

# UCSF

## UC San Francisco Previously Published Works

### Title

Hematopoietic Stem Cell Transplantation for Severe Combined Immunodeficiency

### Permalink

<https://escholarship.org/uc/item/61k712m7>

### Journal

Current Pediatrics Reports, 3(1)

### ISSN

2167-4841

### Authors

Wahlstrom, Justin T  
Dvorak, Christopher C  
Cowan, Morton J

### Publication Date

2015-03-01

### DOI

10.1007/s40124-014-0071-7

Peer reviewed



Published in final edited form as:

*Curr Pediatr Rep.* 2015 March 1; 3(1): 1–10. doi:10.1007/s40124-014-0071-7.

## Hematopoietic Stem Cell Transplantation for Severe Combined Immunodeficiency

**Justin T. Wahlstrom, M.D.**

505 Parnassus Ave, Room M-659, San Francisco, CA 94143, Office: (415) 476-2188, Fax: Fax: (415) 502-4867

**Christopher C. Dvorak, M.D.**, and

505 Parnassus Ave, Room M-659, San Francisco, CA 94143, Office: (415) 476-2188, Fax: Fax: (415) 502-4867

**Morton J. Cowan, M.D**

505 Parnassus Ave, Room M-659, San Francisco, CA 94143, Office: (415) 476-2188, Fax: Fax: (415) 502-4867

Justin T. Wahlstrom: wahlstromj@peds.ucsf.edu; Christopher C. Dvorak: dvorakc@peds.ucsf.edu; Morton J. Cowan: mcowan@peds.ucsf.edu

### Abstract

Hematopoietic stem cell transplantation (HSCT) is an effective approach for the treatment of severe combined immunodeficiency (SCID). However, SCID is not a homogeneous disease, and the treatment required for successful transplantation varies significantly between SCID subtypes and the degree of HLA mismatch between the best available donor and the patient. Recent studies are beginning to more clearly define this heterogeneity and how outcomes may vary. With a more detailed understanding of SCID, new approaches can be developed to maximize immune reconstitution, while minimizing acute and long-term toxicities associated with chemotherapy conditioning.

### Introduction

For patients with severe combined immunodeficiency (SCID), there are several treatment options. Hematopoietic stem cell transplantation (HSCT) is potentially curative for all patients with SCID. Gene therapy (GT) may also be curative and currently is available on an experimental basis for eligible patients with IL2RG and adenosine deaminase deficiency (ADA) SCID (see Calero et al., review in this series). Finally, enzyme replacement therapy (ERT) is supportive therapy for patients with ADA-SCID and is often used while awaiting HSCT or GT. For the majority of patients meeting the diagnostic criteria for SCID,(1, 2)

**Compliance with Ethics Guidelines: Conflicts of Interest:** Justin T. Wahlstrom and Christopher C. Dvorak declare that they have no conflict of interest.

Mortan J.Cowan reports personal fees from Bluebird Bio, Inc. and served on the Scientific Advisory board for Exogen Bio, Inc.

**Human and Animal Rights and Informed Consent:** This article does not contain any studies with human or animal subjects performed by any of the authors.

HSCT offers the most widely available approach to an effective cure and will be the focus of this chapter.

SCID is a rare disease with an incidence of 1:58000 (3) and most centers treating patients with SCID only see on average one patient per year. Thus, it is very difficult if not impossible for any single center to develop optimal treatment approaches for this disorder. Many of the recent advances in our understanding of the diagnosis of SCID and issues surrounding treatment for SCID are a result of collaborative working groups coordinated by the The Inborn Errors Working Party (IEWP) of the European Society for Blood and Marrow Transplant (EBMT), (IEWP/EBMT) and PIDTC (Primary Immune Deficiency Treatment Consortium – a group of 44 centers in North America established in 2009 to study the definitive treatment of PIDs). These issues and their effects on immunologic and survival outcomes are discussed below. Specific considerations for SCID subtypes are outlined in Table 1.

## Timing of HSCT

In general, SCID patients who proceed to HSCT earlier in life have superior outcomes compared to those transplanted later.(5-7) The PIDTC recently performed a retrospective analysis of the largest cohort of SCID patients in North America published to date.(8) In this study, the likelihood of having an active infection at the time of transplant was significantly higher (52%) for patients transplanted at >3.5 months of age (old), compared to those transplanted at <3.5 months of age (young) (22%). Old infants with active infection had significantly ( $p<0.001$ ) poorer 5 year survival (50%) versus young infants with/without infection (94%), old infants with no infection (90%), or infants whose infection cleared by the time of HSCT (82%).

The widespread use of newborn screening (NBS) by T-cell receptor excision circle (TREC) quantification has greatly shortened time to diagnosis, making treatment possible at a very early age.(3, 9, 10) (see Kwan et al., review in this series). In one prospective study, patients diagnosed by family history or NBS proceeded to HSCT at a much younger median age of 67 days (and 74% received HSCT at <3.5 months of age), compared to a median of 214 days (with only 17% receiving HSCT at <3.5 months of age) when diagnosed by clinical signs. (11). In cases where chemotherapy-based conditioning regimens are required, caution is recommended, as there is a likely risk of increased early and late toxicities (discussed below) in these very young patients treated with alkylating agents. However, available data indicate that, regardless of age, patients should be transplanted before the development of an infection if at all possible.

## Conditioning for HSCT

In addition to recipient age, donor type and SCID type may influence the decision regarding conditioning. For example, matched sibling donors (MSDs), matched unrelated donors (URDs), and haploidentical maternal donors in which there is maternal chimerism in the recipient, all engraft readily without conditioning, although the likelihood of reconstituting B cell immunity decreases with each kind of donor as HLA mismatch increases.(12, 13) Additionally, SCID type may influence the decision regarding conditioning; some types are

much more likely than others to recover B cell function without conditioning, as host B-cells in some SCID types can regain function with competent T-cell help. In a cohort of patients receiving unconditioned haploidentical or matched sibling donor transplants, patients with defects in IL7R $\alpha$ , ADA, and CD3 were all more likely to recover B cell immunity compared to those with IL2RG or RAG1/2 defects.(14) Conversely, in IL2RG and JAK3-deficiencies, phenotypically normal B cells are produced but are unable to function despite adequate T-cell help.(15) An advantage of using conditioning, particularly in uninfected patients, is the increased chance of achieving donor myeloid chimerism (62% vs 20%, p=0.007), and being off immunoglobulin replacement therapy in patients surviving beyond 2 years post HSCT (O.R. of 8.9; 84% vs 41%, p<0.001).(8) Overall, the highest rates of B cell function have consistently been observed in studies using myeloablative conditioning (MAC).(16-18)

While not absolutely necessary for T cell reconstitution, myeloid chimerism is also known to strongly predict recovery of thymic function as measured by both TREC levels and CD45RA+ circulating T cells, which reflect thymopoiesis and T-cell receptor (TCR) diversity.(19, 20) This was observed in a subset of PIDTC study patients analyzed for naïve CD4+CD45RA+ reconstitution; 100% of patients who received conditioning developed naive T cells >100/uL, but this was less common in those who did not receive conditioning (approximately 50%).(8) In addition, the use of a conditioning regimen was associated with an 8.8-fold increased chance of attaining a CD3 count >1000/mm<sup>3</sup> at 2-5 years (89% vs 62%, p=0.007).(8) However, in some SCID subtypes (i.e. IL2RG and ADA defects) T cell recovery appears to be equivalent in MSD versus URD transplants even without the use of conditioning.(12)

In terms of which chemotherapy agents are most optimal for SCID patients who may require conditioning, there are several options. Busulfan-based regimens are most commonly used in both reduced-intensity (RIC) and myeloablative (MAC) transplants for SCID, although alternatives, i.e., melphalan-based regimens, have been used with comparable results.(8) Melphalan has been used successfully in a reduced intensity regimen, although metabolism of melphalan in infants is not well defined, and should be used with caution, if at all, in patients <6 months of age (P. Veys, personal communications, European group for Blood and Marrow Transplant guidelines).(21, 22) Treosulfan (not yet approved in the US), which is associated with less veno-occlusive disease and CNS toxicity than busulfan, has also been used in infants, although its metabolism in this age has not been well-studied.(22) Fludarabine is often used in combination with alkylating agents to provide T lymphocyte ablation, but it may not be sufficient to fully eradicate host NK cells in mismatched HSCT and should be used with some care in infants <6 months old.(23)

Serotherapy (anti-thymocyte globulin [ATG] or alemtuzumab [CAMPATH-1H]) is also frequently administered prior to stem cell infusion as part of conditioning, with or without chemotherapy. With its relatively long half-life, serotherapy provides host immunoablation (reducing the risk of rejection), as well as GVHD prophylaxis. Serotherapy with post- HSCT GVHD prophylaxis is essential in URD transplants for SCID patients to reduce the incidence and severity of GVHD.(12) It may be used in combination with ex vivo T cell depletion (TCD) prior to haploidentical HCT, but the dosing must be carefully considered to avoid prolonged T cell recovery.(24)

In summary, the use of conditioning remains controversial. It is clearly associated with poorer survival in patients with infection at the time of HSCT(8), and the safety in uninfected patients is not fully established. While it may result in enhanced T and B cell reconstitution, this may be at the cost of early toxicity and late effects, a question which has not been carefully studied.

## Donor Selection

An approach at our center in UCSF for determining the most appropriate donor and conditioning regimen for patients with SCID is outlined in Figure 1. Regardless of SCID type, all siblings should be HLA-typed as soon as the diagnosis is made. Because of the high incidence of shared HLA haplotypes within some SCID populations, and the potential need for haploidentical donors, parents should also be typed.

## Matched Sibling Donors

An HLA-matched sibling is the donor of choice for all SCID types. In the 2014 PIDTC analysis, for patients receiving a MSD transplant, 5 year survival far exceeded that of any other donor type, at 97%.(8)

For patients with typical SCID (T cells  $<300/\mu\text{L}$  and T cell function  $<10\%$  lower limit of normal),(2) most centers do not use a conditioning regimen for MSD transplants, regardless of SCID type, as the risk of rejection and severe GVHD are quite low. As shown in Table 2, excellent survival rates have been observed in studies where a conditioning regimen was not used, ranging from 89-95%.(7, 12, 25) Even if maternally engrafted cells are present, eradication of these cells prior to unconditioned MSD HSCT is not necessary.(12) Generally, for MSD, conditioning is only required for patients with reticular dysgenesis, leaky SCID (T cells  $<1000/\mu\text{L}$ , T cell function  $<30\%$  lower limit of normal, and absence of maternal engraftment) or Omenn syndrome, which often requires some degree of myeloablation to correct the associated inflammatory and autoimmune manifestations, regardless of donor type. To date, there are no data comparing conditioned vs unconditioned HSCT for patients with SCID receiving a MSD.

Immune reconstitution for patients receiving MSD transplants is largely determined by SCID subtype, as noted above. In the PIDTC analysis, where over 87% of MSD HSCT's were unconditioned, recovery of T cells ( $>1000/\text{mm}^3$  CD3) was achieved in 76% of patients, and B cell function was observed in 81% of patients.(8)

Acute GVHD, a reaction of donor immune cells against host tissues which classically occurs within 100 days of stem cell infusion, is not uncommon in transplants for SCID. Its incidence is modified primarily by donor-recipient HLA disparity as well as the use of GVHD prophylaxis. In MSD transplants, some centers omit GVHD prophylaxis entirely, while others administer a post-transplant calcineurin inhibitor with or without methotrexate, with the rationale that GVHD is of little benefit to a patient with SCID. Although the overall incidence of acute GVHD ranges from 23–50% in MSD HSCT for SCID, Grade III-IV aGVHD and chronic GVHD are uncommon even in two studies where many patients received no GVHD prophylaxis (3-6%).(8, 12)

## Unrelated Donors

In the absence of a MSD, an URD search should be performed as soon as possible. Unrelated umbilical cord blood (UCB) sources should also be considered, and they are discussed below. Survival rates are typically lower with URD's compared to MSD's. This is likely due to a delay in proceeding to HSCT (by approximately 1-3 months) and subsequent immune reconstitution, an increased incidence of GVHD, and possibly due to increased use of chemotherapy-based conditioning. In considering an URD versus an available haploidentical donor, infection status at the time of transplant is an important determinant of survival. In infants of any age without an active infection, overall survival for URD HSCT is equivalent to that of haploidentical HSCT without conditioning (93% vs 91% respectively). (8) However, in the setting of an active infection at the time of transplant, overall survival for an URD HSCT is inferior compared to haploidentical HSCT without conditioning (53% versus 65%,  $p=0.006$ ). In such cases, or when the likelihood of finding a matched URD is very low (rare ethnic background, etc.), haploidentical donors without conditioning should be considered as initial therapy.

Recipients of URD transplants may receive no conditioning, reduced intensity conditioning (RIC), or myeloablative conditioning (MAC), as shown in Table 2. Overall survival appears equivalent in patients receiving URD transplants with or without conditioning. In a multicenter survey of transplants for patients with SCID (mainly IL2RG and ADA) using URD's and UCB without RIC or MAC, 5-year overall survival for URD/UCB recipients was 71%, comparable to published reports for those receiving RIC or MAC.(12) In the PIDTC cohort, HSCT using adult URDs typically included conditioning, and overall survival was 74%.(8) In a direct comparison of RIC vs MAC, a small cohort of infants with SCID (and other immunodeficiencies) receiving URD HSCT's was analyzed: RIC was associated with 94% overall survival, significantly better than with MAC (53%,  $p=0.014$ ). (21)

Overall, to determine the appropriate use of conditioning for an URD transplant, SCID type and infection risk should be considered. For patients without an intrinsic B cell defect, such as IL7R $\alpha$  deficiency, there is little advantage to using conditioning; likewise, for patients with ADA deficiency or DNA repair defects, use of conditioning has been associated with poor outcomes and is not recommended.(26, 27) However, for other patients without active infection, a RIC regimen may be considered to promote B cell reconstitution, although the late effects of chemotherapy exposure in these babies have not been well-characterized.

For URD transplants without conditioning, acute GVHD occurs in up to 73% of patients if serotherapy is not used, and 32% may experience Grade III-IV aGVHD; this is reduced to 50% (8% with Grade III-IV) with serotherapy.(12) Standard GVHD prophylaxis with a calcineurin inhibitor and methotrexate (or mycophenolate mofetil or corticosteroids in the case of UCB) is recommended for these patients. The incidence of chronic GVHD (39%) was significantly higher in URD's compared to MSD recipients (5%;  $p<0.01$ ), and was also partially abrogated by the use of serotherapy (47% vs 27%).

### Umbilical Cord Blood Donors

An alternative stem cell source for a SCID patient without a conventional MSD or adult URD is UCB. In one retrospective comparison of UCB vs haploidentical related donors, no difference in 5-year overall survival was detected (57% vs. 62% respectively,  $p=0.68$ ).<sup>(28)</sup> Although this study showed increased myeloid engraftment and IVIG independence and lower rates of second transplant (for non-engraftment) in the patients receiving UCB, this may have been related to the higher-intensity conditioning that these patients received. Of note, the degree of HLA mismatch in the UCB unit was a strong determinant of inferior overall survival. The PIDTC analysis also demonstrated that for patients <3.5 months old, UCB recipients had the lowest 5-year overall survival compared to MSD (58%,  $p=0.01$ ).<sup>(8)</sup> Based on these and prior studies<sup>(29)</sup> demonstrating that outcomes using a 4/6 HLA matched UCB unit are inferior to those with a TCD haploidentical donor, this approach is only recommended when at least a 5/6 allele-matched unit is available.

### Haploidentical Donors

If a MSD or a well-matched URD (adult or UCB) cannot be identified in a timely fashion, or if the patient has an active infection, a familial haploidentical transplant should be considered. As described above, survival using haploidentical donors without conditioning is comparable to that obtained with URD's, unless the patient has an active infection at the time of transplant, when haploidentical HCT is superior.<sup>(8)</sup> A European retrospective analysis demonstrated equivalent survival using a T cell depleted haploidentical relative (66%) compared to a URD (69%), although patients with active infection were not analyzed separately.<sup>(30)</sup>

An important parameter to consider in haploidentical transplants is the presence of transplacental maternal engraftment (TME). Maternally engrafted cells can be identified in approximately 40%<sup>(31)</sup> of SCID patients requiring transplant, and can interfere with engraftment of a paternal donor<sup>(32)</sup>. Although these maternally-derived T cells are typically functionally incompetent, often expressing a limited TCR repertoire<sup>(33)</sup> with limited or no proliferative response to mitogen<sup>(34)</sup>, they can be associated with pre-transplant GVHD.<sup>(32)</sup> If TME is present without GVHD, fetal-maternal tolerance is inferred and an unconditioned maternal T cell depleted transplant can be performed readily.<sup>(13)</sup> Engraftment is less likely if a non-maternal donor is used without conditioning in patients with circulating maternal T cells.<sup>(35)</sup> In addition, if TME is not present, at a minimum serotherapy should be considered to try to decrease the risk of rejection due to host NK cells and/or small numbers of host T cells.

For haploidentical transplants, use of any conditioning beyond serotherapy has a significant adverse effect on overall survival when used in the context of active infections (overall survival of 65% without conditioning vs 39% with conditioning,  $p=0.006$ ); this difference was diminished for infants without infection (overall survival of 91% vs 81%,  $p=0.16$ ).<sup>(8)</sup> Therefore, withholding conditioning is appropriate in most haploidentical HCTs for SCID, especially when performed for defects in IL2RG or JAK3<sup>(18)</sup> or when TME is present. However, without conditioning, engraftment is less likely in cases where early T cells or NK cells are present and there is no TME. For defects involving RAG1/2 or radiosensitive SCID

(19, 36) graft resistance may be higher, and these patients often require some immunosuppression to achieve T cell engraftment.(13) In an attempt to promote engraftment and immune reconstitution, high CD34 cell doses without busulfan or melphalan-containing conditioning was evaluated in a single-center study.(13) Out of 15 SCID patients who received at least  $20 \times 10^6$  CD34+ cells/kg, 11 engrafted without the need for 2<sup>nd</sup> transplant; 3-year overall survival was 87%, although the 4 patients who rejected received conditioning prior to a second transplant using an unrelated donor. All patients with IL2RG-deficiency or pre-HCT evidence of TME engrafted, while only 3 out of 7 patients with NK+ SCID without TME engrafted.

T-cell reconstitution in recipients of haploidentical transplants is variable. In the PIDTC study, 66% had CD3 recovery ( $>1000/\text{mm}^3$ ) at 2-5 years, compared to 76% of MSD recipients ( $p=0.01$ ) and 76% of URD recipients ( $p=0.84$ ).<sup>(8)</sup> Another study showed that the median time to T cell reconstitution (development of a PHA response  $>50\%$  of control) was 4.9 months, and that to development of a PHA response 90% of control was 9.8 months.<sup>(13)</sup> This is consistent with prior studies showing that TCD is associated with a median time to regain functional T-cells of 5-9 months.<sup>(37, 38)</sup>

Recovery of B cell function following haplocompatible HCT is also heterogeneous and highly dependent upon SCID genotype.<sup>(6, 38)</sup> In an unconditioned cohort primarily consisting of haploidentical donors (and also including some MSDs), Buckley et al. observed long-term independence from immunoglobulin supplementation in 63% of patients. This is comparable to results obtained by Haddad et al in a study of only haploidentical HSCT's, where conditioning was variable: 69% of these patients recovered B cell function at last follow up. This compares favorably to the rate of B cell reconstitution in unconditioned URD's (including UCB) where only 30% of patients were IVIG-independent at last follow-up<sup>(12)</sup>, though this latter result may be partly due to inclusion of only certain SCID genotypes.

The incidence of aGVHD in TCD haploidentical transplants, where no additional GVHD prophylaxis is given, ranges from 20% (grade II-IV) to 58% (all grade I-II).<sup>(8, 13)</sup> Chronic GVHD has been observed in up to 40% of these patients at 6 months post-transplant, but incidence at two years is relatively low (13-18%).<sup>(38)</sup>

## Toxicity

Treatment-related mortality (TRM) occurred in 26% of patients analyzed in the PIDTC series, mostly in patients diagnosed and transplanted at  $>3.5$  months of age, and 38% of the mortality was due to infection.<sup>(8)</sup> However, much of the infection-related mortality seen in HSCT for SCID may be due to infections that occurred prior to diagnosis and HSCT. With the advent of NBS for SCID, TRM due to infection should decrease significantly. Pulmonary toxicity was the other primary contributor to TRM in this study, accounting for 37% of mortality in the entire cohort, and 64% of mortality in patients receiving MAC with busulfan-based regimens.<sup>(8)</sup> A primary risk factor for pulmonary toxicity in SCID patients is pre-existing lung infection (CMV pneumonitis, PCP pneumonia). The pulmonary toxicity



of busulfan, cyclophosphamide or fludarabine on young infants <6 months of age is not yet fully understood and requires further study.(39)

Long-term survival has increased over the last 5 decades among all SCID types.(30, 40) If good T-cell function is achieved within 1-2 years after HSCT, function is very likely to be maintained long-term (20, 41) although some reports suggest that T cell immunity may decline over time in some patients not receiving conditioning.(5) Because long-term survival is expected for most SCID patients, late effects need to be considered carefully. At least two studies have evaluated the late effects of conditioning in SCID patients overall (41) and in patients with RAG- or Artemis-deficient SCID.(42) In the latter study, patients with radiosensitive SCID due to Artemis-deficiency had significantly more problems beyond 2 years post-transplant with growth, tooth development and endocrinopathies when exposed to alkylating chemotherapy prior to HSCT. Organ toxicity (specifically to heart, lungs, and the central nervous system) should be monitored regularly for patients with SCID who have received chemotherapy. In patients transplanted for a variety of primary immunodeficiencies, true secondary malignancies occurred with an incidence of only 0.2% at 5-15 years.(43) However, late mortality (>2 years post-HSCT) due to infection, organ failure, and chronic GVHD still occurred in 7% of patients.(44)

The long-term cognitive function of SCID patients treated with HSCT is a subject of ongoing research. Cognitive effects are related to many factors, including the intrinsic genetic defect (e.g., ADA-SCID and some types of RS-SCID), psychosocial factors such as socioeconomic status, the duration and severity of illness / hospitalization, and possibly direct CNS toxicity from conditioning.(45, 46) Given the general observation that the incidence and severity of cognitive deficits increases when radiation is given to very young infants,(47) alkylating chemotherapy such as busulfan, which readily crosses the blood-brain-barrier, should be administered with caution. Close follow-up, including a full battery of age-appropriate neurocognitive testing, should be done biannually through childhood.

A diagnosis of SCID can also have significant psychosocial effects. Both the disease and the transplant process itself pose stressors to the patient and family. For example, since CMV may be transmitted through breastfeeding, mothers are advised to discontinue breastfeeding if they are CMV-seropositive, which can be very distressing for new mothers; early cessation of breastfeeding has been associated with post-partum depression.(48) Prolonged hospitalization after initial diagnosis is sometimes necessary, especially if the home environment poses a significant infectious risk to the infant. In addition, depending on the type of transplant, the duration of hospital admission for transplant for SCID patients can be months, adding significant stress to families. The social isolation required as part of HSCT for SCID may contribute to long-term deficits in both cognitive and motor development.(49)

## Conclusions

HSCT is a life-saving treatment for patients with SCID, especially if therapy is instituted early, prior to onset of infections. Outcomes for all types of SCID have continued to steadily improve, and collaborative studies are underway that will help to further identify areas where improvement is still needed. While much progress has been made over the years to

reduce side effects and improve survival, more studies are needed to address the questions that remain. For example, identifying the minimum dose of myeloablative conditioning that can achieve myeloid chimerism with reliable T and B-cell reconstitution is especially important for patients who are otherwise unlikely to reject even a mismatched donor graft. This question is especially salient in the era of NBS, as patients are more frequently being diagnosed at <1 month of age, and would clearly benefit from immediate transplant, were it not for the potential risks posed by alkylating chemotherapy. Newer agents, such as the stem cell targeting monoclonal antibodies against CD117 (c-kit) and CD45, hold the promise of allowing early HSCT without the risks of conventional chemotherapy, but need to be evaluated in prospective multicenter studies.(50, 51) In addition to refinement of conditioning approaches for newborns with SCID, the optimal GVHD prophylaxis regimen for both matched and mismatched donors requires further study, since GVHD is of little benefit to these patients, but GVHD prophylaxis can interfere with functional immune recovery. Future prospective studies of new conditioning regimens, GVHD prophylaxis strategies, and late effects will continue to refine HSCT as a therapeutic approach for patients with SCID.

## References

Papers of particular interest, published recently, have been highlighted as:

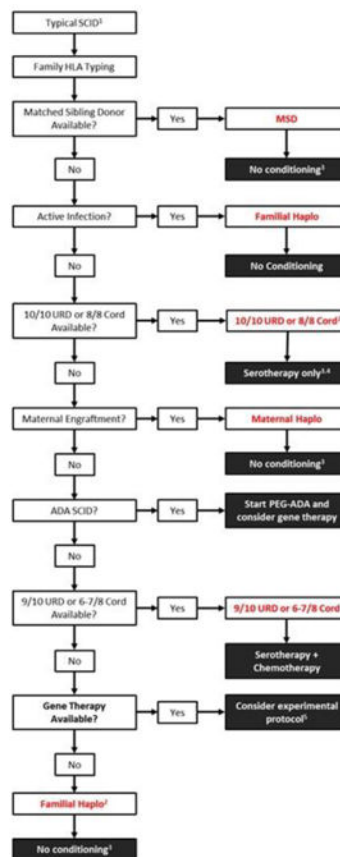
- Of importance
  - Of major importance
1. Geha RS, et al. Primary immunodeficiency diseases: an update from the International Union of Immunological Societies Primary Immunodeficiency Diseases Classification Committee. *The Journal of allergy and clinical immunology*. Oct.2007 120:776. [PubMed: 17952897]
  - 2\*. Shearer WT, et al. Establishing diagnostic criteria for severe combined immunodeficiency disease (SCID), leaky SCID, and Omenn syndrome: the Primary Immune Deficiency Treatment Consortium experience. *The Journal of allergy and clinical immunology*. Apr.2014 133:1092. Provides the rationale and detailed analysis of criteria for making a diagnosis of SCID. [PubMed: 24290292]
  3. Kwan A, et al. Newborn screening for severe combined immunodeficiency in 11 screening programs in the United States. *JAMA : the journal of the American Medical Association*. Aug 20.2014 312:729.
  4. Griffith LM, et al. Primary Immune Deficiency Treatment Consortium (PIDTC) report. *The Journal of allergy and clinical immunology*. Feb.2014 133:335. [PubMed: 24139498]
  5. Buckley RH. Transplantation of hematopoietic stem cells in human severe combined immunodeficiency: longterm outcomes. *Immunologic research*. Apr.2011 49:25. [PubMed: 21116871]
  6. Buckley RH, et al. Hematopoietic stem-cell transplantation for the treatment of severe combined immunodeficiency. *N Engl J Med*. Feb 18.1999 340:508. [PubMed: 10021471]
  7. Railey MD, Lokhnygina Y, Buckley RH. Long-term clinical outcome of patients with severe combined immunodeficiency who received related donor bone marrow transplants without pretransplant chemotherapy or post-transplant GVHD prophylaxis. *The Journal of pediatrics*. Dec. 2009 155:834. [PubMed: 19818451]
  - 8\*\*. Pai SY, et al. Transplantation outcomes for severe combined immunodeficiency, 2000-2009. *N Engl J Med*. Jul 31.2014 371:434. This is the most recent retrospective study of a multi-institutional PIDTC cohort of over 200 SCID patients. The impact of SCID type, infection status, age at transplant, donor type, and conditioning regimen on immune reconstitution, GVHD, and

survival are reported in detail. It showed that transplantation prior to infection is essential to maximize likelihood of survival. [PubMed: 25075835]

9. Kwan A, et al. Newborn screening for severe combined immunodeficiency and T-cell lymphopenia in California: results of the first 2 years. *The Journal of allergy and clinical immunology*. Jul.2013 132:140. [PubMed: 23810098]
10. Puck JM. Laboratory technology for population-based screening for severe combined immunodeficiency in neonates: the winner is T-cell receptor excision circles. *The Journal of allergy and clinical immunology*. Mar.2012 129:607. [PubMed: 22285280]
- 11\*. Dvorak CC, et al. The natural history of children with severe combined immunodeficiency: baseline features of the first fifty patients of the primary immune deficiency treatment consortium prospective study 6901. *Journal of clinical immunology*. Oct.2013 33:1156. This report of the first 50 patients enrolled on the prospective multi-institutional PIDTC SCID study details the characteristics of SCID patients in the era of routine newborn screening and how this affects has affected the timing of HSCT. [PubMed: 23818196]
- 12\*. Dvorak CC, et al. Comparison of outcomes of hematopoietic stem cell transplantation without chemotherapy conditioning by using matched sibling and unrelated donors for treatment of severe combined immunodeficiency. *The Journal of allergy and clinical immunology*. Aug 7.2014 This study of over 100 unconditioned SCID transplants reported the effects of SCID type, donor type, and serotherapy on outcomes including immune reconstitution, GVHD, and survival. It showed that unconditioned transplants for unrelated donors result in acceptable rates of GVHD and survival.
13. Dvorak CC, et al. Megadose CD34(+) cell grafts improve recovery of T cell engraftment but not B cell immunity in patients with severe combined immunodeficiency disease undergoing haplocompatible nonmyeloablative transplantation. *Biology of blood and marrow transplantation : journal of the American Society for Blood and Marrow Transplantation*. Oct.2008 14:1125.
- 14\*. Buckley RH, et al. Post-transplantation B cell function in different molecular types of SCID. *Journal of clinical immunology*. Jan.2013 33:96. This study provides a deeper analysis of how SCID type affects post-transplant immune reconstitution. [PubMed: 23001410]
15. Recher M, et al. IL-21 is the primary common gamma chain-binding cytokine required for human B-cell differentiation in vivo. *Blood*. Dec 22.2011 118:6824. [PubMed: 22039266]
16. Mazzolari E, et al. Long-term immune reconstitution and clinical outcome after stem cell transplantation for severe T-cell immunodeficiency. *The Journal of allergy and clinical immunology*. Oct.2007 120:892. [PubMed: 17825895]
17. Grunebaum E, et al. Bone marrow transplantation for severe combined immune deficiency. *JAMA : the journal of the American Medical Association*. Feb 1.2006 295:508.
- 18\*. Haddad E, Leroy S, Buckley RH. B-cell reconstitution for SCID: should a conditioning regimen be used in SCID treatment? *The Journal of allergy and clinical immunology*. Apr.2013 131:994. This review discusses the advantages and disadvantages of using conditioning in patients with SCID. [PubMed: 23465660]
19. Cavazzana-Calvo M, et al. Long-term T-cell reconstitution after hematopoietic stem-cell transplantation in primary T-cell-immunodeficient patients is associated with myeloid chimerism and possibly the primary disease phenotype. *Blood*. May 15.2007 109:4575. [PubMed: 17272510]
20. Sarzotti M, et al. T cell repertoire development in humans with SCID after nonablative allogeneic marrow transplantation. *Journal of immunology*. Mar 1.2003 170:2711.
21. Rao K, et al. Improved survival after unrelated donor bone marrow transplantation in children with primary immunodeficiency using a reduced-intensity conditioning regimen. *Blood*. Jan 15.2005 105:879. [PubMed: 15367433]
22. Slatter MA, et al. Treosulfan-based conditioning regimens for hematopoietic stem cell transplantation in children with primary immunodeficiency: United Kingdom experience. *Blood*. Apr 21.2011 117:4367. [PubMed: 21325599]
23. Robertson LE, et al. Natural killer cell activity in chronic lymphocytic leukemia patients treated with fludarabine. *Cancer Chemother Pharmacol*. 1996; 37:445. [PubMed: 8599867]

24. Dvorak CC, et al. A trial of alemtuzumab adjunctive therapy in allogeneic hematopoietic cell transplantation with minimal conditioning for severe combined immunodeficiency. *Pediatric Transplantation*. Sep.2014 18:609. [PubMed: 24977928]
25. Myers LA, Patel DD, Puck JM, Buckley RH. Hematopoietic stem cell transplantation for severe combined immunodeficiency in the neonatal period leads to superior thymic output and improved survival. *Blood*. Feb 1.2002 99:872. [PubMed: 11806989]
26. Gaspar HB, et al. How I treat ADA deficiency. *Blood*. Oct 22.2009 114:3524. [PubMed: 19638621]
27. Dvorak CC, Cowan MJ. Radiosensitive severe combined immunodeficiency disease. *Immunology and allergy clinics of North America*. Feb.2010 30:125. [PubMed: 20113890]
28. Fernandes JF, et al. Transplantation in patients with SCID: mismatched related stem cells or unrelated cord blood? *Blood*. Mar 22.2012 119:2949. [PubMed: 22308292]
29. Gluckman E, et al. Milestones in umbilical cord blood transplantation. *Br J Haematol*. Aug.2011 154:441. [PubMed: 21726206]
30. Gennery AR, et al. Transplantation of hematopoietic stem cells and long-term survival for primary immunodeficiencies in Europe: entering a new century, do we do better? *The Journal of allergy and clinical immunology*. Sep.2010 126:602. [PubMed: 20673987]
31. Muller SM, et al. Transplacentally acquired maternal T lymphocytes in severe combined immunodeficiency: a study of 121 patients. *Blood*. Sep 15.2001 98:1847. [PubMed: 11535520]
32. Palmer BE, Mack DG, Martin AK, Maier LA, Fontenot AP. CD57 expression correlates with alveolitis severity in subjects with beryllium-induced disease. *The Journal of allergy and clinical immunology*. Jul.2007 120:184. [PubMed: 17416406]
33. Scaradavou A, Carrier C, Mollen N, Stevens C, Rubinstein P. Detection of maternal DNA in placental/umbilical cord blood by locus-specific amplification of the noninherited maternal HLA gene. *Blood*. Aug 15.1996 88:1494. [PubMed: 8695871]
34. Knobloch C, Goldmann SF, Friedrich W. Limited T cell receptor diversity of transplacentally acquired maternal T cells in severe combined immunodeficiency. *Journal of immunology*. Jun 15.1991 146:4157.
35. Palmer K, et al. Unusual clinical and immunologic manifestations of transplacentally acquired maternal T cells in severe combined immunodeficiency. *The Journal of allergy and clinical immunology*. Aug.2007 120:423. [PubMed: 17481714]
36. Gaspar HB, et al. How I treat severe combined immunodeficiency. *Blood*. Nov 28.2013 122:3749. [PubMed: 24113871]
37. Dror Y, et al. Immune reconstitution in severe combined immunodeficiency disease after lectin-treated, T-cell-depleted haplocompatible bone marrow transplantation. *Blood*. Apr 15.1993 81:2021. [PubMed: 8471764]
38. Haddad E, et al. Long-term immune reconstitution and outcome after HLA-nonidentical T-cell-depleted bone marrow transplantation for severe combined immunodeficiency: a European retrospective study of 116 patients. *Blood*. May 15.1998 91:3646. [PubMed: 9573000]
39. Long-Boyle JR, et al. High fludarabine exposure and relationship with treatment-related mortality after nonmyeloablative hematopoietic cell transplantation. *Bone Marrow Transplantation*. Jan. 2011 46:20. [PubMed: 20383215]
40. Antoine C, et al. Long-term survival and transplantation of haemopoietic stem cells for immunodeficiencies: report of the European experience 1968-99. *Lancet*. Feb 15.2003 361:553. [PubMed: 12598139]
41. Neven B, et al. Long-term outcome after hematopoietic stem cell transplantation of a single-center cohort of 90 patients with severe combined immunodeficiency. *Blood*. Apr 23.2009 113:4114. [PubMed: 19168787]
42. Schuetz C, et al. SCID patients with ARTEMIS vs RAG deficiencies following HCT: increased risk of late toxicity in ARTEMIS-deficient SCID. *Blood*. Jan 9.2014 123:281. [PubMed: 24144642]
43. Kamani NR, et al. Malignancies after hematopoietic cell transplantation for primary immune deficiencies: a report from the Center for International Blood and Marrow Transplant Research.

- Biology of blood and marrow transplantation : journal of the American Society for Blood and Marrow Transplantation. Dec.2011 17:1783.
44. Eapen M, et al. Long-term survival and late deaths after hematopoietic cell transplantation for primary immunodeficiency diseases and inborn errors of metabolism. *Biology of blood and marrow transplantation : journal of the American Society for Blood and Marrow Transplantation*. Sep.2012 18:1438.
  45. Shah AJ, Kohn DB. Neurocognitive function of patients with severe combined immunodeficiency. *Immunology and allergy clinics of North America*. Feb.2010 30:143. [PubMed: 20113891]
  46. Titman P, et al. Cognitive and behavioral abnormalities in children after hematopoietic stem cell transplantation for severe congenital immunodeficiencies. *Blood*. Nov 1.2008 112:3907. [PubMed: 18645040]
  47. Walter AW, et al. Survival and neurodevelopmental outcome of young children with medulloblastoma at St Jude Children's Research Hospital. *Journal of Clinical Oncology*. Dec.1999 17:3720. [PubMed: 10577843]
  48. Mathisen SE, Glavin K, Lien L, Lagerlov P. Prevalence and risk factors for postpartum depressive symptoms in Argentina: a cross-sectional study. *International journal of women's health*. 2013; 5:787.
  49. Lin M, et al. Long-term neurocognitive function of pediatric patients with severe combined immune deficiency (SCID): pre- and post-hematopoietic stem cell transplant (HSCT). *Journal of clinical immunology*. Mar.2009 29:231. [PubMed: 18807155]
  50. Czechowicz A, Kraft D, Weissman IL, Bhattacharya D. Efficient transplantation via antibody-based clearance of hematopoietic stem cell niches. *Science*. Nov 23.2007 318:1296. [PubMed: 18033883]
  51. Straathof KC, et al. Haemopoietic stem-cell transplantation with antibody-based minimal-intensity conditioning: a phase 1/2 study. *Lancet*. Sep 12.2009 374:912. [PubMed: 19729196]
  52. Hassan A, et al. Outcome of hematopoietic stem cell transplantation for adenosine deaminase-deficient severe combined immunodeficiency. *Blood*. Oct 25.2012 120:3615. [PubMed: 22791287]
  53. Honig M, et al. Patients with adenosine deaminase deficiency surviving after hematopoietic stem cell transplantation are at high risk of CNS complications. *Blood* 109. Apr 15.2007 :3595.
  54. O'Marcaigh AS, et al. Bone marrow transplantation for T-B- severe combined immunodeficiency disease in Athabascan-speaking native Americans. *Bone Marrow Transplantation*. Apr.2001 27:703. [PubMed: 11360109]
  55. Bertrand Y, et al. Influence of severe combined immunodeficiency phenotype on the outcome of HLA non-identical, T-cell-depleted bone marrow transplantation: a retrospective European survey from the European group for bone marrow transplantation and the european society for immunodeficiency. *The Journal of pediatrics*. Jun.1999 134:740. [PubMed: 10356144]
  56. Dalal I, et al. Matched unrelated bone marrow transplantation for combined immunodeficiency. *Bone Marrow Transplantation*. Mar.2000 25:613. [PubMed: 10734295]
  57. Knutsen AP, Wall DA. Umbilical cord blood transplantation in severe T-cell immunodeficiency disorders: two-year experience. *Journal of clinical immunology*. Nov.2000 20:466. [PubMed: 11202237]
  58. Bhattacharya A, et al. Single centre experience of umbilical cord stem cell transplantation for primary immunodeficiency. *Bone Marrow Transplantation*. Aug.2005 36:295. [PubMed: 15968287]



**Figure 1.**

Donor selection and conditioning regimen in patients with typical SCID (UCSF approach).

<sup>1</sup>Algorithm excludes Omenn's syndrome and leaky SCID that would be classified as atypical (2). Also excludes patients with DNA sensitivity, as donor selection and conditioning for these patients are variable due to the high risk of rejection and potential for chemotherapy toxicity

<sup>2</sup>Based on availability, CMV status, donor age, or other variables. Patients with RAG SCID receiving a haploidentical transplant will generally require non-myeloablative chemotherapy and serotherapy.

<sup>3</sup>May consider chemotherapy-based conditioning for enhanced B cell and/or T cell reconstitution

<sup>4</sup>Patients with T-B-NK+ SCID receiving an URD transplant generally require a conditioning regimen with serotherapy

<sup>5</sup>Experimental gene therapy protocols are becoming more widely available and should be considered in cases where no appropriate donor can be identified

**Table 1**

SCID subtypes and relevant considerations for HSCT.

T-B+NK- SCID-XL (IL2RG) JAK3	<ul style="list-style-type: none"> <li>• Generally reconstitute T cell immunity from any type of donor without any conditioning required.</li> <li>• When using donors other than matched siblings, usually fail to regain B cell function, unless conditioning is provided.</li> <li>• Increased risk of late HPV infection, which is likely related to abnormal <math>\gamma c</math> / JAK3-dependent signaling in keratinocytes.(41).</li> <li>• Gene therapy (GT) is being evaluated in SCID-XL patients.</li> </ul>
T-B-NK- ADA	<ul style="list-style-type: none"> <li>• When a MSD is not available, enzyme replacement therapy (ERT) is a potential option and should be pursued over a mismatched unrelated HSCT.(26)</li> <li>• The underlying defect in purine metabolism may lead to increased susceptibility to certain chemotherapeutic agents, which may explain why survival is lower in those patients getting conditioning.(26)</li> <li>• Matched related or unrelated donors may not require a conditioning regimen (12). For mismatched related or unrelated donors, most centers have used a conditioning regimen, as rejection is seen more commonly without conditioning in these transplants compared to IL2RG-deficiency patients, despite absence of NK cells.(52)</li> <li>• B cell function is often recovered after matched sibling donor transplant, even without conditioning, but is more variable in unconditioned unrelated donors or mismatched related donors.(12)</li> <li>• 50% of patients will have cognitive abnormalities following either transplantation or ERT (46, 53). This has not been linked to either the use of conditioning or the degree of myeloid engraftment.</li> <li>• For patients without a MSD, GT has a lower risk of immediate complications, but it is not widely available</li> </ul>
T-B+NK+ IL7R $\alpha$ CD45 CD3 subunits	<ul style="list-style-type: none"> <li>• Conditioning is not required if a MSD donor is available.</li> <li>• Limited data exist on the role of conditioning for HLA-matched unrelated donors.</li> <li>• For haploidentical transplants, conditioning is not usually necessary, especially if using a maternal graft in the setting of transplacental maternal chimerism.</li> <li>• As there is no intrinsic B cell defect, B cell recovery in these patients is not dependent on conditioning and is expected in most cases providing T cell reconstitution is achieved.(14)</li> </ul>
T-B-NK+ RAG1/2 Artemis DNA ligase IV Cernunnos-XLF DNA-PKcs	<ul style="list-style-type: none"> <li>• RAG1/2 defects can present with a clinical spectrum dependent on the functionality of the mutations. Complete loss of function results in typical SCID, while hypomorphic mutations can produce a leaky SCID phenotype, with detectable non-maternal circulating T cells that, if autoreactive, result in Omenn Syndrome.</li> <li>• In patients with RAG1 or RAG2-SCID, there is no increased sensitivity to alkylating chemotherapy as seen in defects of DNA repair.(42)</li> <li>• For donors other than MSDs, there is an increased risk of rejection if no conditioning is used.(7, 13)</li> <li>• For donors other than MSD's, there is very poor B cell recovery if conditioning is not used.</li> <li>• Hypomorphic mutations typically require conditioning, regardless of donor type.</li> </ul> <hr/> <ul style="list-style-type: none"> <li>• DNA repair defects are reviewed elsewhere.(27)</li> <li>• If maternal engraftment is present, a maternal donor is strongly preferred, since T-cell engraftment will generally occur without the need for alkylating chemotherapy.</li> <li>• High rate of regimen-related toxicity and late effects when DNA-damaging agents are used.(42, 54)</li> <li>• In order to avoid alkylating therapy, it is advisable to proceed with an unconditioned transplant, even with haploidentical or matched unrelated donors. If conditioning is used, attempts to minimize the number of alkylators and the dosing are recommended.</li> <li>• B cell recovery is rare if conditioning is not used.</li> <li>• Reduced intensity conditioning that includes fludarabine may be used if rejection occurs following unconditioned HSCT.(13)</li> </ul> <hr/> <ul style="list-style-type: none"> <li>• The number of patients with other radiation sensitive defects is too small for survival analysis.</li> <li>• Use of MAC is a risk factor for inferior survival (A Gennery, personal communication).</li> </ul>

- For DNA Ligase IV and Cernunnos-XLF deficient patients, very low dose cyclophosphamide and fludarabine are recommended (<http://esid.org/layout/set/print/Working-Parties/Inborn-Errors-Working-Party-IEWP/Resources/>).

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript



Table 2

Overall survival in studies of SCID patients by donor and conditioning regimen.

First Author	Year	Ref	MRD (N)	Haplo (N)		URD (N)			UCB (N)	
			None	None	RIC/MAC	None	MAC	RIC	MAC	
Conditioning										
Bertrand	1999	(55)		46% (50)	54% (129)					
Dalal	2000	(56)					67% (9)			
Knutsen	2000	(57)								88% (8)
Antoine	2003	(40)	81% (104)				63% (28)			
Rao	2005	(21)					71% (7)	83% (6)		
Bhattacharya	2005	(58)								80% (10)
Grunebaum	2006	(17)	92% (13)		53% (40)		81% (41)			
Dvorak	2008	(13)		87% (15)						
Gennery	2010	(30)	84% (135)	54% (415)			66% (81)			
Buckley	2011	(5)	100% (17)	73% (149)						
Femandes	2012	(28)		62% (175)						57% (74)
Dvorak	2014	(12)	92% (66)				73% (37)			
Pai	2014	(8)	97% (32)	79% (87+16 IS only)	66% (35)		74% (19)			58% (43)