantibodies, little is known about the pathophysiological mechanisms of blister formation in mucous membrane pemphigoid, and targeted treatments are lacking [78].

High-parameter omics to identify diagnostic and therapeutic predictive biomarkers

Oral mucosal pathologies are a heterogeneous group of diseases, ranging from rare to highly prevalent conditions that have serious consequences to health and survival. For example, they can promote tumorigenesis towards the development of OSCC, exacerbate other diseases throughout the body, or cause chronic pain and functional restrictions. Despite these devastating consequences, they achieve relatively little notice in research and science. In recent years, the development of high-dimensional and single-cell technologies has enabled the assessment of cytomic, proteomic, transcriptomic, and metabolomic alterations with unprecedented resolution (Table 1). Application of these emerging omic technologies, routinely utilized to investigate other

Table 1 Overview: characteristics of existing omic methods. Transcriptomics, proteomics, metabolomics, and (spatial) cytomics capture biology at different levels of cellular function. While omic methods differ in their advantages and disadvantages, integrative multiomic studies can strengthen and empower the information content from each omic by describing biologically and clinically relevant interomic interconnectivity

Omic	Transcriptomics	Proteomics	Metabolomics	Cytomics	Spatial cytomics
Most common technologies	Single cell RNAseq [79] Bulk RNAseq [80] Microarrays [81]	Antibody-[82] or aptamer-based [83] detection MALDI-TOF [84]	Mass spectrometry [85]	Multiplex flow cytometry Mass cytometry [86]	Imaging mass cytometry [87] MIBI-TOF [88] CODEX [89]
Analytes	RNA transcripts (microRNA, messenger RNA, long non-coding RNA)	Soluble proteins	Molecules from cell metabolism	Protein expression in single cells	Protein expression (in situ)
Investigated specimen	Lysed single cells/bulk cells	Body fluids (e.g., saliva, plasma, gingival crevicular fluid [90])	Body fluids (e.g., saliva [91], plasma, gingival crevicular fluid)	Cell suspensions (e.g., whole blood [61], PBMCs)	FFPE tissue or fresh frozen tissue (e.g., oral mucosal biopsies)
Advantages	In-depth information about gene expression on a single cell level	Easy translation to clinical biomarker, sample accessibility,	Agnostic measurement of cellular function, downstream knowledge about cellular activity	Proteomic readout of cell phenotype and function, interpretable single cell information	Interpretation of cell distribution in its spatial context, interpretable single cell information
Disadvantages	mRNA instability, posttranscriptional modifications	Need for validation of antigen capture, no interpretability on a single cell level	Need for validation/ curation of putative metabolites, sensitive to environmental variations	Sensitive to sample processing, limited number of markers measured per cell	Sensitive to sample processing, comparatively few markers, variable spatial resolution
Number of analytes	Untargeted or targeted, tens of thousands of gene transcripts	Up to 7,000 proteins [92]	Untargeted, 220,000 analytes documented in the human metabolome database [93]	Up to 50 proteins	Up to 60 proteins [94]

malignant, infectious, or auto-immune disease processes, is urgently needed to develop an integrative view of complex pathophysiological processes underlying oral mucosal pathologies [95].

CyTOF is a powerful analytic platform for the assessment of whole system's immune alterations in neoplastic, infectious, and autoimmune oral pathologies. CyTOF, in contrast to conventional fluorescence-based flow cytometry, uses metal isotope-conjugated antibodies to measure over 50 parameters without significant spectral overlap on a single-cell level. Our previous work on chronic periodontitis illustrates the use of CyTOF to quantify over 800 immune cell phenotypic and functional features for an in-depth characterization of systemic immune perturbations in patients with chronic periodontitis before and after conventional treatment [61]. This longitudinal, prospective analysis identified an exaggerated proinflammatory response to *P. gingivalis*-derived LPS in neutrophils and monocytes as a main characteristic of systemic inflammation. Importantly, the differences between controls and patients with chronic periodontitis identified by a cell-signaling elastic net algorithm (csEN) markedly diminished after total-mouth disinfection treatment. In studies with larger cohorts, these findings should be tested for their generalizability, and cytomic immune profiling should be used to measure the success of new targeted treatment options.

Complementing the multiplex analysis of circulating immune cells with CyTOF, high-dimensional imaging technologies have emerged and combine cell-level proteomic data with spatial information about the in situ location of single cells. Imaging mass cytometry (IMC) [87], multiplexed ion beam imaging by time of flight (MIBI-TOF) [88], or co-detection by indexing (CODEX/PhenoCycler) [89, 96] allow the simultaneous detection of up to 60 protein markers for phenotype and function in tissues. With refined deep-learning cell segmentation algorithms, raw images can be converted into single-cell data for downstream analysis that might comprise supervised manual clustering, unsupervised clustering approaches, and spatial arithmetics such as neighborhood or distance-to-border analyses (Fig. 3) [97–99]. Additionally, these platforms can enable the simultaneous detection of protein and mRNA (RNAscope) targets, as recently demonstrated by Schulz et al. for the tumor microenvironment of breast cancer and melanoma [100, 101].
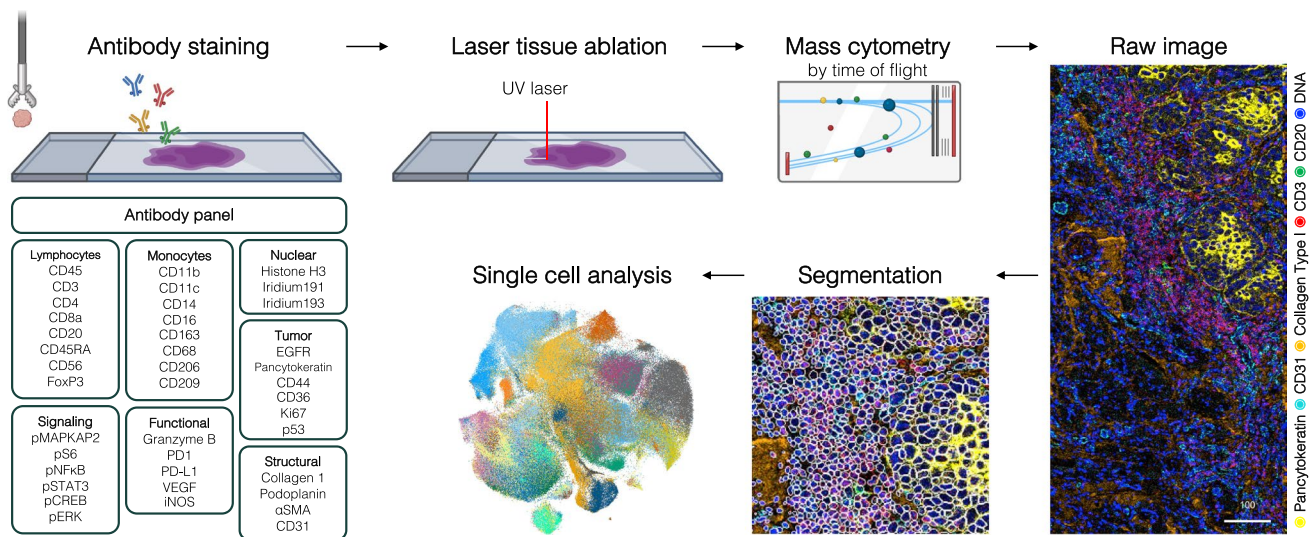
Fig. 3 Spatial single-cell immunome studies on the horizon. Imaging mass cytometry (IMC) allows single-cell proteomics with up to 50 markers of fresh-frozen or formalin-fixed paraffin-embedded tissue slices. Pixel by pixel, the stained tissue is ablated using a UV laser and analyzed using mass spectrometry by time of flight. The raw images of marker intensities can be segmented using various cell segmentation techniques to produce single-cell data, which can be analyzed using existing bioinformatics tools for cytomic datasets

While cytomic approaches (i.e., single-cell proteomics) aim to measure protein expression within individual cells, bulk proteomic assays enable the detection of soluble proteins in bodily fluids. Proteomic assays are often performed in serum or plasma allowing the identification of peripheral-blood proteomic signatures that can ultimately guide clinical decisions, for example, by differentiating between metastatic head and neck squamous cell carcinoma and primary squamous cell lung cancer [102]. However, proteomic assays are readily amenable to the analysis of other compartments, such as gingival crevicular fluid [103] or saliva [104, 105]. These oral compartments are of particular interest in studies of oral pathologies as they provide non-invasive liquid biopsies that contain biologically relevant proteomic markers, often at higher concentrations than in the circulation. Recent advances in multiplex proteomics allow the simultaneous detection of thousands of proteins using modern antibody-based (proximity extension assay, Olink) or aptamer-based (Somalogic) platforms from a small biological sample size (< 100 μl), overcoming previous constraints of limited simultaneous detection capabilities [82, 83]. The gingival crevicular fluid proteome holds important prognostic information for periodontitis progression [90], and the salivary proteome has been targeted in the search for OSCC biomarkers, yielding promising candidate biomarkers, such as elevated interleukin levels (IL-6 and IL-8), tumor antigens (CA125 and CD44), and functional proteins (Ki67 and MMP9) [106]. Hu et al. found a set of five proteomic salivary markers (M2BP, MRP14, CD59, catalase, and profilin) that identified patients with OSCC with an AUC of 93% [107]. Although limited in

sample size, these studies offer promising proof-of-concept information for the use of proteomic analyses of salivary proteins to detect OSCC. Further studies will be needed to validate the findings and to integrate them with metabolomic and cytomic data.

In contrast to cytomic and proteomic platforms that provide a targeted analysis of pre-selected protein analytes, RNA sequencing (RNAseq) platforms offer agnostic detection of over 20,000 gene transcripts in bulk analysis of pooled (bulk RNAseq) or single cells (scRNAseq). A scRNAseq analysis of head and neck squamous cell carcinoma by Puram et al. described distinct transcriptional patterns in the epithelial-to-mesenchymal transition of tumor cells that are linked to nodal metastasis and histological tumor grade [108]. In other malignancies, scRNAseq not only helps elucidate key hallmarks of tumor pathogenesis and progression [109] but also optimize therapeutic strategies [110]. RNAseq approaches are also commonly utilized to characterize transcriptomic dysregulations in autoimmune diseases. In rare diseases such as pemphigus vulgaris, scRNAseq analysis can be a rewarding first approach to guide follow-up, targeted investigations. For example, a bulk RNAseq approach that was recently employed to study the dysregulated peripheral immune system of patients with pemphigus vulgaris unveiled that B cells express increased levels of IL-1β, IL-23, and IL-12 and that different treatment approaches correct these dysregulations differently [111]. Transcriptomic technologies have also evolved to enable spatial resolution of gene expression patterns in tissues. Spatial transcriptomic platforms using next-generation sequencing offer

untargeted mRNA detection and can measure hundreds or thousands of genes per pixel [112, 113]. However, a technical limitation of most existing spatial RNAseq approaches is the requirement of fresh frozen tissue as these techniques are generally incompatible with formalin-fixed paraffin-embedded tissue.

In addition to proteomic and transcriptomic assays, untargeted mass spectrometry analyses of salivary metabolites have been an active field of research in the search for prognostic biomarkers in oral mucosal pathologies. In periodontitis, the metabolites cadaverine and hydrocinnamate positively correlate with the area of active inflammation, whereas uric acid and ethanolamine indicate resolution of inflammation [91]. Some of the identified metabolites in periodontitis are also predictive of OSCC suggesting a pathophysiological link and emphasizing the mutagenic potential of chronic inflammation [114–116]. Ultimately, metabolomic markers can point towards altered cellular biology and functionality in OSCC and can be used to

predict overall survival [117] or response to treatment, such as chemotherapy [118].

Integrated, multiomic modeling to identify new biological crosstalk and improve biomarker discovery

Capturing multiple high-dimensional and/or single-cell modalities in a multiomic approach allows the integration of multiple biological layers into a unified biological representation of the investigated scientific question (Fig. 4). A multiomic approach also enables analysis of inter-omic crosstalk, which can be useful for confirming the validity of identified biological processes when observed in multiple data layers. However, the integration of multiple high-dimensional omic data layers poses certain statistical challenges, including differences in omic data layer dimensions, and information content. Emerging machine
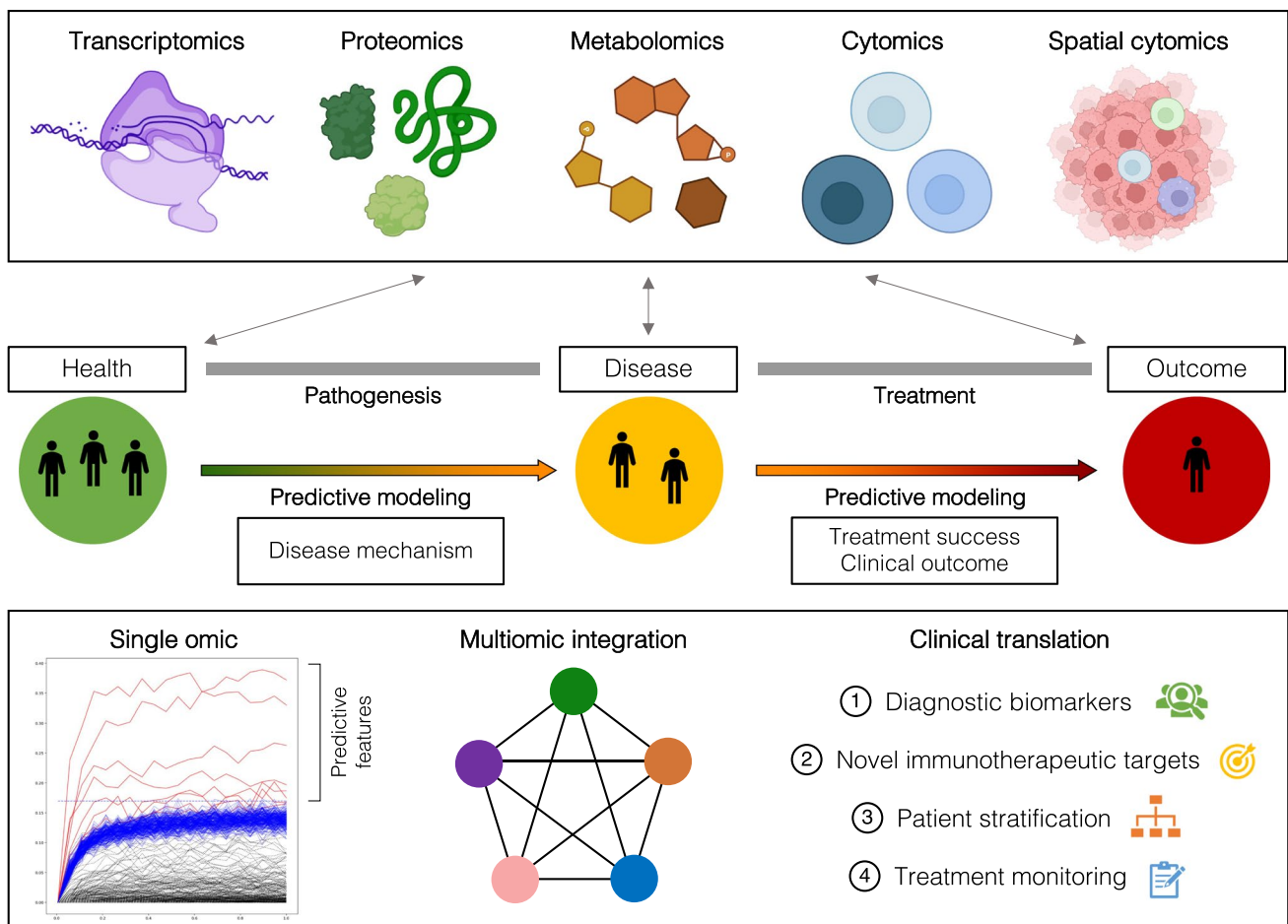

Fig. 4 Towards integration of multiomic data layers to improve classification of oral pathologies and outcomes. Integrative studies of transcriptomic, proteomic, and cytomic biology that contribute to oral mucosal pathologies will be instrumental in defining predictive models of disease mechanism, treatment success, and outcomes. Using machine learning approaches to analyze the individual data layers and combining the predictive power of interlinked biological features will advance the discovery of diagnostic biomarkers and novel immunotherapeutic targets and enable improved patient risk stratification and treatment monitoring

learning approaches have recently been developed that provide elegant solutions to concatenate multiomic information about gene expression, protein abundance, single-cell signaling activity, and phenotype as well as metabolism [119]. The timing in which the datasets are combined (early-fusion vs. late-fusion) and the applied regularization and penalization differ between analysis approaches. A commonly used approach is based on stacked generalization in which predictive models are built on each of the individual data layers before incorporating the most predictive features from each omic dataset into one overarching model [1, 3, 120]. Establishing predictive models for each data layer prior to combining them into an integrated model using stacked generalization can increase the predictive power, as this approach can account for feature intercorrelation within data layers and differences in the number of measurements between data layers. Subsequently, a post hoc correlation network analysis of selected model features can reveal relationships between features from different data layers and inform about biological cross-talks. Future challenges in computational analyses of multiomic data that are particularly relevant to complex, cross-tissue analyses of oral mucosal pathologies include the integration of spatial information such as cell–cell or cell-stroma interactions and the optimization of objective feature selection algorithms to aid in the biological interpretation of multivariate models and facilitate the biomarker discovery process.

Conclusions

Patients with neoplastic, infectious, and autoimmune oral mucosal pathologies currently face limited and insufficient clinical treatment options. Recent proteomic, cytomic, and transcriptomic approaches provide promising avenues to elucidate mechanisms of pathogenesis and allowed for discovery of clinically relevant predictive biomarkers in distinct immune compartments. However, the implementation of single-cell and spatial omic technologies to study oral mucosal pathologies is still in its infancy. Future studies that integrate multiple omic modalities are needed to provide a comprehensive characterization of the immunological compartments that interact and contribute to disease development and resolution. Ultimately, large-scale multiomic studies in diverse patient populations will be necessary to identify and validate robust and biologically plausible signatures of clinical outcomes for the targeted development of novel (immuno)therapeutics to improve and personalize the treatment of patients with oral mucosal pathologies.

Data availability Data sharing not applicable to this review article as no datasets were generated or analyzed.

Declarations

Conflict of interest C. M. S. is a scientific advisor to, has stock options in, and has received research funding from Enable Medicine, Inc., all outside of this work. B. G. is a scientific advisor to surge2surgery. The remaining authors have no conflicts of interest to declare.

References

1. Feyaerts D, Hédou J, Gillard J et al (2022) Integrated plasma proteomic and single-cell immune signaling network signatures demarcate mild, moderate, and severe COVID-19. Cell Rep Med 3:100680. https://doi.org/10.1016/j.xcrm.2022.100680

2. Stelzer IA, Ghaemi MS, Han X et al (2021) Integrated trajectories of the maternal metabolome, proteome, and immunome predict labor onset. Sci Transl Med 13:eabd9898. https://doi.org/10.1126/scitranslmed.abd9898

3. Rumer KK, Hedou J, Tsai A et al (2022) Integrated single-cell and plasma proteomic modeling to predict surgical site complications: a prospective cohort study. Ann Surg 275:582–590. https://doi.org/10.1097/SLA.0000000000005348

4. Aghaeepour N, Ganio EA, Mcilwain D et al (2017) An immune clock of human pregnancy. Sci Immunol 2:eaan2946. https://doi.org/10.1126/sciimmunol.aan2946

5. The Cancer Genome Atlas Research Network, Weinstein JN, Collisson EA et al (2013) The Cancer Genome Atlas Pan-Cancer analysis project. Nat Genet 45:1113–1120. https://doi.org/10.1038/ng.2764

6. Cramer JD, Burtness B, Le QT, Ferris RL (2019) The changing therapeutic landscape of head and neck cancer. Nat Rev Clin Oncol 16:669–683. https://doi.org/10.1038/s41571-019-0227-z

7. Sung H, Ferlay J, Siegel RL et al (2021) Global Cancer Statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 71:209–249. https://doi.org/10.3322/caac.21660

8. Bose P, Brockton NT, Dort JC (2013) Head and neck cancer: from anatomy to biology: biology of head and neck cancer. Int J Cancer 133:2013–2023. https://doi.org/10.1002/ijc.28112

9. Gholizadeh P, Eslami H, Yousefi M et al (2016) Role of oral microbiome on oral cancers, a review. Biomed Pharmacother 84:552–558. https://doi.org/10.1016/j.biopha.2016.09.082

10. Tezal M, Sullivan MA, Hyland A et al (2009) Chronic periodontitis and the incidence of head and neck squamous cell carcinoma. Cancer Epidemiol Biomark Prev 18:2406–2412. https://doi.org/10.1158/1055-9965.EPI-09-0334

11. Listl S, Jansen L, Stenzinger A et al (2013) Survival of patients with oral cavity cancer in Germany. PLoS ONE 8:e53415. https://doi.org/10.1371/journal.pone.0053415

12. Wang B, Zhang S, Yue K, Wang X-D (2013) The recurrence and survival of oral squamous cell carcinoma: a report of 275 cases. Chin J Cancer 32:614–618. https://doi.org/10.5732/cjc.012.10219

13. Zini A, Czerninski R, Sgan-Cohen HD (2010) Oral cancer over four decades: epidemiology, trends, histology, and survival by anatomical sites. J Oral Pathol Med 39:299–305. https://doi.org/10.1111/j.1600-0714.2009.00845.x

14. Binnewies M, Roberts EW, Kersten K et al (2018) Understanding the tumor immune microenvironment (TIME) for effective therapy. Nat Med 24:541–550. https://doi.org/10.1038/s41591-018-0014-x

15. Ptacek J, Locke D, Finck R et al (2020) Multiplexed ion beam imaging (MIBI) for characterization of the tumor microenvironment across tumor types. Lab Invest 100:1111–1123. https://doi.org/10.1038/s41374-020-0417-4

16. Beachler DC, D'Souza G (2013) Oral human papillomavirus infection and head and neck cancers in HIV-infected individuals. Curr Opin Oncol 25:503–510. https://doi.org/10.1097/CCO.0b013e32836242b4

17. Katsanos KH, Roda G, Brygo A et al (2015) Oral cancer and oral precancerous lesions in inflammatory bowel diseases: a systematic review. ECCOJC 9:1043–1052. https://doi.org/10.1093/ecco-jcc/jjv122

18. Winck FV, Prado Ribeiro AC, Ramos Domingues R et al (2015) Insights into immune responses in oral cancer through proteomic analysis of saliva and salivary extracellular vesicles. Sci Rep 5:16305. https://doi.org/10.1038/srep16305

19. Kreimer AR, Johansson M, Waterboer T et al (2013) Evaluation of human papillomavirus antibodies and risk of subsequent head and neck cancer. JCO 31:2708–2715. https://doi.org/10.1200/JCO.2012.47.2738

20. Ahn SM, Chan JYK, Zhang Z et al (2014) Saliva and plasma quantitative polymerase chain reaction–based detection and surveillance of human papillomavirus–related head and neck cancer. JAMA Otolaryngol Head Neck Surg 140:846. https://doi.org/10.1001/jamaoto.2014.1338

21. Rettig EM, Wentz A, Posner MR et al (2015) Prognostic implication of persistent human papillomavirus type 16 DNA detection in oral rinses for human papillomavirus–related oropharyngeal carcinoma. JAMA Oncol 1:907. https://doi.org/10.1001/jamaoncol.2015.2524

22. Zhou C, Diao P, Wu Y et al (2020) Development and validation of a seven-immune-feature-based prognostic score for oral squamous cell carcinoma after curative resection. Int J Cancer 146:1152–1163. https://doi.org/10.1002/ijc.32571

23. Lim KP, Chun NAL, Ismail SM et al (2014) CD4+CD25hiCD127low regulatory T cells are increased in oral squamous cell carcinoma patients. PLoS ONE 9:e103975. https://doi.org/10.1371/journal.pone.0103975

24. Zhang Y, Guo J, Jia R (2021) Treg: a promising immunotherapeutic target in oral diseases. Front Immunol 12:667862. https://doi.org/10.3389/fimmu.2021.667862

25. Aziz S, Ahmed SS, Ali A et al (2015) Salivary immunosuppressive cytokines IL-10 and IL-13 are significantly elevated in oral squamous cell carcinoma patients. Cancer Invest 33:318–328. https://doi.org/10.3109/07357907.2015.1041642

26. Schuler PJ, Harasymczuk M, Schilling B et al (2013) Effects of adjuvant chemoradiotherapy on the frequency and function of regulatory T cells in patients with head and neck cancer. Clin Cancer Res 19:6585–6596. https://doi.org/10.1158/1078-0432.CCR-13-0900

27. Long L, Zhang X, Chen F et al (2018) The promising immune checkpoint LAG-3: from tumor microenvironment to cancer immunotherapy. Genes Cancer 9:176–189. https://doi.org/10.18632/genesandcancer.180

28. Lines JL, Pantazi E, Mak J et al (2014) VISTA is an immune checkpoint molecule for human T cells. Can Res 74:1924–1932. https://doi.org/10.1158/0008-5472.CAN-13-1504

29. Anderson AC (2014) Tim-3: an emerging target in the cancer immunotherapy landscape. Cancer Immunol Res 2:393–398. https://doi.org/10.1158/2326-6066.CIR-14-0039

30. Burtness B, Harrington KJ, Greil R et al (2019) Pembrolizumab alone or with chemotherapy versus cetuximab with chemotherapy for recurrent or metastatic squamous cell carcinoma of the head and neck (KEYNOTE-048): a randomised, open-label, phase 3 study. Lancet 394:1915–1928. https://doi.org/10.1016/S0140-6736(19)32591-7

31. Ritprajak P, Azuma M (2015) Intrinsic and extrinsic control of expression of the immunoregulatory molecule PD-L1 in epithelial cells and squamous cell carcinoma. Oral Oncol 51:221–228. https://doi.org/10.1016/j.oraloncology.2014.11.014

32. Takahashi H, Sakakura K, Arisaka Y et al (2019) Clinical and biological significance of PD-L1 expression within the tumor microenvironment of oral squamous cell carcinoma. Anticancer Res 39:3039–3046. https://doi.org/10.21873/anticanres.13437

33. Gao A, Pan X, Yang X, Lin Z (2021) Predictive factors in the treatment of oral squamous cell carcinoma using PD-1/PD-L1 inhibitors. Invest New Drugs 39:1132–1138. https://doi.org/10.1007/s10637-021-01082-w

34. Watanabe Y, Katou F, Ohtani H et al (2010) Tumor-infiltrating lymphocytes, particularly the balance between CD8+ T cells and CCR4+ regulatory T cells, affect the survival of patients with oral squamous cell carcinoma. Oral Surg Oral Med Oral Pathol Oral Radiol Endodontol 109:744–752. https://doi.org/10.1016/j.tripleo.2009.12.015

35. Zhang Q, Liu L, Gong C et al (2012) Prognostic significance of tumor-associated macrophages in solid tumor: a meta-analysis of the literature. PLoS ONE 7:e50946. https://doi.org/10.1371/journal.pone.0050946

36. Petruzzi MNMR, Cherubini K, Salum FG, de Figueiredo MAZ (2017) Role of tumour-associated macrophages in oral squamous cells carcinoma progression: an update on current knowledge. Diagn Pathol 12:32. https://doi.org/10.1186/s13000-017-0623-6

37. Mandal R, Şenbabaoğlu Y, Desrichard A et al (2016) The head and neck cancer immune landscape and its immunotherapeutic implications. JCI Insight 1:e89829. https://doi.org/10.1172/jci.insight.89829

38. Nishii N, Hirotsu Y, Takahashi Y et al (2022) Observation of dynamic changes in neutrophil-to-lymphocyte ratio is useful for evaluating treatment response to nivolumab in PD-L1-negative advanced oral cancer. J Oral Maxillofac Surg Med Pathol 34:833–841. https://doi.org/10.1016/j.ajoms.2022.06.003

39. Almangush A, Leivo I, Mäkitie AA (2021) Biomarkers for immunotherapy of oral squamous cell carcinoma: current status and challenges. Front Oncol 11:616629. https://doi.org/10.3389/fonc.2021.616629

40. Deo PN, Deshmukh R (2019) Oral microbiome: unveiling the fundamentals. J Oral Maxillofac Pathol 23:122–128. https://doi.org/10.4103/jomfp.JOMFP_304_18

41. How KY, Song KP, Chan KG (2016) Porphyromonas gingivalis: an overview of periodontopathic pathogen below the gum line. Front Microbiol 7:53. https://doi.org/10.3389/fmicb.2016.00053

42. Hernández M, Dutzan N, García-Sesnich J et al (2011) Host-pathogen interactions in progressive chronic periodontitis. J Dent Res 90:1164–1170. https://doi.org/10.1177/0022034511401405

43. Eke PI, Dye BA, Wei L et al (2015) Update on prevalence of periodontitis in adults in the United States: NHANES 2009 to 2012. J Periodontol 86:611–622. https://doi.org/10.1902/jop.2015.140520

44. Morishita M, Ariyoshi W, Okinaga T et al (2013) A. actinomycetemcomitans LPS enhances foam cell formation induced by LDL. J Dent Res 92:241–246. https://doi.org/10.1177/0022034512473309

45. Nguyen CM, Kim JWM, Quan VH et al (2015) Periodontal associations in cardiovascular diseases: the latest evidence and understanding. J Oral Biol Craniofac Res 5:203–206. https://doi.org/10.1016/j.jobcr.2015.06.008

46. Ajita M, Karan P, Vivek G et al (2013) Periodontal disease and type 1 diabetes mellitus: associations with glycemic control and complications: an Indian perspective. Diabetes Metab Syndr 7:61–63. https://doi.org/10.1016/j.dsx.2013.03.001

47. Bassani DG, Olinto MTA, Kreiger N (2007) Periodontal disease and perinatal outcomes: a case-control study. J Clin Periodontol 34:31–39. https://doi.org/10.1111/j.1600-051X.2006.01012.x

48. Cardoso EM, Reis C, Manzanares-Céspedes MC (2018) Chronic periodontitis, inflammatory cytokines, and interrelationship with other chronic diseases. Postgrad Med 130:98–104. https://doi.org/10.1080/00325481.2018.1396876

49. Cheng R, Billet S, Liu C et al (2020) Periodontal inflammation recruits distant metastatic breast cancer cells by increasing myeloid-derived suppressor cells. Oncogene 39:1543–1556. https://doi.org/10.1038/s41388-019-1084-z

50. Nabet C, Lelong N, Colombier M-L et al (2010) Maternal periodontitis and the causes of preterm birth: the case-control Epipap study. J Clin Periodontol 37:37–45. https://doi.org/10.1111/j.1600-051X.2009.01503.x

51. Peng C-H, Yang Y-S, Chan K-C et al (2017) Periodontal treatment and the risks of cardiovascular disease in patients with type 2 diabetes: a retrospective cohort study. Intern Med 56:1015–1021. https://doi.org/10.2169/internalmedicine.56.7322

52. Konopka T, Zakrzewska A (2020) Periodontitis and risk for preeclampsia - a systematic review. Ginekol Pol 91:158–164. https://doi.org/10.5603/GP.2020.0024

53. Silva N, Dutzan N, Hernandez M et al (2008) Characterization of progressive periodontal lesions in chronic periodontitis patients: levels of chemokines, cytokines, matrix metalloproteinase-13, periodontal pathogens and inflammatory cells. J Clin Periodontol 35:206–214. https://doi.org/10.1111/j.1600-051X.2007.01190.x

54. Wei D, Zhang X-L, Wang Y-Z et al (2010) Lipid peroxidation levels, total oxidant status and superoxide dismutase in serum, saliva and gingival crevicular fluid in chronic periodontitis patients before and after periodontal therapy. Aust Dent J 55:70–78. https://doi.org/10.1111/j.1834-7819.2009.01123.x

55. Sorsa T, Gursoy UK, Nwhator S et al (2000) (2016) Analysis of matrix metalloproteinases, especially MMP-8, in gingival crevicular fluid, mouthrinse and saliva for monitoring periodontal diseases. Periodontol 70:142–163. https://doi.org/10.1111/prd.12101

56. Berglundh T, Donati M, Zitzmann N (2007) B cells in periodontitis? Friends or enemies? Periodontol 2000 45:51–66. https://doi.org/10.1111/j.1600-0757.2007.00223.x

57. Jing L, Kim S, Sun L et al (2019) IL-37- and IL-35/IL-37-producing plasma cells in chronic periodontitis. J Dent Res 98:813–821. https://doi.org/10.1177/0022034519847443

58. Zouali M (2017) The emerging roles of B cells as partners and targets in periodontitis. Autoimmunity 50:61–70. https://doi.org/10.1080/08916934.2016.1261841

59. Ebersole JL, Nagarajan R, Kirakodu S, Gonzalez OA (2021) Transcriptomic phases of periodontitis lesions using the nonhuman primate model. Sci Rep 11:9282. https://doi.org/10.1038/s41598-021-88803-6

60. Ide M, McPartlin D, Coward PY et al (2003) Effect of treatment of chronic periodontitis on levels of serum markers of acute-phase inflammatory and vascular responses: acute-phase proteins and periodontal treatment. J Clin Periodontol 30:334–340. https://doi.org/10.1034/j.1600-051X.2003.00282.x

61. Gaudilliere DK, Culos A, Djebali K et al (2019) Systemic immunologic consequences of chronic periodontitis. J Dent Res 98:985–993. https://doi.org/10.1177/0022034519857714

62. Riordain RN, Meaney S, McCreary C (2011) Impact of chronic oral mucosal disease on daily life: preliminary observations from a qualitative study: a qualitative study of chronic oral mucosal conditions. Oral Dis 17:265–269. https://doi.org/10.1111/j.1601-0825.2010.01734.x

63. Thongprasom K, Carrozzo M, Furness S, Lodi G (2011) Interventions for treating oral lichen planus. Cochrane Database Syst Rev. https://doi.org/10.1002/14651858.CD001168.pub2

64. Lavanya N, Jayanthi P, Rao UK, Ranganathan K (2011) Oral lichen planus: an update on pathogenesis and treatment. J Oral Maxillofac Pathol 15:127–132. https://doi.org/10.4103/0973-029X.84474

65. Sugerman PB, Savage NW, Zhou X et al (2000) Oral lichen planus. Clin Dermatol 18:533–539. https://doi.org/10.1016/S0738-081X(00)00142-5

66. Olson MA, Rogers RS, Bruce AJ (2016) Oral lichen planus. Clin Dermatol 34:495–504. https://doi.org/10.1016/j.clindermatol.2016.02.023

67. Wang H, Deng Y, Peng S et al (2021) RNA-Seq based transcriptome analysis in oral lichen planus. Hereditas 158:39. https://doi.org/10.1186/s41065-021-00202-z

68. Bruch JM, Treister NS (2010) Clinical oral medicine and pathology. Humana Press, New York

69. Chiang C-P, Yu-Fong Chang J, Wang Y-P et al (2019) Recurrent aphthous stomatitis – etiology, serum autoantibodies, anemia, hematinic deficiencies, and management. J Formos Med Assoc 118:1279–1289. https://doi.org/10.1016/j.jfma.2018.10.023

70. Grando SA, Bystryn J-C, Chernyavsky AI et al (2009) Apoptolysis: a novel mechanism of skin blistering in pemphigus vulgaris linking the apoptotic pathways to basal cell shrinkage and suprabasal acantholysis. Exp Dermatol 18:764–770. https://doi.org/10.1111/j.1600-0625.2009.00934.x

71. Holstein J, Solimani F, Baum C et al (2021) Immunophenotyping in pemphigus reveals a TH17/TFH17 cell–dominated immune response promoting desmoglein1/3-specific autoantibody production. J Allergy Clin Immunol 147:2358–2369. https://doi.org/10.1016/j.jaci.2020.11.008

72. Huang Z, Qu P, Wang K et al (2022) Transcriptomic profiling of pemphigus lesion infiltrating mononuclear cells reveals a distinct local immune microenvironment and novel lncRNA regulators. J Transl Med 20:182. https://doi.org/10.1186/s12967-022-03387-7

73. Ellebrecht CT, Payne AS (2017) Setting the target for pemphigus vulgaris therapy. JCI Insight 2:e92021. https://doi.org/10.1172/jci.insight.92021

74. Popescu I, Statescu L, Vata D et al (2019) Pemphigus vulgaris - approach and management (Review). Exp Ther Med. https://doi.org/10.3892/etm.2019.7964

75. Murrell DF, Patsatsi A, Stavropoulos P et al (2021) Proof of concept for the clinical effects of oral rilzabrutinib, the first Bruton tyrosine kinase inhibitor for pemphigus vulgaris: the phase II BELIEVE study. Br J Dermatol 185:745–755. https://doi.org/10.1111/bjd.20431

76. Buonavoglia A, Leone P, Dammacco R et al (2019) Pemphigus and mucous membrane pemphigoid: an update from diagnosis to therapy. Autoimmun Rev 18:349–358. https://doi.org/10.1016/j.autrev.2019.02.005

77. Xu H-H, Werth VP, Parisi E, Sollecito TP (2013) Mucous membrane pemphigoid. Dent Clin North Am 57:611–630. https://doi.org/10.1016/j.cden.2013.07.003

78. Kamaguchi M, Iwata H (2019) The diagnosis and blistering mechanisms of mucous membrane pemphigoid. Front Immunol 10:34. https://doi.org/10.3389/fimmu.2019.00034

79. Patel AP, Tirosh I, Trombetta JJ et al (2014) Single-cell RNA-seq highlights intratumoral heterogeneity in primary glioblastoma. Science 344:1396–1401. https://doi.org/10.1126/science.1254257

80. Ozsolak F, Milos PM (2011) RNA sequencing: advances, challenges and opportunities. Nat Rev Genet 12:87–98. https://doi.org/10.1038/nrg2934

81. Zhao S, Fung-Leung W-P, Bittner A et al (2014) Comparison of RNA-Seq and microarray in transcriptome profiling of activated T cells. PLoS ONE 9:e78644. https://doi.org/10.1371/journal.pone.0078644

82. Lundberg M, Eriksson A, Tran B et al (2011) Homogeneous antibody-based proximity extension assays provide sensitive and specific detection of low-abundant proteins in human blood. Nucleic Acids Res 39:e102–e102. https://doi.org/10.1093/nar/gkr424

83. Rohloff JC, Gelinas AD, Jarvis TC et al (2014) Nucleic acid ligands with protein-like side chains: modified aptamers and their use as diagnostic and therapeutic agents. Mol Ther Nucleic Acids 3:e201. https://doi.org/10.1038/mtna.2014.49

84. Aebersold R, Mann M (2003) Mass spectrometry-based proteomics. Nature 422:198–207. https://doi.org/10.1038/nature01511

85. Dettmer K, Aronov PA, Hammock BD (2007) Mass spectrometry-based metabolomics: mass spectrometry-based metabolomics. Mass Spectrom Rev 26:51–78. https://doi.org/10.1002/mas.20108

86. Bendall SC, Simonds EF, Qiu P et al (2011) Single-cell mass cytometry of differential immune and drug responses across a human hematopoietic continuum. Science 332:687–696. https://doi.org/10.1126/science.1198704

87. Giesen C, Wang HAO, Schapiro D et al (2014) Highly multiplexed imaging of tumor tissues with subcellular resolution by mass cytometry. Nat Methods 11:417–422. https://doi.org/10.1038/nmeth.2869

88. Angelo M, Bendall SC, Finck R et al (2014) Multiplexed ion beam imaging of human breast tumors. Nat Med 20:436–442. https://doi.org/10.1038/nm.3488

89. Goltsev Y, Samusik N, Kennedy-Darling J et al (2018) Deep profiling of mouse splenic architecture with CODEX multiplexed imaging. Cell 174:968-981.e15. https://doi.org/10.1016/j.cell.2018.07.010

90. Kinney JS, Morelli T, Oh M et al (2014) Crevicular fluid biomarkers and periodontal disease progression. J Clin Periodontol 41:113–120. https://doi.org/10.1111/jcpe.12194

91. Sakanaka A, Kuboniwa M, Hashino E et al (2017) Distinct signatures of dental plaque metabolic byproducts dictated by periodontal inflammatory status. Sci Rep 7:42818. https://doi.org/10.1038/srep42818

92. Candia J, Daya GN, Tanaka T et al (2022) Assessment of variability in the plasma 7k SomaScan proteomics assay. Sci Rep 12:17147. https://doi.org/10.1038/s41598-022-22116-0

93. Wishart DS, Guo A, Oler E et al (2022) HMDB 5.0: the Human Metabolome Database for 2022. Nucleic Acids Res 50:D622–D631. https://doi.org/10.1093/nar/gkab1062

94. Hickey JW, Neumann EK, Radtke AJ et al (2022) Spatial mapping of protein composition and tissue organization: a primer for multiplexed antibody-based imaging. Nat Methods 19:284–295. https://doi.org/10.1038/s41592-021-01316-y

95. Hasin Y, Seldin M, Lusis A (2017) Multi-omics approaches to disease. Genome Biol 18:83. https://doi.org/10.1186/s13059-017-1215-1

96. Schürch CM, Bhate SS, Barlow GL et al (2020) Coordinated cellular neighborhoods orchestrate antitumoral immunity at the colorectal cancer invasive front. Cell 182:1341-1359.e19. https://doi.org/10.1016/j.cell.2020.07.005

97. Greenwald NF, Miller G, Moen E et al (2022) Whole-cell segmentation of tissue images with human-level performance using large-scale data annotation and deep learning. Nat Biotechnol 40:555–565. https://doi.org/10.1038/s41587-021-01094-0

98. Windhager J, Bodenmiller B, Eling N (2021) An end-to-end workflow for multiplexed image processing and analysis. bioRxiv 2021.11.12.468357. https://doi.org/10.1101/2021.11.12.468357

99. Lee MY, Bedia JS, Bhate SS et al (2022) Cell Seg: a robust, pre-trained nucleus segmentation and pixel quantification software for highly multiplexed fluorescence images. BMC Bioinformatics 23:46. https://doi.org/10.1186/s12859-022-04570-9

100. Schulz D, Zanotelli VRT, Fischer JR et al (2018) Simultaneous Multiplexed Imaging of mRNA and Proteins with Subcellular Resolution in Breast Cancer Tissue Samples by Mass Cytometry. Cell Syst 6:25-36.e5. https://doi.org/10.1016/j.cels.2017.12.001

101. Hoch T, Schulz D, Eling N et al (2022) Multiplexed imaging mass cytometry of the chemokine milieus in melanoma characterizes features of the response to immunotherapy. Sci Immunol 7:eabk1692. https://doi.org/10.1126/sciimmunol.abk1692

102. Bohnenberger H, Kaderali L, Ströbel P et al (2018) Comparative proteomics reveals a diagnostic signature for pulmonary head-and-neck cancer metastasis. EMBO Mol Med 10:e8428. https://doi.org/10.15252/emmm.201708428

103. Khurshid Z, Mali M, Naseem M et al (2017) Human gingival crevicular fluids (GCF) proteomics: an overview. Dent J 5:12. https://doi.org/10.3390/dj5010012

104. Pillai J, Chincholkar T, Dixit R, Pandey M (2021) A systematic review of proteomic biomarkers in oral squamous cell cancer. World J Surg Oncol 19:315. https://doi.org/10.1186/s12957-021-02423-y

105. Chu H-W, Chang K-P, Hsu C-W et al (2019) Identification of salivary biomarkers for oral cancer detection with untargeted and targeted quantitative proteomics approaches. Mol Cell Proteomics 18:1796–1806. https://doi.org/10.1074/mcp.RA119.001530

106. Cheng YL, Rees T, Wright J (2014) A review of research on salivary biomarkers for oral cancer detection. Clinical and Translational Medicine 3:3. https://doi.org/10.1186/2001-1326-3-3

107. Hu S, Arellano M, Boontheung P et al (2008) Salivary proteomics for oral cancer biomarker discovery. Clin Cancer Res 14:6246–6252. https://doi.org/10.1158/1078-0432.CCR-07-5037

108. Puram SV, Tirosh I, Parikh AS et al (2017) Single-cell transcriptomic analysis of primary and metastatic tumor ecosystems in head and neck cancer. Cell 171:1611-1624.e24. https://doi.org/10.1016/j.cell.2017.10.044

109. Peng J, Sun B-F, Chen C-Y et al (2019) Single-cell RNA-seq highlights intra-tumoral heterogeneity and malignant progression in pancreatic ductal adenocarcinoma. Cell Res 29:725–738. https://doi.org/10.1038/s41422-019-0195-y

110. Kim K-T, Lee HW, Lee H-O et al (2016) Application of single-cell RNA sequencing in optimizing a combinatorial therapeutic strategy in metastatic renal cell carcinoma. Genome Biol 17:80. https://doi.org/10.1186/s13059-016-0945-9

111. Hébert V, Petit M, Maho-Vaillant M et al (2019) Modifications of the transcriptomic profile of autoreactive B Cells from pemphigus patients after treatment with rituximab or a standard corticosteroid regimen. Front Immunol 10:1794. https://doi.org/10.3389/fimmu.2019.01794

112. Stickels RR, Murray E, Kumar P et al (2021) Highly sensitive spatial transcriptomics at near-cellular resolution with Slide-seqV2. Nat Biotechnol 39:313–319. https://doi.org/10.1038/s41587-020-0739-1

113. Williams CG, Lee HJ, Asatsuma T et al (2022) An introduction to spatial transcriptomics for biomedical research. Genome Med 14:68. https://doi.org/10.1186/s13073-022-01075-1

114. Chen X, Yu D (2019) Metabolomics study of oral cancers. Metabolomics 15:22. https://doi.org/10.1007/s11306-019-1483-8

115. Romano F, Meoni G, Manavella V et al (2018) Analysis of salivary phenotypes of generalized aggressive and chronic periodontitis through nuclear magnetic resonance-based metabolomics: salivary metabolomics in periodontitis. J Periodontol 89:1452–1460. https://doi.org/10.1002/JPER.18-0097

116. Sugimoto M, Wong DT, Hirayama A et al (2010) Capillary electrophoresis mass spectrometry-based saliva metabolomics identified oral, breast and pancreatic cancer-specific profiles. Metabolomics 6:78–95. https://doi.org/10.1007/s11306-009-0178-y

117. Ishikawa S, Sugimoto M, Konta T et al (2022) Salivary metabolomics for prognosis of oral squamous cell carcinoma. Front Oncol 11:789248. https://doi.org/10.3389/fonc.2021.789248

118. Ye G, Liu Y, Yin P et al (2014) Study of induction chemotherapy efficacy in oral squamous cell carcinoma using pseudotargeted metabolomics. J Proteome Res 13:1994–2004. https://doi.org/10.1021/pr4011298

119. Ding DY, Li S, Narasimhan B, Tibshirani R (2022) Cooperative learning for multiview analysis. Proc Natl Acad Sci USA 119:e2202113119. https://doi.org/10.1073/pnas.2202113119

120. Ghaemi MS, DiGiulio DB, Contrepois K et al (2019) Multiomics modeling of the immunome, transcriptome, microbiome, proteome and metabolome adaptations during human pregnancy. Bioinformatics 35:95–103. https://doi.org/10.1093/bioinformatics/bty537