

UCSF

UC San Francisco Previously Published Works

Title

Performance of Cervical Screening a Decade Following HPV Vaccination: The Costa Rica Vaccine Trial.

Permalink

<https://escholarship.org/uc/item/60g0z9ss>

Journal

Journal of the National Cancer Institute, 114(9)

Authors

Hu, Shang-Ying
Kreimer, Aimée
Porras, Carolina
[et al.](#)

Publication Date




2022-09-09

DOI

10.1093/jnci/djac107

Peer reviewed

Performance of Cervical Screening a Decade Following HPV Vaccination: The Costa Rica Vaccine Trial

Shang-Ying Hu, PhD ^{1,2,*}, Aimée R. Kreimer, PhD,² Carolina Porras, MSc,³ Diego Guillén, MD ³, Mario Alfaro, MD,³ Teresa M. Darragh, MD,⁴ Mark H. Stoler, MD,⁵ Luis F. Villegas, MD,³ Rebecca Ocampo, MD,³ Ana Cecilia Rodriguez, MD,⁶ Mark Schiffman, MD, PhD,² Sabrina H. Tsang, PhD, MPH,² Douglas R. Lowy, MD,^{2,7} John T. Schiller, PhD,⁷ John Schussler, BSc,⁸ Wim Quint, PhD,⁹ Mitchell H. Gail, MD, PhD,² Joshua N. Sampson, PhD ², Allan Hildesheim, PhD,² Rolando Herrero, MD, PhD^{3,10}; on behalf of the Costa Rica HPV Vaccine Trial (CVT) Group

¹Department of Cancer Epidemiology, National Cancer Center/National Clinical Research Center for Cancer/Cancer Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China; ²Division of Cancer Epidemiology and Genetics, National Cancer Institute, National Institutes of Health, Bethesda, MD, USA; ³Agencia Costarricense de Investigaciones Biomédicas (ACIB), Fundación INCIENSA, San José, Costa Rica; ⁴School of Medicine, University of California, San Francisco, CA, USA; ⁵Department of Pathology, University of Virginia, Charlottesville, VA, USA; ⁶Independent Consultant, San José, Costa Rica; ⁷Laboratory of Cellular Oncology, Center for Cancer Research, National Cancer Institute, Rockville, MD, USA; ⁸Information Management Services, Silver Spring, MD, USA; ⁹Viroclinics-DDL, Rijswijk, Netherlands; and ¹⁰Early Detection Prevention and Infections, International Agency for Research on Cancer, Lyon, France

*Correspondence to: Shang-Ying Hu, PhD, Department of Cancer Epidemiology, National Cancer Center/National Clinical Research Center for Cancer/Cancer Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College, 17 South Panjiyuan Lane, PO Box 2258, Beijing 100021, China (e-mail: shangyinghu@cicams.ac.cn).

Abstract

Background: We investigated the impact of human papillomavirus (HPV) vaccination on the performance of cytology-based and HPV-based screening for detection of cervical precancer among women vaccinated as young adults and reaching screening age. **Methods:** A total of 4632 women aged 25–36 years from the Costa Rica HPV Vaccine Trial were included (2418 HPV-vaccinated as young adults and 2214 unvaccinated). We assessed the performance of cytology- and HPV-based cervical screening modalities in vaccinated and unvaccinated women to detect high-grade cervical precancers diagnosed over 4 years and the absolute risk of cumulative cervical precancers by screening results at entry. **Results:** We detected 95 cervical intraepithelial neoplasia grade 3 or worse (52 in unvaccinated and 43 in vaccinated women). HPV16/18/31/33/45 was predominant (69%) among unvaccinated participants, and HPV35/52/58/39/51/56/59/66/68 predominated (65%) among vaccinated participants. Sensitivity and specificity of cervical screening approaches were comparable between women vaccinated as young adults and unvaccinated women. Colposcopy referral rates were lower in the vaccinated group for HPV-based screening modalities, but the positive predictive value was comparable between the 2 groups. **Conclusions:** Among women approaching screening ages, vaccinated as young adults, and with a history of intensive screening, the expected reduction in the positive predictive value of HPV testing, associated with dropping prevalence of HPV-associated lesions, was not observed. This is likely due to the presence of high-grade lesions associated with nonvaccine HPV types, which may be less likely to progress to cancer.

Cervical cancer (CC) is the fourth most common cancer among women worldwide (1), is causally associated with persistent human papillomavirus (HPV) infection (2), and is preventable by vaccination against HPV and screening/treatment of precancers (3). Vaccines have been adopted into immunization programs in more than 100 countries worldwide (4).

CC screening approaches in countries with high vaccination coverage must be carefully considered as vaccinated cohorts

attain screening age. Reductions in vaccine-preventable HPV types and associated lesions could diminish the positive predictive value (PPV) and other characteristics of screening strategies (5–8). Lengthening of screening intervals may be possible according to guidelines benchmarking against precancer risk (9). Data on performance of screening and triage methods are required to define programmatic approaches, which may differ in high- and low-resource settings.

There are limited data from HPV-vaccinated cohorts reaching screening ages, and current guidelines do not differentiate programmatic recommendations based on the existence of vaccination programs (10-12). Thus far, most analyses from clinical and population-based studies focused on changes in cytology reporting profiles, colposcopy workload, and clinical activities in vaccinated women (5,13-17) or cost-effectiveness using health decision models (18-20). A randomized trial in Australia in a population with substantial uptake of HPV vaccination demonstrated a statistically significantly higher detection rate of high-grade precancers in HPV-screened women compared with cytology-screened women (21).

To inform cervical screening guidelines for HPV-vaccinated populations, we analyzed data from the Costa Rica (CR) HPV Vaccine Trial (CVT) and report performance of cytology- and HPV-based screening methods for predicting high-grade precancer and absolute risk of cumulative precancers by screening results, comparing unvaccinated women with women vaccinated as young adults whose attained age ranged from 25 to 36 years.

Methods

Study Population and Procedure

CVT was a phase III trial evaluating the efficacy of the HPV16/18 vaccine (Cervarix, GSK, Belgium) against cervical HPV16/18 infection and related precancers (NCT00128661), as previously described (22,23). Between 2004 and 2005, 7466 women in CR aged 18 to 25 years were randomly assigned (1:1) to receive 3 doses of HPV16/18 vaccine (HPV-vaccinated arm) or hepatitis A vaccine (control arm) and were followed-up annually for 4 years. At enrollment and follow-up, sexually active women had cervical cells collected for liquid-based cytology and HPV testing. Cytology guided colposcopy referral. Women with minor cytological abnormalities (including low-grade squamous intraepithelial lesion, atypical squamous cells of undetermined significance with HPV positive [ASC-US/HPV+], or inadequate cytology) were followed-up every 6 months, and women with high-grade squamous intraepithelial lesion (HSIL) or repeated minor abnormalities had colposcopy, biopsy, and treatment as needed. After colposcopy and/or treatment, women were followed-up every 6 months.

After the initial 4-year follow-up, women had cross-over vaccination and the HPV-vaccinated arm was invited to a long-term follow-up observational study for 7 additional years. To replace controls, a new screening-only unvaccinated control group (UCG, n = 2836) was recruited from identical birth cohorts and geographical regions. At enrollment, UCG women were screened using cytology and HPV testing and followed strict colposcopic algorithms to increase comparability with CVT women, who were previously screened for 4 years. UCG women with negative screening tests or HPV-negative ASC-US were rescreened in 2 years. Women with cytologic HSIL had colposcopy or treatment as necessary, and those with minor cytological abnormalities or positive or insufficient HPV test had cotesting 6 months later. If both tests were negative, they reverted to biennial screening; otherwise, they attended colposcopy. Comparability between control groups regarding women's characteristics and future HPV acquisition risk has been reported (23).

Since year 7 after vaccination (HPV-vaccinated group) and after 1 round of intensive screening (UCG), namely since the baseline visit of this analysis, both groups had 2 liquid-based

cytology tests at 2-year intervals. Women with minor abnormalities had 6-month follow-up with cotesting. If tests were negative, women returned to cytologic biennial screening. If cytology was abnormal, they had colposcopy; if only HPV was positive, they had a second 6-month follow-up; and, if positive, they had colposcopy, as did women with HSIL cytology. At the last screening visit 4 years after the baseline visit of this analysis (11 years after vaccination for vaccinated group), women had cotesting with cytology and HPV testing. Those with minor abnormalities or only HPV positive had a 6-month cotesting follow-up, and if positive in either test they had colposcopy; otherwise, they were exited. Figure 1 shows the screening and follow-up procedures involved in this analysis.

Histologically confirmed cervical intraepithelial neoplasia grade 2 or worse (CIN2+; including CIN2, CIN3, adenocarcinoma in situ, and cancer) were treated mainly with large loop excision of transformation zone. For women with cytologic abnormalities or worrisome virologic patterns, a review panel including epidemiologists, clinicians, and pathologists assessed virologic, cytologic, colposcopic, and histologic findings for clinical safety and return to usual care. In some cases, diagnostic large loop excision of transformation zone was performed (eg, persistent HPV16 infection with cytologic abnormalities not histologically confirmed). CIN2+ diagnoses during colposcopic follow-up until December 2019 were included. The study was approved by Instituto Costarricense de Investigación y Enseñanza en Nutrición y Salud (INCIENSA) institutional review board in CR and National Cancer Institute institutional review board in the United States; all participants signed informed consent.

Clinical Diagnosis and HPV Testing

Cytology was reported using the Bethesda system (24) at a central laboratory in CR. During the blinded CVT portion, abnormal slides plus 10% random negatives were reread in the United States. Upgrades triggered colposcopic referral. Given good agreement ($\kappa = 0.68$, 95% CI = 0.66 to 0.69), this process stopped during long-term follow-up observational study. Histology was initially interpreted by a CR pathologist (Haematoxylin and Eosin, CIN classification), and a US pathologist reviewed slides blinded to the CR diagnoses. Discrepant results were reviewed by another US pathologist, and majority rule defined the final diagnosis; p16 immunostaining was not used.

Hybrid Capture 2 (HC2; Qiagen, MD, USA) was used for DNA detection of any of 13 oncogenic types (HPV16/18/31/33/35/39/45/51/52/56/58/59/68) for clinical management and ASC-US triage. HC2 also detects high viral loads of genetically related types (25). At year 11, this test was replaced by Aptima HPV (Hologic, CA, USA), which detects HPV E6/E7 mRNA from 13 HPV types (HPV16/18/31/33/35/39/45/51/52/56/58/59/68) and HPV66 (26).

During the blinded phase of CVT, HPV genotyping was performed using SPF10/DEIA/LiPA25 assay (version 1, Viroclinics-DDL, the Netherlands), detecting 14 oncogenic (HPV16/18/31/33/35/39/45/51/52/56/58/59/66, and HPV68/73) and 11 nononcogenic (HPV6/11/34/40/42/43/44/53/54/70/74) HPV types. HPV68/73 was treated as HPV68 for statistics because the 2 types cannot be discriminated. The test was replaced by the National Cancer Institute-developed, TypeSeq HPV method after demonstrating comparability with SPF10/DEIA/LiPA25 (27). TypeSeq is a next-generation sequencing-based assay using TypeSeq 3-PCR stage workflow and a custom Torrent Suite plugin. It provides a binary result (positive or negative) for 51 HPV types and a human DNA control.

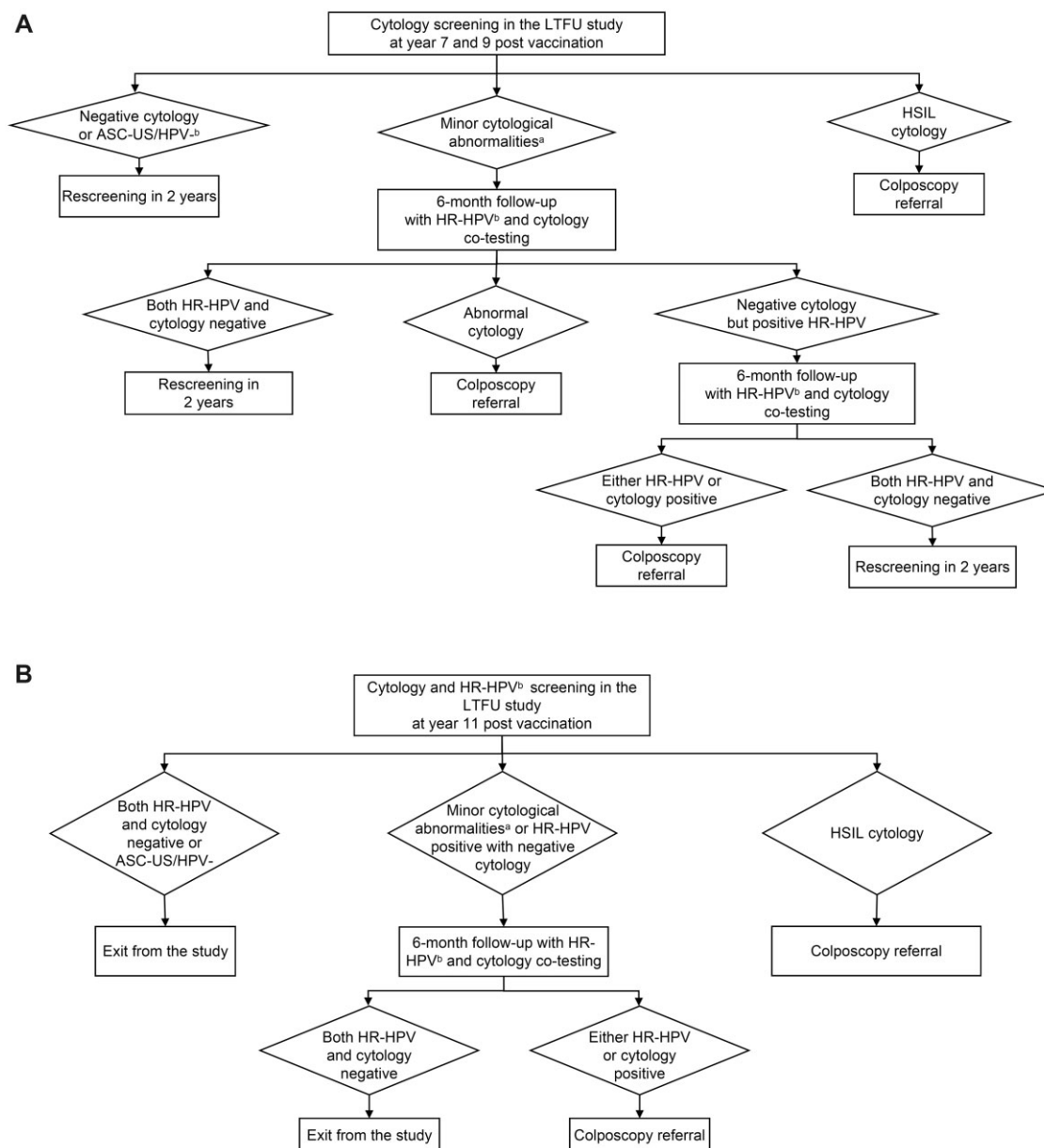


Figure 1. Flowchart of screening procedure in the long-term follow-up (LTFU) study. **A)** Flowchart of the screening procedure at year 7 and 9 post vaccination is shown. **B)** Flowchart of the screening procedure at year 11 post vaccination is shown. ^aMinor cytological abnormalities included low-grade squamous intraepithelial lesion, atypical squamous cells of undetermined significance with a positive high-risk human papillomavirus (HR-HPV) test (ASC-US/HPV+), or inadequate cytology. ^bHybrid Capture 2 (Qiagen, MD, USA) was used for DNA detection of any of 13 oncogenic types (HPV16/18/31/33/35/39/45/51/52/56/58/59/68) for clinical management and ASC-US triage at year 7 and 9. At year 11, this test was replaced by Aptima HPV (Hologic, CA, USA), detecting HPV E6/E7 mRNA from 13 HPV types (HPV16/18/31/33/35/39/45/51/52/56/58/59/68) and HPV66. HSIL = high-grade squamous intraepithelial lesion.

Analytical Cohorts

Our analytical population included 4632 women who attended the baseline screening visit (year 7 after vaccination or first visit after initial screening for UCG), were 25 years or older, and had at least 1 follow-up visit. Women with CIN2+ before baseline were excluded (Figure 2). A total of 2418 were vaccinated and 2214 were UCG. Analyses start at year 7 of the vaccinated cohort because 1) participants were aged 25 years to 36 years, within the targeted screening ages (172 women younger than 25 years were excluded); 2) the impact of vaccination is more pronounced as time passes after vaccination (14,28); and 3) intensive screening of the UCG had been completed and validation of comparability

of the groups performed. Follow-up time was stopped at diagnosis of CIN2+ or censored at the last follow-up.

Statistical Analysis

We evaluated the performance of the cytology algorithm (with referral of ASC-US/HPV+ or more severe) and alternative approaches simulated using available data, separately for unvaccinated and vaccinated women, as follows: primary HPV screening methods, including 14 high-risk HPV (HR-HPV; HPV16/18/31/33/35/39/45/51/52/56/58/59/66/68) alone and 8-HPV typing (HPV16/18/31/33/35/45/52/58) alone; cotesting with HR-HPV and cytology; and triage methods

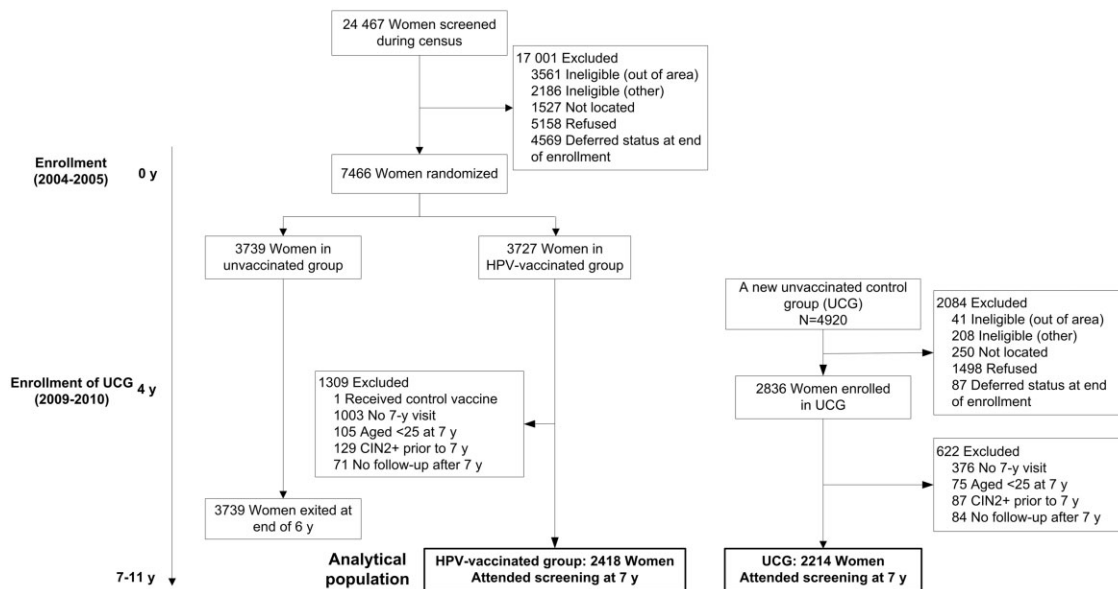


Figure 2. Selection of analytic cohorts. CIN2+ = cervical intraepithelial neoplasia grade 2 or worse; HPV = human papillomavirus; UCG = unvaccinated control group.

for HR-HPV-positive women: HR-HPV triaged with cytology (ASC-US+ threshold without knowledge of HPV status), with HPV16/18 and reflex cytology, and with 8-HPV typing and reflex cytology. We evaluated the HPV16/18/31/33/35/45/52/58 group as primary screening or triage for HR-HPV-positive women because these HPV types have high 3-year CIN3+ (including CIN3, adenocarcinoma in situ, and cancer) risk (29), whereas other HR-HPV types have lower risk despite high prevalence in some populations.

We assessed the performance of screening modalities at entry into the analytical cohort, including referral rate, sensitivity, specificity, PPV, and negative predictive value (NPV). The cumulative cervical precancer cases from year 7 to 11 were defined as the gold standard, with CIN3+ as primary and CIN2+ as secondary outcome. Women who always tested negative were considered negative. Mid-p corrected exact binomial 95% confidence intervals (CIs) of the above metrics were calculated. Ratios of the metrics and 95% confidence intervals between groups were reported by an asymptotic Wald method. Moreover, we performed sensitivity analysis using CIN3+ within 1 year after first colposcopy to assess the impact of repeated colposcopies on our assessments.

We also assessed cumulative risks of CIN2+/CIN3+ during follow-up by test results at entry and compared risks in the vaccinated group with those in the UCG. This statistic is calculated by dividing the number of outcomes in the strata by the total number of women in the strata. Ratios and 95% confidence intervals between groups were calculated by an asymptotic Wald method. Meanwhile, χ^2 tests were used to compare HPV distributions among cumulative cervical precancers between groups. Statistical analyses were performed with SAS 9.4. All statistical tests were 2-sided, and P values less than .05 were considered statistically significant.

Results

Participant Characteristics

The median follow-up time was 4.3 years after entry in the analytical cohort (interquartile range = 3.7–4.9), and it was similar between the HPV-vaccinated group and the UCG

(Supplementary Table 1, available online). UCG women were similar with respect to age and lifetime number of sexual partners at entry compared with the vaccinated group but had more pregnancies, slightly lower reported oral contraceptive use, and a slightly lower rate of smoking. As expected, compared with the vaccinated group, the UCG had higher HR-HPV (23.0% vs 20.2%) and HPV16/18 positivity (6.2% vs 1.0%) at entry.

Clinical Performance of Different Screening Modalities by Vaccination Status

When comparing each specific screening modality between the HPV-vaccinated and the UCG (Tables 1 and 2), somewhat higher sensitivities for detection of cumulative CIN3+ were observed in the vaccinated group for cytology alone (50.0% vs 40.8%; ratio = 1.22, 95% CI = 0.78 to 1.93) and consequently HR-HPV triaged with cytology (47.6% vs 40.8%, ratio = 1.17, 95% CI = 0.73 to 1.85), along with similar specificity. The PPVs and NPVs for all test combinations were comparable between the 2 groups. Referral rates were lower in vaccinated women when considering HPV-based modalities but not cytology-based ones.

The results for CIN3+ were generally comparable with those for CIN2+ (Supplementary Tables 2 and 3, available online) or restricting to the first year of colposcopy (Supplementary Tables 4 and 5, available online) or restricting to women aged 30 years and older (data not shown). The performance of the screening modalities in the vaccinated group when restricted to women who were HPV negative at vaccination (Supplementary Tables 6 and 7, available online) was similar to that of the vaccinated group including both HPV-positive and HPV-negative women (Tables 1 and 2).

Cumulative Risk of Cervical Precancers by Vaccination Status and Screening Results

Cumulative risk of cervical precancers over the 4 years including repeated colposcopic evaluations was higher in the UCG

Table 1. Comparison of screening performance across groups (HPV-vaccinated group vs UCG) using primary screening tests for CIN3+ detection over 4 years

Primary screening method	Group	Total screened, No.	CIN3+ detection, No. (%)	Sensitivity, % (95% CI)	Specificity, % (95% CI)	PPV, % (95% CI)	NPV, % (95% CI)	Referral rate, % (95% CI)
Cytology ^a	UCG	2117	20 (0.9)	40.8 (27.8 to 54.9)	91.9 (90.7 to 93.0)	10.7 (6.8 to 15.8)	98.5 (97.9 to 99.0)	8.8 (7.7 to 10.1)
	HPV vaccinated	2320	21 (0.9)	50.0 (35.1 to 64.9)	92.4 (91.2 to 93.4)	10.8 (7.0 to 15.7)	99.0 (98.5 to 99.4)	8.4 (7.3 to 9.6)
Ratio compared with UCG				1.22 (0.78 to 1.93)	1.00 (0.99 to 1.02)	1.01 (0.56 to 1.80)	1.01 (1.00 to 1.01)	0.95 (0.79 to 1.15)
HR-HPV ^b	UCG	2213	38 (1.7)	73.1 (59.9 to 83.8)	78.2 (76.4 to 79.8)	7.5 (5.4 to 10.0)	99.2 (98.7 to 99.5)	23.0 (21.3 to 24.8)
	HPV vaccinated	2413	32 (1.3)	74.4 (59.9 to 85.7)	80.8 (79.2 to 82.3)	6.6 (4.6 to 9.0)	99.4 (99.0 to 99.7)	20.2 (18.6 to 21.8)
Ratio compared with UCG				1.02 (0.80 to 1.30)	1.03 (1.00 to 1.06)	0.88 (0.56 to 1.39)	1.00 (1.00 to 1.01)	0.88 (0.78 to 0.98)
8-HPV genotyping ^c	UCG	2213	31 (1.4)	59.6 (45.9 to 72.3)	85.7 (84.1 to 87.1)	9.1 (6.4 to 12.5)	98.9 (98.3 to 99.3)	15.4 (14.0 to 17.0)
	HPV vaccinated	2413	26 (1.1)	60.5 (45.4 to 74.2)	91.2 (90.0 to 92.3)	11.1 (7.5 to 15.6)	99.2 (98.8 to 99.5)	9.7 (8.6 to 10.9)
Ratio compared with UCG				1.01 (0.73 to 1.41)	1.07 (1.04 to 1.09)	1.22 (0.75 to 2.00)	1.00 (1.00 to 1.01)	0.63 (0.54 to 0.74)
Cotesting with HR-HPV ^b and cytology (ASC-US+)	UCG	2116	39 (1.8)	79.6 (66.6 to 89.1)	73.1 (71.1 to 75.0)	6.6 (4.8 to 8.8)	99.3 (98.8 to 99.7)	28.1 (26.2 to 30.1)
	HPV vaccinated	2319	33 (1.4)	78.6 (64.3 to 89.0)	75.5 (73.6 to 77.2)	5.6 (3.9 to 7.7)	99.5 (99.0 to 99.7)	25.5 (23.8 to 27.3)
Ratio compared with UCG				0.99 (0.80 to 1.22)	1.03 (1.00 to 1.07)	0.85 (0.54 to 1.33)	1.00 (1.00 to 1.01)	0.91 (0.82 to 1.00)

^aASC-US+ = atypical squamous cells of undetermined significance or worse; CI = confidence interval; CIN3+ = cervical intraepithelial neoplasia grade 3 or worse; HC2 = Hybrid Capture 2; HR-HPV = high-risk human papillomavirus; NPV = negative predictive value; PPV = positive predictive value; UCG = unvaccinated control group. Cytology screening algorithm refers to cytology plus HC2 triage of ASC-US.

^bHR-HPV includes all 14 oncogenic HPV types: HPV16/18/31/33/35/39/45/51/52/56/58/59/66/68.

^c8-HPV genotyping refers to HPV16/18/31/33/35/45/52/58.

Table 2. Comparison of screening performance across groups (HPV-vaccinated group vs UCG) using screening algorithms with triage for HR-HPV^a positive women for CIN3+ detection over 4 years

Screening method (Primary test/ triage test)	Group	Total screened, No.	CIN3+ detection, No. (%)	Sensitivity, % (95% CI)	Specificity, % (95% CI)	PPV, % (95% CI)	NPV, % (95% CI)	Referral rate, % (95% CI)
HR-HPV ^a / cytology (ASC-US+)	UCG	2116	20 (0.9)	40.8 (27.8 to 54.9)	93.5 (92.4 to 94.5)	13.0 (8.3 to 19.0)	98.5 (97.9 to 99.0)	7.3 (6.2 to 8.4)
	HPV vaccinated	2319	20 (0.9)	47.6 (32.9 to 62.6)	94.0 (93.0 to 94.9)	12.8 (8.2 to 18.8)	99.0 (98.5 to 99.3)	6.7 (5.8 to 7.8)
Ratio compared with UCG				1.17 (0.73 to 1.85)	1.01 (0.99 to 1.02)	0.99 (0.55 to 1.76)	1.00 (1.00 to 1.01)	0.92 (0.75 to 1.15)
HR-HPV ^a / HPV16/18 and reflex cytology (ASC-US+)	UCG	2116	24 (1.1)	49.0 (35.2 to 62.8)	89.3 (87.9 to 90.6)	9.8 (6.5 to 14.0)	98.7 (98.1 to 99.1)	11.6 (10.3 to 13.0)
	HPV vaccinated	2319	23 (1.0)	54.8 (39.6 to 69.2)	93.5 (92.4 to 94.4)	13.4 (8.9 to 19.1)	99.1 (98.6 to 99.5)	7.4 (6.4 to 8.5)
Ratio compared with UCG				1.12 (0.75 to 1.66)	1.05 (1.03 to 1.07)	1.37 (0.80 to 2.34)	1.00 (1.00 to 1.01)	0.64 (0.53 to 0.77)
HR-HPV ^a / 8-HPV genotyping ^b and reflex cytology (ASC-US+)	UCG	2116	34 (1.6)	69.4 (55.5 to 81.0)	83.4 (81.7 to 84.9)	9.0 (6.4 to 12.2)	99.1 (98.6 to 99.5)	17.8 (16.2 to 19.5)
	HPV vaccinated	2319	31 (1.3)	73.8 (59.0 to 85.4)	87.9 (86.5 to 89.2)	10.1 (7.1 to 13.9)	99.5 (99.1 to 99.7)	13.2 (11.9 to 14.7)
Ratio compared with UCG				1.06 (0.82 to 1.38)	1.05 (1.03 to 1.08)	1.12 (0.70 to 1.78)	1.00 (1.00 to 1.01)	0.74 (0.65 to 0.85)

^a8-HPV genotyping refers to HPV16/18/31/33/35/45/52/58. ASC-US+ = atypical squamous cells of undetermined significance or worse; CI = confidence interval; CIN3+ = cervical intraepithelial neoplasia grade 3 or worse; HPV = human papillomavirus; HR-HPV = high-risk human papillomavirus; NPV = negative predictive value; PPV = positive predictive value; UCG = unvaccinated control group. HR-HPV includes all 14 oncogenic HPV types: HPV16/18/31/33/35/39/45/51/52/56/58/59/66/68.

^b8-HPV genotyping refers to HPV16/18/31/33/35/45/52/58.

Table 3. HPV type distribution in CIN2 and CIN3+ diagnosed over 4 years by vaccination status^a

HPV genotypes	CIN2 (n = 66)		CIN3+ (n = 95)	
	UCG No. (%)	HPV-vaccinated group No. (%)	UCG No. (%)	HPV-vaccinated group No. (%)
HPV16/18	18 (38.3)	1 (5.3)	22 (42.3)	4 (9.3)
HPV31/33/45	12 (25.5)	1 (5.3)	14 (26.9)	9 (20.9)
HPV35/52/58	9 (19.1)	7 (36.8)	11 (21.2)	18 (41.9)
HPV39/51/56/59/66/68	7 (14.9)	5 (26.3)	3 (5.8)	10 (23.3)
Nononcogenic HPV	0 (0.0)	2 (10.5)	1 (1.9)	1 (2.3)
HPV negative/unknown	1 (2.1)	3 (15.8)	1 (1.9)	1 (2.3)

^aCIN2 = cervical intraepithelial neoplasia grade 2; CIN3+ = cervical intraepithelial neoplasia grade 3 or worse; HPV = human papillomavirus; UCG = unvaccinated control group.

than in the vaccinated group (CIN3+: 2.3% vs 1.8%, $P = .17$; CIN2+: 4.5% vs 2.6%, $P < .001$). When restricting to the first year of colposcopy, these estimates decreased by approximately one-third in the UCG and by one-fourth in the vaccinated group.

For the 95 CIN3+, HPV16/18/31/33/45 were predominant (69.2%) among the UCG cases, whereas HPV35/52/58/39/51/56/59/66/68 predominated (65.2%) among the vaccinated participants (Table 3). A similar pattern was observed in the 66 CIN2. All lesions associated with HPV16/18 in the vaccinated group occurred in women HPV16/18 positive before vaccination.

Compared with the UCG, vaccinated women with negative cytology, negative HR-HPV, or both cytology and HR-HPV negative at entry had a statistically non-significant approximately 20% to 30% decreased risk of cumulative CIN3+ (Table 4) and at least 50% decreased risk of cumulative CIN2+ (Supplementary Table 8, available online). Comparable decreased risks of cumulative CIN3+ and CIN2+ in vaccinated women were observed for women with only HR-HPV positive or with only non-HPV16/18 positive. There were no reductions in cumulative risks among women with cytological HSIL+, or HPV16/18 or non-HPV16/18 plus ASC-US+. These patterns were similar for diagnoses within 1 year of colposcopy, with lower incidences (Supplementary Table 9, available online). Cumulative risk of CIN3+ among vaccinated women who were HPV negative at vaccination (Supplementary Table 10, available online) was comparable with that in the entire group including HPV-positive and -negative participants. The patterns were again comparable for women 30 years and older (data not shown).

Discussion

We modeled performance of cytology- and HPV-based screening modalities over 4 years. Among previously screened young women, sensitivity and specificity of screening modalities were comparable between vaccinated and unvaccinated women. Interestingly, the PPV for detection of CIN3+ was similar in the 2 groups for all screening approaches, including those where triage methods were used. However, HPV types in lesions were very different, with predominance of vaccine and cross-protected types in the unvaccinated and of other types in the vaccinated. Nonvaccine HPV types are less frequent in CC, and approximately 50% of precancers are caused by non-16/18 HPV types compared with approximately 30% of cancers (30). Thus, in vaccinated women, a positive screening test result, despite predicting histologically confirmed CIN3+, may be associated with lower cancer risk than in unvaccinated women, given different progression risks associated with different HPV types. This has implications for screening vaccinated cohorts because

numbers of lesions currently considered cancer precursors may not be reduced as expected. Additional natural history studies of nonvaccine HPV types and associated lesions are required, despite design difficulties. NPV was similar, but vaccinated women had lower colposcopy referral using HPV-based screening approaches due to lower detection of HPV infections.

As previously observed in unvaccinated women (31-33), cytology was less sensitive than HPV for precancer detection and had limited value as an addition to HPV. We observed somewhat higher sensitivity of cytology in vaccinated compared with unvaccinated women, but sexual behavior, size of lesions, and virology did not appear to explain it (data not shown). Overall, using HC2, the HPV positivity of ASC-US was 52%, consistent with cytology with proper sensitivity and specificity balance.

A global meta-analysis reported the sensitivity of HPV testing and cytology for CIN3+ detection was greater than 90% and greater than 70% (31), respectively, but reported sensitivity of cytology from Latin American laboratories has been lower (34). Sensitivity of HPV testing in our study was lower than expected, possibly resulting from the extensive management of abnormalities and detection of cumulative precancers. However, performance was similar when restricting to cases detected during the first year of colposcopy. Also, both groups had been extensively screened, and many of the HPV infections are likely incident, possibly affecting test performance. Furthermore, genotyping was conducted with a more sensitive test than those in clinical use, possibly influencing modalities including genotyping. Nevertheless, the lower sensitivity observed should not have an impact on the comparison of clinical performance of each screening modality between the unvaccinated and the vaccinated groups, which was the main objective of our study, and no impact on the ranking of metrics among screening modalities.

Part of the explanation of the similar PPV in 2 groups is probably that women were vaccinated between ages 18 and 25 years, emulating catch-up, and most vaccinated women (82%) were sexually active at vaccination, with 23.0% of HR-HPV positivity, 6.2% of HPV16/18 infection and thus higher risk of precancer than cohorts of girls vaccinated younger. Some studies have demonstrated that PPV of cytology decreases among women vaccinated at younger ages (5,7,15).

Restricting to the 8 most carcinogenic HPV types (8-HPV genotyping) for primary screening and 3 triage methods using cytology alone or combined with HPV16/18 or 8-HPV genotyping for HPV-positive women yielded increased specificity and a decreased colposcopy referral compared with HR-HPV testing in vaccinated and unvaccinated women. However, as expected, 8-

Table 4. Cumulative risks of CIN3+ over 4 years by screening results and vaccination status^a

Screening results	UCG		HPV-vaccinated group		Ratio compared with UCG (95% CI)
	No. of CIN3+/No. of women	% (95% CI)	No. of CIN3+/No. of women	% (95% CI)	
Negative cytology	26/1857	1.4 (0.9 to 2.0)	21/2043	1.0 (0.7 to 1.5)	0.73 (0.41 to 1.30)
ASC-US/HC2-	3/73	4.1 (1.1 to 10.8)	0/82	0.0 (0.0 to 3.6)	0.00 (NA)
ASC-US/HC2+	8/84	9.5 (4.5 to 17.3)	7/85	8.2 (3.7 to 15.6)	0.86 (0.33 to 2.28)
LSIL	8/90	8.9 (4.2 to 16.2)	9/98	9.2 (4.6 to 16.2)	1.03 (0.42 to 2.56)
HSIL+	4/13	30.8 (10.6 to 58.7)	5/12	41.7 (17.2 to 69.8)	1.35 (0.47 to 3.89)
HR-HPV negative	14/1703	0.8 (0.5 to 1.3)	11/1926	0.6 (0.3 to 1.0)	0.69 (0.32 to 1.53)
HR-HPV negative and ASC-US+	3/105	2.9 (0.7 to 7.6)	1/121	0.8 (0.0 to 4.0)	0.29 (0.03 to 2.74)
Both HR-HPV and cytology negative	10/1521	0.7 (0.3 to 1.2)	9/1727	0.5 (0.3 to 1.0)	0.79 (0.32 to 1.95)
HR-HPV positive and ASC-US+	20/154	13.0 (8.3 to 19.0)	20/156	12.8 (8.2 to 18.8)	0.99 (0.55 to 1.76)
HR-HPV positive and cytology negative	16/336	4.8 (2.8 to 7.5)	12/315	3.8 (2.1 to 6.4)	0.80 (0.38 to 1.66)
HPV16/18 positive	16/138	11.6 (7.0 to 17.8)	4/24	16.7 (5.5 to 35.5)	1.44 (0.53 to 3.93)
Non-HPV16/18 HR-HPV positive and ASC-US+	9/110	8.2 (4.1 to 14.5)	19/149	12.8 (8.1 to 18.9)	1.56 (0.73 to 3.31)
Non-HPV16/18 HR-HPV positive and cytology negative	12/245	4.9 (2.7 to 8.2)	9/299	3.0 (1.5 to 5.5)	0.61 (0.26 to 1.43)

^aASC-US+ = atypical squamous cells of undetermined significance or worse; CI = confidence interval; CIN3+ = cervical intraepithelial neoplasia grade 3 or worse; HC2 = Hybrid Capture 2; HPV = human papillomavirus; HR-HPV = high-risk human papillomavirus; HSIL+ = high-grade squamous intraepithelial lesion or worse; LSIL = low-grade squamous intraepithelial lesion; NA = not applicable; UCG = unvaccinated control group.

HPV genotyping had lower sensitivity compared with HR-HPV testing because it misses precancers by other HPV types.

Triaging using 8-HPV genotyping and reflex cytology had better sensitivity than other approaches, with the expected loss in specificity and higher referrals. Yet, HPV genotyping needs reassessment among girls vaccinated before sexual debut attaining screening age, where reductions in infections and precancers are likely more pronounced (35-37). In fact, in our study, all HPV16/18-associated CIN3s in the vaccinated cohort were present at vaccination. More sensitive and specific biomarkers for triage, such as p16/Ki-67 dual staining and host and viral methylation, among others, are needed (21,38-43). Artificial intelligence algorithms using cervical images offer promise for triage and possibly for primary screening (44).

Cumulative risk of CIN3+ among vaccinated women was approximately 20% to 30% lower compared with unvaccinated women following negative cytology, negative HR-HPV, or both cytology and HR-HPV negative. However, absolute cumulative risk of CIN3+ appears relatively high, likely because our participants were at peak ages of CIN3 incidence (45) and some underwent colposcopic follow-up for years after initial referral. Due to this possible overestimate, we could not assess the impact of extending screening intervals for screen-negative women (9). Interestingly, cumulative risk estimates were comparable restricting to HPV-negative women at vaccination.

The main strengths of this study include comprehensive evaluation of screening modalities in a relatively large cohort of vaccinated and unvaccinated women reaching screening ages and a reliable gold standard with expert-reviewed diagnoses. However, groups were not randomly assigned and had a different screening history that we attempted to make equal by intensively screening the UCG (23). In addition, colposcopic referral was triggered during initial follow-up by cytology and not HPV test, possibly introducing verification bias. However, at the last visit, women were also referred based on HPV results, allowing capture of previously missed cases in women with only HR-HPV positive, to reduce the possibility of bias (46,47). Some verification bias may remain if probability of colposcopic referral is higher for women with abnormal cytology compared with HPV.

Our findings concur with recommendations favoring HPV testing for primary screening among women vaccinated as young adults. More research is needed to inform screening guidelines in HPV-vaccinated cohorts, especially among those vaccinated as young girls, because differences in baseline cytologic abnormalities, vaccination coverage, vaccination age, vaccine types, and sexual behaviors will affect performance of screening methods. The finding of limited impact of vaccination on the PPV of screening warrants further exploration in other datasets and can provide important clues to define research priorities and for design of future screening programs.

Funding

The Costa Rica HPV Vaccine Trial is a long-standing collaboration between investigators in Costa Rica and the National Cancer Institute (NCI). The trial is sponsored and funded by the NCI (contract N01-CP-11005), with funding support from the National Institutes of Health Office of Research on Women's Health. GlaxoSmithKline Biologicals provided vaccine and support for aspects of the trial associated with regulatory submission needs of the company under a Clinical Trials Agreement (FDA BB-IND 7920) during the 4-year, randomized blinded phase of our study. The long-term follow-up was funded by the NCI with support from the National Institutes of Health Offices of Research on Women's Health. Registered with Clinicaltrials.gov NCT00128661. SYH was supported by the Short Term Scientist Exchange Program (STSEP) at the NCI Center for Global Health (CGH) and by the grant from China Scholarship Council (CSC, grant No. 201808110125) during her collaborative research exchange visit at the NCI.

Notes

Role of the funder: The funders had no role in the design and conduct of the study; collection, management, analysis, and

interpretation of the data; and preparation of the manuscript. The NCI approved a final version of this manuscript.

Disclosures: JTS and DRL report that they are named inventors on US Government-owned HPV vaccine patents that are licensed to GlaxoSmithKline and Merck and for which the NCI receives licensing fees. They were entitled to limited royalties as specified by federal law. The NCI has received screening results, equipment, and supplies at no cost from Roche and BD for independent evaluations of screening test performance. SHT is now an employee of Merck Sharp & Dohme (MSD) Corp., a subsidiary of Merck & Co., Inc., Kenilworth, NJ, USA, but completed all work associated with this manuscript while employed at the NCI. MSD was not involved in this work. The other authors declare that they have no conflicts of interest. DRL and MHG, who are JNCI Associate Editors, were not involved in the editorial review or decision to publish this manuscript.

Author contributions: SYH: conceptualization; formal analysis; investigation; methodology; project administration; writing - original draft; writing - review & editing. ARK: conceptualization; data curation; formal analysis; funding acquisition; investigation; methodology; project administration; supervision; writing - review & editing. CP: data curation; investigation; methodology; writing - review & editing. TMD: histopathology interpretation, data curation; writing - review & editing. ACR: data curation; supervision; writing - review & editing. MHS: histopathology interpretation, data curation; writing - review & editing. DG: histopathology interpretation, data curation; writing - review & editing. MA: cytopathology interpretation, data curation; writing - review & editing. LFV: colposcopy, data curation; writing - review & editing. RO: supervision, data curation; writing - review & editing. MS: histopathology interpretation, supervision; writing - review & editing. SHT: writing - review & editing. DRL: funding acquisition; supervision; writing - review & editing. JTS: funding acquisition; investigation; supervision; writing (review and editing). JS: data curation; formal analysis; methodology; project administration; writing - review & editing. WQ and MHG: methodology; writing - review & editing. JNS: conceptualization; formal analysis; investigation; methodology; supervision; writing - review & editing. AH: conceptualization; formal analysis; funding acquisition; investigation; methodology; project administration; supervision; writing - review & editing. RH: conceptualization; formal analysis; methodology; supervision; project administration; writing - original draft; writing - review & editing.

Disclaimers: Where authors are identified as personnel of the International Agency for Research on Cancer (IARC)-WHO, the authors alone are responsible for the views expressed in this article and they do not necessarily represent the decisions, policy, or views of the IARC-WHO.

Acknowledgements: We dedicate this work to the memory of our beloved colleague and friend Paula Gonzalez, the principal investigator of the Costa Rica Vaccine Trial long-term follow-up study. We extend a special thanks to the women of Guanacaste and Puntarenas, Costa Rica, who gave of themselves in participating in this effort. In Costa Rica, we acknowledge the tremendous effort and dedication of the staff involved in this project; we would like to specifically acknowledge the meaningful contributions by Carlos Avila, Loretto Carvajal, and Cristian Montero. In the United States, we extend our appreciation to the team from Information Management Services (IMS) responsible for the development and maintenance of the data system

used in the trial and who serve as the data management center for this effort, especially Jean Cyr, Julie Buckland, and Brian Befano. We thank Dr Diane Solomon (CVT: medical monitor & QC pathologist) for her invaluable contributions during the trial and Nora Macklin and Kate Torres for the expertise in coordinating the study.

The Costa Rica HPV Vaccine Trial (CVT) Group members: Bernal Cortés, Paula González (deceased), Rolando Herrero, Silvia E. Jiménez, Carolina Porras, Ana Cecilia Rodríguez (Agencia Costarricense de Investigaciones Biomédicas, ACIB, formerly Proyecto Epidemiológico Guanacaste, PEG, Fundación INCIENSA, San José, Costa Rica); Allan Hildesheim, Aimée R. Kreimer, Douglas R. Lowy, Mark Schiffman, John T. Schiller, Mark Sherman, Sholom Wacholder (deceased) (United States National Cancer Institute, Bethesda, MD, USA); Ligia A. Pinto, Troy J. Kemp (Leidos Biomedical Research, Inc., Frederick National Laboratory for Cancer Research, Frederick, MD, USA); Mary K. Sidawy, (Georgetown University, Washington, DC, USA); Wim Quint, Leen-Jan van Doorn, Linda Struijk (Viroclinics-DDL, Netherlands); Joel M. Palefsky, Teresa M. Darragh (University of California, San Francisco, CA, USA); Mark H. Stoler (University of Virginia, Charlottesville, VA, USA).

Data Availability

Outside collaborators can apply to access our protocols and data from the blinded phase of the Costa Rica Vaccine Trial (CVT, NCT00128661) for research purposes in accordance with institutional review boards on reasonable request to the Working Group of CVT. Data for the long term follow-up phase are not yet available. A trial summary, current publications, and contact information are available online at <https://dceg.cancer.gov/research/who-we-study/cohorts/costa-rica-vaccine-trial>.

References

1. Ferlay J, Ervik M, Lam F, et al. Global cancer observatory: cancer today. <https://gco.iarc.fr/today>. Accessed December 24, 2020.
2. Marks MA, Castle PE, Schiffman M, et al. Evaluation of any or type-specific persistence of high-risk human papillomavirus for detecting cervical precancer. *J Clin Microbiol*. 2012;50(2):300-306.
3. Brisson M, Kim JJ, Canfell K, et al. Impact of HPV vaccination and cervical screening on cervical cancer elimination: a comparative modelling analysis in 78 low-income and lower-middle-income countries. *Lancet*. 2020;395(10224):575-590.
4. World Health Organization. Introduction of HPV (Human Papilloma Virus) vaccine. https://immunizationdata.who.int/pages/vaccine-intro-by-antigen/hpv.html?ISO_3_CODE=&YEAR=. Accessed March 19, 2022.
5. Lei J, Ploner A, Lehtinen M, et al. Impact of HPV vaccination on cervical screening performance: a population-based cohort study. *Br J Cancer*. 2020;123(1):155-160.
6. Palmer TJ, McFadden M, Pollock KG, et al. HPV immunisation and cervical screening—confirmation of changed performance of cytology as a screening test in immunised women: a retrospective population-based cohort study. *Br J Cancer*. 2016;114(5):582-589.
7. Hestbech MS, Lynge E, Kragstrup J, et al. The impact of HPV vaccination on future cervical screening: a simulation study of two birth cohorts in Denmark. *BMJ Open*. 2015;5(8):e007921.
8. Leeftang MM, Rutjes AW, Reitsma JB, et al. Variation of a test's sensitivity and specificity with disease prevalence. *CMAJ*. 2013;185(11):E537-E544.
9. Cheung LC, Egemen D, Chen X, et al. 2019 ASCCP risk-based management consensus guidelines: methods for risk estimation, recommended management, and validation. *J Low Genit Tract Dis*. 2020;24(2):90-101.
10. WHO. *WHO Guideline for Screening and Treatment of Cervical Pre-Cancer Lesions for Cervical Cancer Prevention*. 2nd ed. Geneva, Switzerland: World Health Organization; 2021.
11. Perkins RB, Guido RS, Castle PE, et al.; 2019 ASCCP Risk-Based Management Consensus Guidelines Committee. 2019 ASCCP risk-based management consensus guidelines for abnormal cervical cancer screening tests and cancer precursors. *J Low Genit Tract Dis*. 2020;24(2):102-131.
12. Fontham ETH, Wolf AMD, Church TR, et al. Cervical cancer screening for individuals at average risk: 2020 guideline update from the American Cancer Society. *CA A Cancer J Clin*. 2020;70(5):321-346.

13. Palmer T, Wallace L, Pollock KG, et al. Prevalence of cervical disease at age 20 after immunisation with bivalent HPV vaccine at age 12–13 in Scotland: retrospective population study. *BMJ*. 2019;365:11161.
14. Rodriguez AC, Solomon D, Herrero R, et al.; for the CVT Group. Impact of human papillomavirus vaccination on cervical cytology screening, colposcopy, and treatment. *Am J Epidemiol*. 2013;178(5):752–760.
15. Castle PE, Xie X, Xue X, et al. Impact of human papillomavirus vaccination on the clinical meaning of cervical screening results. *Prev Med*. 2019;118:44–50.
16. Thamsborg LH, Napolitano G, Larsen LG, et al. Impact of HPV vaccination on outcome of cervical cytology screening in Denmark—a register-based cohort study. *Int J Cancer*. 2018;143(7):1662–1670.
17. Cruickshank ME, Pan J, Cotton SC, et al. Reduction in colposcopy workload and associated clinical activity following human papillomavirus (HPV) catch-up vaccination programme in Scotland: an ecological study. *BJOG*. 2017;124(9):1386–1393.
18. Kim JJ, Burger EA, Sy S, et al. Optimal cervical cancer screening in women vaccinated against human papillomavirus. *JNCI J Natl Cancer Inst*. 2017;109(2):djw216.
19. Lew JB, Simms KT, Smith MA, et al. Primary HPV testing versus cytology-based cervical screening in women in Australia vaccinated for HPV and unvaccinated: effectiveness and economic assessment for the National Cervical Screening Program. *Lancet Public Health*. 2017;2(2):e96–e107.
20. Pedersen K, Burger EA, Nygard M, et al. Adapting cervical cancer screening for women vaccinated against human papillomavirus infections: the value of stratifying guidelines. *Eur J Cancer*. 2018;91:68–75.
21. Canfell K, Caruana M, Gebski V, et al. Cervical screening with primary HPV testing or cytology in a population of women in which those aged 33 years or younger had previously been offered HPV vaccination: results of the Compass pilot randomised trial. *PLoS Med*. 2017;14(9):e1002388.
22. Herrero R, Hildesheim A, Rodriguez AC, et al.; Costa Rica Vaccine Trial (CVT) Group. Rationale and design of a community-based double-blind randomized clinical trial of an HPV 16 and 18 vaccine in Guanacaste, Costa Rica. *Vaccine*. 2008;26(37):4795–4808.
23. Gonzalez P, Hildesheim A, Herrero R, et al.; Costa Rica HPV Vaccine Trial (CVT) Group. Rationale and design of a long term follow-up study of women who did and did not receive HPV 16/18 vaccination in Guanacaste, Costa Rica. *Vaccine*. 2015;33(18):2141–2151.
24. Solomon D, Davey D, Kurman R, et al.; Bethesda 2001 Workshop. The 2001 Bethesda System: terminology for reporting results of cervical cytology. *JAMA*. 2002;287(16):2114–2119.
25. Castle PE, Solomon D, Wheeler CM, et al. Human papillomavirus genotype specificity of hybrid capture 2. *J Clin Microbiol*. 2008;46(8):2595–2604.
26. Iftner T, Neis KJ, Castanon A, et al. Longitudinal clinical performance of the RNA-based aptima human papillomavirus (AHPV) assay in comparison to the DNA-based hybrid capture 2 HPV test in two consecutive screening rounds with a 6-year interval in Germany. *J Clin Microbiol*. 2019;57(1):e01177–e01118.
27. Wagner S, Roberson D, Boland J, et al.; CVT Group. Evaluation of TypeSeq, a novel high-throughput, low-cost, next-generation sequencing-based assay for detection of 51 human papillomavirus genotypes. *J Infect Dis*. 2019;220(10):1609–1619.
28. Drolet M, Benard E, Perez N, et al.; HPV Vaccination Impact Study Group. Population-level impact and herd effects following the introduction of human papillomavirus vaccination programmes: updated systematic review and meta-analysis. *Lancet*. 2019;394(10197):497–509.
29. Schiffman M, Hyun N, Raine-Bennett TR, et al. A cohort study of cervical screening using partial HPV typing and cytology triage. *Int J Cancer*. 2016;139(11):2606–2615.
30. Smith JS, Lindsay L, Hoots B, et al. Human papillomavirus type distribution in invasive cervical cancer and high-grade cervical lesions: a meta-analysis update. *Int J Cancer*. 2007;121(3):621–632.
31. Koliopoulos G, Nyaga VN, Santesso N, et al. Cytology versus HPV testing for cervical cancer screening in the general population. *Cochrane Database Syst Rev*. 2017;8(8):CD008587.
32. Wright TC Jr, Stoler MH, Behrens CM, et al. Interlaboratory variation in the performance of liquid-based cytology: insights from the ATHENA trial. *Int J Cancer*. 2014;134(8):1835–1843.
33. Pan QJ, Hu SY, Guo HQ, et al. Liquid-based cytology and human papillomavirus testing: a pooled analysis using the data from 13 population-based cervical cancer screening studies from China. *Gynecol Oncol*. 2014;133(2):172–179.
34. Ferreccio C, Barriga MI, Lagos M, et al. Screening trial of human papillomavirus for early detection of cervical cancer in Santiago, Chile. *Int J Cancer*. 2013;132(4):916–923.
35. Herrero R, Wacholder S, Rodriguez AC, et al.; Costa Rica Vaccine Trial Group. Prevention of persistent human papillomavirus infection by an HPV16/18 vaccine: a community-based randomized clinical trial in Guanacaste, Costa Rica. *Cancer Discov*. 2011;1(5):408–419.
36. Bhatia R, Kavanagh K, Cubie HA, et al. Use of HPV testing for cervical screening in vaccinated women—insights from the SHEVa (Scottish HPV Prevalence in Vaccinated Women) study. *Int J Cancer*. 2016;138(12):2922–2931.
37. Arbyn M, Xu L, Simoons C, et al. Prophylactic vaccination against human papillomaviruses to prevent cervical cancer and its precursors. *Cochrane Database Syst Rev*. 2018;5(5):CD009069.
38. Stanczuk GA, Baxter GJ, Currie H, et al. Defining optimal triage strategies for hrHPV screen-positive women—an evaluation of HPV 16/18 genotyping, cytology, and p16/Ki-67 cytoimmunochemistry. *Cancer Epidemiol Biomarkers Prev*. 2017;26(11):1629–1635.
39. Verhoef VM, Bosgraaf RP, van Kemenade FJ, et al. Triage by methylation-marker testing versus cytology in women who test HPV-positive on self-collected cervicovaginal specimens (PROHTECT-3): a randomised controlled non-inferiority trial. *Lancet Oncol*. 2014;15(3):315–322.
40. Clarke MA, Luhn P, Gage JC, et al. Discovery and validation of candidate host DNA methylation markers for detection of cervical precancer and cancer. *Int J Cancer*. 2017;141(4):701–710.
41. Hernandez-Lopez R, Lorincz AT, Torres-Ibarra L, et al.; FRIDA Study Group. Methylation estimates the risk of precancer in HPV-infected women with discrepant results between cytology and HPV16/18 genotyping. *Clin Epigenet*. 2019;11(1):140.
42. Rezhake R, Hu SY, Zhao S, et al. Eight-type human papillomavirus E6/E7 oncoprotein detection as a novel and promising triage strategy for managing HPV-positive women. *Int J Cancer*. 2019;144(1):34–42.
43. Ferrera A, Valladares W, Cabrera Y, et al. Performance of an HPV 16/18 E6 oncoprotein test for detection of cervical precancer and cancer. *Int J Cancer*. 2019;145(8):2042–2050.
44. Hu L, Bell D, Antani S, et al. An observational study of deep learning and automated evaluation of cervical images for cancer screening. *J Natl Cancer Inst*. 2019;111(9):923–932.
45. Bruni L, Albero G, Serrano B, et al. ICO/IARC Information Centre on HPV and Cancer (HPV Information Centre). Human Papillomavirus and Related Diseases in Costa Rica. Summary Report 22 October 2021. <https://hvpcentre.net/statistics/reports/CRI.pdf?t=1654948289376>. Accessed December 11, 2021.
46. Ogilvie GS, van Niekerk D, Krajden M, et al. Effect of screening with primary cervical HPV testing vs cytology testing on high-grade cervical intraepithelial neoplasia at 48 months: the HPV FOCAL randomized clinical trial. *JAMA*. 2018;320(1):43–52.
47. Zhang J, Zhao Y, Dai Y, et al. Effectiveness of high-risk human papillomavirus testing for cervical cancer screening in China: a multicenter, open-label, randomized clinical trial. *JAMA Oncol*. 2021;7(2):263–270.