Title: Plagiarism or not? Investigation of Turnitin®-detected similarity hits in biology lab reports

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ABSTRACT

In undergraduate biology laboratory courses, lab reports can be a useful tool for teaching scientific writing, integration of source material, and information literacy, however these teaching objectives are at times undermined by students’ plagiarism. Lab instructors often use similarity-matching software to detect plagiarism in lab reports, yet similarity hits detected with such software remain poorly characterized. In the upper-division molecular biology lab course described here, Turnitin® routinely detected dozens of similarity hits in lab reports. To determine whether this abundance of similarity hits was indicative of widespread plagiarism, we analyzed similarity hits detected in 255 lab reports written by 135 students. Only a small minority of Turnitin® similarity matches were problematic, but over half of the lab reports contained at least one problem with incorporation of scientific sources (e.g., lab manual, scientific articles). We identified four common types of such writing problems: patchwriting, technical parroting, copying, and falsification of sources. In 18% of the lab reports, we detected an alarmingly superficial use of primary literature. Most of the source incorporation problems did not rise to the level of plagiarism. As a result of this study, we recommend changes in scientific writing instruction and a transition to labs providing more authentic research experiences.
INTRODUCTION

In undergraduate biology classes, laboratory reports whose structure parallels that of scientific articles have been seen as a standard tool for acquiring writing competency, as they aim to guide students towards critical and evaluative thinking, information literacy, clearly-written scientific communication, and appropriate integration of source material. It has been suggested that the process of writing a lab report enables students to make sense of their lab experience within the context of scientific inquiry [1].

A traditional undergraduate biology lab report includes several standard sections that mirror the structure of a scientific paper: a brief Abstract summarizes the most important findings, an Introduction explains the background of the experiment, a Materials and Methods section outlines and details experimental procedures, a Results section presents data, and a Discussion section elaborates on data analysis. This traditional format was implemented in the upper-division molecular biology lab classes that were the focus of this study. For the past seven years of teaching these labs, similarity-matching software, Turnitin®, has been used to detect potential instances of plagiarism. Over the years, instructors have been aware of the large number of similarities detected by Turnitin® in many of the lab reports. The majority of these matches linked to lab reports by other students, which raised the possibility of our students having access to, and plagiarizing from, these lab reports.

A growing incidence of plagiarism among science students plagiarism, a deliberate use of someone else’s ideas and language without acknowledgment of their source [2], has been a source of great concern in academia [3–7].

Since the literature on plagiarism in lab reports has been limited to surveying students’ self-reported behaviors such as copying from other students or falsifying data [8], we decided to conduct an examination of a large number of lab reports in order to understand the nature of the matches detected by Turnitin®. Specifically, we asked: do the abundant similarity matches identified by Turnitin® in student lab reports indicate plagiarism or
misuse of sources? Another possibility is that these matches result from unavoidable repetition of technical terms and protocol details, resulting when hundreds of students write reports on the same experimental procedure. To answer this question, we conducted a detailed analysis of Turnitin®-detected similarity matches in 255 lab reports written by 135 students in this course.

METHODS

*Context of the study*

This study was conducted at a large, research-intensive, public university where about 800 students take the upper-division molecular biology lab course annually. The participants of this study were 135 students, mostly seniors majoring in Biological Sciences (92%), enrolled in three of the lab sections during the 2012-2013 academic year. One instructor (G.B.) taught two of the sections (35 and 49 students, respectively), and another instructor (E.T.) taught the third section (51 students).

Students typically wrote three to four lab reports, each lab report addressing a project that spanned three or more labs. Students submitted the lab reports were submitted to Turnitin®. The course lab manual provided students with the background to the experiments and with detailed, step-by-step protocol instructions. Two lab reports from each student were examined in this study: Lab Report 1, based on the cloning of the *lux* operon from the bioluminescent bacterium *Vibrio fischeri*, and Lab Report 3, based on phenotypic observation and qRT-PCR analysis of an RNAi-mediated knockdown of *unc-22* gene expression in *C. elegans* The two instructors used the same guidelines for the lab reports. The same experiments had been conducted for at least five (Lab Report 3) to eight (Lab Report 1) years, so the pool of existing lab reports written by former students and submitted to Turnitin® was extensive. Information about academic integrity was provided to students in the syllabus and was posted on the course website (Supplemental Materials, Appendix A).
Detailed description of the analysis of Turnitin®-generated reports is provided in the Supplemental Materials, Appendix B. Briefly, we analyzed 255 Turnitin® originality reports (Lab Report 1: 130 reports, Lab Report 3: 125 reports). All matches were examined by a team of two biology instructors (E.T. and G. B.) with the goal to determine if a match was significant or if the similarity could be attributed to chance alone. Others have deemed the appropriation of five or more consecutive words from a source as constituting an incident of plagiarism [9,10]. However, in this study, the lab reports contained a substantial amount of technical terminology that could not have been expressed differently. Therefore, in considering if a match was significant, we asked, “In how many other ways could the same information be communicated? When the length of the match was relatively short and the information very technical or included commonly used terminology, we considered the match insignificant (Table 1). After several joint norming sessions, the two biology instructors made the determination of significance independently and then met and discussed any disparities, until consensus was reached.

The next task was to determine the source text of each match. Turnitin® very reliably identified matches to journal articles and websites, however, the majority of the similarity matches were attributed by Turnitin® to other students’ papers and for these matches, the source text was not readily available. Furthermore, we determined that most of such matches were actually matches to the lab manual. Table 2 shows an example from one student’s paper, where Turnitin® found matches to 47 different students’ lab reports, all from our institution. All 47 of these matches had similarities to the lab manual text as well, making it by far the most likely source. When this type of source misattribution was encountered, we treated all of such matches as one match, to the lab manual. In the majority of the cases, a consensus was reached on significant matches and their likely sources. Cases where no consensus was reached were excluded from the analysis.
After significant matches were determined, the second coding team (M.P., a writing expert and T.B.G., a plagiarism expert) categorized these matches into types of source material incorporation problems (Table 3). A source incorporation problem was described as “extended” if it incorporated two or more sentences from the original source. The second coding team first determined the type of source incorporation problem independently, then compared the codes and discussed and resolved any disagreements.

This study was determined to be exempt from IRB approval by the UC San Diego Human Research Protections Program, project #120621XX.

RESULTS

In the 255 lab reports included in this study, Turnitin® identified, on average, 29 similarity hits to other texts in Lab report 1 and 28 matches in Lab Report 3. However, after discarding the matches we deemed insignificant and accounting for Turnitin® misattribution problem (matches to the lab manual were misattributed to lab reports by other students, see Methods and Table 2), we detected an average of two significant source incorporation problems per lab report (1.6 in Lab report 1 and 2.2 in Lab report 3).

Over half of the lab reports (53%) exhibited at least one source incorporation problem and 8% of the lab reports contained five or more source incorporation problems (Table 4). Among the 341 source incorporation problems we detected, 41.5% extended beyond a single sentence. The source incorporation problems occurred most often within sections rich in technical details: Materials and Methods and the parts of Results where students wrote about procedures (50.5% of source incorporation problems), or in the Introduction section, where students wrote about the background to the experiments (47% source incorporation problems).

We categorized source incorporation problems into four types (see Table 3 for examples):
1. Patchwriting: a match to the source that reproduces the original language, but includes synonym substitutions, word or phrase omissions, and sentence restructuring [11]

2. Technical parroting [12]: a match rich in technical details (i.e., volumes, concentrations), reproduced from the lab manual or lecture slides with little or no change. Such matches typically occurred in the Materials and Methods and the Results sections of lab reports.

3. Copying: a verbatim match to an entire sentence in a source (may include minor word substitutions or omissions)

4. Falsification of a source: a copied or patchwritten text contains a citation that attributes the information to one scientific source (typically, a journal article), while the source of the information is different (typically, the lab manual).

**Patchwriting and technical parroting.** The predominant source incorporation problems were patchwriting and technical parroting (Table 3), accounting for 87% of such problems (Table 4). Patchwriting, an excessive use of a source’s text with the source text modified slightly through word substitutions or deletions [13] contributed 59% of the source incorporation problems (Table 4). Among the instances of patchwriting, some showed a clear (but unsuccessful) attempt to restructure the original sentence and “make it their own,” whereas in other instances, this attempt was so minimal, and the use of the source text so extensive, that it bordered on copying (Supplemental Table S2). One-third of the patchwriting problems extended to more than one sentence (Table 4).

A related problem was technical parroting: the repetition of methods, processes, or procedures from the lab manual, with little or no change from the original (Table 3). The term “technical parroting” is drawn from Moskovitz and Kellogg, who have suggested that when a procedure is already outlined in detail in the lab manual, there is “little for [the students] to do but parrot back selected details from the [lab] manual”[12]. Technical
details, such as volumes of reagents, temperatures, and times of incubation, were abundant in these matches, and the source was almost invariably the lab manual (Table 3). Despite the multitude of technical terms, many of which could not be expressed differently, we still considered technical parroting a source incorporation problem because in these instances, instead of distilling the most pertinent information from the protocol description, students were simply repeating the general instructions of the lab manual. Most (78%) of the technical parroting extended to more than one sentence (Table 4).

Since lab reports are typically rich in specific terminology and technical details, is it perhaps impossible to avoid substantial similarity to the lab manual or to what has been written before by other students? This does not seem to be the case: we found that many students (29%) managed to successfully present the experimental purpose, definitions, procedures, results, and discussion without yielding any significant Turnitin® similarity matches (Table 5).

**Copying.** Direct copying – exact word-for-word replication of the source material – was found in only 9% of the total instances of source incorporation problems; we found 30 instances of copying in 23 papers written by 20 different students. The sources of copied material included the lab manual (33%), another student’s lab report (30%), journal articles (23%), websites (10%), and a book (3%). The majority of copied text was contained in isolated single sentences (81% of instances of copying) used to supplement background information (21 instances) or methods (9 instances). Examples of copying are shown in Table 3 and Supplemental table S3.

**Falsification.** Falsification of sources – a citation of a paper that did not contain the information the student attributed to it – occurred in 5% of the instances of source incorporation problems (Table 4) and was found in 13 papers. In all these cases, the student falsely attributed material that came from a website or the lab manual to a journal article (see examples in Table 3 and Supplemental table S4). The majority of instances of
falsification of sources (13 out of 16) occurred in conjunction with copying or patchwriting. For example, a text might have been patchwritten from the lab manual, but attributed to a scientific article which in fact did not contain such information (Table 3). In the instructions for writing these lab reports, students were asked to incorporate information from journal articles. It is likely that falsification of sources was an attempt to make it appear as if this requirement was satisfied.

**Superficial use of primary literature.** 18% of the lab reports contained instances of patchwriting or copying that occurred when students attempted to use scientific literature as source material. Frequently, the superficial changes students introduced into the patchwritten text resulted in a loss of the biological meaning of the original text (Table 6). The sentences selected by students for such superficial use were predominantly from a paper’s Abstract or the beginning of its Introduction section.

**DISCUSSION**

**Plagiarism or misuse of sources?**

This study was motivated by the authors’ concern about the large numbers of similarity matches detected by Turnitin® in the lab reports written by our students. We sought to determine whether these matches were a result of chance alone (that is, numerous students writing on the same topic and using the same terminology) or whether these matches indicated genuine source incorporation problems: instances of significant similarity between student’s lab report and another text, such as lab manual, website, journal article. If these matches were indeed source incorporation problems, did they rise to the definition of plagiarism?

Source incorporation problems were present in more than half of the lab reports we examined. However, we consider only a small minority of those as plagiarism: copying (14%) or falsification of sources (5%). Most of source incorporation problems we encountered involved patchwriting and “parroting” protocol details from the lab manual.
While patchwriting and “parroting” can be considered plagiarism [14,15], we reasoned that students were not fraudulently trying to present the ideas from the lab manual as their own, since the lab manual’s text was available and well known to the instructor. Therefore, we consider patchwriting and technical parroting a misuse of sources, rather than plagiarism.

Our findings about the prevalence of source incorporation problems agree with a multi-institutional study conducted by Jamieson and Howard (2013) that examined the use of source material in 174 research papers written by freshmen enrolled in writing courses. At least one instance of patchwriting from sources was found in 52% of the papers [16]. Similarly, Flaspohler and colleagues reported that 50% of biology students enrolled in an upper-division elective course committed what the authors called “knowing or unknowing plagiarism” when writing evaluative annotations of research articles [17]. The incidence of direct copying we detected was higher than has been previously reported in freshman writing courses: 9% in our study versus 4.3% previously reported [16] perhaps reflecting differences between the contexts of the writing assignments: biology labs in this study versus writing courses and Jamieson and Howard’s study.

**Superficial use of primary literature**

In 18% of the lab reports, we detected patchwriting from the primary literature, where students cited the source paper, but patchwrote sentences from the scientific articles in ways that very often distorted the meaning of the original sentence. In most instances, only sentences from the Abstract or the beginning of the Introduction sections of the source articles were used, suggesting that students did not read the source articles beyond these sections. These findings are in agreement with a study by Jamieson and Howard (2013) that found that the majority (69%) of citations in 174 papers written in first-year writing courses from a variety of colleges and universities came from the first two pages of the cited sources. Science students also frequently rely on abstracts alone while writing about scientific articles [17].
Howard and colleagues described this phenomenon as “writing from sentences”: an attempt to paraphrase or summarize based on just a few sentences from a source [11]. Without the context and deeper understanding of the subject of the article, this task is very challenging for students, leading to a heavy reliance on the original wording of the text and to misrepresentation of the original meaning.

**Limitations and future directions**

The strength of this study is that it examines actual student writing rather than relying on the student self-reports that dominate plagiarism studies (noteworthy exception is Rebecca Moore Howard’s Citation Project, found at http://site.citationproject.net/). The ability to generalize our results is limited, since this study was conducted at one institution, in one discipline (biology), and in one type of lab report (the expository, traditional lab report). Further research in other lab science courses is needed to gain a comprehensive view of how undergraduates develop their source incorporation skills.

Our research also has a limitation in common with the research conducted within the Citation Project [16]: while we can identify problems with use of sources, we can only hypothesize as to the causes of such problems. The Council of Writing Program Administrators suggests a number of possible causes of plagiarism and misuse of sources: students may lack the training of appropriate integration of sources, mistakes in integrating sources are expected part of the learning process, college instructors may underestimate the difficulty of such integration [2]. We agree that patchwriting and technical parroting can result from lack of knowledge of writing conventions in biology. It is also possible that students did not choose to spend enough time to carefully read and paraphrase the sources. Interviews with the students whose lab reports exhibit source incorporation problems would be very informative in elucidating the underlying reasons of these problems.
RECOMMENDATIONS

Reconsideration of traditional lab reports

Alaimo and colleagues (2009) and Moskovitz and Kellog (2011) argue that the traditional lab report is an artificial genre and is not an optimal medium for developing competency in scientific writing [12,18]. Our study offers evidence to substantiate these claims. In the Introduction section of the lab report, we asked students to find and write about relevant background literature. This is an important skill since many of our students hope to become physicians or researchers and the ability to read and analyze primary literature has been identified as one of the entry-level competencies for aspiring physicians [19]. However, our findings show that many students instead patchwrote or copied information provided in the lab manual. We suggest providing students with less background information in the lab manual, so that they explore the relevant background literature.

Technical parroting, another frequent problem, was mostly characterized by a mere repetition of the lab manual’s information, a “text dump” with limited attempt to understand relevance (no prioritizing, sequencing, or other evidence of deeper understanding). We concur with the argument that lab reports can become a more useful and meaningful exercise in the authentic research context: labs where students develop protocols and communicate them in a formal written format, so that others can replicate the process [12,18]. A call for discovery-based research in undergraduate laboratory courses in STEM disciplines was issued by the President’s Council of Advisors on Science and Technology [20]. Course-based undergraduate research experiences, or CURE’s, offer a promising model for allowing students to engage in authentic research that is also of interest to the scientific community [21,22]. Indeed, the curriculum of the lab course described in our study now includes an authentic research project that has replaced one of the two large projects that engendered the lab reports examined here.
This recently implemented course module involves students in hypothesis building and in experimental design. It will be interesting to examine whether such labs will reduce the incidence of mindless patchwriting and parroting from the lab manual observed in this study.

In addition, to reduce technical parroting, one of the instructors (E.T.) replaced large-scale, high-stakes lab reports with smaller and more frequent assignments in which students practice summarizing procedures, as well as presenting and discussing their results. In such assignments, students are provided with a rubric that includes explicit expectations for summarizing the procedures, as well as with information about grade deduction for parroting the lab manual (an example rubric is provided in the Supplemental Materials, Appendix C). These “mini-lab reports” are graded by graduate instructional assistants, who also provide students with feedback on their writing. Our preliminary observations indicate that technical parroting was reduced in students’ writing, following this curriculum change (E.T., unpublished observations). However, we doubt that authentic research or more frequent, smaller-scale writing assignments will solve the problems we see when students attempt to write relying on primary literature. Below, we offer recommendations for dealing with this problem.

**Training for students**

Multiple studies have suggested that undergraduates often lack not only the skills of paraphrasing and summarizing, but also of conducting an informed literature search and engaging with the ideas in their sources [10,11,13,16,17,23–27]. Struggles with source-based scientific writing are to be expected as novices master disciplinary conventions [28], as students grapple with new vocabulary, concepts, scientific writing conventions, and – when writing from research articles – with complex scientific content [16]. Novices often start with patchwriting, advancing toward paraphrasing and then summarizing as they become more knowledgeable and confident in the discipline [11,29]. When asked to paraphrase or summarize content from an unfamiliar field, not
only students, but also faculty often patchwrite [10,30]. Therefore, rather than viewing patchwriting as an ethical failure, we suggest it is a problem that needs to be addressed with better-scaffolded instruction in discipline-specific reading and writing.

Such instruction should include training in conducting a literature search, critical reading of scientific articles, and appropriately paraphrasing and summarizing scientific ideas from sources. Training in scientific writing should also include explicit instruction in identifying and avoiding plagiarism [2,9,31,32]. The accompanying manuscript by Yang and colleagues offers a successful approach to addressing these issues in the context of our labs. The process of writing about scientific background should include multiple drafts, reviewed by the instructor or appropriately trained teaching assistants and examined via similarity-detection software [2]. Students who struggle with comprehension, paraphrasing, or summarizing should be provided with additional instruction (on reading scientific articles, disciplinary writing, or the ethics of scientific writing) and opportunities to revise their writing. Conversations between instructors and these students could inform a decision regarding which interventions would be most appropriate. Because of its scope, such training should begin early in the undergraduate program.

*Changes in Turnitin®*

Turnitin® similarity reports were helpful in the initial identification of possible source incorporation problems, although some current features of Turnitin® made the determination of the extent of plagiarism and the source of patchwriting difficult in our context. For example, we found many cases in which similarities that were clearly derived from one text available to all students (the lab manual) were identified by Turnitin® as small individual matches to a large number (sometimes dozens) of student papers. This presents a substantial difficulty for instructors in interpreting Turnitin® results: they are left to puzzle if such matches are evidence that the student is plagiarizing or are mere coincidences.
Interpretation of Turnitin® results would be simplified if instructors could indicate which texts (e.g., lab manuals, scientific papers) Turnitin® should consider the “preferred,” most likely source material. Alternatively, the Turnitin® algorithm should be changed so that it can seek the most likely source of plagiarism. It would also be very useful for instructors to have a means for indicating phrases that Turnitin® should ignore (e.g., terminology, technical definitions) because there are no other ways to express the information. Such options are currently unavailable on Turnitin® (to our knowledge).

**Faculty training in using Turnitin®**

Based on our findings, we recommend that lab instructors should be trained to use the existing version of Turnitin® in the following ways:

1. Before students submit their lab reports to Turnitin®, the instructor should upload the lab manual or instructions provided to students so that it will become “the oldest” and thus the “original” source for the Turnitin® algorithm.

2. After the student reports are submitted, the instructor should review each Turnitin® originality report to assess which identified matches are of concern (e.g., a match to a journal article or extensive matches to another student’s paper). We recommend using the “text-only” version of the originality report as it breaks down the suspected sources rather than showing all “student papers” together as one source.

3. If patchwriting or technical parroting problems are detected, we recommend addressing them first as writing problems rather than as plagiarism.

4. Copying and/or falsification of sources should be considered plagiarism and dealt with according to the institution’s procedures for reporting academic integrity violations.

We hope that our recommendations will help instructors who use Turnitin® to recognize and differentiate between instances of plagiarism and misuse of sources. Such differentiation will help them in a difficult task of meeting the needs of their science
program to conduct honest assessments and the needs of students who at times are facing expected challenges in learning scientific writing.

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The authors declare no conflict of interest.

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TABLES LEGENDS

18
Table 1. Examples of Turnitin® matches that were determined to be insignificant. Identical text is indicated in bold. Even though both examples contain a sequence of words that is identical to the matched source (12 identical words in a row in the first example, 9 in the second), we determined that it would be very difficult to deliver the same information in other ways and therefore designated these and similar matches as insignificant.

Table 2. An example of Turnitin® mistakenly attributing the source text to other students’ papers, instead of the lab manual. Matching text is indicated in bold. Only the relevant parts of the lab manual (Butler and Noree, 2012) are shown. The numbers in parentheses in the left column (1, 9, 7, and 64) that precede the matched text indicate the different sources to which Turnitin® attributed these matches. Match 1 was attributed by Turnitin® to the lab manual, while matches 9, 7, and 64 were attributed by Turnitin® to lab reports by other students. However, all of the matched text is also present in the lab manual (right column, bold), making patchwriting from the lab manual a more parsimonious source of the matches. Here and in other tables, we present students’ texts without correcting grammatical or spelling mistakes; only the scientific names of organisms (e.g., *C. elegans*) and of genes were italicized by us.

Table 3. Definitions and illustrations of the four different types of source incorporation problems found in students’ lab reports. Matching text is indicated in bold.

Table 4. Frequency of the types of source incorporation problems. N = 255 lab reports. a Total number of source incorporation problems in all lab reports. b Proportion of each type of source incorporation problems, out of the total instances of source incorporation problems (N= 341). * Note that, because of the rounding, percentages in rows 2-5 add to more than 100%. c Proportion of extended source incorporation problems in each category. # Extended problem refers to a source incorporation problem spanning more than one sentence. d Proportion of extended problems in the total number of source
incorporation problems (N= 341). Note that, because one lab report could have more than one type of source incorporation problem, the percentage numbers in rows 2-5 do not add to 53%.

**Table 5:** Difference between parroting or patchwriting and writing in one’s own words. Left column: source text from which the problematic writing (center column) was derived. Right column: a corresponding part from a lab report written by a different student, demonstrating writing in their own words.

**Table 6.** Examples of patchwriting from the primary literature that distorted the meaning of the original text.
**TABLES**

Table 1. Examples of Turnitin® matches that were determined to be insignificant

<table>
<thead>
<tr>
<th>Student’s text</th>
<th>Matched source text</th>
</tr>
</thead>
<tbody>
<tr>
<td>As it is found predominantly in symbiotic relationships with marine organisms, the <strong>symbiosis between V. fischeri and the Hawaiian bobtail squid Euprymna scolopes has</strong> been carefully studied by scientists for many years.</td>
<td>Interest in the light-organ <strong>symbiosis between V. fischeri and the Hawaiian bobtail squid Euprymna scolopes</strong> has led several researchers to adopt strain ES114 (4)... Source: journal article, Lyell et al., 2008</td>
</tr>
<tr>
<td>Inside the cells, the dsRNA is chopped up with the <strong>enzyme Dicer into double stranded small interfering RNAs (siRNA)</strong> that group together into what is called a RNA-induced silencing (RISC) complex, held together by argonaute proteins.</td>
<td>Once dsRNA enters the cell, it is cleaved by an RNase III –like <strong>enzyme, Dicer, into double stranded small interfering RNAs (siRNA)</strong> 21-23 nucleotides in length that contain 2 nucleotide overhangs on the 3’ ends (9-11). Source: website, <a href="http://www.genelink.com/sirna/RNAicustom.asp">http://www.genelink.com/sirna/RNAicustom.asp</a></td>
</tr>
</tbody>
</table>
Table 2. An example of Turnitin® mistakenly attributing the source text to other students’ papers, instead of the lab manual

<table>
<thead>
<tr>
<th>Student’s text</th>
<th>Lab manual</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Excerpt from the Introduction:</strong></td>
<td><strong>Basically, they found that the introduction of double-stranded RNA (dsRNA) that includes coding sequences of a specific gene can specifically disrupt the function of that gene by inducing the destruction of the mRNA.</strong></td>
</tr>
<tr>
<td>Fire and Mello discovered (1) that the introduction of double-stranded RNA (dsRNA) including coding sequences of a specific gene can disrupt the function of that gene by inducing the degradation of a target mRNA.</td>
<td></td>
</tr>
<tr>
<td><strong>Excerpt from the Materials and Methods:</strong></td>
<td></td>
</tr>
<tr>
<td>The <em>C. elegans</em> strain used, (1) strain NL2099-rrf-3, is particularly sensitive to RNAi. Each plate contained three to four large worms on them. The (9) worms were transferred to plates coated with a specific strain of bacteria HT115 (DE3) that has the gene for T7 polymerase. T7 polymerase expression is under the control of a lac promoter and operator. The HT115 (DE3) bacteria also contain a plasmid (L440 double T7 vector) (7) that has an amp resistance gene, and will either contain no RNAi insert (control), or a 800 bp sequence for the unc-22 gene in the polylinker region. Two T7 promoters flank the (64) unc-22 gene sequence and control the transcription of the insert and the product is a dsRNA.</td>
<td>You will be given untreated plates of gravid <em>C. elegans</em> (strain NL2099-rrf-3—this strain is particularly sensitive to RNAi). Three to four worms will be transferred to plates coated with a specific strain of bacteria HT115(DE3) which contains the gene for T7 polymerase. The expression of the T7 polymerase is under the control of a lac promoter and operator (so just like the pET vector system). The HT115(DE3) bacteria also contain a plasmid that has an amp resistance gene, and will either contain no RNAi insert (control), or a 800 bp sequence for the unc-22 gene in the polylinker region. Transcription of the unc-22 sequence is controlled by two T7 promoters that flank the insert on both ends, so the product is a dsRNA.</td>
</tr>
</tbody>
</table>
Table 3. Definitions and illustrations of the four different types of source incorporation problems found in students’ lab reports

<table>
<thead>
<tr>
<th>Code</th>
<th>Definition</th>
<th>Student Text</th>
<th>Source Text</th>
</tr>
</thead>
<tbody>
<tr>
<td>Copying</td>
<td>Verbatim match to an entire sentence (words and structure) in a source (may include minor word substitutions or omissions, such as replacing “will” for “would” or omitting articles or conjunctions, such as “and, but, when, or”)</td>
<td>These digested pieces of DNA were then ligated together using T4 ligase to create recombinant vectors containing pieces of <em>Vibrio</em> DNA with Sal I “sticky” ends ligated into cut pGEM with Sal I “sticky” ends.</td>
<td>To create recombinant vectors containing pieces of <em>Vibrio</em> DNA with Sal I “sticky” ends ligated into cut pGEM with Sal I “sticky” ends. Source: Lab manual (Butler and Noree, 2012)</td>
</tr>
<tr>
<td>Patchwriting</td>
<td>A match to the source that reproduces the original language, but includes synonym substitutions, word/phrase omissions, and sentence restructuring (Howard, et al. 2010)</td>
<td>The genes coding for a and b subunits of luciferase are <em>luxA</em> and <em>luxB</em>, while <em>luxC</em>, <em>D</em>, and <em>E</em> genes code for polypeptides that are required for the conversion of fatty acids into the long-chain aldehyde required for the luminescent reaction...</td>
<td>The genes coding for the bacterial luciferase enzyme subunits which catalyze the bioluminescence reaction are <em>luxA</em> and <em>luxB</em>. The <em>luxC</em>, <em>D</em>, and <em>E</em> genes code for polypeptides (transferase, esterase, and reductase) that are required for the conversion of fatty acids into the long-chain aldehyde required for the luminescent reaction. Source: Lab manual (Butler and Noree, 2012)</td>
</tr>
<tr>
<td>Technical Parroting</td>
<td>A match was coded as technical parroting if it met three criteria: 1) It contained repeated material from the lab manual or lecture slides, with essentially little or no change from the original 2) It was found in the Materials and Methods or the Results section 3) It was rich in technical details such as temperatures, names of reagents, concentrations, or volumes</td>
<td>RNA samples were then diluted to a final concentration of 20 ng/ul using RNase free water, to which a mixture of master mix from Biorad containing SYBR Green Dye, AmpliTaq Gold DNA Polymerase, dNTPS, reverse transcriptase enzyme, and buffer components were added.</td>
<td>Master Mix from Biorad containing SYBR Green Dye, AmpliTaq Gold DNA Polymerase, dNTPs, reverse transcriptase enzyme and buffer components were added. Source: Lab manual (Butler and Noree, 2012)</td>
</tr>
<tr>
<td>Falsification</td>
<td>The student falsifies a citation, suggesting the text is from one source when</td>
<td>dsRNA can move freely from cell to cell in <em>C. elegans</em> through a pore</td>
<td>Amazingly, dsRNA moves freely from cell to cell in <em>C. elegans</em> through a pore formed by a</td>
</tr>
</tbody>
</table>
really it is from another. In the example on the right, the student cited the material as having come from Winston et al., (2002), but it actually came from the lab manual. Note that this falsification is combined with very close patchwriting (almost copying) from the lab manual.

**Table 4. Frequency of the types of source incorporation problems**

<table>
<thead>
<tr>
<th>Types of source incorporation problems</th>
<th>Number of source incorporation problems</th>
<th>% of total source incorporation problems</th>
<th>% extended source incorporation problems in this category</th>
<th>Number of lab reports containing this source incorporation problem</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patchwriting</td>
<td>201</td>
<td>59%</td>
<td>33%</td>
<td>101 (40%)</td>
</tr>
<tr>
<td>Technical parroting</td>
<td>94</td>
<td>28%</td>
<td>78%</td>
<td>64 (25%)</td>
</tr>
<tr>
<td>Copying</td>
<td>30</td>
<td>9%</td>
<td>20%</td>
<td>23 (9%)</td>
</tr>
<tr>
<td>Falsification</td>
<td>16</td>
<td>5%</td>
<td>19%</td>
<td>13 (5.5%)</td>
</tr>
<tr>
<td>Total instances of source incorporation problems</td>
<td>341</td>
<td>100%*</td>
<td>41.5%*</td>
<td>136 (53%)</td>
</tr>
</tbody>
</table>
Table 5. Difference between parroting or patchwriting and writing in one’s own words.

<table>
<thead>
<tr>
<th>Source text</th>
<th>Student text with problematic source incorporation</th>
<th>Student text in their own words</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Three to four worms</strong> will be transferred to plates coated with a specific strain of bacteria HT115(DE3) which contains the gene for T7 polymerase. The expression of the T7 polymerase is under the control of a lac promoter and operator (so just like the pET vector system). The HT115(DE3) bacteria also contain a plasmid that has an amp resistance gene, and will either contain no RNAi insert (control), or a 800 bp sequence for the unc-22 gene in the polylinker region. Source: Lab manual (Butler and Noree, 2012)</td>
<td>About 3-4 worms were put on plates coated with a strain of bacteria HT115 (DE3) that contains the gene for T7 polymerase. The bacteria contained a plasmid that had an amp resistance gene and either no RNAi insert or a 800 bp sequence for the unc-22 gene in the polylinker region.</td>
<td>On plates of <em>C. elegans</em> including bacterial strain HT115(DE3) containing a <em>T7 polymerase</em> gene, samples have either plasmids with no RNAi insert (control) or the <em>unc-22</em> sequence between the two T7 promoters of a L440 double-T7 vector (experimental). These plates were confirmed to have three to four large worms, with at least two alive.</td>
</tr>
</tbody>
</table>
| **“LuxI and LuxR form a “quorum sensing” regulatory circuit that induces bioluminescence at high cell density”**
Source: journal article, Bose et al., 2008, p. 26.                                                                                                                                                                           | Luminescent expression in bacteria is dependent on cell density and the *luxR* and *luxI* form the quorum sensing regulatory circuit that induces bioluminescence at high cell density. 
*Quororn sensing is a system of stimulus and response correlated to population density.*)                                                                 | The *lux* operon is a very intricate system, that is responsible for the emission of light. …*Lux I* codes for the autoinducer, which when bound by the regulator protein produced by *luxR*, binds to the promoter region to greatly increase transcription of the lux operon. *LuxR* happens to be located directly to the left of the *lux* operon, and contains its own promoter region. |
| **“…system of stimulus and response correlated to population density “**
Table 6. Examples of patchwriting from the primary literature that distorted the meaning of the original text.

<table>
<thead>
<tr>
<th>Source text</th>
<th>Problematic student text</th>
<th>Notes</th>
</tr>
</thead>
<tbody>
<tr>
<td>The host organism can use light emitted by bacteria <strong>for attraction of prey, escape from predators or intra species communication</strong> ... From: Czyz et al., 2000 (Introduction section, 2nd paragraph)</td>
<td>Bacterial species use luminescence either for the attraction of prey, escape from predators or even communication between species.</td>
<td>In the original text, it is the host and not the bacteria that use bioluminescence for the described purposes.</td>
</tr>
<tr>
<td>A polyclonal antibody raised against an <em>Escherichia coli</em> beta-galactosidase-unc-22 fusion protein recognizes a polypeptide in nematode extracts that is between 500,000 and 600,000 daltons and labels the muscle A-band in indirect immunofluorescent microscopy. From: Moerman et al., 1988 (Abstract)</td>
<td><em>unc</em>-22 gene yields a polypeptide <strong>that is between 500,000 and 600,000 Daltons</strong> and immunofluoresces at the muscle-A band.</td>
<td>In the original article it is the fluorescently labeled antibody that labels <em>unc</em>-22 protein and allows detection. In the student’s paper the protein itself immunofluoresces.</td>
</tr>
</tbody>
</table>