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Fever, febrile seizures and epilepsy.

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Authors
Dubé, Céline M
Brewster, Amy L
Richichi, Cristina
et al.

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Seizures induced by fever (febrile seizures) are the most common type of pathological brain activity in infants and children. These febrile seizures and their potential contribution to the mechanisms of limbic (temporal lobe) epilepsy have been a topic of major clinical and scientific interest. Key questions include the mechanisms by which fever generates seizures, the effects of long febrile seizures on neuronal function and the potential contribution of these seizures to epilepsy. This review builds on recent advances derived from animal models and summarizes our current knowledge of the mechanisms underlying febrile seizures and of changes in neuronal gene expression and function that facilitate the enduring effects of prolonged febrile seizures on neuronal and network excitability. The review also discusses the relevance of these findings to the general mechanisms of epileptogenesis during development and points out gaps in our knowledge, including the relationship of animal models to human febrile seizures and epilepsy.

Introduction
Fever provokes seizures in one out of 20–50 children, so that these convulsions, termed ‘ebrile seizures’, are the most common form of pathological brain activity during development [1,2]. In addition, although there is little evidence for an enduring adverse impact of short febrile seizures on the developing brain [1], prolonged or focal febrile seizures (see Glossary) have been associated statistically with the development of intractable epilepsy that involves the limbic circuit (temporal lobe epilepsy; reviewed in [3]). Not surprisingly, these seizures, their underlying mechanisms and their consequences have been a focus of interest for pediatricians and for physicians who treat individuals with epilepsy. In addition, as eloquent examples of perturbed neuronal circuit activity early in life and of activity-dependent plasticity, these seizures have attracted the attention of developmental and systems neurobiologists. To facilitate investigation of these seizures, several animal models of prolonged febrile seizures have been developed (e.g. [4–11]). Over recent years, these models have led to fundamental discoveries about the mechanisms of these seizures, their effects on neuronal excitability [12–18] and their relationship to epilepsy (epileptogenesis, see Box 1). These recent discoveries and the many remaining gaps in our knowledge are the focus of this review.

Mechanisms by which fever leads to ‘febrile’ seizures

Genetic susceptibility to febrile seizures
Febrile seizures occur sporadically and also run in families, so that the contribution of genetic background to their onset has been an active topic of discussion [19]. In immature rodents, hyperthermia provokes seizures in virtually all subjects, suggesting that genetic susceptibility is not a prerequisite for their generation [4,5,7,13]. However, mouse strains differ in their seizure-threshold temperature (a measure of susceptibility [20]), suggesting the involvement of genotype in these seizures. In addition, specific mutations of several ion channels that predispose to febrile seizures have been described in humans [21–24] and rodent models [25]. Among these, mutations of sodium [21,22] and chloride (GABA A [gamma aminobutyric acid A] receptor) [23–25] channels have been prominent. Other single gene mutations might also render individuals more
Fever, febrile seizures and epilepsy: take-home messages

- Febrile seizures are an excellent model of abnormal network activity during development because they do not occur later in life. In addition, they provide biologically relevant examples of activity-dependent, enduring plasticity [14–16,52,56]. The additional importance of these seizures derives from the statistical association of prolonged febrile seizures to human temporal-lobe epilepsy [3]. Animal models of these seizures [4,5,7,11,13,17] offer opportunities to investigate potential causal relationships of these seizures and human epilepsy.
- Studies of the evolution of epilepsy after experimental prolonged febrile seizures suggest that epileptogenesis early in postnatal life might not require cell death [17,52,53]. Rather, enduring functional changes in neuronal and network behavior might provoke episodic hyper-excitability and seizures. These changes are governed by enduring alteration of expression, post-translational modification and function of molecules that govern neuronal excitability (e.g. ion channels and receptors) [12,14,16,52,54–56,59,60]. Among the many changes described (Table 1), altered h, and altered endocannabinoid signaling have been characterized to date and promote transformation of the hippocampal network into one where seizures can be triggered easily.
- The role of genetic factors in both the generation of febrile seizures and their consequences is an important area for investigation. Mutations of ion-channel genes predispose to febrile seizures and influence their relationship to epilepsy [21–24]. The involvement of HCN channels and cannabinoid receptors in the epileptogenic process that follows long experimental febrile seizures suggests that certain mutations in these genes should be epileptogenic.

Susceptible to febrile seizures (see below) and multigene interactions might contribute to the occurrence of these seizures in a more complex and subtle way. In summary, the contribution of the gene–environment interaction to the generation of febrile seizures and to their potential causal relationship to epilepsy [26] is a topic of active investigation. The availability of increasingly sophisticated experimental tools predicts that major breakthroughs in our understanding of the issues involved should emerge in the next few years.

Does fever generate seizures by elevating brain temperature?
Temperature influences numerous cellular processes, including the electrical activity of neurons [27]. The functions of several neuronal ion channels are dependent markedly on temperature in the physiological and fever ranges, approximately 36–42°C (e.g. TRPV4 [transient receptor potential vaniloid 4] [28]), and temperature also modulates the amplitude and kinetics of major ionic currents [29]. These facts suggest that an increase in the temperature of neuronal tissue could enhance the rate, magnitude or synchrony of neuronal firing, leading to seizures; this notion is supported by the fact that, in children, hyperthermia induced by hot baths or anticholinergic medications might also provoke seizures [30]. Measuring brain temperature is not feasible in children with fever; however, direct measurements of intradural temperatures and their correlation with core-body values under normal conditions and during hyperthermia have been carried out in rodent models [31], confirming that, during hyperthermia, brain temperature rises immediately preceding the onset of seizures.

However, whether increased temperature per se suffices to generate electrophysiological and behavioral seizures has remained unclear. Heating brain slices in a dish altered the electrophysiological properties of the hippocampal network but did not provoke clear seizure-like events [32]. By contrast, hyperthermia has been shown recently to alter the activity of certain mutated ion channels, enhancing circuit excitability [25]. Thus, further work is required to determine the direct role of increased temperature in the generation of febrile seizures.

Fever mediators contribute to the generation of febrile seizures
Fever involves the release of cytokines and other inflammatory mediators in the body and within the brain itself [33,34]. Certain cytokines, and specifically interleukin (IL)-1β, enhance neuronal excitability, in part by augmenting glutamate-receptor function [35]. In vivo, these actions of IL-1β enhance the actions of seizure-provoking agents [35]. The possibility that brain hyperthermia might elicit rapid release of endogenous IL-1β [34], which, in turn, contributes to the generation of seizures, was supported by the markedly increased temperature required to induce experimental febrile seizures in mice lacking the IL-1β receptor [20]. Interestingly, mutations in the IL-1β gene promoter that result in increased production of the cytokine have been reported in individuals with febrile seizures [36], although the significance of this finding has been debated [37]. Thus, available data support a significant role for temperature-induced release of endogenous IL-1β in the mechanisms by which hyperthermia – and fever – generates seizures in rodents and humans [9,38]. However, other factors that characterize selectively specific stages of brain development probably contribute in a major way to the age specificity of human and experimental febrile seizures [39,40], as discussed below.

The possibility that fever of specific infectious etiologies might contribute to the generation of human febrile seizures is intriguing in this context: a disproportionate number of febrile seizures have been associated with infection with HHV6 (human herpes virus 6) in some [41] but not all studies [42]. This suggests that mechanisms specific to this virus, including perhaps a unique profile of cytokine induction, might augment neuronal excitability selectively and thus provoke seizures preferentially.

Hyperthermia-induced hyperventilation and alkalosis
Alkalosis of the neuronal environment promotes excitability by several mechanisms [43,44]. The notion that alkalosis might be a key determinant of experimental and human febrile seizures has been proposed [11,45]. In a novel model of febrile seizures, immature rats placed in a heated chamber developed hyperventilation, alkalosis and eventual seizures (Figure 1). In an elegant series of experiments, the alkalosis governed the onset of the seizures; these developed with a latency of 30 min. In other models of febrile seizures, the delay from hyperthermia to seizure-onset was shorter [6,8,46] and did not seem to evoke hyperventilation (46), see Figure 1 for a comparative analysis of the models). Whether alkalosis is instrumental in human febrile seizures
approximately 15 min before the onset of the seizures promotes brain alkalosis and the increased respiratory rate over the minute preceding seizure onset \( p = 0.37 \) versus baseline (values in the graph are shown as mean \( \pm SEM \); derived from [31,46]). In the second model, in which rats are placed in a heated chamber, seizures commence approximately 31 min after initiation of the heating procedure, when body temperature rises from 33.4 \( \pm 0.13 \) \( ^\circ \)C (the seizure-threshold temperature) in 2.9 min, with little change in respiratory rates: 161.8 \( \pm 2.6 \) (n = 12) at baseline and 166.9 \( \pm 4.9 \) during the minute preceding seizure onset \( p = 0.13 \) versus baseline (values in the graph are shown as mean \( \pm SEM \); derived from [11]).

remains unresolved [47]; the presence of alkalosis already at the onset of febrile seizures will support its contribution to their generation. This will require arterial pH measurements at the onset of febrile seizures, yet these seizures rarely commence in settings permitting this analysis.

**Effects of prolonged febrile seizures on neuronal structure and function**

*Do prolonged febrile seizures lead to epilepsy? Clinical issues and experimental approaches*

In prospective human studies, short or simple febrile seizures do not seem to have significant consequences on neuronal function (measured by cognitive tests) or on the probability of epilepsy development [1], although, in very young children, subtle deficits in hippocampus-dependent learning functions might occur [48]. However, the epileptogenic potential of prolonged and/or focal febrile seizures has remained unclear. Prospective studies find a small but significant increased risk for epilepsy and a history of long febrile seizures in individuals who already have temporal lobe epilepsy is common (approximately 30–70%; reviewed in [3]). Human studies are correlative and cannot distinguish between a causal effect of long febrile seizures on epilepsy versus the possibility that pre-existing factors (genetic or acquired) might incite febrile seizures and epilepsy independently or influence the probability that these seizures lead to limbic epilepsy.

Animal models of prolonged febrile seizures have addressed these issues [8,15,17]. Studies of rats subjected to experimental prolonged febrile seizures at the stage of hippocampal development that corresponds to that of human infants [49] followed by chronic video-EEG monitoring demonstrated that a minority of these animals developed limbic epilepsy [17]. Epilepsy or abnormal EEG were absent in controls and in rats that experienced early-life hyperthermia without seizures, supporting the notion that the epilepsy was a direct consequence of the inciting ‘febrile’ seizures. Thus, at least in animal models, prolonged ‘febrile’ seizures can evoke epilepsy, providing a tool for elucidating the responsible mechanisms, for discovering biomarkers for epileptogenesis and for defining windows of opportunity for therapeutic and/or preventive interventions. Clearly, whether epileptogenic processes occurring in the immature rodent brain are analogous directly to those in children remains largely unknown. Parallel human and animal studies, including imaging and molecular analyses, should delineate similarities and divergences among children and immature rats. Ultimately, this issue will be resolved by interventional studies using molecular targets discovered in the animal model and by aiming to prevent human epileptogenesis.

**Experimental prolonged febrile seizure-induced epilepsy provides clues to the puzzle of early-life epileptogenesis**

Although the relevance of experimental long febrile seizures to human disease is not elucidated fully, the occurrence of spontaneous seizures (epilepsy) after these seizures provides a window into the age-specific processes underlying epileptogenesis during the developmental period. For example, a classical mechanism of epileptogenesis in the mature hippocampus involves loss of vulnerable populations of neurons and reorganization of the remaining circuitry [50]. However, significant neuronal loss was not found following single or repeated episodes of experimental febrile seizures [7,15,51], including in rats that became epileptic [17]. This is consistent with the demonstration of epileptogenesis without significant ‘damage’ in other developmental models [52,53] and introduces several concepts regarding early-life epileptogenesis in general. First, the epileptogenic process initiated during development might depend on altered neuronal function, rather than neuronal demise (as discussed later). Second, in human temporal lobe epilepsy with a history of early-life febrile seizures, in which cell loss [mesial temporal sclerosis (MTS)] is often found, the epilepsy might precede – and provoke – MTS.

**Changes evoked by prolonged experimental febrile seizures promote neuronal excitability and seizures**

As mentioned earlier, experimental febrile seizures as well as developmental lithium-pilocarpine [53] or tetanus toxin [52] seizures share a common general mechanism for enhancing hippocampal network excitability and promoting epilepsy. This process involves enduring changes at the molecular and functional levels, such as alterations in neurotransmitter receptors [54,55] or voltage-gated ion channels [14,56]. A wide spectrum of molecular changes has been described after experimental febrile seizures (Table 1); however, the contribution of a given alteration to neuronal excitability and, specifically, to the generation of episodic seizures has been elucidated only for two. These are discussed in the following paragraphs.
The contribution of altered Ih and HCN channel expression to neuronal excitability and seizures

Following experimental prolonged febrile seizures, the properties of Ih, a hyperpolarization-triggered cationic current that contributes to the maintenance of neuronal membrane potential, sub-threshold oscillations and dendritic integration [57–59], were altered in hippocampal pyramidal cells [11,14]. The altered properties of Ih led to the increased probability of frequency-dependent rebound depolarization in response to hyperpolarizing input [14] (which itself was augmented after the seizures [12]). Interestingly, this increased firing probability occurred only in response to hyperpolarizing pulses at a limited frequency range, a fact that helps explain the

Table 1. Changes occurring after experimental or human febrile seizures and their relevance to human epilepsy

<table>
<thead>
<tr>
<th>Change</th>
<th>Animal models</th>
<th>Humans</th>
<th>Significance for epileptogenesis</th>
</tr>
</thead>
<tbody>
<tr>
<td>Structural</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Neuronal death</td>
<td>None in hippocampus [7,17,51] None [15] None for short FS; neuronal loss after long (median 4 h) seizures [8]</td>
<td>Unknown MRI changes, see below</td>
<td>Epileptogenesis found without cell death in rat model [17]</td>
</tr>
<tr>
<td>Sprouting of mossy fibers</td>
<td>Minimal, delayed (3 months) [51] Modest, at 6 months [8] None at 2 months [15]</td>
<td>Unknown Found in hippocampi of TLE patients with history of FS [3,50]</td>
<td>Not always found in human TLE with FS history; absent in FS-evoked epilepsy in models</td>
</tr>
<tr>
<td>Neurogenesis</td>
<td>None at 3, 7, or 28 days [51] None at a week; increased at 7 weeks [10]</td>
<td>Unknown Granule cell dispersion in TLE [72]</td>
<td>Not clear</td>
</tr>
<tr>
<td>MRI changes after prolonged febrile seizures</td>
<td>T2 signal in hippocampus and other limbic structures at a day to 4 weeks [73]</td>
<td>T2 hippocampal signal within days [69] Transient ↑ hippocampal T2 and volume at 2 days [70,71] ↑ hippocampal volume, and DWI intensity within 5 days [74]</td>
<td>Potential biomarker</td>
</tr>
<tr>
<td>Molecular</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Interleukin-1β (IL-1β)</td>
<td>↑ seizure threshold temperature in IL-1RI deficient mice [20] ↑ hippocampal IL-1β levels within hours [9]</td>
<td>↑ frequency of allele promoting IL-1β production in children with FS [36] CSF: ↑ IL-1β in children with FS [75,76] ↑ frequency of allele promoting IL-1β synthesis in TLE [77]</td>
<td>Not clear</td>
</tr>
<tr>
<td>Other cytokines</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Interleukin 6</td>
<td>↑ experimental febrile seizure latency [78]</td>
<td>CSF: ↑ IL-6 in children with FS [36,75]</td>
<td>Not required</td>
</tr>
<tr>
<td>Neuropeptide Y</td>
<td>↑ hippocampal NPY hours after FS [31]</td>
<td>Unknown Unknown</td>
<td></td>
</tr>
<tr>
<td>NMDA receptors</td>
<td>↓ NR2A-mediated ERK1/2 phosphorylation after learning tasks at a month after 9 ‘FS’ [79]</td>
<td>Unknown Unknown</td>
<td></td>
</tr>
<tr>
<td>AMPA receptors</td>
<td>Transient ↓ of GluR2 at 1 day</td>
<td>Unknown Unknown</td>
<td></td>
</tr>
<tr>
<td>HCN channels</td>
<td>↓ HCN1 expression ↑ heteromeric HCN1/HCN2, altered Ih [56,60]</td>
<td>Increased hippocampal HCN1 expression in TLE with MTS [61] Defined in model, not clear in human</td>
<td></td>
</tr>
<tr>
<td>CB1 receptors</td>
<td></td>
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<td></td>
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<tr>
<td>Fos</td>
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<td></td>
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<tr>
<td>CREB</td>
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<tr>
<td>Functional</td>
<td></td>
<td></td>
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<tr>
<td>GABAergic neurotransmission</td>
<td>↑ PKA dependent presynaptic GABAergic inhibition in CA1 at 1 week [12] Mutations in GABA_A receptors γ2 subunits promote experimental FS [25] ↓ GABA_A receptor inhibition, hippocampus [82]</td>
<td>Mutations in GABA_A receptors γ2 found in individuals with FS+ (GEFS +) [23,24] Low CSF GABA in FS children [83]</td>
<td>Not clear</td>
</tr>
<tr>
<td>Ih</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Endocannabinoid signaling</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Seizure susceptibility</td>
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The contribution of altered Ih and HCN channel expression to neuronal excitability and seizures

Following experimental prolonged febrile seizures, the properties of Ih, a hyperpolarization-triggered cationic current that contributes to the maintenance of neuronal membrane potential, sub-threshold oscillations and dendritic integration [57–59], were altered in hippocampal pyramidal cells [11,14]. The altered properties of Ih led to the increased probability of frequency-dependent rebound depolarization in response to hyperpolarizing input [14] (which itself was augmented after the seizures [12]). Interestingly, this increased firing probability occurred only in response to hyperpolarizing pulses at a limited frequency range, a fact that helps explain the
absence of continuous seizures but the generation of sporadic ones (epilepsy). The abnormal firing of a neuron with altered Ih, a firing that might incite a seizure, would only occur under specific circumstances, i.e., when the neuron receives inhibitory input that is within the vulnerable frequency range. At the molecular level, these changes were a result of enduring (for months) changes in the expression of hyperpolarization-activated cyclic-nucleotide-gated (HCN) channels that conduct this current and perhaps also of increased formation of HCN1/HCN2 heteromeric channels [56,60]. HCN channels are encoded by several genes; HCN1 and HCN2 are expressed the most abundantly in mature hippocampus. Each functional HCN channel consists of four subunits assembled as homomeric or heteromeric tetramers and the composition and relative abundance of the HCN-channel isoform within a given neuron govern the properties of cellular Ih (reviewed in [58,59]). The relevance of the alterations in HCN channels and Ih to human epileptogenesis remains to be defined fully. Altered HCN1 channel expression was found in hippocampi from a subset of humans with temporal-lobe epilepsy and MTS [61]. However, whether mutations in HCN channel genes that alter Ih cause human epilepsy remains unknown.

The contribution of cannabinoid receptors and augmented depolarization-induced suppression of inhibition to network excitability and seizures

Prolonged experimental febrile seizures also promoted network hyperexcitability by leading to a selective reduction of inhibition onto hippocampal principal cells. This was a result of enduring augmentation of depolarization-induced suppression of inhibition (DSI) [16]. DSI is mediated by cannabinoid receptors (CB1) residing in presynaptic neurons [62–65]; in essence, DSI occurs when postsynaptic depolarization elicits the local release of endocannabinoids. When these bind CB1 receptors within presynaptic GABAergic interneurons, GABA release is blocked from these terminals [64], reducing inhibitory input from these interneurons to pyramidal cells. The numbers of pre-synaptic, interneuronal CB1 receptors increased after experimental febrile seizures, which led to larger DSI [11,16], thus selectively reducing inhibition in the involved hippocampal circuit. The initiation of DSI requires depolarization of principal cells, so that its destabilizing effects on the network should be episodic, promoting epilepsy rather than continuous seizures [66]. Functional CB1 receptors exist in human hippocampal formation [67]; whether their expression or function is altered in temporal-lobe epilepsy has not been reported. CB1 receptors also reside in glutamatergic neurons, where they might promote neuroprotection [65]. Therefore, the potential contributions of human CB1 gene mutations to epilepsy are difficult to predict.

Obviously, the two examples discussed here are the ‘tip of the iceberg’. Gene-array and functional studies (Table 1) indicate that prolonged experimental febrile seizures lead to broad changes in the gene-expression programs of numerous molecules (ion channels, receptors, transcription factors) that govern neuronal excitability and network responses. Further work is required to examine these changes and discern whether they contribute to hyperexcitability, are compensatory or are merely epiphenomena, as well as to determine their relevance to febrile seizures and epileptogenesis in children.

Summary

Studying febrile seizures, their mechanisms and their consequences has benefited from animal models, leading to novel discoveries. However, significant gaps in our knowledge remain, including the role of genetic susceptibility in the occurrence of febrile seizures and the enduring effects of these seizures on neuronal function. Fever might promote neuronal excitability through one or a combination of several mechanisms, including hyperthermia per se, inflammatory cytokines and alkalosis, perhaps in the setting of genetic susceptibility. Although short febrile seizures do not seem to elicit long-lasting changes in neuronal function, prolonged experimental febrile seizures might lead to limbic epilepsy. This occurs without cell death, suggesting that, during development, the epileptogenic process does not require neuronal loss. The mechanisms mediating early-life epileptogenesis remain unknown but might involve enduring changes in the expression of key molecules that govern neuronal network function. The relevance of the concepts and mechanisms derived from animal models of prolonged febrile seizures to febrile seizures and epileptogenesis in children remains largely unknown. Parallel human and animal studies, including imaging and molecular analyses, should delineate similarities and divergences among children and immature rats. Ultimately, this issue will be resolved by interventional studies using molecular targets discovered in the animal model and aiming to prevent epileptogenesis in susceptible children who have experienced prolonged febrile seizures.

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