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Publication Date

2018-10-01

DOI

10.1016/j.cobeha.2018.06.016

Peer reviewed



Published in final edited form as:

Curr Opin Behav Sci. 2018 October ; 23: 143–149. doi:10.1016/j.cobeha.2018.06.016.

Inclusion of females does not increase variability in rodent research studies

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Abstract

The underrepresentation of female subjects in animal research has gained attention in recent years, and new NIH guidelines aim to address this problem early, at the grant proposal stage. Many researchers believe that use of females will hamper research because of the need for increased sample sizes, and increased costs. Here I review empirical research across multiple rodent species and traits that demonstrates that females are not more variable than males, and that for most traits, female estrous cyclicity need not be considered. I present statistical simulations illustrating how factorial designs can reduce the need for additional research subjects, and discuss cultural issues around the inclusion of male and female subjects in research.

Keywords

Sex; sex differences; variability; variance; male; female; sexual dimorphism

Introduction

Female subjects are underrepresented in animal research across disciplines[1]. Yet studies incorporating females have revealed marked differences in basic biological processes such as pain processing[2,3], and lack of pre-clinical research on female subjects has likely resulted in poorer treatment outcomes for women[4,5]. In 2014, noting the potential negative consequences of this sex bias for human health, the NIH instituted policies to encourage use of both male and female animal research subjects, and consideration of sex as a biological variable[6,7]. Biological sex, or classification as generally male or female based on genetic and physiological features is typically distinguished from gender, considered to be a human's self-representation as male, female or other. Inclusion of both sexes in animal research studies should drive important discoveries in both basic and clinically relevant research[5].

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The call for inclusion of females has met with some criticism[8,9]. One oft-used justification for focusing on males is that females are presumed to be more variable, in part due to estrous cyclicity. Other pushback comes from concern that where sex differences exist, use of males and females will reduce statistical power because of greater spread of pooled data or smaller subsamples of each sex[8]. Finally, some are concerned that increased attention paid to sex differences in preclinical studies will overemphasize what are sometimes small differences in the midst of fundamental similarities, or fail to model sex/gender differences in human health that may have important sociocultural components[9].

This review describes the extent of sex bias and presents the findings of empirical studies and analyses that address these concerns surrounding the inclusion of female subjects.

How bad is the status quo?

Sex bias

The use of predominantly male animal research subjects has been documented in many fields. In a survey across biological disciplines, we found male bias in 8 of 10 fields (general biology, neuroscience, physiology, pharmacology, endocrinology, behavioral physiology, behavior, and zoology—reproduction and immunology were the exceptions)[1]. Similar bias toward the use of male subjects has been found in surveys of pre-clinical animal research on pain, cardiovascular disease, diabetes, and surgical methods[10–13]. In the surgical literature, 80% of studies that specified sex used only male subjects.[11]

We found neuroscience to be one of the worst offenders, with over five studies on males for each study on females; only ~20% of studies used both sexes, and 25% did not specify the sex of study subjects (figure 1A). Lack of reporting of subject sex has been documented at similar rates (20, 22, and 26%) in other surveys[13,11,14]. Even when subject sex is reported, it is sometimes not evident until late in the results, or may require accessing online supplementary material. Subsequent analysis of the neuroscience literature suggests that omission of subject sex has decreased in recent years, but the number of male-only rodent studies has increased, and analysis by subject sex in mixed-sex studies remains infrequent[15].

The bias in animal subject use does not reflect differences in rates of men and women presenting with diseases or disorders; the percentage of women diagnosed with a given condition exceeded the percentage of non-human female subjects in research studies on that condition, in each of several disorders sampled[16]. However, the gender of study authors appears to play a role, as female authorship was significantly positively correlated with the inclusion of both sexes and analysis by subject gender/sex[17].

Analysis by subject sex is not the norm

Even when studies include male and female subjects, analysis by sex of the subjects is infrequent. In a survey of surgical research on animals, only 1% of papers analyzed results by sex[11]. In the aforementioned neuroscience example, 6% of the human and animal research papers sampled used both sexes and analyzed results by sex[1]; other fields showed even lower rates.

In 1993, the NIH introduced the Revitalization Act, requiring women be included in NIH-funded clinical research[18], and requiring sub-group analysis by sex to be enabled. A 2011 analysis found less than a third of the studies required by the NIH to analyze results by subject sex/gender had published analysis by this factor[19]. In November 2017, the NIH announced an amendment [NOT-OD-18-014] to the NIH Policy and Guidelines on the Inclusion of Women and Minorities as Subjects in Clinical Research, stipulating the results of valid subgroup analysis (including by subject sex/gender) must be submitted to clinicaltrials.gov. Similar absences of mandatory subgroup analyses have occurred in drug safety reporting to the FDA[20]. At present, the NIH instructs preclinical investigators to report subject sex and consider sex as a biological variable, but analysis of sex differences need not be performed[7]. Surveys of researchers revealed that how subject sex should best be considered in analysis is not uniformly agreed upon, and that researcher discretion in selecting analyses appropriate to the sample is preferred.[21]

Why not use both sexes? Countering assumptions surrounding the use of female subjects

Although good reasons exist to study females or males alone, the rationale for use of only one sex often stems from unfounded concerns about the use of females or both sexes. These assumptions: that females are more variable than males, that females must be tested across the estrous cycle, and that inclusion of both sexes increases variability, are each countered below.

In considering variability, some common statistical terms and principles will be of service. Raw variation in a data set can be described in terms of *standard deviation* (SD), a measure of the dispersion or spread of data. The *coefficient of variation* describes this standard deviation as a fraction of the mean value (SD/mean) so that it is scaled relative to the data. For example, a sample with mean \pm SD of 1 ± 0.05 and one with 1000 ± 50 have equivalent levels of variability relative to their respective means. *Statistical power* refers to the likelihood of correctly rejecting the null hypothesis, for instance to detect differences between samples at specific conventionally defined statistical thresholds (i.e. $p < 0.05$). For a given sample size, power is greater when mean differences between groups are higher. For a given mean difference between groups, power is greater when sample sizes are higher, allowing better estimation of differences in sample means.

Assumption 1: Females are more variable than males

Even if females were more variable than males, it would not diminish the *importance* of studying both sexes, only the difficulty. Fortunately, researchers do not have to contend with that scenario, as recent empirical analyses of variability in males and females show that unstaged females are not more variable than males across diverse traits, from gene expression to hormone levels in multiple species [22–27]. Indeed, male mice appeared to exhibit slightly, but significantly, greater mean variability than female mice (figure 1B), with substantially higher coefficients of variation (CVs) for hormone measures, metabolism-related traits, and morphology. Females did not have significantly greater variability in any category[22]. Slightly greater male variability was also found in analysis of microarray

datasets in both mice and humans[25]. In rats no sex difference in overall variability was found, but males exhibited significantly higher variability on nervous system measures including neurochemistry and electrophysiology measures, while females exhibited higher variability in the category of non-brain measures[23]. In hamsters, no significant differences in variability were found between male, female, and ovariectomized (non-cycling) females[27].

Assumption 2: Females must be tested across the estrous cycle

Predictable variation in hormone circulation across the estrous cycle contributes to some variation in physiology and behavior, but how much? The similar variability between males and *unstaged* females, described above, suggests it is not more than intrinsic variability in males. But perhaps this similar overall variability comes from different sources: it is plausible that females might exhibit consistent estrous-cycle dependent variability in several traits, while males exhibit variability on different time-scales, or more variability between individuals. In depth analysis of a particular trait (body temperature in mice) revealed differences of this sort, with equivalent overall variability in males and females, but different time-scales of variability[24]. Analysis of a variety of traits tested over 2 consecutive estrous cycles in intact female, ovariectomized female, and male hamsters, revealed no consistent timing differences across traits. Furthermore, staging results by phase of the estrous cycle offered no reduction in variability, even for traits such as sucrose preference tests where estrous cycle variation can be detected under circumstances that are optimized to find it[27]. Thus, testing across the estrous cycle can be considered a specific tool for use in a small subset of specific instances rather than a necessary procedure in most studies[28].

While estrous cyclicity is not a major source of variability, other documented factors may provide avenues for researchers to reduce variability and increase statistical power, including the number of animals/cage and rodent strain[22,23]. Consideration of frequently overlooked study details including bedding type[29], biological sex of the experimenter[30], overlap of animal shipment (a stressor) with puberty[31,32], may also increase consistency across studies. That substantial differences between laboratories remain despite careful efforts at duplication of conditions suggests that more factors may be important than currently realized[33].

Assumption 3: Use of both sexes reduces statistical power and slows progress

Thus far we've considered variability of males and females within each group. But for traits in which a robust sex difference exists, pooling males and females in one group would increase the variability around a combined mean. This gives rise to the concern that sample sizes might need to be doubled to identify treatment effects in studies using both females and males[8]. Factorial designs, however, can evaluate main effects of treatment and subject sex with effectively the same power as pair-wise tests, without increased sample size[34]. Additional factors can also be added at the same sample size with approximately the same power, provided the effect sizes of each new factor are no smaller than the effect sizes used to generate the original estimate of the number of subjects needed; in many cases, five or six factors can be evaluated adequately using the sample size needed for only one factor[34,35].

Illustrations of the statistical ramifications of analyzing results with subject sex as a factor are explored in greater detail in *box 1*. The only circumstance in which a notable reduction in power occurs is when there is an interaction between treatment and sex – which is to say that males and females respond very differently to the treatment. In that case, follow-up testing that is methodologically designed to capture sex differences and their origins[28] will be biologically meaningful and important. Larger samples always provide better potential for analysis, and many have called for fewer studies performed on more individuals, particularly in neuroscience[36]. Sub-division of this sample into males and females and use of factorial designs is an effective method of analysis both in theory and in practice[37].

Much ado about sex:

Sex differences can be small or large, insignificant or critical. One concern about the reporting of sex differences in animal studies is that it may lead to overestimation of human sex/gender differences, especially in brain and behavior[9]. One action researchers can take to address this concern is to indicate the effect size of any sex difference that is reported[5]. It is also critical that in translating findings to humans, we are aware that rodents share only some traits with people, and that both sex and gender play a role in differences between women and men.

On the other hand, we shouldn't underestimate the potential importance of sex differences. In some cases, small phenotypic differences stem from fundamental differences in underlying mechanisms. For example, male and female mice exhibit significant differences in pain sensitivity but have largely overlapping distributions. These “small but significant” differences may arise in part based on pain modulatory pathways that differ with subject sex and hormone exposure[2,3]. Sex-specific mechanisms of synaptic inhibition have been discovered in the hippocampus, demonstrating that even basic mechanisms of neuromodulation can vary with sex[38]. Sex differences in the pathways underlying social behaviors in males and females have been documented, even when males and females exhibit similar behaviors, such as pair-bonding with a mate. One consequence of pathway differences underlying similar behaviors is that the same perturbations of neurochemicals or environmental factors can have opposing effects in males and females[39–41]. Recent work on parental behavior reveals that some pathways contribute similarly to both maternal and paternal behaviors in mice[42,43], while other circuits and neuropeptide expression patterns differ in important ways[44]. Perhaps a more important view is that studies assessing mechanisms underlying behavior in both sexes are so rare that we often have little idea of the magnitude and relevance of differences[45]. There are likely many more sex-specific differences in fundamental physiological processes we have missed.

Another concern is that preclinical research on sex differences may not be beneficial enough to humans to merit study, as some differences between men and women may be based on factors related more to culture and gender than to biological sex[9]. The concern here is that if there is a statistical or conceptual cost to including females, there needs to be a clear benefit[9]. While others have discussed benefits at length[5], this article seeks to mitigate the issue of cost. And many basic sex differences have already been discovered in humans. For instance, rodent research on the sex differences in dopamine fiber densities and modulation

of dopamine circuitry by estradiol[46] led to the discovery of effects of endogenous fluctuation in estradiol on prefrontal cortex-dependent cognitive tasks in women[47]. The role of melanocortin receptor 1 in pain processing in female (but not male) mice was recapitulated in humans, in whom only females with melanocortin 1 receptors exhibit altered pain processing[2].

The discovery of differences such as these, despite limitations in the ability to assess mechanistic variation in humans *in vivo*, suggests that many more human sex differences in underlying mechanisms will ultimately be detected. While there is no guarantee that specific animal research findings or specific sex differences will translate to humans, this can be improved by conducting comparative research across species, as well as corresponding research in humans. The unacceptable alternative is that we adopt the rodent equivalent of considering “man” to stand for “human.”

ACKNOWLEDGEMENTS

I am grateful to David Soergel, Bob Calin-Jageman, and Mary Harrington for feedback on the statistical simulations presented in this manuscript, to Irv Zucker for helpful commentary on the manuscript, and to Brian Prendergast, Kenneth Onishi, and Irv Zucker for material in figure 1B. Funding: this work was supported by the National Institutes of Health [award R15MH113085].

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Box 1.**Including females does not necessitate larger sample sizes**

Some researchers are concerned that mixing males and females in fixed-sized groups will result in increased group variance, reduced power, and the need to test more animals (e.g. Fields, 2014). However, statistical methods appropriate to the analysis of multiple factors result in little to no loss of statistical power (Collins et al. 2009, 2014), except when there is a sizable interaction between sex and treatment. In that case, it is especially important to initially study both sexes anyway, and measurement of the interaction effect may indicate that further, sex-specific analysis or studies are needed.

For illustration, I present the statistical outcomes of 2-group and factorial tests using simulated data. Consider an experiment with two treatments (call them “A” and “B”, e.g. hot vs. cold room temperature) and an outcome measure (e.g. distance traveled). Below are results of tests using single sex samples—12 males (m) or 12 females (f), or samples with 6f and 6m—in the presence and absence of sex differences and interaction effects. In each scenario there is a $\pm 5\%$ change in mean and SD in females and males.

In scenario 1 there is no sex difference between males and females (figure 2a). This is a common result, and in this case there is no cost to mixing sexes. All analysis methods yield equivalent effects of treatment, and 2-way ANOVA on sex and treatment indicates no interaction effect. In scenario 2, there is a large sex difference and a moderate treatment difference (figure 2b). This is the oft-feared scenario in which simply pooling males and females reduces statistical power. Whereas loss of power occurs when a *t*-test is used to compare across treatments, 2-way ANOVA results in no loss of power to detect treatment effects, as the test quantifies treatment differences relative to the mean of each sub-group. Finally, scenario 3 represents a possible “worst-case scenario” where there is a large treatment effect in females and an equally sizable *but opposite* effect in males. Pooling males and females results in the eradication of a treatment difference, but the ANOVA interaction effect will very likely be significant, and signals that sex-specific follow-up is strongly indicated.

Although the factorial approach is powerful, it is not without potential weaknesses. ANOVA on sex*treatment generates 3 F values at $\alpha = 0.05$. This leads to a higher collective type I error rate than one *t*-test (explaining why the “treatment” factor performs as well in ANOVA as the *t*-test in scenario

1). This is important to keep in mind if additional factors are added. Also, while scenario 3 is extreme, when interaction effects are more intermediate, they will be less easily detected. If assessing sex differences is a primary rather than secondary goal, increased sample size will improve detection of interactions. Simulation details are described in the figure caption below. Specific parameters used as well as the R code that generates the simulated data and graphs are available at [*insert OSF url here upon article acceptance*].

Highlights

- Female subjects are underrepresented in animal research
- Recent studies demonstrate that unstaged female rodents are no more variable than males
- Factorial designs allow analysis of both sexes without necessitating increased sample sizes
- Sex differences should be presented with information on effect size

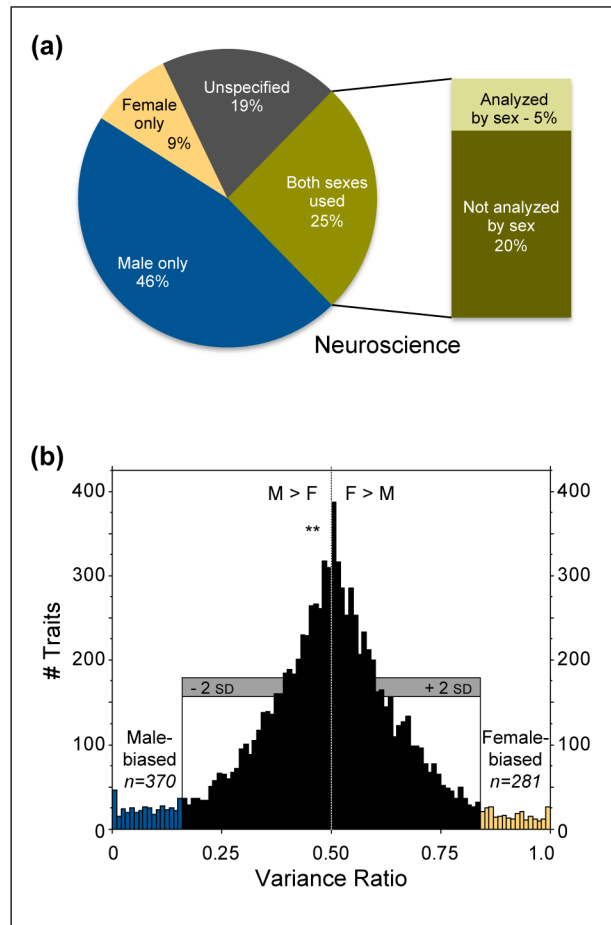


Figure 1.

Data on sex bias and trait variability. A. Inclusion of male and female animal research subjects was surveyed across 10 biological disciplines (neuroscience shown here). Even when both sexes were used, analyses typically did not consider subject sex. Analysis of data from [1]. B. Coefficients of variation were assessed in male and female mice across >9,900 measurements of traits. Variability was similar in males and females, with more male-biased than female-biased traits, and a mean variance ratio significantly lower than 0.5. Modified (with permission) from [22].

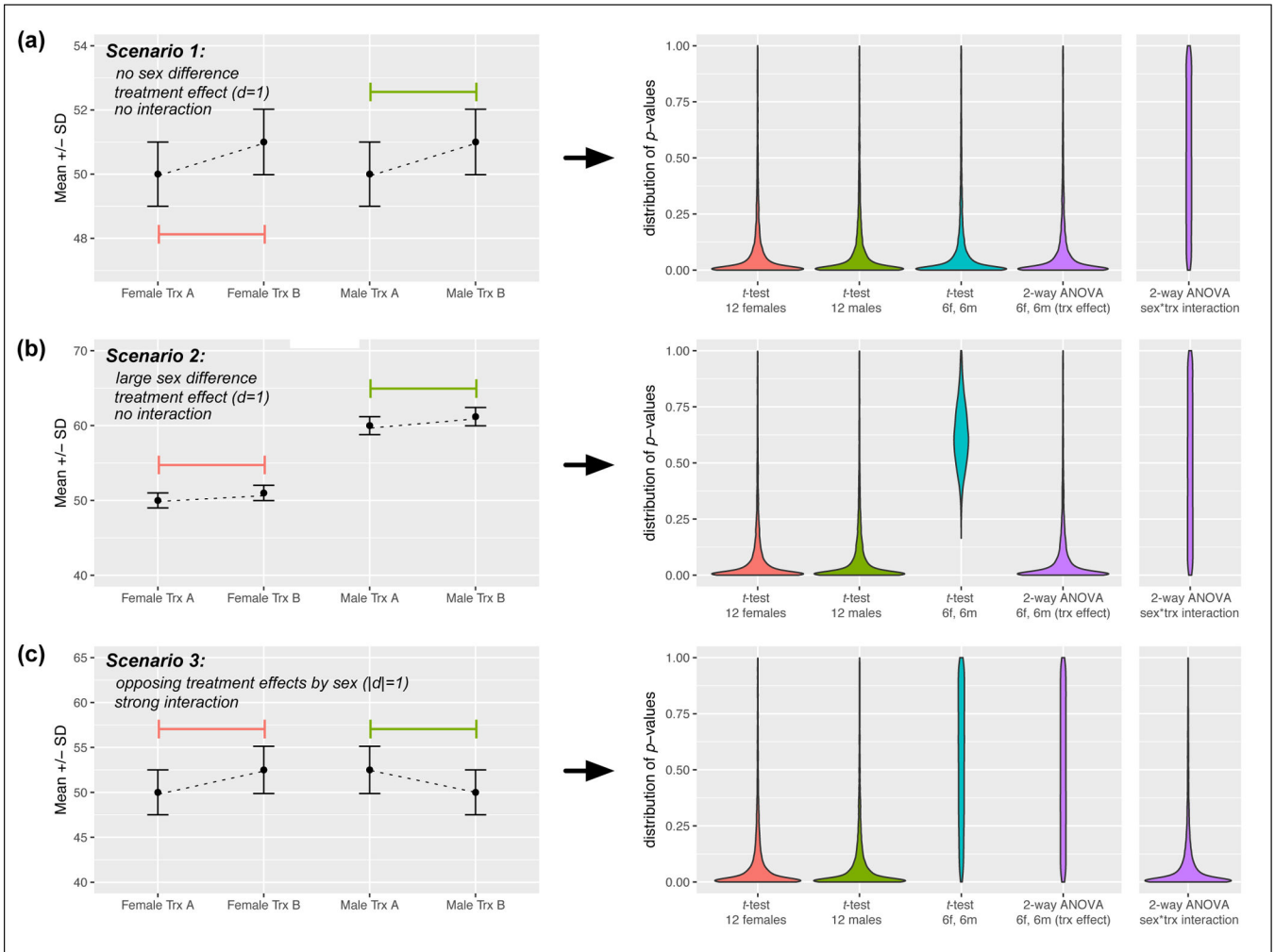


Figure 2. Simulated p -value distributions for different group compositions and treatment effects. **Left panels:** mean and standard deviation of each group, used to generate 10,000 possible samples of subjects (12f, 12m, or 6f and 6m) in each treatment, according to a gaussian distribution. Coefficients of variation (SD/mean) were constant for each sex/treatment combination so that the spread of groups with different means would be equivalent. Effect sizes for comparisons of treatment A vs. B in all male or female groups were matched (Cohen’s $d=1$ using SD of lower group, or .97-.99 using pooled SD). **Right panels:** violin plots of p -values generated for t -tests between different kinds of groups in treatments A vs. B (first three datasets), or from the treatment factor from 2-way ANOVA on mixed sex groups (purple plot). ANOVAs were run with sex and treatment as factors, and an interaction term. The fifth plot (also in purple) represents the distribution of p -values of the ANOVA’s interaction term across runs. Even with a large sex difference, there is no loss from testing half males and half females when a factorial analysis is used, as long as there is no interaction. When an interaction is present, factorial analysis cannot detect a unified

treatment effect, but the strong interaction effect will indicate that subgroup analysis by sex and possible follow-up experiments are merited.

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