UCSF UC San Francisco Previously Published Works

Title

A Comprehensive Review of Immunoreceptor Regulation of Osteoclasts

Permalink https://escholarship.org/uc/item/5z43q7bg

Journal Clinical Reviews in Allergy & Immunology, 51(1)

ISSN 1080-0549

Authors Humphrey, Mary Beth Nakamura, Mary C

Publication Date 2016-08-01

DOI 10.1007/s12016-015-8521-8

Peer reviewed



HHS Public Access

Author manuscript *Clin Rev Allergy Immunol*. Author manuscript.

A Comprehensive Review of Immunoreceptor Regulation of Osteoclasts

Mary Beth Humphrey and

Department of Medicine, Division of Rheumatology, Immunology, and Allergy, University of Oklahoma Health Sciences Center, 975 NE 10th St., BRC209, Oklahoma City, OK 73104, Office: 405-271-8001 x35290, marybeth-humphrey@ouhsc.edu

Mary C. Nakamura^{*}

Department of Medicine, Division of Rheumatology, University of California, San Francisco, Arthritis/Immunology Section, San Francisco Veterans Administration Medical Center, 4150 Clement St 111R San Francisco, CA 94121

Abstract

Osteoclasts require coordinated co-stimulation by several signaling pathways to initiate and regulate their cellular differentiation. RANKL (Receptor Activator for NFkB ligand or TNFSF25), a TNF (tumor necrosis factor) superfamily member, is the master cytokine required for osteoclastogenesis with essential co-stimulatory signals mediated by ITAM (immunoreceptor tyrosine-based activation motif)-signaling adaptors, DAP12 (DNAX associated protein 12kD size) and FcRy (FccR1 gamma chain). The ITAM-signaling adaptors do not have an extracellular ligand-binding domain, and therefore must pair with ligand-binding immunoreceptors to interact with their extracellular environment. DAP12 pairs with a number of different immunoreceptors including TREM-2 (Triggering Receptor Expressed on Myeloid Cells 2), MDL-1 (Myeloid Dap-12 associated Lectin) and Siglec-15 (Sialic acid-binding immunoglobulin-type lectin15), while FcRy pairs with a different set of receptors including OSCAR (Osteoclast Specific Activating Receptor), PIR-A (Paired Immunoglobulin Receptor A), and Fc Receptors. The ligands for many of these receptor in the bone microenvironment remain unknown. Here we will review immunoreceptors known to pair with either DAP12 or FcRy that have been shown to regulate osteoclastogenesis, However, co-stimulation and the effects of ITAM-signaling have turned out to be complex including paradoxical findings that ITAM-signaling adaptor-associated receptors can inhibit osteoclastogenesis and ITIM (immunoreceptor tyrosine-based inhibitory motif) receptors can promote osteoclastogenesis. Thus, co-stimulation of osteoclastogenesis continues to reveal additional complexities that are important in the regulatory mechanisms that seek to maintain bone homeostasis.

Keywords

Osteoclast; Immunoreceptor; ITAM; Fc Receptor; RANK; Osteoimmunology

^{*} corresponding author Phone: 415-750-2104, Mary.nakamura@ucsf.edu.

Mary Beth Humphrey and Mary C. Nakamura declare that they have no conflict of interest.

Immunoreceptor Regulation of Osteoclasts

Osteoclast differentiation is a highly regulated process, meant to ensure that osteoclasts are formed on bone surfaces and specifically respond to bone remodeling stimuli (1, 2). Osteoclast differentiation and activity must be closely coupled to processes of bone formation and require cell-cell communication through both intercellular interactions and soluble cytokines (3). Like many immune cells, osteoclasts require coordinated costimulation by several signaling pathways to initiate cellular differentiation at defined places and times. RANKL (Receptor Activator for NFkB ligand or TNFSF25), a TNF (tumor necrosis factor) superfamily member, is the essential cytokine required for osteoclastogenesis (4). The critical co-stimulatory signals are directed by immunoreceptors that signal through ITAM (immunoreceptor tyrosine-based activation motif)-signaling adaptors, DAP12 (DNAX associated protein 12kD size) and FcRy (Fc[.epsilon]R1 gamma chain) (5, 6). ITAM-signaling adaptors initiate a calcium flux that is necessary for NFATc1 activation to drive the differentiation process (7). NFATc1 is known as the master transcriptional regulator required for osteoclastogenesis (7). While a number of ITAMsignaling adaptor-associated receptors have been identified as regulators of osteoclastogenesis, the exact roles of each of these receptors in basal or pathological bone remodeling remain incompletely understood (8). In addition, at specific sites and/or conditions, osteoclastogenesis can be initiated in the absence of the ITAM signaling adaptors suggesting that alternative costimulatory pathways can be utilized (9).

The role of these immunoreceptors in innate immune cells is primarily to enable or limit the response of cells to microenvironmental change (10). However in the bone environment we have limited understanding of the receptors utilized by osteoclasts to respond and the ligands that are sensed. Here we review immunoreceptors that have been identified to activate or inhibit osteoclastogenesis and discuss their effects on bone homeostasis. While the list of potential costimulatory receptors is likely not complete, we have learned that for osteoclasts, like other innate cells, it is the combined input of arrays of receptors that fine tunes the differentiation and function of these unique cells to carry out bone resorption.

Costimulation by ITAM-signaling adaptor-associated immunoreceptors

The evidence that RANK requires a co-stimulatory signal to initiate osteoclastogenesis was first demonstrated by examining mice deficient in DAP12 and the FcR γ chains (5, 6). These ITAM-signaling adaptors present in myeloid cells are known to contain ITAMs (immunoreceptor tyrosine based activation motifs) that initiate a tyrosine kinase based signaling cascade. The finding that mice deficient in both ITAM-signaling adaptors are severely osteopetrotic despite the presence of intact RANK and MCSF (macrophage colony stimulating factor) signaling pathways serves as the critical evidence for the requirement for ITAM costimulation during osteoclastogenesis. Mice deficient in both the DAP12 and the FcR γ ITAM-signaling adaptors are unable to form osteoclasts *in vitro* or *in vivo* (5, 6). Koga et. al. proposed the model of co-stimulation in osteoclastogenesis in which ITAM signals are required to cooperate with RANK signals to induce the master transcription factor for osteoclastogenesis, NFATc1 (5, 7).

The ITAM-signaling adaptors do not have an extracellular ligand-binding domain, and are known to pair with a number of different receptors through paired charged residues in their transmembrane domains (11) (Figure 1). Several immunoreceptors that pair with either DAP12 or FcR γ have been identified as costimulatory molecules for RANKL and while they share the ability to stimulate osteoclastogenesis, their expression is differentially regulated and responsive to distinct stimuli (2, 12). Ligands identified for some of these receptors are also differentially regulated, though most stimulatory ligands in the bone microenvironment are not well understood. In combination, these regulatory mechanisms serve to locally modulate osteoclastogenesis and maintain bone homeostasis. ITAM-signaling adaptorassociated receptors in immune cells often activate cellular differentiation or function which can be inhibited by related receptors with ITIM-signaling (immunoreceptors tyrosine-based inhibitory motif) domains. It was initially thought that the balance between ITAM and ITIM receptor expression might be the primary means to regulate co-stimulation during osteoclastogenesis, However, co-stimulation and the effects of ITAM and ITIM-signaling have turned out to be much more complex.

DAP12-associated receptors

DAP12 was originally identified in NK cells and some lymphocytes, was found to be expressed in myeloid cells, including osteoclast precursors, and became of significant interest in osteoclasts due to the consequences of DAP12 loss-of-function mutations in humans (13). The rare disorder of Nasu-Hakola disease or PLOSL (Polycystic lipomembranous osteodysplasia with sclerosing leukoencephalopathy) is a recessive genetic disorder characterized by rapidly progressive dementia with large bony cysts, osteopenia and early degenerative arthritis in the wrists and ankles. Patients with PLOSL have either functional deficiency of DAP12 or a DAP12-associated receptor, TREM-2 (Triggering receptor expressed on myeloid cells-2) (13). Peripheral blood mononuclear cells isolated from PLOSL patients (DAP12- or TREM2-deficient humans) fail to differentiate normally into osteoclasts when stimulated with RANKL and MCSF (14, 15). Instead cells from PLOSL patients show inefficient and delayed differentiation of osteoclasts with a remarkably reduced bone resorption capability in vitro (14, 15). DAP12^{-/-} mice show a mild increase in bone mass in vivo while DAP12^{-/-} OC (osteoclasts) have defects in cytoskeletal organization with impaired formation of multinucleated OC and impaired resorption (16-20). Interestingly, DAP12^{-/-} mice have recently been shown to have impaired fracture healing, suggesting a role for DAP12 in the inflammatory and bone regeneration response of fracture repair (21). DAP12 pairs with a number of different immunoreceptors in myeloid cells and a few of these individual receptors have been examined in osteoclastogenesis. While TREM2 is the DAP12-associated receptor of importance in the PLOSL phenotype, it is not the only DAP12-associated receptor that regulates osteoclastogenesis.

TREM 2

Due to its known role in PLOSL, TREM2 was the first DAP12-associated receptor examined in osteoclasts. TREM2 had also been studied in other immune cells and interestingly plays dual roles in immune responses. While TREM2 stimulates dendritic cell differentiation (22), it also inhibits inflammatory cytokine production by macrophages and

dendritic cells in response to TLR (toll like receptor) stimulation by LPS (lipopolysaccharide) or CpG DNA (23, 24). TREM2 crosslinking on mouse osteoclasts inhibits resorption while promoting multinucleation, migration, and osteoclast differentiation (25). While TREM2 crosslinking on mouse osteoclasts leads to syk phosphorylation and osteoclast differentiation (25), TREM2 crosslinking also to lead to association of the SHIP1 (SH2-domain inositol phosphatase) with DAP12, which leads to inhibitory signals (26). TREM2 also associates with DAP10 leading to strong activation of PI3K (26). Thus, the TREM2 receptor can activate and inhibit osteoclastogenesis (25, 27). The dual action of the TREM2–DAP12 complex may be regulated by differing TREM2 ligands, differing strength of ligand signals or combinations of different costimulatory signals, but this is not yet delineated (27).

The ligand specificity of TREM2 has been a topic of considerable interest and a number of studies suggest that TREM2 functions as a pattern recognition receptor, able to bind to a variety of anionic ligands including Gram-negative and -positive bacteria, heparin and dextran sulfate, HSP60 and apoptotic cells (28-31). TREM2 has also been shown to associate with neighboring Plexin-A1 receptors and interaction of Plexin-A1 with its ligand, Semaphorin 6D, leads to phosphorylation of the associated DAP12 ITAM (32). TREM2 expression is upregulated by RANKL stimulation but downregulated by LPS and IL-10 (25, 28, 33). TREM2 is highly upregulated in active RA (rheumatoid arthritis) synovium and subsequently downregulated in inactive RA suggesting a role in RA-induced inflammation (34). Human TREM2-deficient osteoclasts fail to multinucleate in vitro (15), however TREM2-deficient mouse osteoclasts show accelerated osteoclastogenesis in vitro due to excess M-CSF induced activation of β-catenin that limits pre-osteoclast proliferation and diverts development towards osteoclasts (35). Despite intensive investigation around TREM2, a number of questions remain unanswered, particularly with regard to the roles for TREM2 in bone. Differences between the mouse and human phenotype in TREM2 deficiency are incompletely explained and the relevant ligands in the bone microenvironment are not clearly identified. In other immune cells, TREM2 plays a role in resolving inflammatory responses and recognition of apoptotic cells, processes that could also be of importance in maintaining bone homeostasis.

MDL-1

MDL-1, (myeloid DAP12 associated lectin or CLEC5A) is a C-type lectin DAP12associated receptor, but in contrast to TREM2, stimulates myeloid cell–associated inflammatory responses (36). MDL-1 associates with signaling adaptors DAP12 and DAP10 (DNAX associated protein 12 kD or 10kD) and is most highly expressed in activated myeloid cells (37). MDL-1 is potently induced by TNF α and IL-23 (38, 39). MDL-1 stimulates osteoclastogenesis *in vitro*, and MDL knockdown in osteoclast precursor cells inhibits osteoclastogenesis. Studies in MDL-1^{-/-} mice, demonstrate that MDL-1 plays a significant role in inflammatory arthritis suggesting that MDL-1 promotes inflammatory bone loss (38). Treatment with an agonist anti-MDL-1 antibody increased both disease incidence and severity in a murine collagen antibody induced arthritis model (CAIA) and in a CIA (collagen induced arthritis) model. CAIA is diminished in severity and incidence in both DAP12^{-/-} and MDL-1^{-/-} animals indicating that the receptor and signaling adaptor are

required to promote bone loss. Blockade of MDL-1 function with a soluble receptor fusion protein (MDL-1-Ig) induces resistance to CAIA or CIA induced arthritis (38). Importantly, MDL-1-Ig treatment down-regulates TRAP, cathepsin K, and MMP9 expression, reducing both clinical signs of autoimmune joint inflammation, and bone destruction (38). These studies suggest that MDL-1 is an important regulator of osteoclastogenesis during inflammation.

Ligands important for activating MDL-1 in inflammatory arthritis are not known. The only identified MDL-1 ligand is a Dengue virus capsid protein (40). Interestingly, Dengue hemorrhagic fever is also known as 'break bone' fever, thus dengue might regulate osteoclastogenesis through MDL-1 which may contribute to osseous symptoms. MDL-1 does regulate the intense inflammatory response to dengue fever because blockade of MDL-1 inhibits the cytokine storm and vascular leakage in a mouse model of dengue fever (40). In RA (rheumatoid arthritis) patients, the frequencies of MDL-1-expressing monocytes and levels of MDL-1 and DAP12 gene expression were significantly increased in active RA patients compared to inactive RA pts or healthy controls (41). Interestingly, the frequency of MDL-1 expression also significantly decreased after effective RA therapy. Concordant overexpression of MDL-1 and DAP12 correlated with increased production of proinflammatory cytokines in RA patients, suggesting that MDL-1 plays a role in regulating articular inflammation as well as osteoclastogenesis in inflammatory conditions (41, 42).

Siglec-15

Siglec-15 (sialic acid-binding immunoglobulin-like lectin 15) is one of a family of receptors that binds to sialylated O-linked sugars. Siglec-15 binds to cells expressing the sialic acid motif Neu5Acalpha2-6GalNAcalpha- structure (43). Siglec-15, initially identified as a markedly overexpressed gene in giant cell tumor of the bone, was subsequently investigated for its role in osteoclastogenesis. Other Siglecs associate with DAP12 and their expression or function have not yet been examined in osteoclasts. Siglec-15 is the only Siglec highly conserved throughout vertebrate evolution (44). Siglec-15 is upregulated in differentiated osteoclasts and anti-Siglec-15 polyclonal antibody inhibits human and mouse osteoclast differentiation *in vitro* (45). Siglec- $15^{-/-}$ mice have mild osteopetrosis with increased trabecular bone mass in the lumbar vertebrae and metaphyseal regions of the femur and tibia. Interestingly, osteoclast numbers are unchanged but a systemic bone resorption marker, urinary deoxypyridinoline, is decreased in Siglec- $15^{-/-}$ mice, suggesting that the increased bone mass in the mutant mice is due to a functional osteoclast defect (46). Indeed, Siglec15^{-/-} osteoclasts have impaired functional resorption in vitro (46). The osteoclast defect is more prominent in the secondary spongiosa, consistent with a localized effect, which is also seen in an antigen induced arthritis model in Siglec $15^{-/-}$ mice (47). In murine antigen-induced arthritis, the degree of periarticular bone loss in the proximal tibia is significantly lower in Siglec-15^{-/-} mice compared to WT mice, however the degree of joint inflammation, and cartilage and subchondral bone destruction is comparable (48). Interestingly, TNFa mediated induction of TRAP-positive multinucleated cells is also impaired in Siglec- $15^{-/-}$ cells and Siglec- $15^{-/-}$ mice are resistant to estrogen deficiencyinduced bone loss compared to WT mice suggesting that Siglec-15 blockade may be useful as a therapeutic strategy (49).

Other DAP12 associated receptors

DAP12 associated receptors such as TREM1, TREM3, SIRP β 1, PILRB, CD200R3, CD200R4, CD300B, CD300LE, and MAIRII are known to be present on myeloid cells but their roles have not been examined or demonstrated in osteoclasts (50). However, DAP12 is known to cooperate with non-immunoreceptors in osteoclasts particularly $\alpha_v\beta_3$ integrin and c-Fms receptor to regulate the osteoclast cytoskeleton, adhesion and proliferation. Integrin β_3 stimulation on osteoclasts leads to activation of syk and src associated with DAP12 during cytoskeletal changes in bone resorption and actin ring formation (51). Similarly, M-CSF stimulation of c-Fms leads to c-Src-induced phosphorylation of DAP12 with recruitment of syk to the DAP12 ITAM domain and activation of Vav leading to osteoclast spreading (17, 52). DAP12^{-/-} osteoclastogenesis is partially rescued by high-dose M-CSF *in vitro* but fails to correct the abnormal cytoskeleton formation (52). Thus DAP12 can participate in intracellular signaling initiated by several different types of ligand/receptor interactions in addition to the immunoreceptors that pair specifically with DAP12.

Fcgamma associated Receptors

The FceR1 gamma chain (FcR γ) is similar to DAP12 in that it contains an ITAM domain and no extracellular domain. FcRy also pairs with ligand-binding receptors through interactions in the transmembrane domains of the ITAM-signaling adaptor and the immunoreceptor. FcR γ -associated receptors are a distinct group from receptors that pair with DAP12 (8). The FcR $\gamma^{-/-}$ mouse does not have a bone phenotype at baseline and forms normal osteoclasts in vivo and in vitro (6). However when combined with DAP12 deficiency, the resulting severe osteoclast deficiency and osteopetrosis suggests that FcR γ chain associated receptors are essential in the absence of DAP12 (5, 6). A recent study of mice with combined deficiency of DAP12 and the beta3 integrin showed osteopetrosis and osteoclast deficiency similar to the combined DAP12/FcRy chain deficiency. Further studies demonstrated that beta 3 integrin requires FcRy association to function in the absence of DAP12 (51). FcRy, named for its association with receptors binding to the Fc portion of immunoglobulin molecules, also pairs with a number of additional immunoreceptors. Some FcRy chain associated receptors identified in other myeloid cells such as DCAR (dendritic cell activating receptor) and ILT/LILR (Immunoglobulin like transcript/ Leukocyte Immunoglobulin-like receptor) have yet to be examined in osteoclasts.

OSCAR

OSCAR (osteoclast activating receptor) is an FcR γ -associated receptor that is a member of the leukocyte receptor complex protein family (53). OSCAR was named for the original finding that it is upregulated during osteoclast differentiation, however subsequent studies have revealed that OSCAR expression is not specific to osteoclasts (54). OSCAR is one of the few ITAM-signaling adaptor-associated receptors shown to have a functional ligand in the bone microenvironment. OSCAR binding sites are found in Collagen I, II and III and recognition of exposed extracellular matrix (ECM) ligands have been shown to mediate costimulation of osteoclastogenesis dependent on FcR γ (55). Thus, OSCAR is a collagen receptor and it can provide costimulatory signals for osteoclast differentiation in DAP12^{-/-} cells (55). *In vivo*, OSCAR deficiency alone fails to alter bone mass, which is not surprising

given that absence of its associated signaling chain, FcR γ , in mice also fails to induce a change in bone phenotype (5, 6, 55). OSCAR gene expression is coordinately regulated by the osteoclastogenic transcription factors PU.1/MITF and is further enhanced by RANK induced MKK6/p38 signals. Thus, while OSCAR can stimulate osteoclast differentiation, it is also upregulated during osteoclast differentiation (56).

OSCAR expression has been associated with both bone loss and inflammation in human disease. A single-nucleotide polymorphism within the promoter of the OSCAR gene has been linked to increased risk of postmenopausal osteoporosis (57). In rheumatoid arthritis (RA), OSCAR expression in monocytes is correlated with disease activity and monocytes with high OSCAR expression exhibited an enhanced potential to differentiate into osteoclasts (58). TNF α is the main inducer of OSCAR expression in monocytes (58). Serum levels of OSCAR are reduced in RA patients compared with healthy controls suggesting that expression and secretion or cleavage of OSCAR to a soluble form is immune modulated (59). Supporting this idea, OSCAR transcript expression and soluble OSCAR release is stimulated by TNF α and IL1- β in cultured endothelial cells. OSCAR is strongly expressed in the microvasculature of active synovial tissue in RA (42). OSCAR has also been demonstrated to be a receptor for surfactant protein D, and the interaction can activate TNF α release from human CCR2⁺ inflammatory monocytes (60). OSCAR expression is also regulated by oxidized low-density lipoprotein, thus OSCAR has been suggested to play a role in oxidative stress-mediated atherogenesis as well as monocyte adhesion (61).

PIR-A

PIR-A (paired immunoglobulin-like receptor A) is an FcR γ associated receptor of the immunoglobulin superfamily present in mice, whose ligands are known to be MHC Class I (major histocompatibility complex class I) molecules. TNF α induces the expression of PIR-A on osteoclasts and PIR-A ligands on osteoblasts and these interactions contribute to TNF α -induced osteoclastogenesis and TNF α -induced bone loss in vivo (62). Transgenic expression of TNF α in mice deficient in FcR γ , leads to inflammation but not bone erosion or osteoporosis suggesting that one or more FcRy-associated receptors is required for TNF α induced bone loss. TNFa induces PIR-A upregulation but not other costimulatory receptors such as OSCAR, TREM2, and SIRP β (62). Mice deficient in expression of PIR-A ligands (β2-microglobulin deficient mice lacking MHC Class I expression) also showed abrogation of bone loss with transgenic TNFa exposure. PIR-A-Fc blocks enhanced TNFa-induced osteoclastogenesis in vitro suggesting that PIR-A is required for enhanced $TNF\alpha$ -induced osteoclastogenesis (62). PIR-A receptors are only present in mice, however there are human orthologues in the ILT/LILR (immunoglobulin-like transcript, leukocyte immunoglobulinlike receptor) receptor families that respond to MHC Class I ligands as well (63). Additional studies are needed to determine whether the FcR γ -associated receptors in these receptor families regulate human osteoclasts.

Fc Receptors

Fc Receptors (FcR) are capable of binding to and responding to the Fc domain of serum immunoglobulin and immune complexes. Fc γ RI, Fc γ RIII and Fc γ RIV are all Fc Receptors that utilize the FcR γ chain and their interactions with immune complexes have been

speculated to promote osteoclastogenesis. Fc Receptors could have a direct effect on osteoclast precursors or an indirect effect due to release of proinflammatory cytokines and chemokines upon stimulation of Fc Receptor-bearing immune cells. The role of autoantibodies and immune complexes in regulating osteoclastogenesis has been difficult to sort out, and is further complicated because both activating and inhibitory Fc receptors are expressed on osteoclast precursors. Fc γ RIIB is an ITIM-signaling receptor and generally inhibits signaling by Fc γ RI, Fc γ RIII or Fc γ RIV (Figure 2A). It was suspected that the relative expression of different activating FcRs (Fc γ RI/III/IV) and inhibitory FcR (Fc γ RIIB) on osteoclast precursors would determines osteoclastogenesis in response to immune complex stimulation.

Recent studies have dissected the roles of individual Fc receptors more carefully and demonstrated several interesting findings. Under basal conditions, osteoclast precursors express Fc γ RIIB and Fc γ RIII (64). Fc γ RIII is well known to respond to immune complexes during infections. As an activating receptor that pairs with the FcR γ chain, it might be expected that Fc γ RIII would lead to activation of osteoclastogenesis. Interestingly, Fc γ RIII is downregulated with RANKL stimulation during osteoclastogenesis and overexpression of Fc γ RIII actually inhibits osteoclastogenesis (64). Fc γ RIII is actually a negative regulator of osteoclastogenesis. Consistent with this finding, inhibition of osteoclastogenesis by immune complexes was observed in Fc γ RIIB^{-/-} precursors but not FcR $\gamma^{-/-}$ precursors suggesting that the inhibition was mediated by an activating ITAM-signaling adapter-associated receptor (65). The mechanism behind the inhibition is thought to be that the presence of Fc γ RIII sequesters the FcR γ chain away from other osteoclastogenic Ig-like receptors (such as PIR-A and OSCAR) preventing them from associating with FcR γ to promote osteoclastogenesis (64) (Figure 2B).

The other ITAM-signaling adapter-associated Fc receptors, FcyRI and FcyRIV, are induced under inflammatory conditions and IgG2 immune complexes binding to these receptors can activate osteoclastogenesis. FcyRIV is also upregulated with RANKL stimulation and crosslinking of FcyRIV enhances osteoclast differentiation (64) (Figure 2C). FcyRIV^{-/-} mice and mice with a targeted deletion of FcyRIV limited to osteoclasts (Cathepsin K-Cre) are protected from autoantibody-dependent bone destruction in a serum transfer arthritis model and have lower numbers of osteoclasts in the inflamed joints (64). Mice with an osteoclast specific deletion of FcyRIV showed decreased bone destruction despite persistence of inflammation, suggesting that FcyRIV on osteoclasts is critical for bone destruction during antibody-dependent inflammatory arthritis in mice (64). Another group attempted to modify these interactions therapeutically by using a staphylococcal protein A complex (SPA) which binds to circulating IgG and generates small immunoglobulin complexes that interact with monocytes, macrophages, and preosteoclasts. Formation of these complexes resulted in polarization of macrophages to a regulatory phenotype and directly inhibited differentiation of preosteoclasts into osteoclasts (66). Administration of SPA in the early stages of disease substantially alleviated the clinical and histologic erosive features of CIA in mice and though it was not determined which FcR was responsible for the effects, the study demonstrates that specific types of immune complexes can be utilized to downregulate inflammation and osteoclastogenesis (66). It should be noted that FcyRIV is not present in

humans and human cells have a unique Fc receptor, FcRIIA that is a single chain receptor with a cytoplasmic ITAM domain. Thus ITAM-signaling receptors that interact with the Fc domains of immunoglobulin differ between mice and humans.

The presence of autoantibodies against citrullinated peptides (ACPA) highly correlates with development of bone loss and erosive disease in patients with rheumatoid arthritis. In vitro, affinity-purified human autoantibodies against mutated citrullinated vimentin (MCV) bind to osteoclasts, stimulate osteoclastogenesis and bone resorption (67). Adoptive transfer of these purified human MCV autoantibodies into mice induces increased osteoclastogenesis and osteopenia (67). Interestingly, desialylated, but not sialylated, immune complexes enhance osteoclastogenesis in vitro and in vivo (68). IgG sialylation is of importance in determining the engagement of pro- or anti-inflammatory Fcy receptors (FcyR) and the type of resulting immune response (69). Recently in RA patients, it was shown that the sialylation state of random IgG and specific anti-IgG autoantibodies correlates with bone volume and trabecular number (68). A direct demonstration of desialyated antibodies in vivo is seen when desialyated or sialylated immune complexes are directly injected into a knee joint of mice. Injection with desialyated immune complexes increases osteoclast number and surface area compared with inject of sialyated immune complexes. However, addition of $TNF\alpha$ to the antibody injection eliminates the differential effect due to immune complex sialylation (68). Treatment of mice with the sialic acid precursor N-acetylmannosamine (ManNAc), which results in increased IgG sialylation in vivo, renders mice less susceptible to development of CIA (68), suggesting that modification of sialyation may be therapeutically useful. ACPA are reported to show less sialyation than random IgG consistent with their proosteoclastogenic capacity (70).

ITIM-signaling Receptors that Positively Regulate Osteoclastogenesis

ITAM domains are defined by a specific amino acid sequence that includes a tyrosine that when phosphorylated leads to recruitment of tyrosine kinases that initiate an activating signaling cascade. Opposing the ITAM signals are signals mediated through receptors with ITIM (immunoreceptor tyrosine based inhibitory motif) domains. ITIM domains are defined by a related amino acid sequence that includes a tyrosine that when phosphorylated leads to recruitment of specific phosphatases that can inhibit cellular activation (71). While some ITIM-signaling receptors have been demonstrated to inhibit osteoclastogenesis as expected, the following ITIM-signaling receptors have been demonstrated to paradoxically promote osteoclastogenesis.

Ly49Q

The Ly49 family of C-type lectin receptors that includes ITIM receptors and DAP12 associated receptors that are primarily expressed on NK (natural killer) cells and interact with Class I MHC ligands (72). Ly49Q is an ITIM receptor present on mouse myeloid cells. Surprisingly, knockdown or deletion of Ly49Q resulted in decreased *in vitro* osteoclastogenesis (72). Ly49Q is upregulated with RANKL stimulation and mechanistically, Ly49Q has been shown to sequester SHP-1 (SH2 domain hematopoietic phosphatase-1) away from PIR-B (paired immunologobulin-like receptor B), a different

ITIM containing receptor, to enable osteoclastogenesis to proceed. Ly49Q deficient mice do not demonstrate a significant bone phenotype *in vivo* (72).

DC-STAMP

DC-STAMP (dendritic cell-specific transmembrane protein), despite its name, is expressed in osteoclasts and has been shown to be an important regulator of cell fusion in osteoclasts (73). DC-STAMP was recently shown to be an ITIM-signaling receptor (74). However antibody against DC-STAMP blocks OC formation *in vitro* suggesting DC-STAMP is a positive regulator of osteoclastogenesis. DC-STAMP(high) freshly isolated monocytes produce a higher number of OC in culture than DC-STAMP(low) cells (74). DC-STAMP has been demonstrated to interact with SHP-1 and CD16 (Fc γ RIII) during osteoclastogenesis (74). Perhaps this interaction also sequesters these negative regulators enabling promotion of osteoclastogenesis but this has not directly examined .

PD-1

PD-1 (Programmed cell death protein 1 or CD279) is a cell surface receptor in the immunoglobulin superfamily. Deficiency of PD-1 leads to T cell activation, but surprisingly, the enhanced immune activation does not lead to bone loss. The cytoplasmic tail of PD-1 contains an ITIM domain and an immunoreceptor tyrosine-based switch motif that can link to additional signaling intermediates. PD-1 deficiency in mice leads to reduced osteoclastogenesis and an osteopetrotic phenotype (75). Micro CT analysis of PD-1^{-/-} bones shows significant increases in trabecular and cortical bone mineral densities of the tibia and histomorphometry reveals a 55% decrease in osteoclast number on bone surfaces compared to WT. Bone formation indices are similar in both groups. Cells from PD-1-deficient mice show decreased osteoclastogenesis *in vitro* but do not demonstrate a significant change in functional bone resorption (75). Thus, PD-1 positively regulates osteoclastogenesis, though the exact mechanism has not been demonstrated. Whether or not PD-1 promotes osteoclastogenesis by sequestering SHP-1 or SHIP phosphatases has not been examined.

ITIM-signaling receptors That Inhibit Osteoclastogenesis

Other ITIM-signaling receptors inhibit osteoclastogenesis. The phosphatases SHP-1 and SHIP-1 are known to associate with ITIM domains and both of these phosphatases function as negative regulators of osteoclastogenesis. Mice expressing mutant SHP-1 show marked osteopenia with an increase in bone resorption. SHP-1 deficient mice have increased numbers of osteoclasts *in vitro* with increased bone resorption (76). The SHIP-1 phosphatase also inhibits osteoclastogenesis and SHIP-1 deficient mice are also osteopenic with increased in vitro formation of osteoclasts and increased bone resorption (77).

While less is known about ITIM-signaling receptors in osteoclastogenesis, those that inhibit osteoclastogenesis *in vitro* include: TLT-1s, CLM-1, and PIRB. TLT-1s, is an alternate transcript of TLT-1 (Triggering receptor expressed on myeloid cells like transcript-1), and the only TREM receptor family member with a cytoplasmic ITIM domain (78). TLT-1s is mainly expressed in macrophages and osteoclast precursor cells. Knockdown of TLT-1s strongly enhances the formation and resorption capacity of osteoclasts (78). CLM-1 (CMRF-35 like molecule-1) is expressed on osteoclast precursors and is down-regulated by

stimulation with RANKL. Overexpression of CLM-1 in the osteoclastogenic cell line RAW264.7 inhibits osteoclast differentiation induced by RANKL (79). PIR-B and four isoforms of its human ortholog leukocyte Ig-like receptor LILR-B are present on cultured osteoclast precursor cells of mouse and human origin, respectively, and these ITIMharboring inhibitory receptors constitutively recruit SHP-1 in the presence of RANKL and M-CSF. Both murine PIR-B and the human ortholog LILR-B1 suppress osteoclast development *in vitro*. These receptors are thought to interact with their MHC class I ligands on the same cell surface and then serve as a basal inhibitory stimulus for osteoclastogenesis (80). While these receptors all can inhibit osteoclastogenesis, the exact *in vivo* role they play particularly in pathological states has not been delineated.

FcyRIIB

Fc γ RIIB is an ITIM containing Fc receptor that is highly expressed on osteoclast precursors and serves to chronically inhibit osteoclastogenesis. Fc γ RIIB^{-/-} mice were recently found to have a markedly osteoporotic phenotype with enhanced osteoclastogenesis consistent with initial in vitro observations (64). The enhanced osteoclast formation effect is not a cell autonomous change due to deficiency of Fc γ RIIB, but rather a response to immune complex stimulation enabled by Fc γ RIIB, deficiency. When Fc γ RIIB^{-/-} cells are cultured in the presence of serum from Fc γ RIIB^{-/-} mice, enhanced osteoclastogenesis is observed due to the unusually high levels of immune complexes that stimulate activating FcR γ -associated receptors in the Fc γ RIIB^{-/-} cells. Enhanced osteoclastogenesis is not observed in mice with combined Fc γ RIIB/Fc γ R deficiency confirming the requirement for activating FcR γ associated receptors for the immune complex stimulation of osteoclastogenesis (64).

Bypassing co-stimulation by ITAM-associated immunoreceptors

We note that the concept of ITAM-signaling as strictly activating and ITIM-signaling as strictly inhibitory was overly simplistic and as these many studies have shown, co-stimulation in osteoclastogenesis is much more complex. In addition, while it is often stated that the ITAM-signaling co-receptors are required for osteoclastogenesis, we also note that this does not apply to all situations where osteoclastogenesis occurs. It is clear that there is a significant difference in the phenotype of the DAP12^{-/-}/FcR $\gamma^{-/-}$ mice and the RANK deficient mice (4-6). Lack of RANK signaling leads to a global defect in all osteoclastogenesis, causing osteopetrosis in the long bones and lack of tooth eruption in the jaw. In contrast, while the DAP12^{-/-}/FcR $\gamma^{-/-}$ mice have similar osteopetrosis in the long bones, they show normal tooth eruption (4-6). In addition, despite the lack of osteoclasts in the long bones of DAP12^{-/-}/FcR $\gamma^{-/-}$ mice at baseline, when these mice are estrogen deficient, post-ovariectomy, they are able to generate osteoclasts *in vivo*, with significant bone loss (9). Thus the requirement for specific ITAM-signaling adapters to provide co-stimulation in osteoclastogenesis appears to be microenvironment specific, and can be bypassed in specific locations and pathological conditions.

Recent studies of the transcription factor RBP-J (Recombinant recognition sequence binding protein at the J κ site) demonstrates that deletion of RBP-J rescues the defect in osteoclastogenesis observed in the DAP12^{-/-}/FcR $\gamma^{-/-}$ mice both *in vivo* and *in vitro* (81). RBP-J deficiency allows RANK or TNFR signaling to induce osteoclastogenesis

independent of ITAM-adapter signals. RBP-J deficiency potentiates cell responses to TGF- β by upregulating TGF β R1 and maintaining downstream PLC γ 2/calcium-CaMKK/PYK2 activity in the absence of ITAM-signaling adaptors (81). Whether or not specific other additional co-receptors are also required under these conditions is not yet clear. Nonetheless deficiency of RBP-J results in increased PLC γ 2 expression and activity as well as calcium oscillations that enable NFATc1 activation (81) indicating that osteoclastogenesis continues to require the master terminal regulators of osteoclastogenesis and verifies that pathways that bypass the ITAM-signaling adaptors can initiate the process.

Thus, all osteoclastogenesis is not likely identical, and like other immune cells, differences in co-receptor and/or cytokine stimulation during differentiation likely leads to functional heterogeneity among cells. While all osteoclasts share the function of bone resorption, the combination of enzymatic and acidic mechanisms used by osteoclasts at different sites has been suggested to vary (82), and it remains to be determined if this is a consequence of conditions of differentiation. Much remains to be determined to better delineate these regulatory mechanisms. However the importance of a requirement for co-receptors in osteoclastogenesis is that it provides a mechanism by which osteoclastogenesis can be fine-tuned in response to specific microenvironments, and holds potential for differential regulation of bone resorption in health and disease.

Acknowledgements

We thank T.P. Quinn for helpful suggestions and editing the manuscript. M.B.H. is supported by NIH grant AR064211. M.C.N is supported by a VA Merit Review, NIH grant AG046282, the UCSF-Stanford Arthritis Center of Excellence funded by the Great Western Region of the Arthritis Foundation, the Rheumatology Research Foundation and the Russell/Engleman Rheumatology Research Center.

References

- Mellis DJ, Itzstein C, Helfrich MH, Crockett JC. The skeleton: a multi-functional complex organ: the role of key signalling pathways in osteoclast differentiation and in bone resorption. J Endocrinol. 2011; 211:131–43. [PubMed: 21903860]
- Long CL, Humphrey MB. Osteoimmunology: the expanding role of immunoreceptors in osteoclasts and bone remodeling. Bonekey Rep. 2012; 1 [PubMed: 23789115]
- Wythe SE, Nicolaidou V, Horwood NJ. Cells of the immune system orchestrate changes in bone cell function. Calcif Tissue Int. 2014; 94:98–111. [PubMed: 23912951]
- 4. Boyce BF, Xing L. Biology of RANK, RANKL, and osteoprotegerin. Arthritis Res Ther 9 Suppl. 2007; 1:S1.
- Koga T, Inui M, Inoue K, Kim S, Suematsu A, Kobayashi E, Iwata T, Ohnishi H, Matozaki T, Kodama T, Taniguchi T, Takayanagi H, Takai T. Costimulatory signals mediated by the ITAM motif cooperate with RANKL for bone homeostasis. Nature. 2004; 428:758–63. [PubMed: 15085135]
- 6. Mocsai A, Humphrey MB, Van Ziffle JA, Hu Y, Burghardt A, Spusta SC, Majumdar S, Lanier LL, Lowell CA, Nakamura MC. The immunomodulatory adapter proteins DAP12 and Fc receptor gamma-chain (FcRgamma) regulate development of functional osteoclasts through the Syk tyrosine kinase. Proc Natl Acad Sci U S A. 2004; 101:6158–63. [PubMed: 15073337]
- Negishi-Koga T, Takayanagi H. Ca2+-NFATc1 signaling is an essential axis of osteoclast differentiation. Immunol Rev. 2009; 231:241–56. [PubMed: 19754901]
- Humphrey MB, Lanier LL, Nakamura MC. Role of ITAM-containing adapter proteins and their receptors in the immune system and bone. Immunol Rev. 2005; 208:50–65. [PubMed: 16313340]

- Wu Y, Torchia J, Yao W, Lane NE, Lanier LL, Nakamura MC, Humphrey MB. Bone microenvironment specific roles of ITAM adapter signaling during bone remodeling induced by acute estrogen-deficiency. PLoS One. 2007; 2:e586. [PubMed: 17611621]
- Hamerman JA, Ni M, Killebrew JR, Chu CL, Lowell CA. The expanding roles of ITAM adapters FcRgamma and DAP12 in myeloid cells. Immunol Rev. 2009; 232:42–58. [PubMed: 19909355]
- 11. Lanier LL, Bakker AB. The ITAM-bearing transmembrane adaptor DAP12 in lymphoid and myeloid cell function. Immunol Today. 2000; 21:611–4. [PubMed: 11114420]
- Zawawi MS, Dharmapatni AA, Cantley MD, McHugh KP, Haynes DR, Crotti TN. Regulation of ITAM adaptor molecules and their receptors by inhibition of calcineurin-NFAT signalling during late stage osteoclast differentiation. Biochem Biophys Res Commun. 2012; 427:404–9. [PubMed: 23000414]
- Paloneva J, Kestila M, Wu J, Salminen A, Bohling T, Ruotsalainen V, Hakola P, Bakker AB, Phillips JH, Pekkarinen P, Lanier LL, Timonen T, Peltonen L. Loss-of-function mutations in TYROBP (DAP12) result in a presenile dementia with bone cysts. Nat Genet. 2000; 25:357–61. [PubMed: 10888890]
- Paloneva J, Mandelin J, Kiialainen A, Bohling T, Prudlo J, Hakola P, Haltia M, Konttinen YT, Peltonen L. DAP12/TREM2 deficiency results in impaired osteoclast differentiation and osteoporotic features. J Exp Med. 2003; 198:669–75. [PubMed: 12925681]
- Cella M, Buonsanti C, Strader C, Kondo T, Salmaggi A, Colonna M. Impaired differentiation of osteoclasts in TREM-2-deficient individuals. J Exp Med. 2003; 198:645–51. [PubMed: 12913093]
- Humphrey MB, Ogasawara K, Yao W, Spusta SC, Daws MR, Lane NE, Lanier LL, Nakamura MC. The signaling adapter protein DAP12 regulates multinucleation during osteoclast development. J Bone Miner Res. 2004; 19:224–34. [PubMed: 14969392]
- Zou W, Reeve JL, Liu Y, Teitelbaum SL, Ross FP. DAP12 couples c-Fms activation to the osteoclast cytoskeleton by recruitment of Syk. Mol Cell. 2008; 31:422–31. [PubMed: 18691974]
- Zou W, Zhu T, Craft CS, Broekelmann TJ, Mecham RP, Teitelbaum SL. Cytoskeletal dysfunction dominates in DAP12-deficient osteoclasts. J Cell Sci. 2010; 123:2955–63. [PubMed: 20720152]
- Kaifu T, Nakahara J, Inui M, Mishima K, Momiyama T, Kaji M, Sugahara A, Koito H, Ujike-Asai A, Nakamura A, Kanazawa K, Tan-Takeuchi K, Iwasaki K, Yokoyama WM, Kudo A, Fujiwara M, Asou H, Takai T. Osteopetrosis and thalamic hypomyelinosis with synaptic degeneration in DAP12-deficient mice. J Clin Invest. 2003; 111:323–32. [PubMed: 12569157]
- Nataf S, Anginot A, Vuaillat C, Malaval L, Fodil N, Chereul E, Langlois JB, Dumontel C, Cavillon G, Confavreux C, Mazzorana M, Vico L, Belin MF, Vivier E, Tomasello E, Jurdic P. Brain and bone damage in KARAP/DAP12 loss-of-function mice correlate with alterations in microglia and osteoclast lineages. Am J Pathol. 2005; 166:275–86. [PubMed: 15632019]
- Kamimura M, Mori Y, Sugahara-Tobinai A, Takai T, Itoi E. Impaired Fracture Healing Caused by Deficiency of the Immunoreceptor Adaptor Protein DAP12. PLoS One. 2015; 10:e0128210. [PubMed: 26030755]
- Bouchon A, Hernandez-Munain C, Cella M, Colonna M. A DAP12-mediated pathway regulates expression of CC chemokine receptor 7 and maturation of human dendritic cells. J Exp Med. 2001; 194:1111–22. [PubMed: 11602640]
- Hamerman JA, Jarjoura JR, Humphrey MB, Nakamura MC, Seaman WE, Lanier LL. Cutting edge: inhibition of TLR and FcR responses in macrophages by triggering receptor expressed on myeloid cells (TREM)-2 and DAP12. J Immunol. 2006; 177:2051–5. [PubMed: 16887962]
- Ito H, Hamerman JA. TREM-2, triggering receptor expressed on myeloid cell-2, negatively regulates TLR responses in dendritic cells. Eur J Immunol. 2012; 42:176–85. [PubMed: 21956652]
- Humphrey MB, Daws MR, Spusta SC, Niemi EC, Torchia JA, Lanier LL, Seaman WE, Nakamura MC. TREM2, a DAP12-associated receptor, regulates osteoclast differentiation and function. J Bone Miner Res. 2006; 21:237–45. [PubMed: 16418779]
- Peng Q, Malhotra S, Torchia JA, Kerr WG, Coggeshall KM, Humphrey MB. TREM2- and DAP12-dependent activation of PI3K requires DAP10 and is inhibited by SHIP1. Sci Signal. 2010; 3:ra38. [PubMed: 20484116]

- 27. Colonna M, Turnbull I, Klesney-Tait J. The enigmatic function of TREM-2 in osteoclastogenesis. Adv Exp Med Biol. 2007; 602:97–105. [PubMed: 17966394]
- Daws MR, Sullam PM, Niemi EC, Chen TT, Tchao NK, Seaman WE. Pattern recognition by TREM-2: binding of anionic ligands. J Immunol. 2003; 171:594–9. [PubMed: 12847223]
- N'Diaye EN, Branda CS, Branda SS, Nevarez L, Colonna M, Lowell C, Hamerman JA, Seaman WE. TREM-2 (triggering receptor expressed on myeloid cells 2) is a phagocytic receptor for bacteria. J Cell Biol. 2009; 184:215–23. [PubMed: 19171755]
- Stefano L, Racchetti G, Bianco F, Passini N, Gupta RS, Panina Bordignon P, Meldolesi J. The surface-exposed chaperone, Hsp60, is an agonist of the microglial TREM2 receptor. J Neurochem. 2009; 110:284–94. [PubMed: 19457124]
- Hsieh CL, Koike M, Spusta SC, Niemi EC, Yenari M, Nakamura MC, Seaman WE. A role for TREM2 ligands in the phagocytosis of apoptotic neuronal cells by microglia. J Neurochem. 2009; 109:1144–56. [PubMed: 19302484]
- 32. Takegahara N, Takamatsu H, Toyofuku T, Tsujimura T, Okuno T, Yukawa K, Mizui M, Yamamoto M, Prasad DV, Suzuki K, Ishii M, Terai K, Moriya M, Nakatsuji Y, Sakoda S, Sato S, Akira S, Takeda K, Inui M, Takai T, Ikawa M, Okabe M, Kumanogoh A, Kikutani H. Plexin-A1 and its interaction with DAP12 in immune responses and bone homeostasis. Nat Cell Biol. 2006; 8:615–22. [PubMed: 16715077]
- 33. Park-Min KH, Ji JD, Antoniv T, Reid AC, Silver RB, Humphrey MB, Nakamura M, Ivashkiv LB. IL-10 suppresses calcium-mediated costimulation of receptor activator NF-kappa B signaling during human osteoclast differentiation by inhibiting TREM-2 expression. J Immunol. 2009; 183:2444–55. [PubMed: 19625651]
- 34. Crotti TN, Dharmapatni AA, Alias E, Zannettino AC, Smith MD, Haynes DR. The immunoreceptor tyrosine-based activation motif (ITAM) -related factors are increased in synovial tissue and vasculature of rheumatoid arthritic joints. Arthritis Res Ther. 14:R245. [PubMed: 23146195]
- 35. Otero K, Turnbull IR, Poliani PL, Vermi W, Cerutti E, Aoshi T, Tassi I, Takai T, Stanley SL, Miller M, Shaw AS, Colonna M. Macrophage colony-stimulating factor induces the proliferation and survival of macrophages via a pathway involving DAP12 and beta-catenin. Nat Immunol. 2009; 10:734–43. [PubMed: 19503107]
- Bakker AB, Baker E, Sutherland GR, Phillips JH, Lanier LL. Myeloid DAP12-associating lectin (MDL)-1 is a cell surface receptor involved in the activation of myeloid cells. Proc Natl Acad Sci U S A. 1999; 96:9792–6. [PubMed: 10449773]
- 37. Inui M, Kikuchi Y, Aoki N, Endo S, Maeda T, Sugahara-Tobinai A, Fujimura S, Nakamura A, Kumanogoh A, Colonna M, Takai T. Signal adaptor DAP10 associates with MDL-1 and triggers osteoclastogenesis in cooperation with DAP12. Proc Natl Acad Sci U S A. 2009; 106:4816–21. [PubMed: 19251634]
- 38. Joyce-Shaikh B, Bigler ME, Chao CC, Murphy EE, Blumenschein WM, Adamopoulos IE, Heyworth PG, Antonenko S, Bowman EP, McClanahan TK, Phillips JH, Cua DJ. Myeloid DAP12-associating lectin (MDL)-1 regulates synovial inflammation and bone erosion associated with autoimmune arthritis. J Exp Med. 2010; 207:579–89. [PubMed: 20212065]
- Shin HS, Sarin R, Dixit N, Wu J, Gershwin E, Bowman EP, Adamopoulos IE. Crosstalk among IL-23 and DNAX activating protein of 12 kDa-dependent pathways promotes osteoclastogenesis. J Immunol. 2015; 194:316–24. [PubMed: 25452564]
- Watson AA, O'Callaghan CA. Crystallization and X-ray diffraction analysis of human CLEC5A (MDL-1), a dengue virus receptor. Acta Crystallogr Sect F Struct Biol Cryst Commun. 2010; 66:29–31. [PubMed: 20057064]
- Chen DY, Yao L, Chen YM, Lin CC, Huang KC, Chen ST, Lan JL, Hsieh SL. A potential role of myeloid DAP12-associating lectin (MDL)-1 in the regulation of inflammation in rheumatoid arthritis patients. PLoS One. 2014; 9:e86105. [PubMed: 24465901]
- Crotti TN, Dharmapatni AA, Alias E, Haynes DR. Osteoimmunology: Major and Costimulatory Pathway Expression Associated with Chronic Inflammatory Induced Bone Loss. J Immunol Res. 2015; 2015:281287. [PubMed: 26064999]

- 43. Ishida-Kitagawa N, Tanaka K, Bao X, Kimura T, Miura T, Kitaoka Y, Hayashi K, Sato M, Maruoka M, Ogawa T, Miyoshi J, Takeya T. Siglec-15 protein regulates formation of functional osteoclasts in concert with DNAX-activating protein of 12 kDa (DAP12). J Biol Chem. 2012; 287:17493–502. [PubMed: 22451653]
- 44. Angata T, Tabuchi Y, Nakamura K, Nakamura M. Siglec-15: an immune system Siglec conserved throughout vertebrate evolution. Glycobiology. 2007; 17:838–46. [PubMed: 17483134]
- Hiruma Y, Hirai T, Tsuda E. Siglec-15, a member of the sialic acid-binding lectin, is a novel regulator for osteoclast differentiation. Biochem Biophys Res Commun. 2011; 409:424–9. [PubMed: 21586272]
- 46. Hiruma Y, Tsuda E, Maeda N, Okada A, Kabasawa N, Miyamoto M, Hattori H, Fukuda C. Impaired osteoclast differentiation and function and mild osteopetrosis development in Siglec-15deficient mice. Bone. 2013; 53:87–93. [PubMed: 23238125]
- 47. Kameda Y, Takahata M, Komatsu M, Mikuni S, Hatakeyama S, Shimizu T, Angata T, Kinjo M, Minami A, Iwasaki N. Siglec-15 regulates osteoclast differentiation by modulating RANKLinduced phosphatidylinositol 3-kinase/Akt and Erk pathways in association with signaling Adaptor DAP12. J Bone Miner Res. 2013; 28:2463–75. [PubMed: 23677868]
- Shimizu T, Takahata M, Kameda Y, Endo T, Hamano H, Hiratsuka S, Ota M, Iwasaki N. Sialic acid-binding immunoglobulin-like lectin 15 (Siglec-15) mediates periarticular bone loss, but not joint destruction, in murine antigen-induced arthritis. Bone. 2015; 79:65–70. [PubMed: 26027508]
- Kameda Y, Takahata M, Mikuni S, Shimizu T, Hamano H, Angata T, Hatakeyama S, Kinjo M, Iwasaki N. Siglec-15 is a potential therapeutic target for postmenopausal osteoporosis. Bone. 2015; 71:217–26. [PubMed: 25460183]
- Lanier LL. DAP10- and DAP12-associated receptors in innate immunity. Immunol Rev. 2009; 227:150–60. [PubMed: 19120482]
- Zou W, Teitelbaum SL. Absence of Dap12 and the alphavbeta3 integrin causes severe osteopetrosis. J Cell Biol. 2015; 208:125–36. [PubMed: 25547154]
- 52. Faccio R, Zou W, Colaianni G, Teitelbaum SL, Ross FP. High dose M-CSF partially rescues the Dap12–/– osteoclast phenotype. J Cell Biochem. 2003; 90:871–83. [PubMed: 14624447]
- Kim N, Takami M, Rho J, Josien R, Choi Y. A novel member of the leukocyte receptor complex regulates osteoclast differentiation. J Exp Med. 2002; 195:201–9. [PubMed: 11805147]
- 54. Merck E, Gaillard C, Gorman DM, Montero-Julian F, Durand I, Zurawski SM, Menetrier-Caux C, Carra G, Lebecque S, Trinchieri G, Bates EE. OSCAR is an FcRgamma-associated receptor that is expressed by myeloid cells and is involved in antigen presentation and activation of human dendritic cells. Blood. 2004; 104:1386–95. [PubMed: 15155468]
- 55. Barrow AD, Raynal N, Andersen TL, Slatter DA, Bihan D, Pugh N, Cella M, Kim T, Rho J, Negishi-Koga T, Delaisse JM, Takayanagi H, Lorenzo J, Colonna M, Farndale RW, Choi Y, Trowsdale J. OSCAR is a collagen receptor that costimulates osteoclastogenesis in DAP12deficient humans and mice. J Clin Invest. 2011; 121:3505–16. [PubMed: 21841309]
- Kim JH, Kim K, Jin HM, Youn BU, Song I, Choi HS, Kim N. Upstream stimulatory factors regulate OSCAR gene expression in RANKL-mediated osteoclast differentiation. J Mol Biol. 2008; 383:502–11. [PubMed: 18762192]
- 57. Kim GS, Koh JM, Chang JS, Park BL, Kim LH, Park EK, Kim SY, Shin HD. Association of the OSCAR promoter polymorphism with BMD in postmenopausal women. J Bone Miner Res. 2005; 20:1342–8. [PubMed: 16007331]
- Herman S, Muller RB, Kronke G, Zwerina J, Redlich K, Hueber AJ, Gelse H, Neumann E, Muller-Ladner U, Schett G. Induction of osteoclast-associated receptor, a key osteoclast costimulation molecule, in rheumatoid arthritis. Arthritis Rheum. 2008; 58:3041–50. [PubMed: 18821671]
- Ndongo-Thiam N, de Sallmard G, Kastrup J, Miossec P. Levels of soluble osteoclast-associated receptor (sOSCAR) in rheumatoid arthritis: link to disease severity and cardiovascular risk. Ann Rheum Dis. 2014; 73:1276–7. [PubMed: 24448348]
- Barrow AD, Palarasah Y, Bugatti M, Holehouse AS, Byers DE, Holtzman MJ, Vermi W, Skjodt K, Crouch E, Colonna M. OSCAR is a receptor for surfactant protein D that activates TNF-alpha release from human CCR2+ inflammatory monocytes. J Immunol. 2015; 194:3317–26. [PubMed: 25716998]

- 61. Goettsch C, Rauner M, Sinningen K, Helas S, Al-Fakhri N, Nemeth K, Hamann C, Kopprasch S, Aikawa E, Bornstein SR, Schoppet M, Hofbauer LC. The osteoclast-associated receptor (OSCAR) is a novel receptor regulated by oxidized low-density lipoprotein in human endothelial cells. Endocrinology. 2011; 152:4915–26. [PubMed: 22009730]
- 62. Ochi S, Shinohara M, Sato K, Gober HJ, Koga T, Kodama T, Takai T, Miyasaka N, Takayanagi H. Pathological role of osteoclast costimulation in arthritis-induced bone loss. Proc Natl Acad Sci U S A. 2007; 104:11394–9. [PubMed: 17592115]
- Dietrich J, Nakajima H, Colonna M. Human inhibitory and activating Ig-like receptors which modulate the function of myeloid cells. Microbes Infect. 2000; 2:323–9. [PubMed: 10758410]
- 64. Negishi-Koga T, Gober HJ, Sumiya E, Komatsu N, Okamoto K, Sawa S, Suematsu A, Suda T, Sato K, Takai T, Takayanagi H. Immune complexes regulate bone metabolism through FcRgamma signalling. Nat Commun. 2015; 6:6637. [PubMed: 25824719]
- 65. Grevers LC, de Vries TJ, Everts V, Verbeek JS, van den Berg WB, van Lent PL. Immune complex-induced inhibition of osteoclastogenesis is mediated via activating but not inhibitory Fcgamma receptors on myeloid precursor cells. Ann Rheum Dis. 2013; 72:278–85. [PubMed: 22918932]
- 66. MacLellan LM, Montgomery J, Sugiyama F, Kitson SM, Thummler K, Silverman GJ, Beers SA, Nibbs RJ, McInnes IB, Goodyear CS. Co-opting endogenous immunoglobulin for the regulation of inflammation and osteoclastogenesis in humans and mice. Arthritis Rheum. 2011; 63:3897–907. [PubMed: 22127707]
- 67. Harre U, Georgess D, Bang H, Bozec A, Axmann R, Ossipova E, Jakobsson PJ, Baum W, Nimmerjahn F, Szarka E, Sarmay G, Krumbholz G, Neumann E, Toes R, Scherer HU, Catrina AI, Klareskog L, Jurdic P, Schett G. Induction of osteoclastogenesis and bone loss by human autoantibodies against citrullinated vimentin. J Clin Invest. 2012; 122:1791–802. [PubMed: 22505457]
- 68. Harre U, Lang SC, Pfeifle R, Rombouts Y, Fruhbeisser S, Amara K, Bang H, Lux A, Koeleman CA, Baum W, Dietel K, Grohn F, Malmstrom V, Klareskog L, Kronke G, Kocijan R, Nimmerjahn F, Toes RE, Herrmann M, Scherer HU, Schett G. Glycosylation of immunoglobulin G determines osteoclast differentiation and bone loss. Nat Commun. 2015; 6:6651. [PubMed: 25825024]
- Bohm S, Schwab I, Lux A, Nimmerjahn F. The role of sialic acid as a modulator of the antiinflammatory activity of IgG. Semin Immunopathol. 2012; 34:443–53. [PubMed: 22437760]
- 70. Scherer HU, van der Woude D, Ioan-Facsinay A, el Bannoudi H, Trouw LA, Wang J, Haupl T, Burmester GR, Deelder AM, Huizinga TW, Wuhrer M, Toes RE. Glycan profiling of anticitrullinated protein antibodies isolated from human serum and synovial fluid. Arthritis Rheum. 2010; 62:1620–9. [PubMed: 20178128]
- 71. Daeron M, Jaeger S, Du Pasquier L, Vivier E. Immunoreceptor tyrosine-based inhibition motifs: a quest in the past and future. Immunol Rev. 2008; 224:11–43. [PubMed: 18759918]
- Hayashi M, Nakashima T, Kodama T, Makrigiannis AP, Toyama-Sorimachi N, Takayanagi H. Ly49Q, an ITIM-bearing NK receptor, positively regulates osteoclast differentiation. Biochem Biophys Res Commun. 2010; 393:432–8. [PubMed: 20153723]
- 73. Yagi M, Miyamoto T, Sawatani Y, Iwamoto K, Hosogane N, Fujita N, Morita K, Ninomiya K, Suzuki T, Miyamoto K, Oike Y, Takeya M, Toyama Y, Suda T. DC-STAMP is essential for cellcell fusion in osteoclasts and foreign body giant cells. J Exp Med. 2005; 202:345–51. [PubMed: 16061724]
- 74. Chiu YH, Mensah KA, Schwarz EM, Ju Y, Takahata M, Feng C, McMahon LA, Hicks DG, Panepento B, Keng PC, Ritchlin CT. Regulation of human osteoclast development by dendritic cell-specific transmembrane protein (DC-STAMP). J Bone Miner Res. 2012; 27:79–92. [PubMed: 21987375]
- Nagahama K, Aoki K, Nonaka K, Saito H, Takahashi M, Varghese BJ, Shimokawa H, Azuma M, Ohya K, Ohyama K. The deficiency of immunoregulatory receptor PD-1 causes mild osteopetrosis. Bone. 2004; 35:1059–68. [PubMed: 15542030]
- 76. Aoki K, Didomenico E, Sims NA, Mukhopadhyay K, Neff L, Houghton A, Amling M, Levy JB, Horne WC, Baron R. The tyrosine phosphatase SHP-1 is a negative regulator of osteoclastogenesis and osteoclast resorbing activity: increased resorption and osteopenia in me(v)/me(v) mutant mice. Bone. 1999; 25:261–7. [PubMed: 10495129]

- 77. Takeshita S, Namba N, Zhao JJ, Jiang Y, Genant HK, Silva MJ, Brodt MD, Helgason CD, Kalesnikoff J, Rauh MJ, Humphries RK, Krystal G, Teitelbaum SL, Ross FP. SHIP-deficient mice are severely osteoporotic due to increased numbers of hyper-resorptive osteoclasts. Nat Med. 2002; 8:943–9. [PubMed: 12161749]
- Yoon SH, Lee YD, Ha J, Lee Y, Kim HH. TLT-1s, alternative transcripts of triggering receptor expressed on myeloid cell-like transcript-1 (TLT-1), Inhibits the triggering receptor expressed on myeloid cell-2 (TREM-2)-mediated signaling pathway during osteoclastogenesis. J Biol Chem. 2012; 287:29620–6. [PubMed: 22761415]
- Chung DH, Humphrey MB, Nakamura MC, Ginzinger DG, Seaman WE, Daws MR. CMRF-35like molecule-1, a novel mouse myeloid receptor, can inhibit osteoclast formation. J Immunol. 2003; 171:6541–8. [PubMed: 14662855]
- Mori Y, Tsuji S, Inui M, Sakamoto Y, Endo S, Ito Y, Fujimura S, Koga T, Nakamura A, Takayanagi H, Itoi E, Takai T. Inhibitory immunoglobulin-like receptors LILRB and PIR-B negatively regulate osteoclast development. J Immunol. 2008; 181:4742–51. [PubMed: 18802077]
- Li S, Miller CH, Giannopoulou E, Hu X, Ivashkiv LB, Zhao B. RBP-J imposes a requirement for ITAM-mediated costimulation of osteoclastogenesis. J Clin Invest. 2014; 124:5057–73. [PubMed: 25329696]
- Everts V, de Vries TJ, Helfrich MH. Osteoclast heterogeneity: lessons from osteopetrosis and inflammatory conditions. Biochim Biophys Acta. 2009; 1792:757–65. [PubMed: 19465118]

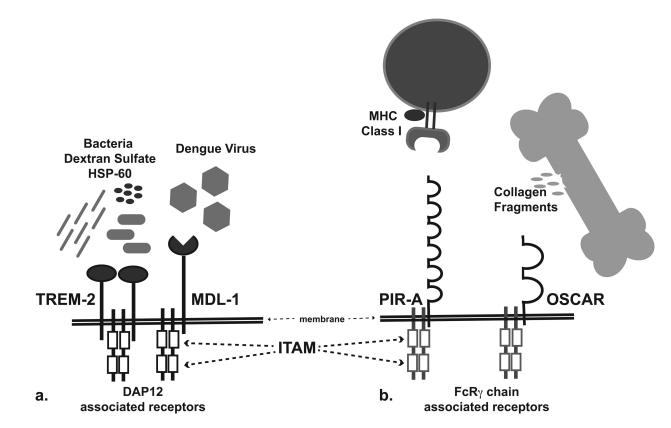


Figure 1. ITAM-signaling adaptor-associated immunoreceptors

A) DAP12-associated receptors TREM-2 and MDL-1. TREM-2 has been shown to interact with bacteria, dextran sulfate and HSP-60 as ligands. Dengue virus is a ligand for MDL-1. B) FcR γ -associated receptor PIR-A interacts with MHC Class I ligands on cell surfaces. OSCAR is also an FcR γ -associated receptor and collagen fragments are functional ligands for OSCAR.

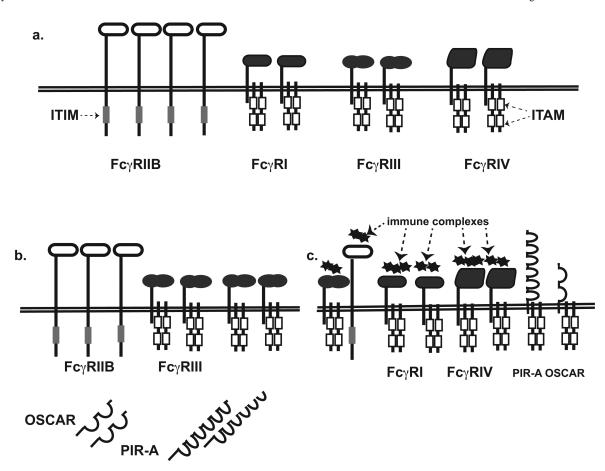


Figure 2. Fc Receptors in Osteoclasts Activate and Inhibit Osteoclastogenesis

A) Multiple Fc Receptors are expressed in osteoclasts. FcγRIIB is an ITIM-signaling recptor. FcγRI, FcγRIII and FcγRIV are all ITAM-signaling adaptor-associated immunoreceptors that associate with the FcRγ adaptor chain. B) Under basal conditions, osteoclast precursors express only FcγRIIB and FcγRIII. Osteoclastogenesis is inhibited. FcγRIII is thought to inhibit osteoclastogenesis by sequestering the FcRγ adaptor chain from PIR-A and OSCAR, which remain intracellular but are not expressed on the cell surface due to the lack of availability of FcRγ adaptor chains. C) RANKL stimulation in the presence of inflammation and immune complex formation leads to downregulation of FcγRIIB and FcγRIII, and upregulation of PIR-A and OSCAR. Osteoclastogenesis is enhanced. (Based on (64)).